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Synthesis and anticancer evaluation of bis(benzimidazoles), bis(benzoxazoles), and benzothiazoles

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Abstract—Four classes of UK-1 analogues were synthesized and their cytotoxicity testing against human A-549, BFTC-905, RD, MES-SA, and HeLa carcinoma cell lines was determined. The results revealed that UK-1 and four of these analogues (15–18) are potent against the cancer cell lines. In particular, compound 16 is more potent than UK-1 against A-549 and HeLa cell lines, and compounds 15, 17, and 18 selectively exhibit potent cytotoxic activity against the BFTV-905 cells (IC₅₀ 9.6 μ M), A-549 cells (IC₅₀ 6.6 μ M), and MES-SA cells (IC₅₀ 9.2 μ M), respectively. © 2006 Elsevier Ltd. All rights reserved.

1. Introduction

Taniguchi et al. isolated the bis(benzoxazole) natural product UK-1 from the mycelial cake of an actinomycete strain 517-02.¹⁻³ Subsequently, Tsuji et al. isolat-AJI9561 from the mycelium extract ed of Streptomyces sp. AJ9561.⁴ UK-1 exhibits significant cvtotoxic activity against B16. HeLa, and P338 cells. but does not exhibit any growth-inhibiting activity against Gram-positive bacteria, yeasts or fungi at up to 100 µg/mL.¹ AJI9561 exhibited cytotoxic activity against both Jurkat and P388 cells with IC₅₀ values of 0.88 and 1.63 µM, respectively. However, the methyl derivative MUK-1 and alkaline hydrolysate demethyl derivative DMUK-1 both exhibit activity against Gram-positive and Gram-negative bacteria.^{2,3} MUK-1 is also active against yeast and filamentous fungi.² Kerwin et al. investigated the metal ion coordination by UK-1 and found that UK-1 binds Zn^{2+} and Mg^{2+} to form stable complexes with an overall stoi-chiometry of 1:1.⁵ Fe³⁺ ions are bound slightly less strongly by UK-1, and Ca²⁺ ions form weak complexes with this ligand. In the presence of Mg^{2+} , UK-1 binds to double-stranded DNA ten times more tightly than in the absence of Mg^{2+} . Like the antitumor agent quinobenzoxazines, which also bind Mg^{2+} and bind to

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DNA in a magnesium ion-dependent fashion, UK-1 inhibits the activity of human topoisomerase II.



Kerwin et al. subsequently reported the activity of UK-1 against a wide range of human cancer cell lines.⁶ UK-1 exhibits a wide spectrum of potent anticancer activity against leukemia, lymphoma, and certain solid tumorderived cell lines, with IC₅₀ values of as low as 20 nM, but is inactive against Staphylococcus aureus. A series of analogues of the bis(benzoxazole) natural product UK-1 in which the carbomethoxy-substituted benzoxazole ring of the natural product was modified were prepared, and their anticancer and antibacterial properties evaluated. An analogue of UK-1, in which the carbomethoxy-substituted benzoxazole ring was replaced by a carbomethoxy-substituted benzimidazole ring (compound **B**), was inactive against human cancer cell lines and the two strains of S. aureus. In contrast, a simplified analogue in which the carbomethoxy-substituted benzoxazole ring was replaced with a carbomethoxy group (compound A) was almost as active as UK-1 against the four cancer cell lines examined but exhibited no activity against S. aureus. The benzylated UK-1 analogue (compound C) is more active than UK-1 against HT-29 cells, about 10-fold less potent than UK-1 against MCF-7 and PC-3 cells, and 20-fold less active than UK-1 against HL60 cells. Metal ion binding investigations of these analogues indicate that they bind both Zn²⁺ and Ca²⁺ approximately as strongly as UK-1. The non-cytotoxic benzimidazole UK-1 analogue binds Mg²⁺ ions 50 times more weakly than UK-1, while the simple benzoxazole analogue binds Mg^{2+} ions almost as effectively as UK-1. These results are evidence of the involvement of Mg²⁺ ion binding in the selective cytotoxicity of UK-1 and its provision of a minimal pharmacophore for the selective cytotoxic activity of the natural product.



Smith et al. synthesized three new analogues of UK-1 and evaluated their efficacies as topoisomerase II inhibitors.⁷ Their results revealed that UK-1 and two of these analogues (compounds **D** and **E**) are catalytic inhibitors of topoisomerase II, whereas compound **F** shows no measurable activity, and identify a critical structural motif required for enzyme inhibition.



A number of structural analogues of this natural product were prepared to understand the structural basis of the selective cytotoxicity of UK-1. A comparison of the anticancer activity of these compounds with that of their structurally simplified analogues of UK-1 elucidates the natural product's selective activity against cancer cells.

2. Chemistry

A number of analogues of UK-1 in which the carbomethoxy-substituted benzoxazole ring was either modified or deleted were prepared. These UK-1 analogues were synthesized in a manner that was analogous to the previously reported total synthesis of the natural product⁶⁻⁸ with some improvements. An analogue of UK-1, in which the carbomethoxy-substituted benzoxazole ring is replaced with a substituted benzimidazole ring, was prepared according to the Schemes 1-3. Compound 6 was obtained in five steps, starting with the esterification of 2-aminobenzoic acid (1) with diazomethane to yield methyl 2-aminobenzoate (2) in 94% yield (Scheme1). The nitration of compound 2 gave 2-acetylamino-3-nitro-benzoic acid methyl ester 3 with a 37% yield, and 2-acetylamino-5-nitro-benzoic acid methyl ester 4 with a 61% yield.⁹ Following separation, the acetyl group of 3 was cleaved using sodium methoxide¹⁰ and treated with diazomethane to esterify partially hydrolyzed ester 5 with a 87% yield. The nitro group of 5 was reduced by hydrogenation on Pd/C to yield 2,3-diaminobenzoic acid methyl ester 6 with a 96% yield.

