A Concise and Diastereoselective Synthesis of Piperidine and Indolizidine Alkaloids via Aza-Prins Cyclization

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Abstract: The synthesis of 2-substituted and 2,4-disubstituted piperidine alkaloids such as (\pm) -coniine, (\pm) -hydroxypipecolic acid, (\pm) -pipecolic acid, (\pm) -coniceine, and (\pm) -4-hydroxy-2-hydroxy-methyl piperidine have been accomplished in a highly diastereoselective manner by employing aza-Prins cyclization as a key step to construct the piperidine core of these alkaloids.

Key words: aza-Prins cyclization, diastereoselective, piperidine, alkaloids, natural products

Piperidine and indolizidine subunits are common structural motifs in naturally occurring alkaloids and constitute the basic skeleton of several drug candidates for the treatment of a wide range of diseases.^{1,2} As a result, various methods have been developed for the stereo- and enantioselective synthesis of piperidine derivatives.³ Of these, the coupling of homoallylic amines with aldehydes - the socalled aza-Prins cyclization - is a versatile and direct method for the synthesis of trans-2,4-disubstituted piperidines.⁴ As part of our ongoing research project on the development of new chemical entities, recently, we have reported the synthesis of 4-iodo- and 4-hydroxypiperidines by means of aza-Prins cyclization.⁵ Herein, we report an extension of this work to the total synthesis of several piperidine and indolizidine alkaloids. The 4-iodoand 4-hydroxypiperidines could be used as key intermediates in the synthesis of five representative alkaloids. To the best of our knowledge, this is the first report on the synthesis of (\pm)-coniine (**A**),⁶ (\pm)-4-hydroxypipecolic acid (**B**),⁷ (\pm)-pipecolic acid (**C**),⁸ (\pm)-coniceine (**D**),⁹ and (\pm)-4-hydroxy-2-hydroxymethylpiperidine (**E**)¹⁰ using aza-Prins cyclization (Scheme 1).

Coniine $(\mathbf{A})^{11}$ is a hemlock alkaloid that was used in ancient Greece to execute criminals. Initially, we prepared the piperidine key intermediate 4-iodopiperidine 1 in 92% yield using aza-Prins cyclization between *N*-tosylhomo-allylic amine and *n*-butyraldehyde,^{5a} which was transformed into coniine by a known reaction sequence.

Piperidine intermediate **2** was achieved by radical reduction of **1** in 95% yield by refluxing with *n*-tributyltinhydride in toluene.¹² The exposure of **2** to a dark-green sodium naphthalenide solution at -78 °C in anhydrous tetrahydrofuran resulted in deprotection of the *N*-tosyl group to give coniine (**A**) in 90% yield (Scheme 2).¹³ The data of product **A** was identical to that reported in the literature.^{6d}

The 4-hydroxypipecolic acid (**B**)¹⁴ motif is the most prevalent subunit of this type in bioactive molecules such as SS20846A¹⁵ and palinavir,¹⁶ which is a highly potent HIV protease inhibitor. Both pipecolic acid and 4-hydroxypipecolic acid, being unnatural amino acids, are useful in peptide synthesis.



Scheme 1 Retrosynthetic analysis for the synthesis of piperidine and indolizidine alkaloids

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Scheme 2 Synthesis of (\pm)-coniine (A). *Reagents and conditions*: (a) GaI₃, I₂, anhydrous CH₂Cl₂, 0 °C to r.t., 7 h, 92%; (b) Bu₃SnH, AIBN, toluene, 4 h, reflux, 95%; (c) Na/naphthalene, anhydrous THF, -78 °C, 6 h, 90%.

The key intermediate, 4-hydroxypiperidine **3**, was synthesized in 85% yield by means of an aza-Prins cyclization between *N*-tosyl homoallylic amine and cinnamaldehyde.^{5b} Oxidative cleavage of olefin **3** with RuCl₃·2H₂O gave the corresponding carboxylic acid **4** in 68% yield without affecting the hydroxy group.^{8a,17} Treatment with sodium/naphthalene at -78 °C in anhydrous tetrahydrofuran deprotected the tosyl group to give the target product **B** in 80% yield (Scheme 3).¹³ The data of product **B** was identical with respect to its ¹H NMR spectrum to that reported in the literature.^{7a}



Scheme 3 Synthesis of (\pm)-4-hydroxypipecolic acid (**B**). *Reagents and conditions*: (a) PMA (10 mol%), anhydrous CH₂Cl₂, reflux, 10 h, 85%; (b) RuCl₃·2H₂O, NaIO₄, MeCN–CCl₄–H₂O (1:1:10), r.t., 8 h, 68%; (c) Na/naphthalene, anhydrous THF, -78 °C, 6 h, 80%.

Pipecolic acid (**C**)¹⁸ is a naturally occurring non-proteinogenic α -amino acid that is frequently found in plants, fungi, and human physiological fluids. It is a key structural unit in many biologically active natural and synthetic molecules, for instance anticonvulsant pipradol,¹⁹ immunosuppressants FK 506²⁰ and rapamycin,²¹ amyloglucosidase inhibitor lentiginosine,²² enzyme inhibitors, and *N*-methyl-D-aspartic acid (NMDA)²³ antagonists.

The key precursor, 4-iodopiperidine **5** was also prepared by means of an aza-Prins cyclization.^{5a} Removal of iodide with *n*-tributyltinhydride in the presence of AIBN gave the (*E*)-2-styryl-*N*-tosylpiperidine **6** in 93% yield.¹² Oxidative cleavage of the olefin was achieved using RuCl₃·2H₂O and NaIO₄ to afford the corresponding

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carboxylic acid **7** in 70% yield.^{8a,17} Deprotection of the *N*-tosyl group was carried out using sodium/naphthalene at -78 °C in anhydrous tetrahydrofuran to afford (±)-pipe-colic acid (**C**) in 89% yield (Scheme 4).¹³ The data for product **C** was in agreement with the those reported in the literature.^{8a}



Scheme 4 Synthesis of (±)-pipecolic acid (C). *Reagents and conditions*: (a) GaI₃, I₂, anhydrous CH₂Cl₂, 0 °C to r.t., 7 h, 90%; (b) Bu₃SnH, AIBN, toluene, 3 h, reflux, 93%; (c) RuCl₃·2H₂O, NaIO₄, MeCN–CCl₄–H₂O (1:1:10), r.t., 8 h, 70%; (d) Na/naphthalene, anhydrous THF, -78 °C, 6 h, 89%.

Bicyclic alkaloid coniceine $(\mathbf{D})^{24}$ is a representative member of the biologically active indolizidine alkaloids. It acts as a non-competitive blocker for muscle and ganglionic nicotinic receptor channels that have been isolated from the skin secretions of neotropical amphibians.

