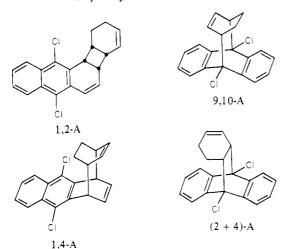
Adiabatic S₁ 9,10-Dichloroanthracene Formation from Electronically Excited 9,10-Dichloroanthracene/1,3-Cyclohexadiene Photoadducts¹

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Abstract: The photochemistry and fluorescence of the two major photoadducts of 9,10-dichloroanthracene (DCA) and 1,3-cyclohexadiene (CHD) are described. The 1,2-adduct, 1,2-A ($[2_{\pi_i} + 2_{\pi_i}]$ to the 1,2-positions of DCA), gives DCA and CHD as main photoproducts in several solvents, $\phi_{-1,2-A} \simeq \phi_{DCA} \simeq \phi_{CHD} \simeq 0.3$. It exhibits fluorescence characteristic of the naphthalene moiety (350-435 nm) and, in addition, gives DCA fluorescence (390-560 nm). The pronounced oxygen quenching effect of the latter shows that cleavage of singlet excited 1,2-A, 1 1,2-A*, is in part adiabatic, $\phi_{^{1}DCA^{*}} \simeq 0.007-0.015$. The 9,10-adduct, 9,10-A ($[4_{\pi_i} + 4_{\pi_i}]$ to the 9,10-position of DCA), is efficiently destroyed by light, $\phi_{-9,10-A} \simeq 1.0$, but gives several products in addition to DCA and CHD, $\phi_{DCA} \simeq \phi_{CHD} \simeq 0.11$. Its fluorescence is identical with DCA fluorescence, corresponding to $\phi_{1pcA^*} \simeq 0.0041-0.0068$, depending on the solvent. The observations are discussed by using the current theoretical model for electrocyclic photochemical reactions. They suggest that if pericyclic minima are involved in adduct photocleavage and formation, they are isolated in the potential energy surface and do not readily interconvert.

Photocycloaddition of 9,10-dichloroanthracene (DCA) to 1,3cyclohexadiene (CHD) yields three major adducts corresponding to allowed $[2_{\pi_s} + 2_{\pi_s}]$ addition of CHD to the 1,2-positions of DCA (1,2-adduct) and $[4_{\pi_s} + 4_{\pi_s}]$ additions of CHD to the 9,10- and



1,4-positions of DCA (9,10- and 1,4-adducts).^{2,3} A reversibly formed nonfluorescing singlet exciplex, \(^1(DCA-CHD)^*\), is probably a common intermediate in these reactions.3 The mechanism for the formation of a minor (<1%) forbidden $[2_{\pi_s} + 4_{\pi_s}]$ adduct [(2 + 4)-adduct] has not been elucidated.3

Photochemically allowed concerted cycloadditions are characterized by correlation between the singly excited (S) states of addends and adduct, correlation of the doubly excited (D) state of addends with the ground (G) state adduct, and correlation of ground-state addends with doubly excited adduct (Figure 1).4

Intended, but avoided, crossing of the D and G states creates a pericyclic minimum on the excited electronic surface at the biradicaloid geometry where new bonds are partly formed and old bonds partly broken.4 At larger intermolecular separation the exciplex energy minimum on S provides a reservoir of addends potentially ready for cycloaddition. If reaction is initiated by electronic excitation of the adduct, leakage from the pericyclic minimum and adiabatic⁵ conversion along the S surface are energetically feasible pathways to electronically excited products, exciplex, and/or addends. Decay through the "funnel" or "hole" at pericyclic minima provides diabatic reaction paths to groundstate addends and adducts.4 Diabatic pathways usually predominate and adiabatic photocleavage reactions are generally inefficient.6,7 Inspection of Figure 1 shows that the photophysical and photochemical properties of [2 + 2] and [4 + 4] cycloadducts may provide a useful approach for examining the theoretical pericyclic minima.8

An earlier failure to detect 1,2-A² was traced to its efficient photocleavage to DCA and CHD.³ Those initial observations and current interest in adiabatic and diabatic photocleavage reactions prompted the following spectroscopic and photochemical study of the 1,2- and 9,10-adducts. The findings reveal significant adiabatic photoreaction channels in both cases and suggest that if pericyclic minima are involved in the diabatic pathways, they differ significantly for the two adducts and do not interconvert.

Spectroscopic Measurements. Fluorescence spectra, uncorrected for nonlinearity of instrumental response, obtained by 320-nm excitation of argon-purged and oxygen-saturated acetonitrile solutions of 1,2-A are shown in Figure 2. Maximum intensities of emission bands are at 365, 383, 405, 428, and 454 nm. Excitation at 254 or 340 nm, where DCA absorbs strongly, rather than at 320 nm, the λ_{max} of 1,2-A, sharply increases the intensity of the 405-, 428-, and 454-nm bands relative to that of the 365and 383-nm bands. The excitation spectrum obtained by monitoring 383-nm emission is analogous to the UV spectrum of 1,2-A, whereas monitoring emission at 454 nm gives an excitation spectrum which resembles a combination of 1,2-A and DCA UV

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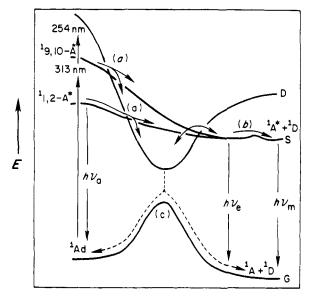
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Reaction Coordinate

Figure 1. Potential energy diagram for [2 + 2] and [4 + 4] cycloreversion illustrating adiabatic ($a \rightarrow b$, solid arrows) and diabatic ($a \rightarrow$ c, broken arrows) photoreaction channels; $h\nu_a$, $h\nu_e$, and $h\nu_m$ designate adduct, exciplex, and addend fluorescence, respectively.