Compound 12 was prepared in five steps, starting with the esterification of salicylic acid 7 by diazomethane to generate methyl salicylate 8 in quantitative yield (Scheme 2). The hydroxyl group of 8 was protected with benzyl ether by treating with benzyl bromide and potassium carbonate to give 9 in 93% yield.¹¹ Compound 9 was transformed to the carboxylic acid 10 in quantitative yield by saponification reaction. Compound 10 was coupled with 6 by treating it with dicyclohexylcarbodiimide to furnish amide 11 with a 83% yield which was then converted to benzimidazole 12 in 87% yield via cyclization-dehydration by refluxing in acetic acid.¹² Compound 12 was saponified by 5 M NaOH to give 13 in 97% yield. The benzyl groups in 12 and 13 were cleaved by hydrogenation to generate the corresponding phenols 14 and 15 in 97% and 60% yields, respectively.

Compound 13 was transformed to acyl chloride by treatment with thionyl chloride and then coupled with



Scheme 1. Synthesis of 2,3-diaminobenzoic acid methyl ester 6.



Scheme 2. Synthesis of benzimidazole analogues 12-15.



Scheme 3. Synthesis of bis(benzimidazole) analogues 17-19.

6 to give **16** in 56% yield (Scheme 3). Compound **16** was refluxed in acetic acid to form the bis(benzimidazole) **17** in 85% yield, and this **17** was converted to **18** in quantitative yield by hydrogenation, and to **19** in 66% yield by saponification.

UK-1 is typically synthesized following the procedure of Kerwin et al. (Scheme 4).⁵ Compound **20** was prepared from 2-amino-3-hydroxybenzoic acid by treating with diazomethane. Compound **10** was transformed to acyl chloride by treating with oxalyl chloride and *N*,*N*-dimethylformamide, and then reacted with **20** to give the amide **22** in 96% yield. Heating **22** to 230 °C produced

a cyclization-dehydration product 23 in 86% yield, which was saponified to give 24 in 84% yield. Compound 24 was converted to acyl chloride by treating it with oxalyl chloride and *N*,*N*-dimethylformamide, and then reacting it with 20 to give 26 in 76% yield. Compound 23 was refluxed with *p*-toluenesulfonic acid in toluene to give compound 25 in 51% yield. Final cyclodehydration and debenzylation were performed by refluxing 26 with *p*-toluenesulfonic acid in *p*-xylene to give the UK-1 in 43% yield.

One of the UK-1 analogues, 35, was prepared from salicylic acid presented in Scheme 5. Salicylic acid



Scheme 4. Synthesis of UK-1.



Scheme 5. Synthesis of 31–35.

was transformed to **27** by treating it with iodomethane and potassium carbonate.¹³ It was then treated with sodium hydroxide to give **28** in 51% yield (in 2 steps). Compound **28** was converted to acyl chloride and coupled with **29** to produce amide **30** in 74% yield. Compound **30** was refluxed with *p*-toluenesulfonic acid in *p*-xylene to give **31** by cyclization-dehydration in 32% yield and a demethylation product **32** in 9% yield. Compound 31 was converted to 33 by saponification. Then, 33 was treated with oxalyl chloride and N,N-dimethylformamide to produce the corresponding acyl chloride, which was coupled with 29 to give the amide 34 in 61% yield (in two steps). Compound 34 was refluxed with *p*-toluenesulfonic acid in *p*-xylene to give 35 by cyclization-dehydration.



Scheme 6. Synthesis of 38-41.

UK-1 analogues with benzothiazole moiety were prepared according to Scheme 6. Compound 10 was treated with oxalyl chloride and N,N-dimethylformamide to produce the corresponding acyl chloride, which was coupled with 2 to give the amide 36 in 81% yield. Then, 36 was converted to thioamide 37 in 51% yield by treatment with Lawesson's reagent.¹⁴ Thioamide 37 was cyclized to benzothiazoles 38 by Jacobson synthesis with alkaline potassium ferricyanide. The crude products contained compounds 38 and 40. Therefore, the crude products were treated with diazomethane and purified by flash column chromatography to give 38 in 21% yield. Compound 39 was prepared by the debenzylation of 38 using ethanethiol and boron trifluoride-diethyl etherate. Compound 38 was converted to acid 40 by saponification. Then, 40 was treated with oxalyl chloride and N,N-dimethylformamide and the corresponding acyl chloride was produced;¹⁵ it was coupled with 2 to give the amide 41 in 51% yield (in two steps). Then, an attempt was made to convert 41 to thioamide by treating it with Lawesson's reagent and Jacobson synthesis with alkaline potassium ferricyanide was used to prepare the corresponding bis(benzothiazole), but the attempt was unsuccessful.

3. Biological assay

The cytotoxicity of UK-1 against P388, B16, and HeLa cell lines has been reported elsewhere.¹ A range of human cancer cell lines A-549 and HeLa cells were investigated to determine the spectrum of cell lines whose growth is inhibited by UK-1. The results, presented in Table 1, indicate that UK-1 exhibits potent anticancer activity against human lung and epithelial carcinoma cell lines. The cytotoxicities of the UK-1 analogues to

Table 1. $IC_{50}{}^a~(\mu M)$ values of tumor cell lines after 72 h continuous exposure to test compounds

Compound	Tumor type/cell line				
	A-549 ^b	BFTC-905°	RD ^d	MES-SA ^e	HeLa ^f
12	70.0	53.8	>100	50.6	67.6
13	87.0	66.4	>100	91.0	67.3
14	74.1	37.9	35.9	33.2	31.5
15	38.2	9.6	17.6	52.7	52.5
16	2.8	11.9	13.8	<5	7.1
17	6.6	36.8	44.9	15.3	nd
18	22.2	29.9	26.5	9.2	14.5
19	46.9	21.4	28.0	33.2	29.3
23	>100	89.2	nd	91.3	82.1
24	>100	98.3	>100	63.9	90.3
25	>100	49.6	58.5	>100	75.8
26	50.8	84.9	87.8	41.0	51.4
31	>100	>100	>100	>100	52.6
32	>100	54.7	>100	89.4	67.6
33	>100	>100	>100	>100	>100
34	nd ^g	nd	nd	nd	nd
39	>100	87.3	39.1	76.7	81.6
41	nd	nd	nd	nd	nd
UK-1	5.1	nd	nd	nd	10.7

 a IC₅₀ is the concentration that induces 50% growth inhibition compared with untreated control cells.