The synthesis of coniceine (D) began from aldehyde 8, which was prepared from 1,4-butanediol by mono-benzylation, followed by oxidation of the free hydroxy group with pyridinium chlorochromate. The resulting aldehyde was subjected to aza-Prins cyclization with homoallylic amine using gallium iodide and iodine to give 4-iodopiperidine 9 in 85% yield. Radical reduction of 9 was achieved by treatment of tributyltin hydride in the presence of AIBN at reflux in toluene to give piperidine 10 in 80% yield.¹² Both N-tosyl and O-benzyl groups underwent smooth cleavage upon treatment with sodium/naphthalene at -78 °C in anhydrous tetrahydrofuran to afford compound 11 in 85% yield.13 Cyclization of 11 was achieved by treatment with iodine, triphenylphosphine and imidazole followed by aqueous sodium bicarbonate to give coniceine (**D**) in 60% yield (Scheme 5).²⁵ The data of product **D** was in agreement with those reported in the literature.9a

4-Hydroxy-2-hydroxymethylpiperidine (E) was prepared from piperidine 13 by tosyl deprotection upon treatment with sodium/naphthalene at -78 °C. Piperidine 13 was synthesized in 65% yield by aza-Prins cyclization between aldehyde 12 and homoallylic amine with PMA at reflux temperature for 18 hours,^{5b} with concomitant debenzylation. It should be noted that the *N*-tosyl group was stable under these conditions.²⁶ Aldehyde 12 was prepared in good yield from ethylene glycol through monobenzylation followed by oxidation of the free alcohol with pyridinium chlorochromate (Scheme 6). The



Scheme 5 Synthesis of (±)-coniceine (D). *Reagents and conditions*: (a) (i) NaH, BnBr, DMF, cat. TBAI, 0 °C to r.t., 12 h, 90%; (ii) PCC–Celite (1:1), anhydrous CH_2CI_2 , r.t., 12 h, 80%; (b) *N*-tosyl homoally-lic amine, GaI₃, I₂, anhydrous CH_2CI_2 , 0 °C to r.t., 11 h, 85%; (c) Bu₃SnH, AIBN, toluene, 10 h, reflux, 80%; (d) Na/naphthalene, an-hydrous THF, -78 °C, 6 h, 85%; (e) I₂, TPP, imidazole, r.t., 4 h, then sat. NaHCO₃, 3 h, 60%.



Scheme 6 Synthesis of (±)-4-hydroxy-2-hydroxymethylpiperidine (E). *Reagents and conditions*: (a) (i) NaH, BnBr, DMF, cat. TBAI, 0 °C to r.t., 14 h, 85%; (ii) PCC–Celite (1:1), anhydrous CH₂Cl₂, r.t., 10 h, 70%; (b) *N*-tosylhomoallylic amine, PMA (10 mol%), anhydrous CH₂Cl₂, reflux, 20 h, 65%; (c) Na/naphthalene, anhydrous THF, -78 °C, 70%.

data of product \mathbf{E} was in agreement with those reported in the literature.¹⁰

In summary, we have demonstrated a modular and concise route to the total synthesis of (\pm) -coniine, (\pm) -4-hydroxypipecolic acid, (\pm) -pipecolic acid, (\pm) -coniceine, and (\pm) -4-hydroxy2-hydroxymethylpiperidine via aza-Prins cyclization.

Commercial reagents were used without further purification and all solvents were purified by standard techniques. IR spectra were recorded with a Perkin–Elmer FT-IR 240-c spectrophotometer using KBr optics. NMR spectra were recorded in $CDCl_3$ with Varian Gemini 200, Bruker 300, or Varian Unity 400 NMR spectrometers. Chemical shifts (δ) are quoted in parts per million (ppm) and are referenced to tetramethylsilane (TMS) as internal standard; coupling constants (*J*) are quoted in hertz. Column chromatographic separations were carried out on silica gel (60–120 mesh) and flash chromatographic separations were carried out using 230–400 mesh

silica gel. MS were recorded with a Micromass VG-7070H for EI and a VG Autospec M for FAB-MS.

trans-4-Iodo-2-propyl-N-tosylpiperidine (1)

A mixture of *N*-tosylhomoallylic amine (0.430 g, 1.91 mmol), *n*-butyraldehyde (0.178 g, 2.48 mmol), iodine (0.485 g, 1.90 mmol) and gallium iodide (0.008 g, 0.019 mmol) in CH₂Cl₂ (25 mL) was stirred at 0 °C for 10 min and then at r.t. for 7 h. After completion of the reaction (indicated by TLC), the reaction mixture was extracted with CH₂Cl₂ (3 × 20 mL) and the combined organic layers were dried over anhydrous Na₂SO₄. Removal of the solvent, followed by purification on silica gel (EtOAc–hexane, 5:95) gave the product.

Yield: 0.715 g (92%); colorless oil.

IR (neat): 2923, 2853, 1739, 1461, 1374, 1216, 1170, 1119, 759, 668 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.67 (d, *J* = 8.3 Hz, 2 H), 7.30 (d, *J* = 7.9 Hz, 2 H), 4.24 (tt, *J* = 12.4, 3.8 Hz, 1 H), 3.89 (br q, 1 H), 3.63 (ddt, *J* = 14.6, 4.4, 2.3 Hz, 1 H), 3.02 (ddd, *J* = 15.2, 12.4, 2.9 Hz, 1 H), 2.40 (s, 3 H), 2.12–2.20 (m, 2 H), 2.02 (ddt, *J* = 13.4, 4.2, 2.1 Hz, 1 H), 1.87 (dq, *J* = 4.6, 12.3 Hz, 1 H), 1.25–1.60 (m, 4 H), 0.90 (t, *J* = 7.1 Hz, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 13.6, 19.6, 21.4, 31.6, 37.9, 41.3, 42.2, 55.1, 126.8, 129.6, 138.1, 143.2.

LC-MS: m/z = 408 [M + H], 430 [M + Na].

(±)-2-Propyl-N-tosylpiperidine (2)

In an oven-dried, single-neck 25 mL round-bottom flask equipped with reflux condenser, magnetic stirring bar, rubber septum and nitrogen inlet, was placed **1** (0.575 g, 1.41 mmol) and anhydrous toluene (20 mL). Bu₃SnH (0.570 mL, 2.12 mmol) was added and the mixture was heated at reflux for 4 h (130 °C), then AIBN (10 mg) was added to the reaction mixture at 80 °C. After completion of the reaction, as indicated by TLC, the mixture was cooled to r.t. and then concentrated to give a syrup. The resulting crude product was purified by column chromatography (EtOAc–petroleum ether, 15:85) to give **2**.

