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## UNEXPECTED DEHALOGENATION OF 3-BROMOPYRAZOLO[3,4-d]PYRIMIDINE NUCLEOSIDES DURING NUCLEOBASE-ANION GLYCOSYLATION

Frank Seela\*, Matthias Zulauf, and Georg Becher

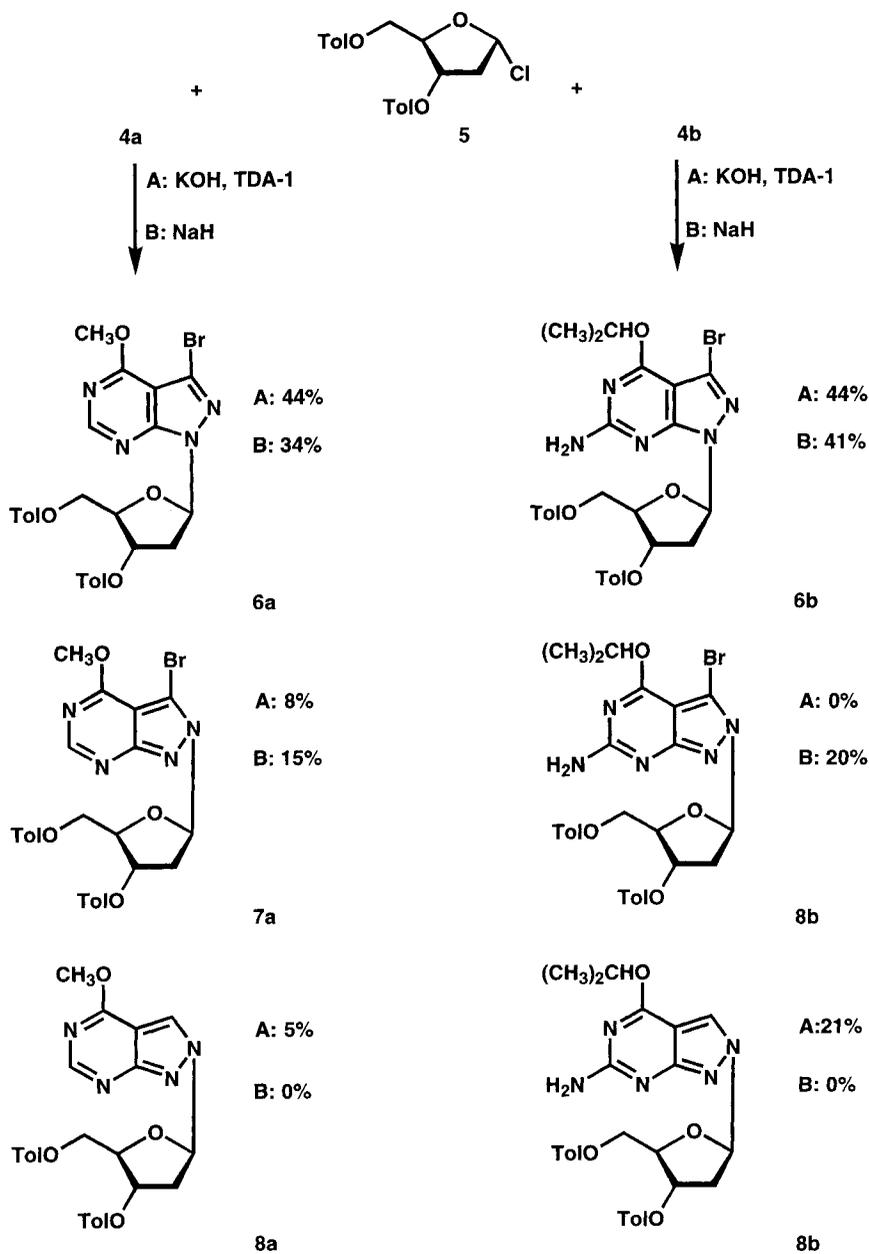
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**ABSTRACT:** The anion-glycosylation (KOH, MeCN, TDA-1) of 3-bromopyrazolo[3,4-d]pyrimidines **4a** and **4b** with 2-deoxy-3,5-di-O-(p-toluoyl)- $\alpha$ -D-erythro-pentofuranosyl chloride (**5**) furnishes the regioisomeric N<sup>1</sup>- $\beta$ -D-2'-deoxyribonucleosides **6a** and **6b** together with the dehalogenated N<sup>2</sup>-regioisomers **8a** and **8b**, stereoselectively. The dehalogenation takes place after the glycosylation and results from the sensitivity of the N-2 nucleosides toward aqueous base. An addition/elimination mechanism is suggested for the dehalogenation reaction.