^b human lung carcinoma cell line.

^c human bladder carcinoma transitional cell line.

^d human rhabdomyosarcoma cell line.

^e human uterine sarcoma cell line.

^f Human epithelial carcinoma cell line.

^g nd, not determined.

na, not acterninea.

cancer cells were examined in order to elucidate the structural basis for selective cytotoxicity of the natural product toward a unique cancer cell. Table 1 summarizes the results of cytotoxicity testing against human

A-549, BFTC-905, RD, MES-SA, and HeLa carcinoma cell lines. UK-1 exhibits potent cytotoxic activity against the A-549 and HeLa cancer cells lines, but compound 16 is more potent than UK-1 against both A-549 cells (IC_{50}) $2.8 \,\mu\text{M}$ vs $5.1 \,\mu\text{M}$ for UK-1) and HeLa cells (IC₅₀ 7.1 µM vs 10.7 µM for UK-1). Compound 16 is the precursor of compound 18, but compound 18 is less potent than 16 against A-549 cells (IC₅₀ 22.2 μ M vs 2.8 μ M for 16), MES-SA (IC₅₀ 9.2 μ M vs <5 μ M for 16), and HeLa cells (IC₅₀ 14.5 μ M vs 7.1 μ M for 16). Only one benzimidazole ring was formed in compound 16, and this skeleton relative to 18 may increase the flexibility to coordinate with Mg^{2+} ion, and enhanced cytotoxicity against cancer cell lines. Compound 26 is the precursor of UK-1, but compound 26 is less potent than UK-1 against A-549 cells (IC₅₀ 50.8 μ M vs 5.1 μ M for UK-1) and HeLa cells (IC₅₀ 151.4 μ M vs 10.7 μ M for UK-1). Even though compound 18 has a bis(benzimidazole) skeleton and UK-1 has the corresponding bis(benzoxazole) skeleton, compound 18 is less potent than UK-1 against A-549 cells (IC₅₀ 22.2 μ M vs 5.1 μ M for UK-1) and HeLa cells (IC₅₀ 14.5 μ M vs 10.7 μ M for UK-1). Kerwin et al. reported the ability of UK-1 to form metal ion complexes that can bind to DNA^{5,17} and inhibit DNA-processing enzymes,⁵ which findings indicate that Mg²⁺ ion binding by UK-1 may lead to biologically relevant complexes with a specific target in cancer cells. Therefore, the interaction between bis(benzoxazole) and the Mg^{2+} ion may be stronger than that of bis(benzimidazole). Interestingly, compounds 15, 17, and 18 selectively exhibit potent cytotoxic activity against the BFTC-905 cells (IC₅₀ 9.6 µM), A-549 cells (IC₅₀ 6.6 μ M), and MES-SA cells (IC₅₀ 9.2 μ M), respectively.

Compounds 14, 25, 32, and 39 have the same skeleton and differ only in their hetero atoms. Interestingly, the compound that has benzimidazole moiety exhibits stronger anticancer activity against cell lines than those that contain benzoxazole or benzothiazole. Compound 18 is less potent than UK-1 against cancer cell lines. Compound 18 contains bis(bezoimidazole) moiety, and UK-1 has bis(benzoxazole) moiety, perhaps because the steric energies¹⁸ of chelating complexes of Mg²⁺ with compounds 18 (11.71 kcal/mol) exceed that of UK-1 (9.29 kcal/mol). When ester 14 was hydrolyzed to form the corresponding acid 15, the power of 15 to inhibit BFTC-905 was increased. The potency of compound 17 against A-549 is stronger than that of compound 18, while the inhibitive potency against BFTC-905, RD, and MES-SA of compound 18 exceeds that of compound 17. The hydroxyl group of compound 18 was converted to benzyl ether in 17. Therefore, the binding sites in compounds 18 and 17 should differ from each other in the cell lines, because compound 17 is large sized and hydrophobic. Compound 16 is the most potent against cancer cell lines herein. The flexibility of 16 may increase its ability to chelate with Mg^{2+} and to promote the interaction at the binding site. Unfortunately, compound 26 is the precursor of UK-1 that corresponds to the compound 16 as the precursor of compound 18. It does not powerfully inhibit activity against cancer cell lines, even though it has good flexibility. Although compounds 16 and 26 have anilide functional groups, the positions of $-NH_2$ groups that form anilide differ, and the conformations of their complexes formed with Mg^{2+} differ. This occurrence may imply that compound **26** is less potent than compound **16** and UK-1, even though it is the precursor of UK-1.

3.1. Conclusion

The natural product of bis(benzoxazole), UK-1, has potential for use against cancer cell lines. Structurally similar bis(benzimidazole) **17** and **18** were also found to exhibit potent anticancer activity. Interestingly, compound **16** is the synthetic precursor of compounds **17**-**19** and is more potent than UK-1 against human lung (A-549) and epithelial (HeLa) carcinoma cell lines. Therefore, further work with UK-1, compounds **15**-**18**, and their analogues, must be conducted to determine whether such targeting is involved in the selective cytotoxicity of UK-1 and compounds **15–18**, and whether the promising spectrum of the in vitro anticancer activity of UK-1 and **15–18** reported herein is also reflected in vivo.

4. Experimental

4.1. Chemistry

4.1.1. General methods. Melting points were recorded using a Yanagimoto Micro Melting Point Apparatus Model-S3 capillary melting point apparatus and are uncorrected. TLC analysis was carried out on silica gel 60 F254 precoated glass sheets and detected under UV light. ¹H and ¹³C NMR spectra were recorded employing a Bruker AVANCE DPE-300 or VARIAN UNI-YTINOVA-500 spectrometer, and chemical shifts (δ) are in ppm relative to CHCl₃ (7.26 ppm or 77 ppm), CD₃OD (δ = 3.30 ppm or 49 ppm), and (CD₃)₂SO (2.49 ppm or 39.7 ppm) as internal standard. Infrared spectra were recorded on a JASCO FT/IR 410 spectrometer. Mass spectra were recorded on a Micromass Platform II or Finnigan/Thermo Quest MAT 95XL spectrometer. In each case proton NMR showed the presence of indicated solvent(s). All chemicals and solvents were purchased from Aldrich Chemical Co., Fisher Scientific, or Lancaster.