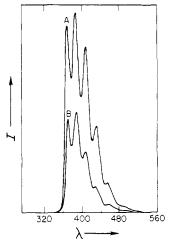


Figure 2. Fluorescence spectra (uncorrected) of argon-purged (curve A) and oxygen-saturated (curve B) CH₃CN solutions of 1,2-A, 7.9 \times 10⁻⁵ M: 320-nm excitation

absorption profiles (Figure 3). The relative intensity of the DCA-like fluorescence region increases rapidly upon irradiation of 1,2-adduct solutions at $\lambda < 370$ nm. The extent of 1,2-A photodegradation in the fluorometer was minimized by using narrow excitation slits, high adduct concentrations, short irradiation times, and fresh solutions following each measurement. 1,2-A samples employed for photochemical and spectroscopic measurements had a zero-time DCA content of $0.15 \pm 0.05\%$ of [1,2-A] (UV, 402 nm). Fluorescence spectra of benzene and pyridine solutions of 1,2-A are red-shifted \sim 2 nm from those in Figure 2.

The fluorescence of acetonitrile and cyclohexane solutions of 9,10-A was examined as a function of excitation wavelength, 240-340 nm, and emission wavelength, 280-560 nm. Only a weak emission identical with DCA fluorescence was detected (Figure 4). Exposure of 9,10-A solutions to the fluorometer excitation beam for prolonged periods of time, e.g., 5-10 min at 270 nm, caused a gradual increase of the intensity of the DCA emission. Due to the presence of trace DCA impurity in 9,10-A samples, excitation of fresh solutions in the 300-390 nm region, where DCA only absorbs, yields DCA fluorescence but at lower intensities than from preirradiated solutions. Comparison of DCA fluorescence intensity from a fresh 2.4×10^{-4} M 9,10-A solution with that from

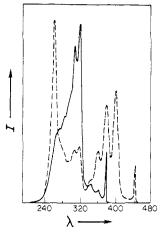


Figure 3. Excitation spectra (uncorrected) of air-saturated solutions of 1,2-A, 3.2 \times 10⁻⁶ M, in CH₃CN: solid curve, monitoring at 383 nm; dashed curve, monitoring at 454 nm.

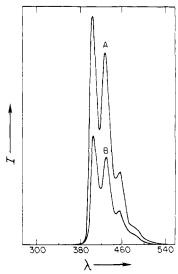


Figure 4. Fluorescence spectra (uncorrected) of DCA (curve A) and 9,10-A (curve B) in air-saturated cyclohexane; 270-nm excitation.

Table I. Fluorescence Quantum Yields

	DCA,a		1,2-A ^b		9,10-A, ^c
solvent	$\phi_{\mathbf{f}}$	$\phi_{ t ft}$	ϕ_{fa}	$\phi_{ extsf{fDCA}}$	$\phi_{ extsf{fDCA}}$
C_6H_6	0.67^{d}	0.037	0.024	0.013	
C_6H_{12}	0.47				0.0032
CH ₃ CN	0.49	0.029	0.017	0.012	0.0020
C_sH_sN	0.70	0.034	0.022	0.012	

 a [DCA] \simeq 6 × 10⁻⁶ M; excitation wavelength 340 nm; degassed solutions. b [1,2-A] \simeq 3 × 10⁻⁵ M; [DCA] \simeq 5 × 10⁻⁴ M; excitation wavelength 320 nm; argon-purged solutions. c [9,10-A] \simeq 2 × 10⁻⁴ M; [DCA] \simeq 4 × 10⁻⁵ M; excitation wavelength 270 nm; argon-purged solutions. d Used as the standard; average value from ref 9-11.

a standard 2.4×10^{-6} M DCA solution in cyclohexane established that the zero-time DCA content of 9,10-A samples used in all measurements was 0.0009% of [9,10-A]. Excitation spectra of fresh 9,10-A solutions are analogous to the adduct's UV absorption spectrum (Figure 5).

Fluorescence quantum yields were determined at 25.0 °C by using DCA fluorescence in degassed benzene as standard, ϕ_f = $0.67 \pm 0.03^{9-11}$ (Table I). The fluorescence spectra were corrected for nonlinearity of instrumental response and changes in refractive

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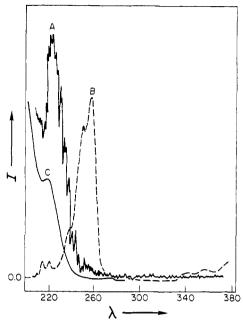


Figure 5. Excitation spectra (uncorrected, but Varian SF-330 fluorometer employed) for air-saturated C_6H_{12} solutions of 9,10-A (curve A) and DCA (curve B), each 2.4×10^{-6} M, monitoring emission at 410 nm. Curve C is 9,10-A's UV absorption profile.3

Table II. Fluorescence Quenching by Oxygen

solute	conditions ^a	$I_{o}/I_{a}{}^{b}$	I_{o}/I_{ox}^{b}
DCA	CH ₃ CN, 53, 428	$1.38 \pm 0.01 (1.39)$	2.83 ± 0.07
DCA	$C_6 H_6, 53, 433$	$1.34 \pm 0.02 (1.30)$	2.76 ± 0.01
DCA	C_6H_{12} , 2.4, 428°	1.15 ± 0.01	
DCA	C, H, N, 53, 434	$1.22 \pm 0.01 (1.17)$	2.00 ± 0.02
1,2-A	CH ₃ CN, 79, 382	1.21 ± 0.01	2.04 ± 0.04
1,2-A	$C_6 H_6$, 76, 383	1.26 ± 0.01	2.34 ± 0.04
1,2-A	C ₅ H ₅ N, 79, 385	1.11 ± 0.01	1.54 ± 0.01
9,10-A	CH ₃ CN, 244, 428 ^c	1.31 ± 0.06	2.85 ± 0.01
9,10-A	$C_6 H_{12}$, 238, 428°	1.15 ± 0.03	

