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Graphic abstract



PF74: R = H; R' = Me

- EC₅₀ = 0.61 μM
 Stabilized CA hexamer
- t_{1/2} (HLMs) = 0.7 min

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Graphic abstract



Highlights

- Design of novel analogs and sub-chemotypes of HIV-1 CA-targeting antiviral **PF74**.
- 5-Hydroxyindole analogs (8,9 and 12) showed much improved potency over PF74
- 2-Indolone analogs (16-24) decreased the T_m of CA hexamers
- The potencies of α and β -naphthyl analogs (33 and 27) were comparable to **PF74**

Novel HIV-1 Capsid-Targeting Small Molecules of the PF74 Binding Site

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Abstract: The PF74 binding site in HIV-1 capsid protein (CA) is a compelling antiviral drug target. Although PF74 confers mechanistically distinct antiviral phenotypes by

competing against host factors for CA binding, it suffers from prohibitively low metabolic stability. Therefore, there has been increasing interest in designing novel sub-chemotypes of **PF74** with similar binding mode and improved metabolic stability. We report herein our efforts to explore the inter-domain interacting indole moiety for designing novel CA-targeting small molecules. Our design includes simple substitution on the indole ring, and more importantly, novel sub-chemotypes with the indole moiety replaced with a few less electron-rich rings. All 56 novel analogs were synthesized and evaluated for antiviral activity, cytotoxicity, and impact on CA hexamer stability. Selected analogs were tested for metabolic stability in liver microsomes. Molecular modeling was performed to verify compound binding to the **PF74** site. In the end, 5-hydroxyindole analogs (**8**,9 and **12**) showed improved potency (up to 20-fold) over **PF74**. Of the novel sub-chemotypes, α - and β -naphthyl analogs (**33** and **27**) exhibited sub micromolar antiviral potencies comparable to that of **PF74**. Interestingly, although only moderately inhibiting HIV-1 (single-digit micromolar EC₅₀s), analogs of the 2-indolone sub-chemotype consistently lowered the melting point (T_m) of CA hexamers, some with improved metabolic stability over **PF74**.

Keywords: HIV-1; capsid-targeting antivirals; PF74; metabolic stability

1. Introduction

Although dozens of antivirals and fixed dose combinations [1] have been approved to treat the infection of human immunodeficiency virus type 1 (HIV-1), these drugs are not curative and HIV-1 remains a global healthcare challenge. Managing HIV-1 infection requires lifelong treatment, and hence, the virus will eventually develop resistance to current drug classes. Combating resistant viruses entails antivirals with novel molecular targets. The multifunctional HIV-1 capsid protein (CA) [2-4], which is a key component of the HIV-1 gag polyprotein [5], represents an emerging and highly attractive target in HIV-1 drug discovery

[6-8]. CA is the building block of the mature HIV-1 capsid core [9, 10], and core stability critically depends on CA-CA interactions. Disrupting these interactions in the early stages of viral replication can perturb core stability to result in premature uncoating, impaired reverse transcription, and loss of infection [11, 12]. In addition, CA also interacts with multiple cellular factors [13] including TRIM5 α [14, 15], cleavage and polyadenylation specific factor 6 (CPSF6) [16, 17], nucleoporins 153 [18-20] and 358 [21, 22] (NUP153, NUP358), MxB [23, 24], and Cyclophilin A (CypA) [25-27]. These CA-host interactions regulate multiple post entry events, such as uncoating, cytoplasmic trafficking, reverse transcription, nuclear transport, integration site distribution, and the evasion of innate immunity [28]. During the late stage of viral replication, CA-CA interactions also drive the assembly and maturation of new infectious viral particles [29]. Therefore, CA-targeting small molecules could confer both early and late stage antiviral phenotypes.



Figure 1. Compound binding sites and chemotypes targeting HIV-1 CA. (A) Ligand binding sites. Three distinct small molecule binding sites at CA_{NTD} are known for C-4, **PF74** and BM-4, respectively. Host factor cyclophilin A binds to the loop on top. Polypeptide CAI binds to the CA_{CTD}. Binding modes were reproduced in Maestro based on PDB 4XFZ [30] with ligands C-4 & BM-4 (PDB: 4E92 [31]) and CAI (PDB: 2BUO [32]) aligned. The picture was created in PyMOL while shadows of binding sites were rendered in PowerPoint; (B) major chemotypes binding to each site: BI-2 and GS-CA1 bind to the **PF74** site; BD-3 and CAP-1 bind to the BM-4 site; arylquinazoline (AQ) and ebselen bind to CA_{CTD}.

Efforts targeting HIV-1 CA have identified a few chemotypes with distinct binding sites (Figure 1) [6, 7, 33]. CA is highly helical consisting of an N terminal domain (CA_{NTD}) and a C terminal domain (CA_{CTD}) with a flexible linker in-between [30, 34, 35]. Although CA_{CTD}-targeting compounds, such as polypeptide CAI [32], arylquinazoline (AQ) [36], and the covalently binding ebselen [37], have been reported, considerably more efforts have been directed toward targeting CA_{NTD}. At the base of the CA_{NTD} is the binding site for three chemical classes represented by BM-4 [31], BD-3 [31] and CAP-1 [38]. Interestingly, the backbone of BM-4 is a benzoimidazole core flanked by a pyrazole ring and a phenyl ring (Figure 1B). However, when the flanking moieties are reversed, the resulting compound C-4 [39] binds to a completely different site near the top of the CA_{NTD} and around the base of the cyclophilin A binding loop [40] (Figure 1A). In general, compounds binding to these two sites perturbed CA assembly *in vitro* and moderately inhibited HIV-1 in cell culture [6]. There was no evidence that they act in the early steps of viral replication, and no direct evidence linking the antiviral activity to the targeting of CA. By contrast, by far the most interesting CA-targeting compounds are the three chemical classes that bind to the **PF74**

binding site, which also include the BI compounds [41] and the GS-CA compounds [42] (Figure 1B). These compounds all inhibited HIV-1 at both the early and the late stages of viral replication. Not surprisingly, there appears to be a positive correlation between the antiviral potency and the structural complexity: the simplest BI compounds exhibited moderate (low micromolar) antiviral activity and the structurally highly complex GS-CA compounds demonstrated sub-nanomolar antiviral activity, whereas the structurally elaborate, yet highly synthetically accessible peptidomimetic **PF74** inhibited HIV-1 in the sub-micromolar range. The binding mode [30] and the activity profile [43, 44] of **PF74** have been particularly well-characterized, revealing a dual antiviral mechanism of action: at low concentrations **PF74** competes against host factors for capsid binding; at high concentrations it induces premature uncoating, and consequently, impairs reverse transcription [11].



Figure 2. Binding mode of **PF74** based on PDB 4XFZ [30] and the design of novel analogs. (A) Detailed molecular interaction network of **PF74** (magenta sticks) with residues (lines) in both the CA_{NTD} (transparent cyan surface) and the adjacent CA_{CTD} (blue dot). The indole

moiety of **PF74** (boxed) interacts with Q63, K70 of CA_{NTD} and Y169, R173, and K182 of the adjacent CA_{CTD} ; (B) Novel analogs designed to explore the two-domain interactions of the indole moiety.

The **PF74** binding pocket is lined with residues in H3 and H4 of the CA_{NTD} (cyan) and H8 and H9 of the adjacent CA_{CTD} (blue), where an extensive network of molecular interactions defines the PF74 binding mode (Figure 2A) [30]. Structurally, PF74 features a phenylalanine core, connected by an aniline moiety at the carboxylate end and an indole-3-acetic acid at the amino end (Figure 2B). Although all three components provide key molecular interactions for CA binding, the indole ring (boxed, Figure 2A) uniquely interacts with both the CA_{NTD} (via H-bond with Q63 and π -cation interaction with K70) and the adjacent CA_{CTD} (via Y169, R173 and K182) [30]. These interactions are lacking with the BI compounds, which do not have a structural equivalent to the indole moiety of **PF74**, likely accounting for their weaker antiviral potency. The core and the aniline moiety of **PF74** engage extensively with the CA_{NTD} [30] (Figure 2A): 1) the phenylalanine core forms two H-bonds with N57, and an additional one with K70; 2) the aniline moiety makes contact with N53 (via the N-methyl group) and A105, T107, and Y130 (via the phenyl ring); and 3) the phenyl ring of the phenylalanine core forms hydrophobic interactions with residues M66 and L69. Since the indole moiety provides the unique inter-domain binding interactions, there have been reported efforts replacing the indole moiety with a simpler and synthetically more accessible 1,2,3-triazole ring, though the replacement resulted in substantially decreased potency (by >10-fold) [45, 46]. However, a recent report showed that replacing the indole ring with a piperazinone moiety yielded compounds with better antiviral activity (up to 6-fold) than PF-74 [47]. We have previously conducted a comprehensive SAR on PF74 with the synthesis of a large number of analogs [48], and have identified a structurally novel and metabolically stable CA-targeting small molecule [49]. In the current report, we describe our

own efforts targeting the two-domain interactions (Figure 2B). We first synthesized a series of **PF74** analogs with a substituent on the indole ring, and then designed and synthesized analogs of a few novel sub-chemotypes, each featuring a distinct ring system to replace the indole. In the end, 56 analogs were synthesized and tested, many of which demonstrated significant anti-HIV-1 activity and interesting effects on CA hexamer stability, and some exhibited improved metabolic stability over **PF74**.

2. Results and discussion

2.1 Chemistry

Briefly, commercially available (*tert*-butoxycarbonyl)-*L*-phenylalanine (**58**) was treated with various amines under a well-established method using T_3P or HATU as the coupling agent in the presence of DIPEA as base to afford **59**. After removal of Boc protecting group using TFA, amine salts **60** were obtained which were further reacted with commercially available acids **61** to produce analogs **1-57**. Details about the synthesis of **59** and **60** were described in the Supporting Information.

Scheme 1. Synthesis of novel PF74 analogs.



Reagents and conditions: (a) amine, HATU (or T₃P), DIPEA, DMF, rt, 12 h; (b) TFA; DCM, rt, 4-6 h; (c) HATU, DIPEA, DMF, rt, 12 h.

2.2 Biological assays and SAR

All analogs were first evaluated in a thermal shift assay, where the effect of a compound on protein stability was measured by the change in protein melting point compared to DMSO control (Δ Tm). A positive value in Δ Tm indicates a stabilizing effect and a negative value a destabilizing effect on the protein. Of note, the CA protein used in the thermal shift assay is in a covalently crosslinked hexameric state. Thus, the Δ Tm values likely reflect local changes that may affect stabilization and exclude inter-hexamer effects, which are important correlates of overall capsid core stability. To simplify presentation of the data, we refer to the effects of these compounds as stabilization or destabilization of "CA hexamer." All compounds were also screened at 20 μ M in a cell-based antiviral assay against HIV-1. Compounds demonstrating significant inhibition were then tested at 2 μ M. Promising compounds were further assessed in dose response fashion for antiviral EC₅₀ values. All compounds were also tested for cytotoxicity either by screening at 100 or 50 μ M, or determination of CC₅₀ values. For some compounds, % inhibition at two concentrations (2 μ M and 20 μ M) was reported instead of the EC₅₀. **PF74** was resynthesized and tested in these assays (1, Δ Tm = 7.4 °C, EC₅₀ = 0.61 μ M, CC₅₀ = 76 μ M).

2.2.1. Substitutions at the indole $(R^1 \text{ and } R^2)$ and aniline ring (R^3)

This series features **PF74** analogs with the indole ring substituted (R¹) with an OMe (**3-7**) or an OH (**8-15**). It is noteworthy that all these analogs lack the methyl group at R² (R² = H). This is because when compared to **PF74** (R² = Me, Δ Tm = 7.4 °C, EC₅₀ = 0.61 μ M) analog **2** (R² = H, Δ Tm = 6.1 °C, EC₅₀ = 0.46 μ M) showed a largely comparable activity profile. Hence, the SAR within this series concerns mainly the effect of the H-bond enabling and electron donating groups at R¹, in combination with the substitution effects on the aniline ring (\mathbb{R}^3). Notably, an OMe substitution at \mathbb{R}^1 by itself did not improve the activity profile (3) vs 2), though slight improvement was observed when R^3 is a *para*-Cl. This chlorine effect was significant as revealed by the direct comparison between analogs 7 ($R^3 = 4$ -Cl, $\Delta Tm =$ 8.0 °C, EC₅₀ = 0.31 μ M) and **3** (R³ = H, Δ Tm = 4.9 °C, EC₅₀ = 0.56 μ M), possibly due to a halogen bond (see molecular modeling for similar analogs). Among the compounds, 8-15, substituted at R^1 with OH, were found the most potent ones of the series. Particularly, analogs 8 (EC₅₀ = 0.053 μ M), 9 (EC₅₀ = 0.035 μ M) and 12 (EC₅₀ = 0.032 μ M) conferred up to ~20-fold higher potency than **PF74** (EC₅₀ = 0.61 μ M) and analog 2 (EC₅₀ = 0.46 μ M). Furthermore, 9 (CC₅₀ > 100 μ M) and 12 (CC₅₀ > 100 μ M) showed lower toxicity than PF74 $(CC_{50} = 76 \mu M)$. These results suggest that an R¹ substituent capable of both H-bond donating and H-bond accepting (e.g. OH) confers better antiviral potency than does an H-bond accepting group (e.g. OMe). However, the significantly improved antiviral potency with 8, 9 and 12 over PF74 is not correlated with results from the thermal shift assay, where very similar Δ Tm (6.3-6.8 °C for 8, 9, 12 vs 7.4 °C for PF74) was observed. It is possible that the afore-mentioned OH group could affect capsid assembly via unknown post-binding molecular mechanisms. In addition, PF74 is known to inhibit both the early and the late stages of HIV-1 replication cycle. Mechanistic studies are currently underway to determine if the most potent compounds (e.g. 8, 9, 12) from this series display similar antiviral profile.

Table 1. Anti-HIV-1 activity, cytotoxicity, and CA hexamer stability profiles of **PF74** analogs (modifications at R^1 , R^2 , R^3).



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	n!	\mathbf{p}^2	D ³		<i>CC</i> ₅₀	TSA ^c
Compound	R	R	R	EC ₅₀ "(μΜ)	$^{b}(\mu M)$	ΔTm (°C)
PF74 (1)	Н	Me	Н	0.61 ± 0.2	76 ± 9	7.4
2	Н	Н	Н	0.46 ± 0.1	> 100	6.1
3	OMe	Н	Н	0.56 ± 0.3	48 ± 5	4.9
4	OMe	Н	4-Me	0.28 ± 0.02	> 50	5.4
5	OMe	Н	4-F	0.57 ± 0.02	> 50	6.6
6	OMe	Н	3-F	0.99 ± 0.002	> 50	4.2
7	OMe	Н	4-Cl	0.31 ± 0.02	> 50	8.0
8	OH	Н	Н	0.053 ± 0.02	55 ± 2	6.8
9	OH	Н	4-Me	0.035 ± 0.004	> 100	6.3
10	OH	Н	4-F	0.46 ± 0.03	> 50	3.1
11	ОН	Н	3-F	0.32 ± 0.02	47 ± 6	4.4
12	ОН	Н	4-Cl	0.032 ± 0.01	> 100	6.6
13	ОН	Н	3-Cl	0.31 ± 0.01	> 100	6.9
14	OH	Н	3-Br	0.38 ± 0.1	> 100	5.8
15	OH	Н	3-CF ₃	0.48 ± 0.02	65 ± 18	6.8

^a Concentration of compound inhibiting HIV-1 replication by 50%, expressed as the mean \pm standard deviation from at least two independent experiments.

 b Concentration of compound causing 50% cell death, expressed as the mean \pm standard deviation from at least two independent experiments.

^c TSA: thermal shift assay. Δ Tm: change of CA hexamer melting point in presence of compound compared to DMSO control.

2.2.2. Novel sub-chemotypes with indole bioisosteres (R^5) and modifications at the aniline ring (R^3 and R^4)

Table 2 summarizes our SAR studies on a few novel sub-chemotypes featuring different ring structures as the indole replacement. The first new sub-chemotype includes nine analogs (16-24) in which the indole moiety is replaced by an indolin-2-one ring. The first four analogs (16-19) bear a 7-methyl group and the next five (20-24) are 4,7-dimethyl substituted. Two prominent SAR trends were observed: the additional 4-methyl group (20-24) led to improved antiviral potency (20 vs 16, 22 vs 19, 23 vs 17); and both 3-Cl and 4-Cl on the aniline moiety (R³) also enhanced potency (18 vs 16, 24 / 23 vs 20). A beneficial chlorine effect conferred by both a 3-Cl and a 4-Cl of the aniline moiety (R³) was observed with this sub-chemotype only (see molecular modeling). With PF74 and all other sub-chemotypes, only the 4-Cl is properly positioned for halogen bonding. Overall, analogs of this series exhibited low micromolar potencies, and more intriguingly, seem to destabilize the CA hexamer. The next series consists of twelve analogs (25-36) with the indole moiety replaced by a naphthyl ring, either via a β substitution (25-30) or an α substitution (31-36). The substitution site (β vs α) did not appear to impact antiviral activity (32 vs 26, 33 vs 27) as prominently as the R^3 group, where a 4-Me (26) or 4-Cl (27) conferred submicromolar activities, though in the α series (31-36) two additional compounds (31 and 36, $R^3 = H$) also demonstrated low micromolar antiviral activity. Next we synthesized seven analogs (37-43) where R^5 is a 1,3-dimethyl-purine-2,6-dione ring system. Unfortunately, none of them inhibited HIV-1 significantly at 2 µM. Discernible impact on CA hexamer stability was not observed either. The last two sub-chemotypes (44-57) synthesized both feature a [6,5] spiro ring system. Overall, analogs of these two sub-chemotypes did not significantly impact CA

hexamer stability. As for antiviral potency, while many of the bicyclic analogs (44-53) inhibited HIV-1, such as compounds 44, 46, 47, 50, and 53, the four tricylic analogs (54-57) did not show any antiviral activity.