4.1.1.1 2-Amino-benzoic acid methyl ester (2). To a solution of 2-aminobenzoic acid (6.00 g, 44.0 mmol) in diethyl ether (100 mL) was added a solution of diazomethane in diethyl ether at 0 °C until ceased to generate nitrogen. After adding 1 mL of acetic acid, the reaction solution was concentrated and the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49–1:19) as the mobile phase producing compound **2** (6.25 g, 41.0 mmol). Yield: 94%. ¹H NMR (300 MHz, CDCl₃, δ): 7.87–7.83 (d, J = 8.1 Hz, 1H), 7.29–7.23 (dd, J = 8.4, 7.0 Hz, 1H), 6.67–6.61 (m, 2H), 5.71 (s, 2H), 3.86 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 168.4, 149.6, 134.0, 131.1, 116.9, 116.7, 111.1, 51.5. IR (KBr): 3481, 3371, 1692, 1435, 1274, 752 cm⁻¹. MS *m/z*: 152 (M⁺+1, 5), 151 (M⁺, 65), 120

(31), 119 (100), 92 (50). HRMS-EI (m/z): [M]⁺ calcd for C₈H₉NO₂, 151.0633; found, 151.0637.

4.1.1.2. 2-Acetylamino-3-nitro-benzoic acid methyl ester (3) and 2-acetylamino-5-nitro-benzoic acid methyl ester (4). A solution of compound 2 (6.30 g, 41.0 mmol) in acetic anhydride (47.0 mL) was stirred at room temperature for 1 h, then a solution of acetic anhydride (9.6 mL), acetic acid (8.4 mL), and concd nitric acid (13.7 mL) was added slowly at 5-15 °C. After stirring at 5-15 °C for 2.5 h, the reaction solution was poured into ice-water (50 mL) and the precipitate was filtered to give compound 4 (5.89 g, 25.0 mmol). The filtrate was diluted with water (50 mL) and stood overnight to give crystalline 3. The mother liquid was extracted with dichloromethane, after concentration the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:3, 1:1) as the mobile phase producing compound 3 (3.68 g, 15.0 mmol, combined with crystalline 3).

4.1.1.3. Compound 3. Yield: 37%. mp: 120–122 °C. ¹H NMR (300 MHz, CDCl₃, δ): 10.34 (s, 1H), 8.22–8.19 (d, J = 6.3 Hz, 1H), 8.11–8.07 (d, J = 8.2 Hz, 1H), 7.36–7.31 (dd, J = 8.1, 6.5 Hz, 1H), 3.96 (s, 3H), 2.23 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 168.6, 166.6, 144.2, 134.8, 132.1, 129.4, 123.8, 122.6, 53.0, 24.1. IR (KBr): 3307, 1683, 1527, 1438, 1365, 1267, 1120 cm⁻¹. MS m/z: 238 (M⁺, 0.58), 196 (100), 192 (15), 165 (12), 164 (53), 90 (14). HRMS-EI (m/z): [M]⁺ calcd for C₁₀H₁₀N₂O₅, 238.0590; found, 238.0591.

4.1.1.4. Compound 4. Yield: 61%. mp: 177–181 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.38 (s, 1H), 8.95–8.92 (m, 2H), 8.39–8.35 (d, J = 9.4 Hz, 1H), 4.0 (s, 3H), 2.29 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 169.4, 167.2, 146.5, 141.6, 129.4, 126.8, 120.3, 114.4, 53.0, 25.5. IR (KBr): 3004, 1685, 1587, 1508, 1436, 1348, 902, 821 cm⁻¹. MS *m*/*z*: 238 (M⁺, 18), 196 (100), 164 (45), 119 (12), 57 (28), 55 (15). HRMS-EI (*m*/*z*): [M]⁺ calcd for C₁₀H₁₀N₂O₅, 238.0590; found, 238.0587.

4.1.1.5. 2-Amino-3-nitro-benzoic acid methyl ester (5). To a solution of compound 3 (0.50 g, 2.1 mmol) in anhydrous methanol (25 mL) was added a sodium methoxide solution (37.0 mL, 0.07 mmol, 1.92 M in methanol) at room temperature. After the reaction was completed, added acetic acid to neutralize the reaction solution. The solvent was removed, and the residue was extracted by ethyl acetate. The extract was dried by anhydrous magnesium sulfate and concentrated. The resulting residue was dissolved in a mixture of diethyl ether and methanol, and added diazomethane at 0 °C until ceased to generate nitrogen. After adding a few drops of acetic acid, the reaction solution was concentrated to produce product 5 (0.40 g, 2.0 mmol). Yield: 87%. mp: 97-100 °C. ¹H NMR (300 MHz, CD₃Cl₃, δ): 8.40–8.37 (d, J = 8.4 Hz, 1H), 8.25-8.22 (d, J = 7.7 Hz, 1H), 6.68-6.63 (dd, J = 8.3, 7.8 Hz, 1H), 3.92 (s, 3H). ¹³C NMR $(125.7 \text{ MHz}, \text{ CDCl}_3, \delta)$: 167.4, 147.2, 139.3, 133.1, 132.2, 114.3, 113.9, 52.2. IR (KBr): 3120, 3028, 953, 882, 847, 706 cm⁻¹. MS *m*/*z*: 197 (M⁺+1, 9), 196 (M⁺, 100), 165 (46), 119 (24), 118 (16), 90 (41). HRMS-EI

(m/z): $[M]^+$ calcd for $C_8H_8N_2O_4$, 196.0484; found, 196.0489.