Certain pyrazolo[3,4-d]pyrimidines exhibit strong therapeutic activity against various diseases.<sup>1-2</sup> Allopurinol is the drug of choice for the treatment of gout.<sup>3</sup> Also pyrazolo[3,4-d]pyrimidine ribonucleosides including those with 3-bromo or 3-iodo substituents develop biological action in particular antiparasitic activity and inhibitory action against adenosine kinases.<sup>4-6</sup> The 2'-deoxyribonucleosides, such as **1** or **2** have been incorporated in oligonucleotides chemically and enzymatically.<sup>7,8</sup>

Recently, the stabilizing effect of pyrrolo[2,3-d]pyrimidines on the oligonucleotide duplex structure was reported.<sup>9</sup> In these cases the 5-membered ring of the base carries a halogen substituent. As the same favorable properties, e.g. stabilization of an oligonucleotide duplex, are expected for 3-halogenated pyrazolo[3,4-d]pyrimidines<sup>2,10,11</sup> the synthesis of corresponding 2'-deoxyribonucleosides was considered. As central intermediates the alkoxy derivatives **6a,b** were chosen. These intermediates should be amenable to conversion to nucleosides with various substituents at carbon-4.





yield) which did not contain bromine. According to analytical and spectroscopic data structure **8a** was established and the compound was found to be identical to a protected nucleoside, which has been synthesized before.<sup>12</sup> The same glycosylation reaction was performed on compound **4b** furnishing only two reaction products. They were also separated chromatographically. The compound of the fast migrating zone was characterized as the bromo nucleoside **6b** (44%); the second zone was identified as the dehalogenated N<sup>2</sup>-isomer **8b**<sup>13</sup> (21%).

In all cases, the position of glycosylation was deduced from the <sup>13</sup>C-NMR-spectra.<sup>12,13</sup> According to the Table (next page) an upfield shift of carbon-3 is observed for the N-2 nucleosides in comparison to the N<sup>1</sup>-isomers which show similar chemical shifts for the base moiety as the free nucleobase. The assignment of the β-D configuration was confirmed by the <sup>1</sup>H-NMR spectra using the characteristic shift differences of the H-C(4') and CH<sub>2</sub>(5') of the toluoylated compounds.<sup>16</sup>

From the experiments described above, it is apparent that the loss of the 3-bromo substituent occurs only in the case of the N-2 deoxyribofuranosides whereas the bromo substituent is stable in the N-1 compounds **6a** and **6b**. The dehalogenation observed during the anion-glycosylation of **4a** and **4b** in the presence of powdered KOH - containing 15% of water - is not observed under anhydrous condition when NaH is used instead of powdered KOH. In this case the desired 3-bromo isomers **6a** (34%) and **7a** (15%) were formed. Correspondingly, the isomers **6b** (41%) and **7b** (20%) were obtained from **4b**. Due to these observations the dehalogenated compounds **8a** and **8b** have to be formed after glycosylation. This was confirmed in a separate experiment by treatment of the N-2 bromo compounds **7a** or **7b** in a mixture of MeCN / powdered KOH but without the halogenose **5**<sup>14</sup>. Indeed, the bromo nucleosides **7a** and **7b** are converted into the dehalogenated compounds **8a** and **8b**.