Table 2. Anti-HIV-1 activity, cytotoxicity, and CA hexamer stability profiles of novel analogs $(\mathbb{R}^3, \mathbb{R}^4, \mathbb{R}^5)$.

Compd Code	R^5	R^4	R ³	inhibition % at 2 μM / 20 μM	EC ₅₀ ^a (μΜ)	CC ₅₀ ^b (µМ)	TSA ^c ΔTm (°C)
PF74 (1)	HN	Me	Н	91/98	0.61 ± 0.2	76 ± 9	7.4
16	HN	Me	Н	0/91	ND	> 50	-1.2
17	HN O	Me	4-Cl	17/96	ND	~ 50	-0.4
18	HN O	Me	3-Cl	14/85 ^d	3.3 ± 0.3	> 100	-1.7
19	HN	Me	3-F	0/93	ND	> 50	-0.9
20	HN for	Me	Н	0/72 ^d	4.9 ± 0.3	> 100	-1.8

$ \begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & $	

21	HN	Me	4-F	0/70 ^d	5.4 ± 0.5	> 100	-1.7
22	HN C	Me	3-F	0/70 ^d	5.2 ± 0.4	60 ± 6	-1.8
23	HN O	Me	4-Cl	10/98 ^d	2.8 ± 0.1	63 ± 13	-1.3
24	HN O	Me	3-C1	27/99 ^d	2.1 ± 0.2	43 ± 2	-2.4
25	, t	Me	н	40/98	ND	< 50	4.8
26	CC &	Me	4-Me	80/99	0.99 ± 0.005	< 50	6.4
27	CO.s.	Me	4-C1	95/100	0.63 ± 0.02	< 50	7.0
28	C ,t	Me	3-Cl	0/94	ND	< 50	4.3
29	C ,t	Me	3-F	0/97	ND	< 50	5.2
30	C , i	Et	Н	46/97	ND	< 50	4.3
31	٤-	Me	Н	66/100	1.1 ± 0.04	< 50	6.0
32	<u>}</u>	Me	4-Me	96/100	1.0 ± 0.2	< 50	5.7

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33	٤-	Me	4-Cl	98/100	0.83 ± 0.03	< 50	7.2	
34	ξ- ξ-	Me	3-Cl	53/96	ND	< 50	4.4	
35	٤-	Me	3-F	44/97	ND	< 50	5.4	
36	ξ- ξ-	Et	Н	53/98	1.8 ± 0.09	< 50	4.9	
37		Me	Н	10/94	ND	> 50	0.9	
38	N N K	Me	4-Me	18/98	ND	> 50	1.6	
39		Me	4-F	14/85	ND	> 50	0	
40		Me	3-F	1/67	ND	> 50	0.5	
41		Me	4-Cl	6/96	ND	> 50	1.5	
42		Me	3-Cl	0/71	ND	> 50	0	

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43	Et	Н	0/94	ND	> 50	0.8
44	Me	Н	-/89	8.9 ± 1.5	>100	1.3
45	Н	Н	-/0	>20	>100	0
46	Me	4-OMe	-/93	5.1 ± 2	>100	1.8
47	Me	4-Me	-/92	1.9 ± 0.5	>100	2.7
48	Me	4-F	9/99	ND	> 50	3.1
49	Me	3-F	24/98	ND	> 50	1.3
50	Me	4-Cl	-/96	2.5 ± 0.5	>100	2.2
51	Me	3-Cl	9/98	ND	< 50	3.3
52	Me	3-Br	-/36	> 20	> 50	1.5
53	Et	Н	-/92	7.6 ± 0.9	>100	2.0
54	Me	Н	-/32	> 20	> 100	0



^a Concentration of compound inhibiting HIV-1 replication by 50%, expressed as the mean \pm standard deviation from at least two independent experiments.

 $^{\rm b}$ Concentration of compound causing 50% cell death, expressed as the mean \pm standard deviation from at least two independent experiments.

^c TSA: thermal shift assay. Δ Tm: change of CA hexamer melting point in presence of compound compared to DMSO control.

^d For these compounds, % inhibition is reported at 1 μ M / 10 μ M.

2.3. Metabolic stability in liver microsomes

Despite its potent antiviral activity, well-characterized mode of binding and easy synthetic accessibility, **PF74** is seriously flawed as an antiviral lead due to its prohibitively poor metabolic stability. In human liver microsomes (HLMs), the half-life $(t_{1/2})$ of **PF74** was less than 1 min [46, 50]. We have previously tested around 20 **PF74** analogs in HLMs and they all showed very poor metabolic stability $(t_{1/2} = 0.6-2.1 \text{ min})$ [48]. However, an analog featuring a quinazoline-2,4-dione as the indole replacement was found to be metabolically stable in HLMs $(t_{1/2} = 31 \text{ min})$ [49]. To gauge the metabolic stability of the sub-chemotypes described herein, we tested selected compounds in HLMs and mouse liver microsomes (MLMs) and the results are

summarized in Table 3. Consistent with literature reports, the $t_{1/2}$ of **PF74** in our metabolic assays was 0.7 min in HLMs and 0.6 min in MLMs. Peptidomimetics, a compound class to which PF74 belongs, are particularly liable toward phase I metabolism, presumably because they are good substrates for liver metabolizing enzyme subfamily cytochrome P450 3A (CYP3A) [51]. This extensive liver metabolism constitutes a major pharmacokinetic (PK) barrier for many FDA-approved HIV-1 protease inhibitors [52]. For example, darunavir was reported to have a half-life $(t_{1/2})$ of 3.6 min in a human liver microsomal incubation system with less CYP protein content compared to ours (0.1 mg/mL vs 0.5 mg/mL) [53]. The main mitigating strategy has been to co-administer the antiviral drugs with a CYP3A inhibitor, such as ritonavir [54] or Cobicistat (Cobi) [55], as a PK enhancer [52]. Therefore, our metabolic stability assays were performed under two distinct sets of conditions: with PK enhancer Cobi or without Cobi (Table 3). Without Cobi, PF74 analogs (3 and 7) exhibited very short half-life (1.2 min and 1.8 min, respectively). The metabolic stability remained poor for analogs of novel sub-chemotypes, though a few of them, such as 16, 24, 37 and 46, did show substantially (7 to 10-fold) improved half-life over **PF74** in HLMs. For the five selected compounds tested in the presence of PK enhancer Cobi, drastically improved metabolic stability was observed with all (Table 3). Collectively, these results demonstrated that replacement of the indole moiety with a less electron-rich ring could lead to metabolically more stable compounds, and that inhibiting CYP3A could mitigate the phase I metabolic liability of this type of compounds.

Table 3. Phase I metabolic stability in liver microsomes $t_{1/2}$ (min).

C		HLM ^a (+Cobi	1	1
Compound	HLM ^a	°)	MLM ^b	MLM ^b (+Cobi ^c)

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PF74 ^c	0.7	91	0.6	34
3	1.2		0.5	
7	1.8		0.6	
16	7.0		1	
20	1.5		0.6	
24	6.7	107	1.4	34
25	1.8		0.5	
27	2.7	>120	1.4	18
31	0.6		0.5	
33	1.1	102	0.6	7.4
37	4.7		1.2	
46	5.2		0.7	
47	3.3		1.2	
48	1.1		0.5	
50	2.5	>120	1.1	20

^a HLM: human liver microsome

^b MLM: mouse liver microsome

^c Microsomal stability measured in the presence of CYP3A inhibitor Cobi.

2.4 Molecular modeling

To evaluate the binding mode of the new analogs, we performed molecular modeling of key compounds based on the co-crystal structure of native HIV-1 capsid protein bound to **PF74** (PDB code: 4XFZ [30]). Analog **12** is found to orient in a similar fashion as the parent compound **PF74** (Figure 3A), and share common key interactions on CA_{NTD} domain including (i) hydrogen bonding with N57 via the carbonyl and NH of phenylalanine core, with Q63 via indole N-H, and with K70 via the other carbonyl moiety; (ii) Cation- π

interaction with K70 via the indole moiety (Figure 3A). However, unlike **PF74**, the two additional substituents of **12**, 4-Cl on the aniline moiety and 5-OH of the indole ring, confer a halogen bond (pink dashed line) and another hydrogen bond with K182 on the adjacent CA_{CTD} domain (black dotted line), respectively (Figure 3A). These additional interactions are consistent with improved target binding affinity (Glide score: -7.6 kcal/mol for 12 vs -5.8 kcal/mol for PF74) and largely increased potency (EC₅₀: 0.032 μ M for 12 vs 0.61 μ M for **PF74**). Importantly, while the halogen bond conferred by the 4-Cl of the aniline moiety appeared to be less important for potency than the hydrogen bond by the 5-OH (2 vs 8 vs 12), moving the Cl from 4 position to 3 (compound 13) led to a 10-fold potency drop, likely due to the potential steric clash posed by the 3-Cl. Similar effects were observed with 3-Br (14) and $3-CF_3$ (15). The potency difference between 3-Cl and 4-Cl aniline analog was prominent throughout SAR studies (27 vs 28, 33 vs 34, and 50 vs 51). The only exception is with the 4,7-dimethylindolin-2-one sub-chemotype where a beneficial halogen effect was observed for both the 4-Cl analog 23 (Figure 3B) and the 3-Cl analog 24 (Figure 3C). In both cases, the loss of the key H-bonding with Q63 and the cation- π interactions with K70, was compensated by a halogen-bond (pink dashed line) with N74 (4-Cl) and N53 (3-Cl), respectively. Similarly, compound 27 having the 2-naphthyl ring in place of indole ring exhibited an identical potency and protein binding affinity to PF74, despite the loss of a key hydrogen-bonding with Q63 (Figure 3D). Possible orientation of the aniline ring of the compound 27 as shown in Figure 3D could improve hydrophobic contacts with N53, G106, and Y130, as a result of halogen bond between G106 and 4-Cl aniline, and could explain the observed potency for this and another similar compound 33.



Figure 3. Docking poses of key compounds based on native HIV-1 capsid protein bound to **PF74** (Glide score: -5.8) (PDB code: 4XFZ [30]). (A) Predicted binding mode of compound **12** (Glide score: -7.6 kcal/mol). (B) Predicted binding mode of **23** (Glide score: -6.4 kcal/mol). (C) Predicted binding mode of **24** (Glide score: -6.8 kcal/mol). (D) Predicted binding mode of **27** (Glide score: -6.1 kcal/mol). Hydrogen-bonding, halogen-bonding and cation- π interactions are depicted as black dotted lines, pink dashed lines, and double headed arrows, respectively. CA with key residues around the binding site, and ligands **12**, **23**, **24**, and **27** are colored grey and violet, respectively. The nitrogen, oxygen, and chlorine atoms are colored blue, red, and green, respectively.

3. Conclusions

In this work, we have designed, synthesized and evaluated 56 compounds consisting of **PF74** analogs and a few novel **PF74**-like chemotypes featuring rings less electron-rich than the indole ring. In the end, 5-hydroxyindole analogs (**8**, **9**, **12**) were up to 20-fold more potent than the prototype **PF74**. Analogs with low to sub micromolar antiviral activities were also identified from all sub-chemotypes, except for the 1,3-dimethyl-purine-2,6-dione sub-chemotype. Of all sub-chemotypes studied herein, the naphthalene ring conferred the best potency as both the α - and β -naphthyl analogs (**33** and **27**) exhibited antiviral potencies comparable to that of **PF74**. Interestingly, the 2-indolone sub-chemotype lowered the T_m of CA hexamers, indicating a unique HIV-1 CA-destabilizing mechanism. The best 2-indolone analog, **24**, also showed 10-fold longer half-life in HLMs than **PF74**. These findings necessitate future medicinal chemistry to further optimize these novel sub-chemotypes, particularly the CA-destabilizing 2-indolone chemotype.

4. Experimental section

4.1 Chemistry

General Procedures. All commercial chemicals were used as supplied unless otherwise indicated. Flash chromatography was performed on a Teledyne Combiflash RF-200 with RediSep columns (silica) and indicated mobile phase. ¹H and ¹³C NMR spectra were recorded on a Varian 600 MHz or Bruker 400 spectrometer. Diastereomeric ratio (dr) was determined by ¹H NMR analysis. Mass data were acquired using an Agilent 6230 TOF LC/MS spectrometer. All NMR and mass spectrometers are located in the shared instrument rooms at the Center for Drug Design, University of Minnesota.

4.1.1. Synthesis of **59** and **60**

Synthesis of intermediates 59 and 60 are described in Supporting Information.

Synthesis

4.1.2.

(S)-N-Methyl-2-(2-(2-methyl-1H-indol-3-yl)acetamido)-N,3-diphenylpropanamide (1)

To a solution of commercially available 2-(2-methyl-1H-indol-3-yl)acetic acid (100 mg, 0.53 mmol, 1 equiv.) in DMF (3 mL), HATU (402 mg, 1.06 mmol, 2 equiv.) and DIPEA (205 mg, 1.59 mmol, 3 equiv.) were added and the mixture was stirred at room temperature for 20 min before (S)-2-amino-N-methyl-N,3-diphenylpropanamide (TFA salt, 235 mg, 0.64 mmol, 1.2 equiv.) was added. The mixture was further stirred overnight at room temperature. Upon completion, H₂O was added and the reaction mixture was extracted with EtOAc (3x30 mL). The organic phases were combined and washed with brine, dried over anhydrous MgSO₄, filtered and concentrated. The product was purified by Combiflash on silica gel using EtOAc/hexane (1:2 to 6:1) as eluent. Yield 70%. ¹H NMR (600 MHz, DMSO- d_6) δ 10.68 (s, 1H), 8.27 (d, J = 7.0 Hz, 1H), 7.38 (dd, J = 17.5, 6.5 Hz, 3H), 7.27 (dd, J = 18.7, 6.7 Hz, 3H), 7.18 (d, J = 7.6 Hz, 1H), 7.12 (s, 3H), 6.98 – 6.89 (m, 1H), 6.84 (d, J = 6.9 Hz, 1H), 6.79 (s, 2H), 4.41 (s, 1H), 3.51 - 3.31 (m, 2H), 3.15 (s, 3H), 2.82 (d, J = 11.5 Hz, 1H), 2.67 (d, J = 11.5 Hz, 1H), 2.11.5 Hz, 1H), 2.23 (s, 3H); 13 C NMR (150 MHz, DMSO- d_6) δ 171.5, 171.0, 143.3, 138.0, 135.4, 133.3, 130.0, 129.2, 128.8, 128.4, 128.2, 128.0, 126.7, 120.3, 118.4, 118.4, 110.5, 105.3, 52.2, 37.6, 37.4, 31.3, 11.7; HRMS (ESI) m/z calcd for $C_{27}H_{26}N_3O_2$ [M – H]⁻ 424.2031, found 424.2029.

(*S*)-2-(2-(1*H*-Indol-3-yl)acetamido)-*N*-methyl-*N*,3-diphenylpropanamide (**2**). The synthetic method was similar to that of compound **1** except that 2-(1*H*-indol-3-yl)acetic acid (100 mg, 0.57 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 252 mg, 0.68 mmol, 1.2 equiv.) were used as starting materials. Yield 65%. ¹H NMR (600 MHz, CD₃OD) δ 7.42 (d, *J* = 7.9 Hz, 1H), 7.37 – 7.31 (m, 4H), 7.18 – 7.05 (m, 5H), 7.03 – 6.94 (m, 3H), 6.75 (d, *J* = 7.3 Hz, 2H), 4.66 (t, *J* = 7.2 Hz, 1H), 3.61 (dd, *J* = 22.4, 15.8 Hz,

2H), 3.16 (s, 3H), 2.87 (dd, J = 13.3, 6.6 Hz, 1H), 2.64 (dd, J = 13.3, 6.6 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 172.7, 171.7, 142.5, 136.7, 136.5, 129.4, 128.7, 128.0, 127.9, 127.2, 127.0, 126.4, 123.5, 121.1, 118.6, 118.0, 110.9, 107.7, 51.9, 37.7, 36.6, 32.2; HRMS (ESI) (-) m/z calcd for C₂₆H₂₄N₃O₂ [M – H]⁻ 410.1874, found 410.1878.

(S)-2-(2-(5-Methoxy-1H-indol-3-yl)acetamido)-N-methyl-N,3-diphenylpropanamide (3).

The synthetic of compound method was similar that except that to 1 2-(5-methoxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (S)-2-amino-N-methyl-N,3-diphenylpropanamide (TFA salt, 217 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. Yield 42%. ¹H NMR (600 MHz, CDCl₃) δ 8.34 (s, 1H), 7.36 -7.31 (m, 3H), 7.26 - 7.24 (m, 1H), 7.13 (t, J = 7.4 Hz, 1H), 7.07 - 7.04 (m, 2H), 6.97 - 6.88(m, 5H), 6.66 (d, J = 7.4 Hz, 2H), 6.30 (d, J = 8.0 Hz, 1H), 4.81 – 4.77 (m, 1H), 3.79 (s, 3H), 3.67 - 3.60 (m, 2H), 3.18 (s, 3H), 2.76 (dd, J = 13.3, 7.0 Hz, 1H), 2.56 (dd, J = 13.3, 7.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 171.3, 171.0, 154.4, 142.4, 136.0, 131.4, 129.7, 129.1, 128.3, 128.2, 127.4, 127.3, 126.7, 124.4, 113.0, 112.1, 108.4, 100.1, 55.8, 51.3, 38.8, 37.6, 33.3; HRMS (ESI) m/z calcd for C₂₇H₂₆N₃O₃ [M – H]⁻ 440.1980, found 440.1983.