^a Solvent and solute concentration in μ M; monitoring wavelength in nm; 25.0 °C throughout; excitation wavelength 320 nm unless noted otherwise. b Error limits are average deviations from the mean of two or more determinations; numbers in parentheses obtained relative to degassed solution I_0 's. C Excitation at 270 nm, filter system 1; see Experimental Section.

index.¹² Identical emission intensities were obtained from degassed or argon-purged DCA solutions. Corrected fluorescence spectra from 1,2-A were decomposed into 11,2-A* and 1DCA* emission contributions by attributing all emission at $\lambda \ge 485$ nm to DCA fluorescence, e.g., Figure 6. This allowed separation of total fluorescence quantum yields, ϕ_{ft} , into 1,2-A, ϕ_{fa} , and DCA, ϕ_{fDCA} , components (Table I).

Relative emission intensities of argon-, I_0 , air-, I_a , and oxygen-purged, I_{ox} , solutions are shown in Table II. In the case of 1,2-A the effect of oxygen depends on the emission wavelength (Table III).

The effect of DCA contamination on the fluorescence yield of 9,10-A was determined by measuring relative emission intensities of air-saturated cyclohexane solutions of 9,10-A with and without added DCA (Table IV). The emission intensity of these solutions for 340-nm excitation where only DCA absorbs was shown to be proportional to [DCA].¹³ For $[9,10-A] = 2.38 \times 10^{-4} \text{ M}$, the undoped solution was found to contain 2.06×10^{-9} M DCA, which increased to 4.94×10^{-9} M under the conditions used in obtaining a single fluorescence spectrum (270 nm, filter 1, \sim 2 min).

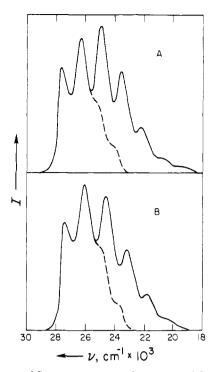


Figure 6. Corrected fluorescence spectra of argon-purged CH3CN (curve A) and C₆H₆ (curve B) solutions; 320-nm excitation. Dashed curves obtained by subtraction of DCA fluorescence from the total spectrum.

Photochemical Observations. (A) Product Studies. Benzene-d₆ and cyclohexane- d_{12} solutions of the adducts were degassed in NMR tubes, flame-sealed at a constriction, and the progress of the photoreactions (~23 °C) was followed by ¹H, NMR, 270 MHz. Tetramethylsilane, ~ 0.003 M, and cyclohexane- d_{11} ¹H resonances were employed as internal standards (Table V). At conversions below 20%, loss of 1,2-A equals DCA and CHD yields within the experimental uncertainty of the measurements (±10%) for either 254- or 313-nm excitation. At higher conversions DCA and CHD yields do not account for adduct loss, probably due to secondary photochemical reactions. The ¹H NMR spectrum of 1.2-A which was irradiated at 254 nm to 55% conversion in C₆D₁₂ showed (in addition to strong DCA and CHD signals) weak multiplets centered at δ 3.2-5.2, δ 4.8-5.2, and δ 7.2 which do not correspond to any of the other known photoadducts. GLC analysis of this solution gave peaks with retention times identical with those of 9-chloroanthracene, MCA (2.2% relative area), DCA (43%), 9,10-A (1.5%), and 1,2-A (49%) as well as two peaks (\sim 2.5% each) with retention times similar to 9,10-A. The multiplets of the unknown photoproduct(s) were not discernible in NMR spectra of 1,2-A solutions irradiated (254 nm) to \sim 25% conversion. NMR spectra of C₆D₁₂ solutions of 9,10-A irradiated (254 nm) to 24-59% conversion show weak resonances corresponding to DCA and CHD (8% of 9,10-A consumed), a multiplet at δ 3.0, and strong broad signals comprised of several overlapping multiplets at δ 5.4–5.6 and δ 6.6–7.5. Once again, these unassigned resonances do not correspond to any of the other known photoadducts. GLC analysis of a 9,10-A C₆D₁₂ solution irradiated (254 nm, filter 3) to $\sim 56\%$ conversion (NMR) showed peaks with retention times identical with those of MCA (3.7%, relative area), DCA (0.9%), 9,10-A (76%), and 1,2-A $(\sim 0.3\%)$, as well as a GLC peak (17%) with a retention time between those of DCA and 9,10-A and three small peaks ($\sim 0.7\%$ each) with retention times similar to that of 1,2-A.

(B) Quantum Yields. 1,2-A loss and DCA quantum yields for 254-nm excitation in CH₃CN and 313-nm excitation in CH₃CN, C_6H_6 , and C_6H_5N were determined for degassed and air-saturated solutions (Table VI). Irradiations were conducted in a Moses merry-go-round apparatus¹⁴ at 25.0 \pm 0.1 °C. The direct pho-

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Table III. Wavelength Dependence of Oxygen Quenching of 1,2-A Emission^a

emission band b λ_{\max}^c		CH₃CN		C ₆ H ₆			C ₅ H ₅ N		
	λ_{\max}^c	$I_{\rm O}/I_{\rm a}$	I_{o}/I_{ox}	λ_{\max}^c	$I_{\rm o}/I_{\rm a}$	I_{o}/I_{ox}	λ_{\max}^c	$I_{ m o}/I_{ m a}$	$I_{\rm o}/I_{\rm ox}$
0-0	365	1.23	1.99	365	1.28	2.33	366	1.11	1.53
0-1	382	1.23	1.99	383	1.26	2.31	385	1.11	1.54
0-0'	404	1.38	2.65	405	1.39	2.81	406	1.18	1.79
0-1'	428	1.50	3.36	429	1.48	3.42	432	1.24	2.07
0-2'	454	1.55	3.59	456	1.52	3.68	456	1.27	2.16

^a Samples and conditions described in Table II. ^b Primed bands assigned to ¹DCA*. ^c Emission wavelength in nm.