4.1.1.6. 2,3-Diamino-benzoic acid methyl ester (6). A mixture of compound 5 (1.00 g, 5.1 mmol) and Pd/C (0.10 g, 10% of Pd on charcoal) in methanol (30 mL) under hydrogen atmosphere (60 psi) was hydrogenated for 3 h. The catalyst was removed by filtration, the filtrate was concentrated, and the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (3:7) as the mobile phase producing compound 6 (0.81 g, 4.9 mmol). Yield: 96%. mp: 63-65 °C. ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3, \delta)$: 7.48–7.45 (d, J = 8.1 Hz, 1 H), 6.86–6.83 (d, J = 7.5 Hz, 1H), 6.62–6.57 (dd, J = 7.9, 7.7 Hz, 1H), 3.87 (s, 3H). ¹³C NMR (125.7 MHz, $CDCl_3$, δ): 168.9, 141.1, 134.1, 122.6, 120.6, 116.6, 111.9, 51.5. IR (KBr): 3367, 1693, 1433, 1287, 1234, 733 cm^{-1} . MS m/z: 167 (M⁺+1, 8), 166 (M⁺, 100), 135 (19), 134 (74), 106 (97), 105 (28). HRMS-EI (*m*/*z*): $[M]^+$ calcd for C₈H₁₀N₂O₂, 166.0742; found, 166.0748.

4.1.1.7. 2-Hydroxy-benzoic acid methyl ester (8). To a solution of compound 7 (7.80 g, 57.0 mmol) in diethyl ether and methanol was added a solution of diazomethane in diethyl ether at 0 °C until ceased to generate nitrogen. After adding a few drops of acetic acid, the reaction solution was concentrated to produce product (8.60 g, 57.0 mmol). Yield: 100%. ¹H NMR 8 $(300 \text{ MHz}, \text{ CDCl}_3, \delta)$: 10.75 (s, 1H), 7.85–7.82 (d, J = 8.0 Hz, 1H), 7.46–7.42 (dd, J = 8.7, 8.4 Hz, 1H), 6.99–6.96 (d, J = 8.4 Hz, 1H), 6.90–6.85 (dd, J = 8.2, 8.1 Hz, 1H), 3.95 (s, 3H). ¹³C NMR (125.7 MHz, $CDCl_3, \delta$): 170.5, 161.5, 135.6, 129.8, 119.1, 117.5, 112.3, 52.2. IR (KBr): 3186, 1678, 1305, 1090, 845, 757 cm⁻¹. MS m/z: 152 (M⁺, 49), 121 (27), 120 (100), 92 (53), 65 (16). HRMS-EI (m/z): $[M]^+$ calcd for C₈H₈O₃, 152.0473; found, 152.0479.

4.1.1.8. 2-Benzyloxy-benzoic acid methyl ester (9). To a solution of compound 8 (10.00 g, 28.0 mmol) in anhydrous dichloromethane (99.0 mL) and anhydrous methanol (99.0 mL) were added potassium carbonate (20.00 g, 145.0 mmol) and benzyl bromide (15.70 mL, 132.0 mmol), and refluxed for 40 h under nitrogen atmosphere. After filtration, the filtrate was concentrated, and the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49) as the mobile phase producing compound 9 (14.79 g, 61.0 mmol). Yield: 93%. ¹H NMR (300 MHz, CDCl₃, δ): 7.84–7.80 (d, J = 7.6 Hz, 1H), 7.51-7.31 (m, 6H), 7.02-6.99 (m, 2H), 5.19 (s, 2H), 3.91 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 166.7, 158.0, 136.6, 133.3, 131.3, 128.4, 127.6, 126.6, 120.6, 120.4, 113.7, 70.4, 51.8. IR (KBr): 2949, 1727, 1599, 1450, 1249, 754 cm⁻¹. MS *m*/*z*: 242 (M⁺, 3), 210 (4), 92 (9), 91 (100), 65 (8). HRMS-EI (m/z): $[M]^+$ calcd for C₁₅H₁₄O₃, 242.0943; found, 242.0951.

4.1.1.9. 2-Benzyloxy-benzoic acid (10). To a solution of compound **9** (5.20 g, 21.0 mmol) in 1,4-dioxane (50 mL) was added 1.4 M sodium hydroxide solution (22 mL) and refluxed for 2 h. After concentration, the

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remained concentrated solution was acidified to pH 1 by adding 1 N hydrochloric acid. The acidic solution was extracted with ethyl acetate, and the extract was dried over anhydrous magnesium sulfate, filtered, and concentrated to produce compound **10** (5.00 g, 22.0 mmol). Yield: 100%. mp: 73–75 °C. ¹H NMR (300 MHz, CDCl₃, δ): 10.8 (s, 1H), 8.23–8.2 (d, *J* = 7.8 Hz, 1H), 7.57–7.56 (m, 1H), 7.45–7.42 (m, 5H), 7.18–7.12 (m, 2H), 5.26 (s, 2H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 165.3, 157.3, 135.0, 134.2, 133.8, 129.16, 129.12, 127.8, 122.4, 117.9, 113.0, 72.1. IR (KBr): 3048, 2952, 1727, 1600, 1450, 1251, 1083, 755 cm⁻¹. MS *m/z*: 228 (M⁺, 1), 92 (11), 91 (100), 69 (9), 57 (15). HRMS-EI (*m/z*): [M]⁺ calcd for C₁₄H₁₂O₃, 228.0786; found, 228.0795.

4.1.1.10. 2-Amino-3-(2-benzyloxy-benzoylamino)-benzoic acid methyl ester (11). To a solution of compound 10 (1.40 g, 6.0 mmol) and compound 6 (0.50 g, 6.0 mmol)3.0 mmol) in anhydrous dichloromethane (15 mL) were added dicyclohexylcarbodiimide (1.23 g, 6.0 mmol) and 4-(dimethylamino)pyridine (12 mg) under argon atmosphere, and stirred at room temperature for 7 h. The reaction mixture was filtered through celite, and the filtrate was concentrated. The residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:19, 1:9, 3:17) as the mobile phase producing compound 11 (0.94 g, 25.0 mmol). Yield: 83%. mp: 115-117 °C. ¹H NMR (300 MHz, CDCl₃, δ): 9.16 (s, 1H), 8.28–8.26 (d, J = 7.8 Hz, 1H), 7.76–7.73 (d, J = 8.1 Hz, 1H), 7.54-7.44 (m, 7H), 7.18-7.13 (m, 2H), 6.67-6.62 (dd, J = 7.9, 7.9 Hz, 1H), 5.56 (s, 2H), 5.26 (s, 2H), 3.85 (s, 3H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 168.4, 164.0, 156.6, 145.0, 135.1, 133.3, 132.6, 130.1, 129.1, 128.8, 128.6, 124.6, 121.9, 121.8, 115.8, 112.7, 112.3, 71.7, 51.6. IR (KBr): 3282, 2935, 1631, 1446, 1243, 1091, MS m/z: 377 (M⁺+1, 5), 376 (M⁺, 24), 211 $746 \, {\rm cm}^{-1}$ (20), 165 (16), 121 (12). HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₀N₂O₄, 376.1423; found, 376.1419.