(*S*)-2-(2-(5-*Methoxy*-1*H*-*indol*-3-*yl*)*acetamido*)-*N*-*methyl*-3-*phenyl*-*N*-(*p*-*tolyl*)*propanamide* (*4*). The synthetic method was similar to that of compound **1** except that 2-(5-methoxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-3-phenyl-*N*-(*p*-tolyl)propanamide (TFA salt, 225 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. Yield 63%. ¹H NMR (600 MHz, CDCl₃) δ 8.60 (s, 1H), 7.22 (d, *J* = 8.7 Hz, 1H), 7.13 – 7.12 (m, 3H), 7.08 – 7.05 (m, 2H), 6.88 – 6.69 (m, 7H), 6.33 (d, J = 8.2 Hz, 1H), 4.82 – 4.79 (m, 1H), 3.79 (s, 3H), 3.66 – 3.59 (m, 2H), 3.15 (s, 3H), 2.77 (dd, J = 13.3, 7.0 Hz, 1H), 2.58 (dd, J = 13.3, 6.9 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.5, 170.9, 154.3, 139.9, 138.1, 136.1, 131.5, 130.3, 129.2, 128.3, 127.4, 127.0, 126.7, 124.5, 112.8, 112.2, 108.2, 100.1, 55.8, 51.2, 38.8, 37.7, 33.4, 21.1; HRMS (ESI) m/z calcd for C₂₈H₂₈N₃O₃ [M – H]⁻ 454.2136, found 454.2139.

(S)-N-(4-Fluorophenyl)-2-(2-(5-methoxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (5). The synthetic method was similar to that of compound 1 except that 2-(5-methoxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.49 mmol, equiv.) and 1 (S)-2-amino-N-(4-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 228 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. Yield 62%. ¹H NMR (600 MHz, CD₃OD) δ 7.24 (d, J = 8.8 Hz, 1H), 7.17 – 7.12 (m, 3H), 7.06 (s, 1H), 7.01 – 6.98 (m, 3H), 6.86 – 6.76 (m, 5H), 4.57 (t, J = 7.4 Hz, 1H), 3.79 (s, 3H), 3.57 (s, 2H), 3.11 (s, 3H), 2.89 (dd, J = 13.3, 7.5 Hz, 1H), 2.67 (dd, J = 13.3, 7.3 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 174.15, 173.20, 163.35 (d, *J*_{CF} = 246.9 Hz), 155.20, 139.96, 137.82, 133.26, 130.67 (d, *J*_{CF} = 7.8 Hz), 130.19, 129.50, 128.75, 127.92, 125.67, 117.37 (d, $J_{CF} = 23.0 \text{ Hz}$), 113.01 (d, $J_{CF} = 9.3 \text{ Hz}$), 108.97, 101.28, 56.27, 53.23, 39.26, 38.07, 33.66; HRMS (ESI) m/z calcd for C₂₇H₂₅FN₃O₃ [M – H]⁻ 458.1885, found 458.1886.

(S)-N-(3-Fluorophenyl)-2-(2-(5-methoxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (6). The synthetic method was similar to that of compound 1 except that 2-(5-methoxy-1H-indol-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (S)-2-amino-N-(3-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 228 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. Yield 59%. ¹H NMR (600 MHz, CDCl₃) δ 8.36 (s, 1H), 7.31 – 7.26 (m, 2H), 7.18 (t, *J* = 7.3 Hz, 1H), 7.12 – 7.10 (m, 2H), 7.03 – 7.00 (m, 2H), 6.90 – 6.87 (m, 2H), 6.77 – 6.73 (m, 3H), 6.28 (d, *J* = 7.9 Hz, 1H), 4.79 – 4.75 (m, 1H), 3.81 (s, 3H), 3.69 – 3.62 (m, 2H), 3.14 (s, 3H), 2.75 (dd, *J* = 13.1, 8.0 Hz, 1H), 2.61 (dd, *J* = 13.1, 6.5 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 171.3, 170.9, 162.8 (d, *J*_{CF} = 249.0 Hz), 154.4, 143.8, 135.8, 131.5, 130.7 (d, *J*_{CF} = 9.0 Hz), 129.2, 128.4, 127.4, 126.9, 124.4, 123.3, 115.2 (d, *J*_{CF} = 21.0 Hz), 114.7 (d, *J*_{CF} = 22.8 Hz), 113.0, 112.2, 108.3, 100.1, 55.8, 51.4, 39.2, 37.5, 33.4; HRMS (ESI) *m*/*z* calcd for C₂₇H₂₅FN₃O₃ [M – H]⁻ 458.1885, found 458.1887.

(S)-N-(4-Chlorophenyl)-2-(2-(5-methoxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (7). The synthetic method was similar to that of compound 1 except that 2-(5-methoxy-1H-indol-3-yl)acetic acid (100 mg, 0.49 equiv.) mmol, 1 and (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 237 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. Yield 80%. ¹H NMR (600 MHz, CD₃OD) δ 7.25 – 7.22 (m, 3H), 7.17 – 7.12 (m, 3H), 7.06 (s, 1H), 6.99 (s, 1H), 6.82 – 6.75 (m, 5H), 4.57 (t, J = 7.4 Hz, 1H), 3.78 (s, 3H), 3.57 (s, 2H), 3.09 (s, 3H), 2.88 (dd, J = 13.2, 7.7 Hz, 1H), 2.67 (dd, J = 13.2, 7.1 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 174.1, 173.0, 155.3, 142.5, 137.8, 135.0, 133.3, 130.8, 130.2, 129.5, 128.8, 127.9, 125.7, 113.0, 113.0, 109.0, 101.4, 56.3, 53.3, 39.4, 37.9, 33.7; HRMS (ESI) m/z calcd for $C_{27}H_{25}CIN_3O_3$ [M - H]⁻ 474.1590, found 474.1595.

(S)-2-(2-(5-Hydroxy-1H-indol-3-yl)acetamido)-N-methyl-N,3-diphenylpropanamide (8). The synthetic method was similar to that of compound 1 except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100)0.52 mmol, equiv.) mg, 1 and (S)-2-amino-N-methyl-N,3-diphenylpropanamide (TFA salt, 232 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 49%. ¹H NMR (600 MHz, CDCl₃) δ 8.42 (s, 1H), 7.30 – 7.27 (m, 3H), 7.13 – 7.11 (m, 2H), 7.08 – 7.06 (m, 2H), 6.90 (s, 1H), 6.86 – 6.77 (m, 4H), 6.68 (d, J = 7.3 Hz, 2H), 6.60 (d, J = 8.1 Hz, 1H), 4.79 (q, J = 7.3 Hz, 1H), 3.55 (s, 2H), 3.17 (s, 3H), 2.79 (dd, J = 13.4, 7.0 Hz, 1H), 2.59 (dd, J = 13.4, 7.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 171.7, 171.6, 150.4, 142.3, 136.0, 131.4, 129.8, 129.2, 128.3, 128.2, 127.8, 127.3, 126.7, 124.7, 112.5, 112.0, 107.6, 103.0, 51.4, 38.7, 37.8, 33; HRMS (ESI) *m/z* calcd for $C_{26}H_{24}N_3O_3 [M - H]^- 426.1823$, found 426.1826.

(*S*)-2-(2-(5-Hydroxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenyl-N-(p-tolyl)propanamide (9). The synthetic method was similar to that of compound **1** except that 2-(5-hydroxy-1H-indol-3-yl)acetic acid (100 mg, 0.52 mmol, 1 equiv.) and (*S*)-2-amino-N-methyl-3-phenyl-N-(p-tolyl)propanamide (TFA salt, 241 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 80%. ¹H NMR (600 MHz, CDCl₃) δ 8.27 (s, 1H), 7.26 (s, 1H), 7.16 – 7.08 (m, 6H), 6.91 (s, 1H), 6.79 – 6.78 (m, 3H), 6.72 (d, *J* = 7.1 Hz, 2H), 6.53 (d, *J* = 8.2 Hz, 1H), 6.27 (s, 1H), 4.83 – 4.79 (m, 1H), 3.57 (s, 2H), 3.15 (s, 3H), 2.80 (dd, *J* = 13.4, 6.9 Hz, 1H), 2.60 (dd, *J* = 13.4, 7.2 Hz, 1H), 2.33 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 171.7, 171.3, 150.2, 139.7, 138.2, 136.1, 131.4, 130.3, 129.2, 128.3, 127.8, 127.0, 126.7, 124.6, 112.5, 112.0, 107.8, 103.1, 51.2, 38.7, 37.8, 33.2, 21.1; HRMS (ESI) *m*/*z* calcd for C₂₇H₂₆N₃O₃ [M – H]⁻ 440.1980, found 440.1984. (S)-N-(4-Fluorophenyl)-2-(2-(5-hydroxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (10). The synthetic method was similar to that of compound 1 except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 0.52 mmol, equiv.) mg, 1 and (S)-2-amino-N-(4-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 241 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 49%. ¹H NMR (600 MHz, CD₃OD) δ 7.19 – 7.14 (m, 5H), 7.03 – 7.00 (m, 3H), 6.86 – 6.82 (m, 4H), 6.69 (d, J = 8.6 Hz, 1H), 4.56 (t, J = 7.4 Hz, 1H), 3.54 (s, 2H), 3.11 (s, 3H), 2.88 (dd, J = 13.2, 7.7 Hz, 1H), 2.68 (dd, J = 13.2, 7.1 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 174.2, 173.2, 163.4 (d, J_{CF} = 246.9 Hz), 151.5, 140.96, 139.94, 137.8, 133.0, 130.7 (d, $J_{CF} = 8.8$ Hz), 130.3, 129.5, 129.2, 127.9, 125.8, 117.4 (d, $J_{CF} = 23.0 \text{ Hz}$), 112.8 (d, $J_{CF} = 9.7 \text{ Hz}$), 108.3, 103.6, 53.2, 39.4, 38.1, 33.7; HRMS (ESI) m/z calcd for C₂₆H₂₃FN₃O₃ [M – H]⁻ 444.1729, found 444.1732.

(*S*)-*N*-(*3*-*Fluorophenyl*)-*2*-(*2*-(*5*-*hydroxy*-*1H*-*indol*-*3*-*yl*)*acetamido*)-*N*-*methyl*-*3*-*phenylprop anamide* (*11*). The synthetic method was similar to that of compound **1** except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.52 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-fluorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 241 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 39%. ¹H NMR (600 MHz, CDCl₃) δ 8.30 (s, 1H), 7.27 – 7.11 (m, 5H), 6.99 – 6.94 (m, 2H), 6.85 – 6.75 (m, 5H), 6.53 (d, *J* = 8.0 Hz, 1H), 6.47 (s, 1H), 6.32 (s, 1H), 4.78 – 4.74 (m, 1H), 3.59 (s, 2H), 3.12 (s, 3H), 2.78 (dd, *J* = 13.2, 8.1 Hz, 1H), 2.63 (dd, *J* = 13.2, 6.6 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 171.6, 171.5, 162.7 (d, *J*_{CF} = 249.4 Hz), 150.4, 143.6, 135.8, 131.4, 130.8 (d, *J*_{CF} = 9.5 Hz), 129.2, 128.4, 127.7, 126.9, 124.6, 123.3, 115.3 (d, *J*_{CF} = 21.2 Hz), 114.6 (d, *J*_{CF} = 20.9 Hz), 112.6, 112.1, 107.7, 103.0, 51.5, 39.0, 37.6, 33.2; HRMS (ESI) *m*/*z* calcd for C₂₆H₂₃FN₃O₃ [M – H]⁻ 444.1729, found 444.1730.

(S)-N-(4-Chlorophenyl)-2-(2-(5-hydroxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (12). The synthetic method was similar to that of compound 1 except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 0.52 mg, mmol. 1 equiv.) and (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 254 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 60%. ¹H NMR (600 MHz, CDCl₃) δ 8.16 (s, 1H), 7.25 – 7.12 (m, 6H), 7.00 (s, 1H), 6.82 – 6.71 (m, 5H), 6.50 (d, J = 8.0 Hz, 1H), 6.05 (s, 1H), 4.74 - 4.70 (m, 1H), 3.60 (s, 2H), 3.12 (s, 3H), 2.79 (dd, J = 13.2, 8.0 Hz, 1H), 2.62 (dd, J = 13.2, 6.7 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 171.5, 171.4, 150.3, 140.8, 135.9, 134.0, 131.3, 129.8, 129.3, 128.7, 128.4, 127.7, 126.9, 124.6, 112.6, 112.0, 107.8, 103.0, 51.3, 38.9, 37.7, 33.2; HRMS (ESI) m/z calcd for C₂₆H₂₃ClN₃O₃ [M – H]⁻ 460.1433, found 460.1436.

(*S*)-*N*-(*3*-*Chlorophenyl*)-*2*-(*2*-(*5*-*hydroxy*-*1H*-*indol*-*3*-*yl*)*acetamido*)-*N*-*methyl*-*3*-*phenylprop anamide* (*13*). The synthetic method was similar to that of compound **1** except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.52 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 254 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 40%. ¹H NMR (600 MHz, CD₃OD) δ 7.23 (d, *J* = 8.1 Hz, 1H), 7.18 (t, *J* = 7.9 Hz, 1H), 7.12 – 7.06 (m, 5H), 6.96 (s, 1H), 6.77 – 6.74 (m, 4H), 6.60 (d, *J* = 8.6 Hz, 1H), 4.47 (t, *J* = 7.4 Hz, 1H), 3.49 – 3.43 (m, 2H), 3.02 (s, 3H), 2.78 (dd, *J* = 13.1, 8.1 Hz, 1H), 2.60 (dd, *J* = 13.1, 6.7 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 174.2, 173.0, 151.5, 145.1, 137.6, 135.9, 133.1, 131.9, 130.2, 129.6, 129.5, 129.2, 128.7, 128.1, 127.2, 125.8, 112.8, 112.8, 108.3, 103.6, 53.4, 39.5, 37.9, 33.7; HRMS (ESI) m/z calcd for C₂₆H₂₃ClN₃O₃ [M – H]⁻ 460.1433, found 460.1430.

(S)-N-(3-Bromophenyl)-2-(2-(5-hydroxy-1H-indol-3-yl)acetamido)-N-methyl-3-phenylprop anamide (14). The synthetic method was similar to that of compound 1 except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.52 mmol, 1 equiv.) and (S)-2-amino-N-(3-bromophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 282 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 65%. ¹H NMR (600 MHz, CD₃OD) δ 7.47 (d, J = 8.0 Hz, 1H), 7.22 – 7.17 (m, 6H), 7.06 (s, 1H), 6.87 – 6.83 (m, 4H), 6.69 (d, J = 8.6 Hz, 1H), 4.57 (t, J = 7.4 Hz, 1H), 3.59 – 3.53 (m, 2H), 3.10 (s, 3H), 2.87 (dd, J = 13.0, 8.2 Hz, 1H), 2.70 (dd, J = 13.1, 6.7 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 174.2, 173.0, 151.5, 145.2, 137.6, 133.0, 132.4, 132.1, 131.5, 130.2, 129.6, 129.2, 128.1, 127.7, 125.8, 123.6, 112.8, 112.8, 108.3, 103.6, 53.4, 39.5, 38.0, 33.7; HRMS (ESI) m/z calcd for $C_{26}H_{23}BrN_{3}O_{3} [M - H]^{-} 504.0928$, found 504.0930.

(*S*)-2-(2-(5-Hydroxy-1H-indol-3-yl)acetamido)-*N*-methyl-3-phenyl-*N*-(3-(trifluoromethyl)ph enyl)propanamide (15). The synthetic method was similar to that of compound **1** except that 2-(5-hydroxy-1*H*-indol-3-yl)acetic acid (100 mg, 0.52 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-3-phenyl-*N*-(3-(trifluoromethyl)phenyl)propanamide (TFA salt, 275 mg, 0.63 mmol, 1.2 equiv.) were used as starting materials. Yield 49%. ¹H NMR (600 MHz, CD₃OD) δ 7.60 (d, *J* = 7.7 Hz, 1H), 7.47 (t, *J* = 7.8 Hz, 1H), 7.18 – 7.12 (m, 6H), 7.04 (s, 1H), 6.86 (s, 1H), 6.81 (d, *J* = 7.2 Hz, 2H), 6.68 (d, *J* = 8.6 Hz, 1H), 4.52 (t, *J* = 7.4 Hz, 1H), 3.58

- 3.52 (m, 2H), 3.13 (s, 3H), 2.87 (dd, J = 12.9, 8.3 Hz, 1H), 2.68 (dd, J = 13.0, 6.7 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 174.2, 173.1, 151.4, 144.6, 137.6, 133.0, 132.7, 131.8, 130.1, 129.6, 129.10, 129.09, 128.0, 126.0, 125.7, 125.4, 124.9 (q, $J_{CF} = 271.2$ Hz), 112.8, 112.8, 112.7, 108.3, 53.4, 39.5, 38.0, 33.7; HRMS (ESI) m/z calcd for C₂₇H₂₃F₃N₃O₃ [M – H]⁻ 494.1697, found 494.1693.