According to these findings, various mechanisms can be discussed for the dehalogenation reaction. Nevertheless, the formation of the dehalogenated N-2 isomers **8a** and **8b** being reduced species of the bromo compounds **7a** and **7b** has to be accompanied by the oxidation of another molecule being present in the reaction mixture. A possible route is given below. In this Scheme, the p-quinoid structure of the N-2 isomer undergoes a nucleophilic addition/elimination reaction. In the first step water is added by an initial attack of hydroxyl ions on the most electrophilic center (C-3). This may lead to an intermediate which undergoes a base-catalyzed elimination in a second step thus restoring the p-quinoid system. According to these findings it is apparent that

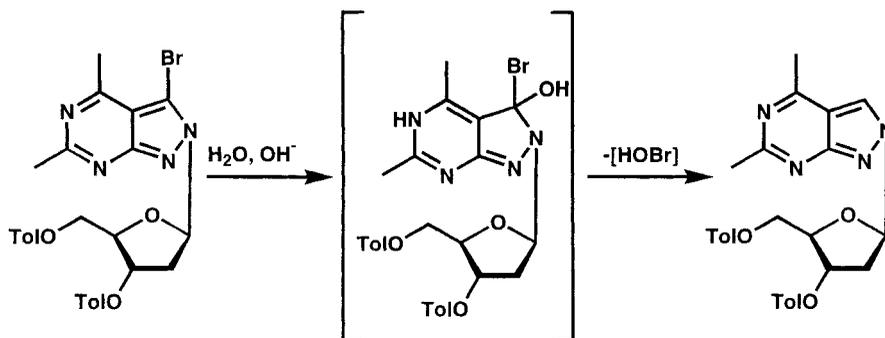
**TABLE.**  $^{13}\text{C}$ -NMR Chemical Shifts of Pyrazolo[3,4-d]pyrimidines 2'-Deoxyribofuranosides .

Comp.	C-3	C-3a	C-4	C-6	C-7a	Me	i-Prop
<b>3a</b>	131.1	101.2	163.2	154.5	154.1	53.6	-
<b>3b</b>	131.5	95.8	162.8	162.0	158.9	-	21.7, 68.2
<b>4a</b>	118.1	101.5	163.1	156.1	156.5	54.5	-
<b>4b</b>	118.4	95.5	162.5	162.5	159.5	-	21.8, 68.9
<b>6a</b>	119.0	102.6	163.0	156.0	155.4	54.1	-
<b>6b</b>	120.0	96.3	162.6	162.5	158.7	-	21.6, 64.0
<b>7a</b>	108.9	103.7	<sup>b</sup>	156.1	159.7	54.4	-
<b>7b</b>	108.2	99.2	163.7	161.7	162.4	-	21.1, 69.1
<b>8a</b>	124.8	102.4	<sup>c</sup>	155.5	161.0	54.0	-
<b>8b</b>	12.5	98.3	163.4	161.6	164.2	-	21.1, 68.6
	C-1'	C-2'	C-3'	C-4'	C-5'		
<b>6a</b>	84.5	35.0	74.2	81.4	63.4		
<b>6b</b>	83.4	34.8	74.9	81.3	64.0		
<b>7a</b>	88.2	36.2	74.1	82.3	63.4		
<b>7b</b>	87.1	36.0	74.4	81.8	63.8		
<b>8a</b>	90.7	37.1	74.5	82.4	64.0		
<b>8b</b>	89.3	36.6	74.8	81.8	64.1		

<sup>a</sup> Measured in ( $\text{D}_6$ )DMSO at 23°. <sup>b</sup> Not detectable.

<sup>c</sup> Superimposed by CO.

3-halogenated pyrazolo[3,4-d]pyrimidine N-1 nucleosides can be incorporated into oligonucleotides chemically while the alkaline deprotection conditions employed during the oligonucleotide synthesis cycle do dehalogenate the N-2 nucleoside residues.