Yield: 0.378 g (95%); colorless oil.

IR (neat): 2930, 2862, 1734, 1456, 1334, 1156, 1093, 931, 815, 654 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.69 (d, *J* = 8.3 Hz, 2 H), 7.25 (d, *J* = 7.5 Hz, 2 H), 3.97–4.05 (m, 1 H), 3.73 (dd, *J* = 3.0, 13.6 Hz, 1 H), 2.95 (ddd, *J* = 15.6, 13.5, 2.2 Hz, 1 H), 2.43 (s, 3 H), 1.20–1.61 (m, 10 H), 0.90 (t, *J* = 6.7 Hz, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 13.7, 18.2, 19.5, 21.3, 24.2, 27.2, 31.4, 40.4, 52.5, 126.7, 129.4, 138.8, 142.5.

LC-MS: *m*/*z* = 282 [M + H], 299 [M + NH₄].

(±)-Coniine (A)

To a stirred solution of naphthalene (0.732 g, 5.72 mmol) in anhydrous THF (20 mL) in a two-neck 50 mL, round-bottom flask equipped with magnetic stirring bar, a rubber septum and a nitrogen inlet, was added sodium (0.132 g, 5.73 mmol) at r.t. and the mixture was stirred until the formation of sodium naphthalide was complete (indicated by the formation of a persistent navy-blue color; sodium naphthalide was formed within 40 min), and then cooled to -78 °C. A solution of **2** (0.268 g, 0.953 mmol) in anhydrous THF (15 mL) was added to the above solution at -78 °C and the resulting mixture was stirred at the same temperature for 45 min. The reaction was quenched by the addition of sat. aq NH₄Cl (15 mL) and the organic layer was washed with H₂O (2 × 10 mL), brine (3 × 10 mL), dried over anhydrous Na₂SO₄ and concentrated in vacuo. Purification of the crude product by column chromatography (EtOAc–petroleum ether, 35:65) gave the target molecule **A**.

Yield: 0.109 g (90%); syrup.

IR (neat): 3447, 2925, 2855, 1735, 1458, 1335, 1155, 1093, 927, 687 $\rm cm^{-1}.$

¹H NMR (300 MHz, CDCl₃): δ = 3.74–3.79 (m, 1 H), 3.65 (dddd, J = 11.5, 4.0, 2.1, 1.7 Hz, 1 H), 3.50 (ddd, J = 11.5, 11.3, 2.4 Hz, 1 H), 2.30–2.36 (m, 1 H), 2.12–2.17 (m, 1 H), 1.77–1.81 (m, 2 H), 1.57–1.70 (m, 3 H), 1.25–1.36 (m, 4 H), 0.93 (dd, J = 7.0, 6.9 Hz, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 13.8, 19.6, 26.1, 32.3, 33.8, 40.6, 46.1, 50.7.

LC-MS: m/z = 150 [M + Na].

trans-2-Styryl-N-tosylpiperidin-4-ol (3)

A mixture of *N*-tosylhomoallylic amine (0.338 g, 1.50 mmol), *trans*-cinnamaldehyde (0.257 g, 1.95 mmol) and phosphomolybdic acid (0.027 g, 0.015 mmol) in CH_2Cl_2 (25 mL) was stirred at reflux temperature for 10 h. After completion of the reaction, as indicated by TLC, the reaction mixture was extracted with CH_2Cl_2 (3 × 20 mL) and the combined organic layers were dried over anhydrous Na₂SO₄. Removal of the solvent followed by purification on silica gel (EtOAc–hexane, 18:82) gave the product.

Yield: 0.456 g (85%); white solid; mp 170-173 °C.

IR (neat): 3452, 2929, 2854, 1598, 1448, 1334, 1156, 1081, 1022, 929, 755, 669 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.64 (d, *J* = 8.2 Hz, 2 H), 7.00– 7.20 (m, 7 H), 6.34 (d, *J* = 16.6 Hz, 1 H), 5.85 (dd, *J* = 5.8, 15.6 Hz, 1 H), 4.78–4.84 (m, 1 H), 3.75–3.83 (m, 2 H), 3.02 (ddd, *J* = 15.1, 13.2, 2.7 Hz, 1 H), 2.35 (s, 3 H), 2.01 (ddd, *J* = 13.0, 3.8, 1.8 Hz, 1 H), 1.80 (dddd, *J* = 11.9, 2.5, 3.7, 3.9 Hz, 1 H), 1.61 (ddd, *J* = 12.8, 11.3, 5.5 Hz, 1 H), 1.35 (dddd, *J* = 12.0, 11.9, 4.5, 3.5 Hz, 1 H).

¹³C NMR (75 MHz, CDCl₃): δ = 21.2, 34.0, 38.9, 40.3, 55.0, 64.3, 125.9, 127.1, 127.6, 128.3, 129.4, 130.5, 132.0, 136.5, 137.1, 143.1.

LC-MS: m/z = 358 [M + H].

HRMS (ESI): m/z [M + H] calcd for C₂₀H₂₄NO₃S: 358.1478; found: 358.1476.

trans-4-Hydroxy-N-tosylpiperidine-2-carboxylic Acid (4)

To a stirred solution of NaIO₄ (0.209 g, 0.976 mmol) in MeCN– CCl₄–H₂O (1:1:10, 12 mL) was added RuCl₃·2H₂O (0.011 g, 0.048 mmol) and the resulting mixture was stirred at r.t. for 1 h. Compound **3** (0.174 g, 0.488 mmol) was dissolved in MeCN (10 mL) and this solution was added to the above reaction mixture, followed by the addition of a second portion of NaIO₄ (0.105 g, 0.490 mmol). The resulting mixture was stirred at r.t. for 30 min and then filtered through Celite. The organic layer was dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography (MeOH–CHCl₃, 1:9 to 2:8) to yield acid **4**.

Yield: 0.099 g (68%); colorless oil.

IR (neat): 3460, 2924, 2855, 1726, 1454, 1273, 1161, 1097, 708 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.73 (d, *J* = 7.3 Hz, 2 H), 7.28 (d, *J* = 8.2 Hz, 2 H), 5.05 (d, *J* = 5.4 Hz, 1 H), 4.90–4.99 (br s, 1 H), 3.96–4.00 (m, 1 H), 3.52 (ddd, *J* = 4.5, 10.9, 13.7 Hz, 1 H), 2.71–2.75 (m, 2 H), 2.49–2.56 (m, 1 H), 2.40–2.49 (m, 4 H).

¹³C NMR (75 MHz, CDCl₃): δ = 20.8, 39.6, 41.5, 41.9, 55.4, 60.3, 127.1, 129.6, 135.8, 143.9, 171.3.