4.1.1.11. 2-(2-Benzyloxy-phenyl)-1H-benzoimidazole-4-carboxylic acid methyl ester (12). A solution of compound 11 (0.48 g, 1.3 mmol) in acetic acid (6 mL) was refluxed at 120-130 °C for 3 h. After cooled to room temperature, the reaction solution was poured into ice water and neutralized by sodium bicarbonate. The aqueous solution was extracted with dichloromethane, and the extract was washed with brine and dried over anhydrous magnesium sulfate. The dry solution was concentrated and dried under vacuum to give compound 12 (0.40 g, 1.1 mmol). Yield: 87%. mp: 103–105 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.45 (s, 1H), 8.62–8.59 (d, J = 6.4 Hz, 1H), 8.03–8 (d, J = 7.7 Hz, 1H), 7.9– 7.87 (d, J = 1 Hz, 1H), 7.56–7.53 (m, 2H), 7.46–7.42 (m, 4H), 7.34–7.29 (m, 1H), 7.21–7.16 (m, 2H), 5.36 (s, 2H), 3.66 (s, 3H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 166.3, 156.4, 150.9, 143.8, 135.7, 134.2, 131.6, 130.5, 128.9, 128.5, 128.0, 124.5, 124.3, 121.9, 121.7, 118.0, 113.4, 113.0, 71.3, 45.1. IR (KBr): 3437, 1719, 1598, 1461, 1286, 994, 746 cm⁻¹. MS m/z: 359 (M⁺+1, 7), 358 (M⁺, 33), 252 (14), 207 (11), 91 (100). HRMS-EI (m/z): [M]⁺ calcd for C₂₂H₁₈N₂O₃, 358.1317; found, 358.1324.

4.1.1.12. 2-(2-Benzyloxy-phenyl)-1H-benzoimidazole-4-carboxylic acid (13). To a solution of compound 12 (0.37 g, 1.0 mmol) in tetrahydrofuran (7.3 mL) was added 5 M sodium hydroxide solution (4.5 mL), and refluxed for 12 h. After cooled to room temperature, the reaction solution was acidified by concd hydrochloric acid to give a white precipitate. The precipitate was filtered and washed with water, and dried under vacuum to produce compound 13 (0.34 g, 1.0 mmol). Yield: 97%. mp: 180–182 °C. ¹H NMR (300 MHz, CD₃OD, δ): 8.22–8.19 (d, J = 7.6 Hz, 1H), 8.07–8.05 (d, J = 8 Hz, 2H), 7.79–7.07 (m, 2H), 7.51–7.48 (m, 3H), (m, 4H), 5.46 (s, 2H). ¹³C NMR 7.36-7.29 (125.7 MHz, CDCl₃, δ): 167.1, 158.9, 150.0, 137.0, 136.8, 133.5, 131.8, 131.5, 129.8, 129.5, 129.2, 128.9, 127.3, 123.1, 119.8, 118.4, 115.7, 112.5, 72.8. IR (KBr): 3351, 2607, 1697, 1128, 848, 622 cm^{-1} . MS *m/z*: 345 $(M^++1, 8), 344 (M^+, 39), 298 (8), 238 (8), 236 (9), 91$ (100). HRMS-EI (m/z): [M]⁺ calcd for C₂₁H₁₆N₂O₃, 344.1161; found, 344.1159.

4.1.1.13. 2-(2-Hydroxy-phenyl)-1H-benzoimidazole-4carboxylic acid methyl ester (14). A mixture of compound 12 (0.21 g, 0.6 mmol) and Pd/C (0.02 g, 10% of Pd on charcoal) in ethyl acetate (10 mL) under hydrogen atmosphere (60 psi) was hydrogenated for 2 h. The catalyst was removed by filtration, and the filtrate was concentrated to give compound 14 (0.15 g, 0.6 mmol). Yield: 97%. mp: 205–207 °C. ¹H NMR (300 MHz, CD_3Cl_3 , δ): 10.72 (s, 1H), 7.96–7.91 (dd, J = 8.2, 7.8 Hz, 2H), 7.70–7.67 (d, J = 7.8 Hz, 1H), 7.43–7.32 (m, 2H), 7.15-7.12 (d, J = 8.1 Hz, 1H), 7.02-6.97 (dd, ¹³C NMR J = 7.8, 7.3 Hz, 1H), 4.04 (s, 3H). (75.4 MHz, CDCl₃, δ): 167, 159.1, 152.2, 142.4, 133.4, 125.0, 123.9, 122.3, 119.2, 118.1, 113.2, 111.7, 52.3. IR (KBr): 3394, 1702, 1463, 1284, 750 cm⁻¹. MS *m*/*z*: 269 $(M^++1, 8), 268 (M^+, 49), 237 (17), 236 (100), 208 (18).$ HRMS-EI (m/z): $[M]^+$ calcd for $C_{15}H_{12}N_2O_3$, 268.0848; found, 268.0843.

4.1.1.14. 2-(2-Hydroxy-phenyl)-1H-benzoimidazole-4carboxylic acid (15). A mixture of compound 13 (0.20 g, 0.6 mmol) and Pd/C (0.02 g, 10% of Pd on charcoal) in methanol (15 mL) under hydrogen atmosphere (60 psi) was hydrogenated for 2 h. The catalyst was removed by filtration, and the filtrate was concentrated, and the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using methanol/ethyl acetate (1:4) as the mobile phase producing compound 15 (0.09 g, 0.4 mmol). Yield: 60%. mp: 310 °C (decomposed). ^TH NMR (300 MHz, DMSO- d_6 , δ): 8.26 (b, 1H), 7.76–7.74 (b, 2H), 7.34– 7.28 (b, 1H), 7.24–7.16 (b, 2H), 6.91 (b, 1H). ¹³C NMR (125.7 MHz, DMSO, δ): 169.1, 157.6, 151.5, 141.2, 134.0, 131.6, 127.7, 124.2, 121.5, 120.3, 119.0, 117.1, 113.5. IR (KBr): 3396, 1585, 1488, 1384, 1292, 1257, 748 cm⁻¹. MS m/z: 255 (M⁺+1, 6), 254 (M⁺, 43), 237 (15), 236 (100), 208 (20). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₁₀N₂O₃, 254.0691; found, 254.0684.