(2S)-N-Methyl-2-(2-(7-methyl-2-oxoindolin-3-yl)acetamido)-N,3-diphenylpropanamide

(*16*). The synthetic method was similar to that of compound **1** except that 2-(7-methyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 215 mg, 0.58 mmol, 1.2 equiv.) were used as starting materials. (Yield 70%, dr 1.1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.36 – 7.31 (m, 3H), 7.22 – 7.17 (m, 3H), 6.99 – 6.95 (m, 2H), 6.88 – 6.83 (m, 3H), 6.75 (t, *J* = 7.6 Hz, 1H), 6.65 (d, *J* = 7.4 Hz, 1H), 4.73 – 4.63 (m, 1H), 3.72 – 3.69 (m, 1H), 3.18 – 3.15 (m, 1H), 2.96 – 2.82 (m, 2H), 2.71 – 2.65 (m, 1H), 2.54 – 2.47 (m, 1H), 2.22 (s, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 181.8, 181.7, 173.1, 172.3, 172.0, 144.0, 143.9, 142.0, 141.9, 138.2, 138.1, 130.8, 130.5, 130.4, 130.3, 130.2, 130.0, 129.5, 129.3, 128.6, 127.9, 127.8, 123.3, 123.3, 122.9, 122.6, 120.5, 120.4, 53.5, 53.1, 39.5, 39.4, 38.1, 38.0, 37.1, 37.0, 16.7; HRMS (ESI) *m*/*z* calcd for C₂₇H₂₆N₃O₃ [M – H]⁻ 440.1980, found 440.1985.

(2S)-N-(4-Chlorophenyl)-N-methyl-2-(2-(7-methyl-2-oxoindolin-3-yl)acetamido)-3-phenylp ropanamide (17). The synthetic method was similar to that of compound 1 except that 2-(7-methyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 238 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. (Yield 70%, dr 1.1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.30 – 7.21 (m, 5H), 6.99 – 6.86 (m, 4H), 6.81 – 6.75 (m, 2H), 6.62 (s, 1H), 4.63 – 4.55 (m, 1H), 3.73 – 3.68 (m, 1H), 3.13 – 3.08 (m, 3H), 2.96 – 2.82 (m, 2H), 2.71 – 2.53 (m, 2H), 2.23 – 2.22 (m, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 181.8, 181.7, 173.1, 173.0, 172.2, 171.9, 142.6, 142.5, 142.2, 142.0, 138.02, 134.92, 130.72, 130.50, 130.4, 130.3, 130.28, 130.19, 130.1, 130.0, 129.60, 128.0, 123.32, 122.8, 122.6, 120.5, 53.4, 52.9, 44.3, 44.2, 39.71, 39.5, 38.0, 37.8, 37.1, 36.9, 16.7; HRMS (ESI) *m/z* calcd for C₂₇H₂₅ClN₃O₃ [M – H]⁻ 474.1590, found 474.1595.

(2*S*)-*N*-(*3*-*Chlorophenyl*)-*N*-*methyl*-2-(2-(7-*methyl*-2-*oxoindolin*-3-*yl*)*acetamido*)-3-*phenylp ropanamide* (*18*). The synthetic method was similar to that of compound **1** except that 2-(7-methyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 238 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. (Yield 72%, dr 1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.31 – 7.24 (m, 6H), 7.01 – 6.88 (m, 5H), 6.81 – 6.77 (m, 1H), 6.67 (s, 1H), 4.62 – 4.55 (m, 1H), 3.76 – 3.71 (m, 1H), 3.13 – 3.09 (m, 3H), 2.96 – 2.84 (m, 2H), 2.73 – 2.69 (m, 1H), 2.55 – 2.51 (m, 1H), 2.23 (s, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 181.75, 173.0, 172.9, 172.3, 172.0, 145.2, 145.0, 142.1, 142.0, 137.9, 135.9, 131.8, 130.5, 130.3, 130.1, 130.0, 129.7, 129.6, 129.4, 128.8, 128.1, 127.24, 123.4, 123.3, 122.9, 122.7, 122.6, 120.5, 53.6, 53.1, 44.3, 39.8, 39.7, 38.0, 37.8, 37.1, 36.9, 16.7; HRMS (ESI) *m*/*z* calcd for C₂₇H₂₅ClN₃O₃ [M – H]⁻ 474.1590, found 474.1594. (2*S*)-*N*-(*3*-*Fluorophenyl*)-*N*-*methyl*-2-(2-(7-*methyl*-2-*oxoindolin*-3-*yl*)*acetamido*)-3-*phenylp ropanamide* (**19**). The synthetic method was similar to that of compound **1** except that 2-(7-methyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.49 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-fluorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 227 mg, 0.59 mmol, 1.2 equiv.) were used as starting materials. (Yield 69%, dr 1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.34 – 7.22 (m, 4H), 7.08 – 7.05 (m, 1H), 7.00 – 6.87 (m, 5H), 6.79 – 6.73 (m, 2H), 6.57 (s, 1H), 4.67 – 4.59 (m, 1H), 3.75 – 3.70 (m, 1H), 3.15 – 3.11 (m, 3H), 2.97 – 2.82 (m, 2H), 2.73 – 2.68 (m, 1H), 2.55 – 2.50 (m, 1H), 2.22 (s, 3H); HRMS (ESI) *m*/*z* calcd for C₂₇H₂₅FN₃O₃ [M – H]⁻ 458.1885, found 458.1890.

(2S)-2-(2-(4,7-Dimethyl-2-oxoindolin-3-yl)acetamido)-N-methyl-N,3-diphenylpropanamide

(20). The synthetic method was similar to that of compound **1** except that 2-(4,7-dimethyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.46 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 203 mg, 0.55 mmol, 1.2 equiv.) were used as starting materials. (Yield 58%, dr 1.2:1). ¹H NMR (600 MHz, CD₃OD) δ 7.30 – 7.24 (m, 2H), 7.20 – 7.14 (m, 4H), 6.92 – 6.82 (m, 3H), 6.69 – 6.62 (m, 1H), 6.45 (s, 1H), 4.58 – 4.52 (m, 1H), 3.68 – 3.67 (m, 1H), 3.14 – 3.06 (m, 3H), 3.01 – 2.81 (m, 3H), 2.65 – 2.59 (m, 1H), 2.23 – 2.16 (m, 6H); ¹³C NMR (100 MHz, CD₃OD) δ 182.1, 182.0, 173.1, 172.9, 171.8, 171.3, 143.9, 143.6, 142.5, 142.2, 138.1, 133.1, 132.8, 130.8, 130.6, 130.5, 130.4, 130.3, 130.2, 129.4, 129.2, 129.1, 128.5, 127.8, 125.0, 124.9, 117.9, 117.8, 53.1, 52.6, 39.8, 39.5, 38.0, 37.8, 36.1, 36.0, 18.6, 16.4; HRMS (ESI) *m*/*z* calcd for C₂₈H₂₈N₃O₃ [M – H]⁻ 454.2136, found 454.2145.

(2S)-2-(2-(4,7-Dimethyl-2-oxoindolin-3-yl)acetamido)-N-(4-fluorophenyl)-N-methyl-3-phen ylpropanamide (21). The synthetic method was similar to that of compound 1 except that 2-(4,7-dimethyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.46 mmol, 1 equiv.) and (S)-2-amino-N-(4-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 212 mg, 0.55 mmol, 1.2 equiv.) were used as starting materials. (Yield 65%, dr 1.1:1). ¹H NMR (600 MHz, CD₃OD) δ 8.27 – 8.14 (m, 1H), 7.31 – 7.18 (m, 3H), 6.99 – 6.82 (m, 6H), 6.70 – 6.64 (m, 1H), 6.31 (s, 1H), 4.53 – 4.46 (m, 1H), 3.69 – 3.65 (m, 1H), 3.10 – 3.01 (m, 3H), 2.99 – 2.85 (m, 3H), 2.65 – 2.61 (m, 1H), 2.23 – 2.17 (m, 6H); HRMS (ESI) *m*/*z* calcd for C₂₈H₂₇FN₃O₃ [M – H]⁻ 472.2042, found 472.2047.

(2S)-2-(2-(4,7-Dimethyl-2-oxoindolin-3-yl)acetamido)-N-(3-fluorophenyl)-N-methyl-3-phen ylpropanamide (22). The synthetic method was similar to that of compound 1 except that 2-(4,7-dimethyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.46 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-fluorophenyl)-*N* $-methyl-3-phenylpropanamide (TFA salt, 212 mg, 0.55 mmol, 1.2 equiv.) were used as starting materials. (Yield 68%, dr 1:1). ¹H NMR (600 MHz, CD₃OD) <math>\delta$ 7.28 – 7.11 (m, 4H), 7.05 – 6.86 (m, 4H), 6.69 – 6.41 (m, 2H), 6.19 – 6.04 (m, 1H), 4.56 – 4.50 (m, 1H), 3.70 – 3.65 (m, 1H), 3.11 – 3.02 (m, 3H), 3.00 – 2.85 (m, 3H), 2.67 – 2.64 (m, 1H), 2.21 – 2.17 (m, 6H); HRMS (ESI) *m/z* calcd for C₂₈H₂₇FN₃O₃ [M – H]⁻ 472.2042, found 472.2046.

(2S)-N-(4-Chlorophenyl)-2-(2-(4,7-dimethyl-2-oxoindolin-3-yl)acetamido)-N-methyl-3-phe nylpropanamide (23). The synthetic method was similar to that of compound 1 except that 2-(4,7-dimethyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.46 mmol, 1 equiv.) and
(*S*)-2-amino-*N*-(4-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 222 mg, 0.55 mmol, 1.2 equiv.) were used as starting materials. (Yield 72%, dr 1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.24 – 7.18 (m, 4H), 7.08 (d, *J* = 8.6 Hz, 1H), 6.92 – 6.85 (m, 3H), 6.70 – 6.63 (m, 2H), 6.23 (s, 1H), 4.51 – 4.45 (m, 1H), 3.68 – 3.63 (m, 1H), 3.09 – 2.99 (m, 3H), 2.91 – 2.84 (m, 3H), 2.65 – 2.60 (m, 1H), 2.23 – 2.16 (m, 6H); ¹³C NMR (100 MHz, CD₃OD) δ 182.1, 181.9, 173.0, 172.7, 171.8, 171.1, 142.7, 142.5, 142.2, 138.0, 134.9, 134.7, 133.0, 132.8, 130.7, 130.6, 130.5, 130.4, 130.3, 130.1, 129.5, 127.9, 127.8, 127.7, 125.0, 124.9, 117.9, 117.8, 53.0, 52.4, 44.2, 44.1, 39.9, 39.6, 37.9, 37.6, 36.0, 18.6, 16.4; HRMS (ESI) *m*/*z* calcd for C₂₈H₂₇ClN₃O₃ [M – H]⁻ 488.1746, found 488.1750.

(2*S*)-*N*-(*3*-*Chlorophenyl*)-*2*-(*2*-(*4*,7-*dimethyl*-*2*-*oxoindolin*-*3*-*yl*)*acetamido*)-*N*-*methyl*-*3*-*phe nylpropanamide* (*24*). The synthetic method was similar to that of compound **1** except that 2-(4,7-dimethyl-2-oxoindolin-3-yl)acetic acid (100 mg, 0.46 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 222 mg, 0.55 mmol, 1.2 equiv.) were used as starting materials. (Yield 79%, dr 1:1). ¹H NMR (600 MHz, CD₃OD) δ 7.27 – 7.07 (m, 6H), 6.91 – 6.85 (m, 3H), 6.68 – 6.63 (m, 1H), 6.29 (s,1H), 4.51 – 4.46 (m, 1H), 3.70 – 3.65 (m, 1H), 3.07 – 2.99 (m, 3H), 2.96 – 2.84 (m, 3H), 2.66 – 2.63 (m, 1H), 2.21 – 2.16 (m, 6H); ¹³C NMR (100 MHz, CD₃OD) δ 182.0, 181.9, 172.9, 172.7, 171.8, 171.3, 145.0, 144.7, 142.5, 142.1, 137.9, 137.8, 135.8, 135.6, 133.0, 132.8, 131.8, 131.6, 130.5, 130.3, 129.6, 129.3, 128.6, 128.0, 127.8, 127.2, 127.1, 125.0, 124.9, 117.9, 117.8, 53.1, 52.6, 44.2, 39.8, 37.9, 37.7, 36.1, 18.6, 16.5; HRMS (ESI) *m/z* calcd for C₂₈H₂₇ClN₃O₃ [M – H]⁻ 488.1746, found 488.1751. (*S*)-*N*-*Methyl*-2-(2-(*naphthalen*-2-*yl*)*acetamido*)-*N*,3-*diphenylpropanamide* (25). The synthetic method was similar to that of compound **1** except that 2-(naphthalen-2-yl)acetic acid (100 mg, 0.54 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 239 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 79%. ¹H NMR (600 MHz, CD₃OD) δ 8.26 (d, *J* = 7.5 Hz, 1H), 7.81 – 7.74 (m, 3H), 7.64 (s, 1H), 7.46 – 7.42 (m, 2H), 7.35 – 7.34 (m, 3H), 7.28 – 7.27 (m, 1H), 7.15 – 7.03 (m, 4H), 6.81 (d, *J* = 7.2 Hz, 2H), 4.69 – 4.65 (m, 1H), 3.66 – 3.61 (m, 2H), 3.19 (s, 3H), 2.95 (dd, *J* = 13.4, 6.4 Hz, 1H), 2.72 (dd, *J* = 13.4, 8.4 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 173.4, 173.3, 144.0, 138.1, 135.0, 134.2, 133.9, 130.9, 130.1, 129.4, 129.4, 129.1, 128.8, 128.7, 128.6, 128.6, 128.3, 127.8, 127.1, 126.7, 53.6, 43.5, 39.0, 38.1; HRMS (ESI) *m*/*z* calcd for C₂₈H₂₅N₂O₂ [M – H]⁻ 421.1922, found 421.1927.

(S)-N-Methyl-2-(2-(naphthalen-2-yl)acetamido)-3-phenyl-N-(p-tolyl)propanamide (26).The synthetic method similar to of compound except that was that 1 2-(naphthalen-2-yl)acetic acid (100)0.54 equiv.) mg, mmol, 1 and (S)-2-amino-N-methyl-3-phenyl-N-(p-tolyl)propanamide (TFA salt, 249 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 58%. ¹H NMR (600 MHz, CDCl₃) δ 7.83 – 7.77 (m, 3H), 7.63 (s, 1H), 7.49 – 7.45 (m, 2H), 7.28 – 7.26 (m, 1H), 7.14 – 7.05 (m, 5H), 6.79 – 6.75 (m, 4H), 6.17 (d, J = 8.2 Hz, 1H), 4.84 – 4.80 (m, 1H), 3.67 – 3.62 (m, 2H), 3.17 (s, 3H), 2.81 (dd, J = 13.4, 6.9 Hz, 1H), 2.62 (dd, J = 13.4, 7.1 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 170.0, 139.9, 138.1, 136.1, 133.6, 132.5, 132.2, 130.4, 129.2, 128.6, 128.3, 128.1, 127.7, 127.7, 127.3, 127.0, 126.7, 126.2, 125.9, 51.0, 43.8, 38.8, 37.7, 21.1; HRMS (ESI) m/z calcd for C₂₉H₂₇N₂O₂ [M – H]⁻ 435.2079, found 435.2083.

(S)-N-(4-Chlorophenyl)-N-methyl-2-(2-(naphthalen-2-yl)acetamido)-3-phenylpropanamide (27). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-2-yl)acetic acid 0.54 (100)mmol, equiv.) and mg, 1 (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 262 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 70%. ¹H NMR (600 MHz, CD₃OD) δ 7.80 – 7.74 (m, 3H), 7.65 (s, 1H), 7.45 – 7.41 (m, 2H), 7.30 – 7.26 (m, 3H), 7.18 – 7.13 (m, 3H), 6.88 - 6.87 (m, 4H), 4.59 (t, J = 7.5 Hz, 1H), 3.65 - 3.61 (m, 2H), 3.12 (s, 3H), 2.95 (dd, J = 13.3, 7.5 Hz, 1H), 2.74 (dd, J = 13.3, 7.5 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 173.3, 173.2, 142.6, 137.9, 135.0, 134.2, 133.9, 130.8, 130.3, 129.5, 129.1, 128.8, 128.7, 128.6, 128.3, 128.0, 127.1, 126.7, 53.5, 43.4, 39.2, 38.0; HRMS (ESI) *m/z* calcd for C₂₈H₂₄ClN₂O₂ $[M - H]^{-}$ 455.1532, found 455.1531.

(S)-N-(3-Chlorophenyl)-N-methyl-2-(2-(naphthalen-2-yl)acetamido)-3-phenylpropanamide (28). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-2-yl)acetic (100)0.54 equiv.) acid mg, mmol, 1 and (S)-2-amino-N-(3-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 262 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 90%. ¹H NMR (600 MHz, CD₃OD) δ 7.79 – 7.74 (m, 3H), 7.67 (s, 1H), 7.44 – 7.40 (m, 2H), 7.32 – 7.24 (m, 3H), 7.19 – 7.13 (m, 3H), 6.93 - 6.76 (m, 4H), 4.58 (t, J = 7.4 Hz, 1H), 3.64 (s, 2H), 3.11 (s, 3H), 2.95 (dd, J =13.1, 7.9 Hz, 1H), 2.75 (dd, J = 13.1, 7.3 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 173.4, 173.1, 145.1, 137.8, 135.9, 135.0, 134.2, 133.9, 131.9, 130.2, 129.6, 129.4, 129.1, 128.8, 128.7, 128.7, 128.6, 128.3, 128.1, 127.2, 127.1, 126.7, 53.6, 43.4, 39.4, 38.0; HRMS (ESI) m/z calcd for C₂₈H₂₄ClN₂O₂ [M – H]⁻ 455.1532, found 455.1533.