LC-MS: m/z = 300 [M + H], 322 [M + Na].

(±)-4-Hydroxypipecolic Acid (B)

Compound 4 (0.049 g, 0.163 mmol) was subjected to tosyl deprotection using the procedure described above (synthesis of A from 2)

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Yield: 0.019 g (80%); viscous oil.

IR (neat): 3430, 2927, 1630, 1028, 764 cm⁻¹.

¹H NMR (300 MHz, DMSO- d_6): $\delta = 10.4$ (s, 1 H), 5.04–5.07 (m, 2 H), 3.74–3.85 (m, 7 H).

LC-MS: m/z = 168 [M + Na].

trans-4-Iodo-2-styryl-N-tosylpiperidine (5)

A mixture of *N*-tosylhomoallylic amine (0.193 g, 0.857 mmol), *trans*-cinnamaldehyde (0.147 g, 1.11 mmol), iodine (0.218 g 0.858 mmol) and gallium iodide (0.0038 g, 0.008 mmol) in CH₂Cl₂ (15 mL) was stirred at 0 °C for 10 min and then at r.t. for 7 h. After completion of the reaction, as indicated by TLC, the reaction mixture was extracted with CH₂Cl₂ (3 × 10 mL) and the combined organic layers were dried over anhydrous Na₂SO₄. Removal of the solvent and purification on silica gel (EtOAc–hexane, 5:95) gave the product.

Yield: 0.360 g (90%); yellow syrup.

IR (KBr): 2918, 2858, 1646, 1492, 1383, 1334, 1189, 1155, 1087, 931, 700, 651 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.65 (d, *J* = 8.7 Hz, 2 H), 7.13–7.25 (m, 7 H), 6.43 (d, *J* = 16.2 Hz, 1 H), 5.92 (dd, *J* = 5.6, 16.1 Hz, 1 H), 4.63–4.73 (m, 1 H), 4.27 (tt, *J* = 12.3, 3.9 Hz, 1 H), 3.67 (ddt, *J* = 14.5, 4.3, 2.3 Hz, 1 H), 3.10 (ddd, *J* = 15.1, 12.2, 2.8 Hz, 1 H), 2.10–2.55 (m, 7 H).

¹³C NMR (75 MHz, CDCl₃): δ = 21.3, 32.1, 33.7, 38.9, 44.3, 55.5, 125.9, 127.2, 128.3, 129.2, 130.5, 133.1, 134.5, 136.2, 137.1, 143.2. LC-MS: *m*/*z* = 490 [M + Na].

HRMS (ESI): m/z [M + Na] calcd for C₂₀H₂₂NO₂SNa: 490.0287; found: 490.0308.

(±)-2-Styryl-N-tosylpiperidine (6)

Compound 5 (0.259 g, 0.555 mmol) was subjected to the same reaction conditions (synthesis of 2 from 1) with Bu_3SnH (0.24 mL, 0.831 mmol) and AIBN (10 mg) to give compound 6.

Yield: 0.176 g (93%); syrup.

IR (neat): 2927, 2864, 1709, 1450, 1340, 1213, 1158, 1054, 964, 815, 696 $\rm cm^{-1}.$

¹H NMR (300 MHz, CDCl₃): δ = 7.64 (d, *J* = 7.5 Hz, 2 H), 7.10–7.25 (m, 7 H), 6.38 (d, *J* = 15.8 Hz, 1 H), 5.93 (dd, *J* = 6.7, 16.6 Hz, 1 H), 4.69 (dd, *J* = 11.5, 4.5 Hz, 1 H), 3.72 (dd, *J* = 12.8, 2.3 Hz, 1 H), 2.98 (ddd, *J* = 15.8, 12.4, 3.7 Hz, 1 H), 2.34 (s, 3 H), 1.79 (ddd, *J* = 3.7, 7.5, 18.1 Hz, 2 H), 1.49–1.66 (m, 4 H).

¹³C NMR (75 MHz, CDCl₃): δ = 18.8, 21.3, 30.1, 34.0, 40.1, 54.9, 125.3, 126.3, 127.1, 128.3, 129.6, 132.7, 143.3.

LC-MS: m/z = 340 [M - H].

(±)-N-Tosylpiperidine-2-carboxylic Acid (7)

Compound **6** (0.131 g, 0.384 mmol) was subjected to the same reaction conditions (synthesis of **4** from **3**) with RuCl₃·2H₂O (0.009 g, 0.040 mmol), NaIO₄ (0.171 g, 0.799 mmol), and a second portion of NaIO₄ (0.086 g, 0.401 mmol) to give **7**.

Yield: 0.092 g (70%); viscous oil.

IR (neat): 3430, 2926, 2856, 1724, 1666, 1595, 1411, 1356, 1282, 168, 1092, 1064, 1039, 939, 812, 686 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): δ = 9.52 (br s, 1 H), 7.62 (d, J = 8.7 Hz, 2 H), 7.22 (d, J = 8.2 Hz, 2 H), 5.19–5.21 (dd, J = 2.9, 5.8 Hz, 1 H), 3.68 (ddt, J = 3.6, 13.1, 17.5 Hz, 1 H), 3.31–3.38 (ddd,

J = 2.9, 11.7, 16.1 Hz, 1 H), 2.41 (s, 3 H), 2.15 (dddd, *J* = 2.9, 13.1, 15.3, 18.3 Hz, 1 H), 1.24–1.67 (m, 5 H).

¹³C NMR (75 MHz, CDCl₃): δ = 13.9, 21.2, 38.8, 40.1, 42.4, 56.3, 127.1, 129.6, 135.2, 143.8, 171.2.

LC-MS: m/z = 282 [M - 1].

(±)-Pipecolic Acid (C)

Compound 7 (0.042 g, 0.148 mmol) was subjected to tosyl deprotection using the same reaction conditions (synthesis of A from 2) with naphthalene (0.114 g, 0.890 mmol) and sodium metal (0.020 g, 0.869 mmol) to give the target molecule C.

Yield: 0.017 g (89%); syrup.

IR (neat): 3452, 2926, 1723, 1655, 1158, 1036, 822 cm⁻¹.

¹H NMR (300 MHz, DMSO- d_6): δ = 9.05 (br s, 1 H), 3.82–3.89 (m, 1 H), 3.71–3.74 (m, 1 H), 3.51–3.54 (m, 1 H), 3.29–3.31 (m, 1 H), 2.78–2.83 (m, 2 H), 2.51–2.63 (m, 3 H).

LC-MS: m/z = 130 [M + H].