## EXPERIMENTAL

**General.** Elemental analyses were performed by Mikroanalytisches Labor Beller (Göttingen, Germany). NMR-Spectra were measured on a AC 250 and AMX 500 spectrometer (Bruker, Germany). Chemical shifts are in ppm relative to TMS as internal standard. UV-spectra were recorded on a U 3200 spectrometer (Hitachi, Japan). Thin-layer chromatography (TLC) was performed on silica gel 60 F<sub>254</sub> plates (Merck, Germany). Column chromatography was performed on silica gel 60 (Merck, Germany).

### 3-Bromo-4-methoxy-1H-pyrazolo[3,4-d]pyrimidine (**4a**).

To a suspension of **3a**<sup>12</sup> (600 mg, 4 mmol) in 1,2-dichloroethane (50 ml) were added N-bromosuccinimide (1.1 g, 6.2 mmol) and azoisobutyronitrile (AIBN, 50 mg). After heating under reflux for 30 min, the solvent was evaporated and the residue applied to a silica gel column. The elution was performed with methylene chloride/methanol (0% → 5% methanol). Crystallization from CH<sub>2</sub>Cl<sub>2</sub>/MeOH afforded colorless crystals (690 mg, 75%) of **4a**: mp 194-196°C (dec.); TLC (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 95:5): R<sub>f</sub> 0.4; UV (MeOH): λ<sub>max</sub> 246, 266 nm (ε = 5200, 3500) <sup>1</sup>H-NMR (250 MHz, (D<sub>6</sub>) DMSO): δ 4.09 (s, 3H, OCH<sub>3</sub>), 8.55 (s, 1H, H-(C6)), 14.26 (s, 1H, NH); Anal. calcd. for C<sub>6</sub>H<sub>5</sub>BrN<sub>4</sub>O (229.0): C 31.47, H 2.20, N 24.46. Found: C 31.26, H 2.28, N 24.49.

### 6-Amino-3-bromo-4-isopropoxy-1H-pyrazolo[3,4-d]pyrimidine (**4b**).

Compound **4b** was prepared as described for **4a** but using the following amounts: **3b**<sup>13</sup>

(1.0 g, 5.2 mmol); NBS (1.0 g, 5.6 mmol; 1.5 h). The solution was evaporated to dryness, the residue dissolved in MeOH, decolorized with charcoal and filtered. Upon addition of ice-water a colorless amorphous solid precipitated (1.1 g, 78%). An analytical sample was crystallized from i-PrOH: mp 221°C (dec.); TLC (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 9:1): R<sub>f</sub> 0.4; UV (MeOH): λ<sub>max</sub> 248, 276 nm (ε = 6000, 7100); <sup>1</sup>H-NMR (250 MHz, (D<sub>6</sub>) DMSO): δ 1.32 (d, J = 6.4 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>), 5.41 (m, 1H, OCH), 6.77 (s, 2H, NH<sub>2</sub>), 13.0 (s, 1H, NH); Anal. calcd. for C<sub>8</sub>H<sub>10</sub>BrN<sub>5</sub>O (272.1): C 35.31, H 3.70, N 25.74. Found: C 35.58, H 3.81, N 25.54.

#### **Nucleobase Anion-Glycosylation of Compounds 4a and 4b with the Halogenose 5<sup>14</sup> in the Presence of KOH/TDA-1 (Method A):**

To a suspension of 4a (1.0 g, 4.4 mmol) in MeCN (60 ml), KOH (85%, 470 mg, 7.1 mmol) and TDA-1 (= tris[2-(2-methoxyethoxy)ethyl]amine; 75 μl) was added. After stirring at r.t. for 10 min 5<sup>14</sup> (2.1 g, 5.4 mmol) was added and stirring continued for another 20 min. Insoluble material was filtered off and after evaporation the residue was subjected to flash chromatography (FC).