(S)-N-(3-Fluorophenyl)-N-methyl-2-(2-(naphthalen-2-yl)acetamido)-3-phenylpropanamide (29). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-2-yl)acetic acid (100)0.54 mmol, equiv.) mg, 1 and (S)-2-amino-N-(3-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 251 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 79%. ¹H NMR (600 MHz, CD₃OD) δ 7.79 – 7.73 (m, 3H), 7.66 (s, 1H), 7.44 – 7.40 (m, 2H), 7.31 – 7.27 (m, 2H), 7.17 – 7.11 (m, 3H), 7.06 (t, J = 7.9 Hz, 1H), 6.87 – 6.81 (m, 3H), 6.62 (s, 1H), 4.63 (t, J = 7.4 Hz, 1H), 3.66 -3.61 (m, 2H), 3.13 (s, 3H), 2.95 (dd, J = 13.2, 7.5 Hz, 1H), 2.74 (dd, J = 13.2, 7.6 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 173.4, 173.2, 164.2 (d, J_{CF} = 247.4 Hz), 145.4 (d, J_{CF} = 9.7 Hz), 137.9, 135.0, 134.2, 133.9, 132.1 (d, *J*_{CF} = 9.1 Hz), 130.2, 129.5, 129.1, 128.8, 128.7, 128.6, 128.3, 128.0, 127.1, 126.7, 124.7, 116.2 (d, $J_{CF} = 21.3 \text{ Hz}$), 115.9 (d, $J_{CF} = 23.1 \text{ Hz}$), 53.6, 43.4, 39.3, 38.0; HRMS (ESI) m/z calcd for C₂₈H₂₄FN₂O₂ [M – H]⁻ 439.1827, found 439.1830.

(*S*)-*N*-*Ethyl*-2-(2-(*naphthalen*-2-*yl*)*acetamido*)-*N*,3-*diphenylpropanamide* (**30**). The synthetic method was similar to that of compound **1** except that 2-(naphthalen-2-yl)acetic acid (100 mg, 0.54 mmol, 1 equiv.) and (*S*)-2-amino-*N*-ethyl-*N*,3-diphenylpropanamide (TFA salt, 249 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 80%. ¹H NMR (600 MHz, CDCl₃) δ 7.83 – 7.78 (m, 3H), 7.64 (s, 1H), 7.50 – 7.45 (m, 2H), 7.36 – 7.34 (m, 3H), 7.27 – 7.26 (m, 1H), 7.14 (t, *J* = 7.4 Hz, 1H), 7.06 (t, *J* = 7.6 Hz, 2H), 6.91 – 6.86 (m, 1H), 6.73 (d, *J* = 7.3 Hz, 2H), 6.08 (d, *J* = 8.1 Hz, 1H), 4.72 – 4.68 (m, 1H), 3.81 – 3.75 (m, 1H), 3.67 – 3.62 (m, 2H), 3.59 – 3.53 (m, 1H), 2.82 (dd, *J* = 13.4, 6.9 Hz, 1H), 2.60 (dd, *J* = 13.4, 7.1 Hz, 1H), 1.06 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.7,

169.9, 140.8, 136.1, 133.6, 132.5, 132.2, 129.7, 129.3, 128.6, 128.4, 128.3, 128.1, 127.7, 127.7, 127.3, 126.8, 126.2, 125.9, 51.4, 44.6, 43.8, 38.8, 12.8; HRMS (ESI) *m/z* calcd for C₂₉H₂₇N₂O₂ [M – H]⁻ 435.2079, found 435.2081.

(*S*)-*N*-*Methyl*-2-(2-(*naphthalen*-1-*yl*)*acetamido*)-*N*,3-*diphenylpropanamide* (*31*). The synthetic method was similar to that of compound **1** except that 2-(naphthalen-1-yl)acetic acid (100 mg, 0.54 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 239 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 70%. ¹H NMR (600 MHz, CD₃OD) δ 7.86 – 7.83 (m, 2H), 7.76 (d, *J* = 8.2 Hz, 1H), 7.46 – 7.42 (m, 2H), 7.39 – 7.36 (m, 1H), 7.33 – 7.29 (m, 4H), 7.17 – 7.10 (m, 3H), 7.00 (m, 2H), 6.77 (d, *J* = 7.3 Hz, 2H), 4.68 – 4.66 (m, 1H), 3.94 (s, 2H), 3.17 (s, 3H), 2.92 (dd, *J* = 13.4, 6.3 Hz, 1H), 2.70 (dd, *J* = 13.4, 8.4 Hz, 2H); ¹³C NMR (150 MHz, CD₃OD) δ 173.3, 173.2, 143.9, 138.0, 135.3, 133.5, 132.8, 130.8, 130.1, 129.6, 129.4, 129.3, 129.0, 128.8, 128.6, 127.8, 127.3, 126.7, 126.5, 124.9, 53.5, 40.9, 39.0, 38.1; HRMS (ESI) *m/z* calcd for C₂₈H₂₅N₂O₂ [M – H]⁻ 421.1922, found 421.1928.

(S)-N-Methyl-2-(2-(naphthalen-1-yl)acetamido)-3-phenyl-N-(p-tolyl)propanamide (32). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-1-yl)acetic (100)0.54 mmol, equiv.) acid 1 and mg, (S)-2-amino-N-methyl-3-phenyl-N-(p-tolyl)propanamide (TFA salt, 249 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials.

Yield 69%. ¹H NMR (600 MHz, CDCl₃) δ 7.87 – 7.85 (m, 2H), 7.80 (d, J = 8.2 Hz, 1H), 7.50 – 7.41 (m, 3H), 7.35 (d, J = 6.9 Hz, 1H), 7.13 – 7.08 (m, 3H), 7.01 (t, J = 7.6 Hz, 2H), 6.75 (s,

1H), 6.60 (d, J = 7.4 Hz, 2H), 6.08 (d, J = 8.3 Hz, 1H), 4.82 – 4.78 (m, 1H), 3.99 – 3.90 (m, 2H), 3.12 (s, 3H), 2.68 (dd, J = 13.3, 6.9 Hz, 1H), 2.51 (dd, J = 13.4, 7.0 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 169.9, 139.8, 138.1, 136.0, 133.9, 132.1, 131.0, 130.4, 129.1, 128.7, 128.3, 128.2, 128.2, 127.0, 126.6, 126.5, 126.0, 125.7, 123.8, 51.0, 41.6, 38.7, 37.6, 21.1; HRMS (ESI) m/z calcd for C₂₉H₂₇N₂O₂ [M – H]⁻ 435.2079, found 435.2085.

(S)-N-(4-Chlorophenyl)-N-methyl-2-(2-(naphthalen-1-yl)acetamido)-3-phenylpropanamide (33). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-1-yl)acetic (100)0.54 acid mmol, 1 equiv.) and mg, (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 262 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 80%. ¹H NMR (600 MHz, CD₃OD) δ 7.90 – 7.84 (m, 2H), 7.77 (d, J = 8.2 Hz, 1H), 7.46 – 7.45 (m, 2H), 7.40 – 7.37 (m, 1H), 7.32 (d, J = 6.9 Hz, 1H), 7.25 - 7.15 (m, 5H), 6.86 - 6.85 (m, 4H), 4.59 (t, J = 7.5 Hz, 1H), 3.98 - 6.85 (t, J = 7.5 Hz, 1H), 3.98 - 6.85 (t, J = 7.5 Hz, 1H), 3.98 + 6.85 (t, J = 7.5 Hz, 1H), 3.98 + 6.85 (t, J = 7.53.93 (m, 2H), 3.12 (s, 3H), 2.93 (dd, J = 13.3, 7.4 Hz, 1H), 2.74 (dd, J = 13.3, 7.6 Hz, 1H);NMR (100 MHz, CD₃OD) δ 173.3, 173.2, 142.6, 138.0, 135.4, 135.0, 133.6, 132.8, 130.8, 130.3, 130.2, 129.7, 129.6, 129.1, 128.9, 128.0, 127.3, 126.8, 126.6, 124.9, 53.5, 40.8, 39.2, 38.0; HRMS (ESI) m/z calcd for C₂₈H₂₄ClN₂O₂ [M – H]⁻ 455.1532, found 455.1534.

(*S*)-*N*-(*3*-*Chlorophenyl*)-*N*-*methyl*-2-(*2*-(*naphthalen*-1-*yl*)*acetamido*)-*3*-*phenylpropanamide* (*34*). The synthetic method was similar to that of compound **1** except that 2-(naphthalen-1-yl)acetic acid (100 mg, 0.54 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 262 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 82%. ¹H NMR (600 MHz, CD₃OD) δ 7.91 – 7.90 (m, 1H), 7.85 – 7.83 (m, 1H), 7.76 (d, *J* = 8.2 Hz, 1H), 7.47 – 7.44 (m, 2H), 7.40 – 7.16 (m, 8H), 6.91 – 6.85 (m, 3H), 4.60 – 4.57 (m, 1H), 3.99 – 3.93 (m, 2H), 3.11 (s, 3H), 2.93 (dd, *J* = 13.1, 7.8 Hz, 1H), 2.74 (dd, *J* = 13.1, 7.3 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 173.3, 173.1, 145.1, 137.8, 135.9, 135.3, 133.6, 132.8, 131.9, 130.2, 129.7, 129.6, 129.4, 129.0, 128.9, 128.7, 128.1, 127.3, 127.2, 126.8, 126.6, 124.9, 53.6, 40.8, 39.3, 38.0; HRMS (ESI) *m*/*z* calcd for C₂₈H₂₄ClN₂O₂ [M – H]⁻ 455.1532, found 455.1535.

(S)-N-(3-Fluorophenyl)-N-methyl-2-(2-(naphthalen-1-yl)acetamido)-3-phenylpropanamide (35). The synthetic method was similar to that of compound 1 except that 2-(naphthalen-1-yl)acetic acid (100)mg, 0.54 mmol, 1 equiv.) and (S)-2-amino-N-(3-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 251 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 65%. ¹H NMR (600 MHz, CD₃OD) δ 8.23 (d, J = 6.7 Hz, 1H), 7.90 – 7.76 (m, 3H), 7.46 – 7.26 (m, 6H), 7.20 – 7.14 (m, 3H), 7.05 (t, J = 7.8 Hz, 1H), 6.85 - 6.80 (m, 3H), 6.61 (s, 1H), 4.66 - 4.63 (m, 1H), 3.96 (s, 2H), 3.13(s, 3H), 2.94 (dd, J = 13.2, 7.3 Hz, 1H), 2.74 (dd, J = 13.1, 7.7 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 173.4, 173.1, 164.2 (d, J = 247.5 Hz), 145.4 (d, J = 9.5 Hz), 137.9, 135.3, 133.6, 132.7, 132.1 (d, J = 9.2 Hz), 130.2, 129.7, 129.5, 129.0, 128.8, 128.0, 127.3, 126.8, 126.5, 124.9, 124.7, 116.2 (d, J = 21.1 Hz), 115.9 (d, J = 22.7 Hz), 53.5, 40.8, 39.2, 37.9; HRMS (ESI) m/z calcd for C₂₈H₂₄FN₂O₂ [M – H]⁻ 439.1827, found 439.1831.

(*S*)-*N*-*Ethyl*-2-(2-(*naphthalen*-1-*yl*)*acetamido*)-*N*,3-*diphenylpropanamide* (**36**). The synthetic method was similar to that of compound **1** except that 2-(naphthalen-1-yl)acetic

acid (100 mg, 0.54 mmol, 1 equiv.) and (*S*)-2-amino-*N*-ethyl-*N*,3-diphenylpropanamide (TFA salt, 249 mg, 0.65 mmol, 1.2 equiv.) were used as starting materials. Yield 69%. ¹H NMR (600 MHz, CDCl₃) δ 7.87 – 7.85 (m, 2H), 7.80 (d, *J* = 8.2 Hz, 1H), 7.50 – 7.41 (m, 3H), 7.36 – 7.32 (m, 4H), 7.09 (t, *J* = 7.4 Hz, 1H), 7.02 – 6.99 (m, 2H), 6.83 (s, 1H), 6.60 (d, *J* = 7.4 Hz, 2H), 6.07 (d, *J* = 8.3 Hz, 1H), 4.70 – 4.66 (m, 1H), 3.99 – 3.90 (m, 2H), 3.74 – 3.71 (m, 1H), 3.53 – 3.47 (m, 1H), 2.69 (dd, *J* = 13.3, 6.9 Hz, 1H), 2.50 (dd, *J* = 13.3, 7.0 Hz, 1H), 1.01 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 169.9, 140.7, 136.0, 133.9, 132.1, 131.0, 129.7, 129.2, 128.7, 128.4, 128.3, 128.2, 128.2, 126.6, 126.5, 126.0, 125.7, 123.8, 51.3, 44.6, 41.6, 38.8, 12.7; HRMS (ESI) *m*/*z* calcd for C₂₉H₂₇N₂O₂ [M – H]⁻ 435.2079, found 435.2083.

(*S*)-2-(2-(1,3-*Dimethyl*-2,6-*dioxo*-1,2,3,6-*tetrahydro*-7*H*-*purin*-7-*yl*)*acetamido*)-*N*-*methyl*-*N* ,3-*diphenylpropanamide* (**37**). The synthetic method was similar to that of compound **1** except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7*H*-purin-7-yl)acetic acid (100 mg, 0.42 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-*N*,3-diphenylpropanamide (TFA salt, 184 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 77%. ¹H NMR (600 MHz, CD₃OD) δ 7.85 (s, 1H), 7.31 – 7.29 (m, 4H), 7.19 – 7.15 (m, 3H), 6.91 – 6.87 (m, 3H), 5.06 – 5.00 (m, 2H), 4.66 (t, *J* = 7.4 Hz, 1H), 3.53 (s, 3H), 3.30 (s, 3H), 3.17 (s, 3H), 2.98 (dd, *J* = 13.4, 7.4 Hz, 1H), 2.75 (dd, *J* = 13.4, 7.5 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 172.9, 168.1, 156.5, 153.2, 149.7, 144.6, 143.8, 137.9, 130.8, 130.2, 129.5, 129.3, 128.6, 127.9, 108.4, 53.4, 49.5, 39.3, 38.1, 30.2, 28.2; HRMS (ESI) *m*/*z* calcd for C₂₅H₂₅N₆O₄ [M – H]⁻ 473.1943, found 473.1945.

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(S)-2-(2-(1,3-Dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetamido)-N-methyl-3phenyl-N-(p-tolyl)propanamide (38). The synthetic method was similar to that of compound **1** except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7*H*-purin-7-yl)acetic acid (100)0.42 mmol, 1 equiv.) and mg, (S)-2-amino-N-methyl-3-phenyl-N-(p-tolyl)propanamide (TFA salt, 191 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 72%. ¹H NMR (600 MHz, CD₃OD) δ 7.82 (s, 1H), 7.14 - 7.07 (m, 5H), 6.87 - 6.77 (m, 4H), 5.00 (s, 2H), 4.66 (t, J = 7.4 Hz, 1H), 3.47 (s, 3H), 3.24 (s, 3H), 3.13 (s, 3H), 2.94 (dd, J = 13.4, 7.3 Hz, 1H), 2.72 (dd, J = 13.4, 7.5 Hz, 1H), 2.27 (s, 3H); 13 C NMR (100 MHz, CD₃OD) δ 172.9, 168.0, 156.4, 153.2, 149.7, 144.6, 141.2, 139.5, 137.9, 131.3, 130.2, 129.4, 128.2, 127.9, 108.4, 53.3, 39.4, 38.1, 30.2, 28.2, 21.1; HRMS (ESI) m/z calcd for C₂₆H₂₇N₆O₄ [M – H]⁻ 487.2100, found 487.2104.

(S)-2-(2-(1,3-Dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetamido)-N-(4-fluoro phenyl)-N-methyl-3-phenylpropanamide (39). The synthetic method was similar to that of compound 1 except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetic acid (100)0.42 mg, mmol, 1 equiv.) and (S)-2-amino-N-(4-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 193 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 67%. ¹H NMR (600 MHz, CD₃OD) δ 8.53 (d, J = 7.5 Hz, 1H), 7.85 (s, 1H), 7.21 – 7.20 (m, 3H), 7.01 – 6.93 (m, 5H), 5.06 – 5.00 (m, 2H), 4.61 (q, J = 7.4 Hz, 1H), 3.52 (s, 3H), 3.29 (s, 3H), 3.13 (s, 3H), 2.99 (dd, J = 13.2, 8.1 Hz, 1H), 2.78 (dd, J = 13.3, 7.0 Hz, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 172.9, 168.0, 163.3 (d, *J* = 247.0 Hz), 156.4, 153.2, 149.7, 144.6, 139.9 (d, *J* = 3.2 Hz), 137.8, 130.6, 130.3,

129.6, 128.0, 117.3 (d, J = 23.0 Hz), 108.4, 53.3, 49.4, 39.4, 38.1, 30.2, 28.2; HRMS (ESI) m/z calcd for C₂₅H₂₄FN₆O₄ [M – H]⁻ 491.1849, found 491.1845.