4-(Benzyloxy)butanal (8)

To a stirred solution of 1,4-butanediol (0.283 g, 3.14 mmol) in anhydrous DMF (35 mL) in a single-necked, round-bottom 100 mL flask equipped with nitrogen inlet, was added NaH (0.090 g, 3.77 mmol) at 0 °C and stirring was continued for 30 min. To the above mixture, benzyl bromide (0.27 mL, 3.11 mmol) was added dropwise at 0 °C, then TBAI (0.0011 g, 0.0031 mmol) was added and the reaction was stirred at r.t. for 12 h. After completion of the reaction, as indicated by TLC, the mixture was quenched with H₂O (5 mL) and the aqueous layer was extracted with EtOAc (3 × 15 mL). The combined organic layers were concentrated in vacuo and the resulting syrup was purified by column chromatography (EtOAc–petroleum ether, 15:85) to give the mono-protected alcohol.

Yield: 0.509 g (90%); syrup.

IR (neat): 3383, 2940, 2865, 1714, 1451, 1364, 1275, 1102, 1064, 742, 706, 611 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.25–7.34 (m, 5 H), 4.49 (s, 2 H), 3.61 (t, *J* = 6.1 Hz, 2 H), 3.49 (t, *J* = 6.1 Hz, 2 H), 1.61–1.75 (m, 4 H).

¹³C NMR (75 MHz, CDCl₃): δ = 26.1, 29.4, 61.9, 70.1, 72.6, 127.7, 128.2, 128.1, 137.8.

LCMS: m/z = 181 [M + H].

HRMS (ESI): m/z [M + Na] calcd for $C_{11}H_{16}O_2Na$: 203.1042; found: 203.1042.

This product (0.410 g, 2.27 mmol) was directly oxidized over PCC (0.979 g, 4.55 mmol) and Celite (0.979 g) in CH_2Cl_2 (40 mL) at r.t. for about 12 h. After completion of the reaction, as indicated by TLC, the reaction mixture poured on silica gel and purified by column chromatography (EtOAc–hexane, 7:93) to give aldehyde **8**.

Yield: 0.324 g (80%); oil.

IR (neat): 2949, 2858, 1720, 1600, 1452, 1274, 1106, 1027, 711, 609 $\rm cm^{-1}$.

¹H NMR (500 MHz, CDCl₃): δ = 10.04 (s, 1 H), 7.29–7.35 (m, 5 H), 4.50 (s, 2 H), 4.35 (t, *J* = 6.8 Hz, 2 H), 3.50–3.56 (m, 2 H), 1.79–1.98 (m, 2 H).

¹³C NMR (75 MHz, CDCl₃): δ = 26.2, 64.6, 69.6, 72.7, 127.4, 128.8, 129.3, 129.5, 134.2, 138.3, 192.1.

trans-2-[3-(Benzyloxy)propyl]-N-tosyl-piperidine-4-ol (9)

A mixture of *N*-tosylhomoallylic amine (0.219 g, 0.973 mmol), 4-(benzoloxy)butanal (**8**; 0.225 g, 1.26 mmol), iodine (0.247 g, 0.973 mmol) and gallium iodide (0.004 g, 0.009 mmol) in CH₂Cl₂ (25 mL) was stirred at 0 °C to r.t. for 11 h. After completion of the reaction,

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as indicated by TLC, the reaction mixture was extracted with CH_2Cl_2 (3 × 20 mL), and the combined organic layers were dried over anhydrous Na_2SO_4 . Removal of the solvent, followed by purification on silica gel (EtOAc–hexane, 10:90) gave the product.

Yield: 0.424 g (85%); yellow syrup.

IR (neat): 2936, 1722, 1598, 1451, 1334, 1157, 1075, 1031, 931, 814, 722, 676 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.67 (d, *J* = 7.9 Hz, 2 H), 7.21– 7.30 (m, 7 H), 4.44 (s, 2 H), 4.34 (tt, *J* = 12.2, 3.7 Hz, 1 H), 4.10– 4.25 (m, 1 H), 3.79 (ddt, *J* = 14.7, 4.6, 2.3 Hz, 1 H), 3.44–3.50 (dd, *J* = 4.8, 10.5 Hz, 2 H), 2.96 (ddd, *J* = 2.5, 9.6, 13.5 Hz, 1 H), 2.40 (s, 3 H), 1.49–1.70 (m, 4 H), 0.82–1.25 (m, 4 H).

¹³C NMR (75 MHz, CDCl₃): δ = 21.4, 26.6, 27.6, 32.4, 36.8, 39.3, 42.1, 56.5, 63.1, 72.7, 126.8, 127.5, 128.3, 129.7, 133.1, 135.2, 138.8, 143.2.

LCMS: m/z = 514 [M + H].

(±)-2-[3-(Benzyloxy)propyl]-N-tosylpiperidine (10)

Compound 9 (0.302 g, 0.588 mmol) was subjected to the procedure described (synthesis of 2 from 1) with Bu_3SnH (0.257 mL, 0.883 mmol) and AIBN (10 mg) to give compound 10.

Yield: 0.183 g (80%); syrup.

IR (neat): 2926, 2855, 1595, 1453, 1336, 1215, 1161, 1058, 934, 735, 660 $\rm cm^{-1}.$

¹H NMR (300 MHz, CDCl₃): δ = 7.69 (d, *J* = 8.3 Hz, 2 H), 7.28– 7.33 (m, 7 H), 4.46 (s, 2 H), 4.20 (dd, *J* = 11.0, 5.5 Hz, 1 H), 3.90 (ddt, *J* = 3.7, 14.3, 18.1 Hz, 1 H), 3.44–3.50 (m, 2 H), 3.10 (ddd, *J* = 2.2, 15.1, 12.8 Hz, 1 H), 2.42 (s, 3 H), 1.99 (dddd, *J* = 2.2, 4.5, 14.3, 16.6 Hz, 2 H), 1.25–1.65 (m, 8 H).

¹³C NMR (75 MHz, CDCl₃): δ = 19.0, 21.5, 26.6, 27.5, 29.8, 32.8, 39.1, 53.5, 69.4, 72.8, 126.9, 127.6, 128.4, 129.8, 138.2, 138.5, 143.3.

LCMS: m/z = 386 [M - H].

Amino Alcohol (11)

Compound **10** (0.133 g, 0.344 mmol) was subjected to tosyl deprotection (synthesis of **A** from **2**) using naphthalene (0.264 g, 2.062 mmol) and sodium metal (0.047 g, 2.043 mmol) to give piperidine **11**.

Yield: 0.042 g (86%); syrup.