Table IV. Relative Fluorescence Intensities of 9,10-A Solutions Doped with DCA^a

[DCA]/[DCA] _o	$I_{\mathbf{d}}/I_{\mathbf{o}}^{\ b}$	
1.00	1.00	
26.4	1.18	
307	3.10	
752	6.15	
	1.00 26.4 307	1.00 1.00 26.4 1.18 307 3.10

^a Air-saturated $C_6 H_{12}$; 25.0 °C; [9,10-A] = 2.38 × 10⁻⁴ M; excitation at 270 nm (filter 1); emission at 405 nm. b Ratio of doped to undoped fluorescence intensity.

Table V. Photoproduct Distribution, 270-MHz ¹H NMR^a

solvent			% product appearance		
	adduct; M	adduct loss, b %	DCA	CHD	
C_6D_6	1,2-A; 0.033 ^c	6. ₅ ± 1	6. ₂ ± 1	6. ₂ ± 1	
C_6D_6	$1,2-A;0.033^c$	12.7 ± 1	$13{2} \pm 2$	10. ₉ ± 2	
C_6D_6	$1,2-A;0.033^c$	47.0 ± 6	$31., \pm 4$	28.0 ± 4	
C_6D_{12}	$1,2-A;0.034^d$	22.6 ± 3	19.7 ± 2	19.7 ± 2	
C_6D_{12}	1,2-A; 0.034 ^d	29., ± 3	22.4 ± 2	22., ± 2	
C_6D_1	$1,2-A;0.034^d$	$54{0} \pm 5$	37., ± 4	37., ± 4	
$C_6 D_{12}$	9,10-A; 0.034 ^e	$59{9}^{4} \pm 6$	$4{4}^{1} \pm 1$	$4{4}^{1} \pm 1$	

^a Degassed NMR tubes; ~23 °C. ^b Relative to initial adduct concentration; error limits reflect precision in NMR integration. c 313 nm, filter 2. d 254 nm, filter 3. e 254 nm, filter 4.

toisomerization of trans-stilbene in degassed acetonitrile was used for actinometry; $\phi_{t\rightarrow c} = 0.51 \pm 0.02$ reported^{15,16} for *n*-pentane, 313 nm, was assumed to apply for acetonitrile. Stationary states for stilbene photoisomerization in CH₃CN were shown to be 45.7 \pm 0.1% cis, 254 nm, 8.92 × 10⁻⁴ M stilbene, and 91.2 \pm 0.1% cis, 313 nm, 4.46×10^{-4} M stilbene. These values are very close to those reported for n-pentane and were used for back-reaction corrections. 15 Corrected conversions to cis-stilbene ranged from 4.49 to 9.72%. When necessary, larger conversions were avoided by irradiating two actinometer solutions in series with the same adduct sample, and the initial [trans-stilbene] was adjusted from 4.46×10^{-4} to 8.92×10^{-4} M.¹⁷ Results from duplicate actinometer samples irradiated in parallel were in excellent agreement. Freshly irradiated solutions were analyzed for 1,2-A loss and DCA appearance by UV spectroscopy, monitoring 1,2-A at $\lambda_{max} \simeq 320$ nm and DCA at $\lambda_{max} \simeq 402$ nm;³ conversions ranged between 5.88 and 13.1%.

Solutions of 9,10-A in degassed and air-saturated cyclohexane were irradiated (254 nm) in parallel with 1,2-A solutions. DCA formation was monitored by UV absorption in this case also. UV spectra of irradiated 9,10-A solutions show, in addition to DCA absorption, two weak bands centered at 306 and 318 nm similar to those of 1,2-A. Assuming that the unknown product has the same absorptivity coefficient as 1,2-A gives $\phi \simeq \frac{1}{3}\phi_{DCA}$. In the presence of air the 306-318 nm region could not be analyzed due to an additional broad shoulder at $\lambda < 340$ nm. Conversions to DCA ranged from 0.0343 to 0.0675%, and by inference from the NMR observations (see above), 9,10-A loss was less than 1%.

Table VI. Adduct Loss and DCA Formation Quantum Yields

adduct ^a	$^{\lambda_{\mathrm{exc}},^{b}}_{\mathrm{nm}}$	solvent	$\phi_{-\mathbf{A}}^{c}$	$\phi_{\mathbf{DCA}}^{\mathbf{c}}$
1,2-A	254	CH ₃ CN	0.32	0.30
1,2-A	254	CH_3CN^d	0.25 ± 0.01	0.25 ± 0.02
1,2-A	313	CH¸CN	0.36 ± 0.02	0.35 ± 0.04
1,2-A	313	CH_3CN^d	0.28 ± 0.01	0.26 ± 0.01
1,2-A	313	C_6H_6	0.27 ± 0.05	0.27 ± 0.03
1,2-A	313	C_5H_5d	0.21 ± 0.01	0.18 ± 0.01
1,2-A	313	C,H,N	0.31 ± 0.01	0.30 ± 0.03
1,2-A	313	$C_5H_5N^d$	0.28 ± 0.01	0.24 ± 0.01
9,10-A	254	$C_{\bullet}H_{\bullet}$	1.0^{e}	0.11 ± 0.01
9,10-A	254	$C_6H_{12}^{12}d$		0.11

Quantum yields are reported in Table VI.