(S)-2-(2-(1,3-Dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetamido)-N-(3-fluoro phenyl)-N-methyl-3-phenylpropanamide (40). The synthetic method was similar to that of compound **1** except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7*H*-purin-7-yl)acetic acid (100)0.42 1 equiv.) and mg, mmol. (S)-2-amino-N-(3-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 193 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 75%. ¹H NMR (600 MHz, CD₃OD) δ 8.57 (d, J = 7.3 Hz, 1H), 7.86 (s, 1H), 7.29 – 7.20 (m, 4H), 7.03 (t, J = 8.3 Hz, 1H), 6.94 – 6.93 (m, 2H), 6.71 – 6.54 (m, 1H), 5.04 (s, 2H), 4.67 – 4.63 (m, 1H), 3.52 (s, 3H), 3.30 (s, 3H), 3.14 (s, 3H), 2.99 (dd, J = 13.1, 8.4 Hz, 1H), 2.79 (dd, J = 13.2, 6.7 Hz, 1H); ¹³C NMR $(150 \text{ MHz}, \text{CD}_3\text{OD}) \delta 172.7, 168.0, 164.1 \text{ (d}, J = 247.4 \text{ Hz}), 156.5, 153.2, 149.7, 145.2 \text{ (d}, J = 247.4 \text{ Hz})$ = 9.6 Hz), 144.6, 137.7, 132.0 (d, J = 9.2 Hz), 130.3, 129.6, 128.0, 124.6, 116.1 (d, J = 21.2) Hz), 115.9 (d, J = 22.6 Hz), 108.4, 53.4, 49.4, 39.6, 37.9, 30.2, 28.2; HRMS (ESI) m/z calcd for $C_{25}H_{24}FN_6O_4 [M - H]^- 491.1849$, found 491.1843.

(S)-N-(4-Chlorophenyl)-2-(2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)ace tamido)-N-methyl-3-phenylpropanamide (41). The synthetic method was similar to that of compound 1 except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetic acid (100 mg, 0.42 mmol, 1 equiv.) and (S)-2-amino-N-(4-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 201 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 90%. ¹H NMR (600 MHz, CDCl₃)

δ 7.67 (d, J = 8.1 Hz, 1H), 7.63 (s, 1H), 7.27 – 7.25 (m, 2H), 7.20 – 7.15 (m, 3H), 6.90 – 6.89 (m, 2H), 6.75 (s, 1H), 4.98 – 4.87 (m, 2H), 4.78 (q, J = 7.7 Hz, 1H), 3.59 (s, 3H), 3.37 (s, 3H), 3.18 (s, 3H), 2.96 (dd, J = 13.4, 7.9 Hz, 1H), 2.80 (dd, J = 13.4, 7.1 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 165.2, 155.5, 151.5, 148.7, 142.3, 140.8, 135.8, 134.1, 129.9, 129.2, 128.7, 128.4, 127.0, 106.8, 51.4, 49.2, 39.0, 37.7, 29.8, 29.7, 28.0; HRMS (ESI) *m/z* calcd for C₂₅H₂₄ClN₆O₄ [M – H]⁻ 507.1553, found 507.1548.

(S)-N-(3-Chlorophenyl)-2-(2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)ace tamido)-N-methyl-3-phenylpropanamide (42). The synthetic method was similar to that of compound 1 except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7*H*-purin-7-yl)acetic acid (100)0.42mmol. 1 equiv.) and mg, (S)-2-amino-N-(3-chlorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 201 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 82%. ¹H NMR (600 MHz, CDCl₃) δ 7.63 (s, 1H), 7.50 (d, J = 8.0 Hz, 1H), 7.28 – 7.17 (m, 5H), 6.90 – 6.89 (m, 3H), 6.58 (s, 1H), 4.96 – 4.84 (m, 2H), 4.79 – 4.75 (m, 1H), 3.59 (s, 3H), 3.38 (s, 3H), 3.17 (s, 3H), 2.95 (dd, J = 13.3, 8.3 Hz, 1H), 2.80 (dd, J = 13.3, 6.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 170.9, 164.9, 155.5, 151.5, 148.8, 143.4, 142.2, 135.6, 135.0, 130.6, 129.1, 128.5, 127.5, 127.1, 125.7, 106.8, 51.4, 49.4, 39.1, 37.6, 29.8, 28.0; HRMS (ESI) m/z calcd for $C_{25}H_{24}CIN_6O_4 [M - H]^- 507.1553$, found 507.1559.

(S)-2-(2-(1,3-Dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetamido)-N-ethyl-N,3 -diphenylpropanamide (43). The synthetic method was similar to that of compound 1 except that 2-(1,3-dimethyl-2,6-dioxo-1,2,3,6-tetrahydro-7H-purin-7-yl)acetic acid (100 mg, 0.42 mmol, 1 equiv.) and (*S*)-2-amino-*N*-ethyl-*N*,3-diphenylpropanamide (TFA salt, 191 mg, 0.50 mmol, 1.2 equiv.) were used as starting materials. Yield 75%. ¹H NMR (600 MHz, CDCl₃) δ 7.57 (s, 1H), 7.37 – 7.33 (m, 4H), 7.15 – 7.09 (m, 3H), 6.90 – 6.81 (m, 3H), 4.95 – 4.76 (m, 2H), 4.74 – 4.70 (m, 1H), 3.86 – 3.80 (m, 1H), 3.62 –3.58 (m, 1H), 3.58 (s, 3H), 3.38 (s, 3H), 2.94 (dd, *J* = 13.7, 6.7 Hz, 1H), 2.73 (dd, *J* = 13.7, 7.8 Hz, 1H), 1.09 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 164.8, 155.5, 151.6, 148.7, 142.0, 140.6, 136.0, 129.7, 129.1, 128.4, 128.4, 128.2, 126.7, 106.7, 51.4, 49.5, 44.7, 38.7, 29.8, 28.0, 12.8; HRMS (ESI) *m*/*z* calcd for C₂₆H₂₇N₆O₄ [M – H]⁻ 487.2100, found 487.2105.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-methyl-N,3-diphenylpropana mide (44). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-N-methyl-N,3-diphenylpropanamide (TFA salt, 195 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 65%. ¹H NMR (600 MHz, DMSO- d_6) δ 8.65 (s, 1H), 8.53 (d, *J* = 7.8 Hz, 1H), 7.45 – 7.32 (m, 3H), 7.20 – 7.12 (m, 4H), 6.82 – 6.77 (m, 2H), 4.42 – 4.38 (m, 1H), 3.95 – 3.82 (m, 2H), 3.13 (s, 3H), 2.81 (dd, *J* = 13.2, 6.2 Hz, 1H), 2.61 (dd, *J* = 13.5, 9.2 Hz, 1H), 1.64 – 1.59 (m, 4H), 1.57 – 1.45 (m, 5H), 1.29 – 1.24 (m, 1H); ¹³C NMR (150 MHz, DMSO- d_6) δ 176.9, 171.0, 166.1, 155.8, 143.1, 137.7, 130.0, 129.2, 129.2, 128.6, 128.3, 127.9, 126.9, 61.5, 52.1, 37.8, 37.6, 33.7, 24.8, 21.2; HRMS (ESI) *m*/*z* calcd for C₂₆H₃₁N₄O₄ [M + H]⁺ 463.2340, found 463.2337.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N,3-diphenylpropanamide.

(45). The synthetic method was similar to that of compound 1 except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and

(*S*)-2-amino-*N*,3-diphenylpropanamide (TFA salt, 188 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 70%. ¹H NMR (600 MHz, DMSO- d_6) δ 10.04 (s, 1H), 8.67 (s, 1H), 8.55 (d, *J* = 7.9 Hz, 1H), 7.54 (d, *J* = 7.8 Hz, 2H), 7.30 – 7.24 (m, 4H), 7.20 – 7.15 (m, 1H), 7.04 (t, *J* = 7.4 Hz, 1H), 4.64 – 4.59 (m, 1H), 4.00 – 3.89 (m, 2H), 3.02 (dd, *J* = 13.7, 5.8 Hz, 1H), 2.88 (dd, *J* = 13.7, 8.5 Hz, 1H), 1.65 – 1.55 (m, 4H), 1.54 – 1.46 (m, 5H), 1.29 – 1.25 (m, 1H); ¹³C NMR (150 MHz, DMSO- d_6) δ 177.0, 170.1, 166.4, 155.9, 139.1, 137.7, 129.6, 129.1, 128.6, 126.9, 123.9, 119.9, 61.5, 55.4, 38.3, 33.7, 33.7, 24.8, 21.2; HRMS (ESI) *m*/*z* calcd for C₂₅H₂₉N₄O₄ [M + H]⁺ 449.2183, found 449.2186.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-(4-methoxyphenyl)-N-methyl-3-phenylpropanamide (46). The synthetic method was similar to that of compound 1 except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(4-methoxyphenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 211 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 62%. ¹H NMR (600 MHz, DMSO- d_6) δ 8.65 (s, 1H), 8.50 (d, *J* = 7.9 Hz, 1H), 7.20 – 7.14 (m, 3H), 7.07 – 6.99 (m, 2H), 6.93 (d, *J* = 8.3 Hz, 2H), 6.84 (d, *J* = 7.0 Hz, 2H), 4.41 – 4.35 (m, 1H), 3.95 – 3.79 (m, 2H), 3.76 (s, 3H), 3.08 (s, 3H), 2.83 (dd, *J* = 13.5, 5.3 Hz, 1H), 2.61 (dd, *J* = 13.4, 8.8 Hz, 1H), 1.64 – 1.58 (m, 4H), 1.54 – 1.42 (m, 5H), 1.30 – 1.23 (m, 1H); ¹³C NMR (150 MHz, DMSO- d_6) δ 176.9, 171.2, 166.0, 158.9, 155.8, 137.7, 135.8, 129.5, 129.1, 128.8, 128.4, 127.1, 115.3, 114.9, 61.5, 55.9, 55.7, 52.0, 51.8, 37.9, 33.7, 24.9, 21.2; HRMS (ESI) *m*/*z* calcd for C₂₇H₃₃N₄O₅ [M + H]⁺ 493.2445, found 493.2449.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-methyl-3-phenyl-N-(p-tolyl)p ropanamide (47). The synthetic method was similar to that of compound 1 except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-*N*-methyl-3-phenyl-*N*-(*p*-tolyl)propanamide (TFA salt, 203 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 61%. ¹H NMR (600 MHz, DMSO-*d*₆) δ 8.65 (s, 1H), 8.50 (d, *J* = 7.9 Hz, 1H), 7.20 – 7.15 (m, 4H), 6.98 (d, *J* = 7.1 Hz, 2H), 6.85 – 6.80 (m, 2H), 4.44 – 4.39 (m, 1H), 3.96 – 3.82 (m, 2H), 3.09 (s, 3H), 2.82 (dd, *J* = 12.5, 7.4 Hz, 1H), 2.61 (dd, *J* = 13.2, 8.9 Hz, 1H), 2.31 (s, 3H), 1.66 – 1.59 (m, 4H), 1.54 – 1.44 (m, 5H), 1.31 – 1.24 (m, 1H); ¹³C NMR (150 MHz, DMSO-*d*₆) δ 176.9, 171.0, 166.0, 155.8, 140.6, 137.7, 137.7, 130.5, 129.7, 129.3, 128.6, 127.6, 126.9, 61.5, 51.9, 38.0, 37.6, 33.7, 24.8, 21.2, 21.1; HRMS (ESI) *m*/*z* calcd for C₂₇H₃₃N₄O₄ [M + H]⁺ 477.2496, found 477.2494.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-(4-fluorophenyl)-N-methyl-3phenylpropanamide (48). The synthetic method was similar to that of compound 1 except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-N-(4-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 205 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 69%. ¹H NMR (600 MHz, CD₃OD) δ 7.32 – 7.23 (m, 5H), 7.03 – 7.00 (m, 2H), 6.98 – 6.96 (m, 2H), 4.60 – 4.58 (m, 1H), 4.16 – 4.04 (m, 2H), 3.13 (s, 3H), 2.97 (dd, *J* = 13.1, 8.4 Hz, 1H), 2.75 (dd, *J* = 13.1, 6.6 Hz, 1H), 1.83 – 1.77 (m, 4H), 1.66 – 1.63 (m, 3H), 1.58 – 1.53 (m, 2H), 1.43 – 1.37 (m, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 172.9, 168.0, 163.3 (d, *J* = 246.8 Hz), 157.7, 139.9, 139.9, 137.9, 130.6 (d, *J* = 7.6 Hz), 130.4, 129.6, 128.0, 117.4 (d, *J* = 23.0 Hz), 63.3, 53.2, 40.9, 39.6, 38.1, 34.6, 25.8, 22.5; HRMS (ESI) *m*/z calcd for C₂₆H₂₈FN₄O₄ [M – H]⁻ 479.2100, found 479.2106. 2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-(3-fluorophenyl)-N-methyl-3phenylpropanamide (**49**). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-N-(3-fluorophenyl)-N-methyl-3-phenylpropanamide (TFA salt, 205 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 70%. ¹H NMR (600 MHz, CD₃OD) δ 7.33 – 7.23 (m, 5H), 7.06 (t, *J* = 8.3 Hz, 1H), 6.97 – 6.96 (m, 2H), 6.73 (s, 1H), 6.47 (s, 1H), 4.64 – 4.61 (m, 1H), 4.17 – 4.05 (m, 2H), 3.14 (s, 3H), 2.97 (dd, *J* = 13.0, 8.6 Hz, 1H), 2.77 (dd, *J* = 13.1, 6.4 Hz, 1H), 1.84 – 1.79 (m, 4H), 1.67 – 1.64 (m, 3H), 1.59 – 1.52 (m, 2H), 1.43 – 1.37 (m, 1H); ¹³C NMR (150 MHz, CD₃OD) δ 178.9, 172.7, 168.1, 164.2 (d, *J* = 247.4 Hz), 157.8, 145.3 (d, *J* = 9.6 Hz), 137.8, 132.1 (d, *J* = 9.1 Hz), 130.4, 129.6, 128.1, 124.6, 116.1 (d, *J* = 21.1 Hz), 115.8 (d, *J* = 22.9 Hz), 63.3, 53.4, 40.9, 39.8, 37.9, 34.6, 25.8, 22.5; HRMS (ESI) *m*/*z* calcd for C₂₆H₂₈FN₄O₄ [M – H]⁻ 479.2100, found 479.2100.

N-(*4*-*Chlorophenyl*)-2-(2-(2,4-*dioxo*-1,3-*diazaspiro*[4.5]*decan*-3-*yl*)*acetamido*)-*N*-*methyl*-3 -*phenylpropanamide* (*50*). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-*y*])acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(4-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 213 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 56%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.67 (s, 1H), 8.60 (d, *J* = 7.7 Hz, 1H), 7.45 (d, *J* = 8.5 Hz, 2H), 7.22 – 7.18 (m, 3H), 7.10 (d, *J* = 8.1 Hz, 2H), 6.91 – 6.87 (m, 2H), 4.40 – 4.32 (m, 1H), 3.98 – 3.82 (m, 2H), 2.90 – 2.83 (m, 1H), 2.66 (dd, *J* = 13.2, 8.3 Hz, 1H), 1.70 – 1.60 (m, 4H), 1.56 – 1.41 (m, 5H), 1.31 – 1.28 (m, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 177.0, 170.9, 166.2, 155.8, 142.0, 137.5, 132.8, 130.0, 129.8, 129.3, 128.7, 127.1, 61.6, 52.0, 38.0, 37.5, 33.7, 24.8, 21.3; HRMS (ESI) *m*/*z* calcd for C₂₆H₃₀ClN₄O₄ [M + H]⁺ 497.1950, found 497.1952.

N-(*3*-*Chlorophenyl*)-2-(2-(2,4-*dioxo*-1,3-*diazaspiro*[4.5]*decan*-3-*yl*)*acetamido*)-*N*-*methyl*-3 -*phenylpropanamide* (*51*). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-chlorophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 213 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 72%. ¹H NMR (600 MHz, CD₃OD) δ 7.32 – 7.25 (m, 7H), 6.98 – 6.96 (m, 2H), 4.60 – 4.58 (m, 1H), 4.17 – 4.05 (m, 2H), 3.12 (s, 3H), 2.97 (dd, *J* = 13.0, 9.0 Hz, 1H), 2.77 (dd, *J* = 13.0, 6.2 Hz, 1H), 1.84 – 1.78 (m, 4H), 1.68 – 1.64 (m, 3H), 1.59 – 1.53 (m, 2H), 1.44 – 1.37 (m, 1H); HRMS-ESI (–) *m/z* calcd for C₂₆H₂₈ClN₄O₄ [M – H]⁻ 495.1805, found 495.1810.

N-(*3*-*Bromophenyl*)-2-(2-(2,4-*dioxo*-1,3-*diazaspiro*[4.5]*decan*-3-*yl*)*acetamido*)-*N*-*methyl*-3*phenylpropanamide* (**52**). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-*N*-(3-bromophenyl)-*N*-methyl-3-phenylpropanamide (TFA salt, 237 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 68%. ¹H NMR (600 MHz, DMSO-*d*₆) δ 8.65 (s, 1H), 8.60 (d, *J* = 7.3 Hz, 1H), 7.54 (d, *J* = 7.9 Hz, 1H), 7.33 (t, *J* = 8.0 Hz, 1H), 7.20 – 7.13 (m, 5H), 6.88 – 6.87 (m, 2H), 4.34 – 4.31 (m, 1H), 3.95 – 3.84 (m, 2H), 3.08 (s, 3H), 2.86 (dd, *J* = 13.4, 5.9 Hz, 1H), 2.66 (dd, *J* = 14.4, 7.2 Hz, 1H), 1.63 – 1.48 (m, 9H), 1.29 – 1.23 (m, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 177.0, 171.0, 166.2, 155.8, 144.6, 137.5, 131.7, 131.3, 130.7, 129.3, 128.8, 127.2, 127.1, 122.3, 61.6, 52.2, 38.1, 37.5, 33.7, 24.9, 21.3; HRMS (ESI) m/z calcd for C₂₆H₂₈BrN₄O₄ [M – H]⁻ 539.1299, found 539.1297.