IR (neat): 3448, 2923, 2853, 1637, 1460, 1375, 770 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): $\delta = 6.23-6.91$ (br s 1 H), 4.46–4.44 (m, 2 H), 4.09–4.18 (m, 1 H), 3.54–3.60 (m, 1 H), 3.29–3.41 (m, 1 H), 3.11–3.25 (m, 1 H), 3.00–3.12 (m, 1 H), 1.53–2.03 (m, 8 H).

¹³C NMR (75 MHz, CDCl₃): δ = 21.1, 26.2, 29.8, 32.2, 32.8, 42.8, 51.9, 63.2.

LCMS: m/z = 166 [M + Na].

(±)-Coniceine (D)

To a stirred solution of **11** (0.021 g, 0.147 mmol) in anhydrous CH_2Cl_2 (15 mL) in a 100 mL single-neck round-bottom flask were added Ph_3P (0.115 g, 0.440 mmol), iodine (0.112 g, 0.440 mmol) and imidazole (0.029 g, 0.440 mmol) at 0 °C. The resulting mixture was stirred at r.t. for 4 h and then quenched with sat. aq NaHCO₃ (10 mL). The aqueous layer was extracted with EtOAc (2 × 10 mL), the combined organic layers were concentrated in vacuo, and the resulting syrup was purified by column chromatography (EtOAc–petroleum ether, 10:90) to give the target molecule **D**.

Yield: 0.011 g (60%); syrup.

¹H NMR (300 MHz, $CDCl_3$): $\delta = 4.06-4.13$ (m, 1 H), 3.57-3.63 (m, 1 H), 2.26-2.33 (m, 1 H), 2.13-2.17 (m, 1 H), 2.02-2.06 (m, 2 H),

1.83-1.92 (m, 1 H), 1.61-1.72 (m, 1 H), 1.47-1.55 (m, 1 H), 1.31-1.41 (m, 1 H), 1.17-1.29 (m, 5 H).

LCMS: m/z = 124 [M - H].

2-(Benzyloxy)acetaldehyde (12)

To a stirred solution of ethylene glycol (0.150 g, 2.41 mmol) in anhydrous DMF (25 mL) in a single-neck, round-bottom 100 mL flask equipped with a nitrogen inlet, was added NaH (0.071 g, 2.95 mmol) at 0 °C and stirring was continued for 30 min. To the above mixture, benzyl bromide (0.21 mL, 2.388 mmol) was added dropwise at 0 °C, then TBAI (0.0017 g, 0.0048 mmol) was added and the mixture was stirred at r.t. for 14 h. After completion of the reaction, as indicated by TLC, the mixture was quenched with H_2O (5 mL) and the aqueous layer was extracted with EtOAc (3×15 mL). The combined organic layers were concentrated in vacuo and the resulting syrup was purified by column chromatography (EtOAc-petroleum ether, 15:85) to give the mono-protected alcohol.

Yield: 0.311 g (85%); syrup.

IR (neat): 3413, 2926, 2863, 1496, 1453, 1358, 1209, 1112, 1067, 891, 741, 698 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.24–7.27 (m, 5 H), 4.47 (s, 2 H), 3.63 (t, J = 4.7 Hz, 2 H), 3.48 (t, J = 4.9 Hz, 2 H), 3.04 (br s, 1 H). ¹³C NMR (75 MHz, CDCl₃): δ = 61.4, 71.2, 73.2, 127.6, 128.2, 137.7.

LCMS: $m/z = 170 [M + NH_4], 175 [M + Na].$

This product (0.216 g, 1.42 mmol) was directly oxidized over PCC (0.611 g, 2.84 mmol) and Celite (0.611 g) in CH₂Cl₂ (30 mL) at r.t. for about 10 h. After completion of the reaction, as indicated by TLC, the reaction mixture was passed through a silica gel column (EtOAc-hexane, 5:95) to give the pure aldehyde 12.

Yield: 0.149 g (70%); oil.

IR (neat): 2867, 1704, 1613, 1498, 1455, 1351, 1310, 1260, 1047, 967, 869, 744, cm⁻¹.

¹H NMR (400 MHz, CDCl₃): δ = 9.28 (s, 1 H), 7.28–7.34 (m, 5 H), 5.14 (d, J = 2.9 Hz, 1 H), 5.09 (d, J = 2.9 Hz, 1 H), 4.90 (s, 2 H).

LCMS: m/z = 149 [M - H].

trans-2-Hydroxymethyl-N-tosylpiperidin-4-ol (13)

A mixture of N-tosylhomoallylic amine (0.2 g, 0.888 mmol), 2-(benzyloxy)acetaldehyde (12; 0.086 g, 0.575 mmol) and phosphomolybdic acid (0.008 g, 0.004 mmol) in CH₂Cl₂ (30 mL) was stirred at reflux temperature for 20 h. After completion of the reaction, as indicated by TLC, the reaction mixture was extracted with CH2Cl2 $(3 \times 20 \text{ mL})$ and the combined organic layers were dried over anhydrous Na₂SO₄. Removal of the solvent followed by purification on silica gel (EtOAc-hexane, 25:75) gave piperidine 13.

Yield: 0.082 g (65%); colorless oil.

IR (neat): 3448, 2988, 2886, 1595, 1342, 1256, 1161, 1098, 1008, 968, 868, 818, 688 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.62 (d, *J* = 8.1 Hz, 2 H), 7.29 (d, J = 7.9 Hz, 2 H), 4.51–4.53 (m, 1 H), 4.36–4.41 (m, 1 H), 3.64–3.70 (m, 1 H), 3.52 (dd, J = 3.5, 9.4 Hz, 1 H), 3.25 (d, J = 9.4 Hz, 1 H), 2.82-2.92 (m, 1 H), 2.45 (s, 3 H), 1.80-1.89 (m, 2 H), 1.63-1.69 (m. 2 H).

¹³C NMR (75 MHz, CDCl₃): δ = 21.3, 30.6, 37.2, 39.8, 55.8, 69.1, 73.6, 126.9, 129.5, 134.8, 143.3.

LCMS: *m*/*z* = 285 [M⁺], 268 [M – OH].

(±)-4-Hydroxy-2-hydroxymethylpiperidine (E)

Compound 13 (0.040 g, 0.140 mmol) was subjected to tosyl deprotection using the same reaction conditions (synthesis of A) with naphthalene (0.107 g, 0.842 mmol) and sodium metal (0.019 g, 0.842 mmol) to give the target molecule E.

Yield: 0.009 g (50%); syrup.

IR (neat): 3471, 2923, 1622, 1615, 1051, 1010, 722 cm⁻¹.