4.1.1.15. 2-Amino-3-{[2-(2-benzyloxy-phenyl)-1*H*-benzoimidazole-4-carbonyl]-amino}-benzoic acid methyl ester (16). A solution of compound 13 (0.34 g, 1.0 mmol) in

thionyl chloride (2 mL) was refluxed for 1 h, and the thionyl chloride was removed by distillation. The remained residue was dissolved in anhydrous dichloromethane, to this solution was added a solution of compound 6 (0.17 g, 1.0 mmol), triethylamine (1.0 mL) in anhydrous dichloromethane (5 mL) by canula. The resulting solution was stirred at room temperature overnight and quenched by adding water. The aqueous solution was extracted with dichloromethane, and the extract was dried over anhydrous magnesium sulfate. The organic solution was concentrated, and the residue was purified by flash column chromatography using silica gel as the stationary phase and using methanol/ethyl acetate (1:1, 3:1) as the mobile phase producing compound 16 (0.27 g, 0.6 mmol). Yield: 56%. mp: 178-181 °C. ¹H NMR (300 MHz, CD₃OD, δ): 11.68 (b, 1H), 10.84 (b, 1H), 8.63–8.60 (d, J = 9.0 Hz, 1H), 8.21-8.18 (d, J = 7.3 Hz, 1H), 8.07-8.05 (d, J = 7.4 Hz, 1H). 7.82–7.79 (d. J = 8.0 Hz. 1H). 7.52–7.14 (m. 10H), 6.81-6.76 (dd, J = 7.4, 7.3 Hz, 1H), 6.32 (b, 2H), 5.32 (s, 2H), 3.91 (s, 3H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 168.8, 164.3, 156.4, 150.2, 144.4, 140.4, 135.7, 133.6, 132.1, 130.4, 129.24, 129.21, 129.0, 127.9, 127.8, 125.6, 124.3, 122.7, 122.4, 122.3, 117.0, 116.0, 114.6, 113.0, 111.9, 71.5, 51.6. IR (KBr): 3392, 1704, 1673, 1292, 1249, 742 cm⁻¹. MS m/z: 493 (M⁺+1, 18), 492 (M⁺, 54), 475 (44), 474 (42), 327 (57). HRMS-EI (*m*/*z*): [M]⁺ calcd for C₂₉H₂₄N₄O₄, 492.1798; found, 492.1792.

4.1.1.16. 2'-(2-Benzyloxy-phenyl)-1H,1'H-[2,4']bibenzoimidazolyl-4-carboxylic acid methyl ester (17). A solution of compound 16 (0.17 g, 0.4 mmol) in acetic acid (3 mL) was refluxed at 120-130 °C for 2 h. After cooled to room temperature, the reaction solution was poured into ice water and neutralized by sodium bicarbonate. The aqueous solution was extracted with dichloromethane, and the extract was washed with brine and dried over anhydrous magnesium sulfate. The dry solution was concentrated and dried under vacuum to give compound 17 (0.14 g, 0.3 mmol). Yield: 85%. mp: 202-206 °C. ¹H NMR (300 MHz, CDCl₃, δ): 13.66 (s, 1H), 10.74 (s, 1H), 9.01–8.98 (d, J = 7.8 Hz, 1H), 8.40–8.37 (m, 1H), 8.07-8.04 (d, J = 8.0 Hz, 1H), 7.99-7.96 (d, J = 7.7 Hz, 1H), 7.54–7.49 (m, 6H), 7.37–7.31 (m, 4H), 7.22–7.19 (d, J = 8.8 Hz, 1H), 5.36 (s, 2H), 4.18 (s, 3H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 166.6, 156.2, 152.4, 150.2, 144.6, 140.5, 135.8, 134.3, 133.8, 131.7, 130.4, 129.1, 128.9, 127.8, 124.7, 124.2, 122.9, 121.8, 121.4, 120.8, 118.5, 117.3, 113.6, 112.6, 112.5, 71.2, 52.0. IR (KBr): 3413, 1720, 1432, 1280, 744 cm⁻¹. MS m/z: 475 (M⁺+1, 33), 474 (M⁺, 100), 473 (20), 352 (25), 351 (33). HRMS-EI (m/z): $[M]^+$ calcd for C₂₉H₂₂N₄O₃, 474.1692; found, 474.1684.

4.1.1.17. 2'-(2-Hydroxy-phenyl)-1*H*,1'*H*-[**2**,4']bibenzoimidazolyl-4-carboxylic acid methyl ester (**18**). A mixture of compound **17** (0.05 g, 0.1 mmol) and Pd/C (2.5 mg, 10% of Pd on charcoal) in methanol (25 mL) under hydrogen atmosphere (1 atm) was hydrogenated overnight. The catalyst was removed by filtration, and the filtrate was concentrated to give compound **18** (0.03 g, 0.07 mmol). Yield: 66%. mp: 283–286 °C. ¹H NMR (300 MHz, CDCl₃, δ): 12.0 (s, 1H), 10.88 (s, 1H), 8.12–8.09 (d, J = 8.1 Hz, 1H), 7.97–7.95 (d, J = 7.9 Hz, 1H), 7.89–7.86 (m, 2H), 7.77–7.74 (d, J = 7.8 Hz, 1H), 7.46–7.37 (m, 3H), 7.17–7.15 (d, J = 7.5 Hz, 1H), 7.09–7.04 (dd, J = 7.8, 7.4 Hz, 1H), 4.06 (s, 3H). ¹³C NMR (125.7 MHz, DMSO- d_6 , δ): 165.9, 156.2, 152.0, 151.6, 144.3, 140.1, 135.8, 133.8, 131.9, 129.1, 124.2, 124.1, 122.4, 121.7, 120.1, 119.6, 116.8, 115.0, 113.3, 52.3. IR (KBr): 3486, 3378, 3189, 3068, 2977, 2817, 1675, 1608, 1583, 784 cm⁻¹. MS *m*/*z*: 385 (M⁺+1, 25), 384 (100), 353 (24), 352 (93), 162 (13). HRMS-EI (*m*/*z*): [M]⁺ calcd for C₂₂H₁₆N₄O₃, 384.1222; found, 384.1219.