2-(2-(2,4-Dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetamido)-N-ethyl-N,3-diphenylpropanami *de* (**53**). The synthetic method was similar to that of compound **1** except that 2-(2,4-dioxo-1,3-diazaspiro[4.5]decan-3-yl)acetic acid (100 mg, 0.44 mmol, 1 equiv.) and (*S*)-2-amino-N-ethyl-N,3-diphenylpropanamide (TFA salt, 203 mg, 0.53 mmol, 1.2 equiv.) were used as starting materials. Yield 58%. ¹H NMR (400 MHz, DMSO- d_6) δ 8.65 (s, 1H), 8.49 (d, *J* = 7.9 Hz, 1H), 7.46 – 7.39 (m, 3H), 7.20 – 7.16 (m, 3H), 7.08 – 7.04 (m, 2H), 6.86 – 6.81 (m, 2H), 4.35 – 4.28 (m, 1H), 3.98 – 3.83 (m, 2H), 3.69 – 3.55 (m, 2H), 2.86 (dd, *J* = 13.4, 5.5 Hz, 1H), 2.62 (dd, *J* = 13.4, 8.5 Hz, 1H), 1.65 – 1.53 (m, 4H), 1.56 – 1.45 (m, 5H), 1.34 – 1.25 (m, 1H), 0.97 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 170.0, 170.4, 166.0, 155.8, 141.3, 137.7, 130.0, 129.4, 128.9, 128.6, 128.5, 127.0, 61.5, 52.3, 44.2, 38.0, 33.7, 24.9, 21.3, 13.1; HRMS (ESI) *m*/*z* calcd for C₂₇H₃₃N₄O₄ [M + H]⁺ 477.2496, found 477.2500.

2-(2-(2,5-Dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetamido)-N -methyl-N,3-diphenylpropanamide (54). The synthetic method was similar to that of compound 1 except that 2-(2,5-dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetic acid (100 mg, 0.36 mmol, 1 equiv.) and (S)-2-amino-N-methyl-N,3-diphenylpropanamide (TFA salt, 158 mg, 0.43 mmol, 1.2 equiv.) were used as starting materials. Yield 52%. ¹H NMR (600 MHz, DMSO- d_6) δ 8.83 (s, 1H), 8.63 (d, J = 8.0 Hz, 1H), 7.46 – 7.32 (m, 3H), 7.31 – 7.04 (m, 9H), 6.84 (d, J = 6.4 Hz, 2H), 4.46 – 4.41 (m, 1H), 4.09 – 3.94 (m, 2H), 3.12 (s, 3H), 2.86 (dd, J = 13.3, 5.3 Hz, 1H), 2.75 – 2.60 (m, 2H), 2.63 – 2.60 (m, 1H), 2.09 – 1.82 (m, 4H); HRMS (ESI) m/z calcd for C₃₀H₃₁N₄O₄ [M + H]⁺ 511.2340, found 511.2340.

2-(2-(2,5-Dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetamido)-N,

3-diphenylpropanamide (55). The synthetic method was similar to that of compound **1** except that 2-(2,5-dioxo-3',4'-dihydro-2'*H*-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetic acid (100 mg, 0.36 mmol, 1 equiv.) and (*S*)-2-amino-*N*,3-diphenylpropanamide (TFA salt, 152 mg, 0.43 mmol, 1.2 equiv.) were used as starting materials. Yield 72%. ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.09 (s, 1H), 8.87 (s, 1H), 8.68 (s, 1H), 7.54 (s, 2H), 7.26 – 6.94 (m, 10H), 4.72 (s, 1H), 4.15 – 4.05 (m, 2H), 3.05 (s, 2H), 2.91 (s, 1H), 2.76 (s, 2H), 2.08 – 1.81 (m, 4H); ¹³C NMR (150 MHz, DMSO-*d*₆) δ 176.6, 170.0, 166.4, 155.7, 139.0, 138.1, 137.7, 134.6, 129.6, 129.6, 129.5, 129.1, 128.6, 128.5, 128.0, 119.9, 126.9, 123.9, 62.8, 55.4, 38.5, 34.1, 28.9, 18.9; HRMS (ESI) *m*/*z* calcd for C₂₉H₂₉N₄O₄ [M + H]⁺ 497.2183, found 497.2182.

2-(2-(2,5-Dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetamido)-N -(4-methoxyphenyl)-N-methyl-3-phenylpropanamide (56). The synthetic method was compound similar that of 1 except that to 2-(2,5-dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetic acid (100 0.36 mmol, 1 equiv.) mg, and (S)-2-amino-N-(4-methoxyphenyl)-N-methyl-3-phenylpropanamide (TFA salt, 171 mg, 0.43 mmol, 1.2 equiv.) were used as starting materials. Yield 67%. ¹H NMR (600 MHz, DMSO- d_6) δ 8.83 (s, 1H), 8.61 – 8.57 (m, 1H), 7.25 – 7.11 (m, 7H), 7.06 – 6.96 (m, 2H), 6.95 - 6.92 (m, 2H), 6.91 - 6.86 (m, 2H), 4.44 - 4.34 (m, 1H), 4.10 - 3.98 (m, 2H), 3.77 (s, 3H),

3.08 (s, 3H), 2.87 (dd, J = 13.1, 5.5 Hz, 1H), 2.77 – 2.75 (m, 2H), 2.66 – 2.61 (m, 1H), 2.15 – 2.02 (m, 2H), 1.92 – 1.80 (m, 2H); HRMS (ESI) m/z calcd for C₃₁H₃₃N₄O₅ [M + H]⁺ 541.2445, found 541.2446.

2-(2-(2,5-Dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetamido)-N -methyl-3-phenyl-N-(p-tolyl)propanamide (57). The synthetic method was similar to that of 1 compound except that 2-(2,5-dioxo-3',4'-dihydro-2'H-spiro[imidazolidine-4,1'-naphthalen]-1-yl)acetic acid (100 mg, 0.36 mmol, 1 equiv.) and (S)-2-amino-N-methyl-3-phenyl-N-(p-tolyl)propanamide (TFA salt, 164 mg, 0.43 mmol, 1.2 equiv.) were used as starting materials. Yield 55%. ¹H NMR (600 MHz, DMSO- d_6) δ 8.83 (s, 1H), 8.60 (d, J = 7.9 Hz, 1H), 7.29 (d, J = 7.8 Hz, 1H), 7.29 – 7.12 (m, 8H), 6.95 (d, *J* = 7.1 Hz, 2H), 6.87 (d, *J* = 7.4 Hz, 2H), 4.46 – 4.40 (m, 1H), 4.13 – 3.92 (m, 2H), 3.08 (s, 3H), 2.87 (dd, J = 13.5, 5.8 Hz, 1H), 2.77 – 2.72 (m, 2H), 2.64 $(dd, J = 13.3, 8.4 Hz, 1H), 2.32 (s, 3H), 2.08 - 1.80 (m, 4H); {}^{13}C NMR (150 MHz, DMSO-d_6)$ δ 176.6, 171.0, 166.0, 155.7, 138.1, 137.7, 137.6, 134.6, 130.4, 129.5, 129.4, 128.6, 128.5, 127.6, 126.9, 62.7, 51.8, 38.1, 37.6, 34.0, 28.9, 21.1, 18.9; HRMS (ESI) m/z calcd for $C_{31}H_{33}N_4O_4 [M + H]^+ 525.2496$, found 525.2498.

4.2 Thermal Shift Assays (TSAs) to test compounds for HIV-1 CA hexamer stability

Compounds were screened for their effect on CA stability using purified covalently-crosslinked hexameric CA^{A14C/E45C/W184A/M185A} (CA121). CA121 cloned in a pET11a expression plasmid was kindly provided by Dr. Owen Pornillos (University of Virginia, Charlottesville, VA). Protein was expressed in *E. coli* BL21(DE3)RIL and purified as reported previously [34]. The TSA has been previously described [56-58]. Briefly, the TSA

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was conducted on the PikoReal Real-Time PCR System (Thermo Fisher Scientific) or the QuantStudio 3 Real-Time PCR system (Thermo Fisher Scientific). Each reaction contained 7.5 μ M CA121, 1x Sypro Orange Protein Gel Stain (Life Technologies), 50 mM sodium phosphate buffer (pH 8.0) and 1% DMSO (control) or 20 μ M compound. The plate was heated from 25 to 95 °C with a heating rate of 0.2 °C every 10 sec. The fluorescence intensity was measured with an Ex range of 475–500 nm and Em range of 520–590 nm. The differences in the melting temperature (ΔT_m) of CA hexamer in DMSO (T₀) verses in the presence of compound (T_m) were calculated using the following formula: $\Delta T_m = T_m - T_0$.

4.3 Virus Production

The wild-type laboratory HIV-1 strain, HIV-1_{NL4-3} [59], was produced using a pNL4-3 vector that was obtained through the NIH AIDS Reagent Program, Division of AIDS, NIAID, NIH. HIV-1_{NL4-3} was generated by transfecting HEK 293FT cells in a T75 flask with 10 μ g of the pNL4-3 vector and FuGENE®HD Transfection Reagent (Promega). Supernatant was harvested 48-72 hours post-transfection and transferred to MT2 cells for viral propagation. Virus was harvested when syncytia formation was observed, which took 3-5 days. The viral supernatant was then concentrated using 8% w/v PEG 8,000 overnight at 4 °C, followed by centrifugation for 40 min at 3,500 rpm. The resulting viral-containing pellet was concentrated 10-fold by resuspension in DMEM without FBS and stored at -80 °C.

4.4 Anti-HIV-1 and Cytotoxicity assays

Anti-HIV-1 activity of **PF74** and **PF74**-related analogs were examined in TZM-GFP cells. The potency of HIV-1 inhibition by a compound was based on its inhibitory effect on viral LTR-activated GFP expression compared with that of compound-free (DMSO) controls. Briefly, TZM-GFP cells were plated at density of 1×10^4 cells per well in a 96-well plate. 24 h later, media was replaced with increasing concentrations of compound. 24 h post treatment,

cells were exposed to an HIV-1 strain (MOI = 1). After incubation for 48 h, anti-HIV-1 activity was assessed by counting the number of GFP positive cells on a CytationTM 5 Imaging Reader (BioTek) and 50% effective concentration (EC₅₀) values were determined.

Cytotoxicity of each compound was also determined in TZM-GFP cells. Cells were plated at a density of 1×10^4 cells per well in a 96-well plate and were continuously exposed to increasing concentrations of a compound for 72 hours. The number of viable cells in each well was determined using a Cell Proliferation Kit II (XTT), and 50% cytotoxicity concentration (CC₅₀) values were determined. All the cell-based assays were conducted in duplicate and in at least two independent experiments.

For the EC₅₀ and CC₅₀ dose responses, values were plotted in GraphPad Prism 5 and analyzed with the *log (inhibitor) vs. normalized response – variable slope* equation. Final values were calculated in each independent assay and the average values were determined. Statistical analysis (calculation of standard deviation) was performed by using Microsoft Excel.

4.5 Microsomal stability assay.

The *in vitro* microsomal stability assay was conducted in duplicate in mouse and human liver microsomal systems, which were supplemented with nicotinamide adenine dinucleotide phosphate (NADPH) as a cofactor. Briefly, a compound (1 μ M final concentration) was pre-incubated, in the absence or presence of 0.5 μ M Cobicistat (CYP 3A inhibitor, purchased from medchemexpress.com and verified with LCMS), with the reaction mixture containing liver microsomal protein (0.5 mg/mL final concentration) and MgCl₂ (1 mM final concentration) in 0.1 M potassium phosphate buffer (pH 7.4) at 37 °C for 15 minutes. The reaction was initiated by addition of 1 mM NADPH, followed by incubation at 37 °C. A negative control was performed in parallel in the absence of NADPH to measure any chemical instability or non-NADPH dependent enzymatic degradation for each compound. At various time points (0, 5, 15, 30, and 60 min), 1 volume

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of reaction aliquot was taken and quenched with 3 volumes of acetonitrile containing an appropriate internal standard and 0.1% formic acid. The samples were then vortexed and centrifuged at 15,000 rpm for 5 min at 4 °C. The supernatants were collected and analyzed by LC-MS/MS to determine the *in vitro* metabolic half-life ($t_{1/2}$).

4.6 Molecular modeling

Molecular modeling was performed using the Schrödinger small molecule drug discovery suite 2019-1 [60]. The crystal structure of native full length HIV-1 capsid protein in complex with **PF74** was retrieved from the protein data bank (PDB code: 4XFZ) [30]. The above structure was analyzed using Maestro [61] (Schrödinger Inc.) and subjected to a docking protocol that involves several steps including preparing the protein of interest, grid generation, ligand preparation, and docking. The crystal structure was refined using the protein preparation wizard [62] (Schrödinger Inc.) in which missing hydrogen atoms, side chains, and loops were added using prime and minimized using the OPLS 3e force field [63] to optimize the hydrogen bonding network and converge the heavy atoms to an rmsd of 0.3Å. The receptor grid generation tool in Maestro (Schrödinger Inc.) was used to define an active site around the native ligand PF74 to cover all the residues within 12 Å. All the compounds were drawn using Maestro and subjected to LigPrep [64] to generate conformers, possible protonation at pH of 7 ± 2 that serves as an input for docking process. All the dockings were performed using Glide XP [65](Glide, version 8.2) with the van der Waals radii of nonpolar atoms for each of the ligands scaled by a factor of 0.8. The solutions were further refined by post docking and minimization under implicit solvent to account for protein flexibility. The residue numbers of HIV-1 capsid protein used in the discussion and the figures were based on the native full length HIV-1 capsid protein.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at

Abbreviations

HIV, human immunodeficiency virus

CA, capsid protein

CPSF6, cleavage and polyadenylation specific factor 6

CypA, Cyclophilin A

CA_{NTD}, CA N-terminal domain

CA_{CTD}, CA C-terminal domain

SAR, structure-activity-relationship

HATU, 1-[Bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxid

hexafluorophosphate

T₃P, 2,4,6-Tripropyl-1,3,5,2,4,6-trioxatriphosphorinane-2,4,6-trioxide

DIPEA, N,N-Diisopropylethylamine

TFA, Trifluoroacetic acid

TSA, thermal shift assay

HLM, human liver microsome

MLM, mouse liver microsome

 EC_{50} , 50% effective concentration

CC₅₀, 50% cytotoxicity concentration

References

- 1 AIDSinfo, 'FDA-Approved HIV Medicines' <https://aidsinfo.nih.gov/understanding-hiv-aids/fact-sheets/21/58/fda-approved-hiv -medicines> [Accessed March 2020.
- 2 E. M. Campbell, and T. J. Hope, HIV-1 capsid: the multifaceted key player in HIV-1 infection, Nat Rev Microbiol, 13 (2015), 471-83.
- 3 V. Le Sage, A. J. Mouland, and F. Valiente-Echeverria, Roles of HIV-1 capsid in viral replication and immune evasion, Virus Res, 193 (2014), 116-29.
- 4 A. Fassati, Multiple roles of the capsid protein in the early steps of HIV-1 infection, Virus Res, 170 (2012), 15-24.
- 5 N. M. Bell, and A. M. Lever, HIV Gag polyprotein: processing and early viral particle assembly, Trends Microbiol, 21 (2013), 136-44.
- 6 S. K. Carnes, J. H. Sheehan, and C. Aiken, Inhibitors of the HIV-1 capsid, a target of opportunity, Curr Opin HIV AIDS, 13 (2018), 359-65.

- 7 S. Thenin-Houssier, and S. T. Valente, HIV-1 Capsid Inhibitors as Antiretroviral Agents, Curr Hiv Res, 14 (2016), 270-82.
- J. Y. Zhang, X. Y. Liu, and E. De Clercq, Capsid (CA) Protein as a Novel Drug Target: Recent Progress in the Research of HIV-1 CA Inhibitors, Mini-Rev Med Chem, 9 (2009), 510-18.
- 9 B. K. Ganser, S. Li, V. Y. Klishko, J. T. Finch, and W. I. Sundquist, Assembly and analysis of conical models for the HIV-1 core, Science, 283 (1999), 80-3.
- 10 S. Li, C. P. Hill, W. I. Sundquist, and J. T. Finch, Image reconstructions of helical assemblies of the HIV-1 CA protein, Nature, 407 (2000), 409-13.
- S. Rankovic, R. Ramalho, C. Aiken, and I. Rousso, PF74 Reinforces the HIV-1
 Capsid To Impair Reverse Transcription-Induced Uncoating, J Virol, 92 (2018).
- 12 Z. Ambrose, and C. Aiken, HIV-1 uncoating: connection to nuclear entry and regulation by host proteins, Virology, 454-455 (2014), 371-9.
- M. Yamashita, and A. N. Engelman, Capsid-Dependent Host Factors in HIV-1 Infection, Trends Microbiol, 25 (2017), 741-55.
- 14 D. M. Sayah, E. Sokolskaja, L. Berthoux, and J. Luban, Cyclophilin A retrotransposition into TRIM5 explains owl monkey resistance to HIV-1, Nature, 430 (2004), 569-73.
- M. Stremlau, C. M. Owens, M. J. Perron, M. Kiessling, P. Autissier, and J. Sodroski,
 The cytoplasmic body component TRIM5alpha restricts HIV-1 infection in Old
 World monkeys, Nature, 427 (2004), 848-53.