The results described above will be discussed in terms of the mechanism

$${}^{1}Ad \xrightarrow{h\nu} {}^{1}Ad*$$
 (1)

$${}^{1}\mathrm{Ad}^{*} \xrightarrow{k_{\mathrm{fa}}} {}^{1}\mathrm{Ad} + h\nu_{\mathrm{a}} \tag{2}$$

$${}^{1}\mathrm{Ad}^{*} \xrightarrow{k_{\mathrm{isa}}} {}^{3}\mathrm{Ad}^{*} \rightarrow {}^{1}\mathrm{Ad} \tag{3}$$

$${}^{1}\text{Ad}* \xrightarrow{k_{\text{da}}} \alpha^{1}\text{DCA} + \alpha^{1}\text{CHD} + \beta\text{P} + (1 - \alpha - \beta)^{1}\text{Ad}$$
 (4)

$${}^{1}\text{Ad}^{*} \xrightarrow{k_{\text{ca}}} {}^{1}(\text{DCA}\cdot\text{CHD})^{*}$$
 (5)

$$^{1}(DCA \cdot CHD)^{*} \xrightarrow{k_{de}} ^{1}DCA + ^{1}CHD$$
 (6)

$$^{1}(DCA\cdot CHD)^{*} \xrightarrow{k_{pc}} adducts^{3}$$
 (7)

$$^{1}(DCA\cdot CHD)^{*} \stackrel{k_{*}}{\longleftrightarrow} ^{1}DCA^{*} + ^{1}CHD$$
 (8)

$${}^{1}DCA^{*} \xrightarrow{k_{fm}} {}^{1}DCA + h\nu_{m}$$
 (9)

$${}^{1}DCA^{*} \xrightarrow{k_{\mathsf{ism}}} {}^{3}DCA^{*} \rightarrow {}^{1}DCA$$
 (10)

where Ad is an adduct, P represents unknown products, ¹(DCA·CHD)* is the singlet exciplex, and all other symbols are self-explanatory. Equations 6-10 have been discussed in accounting for the DCA/CHD cycloaddition reactions.³ Reference to Figure 1 shows that pericyclic minima on the D surface are proposed intermediates for adiabatic cleavage. Our results do not require involvement of these intermediates, so they are not explicitly included in the mechanism.

Spectroscopic Observations. The emission and excitation spectra of 9,10-A reveal unequivocally the existence of an adiabatic reaction channel in its photocleavage (Figures 4 and 5). Furthermore, the DCA doping experiments (Table IV) show that all fluorescence observed following its excitation is due to ¹DCA* formed via this channel and not by excitation of the miniscule amount of DCA present as a contaminant.

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⁽¹⁷⁾ A detailed description of the results is given in Smothers, W. K., Ph.D. Dissertation, The Florida State University, Tallahassee, Florida, 1981.

 $^{^{}a}$ [1,2-A]_o = 4.55 × 10⁻⁴ or 4.6 × 10⁻⁴ M; [9,10-A]_o = 0.0385 M. b Excitation at 254 nm, filter 5; excitation at 313 nm, filter 6. ^c Uncertainties are average deviations from the mean of two or more independent determinations. d Air-saturated solutions. ^e From ϕ_{-A}/ϕ_{DCA} from NMR experiments.

DCA contamination of 1,2-A is more serious and significantly contributes to the fluorescence measurements especially when excitation is in regions where DCA absorbs strongly (Figures 2 and 3). The excitation spectrum obtained by monitoring emission at 383 nm is analogous to the UV absorption profile of 1,2-A as expected for emission originating entirely from ¹1,2-A*. On the other hand, monitoring DCA emission at 454 nm gives an excitation spectrum which resembles a mixture of DCA and 1,2-A absorption spectra. Since 11,2-A* does not emit at 454 nm (Figure 6), the appearance of the adduct-like absorption profile in the 300-330 nm region of the excitation spectrum suggests strongly an adiabatic pathway to ¹DCA* from ¹1,2-A*. This conclusion is confirmed by the influence of molecular oxygen on the total fluorescence envelope of 1,2-A solutions. Inspection of Figure 2 and Table III shows that ¹DCA* fluorescence quenching is more pronounced than 11,2-A* fluorescence quenching. Furthermore, since ¹DCA* fluorescence generated by excitation of 1,2-A solutions is quenched more strongly than fluorescence generated by exciting DCA directly (compare Tables II and III), at least part of the DCA fluorescence from the 1,2-A solutions must derive from adiabatic cleavage of ¹1,2-A*. This conclusion can be understood by including the following quenching steps in the mechanism:

$${}^{1}\text{Ad*} + \text{O}_{2} \xrightarrow{k_{qa}} \text{quenching}$$
 (11)

$$^{1}DCA^* + O_2 \xrightarrow{k_{qm}} quenching$$
 (12)

$$^{1}(DCA \cdot CHD)^{*} \xrightarrow{k_{qe}} quenching$$
 (13)

Application of the steady-state approximation to all excited species in eq 1-13 gives

$$\left(\frac{I_{o}}{I}\right)_{\lambda} = \left[\frac{\sigma_{\lambda}}{1 + k_{qa}\tau_{a}[O_{2}]} + \frac{1 - \sigma_{\lambda}}{(1 + k_{qa}\tau_{a}[O_{2}])(1 + k_{qe}\tau_{e}[O_{2}])(1 + k_{qm}\tau_{m}[O_{2}])}\right]^{-1} (14)$$

where $\tau_a = (k_{\rm fa} + k_{\rm isa} + k_{\rm da} + k_{\rm ea})^{-1}$, $\tau_{\rm e} = (k_{\rm e} + k_{\rm de} + k_{\rm pe})^{-1}$ and $\tau_{\rm m} = (k_{\rm fm} + k_{\rm ism})^{-1}$ are the lifetimes of $^{\rm I}{\rm Ad}^*$, $^{\rm I}({\rm DCA\cdot CHD})^*$ at [CHD] = 0, and $^{\rm I}{\rm DCA}^*$, respectively, and σ_{λ} is the fraction of light emitted by $^{\rm I}{\rm Ad}^*$ at a specific wavelength λ in the absence of ${\rm O}_2$. This expression can be simplified since the exciplex, to the extent that it participates in the cycloaddition reaction, is nonemitting and too short-lived to be quenched by oxygen, i.e., $k_{\rm qe}\tau_{\rm e}[{\rm O}_2]\simeq 0.3$ Inspection of Figure 6 shows that $\sigma_{\lambda}=1$ for 0,0 and 0,1 bands of $^{\rm I}{\rm Ad}^*$ fluorescence. It follows that at those wavelengths eq 14 reduces to