¹H NMR (300 MHz, DMSO- d_6): $\delta = 4.27$ (br s, 1 H), 4.19 (dd, J = 9.06, 11.2 Hz, 1 H), 3.86–3.88 (m, 1 H), 3.62–3.72 (ddd, J = 13.1, 3.5, 2.3 Hz, 1 H), 2.56–2.69 (m, 2 H), 2.37–2.40 (m, 1 H), 1.74-1.76 (m, 1 H), 1.57-1.59 (m, 1 H), 1.23-1.25 (m, 1 H), 0.86-0.89 (m, 1 H).

LCMS: m/z = 149 [M + Na].

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References and Notes

- (1) (a) Jones, T. H.; Blum, M. S. In Alkaloids: Chemical and Biological Perspectives, Vol. 1; Pelletier, S. W., Ed.; Wiley: New York, 1983, Chap. 2, 33. (b) Fodor, G. B.; Colasanti, B. In Alkaloids: Chemical and Biological Perspectives, Vol. 3; Pelletier, S. W., Ed.; Wiley: New York, 1985, Chap. 1, 1-90. (c) Schneider, M. J. In Alkaloids: Chemical and Biological Perspectives, Vol. 10; Pelletier, S. W., Ed.; Wiley: New York, 1986, Chap. 3, 155. (d) Trunz, G. M.; Findlay, J. A. In The Alkaloids; Brossi, A., Ed.; Academic: London, 1985, Chap. 3, 89. (e) Numata, A.; Ibuka, I. In The Alkaloids, Vol. 31; Brossi, A., Ed.; Academic: New York, 1987, 193. (f) Angle, R. S.; Breitenbucher, J. G. Studies in Natural Products Chemistry, In Stereoselective Synthesis (Part J), Vol. 16; Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, 1995, 453. (g) Gershwin, M. E.; Terr, A. Clin. Rev. Allergy Immunol. 1996, 14, 241. (h) Micel, K. H.; Sandberg, F.; Haglid, F.; Norin, T. Acta Pharm. Suec. 1967, 4, 97. (i) Micel, K. H.; Sandberg, F.; Haglid, F.; Norin, T.; Chan, R. P. K.; Craig, J. C. Acta Chem. Scand. 1969, 23, 3479. (j) Kallstrom, S.; Leino, R. Bioorg. Med. Chem. 2008, 16,601.
- (2) (a) Wenzel, B.; Sorger, D.; Heinitz, K.; Scheunemann, M.; Schliebs, R.; Steinbach, J.; Sabri, O. Eur. J. Med. Chem. 2005, 40, 1197. (b) Guzikowski, A. P.; Tamiz, A. P.; Acosta-Burruel, M.; Hong-Bae, S.; Cai, S. X.; Hawkinson, J. E.; Keana, J. F.; Kesten, S. R.; Shipp, C. T.; Tran, M.; Whittermore, E. R.; Woodward, R. M.; Wright, J. L.; Zhou, Z. L. J. Med. Chem. 2000, 43, 984; and references cited therein. (c) Watson, A. A.; Fleet, G. W. J.; Asano, N.; Molyneux, R. J.; Nash, R. J. Phytochemistry 2001, 56, 265. (d) Daly, J. W.; Spande, T. F.; Garraffo, H. M. J. Nat. Prod. 2005, 68, 1556. (e) Asano, N. Curr. Top. Med. Chem. 2003, 3, 471. (f) Howard, N.; Abell, C.; Blakemore, W.; Chessari, G.; Congreve, M.; Howard, S.; Jhoti, H.; Murray, C. W.; Seavers, L. C. A.; Van Montfort, R. L. M. J. Med. Chem. 2006, 49, 1346. (g) Zhou, Y.; Gregor, V. E.; Ayida, B. K.; Winters, G. C.; Sun, Z.; Murphy, D.; Haley, G.; Bailey, D.;

Froelich, J. M.; Fish, S.; Webber, S. E.; Hermann, T.; Wall,
D. *Bioorg. Med. Chem. Lett.* 2007, *17*, 1206. (h) Baliah, V.;
Jeyaraman, R.; Chandrasekaran, L. *Chem. Rev.* 1983, *83*, 379. (i) Horton, D. A.; Bourne, G. T.; Smythe, M. L. *Chem. Rev.* 2003, *103*, 893. (j) Michael, J. P. *Nat. Prod. Rep.* 2008, 25, 139. For reviews, see: (k) Peters, R.; Jautze, S. *Synthesis* 2010, 365. (l) Hiemstra, H.; Marcelli, T. *Synthesis* 2010, 1229. (m) Huang, Y.; Domling, A. *Synthesis* 2010, 2859.

- (3) (a) Felpin, F. X.; Lebreton, J. Eur. J. Org. Chem. 2003, 3693. (b) Couty, F. Amino Acids 1999, 16, 297. (c) Buffat, M. G. P. Tetrahedron 2004, 60, 1701; and references therein. (d) Speckamp, W. N.; Hiemstra, H. Tetrahedron 1985, 41, 4367. (e) Shipman, M.; Clarkson, G. J.; Wynne, E. Tetrahedron Lett. 2008, 49, 250. (f) Bailey, P. D.; Millwood, P. A.; Smith, P. D. Chem. Commun. 1998, 633. (g) Cortez, G. A.; Schrock, R. R.; Hoveyda, A. H. Angew. Chem. Int. Ed. 2007, 46, 4534. (h) Watson, P. S.; Jiang, B.; Scott, B. Org. Lett. 2000, 2, 3679.
- (4) (a) Carballo, R. M.; Ramirez, M. A.; Rodriguez, M. L.; Martin, V. S.; Padron, J. I. *Org. Lett.* **2006**, *8*, 3837.
 (b) Kishi, Y.; Nagura, H.; Inagi, S.; Fuchigami, T. *Chem. Commun.* **2008**, 3876.
- (5) (a) Yadav, J. S.; Reddy, B. V. S.; Chaya, D. N.; Kumar, G. G. K. S. N.; Naresh, P.; Aravind, S.; Kunwar, A. C.; Madavi, C. *Tetrahedron Lett.* **2008**, *49*, 3330. (b) Yadav, J. S.; Reddy, B. V. S.; Chaya, D. N.; Kumar, G. G. K. S. N.; Naresh, P.; Jagadeesh, B. *Tetrahedron Lett.* **2009**, *50*, 1799.
- (6) (a) Guerrier, L.; Royer, J.; Grierson, D. S.; Husson, H. J. Am. Chem. Soc. 1983, 105, 7754. (b) Moody, C. J.; Lightfoot, A. P.; Gallagher, P. T. J. Org. Chem. 1997, 62, 746.
 (c) Klegraf, E.; Follmann, M.; Schollmeyer, D.; Kunz, H. Eur. J. Org. Chem. 2004, 3346. (d) Airiau, E.; Girard, N.; Pizzeti, M.; Salvadori, J.; Taddei, M.; Mann, A. J. Org. Chem. 2010, 75, 8670.
- (7) (a) Chattopadhyay, S. K.; Biswas, T.; Biswas, T. *Tetrahedron Lett.* 2008, 49, 1365. (b) Vink, M. K. S.; Schortinghuis, C. A.; Luten, J.; van Maarseveen, J. H.; Schoemaker, H. E.; Hiemstra, H.; Rutjes, F. P. J. T. *J. Org. Chem.* 2002, 67, 7869. (c) Rutjes, F. P. J. T.; Veerman, J. J. N.; Meester, W. J. N.; Hiemstra, H.; Schoemaker, H. E. *Eur. J. Org. Chem.* 1999, 1127.
- (8) (a) Lemire, A.; Charette, A. B. J. Org. Chem. 2010, 75, 2077. (b) Fadel, A.; Lahrache, N. J. Org. Chem. 2007, 72, 1780.
- (9) (a) Sibi, M. P.; Christensen, J. W. J. Org. Chem. 1999, 64, 6434. (b) Wilson, S. R.; Sawicki, R. A. J. Org. Chem. 1979, 44, 330. (c) Hart, D. J.; Tsai, Y.-M.; Choi, J.-K.; Burnett, D. A. J. Am. Chem. Soc. 1984, 106, 8201.
- (10) For the data of compound E, see: Takahata, H.; Banba, Y.;
 Ouchi, H.; Nemoto, H.; Kato, A.; Adachi, I. *J. Org. Chem.* 2003, *68*, 3603.
- (11) (a) Lebrun, S.; Couture, A.; Deniau, E.; Grandclaudon, P. Tetrahedron: Asymmetry 2008, 19, 1245. (b) Vetter, J. Food Chem. Toxicol. 2004, 42, 1373. (c) Reynolds, T. R. Phytochemistry 2005, 66, 1399.