#### **3-Bromo-1-[2-deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-methoxy-1H-pyrazolo[3,4-d]pyrimidine (6a).**

From the fast migrating main zone (petroleum ether/ethyl acetate 2:1) 6a was isolated. Crystallization from petroleum ether/ethyl acetate yielded colorless needles (1.13 g, 44%): mp 177-178°C (dec.); TLC (petroleum ether/ethyl acetate, 1:1): R<sub>f</sub> 0.6; UV (MeOH): λ<sub>max</sub> 240, 270 nm (ε = 34300, 7500); <sup>1</sup>H-NMR (500 MHz, (D<sub>6</sub>) DMSO): δ 2.41, 2.51 (2s, 6H, 2CH<sub>3</sub>), 2.81 (m, 1H, H<sub>α</sub>-(C2')), 3.27 (m, 1H, H<sub>β</sub>-(C2')), 4.14 (s, 3H, OCH<sub>3</sub>), 4.48 (m, 2H, H-(C5')), 4.58 (m, 1H, H-(C4')), 5.81 (m, 1H, H-(C3')), 6.83 (t, J = 6.2 Hz, 1H, H-(C1')), 7.35, 7.92 (4d, J = 7.8, 7.9 Hz, 8H, 2C<sub>6</sub>H<sub>4</sub>), 8.67 (s, 1H, H-(C6)). Anal. calcd. for C<sub>27</sub>H<sub>25</sub>BrN<sub>4</sub>O<sub>6</sub> (581.4): C 55.78, H 4.33, N 9.64. Found: C 55.98, H 4.43, N 9.53.

#### **3-Bromo-2-[2-deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-methoxy-1H-pyrazolo[3,4-d]pyrimidine (7a).**

From the second zone (petroleum ether/ethyl acetate 1:1) a colorless foam was obtained (205 mg, 8%): TLC (petroleum ether/ethyl acetate, 1:1): R<sub>f</sub> 0.4; UV (MeOH): λ<sub>max</sub> 240 nm (ε = 33900); <sup>1</sup>H-NMR (500 MHz, (D<sub>6</sub>) DMSO): δ 2.30, 2.35 (2s, 6H, 2CH<sub>3</sub>),

2.82 (m, 1H, H<sub>α</sub>-(C2')), 3.39 (m, 1H, H<sub>β</sub>-(C2')), 4.06 (s, 3H, OCH<sub>3</sub>), 4.33, 4.48 (2m, 2H, H-(C5')), 4.59 (m, 1H, H-(C4')), 5.90 (m, 1H, H-(C3')), 6.68 (dd, J = 6.3, 6.5 Hz, 1H, H-(C1')), 7.24, 7.80 (4d, J = 7.9, 8.0 Hz, 8H, 2C<sub>6</sub>H<sub>4</sub>), 8.56 (s, 1H, H-(C6)). Anal. calcd. for C<sub>27</sub>H<sub>25</sub>BrN<sub>4</sub>O<sub>6</sub> (581.4): C 55.78, H 4.33, N 9.64. Found: C 55.79, H 4.43, N 9.65.

**2-[2-Deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-methoxy-1H-pyrazolo[3,4-d]pyrimidine (8a).**

Evaporation of the slow migrating zone (petroleum ether/ethyl acetate 1:2) yielded a colorless foam of **8a**<sup>12</sup> (110 mg, 5%) upon evaporation. UV (MeOH): λ<sub>max</sub> 242 nm (ε = 35600) (Lit.<sup>12</sup> 242 nm (ε = 36000)). All other data were identical with the literature.<sup>12</sup>

**6-Amino-3-bromo-1-[2-deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-isopropoxy-1H-pyrazolo[3,4-d]pyrimidine (6b).**