$$\left(\frac{I_o}{I}\right)_{0,0} = \left(\frac{I_o}{I}\right)_{0,1} = 1 + k_{qa}\tau_a[O_2]$$
 (15)

the ordinary Stern-Volmer relationship, as is borne out by the results in Table III. Similarly, it can be seen that emission at the 0,2' band corresponds to the limiting condition of $\sigma_{\lambda}=0$ which when applied to eq 14 along with $k_{\rm qc} \tau_{\rm e}[{\rm O}_2]=0$, gives

$$\left(\frac{I_o}{I}\right)_{0.2} = (1 + k_{qa}\tau_a[O_2])(1 + k_{qm}\tau_m[O_2])$$
 (16)

Should other intermediates, such as pericyclic minima of the D state(s), lie in the path to $^1\mathrm{DCA}^*$ and should these also be quenched by oxygen, the oxygen effect would be larger than predicted by eq 16. Since the first term in eq 16 is known from eq 15 and the second term describes the effect of oxygen on directly excited DCA, eq 16 can be expressed as $(I_o/I)_{0.2'} = (I_o/I)_{0.0'}$ ($I_o/I)_{DCA}$ which are quantities given in Tables II and III. Values calculated in this way (Table VII) are generally larger than the observed values which are nevertheless significantly larger than experimental values for directly excited $^1\mathrm{DCA}^*$. While these observations establish an adiabatic pathway to $^1\mathrm{DCA}^*$, they require that part of the $^1\mathrm{DCA}^*$ emission from 1,2-A solution excited

Table VII. Calculated Oxygen Effects on Fluorescence of ¹DCA* Formed Adiabatically from ¹1,2-A* ^a

solvent	$I_{\rm o}/I_{\rm a}$	ξab	I_{o}/I_{ox}	ξox
CH,CN	1.68 ± 0.02	0.63	5.77 ± 0.18	0.42
C, H,	1.66 ± 0.03	0.57	6.46 ± 0.11	0.44
C,H,N	1.33 ± 0.04	0.41	3.08 ± 0.04	0.21

^a Using eq 16 and the observations in Tables II and III; ranges are standard deviations.
^b See ref 18.

at 320 nm arise by direct excitation of DCA present as an impurity. More specifically, simple algebraic manipulation of the expected and observed $I_{\rm o}/I$ values in Tables II, III, and VII for air-saturated solutions gives 0.63, 0.57, and 0.41 as the fractions of $^{\rm 1}DCA^*$ emission observed from 1,2-A solutions in CH₃CN, C₆H₆, and C₅H₅N, respectively, which originates from 1,2-A excitation. Somewhat lower DCA fluorescence contributions from 1,2-A excitation are predicted from the observations in oxygen-saturated solutions. It follows that approximately half of the $\phi_{\rm IDCA}$ values given for 1,2-A in Table I are due to adiabatic cleavage of $^{\rm 1}$ 1,2-A*. Sequential oxygen quenching effects have also been observed in the intramolecular adiabatic photocycloreversion of dimethyllepidopterene. To

In contrast to the results with 1,2-A, relative ¹DCA* emission intensities of argon-purged air- and oxygen-saturated 9,10-A solutions are, within the precision of the measurements, identical with those obtained by direct excitation of ¹DCA (Table II). It follows that the lifetime of the obligatory precursor 19,10-A*, as well as the lifetimes of any other excited-state intermediates in the $^{1}9,10\text{-A*} \rightarrow ^{1}DCA*$ path, is too short $(\tau < 1 \text{ ns})$ to be quenched by oxygen; i.e., $k_{qa}\tau_a[O_2]\ll 1$ in eq 16. This conclusion is consistent with measurements made on the 9,10-adduct obtained from 9-cyanoanthracene, CNA, and CHD. In this system ¹CNA* is observed in absorption within 33 ps of adduct excitation.7c It is also consistent with our inability to detect fluorescence directly from $^{19},10$ -A*, $\phi_{fa} < 10^{-4}$. The oxygen effect on 9,10-A-derived ¹DCA* fluorescence is reminiscent of observations on the photocleavage of 9-methylanthracene, ¹MA, dimer. In that case too, I_o/I_{ox} ratios for fluorescence from ${}^1{\rm MA}^*$ formed adiabatically from the singlet excited dimer, ¹MA*₂, or by direct ¹MA excitation are identical in CH₃CN.¹⁹ However, since a fluorescent excimer also forms adiabatically from ¹MA*₂ and should also be quenched by oxygen (the magnitude of this effect was, unfortunately, not reported), enhanced oxygen quenching of ¹MA* fluorescence would be expected on the basis of the ${}^{1}MA^{*}_{2} \rightarrow {}^{1}(MA \cdot MA)^{*} \rightarrow$ ¹MA* + MA sequence. ¹⁹ In the absence of such enhancement it was concluded that singlet excimer and ¹MA* form from ¹MA*₂ by independent adiabatic channels.¹⁹ Further elaboration on this mechanism has been based on the temperature dependencies of fluorescence quantum yields and lifetimes of ¹MA*₂, ¹(MA·MA)*, and ¹MA* in hydrocarbon solvents.²⁰ Since the DCA-CHD exciplex is nonfluorescent and short-lived,3 our observations do not explicitly address the interesting possibility of an independent adiabatic channel from either adduct S₁ state to ¹DCA*; since it is not needed in accounting for the results, it has been excluded from the mechanism.