- (12) Davis, F. A.; Song, M.; Augustine, A. J. Org. Chem. 2006, 71, 2779.
- (13) Lu, Z.-H.; Zhou, W.-S. *Tetrahedron* **1993**, *49*, 4659.
- (14) (a) Lamarre, D.; Croteau, G.; Bourgon, L.; Thibeault, D.; Wardrop, E.; Clouette, C.; Vaillancourt, M.; Cohen, E.; Pargellis, C.; Yoakim, C.; Anderson, P. *Antimicrob. Agents Chemother.* 1997, *41*, 965. (b) Anderson, P. C.; Soucy, F.; Yoakim, C.; Lavallee, P.; Beaulieu, P. L. US Patent 5 614 533, 1997; *Chem. Abstr.* 1997, 2131185. (c) Wu, W. J.; Raleigh, D. P. *J. Org. Chem.* 1998, *63*, 6689. (d) Cowell, S. M.; Lee, Y. S.; Cain, J. P.; Hurby, V. J. *Curr. Med. Chem.* 2004, *11*, 2785.
- (15) (a) Komoto, T.; Yano, K.; Ono, J.; Okawa, J.; Nakajima, T. JP Patent 61035788, **1986**. (b) Takemoto, Y.; Ueda, S.; Takeuchi, J.; Nakmoto, T.; Iwata, C. *Tetrahedron Lett.* **1994**, *35*, 8821.
- (16) (a) Beaulieu, P. L.; Lavallee, P.; Abraham, A.; Anderson, P. C.; Boucher, C.; Bousquet, Y.; Duceppe, J.; Gillard, J.; Gorys, V.; Grand-Maitre, C.; Grenier, L.; Guindon, Y.; Guse, I.; Plamondon, L.; Soucy, F.; Valois, S.; Wernic, D.; Yoakim, C. *J. Org. Chem.* **1997**, *62*, 3440. (b) Anderson, P. C.; Souey, F.; Yoakim, C.; Lavallee, P.; Beaulieu, P. L. US Patent 5.545.640, **1996**.
- (17) (a) Liang, N.; Datta, A. J. Org. Chem. 2005, 70, 10182.
 (b) Nair, L. G.; Saksena, A.; Lovey, R.; Sannigrahi, M.; Wong, J.; Kong, J.; Fu, X.; Girijavallabhan, V. J. Org. Chem. 2010, 75, 1285. (c) Confalonieri, G.; Marrotta, E.; Rama, F.; Righi, P.; Rosini, G.; Serra, R.; Venturelli, F. Tetrahedron 1994, 50, 3235.
- (18) (a) Morrison, R. I. *Biochem. J.* **1953**, *53*, 474.
 (b) Pastuszak, I.; Molyneux, R. J.; James, L. F.; Elbein, A. D. *Biochemistry* **1990**, *29*, 1886.
- (19) Portoghese, P. S.; Pazdernik, T. L.; Kuhn, W. L.; Hite, G.; Shafiee, A. J. Med. Chem. 1968, 11, 12.
- (20) (a) Tanaka, H.; Kuroda, A.; Marusawa, H.; Hatanaka, H.; Kino, T.; Goto, T.; Hashimoto, M.; Taga, T. J. Am. Chem. Soc. 1987, 109, 5031. (b) Ireland, R. E.; Gleason, J. L.; Gegnas, L. D.; Highsmith, T. K. J. Org. Chem. 1996, 61, 6856.
- (21) (a) Swindells, D. C. N.; White, P. S.; Findlay, J. A. *Can. J. Chem.* **1978**, *56*, 2491. (b) Smith, A. B. III; Condon, S. M.; McCauley, J. A.; Leazer, J. L. Jr.; Leahy, J. W.; Maleczka, R. E. Jr. *J. Am. Chem. Soc.* **1997**, *119*, 962.
- (22) Pastuszak, I.; Molyneux, R. J.; James, L. F.; Elbein, A. D. *Biochemistry* **1990**, *29*, 1886.
- (23) Skiles, J. W.; Giannousis, P. P.; Fales, K. R. Bioorg. Med. Chem. Lett. 1996, 6, 963.
- (24) (a) Michael, J. P. *Nat. Prod. Rep.* 2005, *22*, 603.
 (b) Erspamer, V.; Melchiorri, P. *Trends Pharmacol. Sci.* 1980, *1*, 391. (c) Aronstam, R. S.; Daly, J. W.; Spande, T. F.; Narayanan, T. K.; Albuquerque, E. X. *Neurochem. Res.* 1986, *11*, 1227.
- (25) (a) Xiaotao, P.; Dawei, M. J. Org. Chem. 2003, 68, 4400.
 (b) Padwa, A.; Harris, J. M. J. Org. Chem. 2003, 68, 4371.
- (26) Baskaran, S.; Kumar, G. D. K. J. Org. Chem. 2005, 70, 4520.