As described for **4a** compound **4b** (1.5 g, 5.5 mmol) was treated in an analogous manner with KOH (85%, 1.46 g, 22 mmol), MeCN (150 ml), TDA-1 (20 μl) and **5**<sup>14</sup> (2.6 g, 6.7 mmol). Column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/acetone 98:2) gave a colorless foam (1.5 g, 44%): TLC (CH<sub>2</sub>Cl<sub>2</sub>/acetone 98:2): R<sub>f</sub> 0.5; UV (MeOH): λ<sub>max</sub> 231, 274 nm (ε = 35400, 7700); <sup>1</sup>H-NMR (500 MHz, (D<sub>6</sub>) DMSO): δ 1.33 (d, J = 6.2 Hz, 6H, CH(CH<sub>3</sub>)<sub>2</sub>), 2.36, 2.38 (2s, 2Ar-CH<sub>3</sub>), 2.67 (m, 1H, H<sub>α</sub>-(C2')), 3.15 (m, 1H, H<sub>β</sub>-(C2')), 4.50 (m, 3H, H-(C5', C4')), 5.43 (m, 1H, OCH), 5.73 (m, 1H, H-(C3')), 6.55 (t, J = 5.9 Hz, 1H, H-(C1')), 7.05 (s, 2H, NH<sub>2</sub>), 7.28-7.92 (4d, J = 8.1 Hz, 8H, 2C<sub>6</sub>H<sub>4</sub>). Anal. calcd. for C<sub>29</sub>H<sub>30</sub>BrN<sub>5</sub>O<sub>6</sub> (624.5): C 55.78, H 4.84, N 11.21. Found: C 56.11, H 4.84, N 10.93.

**6-Amino-2-[2-deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-isopropoxy-1H-pyrazolo[3,4-d]pyrimidine (8b).**

From the second zone compound **8b**<sup>13</sup> (630 mg, 21%) was obtained. UV (MeOH): λ<sub>max</sub> 224 nm (ε = 42200) (Lit.<sup>13</sup> 224 nm (ε = 42500)). All other data were identical with the literature.<sup>13</sup>

**Nucleobase Anion-Glycosylation of Compounds 4a and 4b with the Halogenose 5<sup>14</sup> in the Presence of NaH (Method B):**

Glycosylation of **4a**: To a suspension of **4a** (1.0 g, 4.4 mmol) in MeCN (60 ml) was added NaH (97%, 163 mg, 6.6 mmol). After stirring at r.t. for 10 min **5**<sup>14</sup> (2.1 g, 5.4 mmol) was added and stirring continued for 30 min. The mixture was filtered and the filtrate

was evaporated. The further work-up was performed as described under Method A. The fast migrating zone furnished compound **6a** (870 mg, 34%). From the second zone compound **7a** was isolated (384 mg, 15%).

The glycosylation of **4b** was performed as described for **4a** using the following amounts: **4b** (1.5 g, 5.5 mmol), NaH (97%, 150 mg, 6.1 mmol), MeCN (100 ml) and **5**<sup>14</sup> (2.6 g, 6.6 mmol). The work-up was the same as described under Method A.

**6-Amino-3-bromo-2-[2-deoxy-3,5-di-O-(p-toluoyl)-β-D-erythro-pentofuranosyl]-4-isopropoxy-1H-pyrazolo[3,4-d]pyrimidine (7b).**