According to the mechanism in eq 1-10, the quantum yields of adiabatically formed ¹DCA* are given by

$$\phi_{^{1}DCA^{*}} = \phi_{fDCA}/\phi_{fm} = k_{ea}\tau_{a}k_{-e}\tau_{e}$$
 (17)

where φ_{fm} is the quantum yield of DCA fluorescence from directly excited DCA.²¹ Values of $\phi_{^{1}DCA^{*}}$, calculated from the data in Tables I and VII, are shown in Table VIII. Stern-Volmer plots for the quenching of $^{1}DCA^{*}$ fluorescence by CHD have yielded

⁽¹⁸⁾ The expression used in these calculations is $(I/I_o)_{obsd} = \xi/(I_o/I)_{adiab} + (1-\xi)/(I_o/I)_{direct}$, where ξ is the fraction of DCA fluorescence originating by 1,2-A excitation.

⁽¹⁹⁾ Menter, J.; Förster, Th. Photochem. Photobiol. 1972, 15, 289. (20) Yamamoto, S.; Grellmann, K.-H.; Weller, A. Chem. Phys. Lett. 1980, 241

^{(21) &}lt;sup>1</sup>DCA* quenching by product CHD is negligible due to the low adduct concentrations and low conversions employed.³

Table VIII. Quantum Yields and Related Parameters for Adiabatic ¹DCA* Formation from the Adducts

solvent	adduct	ϕ_1 DCA* a	$\tau_{\mathbf{a}},\mathrm{ns}^{b}$	$k_{-e}\tau_{e}^{c}$	$k_{\mathbf{e}\mathbf{a}}\tau_{\mathbf{a}}^{d}$
CH, CN	1,2-A	0.015	$5.8_6 \pm 0.15$	0.92	0.017
$C_{\ell}H_{\ell}$	1,2-A	0.011	$7.7^{\circ}_{0} \pm 0.26$	0.98	0.011
C,H,N	1,2-A	0.0070	$5.0^{\circ}_{6} \pm 0.06$	0.76	0.009
CH, CN	9,10-A	0.0041	· ·	0.92	0.004
$C_6 H_{12}$	9,10-A	0.0068		0.98^{e}	0.006

 a Values of $\phi_{\rm fDCA}$ for 1,2-A corrected by using $\xi_{\rm a}$ from Table VII. b Based on O₂ quenching of ¹1,2-A* fluorescence; see the text. c From ref 3. d Using eq 17. e Assumed identical with the C₆ H₆ value.

 $pk_e\tau_m$ values, where p is the fraction of exciplexes which do not regenerate $^1\mathrm{DCA}^{*,3}$ Lower limits for p, estimated from the deviation of pk_e from values of diffusion-controlled rate constants, were used to calculate $k_{-e}\tau_e=1-p$ and, by using eq 17, $k_{\mathrm{ea}}\tau_a$ values shown in Table VIII. Also shown in Table VIII are τ_a values for 1,2-A obtained from I_0/I_a and I_0/I_{ox} data by assuming $k_{\mathrm{qa}}=k_{\mathrm{qm}}$ in eq 11 and 12. It follows that $k_{\mathrm{ea}}=2.90\times10^6$, 1.4 \times 106, and 1.8 \times 106 s⁻¹ in CH₃CN, C₆H₆, and C₅H₅N, respectively. Of mechanistic significance is the observation that the effect of changing the solvent from benzene to pyridine on $\phi_{^1\mathrm{DCA}^*}$ appears to be dictated primarily by the change in $k_{-e}\tau_e$, consistent with sequential formation of $^1(\mathrm{DCA}\cdot\mathrm{CHD})^*$ and $^1\mathrm{DCA}^*$ from 11 ,2-A*. Increasing the polarity of the solvent to acetonitrile causes a modest increase in k_{ea} . A similar, but much more pronounced, effect may be operating in the case of the $^1\mathrm{MA}$ dimer, for which $^1\mathrm{MA}^*_2$ fluorescence is observed in hydrocarbon solvents $^2\mathrm{MB}$ 0 but is absent in acetonitrile.

Photochemical Observations. Irradiation of a 1:1 mixture of 1,2-A and 1,4-A in cyclohexane at 366 nm has been reported to selectively destroy 1,2-A (yield DCA: $72 \pm 2\%$).³ Product distributions (Table V) and quantum yields (Table VI) obtained in this work show that DCA and CHD are obtained in 1:1 relative yield from 1,2-A (313 nm) or 9,10-A (254 nm) but with markedly different efficiencies. At low conversions, cleavage of 1,2-A to DCA and CHD proceeds quantitatively for 313-nm excitation and nearly quantitatively (\sim 90%) for excitation at 254 nm. In the case of 9,10-A, a complex mixture of products is obtained and cleavage to DCA and CHD accounts for only 8-11% of the consumed adduct. Photochemical conversion of 1,2-A or 9,10-A to the other photoadducts could not be detected by NMR analysis even at high conversions, but UV absorption spectra reveal the possible formation of 1,2-A as a minor photoproduct, $\phi_{1,2-A} \le$ 0.037, from 9,10-A.