- V. Achuthan, J. M. Perreira, G. A. Sowd, M. Puray-Chavez, W. M. McDougall, A.
 Paulucci-Holthauzen, X. Wu, H. J. Fadel, E. M. Poeschla, A. S. Multani, S. H.
 Hughes, S. G. Sarafianos, A. L. Brass, and A. N. Engelman, Capsid-CPSF6
 Interaction Licenses Nuclear HIV-1 Trafficking to Sites of Viral DNA Integration,
 Cell Host Microbe, 24 (2018), 392-404 e8.
- 17 D. A. Bejarano, K. Peng, V. Laketa, K. Borner, K. L. Jost, B. Lucic, B. Glass, M. Lusic, B. Mueller, and H. G. Krausslich, HIV-1 nuclear import in macrophages is regulated by CPSF6-capsid interactions at the nuclear pore complex, Elife, 8 (2019).
- 18 C. L. Woodward, S. Prakobwanakit, S. Mosessian, and S. A. Chow, Integrase interacts with nucleoporin NUP153 to mediate the nuclear import of human immunodeficiency virus type 1, J Virol, 83 (2009), 6522-33.
- 19 K. A. Matreyek, S. S. Yucel, X. Li, and A. Engelman, Nucleoporin NUP153 phenylalanine-glycine motifs engage a common binding pocket within the HIV-1 capsid protein to mediate lentiviral infectivity, PLoS Pathog, 9 (2013), e1003693.
- C. Buffone, A. Martinez-Lopez, T. Fricke, S. Opp, M. Severgnini, I. Cifola, L. Petiti,
 S. Frabetti, K. Skorupka, K. K. Zadrozny, B. K. Ganser-Pornillos, O. Pornillos, F. Di
 Nunzio, and F. Diaz-Griffero, Nup153 Unlocks the Nuclear Pore Complex for HIV-1
 Nuclear Translocation in Nondividing Cells, J Virol, 92 (2018).
- A. Dharan, S. Talley, A. Tripathi, J. I. Mamede, M. Majetschak, T. J. Hope, and E. M.
 Campbell, KIF5B and Nup358 Cooperatively Mediate the Nuclear Import of HIV-1
 during Infection, PLoS Pathog, 12 (2016), e1005700.

- A. M. Meehan, D. T. Saenz, R. Guevera, J. H. Morrison, M. Peretz, H. J. Fadel, M. Hamada, J. van Deursen, and E. M. Poeschla, A cyclophilin homology domain-independent role for Nup358 in HIV-1 infection, PLoS Pathog, 10 (2014), e1003969.
- T. Fricke, T. E. White, B. Schulte, D. A. de Souza Aranha Vieira, A. Dharan, E. M.
 Campbell, A. Brandariz-Nunez, and F. Diaz-Griffero, MxB binds to the HIV-1 core and prevents the uncoating process of HIV-1, Retrovirology, 11 (2014), 68.
- B. Xu, Q. Pan, and C. Liang, Role of MxB in Alpha Interferon-Mediated Inhibition of
 HIV-1 Infection, J Virol, 92 (2018).
- E. K. Franke, H. E. Yuan, and J. Luban, Specific incorporation of cyclophilin A into HIV-1 virions, Nature, 372 (1994), 359-62.
- M. Thali, A. Bukovsky, E. Kondo, B. Rosenwirth, C. T. Walsh, J. Sodroski, and H. G.
 Gottlinger, Functional association of cyclophilin A with HIV-1 virions, Nature, 372 (1994), 363-5.
- K. Kim, A. Dauphin, S. Komurlu, S. M. McCauley, L. Yurkovetskiy, C. Carbone, W.
 E. Diehl, C. Strambio-De-Castillia, E. M. Campbell, and J. Luban, Cyclophilin A protects HIV-1 from restriction by human TRIM5alpha, Nat Microbiol, 4 (2019), 2044-51.
- 28 M. Novikova, Y. Zhang, E. O. Freed, and K. Peng, Multiple Roles of HIV-1 Capsid during the Virus Replication Cycle, Virol Sin, 34 (2019), 119-34.
- E. O. Freed, HIV-1 assembly, release and maturation, Nat Rev Microbiol, 13 (2015),
 484-96.

- 30 A. T. Gres, K. A. Kirby, V. N. KewalRamani, J. J. Tanner, O. Pornillos, and S. G. Sarafianos, STRUCTURAL VIROLOGY. X-ray crystal structures of native HIV-1 capsid protein reveal conformational variability, Science, 349 (2015), 99-103.
- C. T. Lemke, S. Titolo, U. von Schwedler, N. Goudreau, J. F. Mercier, E. Wardrop,
 A. M. Faucher, R. Coulombe, S. S. Banik, L. Fader, A. Gagnon, S. H. Kawai, J.
 Rancourt, M. Tremblay, C. Yoakim, B. Simoneau, J. Archambault, W. I. Sundquist,
 and S. W. Mason, Distinct effects of two HIV-1 capsid assembly inhibitor families
 that bind the same site within the N-terminal domain of the viral CA protein, J Virol,
 86 (2012), 6643-55.
- 32 F. Ternois, J. Sticht, S. Duquerroy, H. G. Krausslich, and F. A. Rey, The HIV-1 capsid protein C-terminal domain in complex with a virus assembly inhibitor, Nat Struct Mol Biol, 12 (2005), 678-82.
- W. S. Blair, C. Pickford, S. L. Irving, D. G. Brown, M. Anderson, R. Bazin, J. A. Cao,
 G. Ciaramella, J. Isaacson, L. Jackson, R. Hunt, A. Kjerrstrom, J. A. Nieman, A. K.
 Patick, M. Perros, A. D. Scott, K. Whitby, H. Wu, and S. L. Butler, HIV Capsid is a
 Tractable Target for Small Molecule Therapeutic Intervention, Plos Pathogens, 6 (2010).
- O. Pornillos, B. K. Ganser-Pornillos, B. N. Kelly, Y. Hua, F. G. Whitby, C. D. Stout,
 W. I. Sundquist, C. P. Hill, and M. Yeager, X-ray structures of the hexameric building block of the HIV capsid, Cell, 137 (2009), 1282-92.
- 35 G. Zhao, J. R. Perilla, E. L. Yufenyuy, X. Meng, B. Chen, J. Ning, J. Ahn, A. M. Gronenborn, K. Schulten, C. Aiken, and P. Zhang, Mature HIV-1 capsid structure by

cryo-electron microscopy and all-atom molecular dynamics, Nature, 497 (2013), 643-6.

- A. Machara, V. Lux, M. Kozisek, K. Grantz Saskova, O. Stepanek, M. Kotora, K. 36 Parkan, M. Pavova, B. Glass, P. Sehr, J. Lewis, B. Muller, H. G. Krausslich, and J. Konvalinka, Specific Inhibitors of HIV Capsid Assembly Binding to the C-Terminal Domain of the Capsid Protein: Evaluation of 2-Arylquinazolines as Potential Antiviral Compounds, J Med Chem, 59 (2016), 545-58.
- S. Thenin-Houssier, I. M. de Vera, L. Pedro-Rosa, A. Brady, A. Richard, B. Konnick, 37 S. Opp, C. Buffone, J. Fuhrmann, S. Kota, B. Billack, M. Pietka-Ottlik, T. Tellinghuisen, H. Choe, T. Spicer, L. Scampavia, F. Diaz-Griffero, D. J. Kojetin, and S. T. Valente, Ebselen, a Small-Molecule Capsid Inhibitor of HIV-1 Replication, Antimicrob Agents Chemother, 60 (2016), 2195-208.
- B. N. Kelly, S. Kyere, I. Kinde, C. Tang, B. R. Howard, H. Robinson, W. I. 38 Sundquist, M. F. Summers, and C. P. Hill, Structure of the antiviral assembly inhibitor CAP-1 complex with the HIV-1 CA protein, J Mol Biol, 373 (2007), 355-66.
- N. Goudreau, C. T. Lemke, A. M. Faucher, C. Grand-Maitre, S. Goulet, J. E. Lacoste, 39 J. Rancourt, E. Malenfant, J. F. Mercier, S. Titolo, and S. W. Mason, Novel inhibitor binding site discovery on HIV-1 capsid N-terminal domain by NMR and X-ray crystallography, ACS Chem Biol, 8 (2013), 1074-82.

- 40 Z. Liu, Q. Pan, Z. Liang, W. Qiao, S. Cen, and C. Liang, The highly polymorphic cyclophilin A-binding loop in HIV-1 capsid modulates viral resistance to MxB, Retrovirology, 12 (2015), 1.
- L. Lamorte, S. Titolo, C. T. Lemke, N. Goudreau, J. F. Mercier, E. Wardrop, V. B.
 Shah, U. K. von Schwedler, C. Langelier, S. S. R. Banik, C. Aiken, W. I. Sundquist,
 and S. W. Mason, Discovery of Novel Small-Molecule HIV-1 Replication Inhibitors
 That Stabilize Capsid Complexes, Antimicrob Agents Ch, 57 (2013), 4622-31.
- S. R. Yant, A. Mulato, D. Hansen, W. C. Tse, A. Niedziela-Majka, J. R. Zhang, G. J. Stepan, D. Jin, M. H. Wong, J. M. Perreira, E. Singer, G. A. Papalia, E. Y. Hu, J. Zheng, B. Lu, S. D. Schroeder, K. Chou, S. Ahmadyar, A. Liclican, H. Yu, N. Novikov, E. Paoli, D. Gonik, R. R. Ram, M. Hung, W. M. McDougall, A. L. Brass, W. I. Sundquist, T. Cihlar, and J. O. Link, A highly potent long-acting small-molecule HIV-1 capsid inhibitor with efficacy in a humanized mouse model, Nat Med, 25 (2019), 1377-84.
- A. Saito, D. Ferhadian, G. A. Sowd, E. Serrao, J. Shi, U. D. Halambage, S. Teng, J. Soto, M. A. Siddiqui, A. N. Engelman, C. Aiken, and M. Yamashita, Roles of Capsid-Interacting Host Factors in Multimodal Inhibition of HIV-1 by PF74, J. Virol., 90 (2016), 5808-23.
- J. Shi, J. Zhou, V. B. Shah, C. Aiken, and K. Whitby, Small-molecule inhibition of human immunodeficiency virus type 1 infection by virus capsid destabilization, J Virol, 85 (2011), 542-9.

- L. Sun, T. Huang, A. Dick, M. E. Meuser, W. A. Zalloum, C. H. Chen, X. Ding, P. Gao, S. Cocklin, K. H. Lee, P. Zhan, and X. Liu, Design, synthesis and structure-activity relationships of 4-phenyl-1H-1,2,3-triazole phenylalanine derivatives as novel HIV-1 capsid inhibitors with promising antiviral activities, Eur J Med Chem, 190 (2020), 112085.
- G. Wu, W. A. Zalloum, M. E. Meuser, L. Jing, D. Kang, C. H. Chen, Y. Tian, F. Zhang, S. Cocklin, K. H. Lee, X. Liu, and P. Zhan, Discovery of phenylalanine derivatives as potent HIV-1 capsid inhibitors from click chemistry-based compound library, Eur J Med Chem, 158 (2018), 478-92.
- L. Sun, A. Dick, M. E. Meuser, T. Huang, W. A. Zalloum, C-H Chen, S. Cherukupalli, S. Xu, X. Ding, P. Gao, D. Kang, E. D. Clercq, C. Pannecouque, S. Cocklin, K-H Lee, X. Liu, and P. Zhan, Design, Synthesis, and Mechanism Study of Benzenesulfonamide-Containing Phenylalanine Derivatives as Novel HIV-1 Capsid Inhibitors with Improved Antiviral Activities, J. Med. Chem., 63 (2020), 4790-810.
- L. Wang, M. C. Casey, S. K. V. Vernekar, H. T. Do, R. L. Sahani, K. A. Kirby, H. Du,
 A. Hachiya, H. Zhang, P. R. Tedbury, J. Xie, S. G. Sarafianos, and Z. Wang,
 Chemical profiling of HIV-1 capsid-targeting antiviral **PF74**, Eur. J. Med. Chem.,
 200 (2020), 112427.
- S. K. V. Vernekar, R. L. Sahani, M. C. Casey, J. Kankanala, L. Wang, K. A. Kirby, H.
 Du, H. Zhang, P. R. Tedbury, J. Xie, S. G. Sarafianos, and Z. Wang, Toward
 Structurally Novel and Metabolically Stable HIV-1 Capsid-Targeting Small
 Molecules, Viruses, 12 (2020).

- J. P. Xu, A. C. Francis, M. E. Meuser, M. Mankowski, R. G. Ptak, A. A. Rashad, G.
 B. Melikyan, and S. Cocklin, Exploring Modifications of an HIV-1 Capsid Inhibitor:
 Design, Synthesis, and Mechanism of Action, J Drug Des Res, 5 (2018).
- V. J. Wacher, J. A. Silverman, Y. Zhang, and L. Z. Benet, Role of P-glycoprotein and cytochrome P450 3A in limiting oral absorption of peptides and peptidomimetics, J Pharm Sci, 87 (1998), 1322-30.
- 52 L. Xu, and M. C. Desai, Pharmacokinetic enhancers for HIV drugs, Curr Opin Investig Drugs, 10 (2009), 775-86.
- 53 N. M. Midde, Y. Q. Gong, T. J. Cory, J. H. Li, B. Meibohm, W. H. Li, and S. Kumar, Influence of Ethanol on Darunavir Hepatic Clearance and Intracellular PK/PD in HIV-Infected Monocytes, and CYP3A4-Darunavir Interactions Using Inhibition and in Silico Binding Studies, Pharm Res-Dordr, 34 (2017), 1925-33.
- 54 N. Buss, P. Snell, J. Bock, A. Hsu, and K. Jorga, Saquinavir and ritonavir pharmacokinetics following combined ritonavir and saquinavir (soft gelatin capsules) administration, Br J Clin Pharmacol, 52 (2001), 255-64.
- L. H. Xu, H. T. Liu, B. P. Murray, C. Callebaut, M. S. Lee, A. Hong, R. G. Strickley,
 L. K. Tsai, K. M. Stray, Y. J. Wang, G. R. Rhodes, and M. C. Desai, Cobicistat (GS-9350): A Potent and Selective Inhibitor of Human CYP3A as a Novel Pharmacoenhancer, Acs Med Chem Lett, 1 (2010), 209-13.
- 56 M. C. Lo, A. Aulabaugh, G. Jin, R. Cowling, J. Bard, M. Malamas, and G. Ellestad, Evaluation of fluorescence-based thermal shift assays for hit identification in drug discovery, Anal Biochem, 332 (2004), 153-9.

- 57 Y. Miyazaki, N. Doi, T. Koma, A. Adachi, and M. Nomaguchi, Novel In Vitro Screening System Based on Differential Scanning Fluorimetry to Search for Small Molecules against the Disassembly or Assembly of HIV-1 Capsid Protein, Front Microbiol, 8 (2017), 1413.
- M. W. Pantoliano, E. C. Petrella, J. D. Kwasnoski, V. S. Lobanov, J. Myslik, E. Graf,
 T. Carver, E. Asel, B. A. Springer, P. Lane, and F. R. Salemme, High-density
 miniaturized thermal shift assays as a general strategy for drug discovery, J Biomol
 Screen, 6 (2001), 429-40.
- A. Adachi, H. E. Gendelman, S. Koenig, T. Folks, R. Willey, A. Rabson, and M. A. Martin, Production of acquired immunodeficiency syndrome-associated retrovirus in human and nonhuman cells transfected with an infectious molecular clone, J Virol, 59 (1986), 284-91.
- 60 Schrödinger Small-Molecule Drug Discovery Suite 2019-1, Schrödinger, LLC, New York, NY, 2019.
- 61 Schrödinger Release 2019-1: Maestro, Schrödinger, LLC, New York, NY, 2019.
- 62 G. M. Sastry, M. Adzhigirey, T. Day, R. Annabhimoju, and W. Sherman, Protein and ligand preparation: parameters, protocols, and influence on virtual screening enrichments, J Comput Aid Mol Des, 27 (2013), 221-34.
- 63 W. L. Jorgensen, D. S. Maxwell, and J. TiradoRives, Development and testing of the OPLS all-atom force field on conformational energetics and properties of organic liquids, J Am Chem Soc, 118 (1996), 11225-36.
- 64 Schrödinger Release 2019-1: LigPrep, Schrödinger, LLC, New York, NY, 2019.

R. A. Friesner, J. L. Banks, R. B. Murphy, T. A. Halgren, J. J. Klicic, D. T. Mainz, M.
P. Repasky, E. H. Knoll, M. Shelley, J. K. Perry, D. E. Shaw, P. Francis, and P. S.
Shenkin, Glide: a new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy, J Med Chem, 47 (2004), 1739-49.

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Highlights

- Design of novel analogs and sub-chemotypes of HIV-1 CA-targeting antiviral PF74. •
- 5-Hydroxyindole analogs (8,9 and 12) showed much improved potency over PF74 •
- 2-Indolone analogs (16-24) decreased the T_m of CA hexamers •
- The potencies of α and β -naphthyl analogs (33 and 27) were comparable to PF74 •

Declaration of interest statement

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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