Flash-chromatography of the reaction product was performed with CH<sub>2</sub>Cl<sub>2</sub>/acetone 98:2 yielding two main zones. From the fast migrating zone **6b** was obtained (1.4 g, 41 %). The second zone yielded **7b** as a colorless foam (680 mg, 20%): TLC (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 95:5): R<sub>f</sub> 0.5; UV (MeOH): λ<sub>max</sub> 268, 303 nm (ε = 11600, 5500); <sup>1</sup>H-NMR (500 MHz, (D<sub>6</sub>) DMSO): δ 1.33 (d, J = 6.7 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>), 2.35, 2.38 (2s, 6H, Ar-CH<sub>3</sub>), 2.67 (m, 1H, H<sub>α</sub>-C(2')), 3.15 (m, 1H, H<sub>β</sub>-C(2')), 4.44 (m, 1H, H-C(4')), 4.51 (m, 2H, H-C(5')), 5.43 (m, 1H, OCH), 5.91 (m, 1H, H-C(3')), 6.52 (t, J = 6.2 Hz, 1H, H-C(1')), 6.64 (s, 2H, NH<sub>2</sub>), 7.28-7.92 (4d, J = 8.1 Hz, 8H, 2C<sub>6</sub>H<sub>4</sub>). Anal. calcd. for C<sub>29</sub>H<sub>30</sub>BrN<sub>5</sub>O<sub>6</sub> (624.5): C 55.78, H 4.84, N 11.21. Found: C 55.89, H 4.80, N 11.24.

**Debromination of Compounds 7a and 7b in MeCN/Powdered KOH:**

To a solution of compound **7a** (200 mg, 0.34 mmol) in MeCN (30 ml) powdered KOH (85%, 240 mg, 3.6 mmol) was added. The suspension was stirred for 30 min at r.t., filtered and the filtrate was evaporated. The residue was subjected to FC (petroleum ether/ethyl acetate 1:1) yielding compound **8a** as a colorless foam (60 mg, 35%). The debromination of compound **7b** was performed analogously furnishing compound **8b**.

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**REFERENCES**

1. Nelson, D. J.; LaFon, S. W.; Tuttle J. V.; Miller, W. H.; Miller, R. L.; Krenitsky, T. A.; Elion, G. B.; Berens, R. L.; Marr, J. J. *J. Biol. Chem.* **1979**, *254*, 11544.

2. Taylor, E. C.; Patel, H. H. *Tetrahedron* **1992**, *48*, 8089.
3. Elion, G. B.; Callahan, S. W.; Nathan, H.; Bieber, S.; Rundles, R. W.; Hitchings, G. H. *Biochem. Pharmacol.* **1963**, *12*, 85.
4. Cottam, H. B.; Petrie, C. R.; McKernan, P. A.; Goebel, R. J.; Dalley, N. K.; Davidson, R. B.; Robins, R. K.; Revankar, G. R. *J. Med. Chem.* **1984**, *27*, 1119.
5. Petrie III, C. R.; Cottam, H. B.; McKernan, P. A.; Robins, R. K.; Revankar, G. R. *J. Med. Chem.* **1985**, *28*, 1010.
6. Cottam, H. B.; Wasson, D. B.; Shih, H. C.; Raychaudhuri, A.; Di Pasquale, G.; Carson, D. A. *J. Med. Chem.* **1993**, *36*, 3424.
7. Seela, F.; Kaiser, K. *Helv. Chim. Acta* **1988**, *71*, 1813.
8. Mersmann, K.; Seela, F. **1996**, unpublished data.
9. Seela, F.; Thomas, H. *Helv. Chim. Acta* **1995**, *78*, 94.
10. Bontems, R. J.; Anderson, J. D.; Smee, D. F.; Jin, A.; Alaghamandan, H. A.; Sharma, B. S.; Jolley, W. B.; Robins, R. K.; Cottam, H. B. *J. Med. Chem.* **1990**, *33*, 2174.
11. Seela, F.; Ramzaeva, N.; Becher, G. *Collect. Czech. Chem. Commun.* **1996**, *61*, 258.
12. Seela, F.; Steker, H. *Helv. Chim. Acta* **1985**, *68*, 563.
13. Dissertation H. Steker, Universität Osnabrück **1988**.
14. Hoffer, M. *Chem. Ber.* **1960**, *93*, 2777.
15. Seela, F.; Winkeler, H.-D. *J. Org. Chem.* **1983**, *48*, 3119.
16. Seela, F.; Steker, H. *Helv. Chim. Acta* **1986**, *69*, 1602.

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