Comparison of $\phi_{^{1}DCA^{*}}$ and ϕ_{DCA} values (Tables II and VIII) shows that for both adducts adiabatic cleavage to $^{1}DCA^{*}$ accounts for only 2-6% of the total ^{1}DCA formed; clearly, diabatic reaction channels to ^{1}DCA predominate. If the exciplex is an essential intermediate in the paths to $^{1}DCA^{*}$ as shown in the mechanism and Figure 1, and to the photoadducts generally, 3 then some adduct photoisomerization via the exciplex would be expected

$$\phi_{\mathrm{Ad_i}} = p \ \phi_{\mathrm{Ad_i}}^{\mathrm{lim}} k_{\mathrm{ea}} \tau_{\mathrm{a}} \tag{18}$$

where $\phi_{\rm Ad}^{\rm int}$ is the limiting appearance quantum yield of adduct i at infinite diene concentration in the DCA/CHD system. Experimentally determined p, 3 $\phi_{\rm Ad}^{\rm im^3}$, and $k_{\rm ea} \tau_{\rm a}$ values (Table VIII) give $\phi_{\rm Ad}$, values 3–5 orders of magnitude smaller than observed adduct loss quantum yields. It follows that adiabatic formation of the singlet exciplex should result in yields of its characteristic products too low to allow their detection by NMR analysis, consistent with our observations. On the other hand, if the UV absorption bands at 306 and 318 nm, observed upon irradiation of 9,10-A in degassed cyclohexane, correspond to 1,2-A, the resulting $\phi_{1,2-A}$ value of 0.037 \pm 0.002 is substantially larger than predicted by eq 18, $\phi_{1,2-A} \simeq 2 \times 10^{-5}$. Though 1,2-A would still

be a minor photoproduct from 9,10-A, the possible involvement of a diabatic reaction channel from ¹9,10-A* to 1,2-A which bypasses the singlet exciplex is intriguing. In any case, it seems safe to conclude that interconversion between pericyclic minima on the **D** surface does not play a significant role in the deactivation of ¹1,2-A* and ¹9,10-A*.

The effect of oxygen on ϕ_{-A} and ϕ_{DCA} is consistent with the mechanism eq 1–13. For 1,2-A, ϕ_{-A}^o/ϕ_{-A}^a and $\phi_{DCA}^o/\phi_{DCA}^a$ ratios (Table VI) are in close agreement with I_o/I_a ratios obtained for ¹1,2-A* fluorescence (Tables II and III). Similarly, for 9,10-A, for which $I_o/I_a=1.00$ was inferred from the effect of air on the fluorescence of adiabatically generated ¹DCA*, $\phi_{DCA}^o/\phi_{DCA}^a=1.00$. Since oxygen quenching should provide an efficient decay channel for adduct triplets, the agreement between ϕ_{-A}^o/ϕ_{-A}^a , $\phi_{DCA}^o/\phi_{DCA}^a$ and I_o/I_a ratios suggests either that they are not involved in the photochemistry of the adducts or that they are unusually short-lived and not measurably quenched by O_2 in air-saturated solutions.

Experimental Section

Materials. All materials used in this work have been described previously.³

Fluorescence Measurements. A Perkin-Elmer Hitachi MPF-2A spectrophotometer was employed in the ratio recording mode as previously described.3 Slit band-passes were typically set at 4-7 and 5-10 nm for excitation and emission, respectively. Streams of argon, air, or oxygen were gently bubbled through the solutions for 120 s (timed) directly in $1 \times 1 \times 5$ cm quartz cells, which were then Teflon stoppered. No change in the oxygen content of the solutions was noted while measurements were being made. Degassed solutions and argon-bubbled solutions of DCA gave identical fluorescence intensities. Relative fluorescence quantum yields were determined at 25.0 ± 0.1 °C by using fresh solutions with carefully matched absorbances (5.0-cm UV cells were employed for absorbance matching). Relative emission intensities, corrected for nonlinearity of instrumental response, were plotted vs. cm⁻¹ and relative areas determined by cutting and weighing Xerox copies of the spectra.¹² A few fluorescence excitation spectra were measured at room temperature (\sim 23 °C) by using a Varian SF-330 spectrofluorometer (courtesy Dr. M. Kasha) which has a flatter response than the Hitachi spectrophotometer over the region of interest.

Irradiation Procedures. A merry-go-round apparatus¹⁴ was employed for quantum yield measurements as previously described.³ Quartz ampules (13-mm o.d.) and NMR tubes (3-mm o.d.) were used for 254-nm irradiations, and Pyrex ampules and NMR tubes were used for 313-nm irradiations.

Lamps and Light Filters. A Nester-Faust low-pressure mercury lamp was used for 254-nm excitation, and Hanovia medium-pressure mercury lamps (200- and 550-W, Ace Glass, Inc.) were used for 313-nm excitation. Filter system 1 consisted of a 1.0 cm pathlength UV cell filled with aqueous potassium iodide, 0.02 g/mL, and a 1.0-cm UV cell filled with 1 atm of chlorine gas. The two cells were arranged in series so that light from the Hitachi fluorometer excitation beam entered the KI solution first. This filter system removes all light at 300-370 nm and <265 nm and was used to minimize inadvertent DCA excitation when exciting 9,10-A solutions with the Hitachi monochromator set at 270 nm. Filter system 2 consisted of a 500-mL aqueous solution of K₂Cr₂O₇ (0.734 g) and K₂CO₃ (15.0 g) transmitting a narrow band of light at 313 nm and visible light at $\lambda > 450$ nm and Corning CS 7-54 glass filter plates which remove visible light at $\lambda > 430$ nm. The solution was contained in a Pyrex Hanovia reactor-probe assembly, 0.2-0.3 cm pathlength, equipped with a 550-W Hanovia lamp. Filter system 3 consisted of 2 atm of Cl₂ sealed in the cooling jacket of a quartz Hanovia probe, 1-cm path length, and a cylindrical Vycor sleeve. Filter system 4 was as in filter system 3 except 1 atm of Cl₂ gas was employed and the Vycor sleeve was omitted. Filter system 5 consisted of Corning CS 7-54 glass filter plates in combination with filter system 3. Filter system 6 was as in filter system 2, except 1.0 g of K₂Cr₂O₇, 15.4 g of K₂CO₃, and a 200-W Hanovia lamp were employed.

Analytical Procedures. NMR, GLC, and UV-visible analytical procedures have been described.^{3,17}

Registry No. 1,2-A, 83929-15-1; 9,10-A, 83929-13-9; 9,10-dichloroanthracene, 605-48-1.