4.1.1.18. 2'-(2-Benzyloxy-phenyl)-1H,1'H-[2,4']bibenzoimidazolyl-4-carboxylic acid (19). To a solution of compound 17 (0.05 g, 0.1 mmol) in tetrahydrofuran (2 mL) was added 5 M sodium hydroxide solution (3 mL), and refluxed for 48 h. After cooled to room temperature, the reaction solution was acidified by concentrated hydrochloric acid to give a white precipitate. The precipitate was filtered and washed with water, and dried under vacuum to produce compound 19 (0.08 g, 0.2 mmol). Yield: 100%. mp: 256 °C (decomposed). 1 H (300 MHz, CD_3OD , δ): 8.55-8.53 NMR (d. J = 7.5 Hz, 1H), 8.02–7.99 (d, J = 7.5 Hz, 2H), 7.88– 7.85 (d, J = 9.0 Hz, 1H), 7.78–7.76 (d, J = 6.0 Hz, 6H), 7.56-7.28 (m, 8H), 7.16-7.13 (d, J = 8.4 Hz, 1H), 7.06-7.037.02 (dd, J = 7.5, 6.0 Hz, 1H), 5.33 (s, 2H). ¹³C NMR (125.7 MHz, *d*-DMSO, δ): 166.6, 155.9, 151.1, 149.8, 140.2, 136.9, 135.4, 132.4, 131.7, 130.3, 128.5, 127.7, 127.1, 126.2, 124.3, 123.0, 121.6, 120.7, 116.9, 115.8, 114.1, 69.1. IR (KBr): 3390, 3200, 3095, 3050, 1625, 1575, 1550, 1095, 750 cm⁻¹. MS m/z: 460 (M⁺, 1.78), 418 (45), 253 (14), 252 (100), 91 (26). HRMS-EI (m/z): $[M]^+$ calcd for C₂₈H₂₀N₄O₃, 460.1535; found, 460.1541.

4.1.1.19. 2-Amino-3-hydroxy-benzoic acid methyl ester (20). To a solution of 2-amino-3-hydroxybenzoic acid (0.15 g, 0.9 mmol) in diethyl ether (10.0 mL) and methanol (2.0 mL) was added a solution of diazomethane in diethyl ether at 0 °C until ceased to generate nitrogen. After adding 1 mL of acetic acid, the reaction solution was concentrated and the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:9) as the mobile phase producing compound 20 (0.15 g, 0.8 mmol). Yield: 92%. mp: 94–97 °C. ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3, \delta)$: 7.50–7.47 (d, J = 8.7 Hz, 1H), 6.83-6.81 (d, J = 6.6 Hz, 1H), 6.52-6.47 (dd, J = 8.0, 7.9 Hz, 1H), 3.87 (s, 3H). ¹³C NMR (125.7 MHz, $CDCl_3$, δ): 168.7, 143.1, 140.5, 123.1, 117.9, 115.0, 111.3, 51.5. IR (KBr): 3409, 1704, 1439, 1293, 743 cm⁻¹. MS m/z: 167 (M⁺, 100), 136 (26), 135 (79), 108 (12), 107 (85). HRMS-EI (m/z): $[M]^+$ calcd for C₈H₉NO₃, 167.0582; found, 167.0589.

4.1.1.20. 2-(2-Benzyloxy-benzoylamino)-3-hydroxybenzoic acid methyl ester (22). A solution of compound 10 (0.28 g, 1.2 mmol) in anhydrous dichloromethane (10 mL) was added oxalyl chloride (0.22 mL, 2.5 mmol), and stirred for 10 min at room temperature; following added 5 drops of N,N-dimethylformamide and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (10 mL). To this solution was added a solution of compound 20 (0.15 g, 0.8 mmol) and pyridine (2 mL) in anhydrous dichloromethane (10 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:19, 1:9) as the mobile phase producing compound **22** (0.30 g, 0.8 mmol). Yield: 96%. mp: 101-102 °C. ¹H NMR (300 MHz, CDCl₃, δ): 12.03 (s, 1H), 9.39 (s, 1H), 8.26–8.23 (d, J = 7.9 Hz, 1H), 7.60–7.57 (d, J = 7.7 Hz, 1H), 7.47– 7.40 (m, 3H), 7.36-7.28 (m, 4H), 7.23-7.18 (m, 1H), 7.11–7.05 (dd, J = 7.9, 7.9 Hz, 1H), 7.04–7.01 (d, J = 8.3 Hz, 1H), 5.47 (s, 2H), 3.78 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 167.5, 165.5, 157.0, 150.7, 136.2, 133.9, 132.7, 128.6, 128.2, 128.0, 126.9, 126.0, 125.7, 122.9, 121.5, 121.3, 120.9, 113.3, 70.8, 52.2. IR (KBr): 1710, 1627, 1455, 1288, 989, 751 cm⁻¹. MS *m*/*z*: 377 (M⁺, 22), 211 (74), 166 (22), 121 (17), 91 (100). HRMS-EI (m/z): $[M]^+$ calcd for $C_{22}H_{19}NO_5$, 377.1263; found, 377.1257.

4.1.1.21. 2-(2-Benzyloxy-phenyl)-benzoxazole-4-carboxylic acid methyl ester (23). Compound **22** (0.11 g, 0.3 mmol) was heated up to 230 °C under vacuum (ca. 40 torr) for 5 h, and purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49, 1:19) as the mobile phase producing compound **23** (0.09 g, 0.3 mmol). Yield: 86%. mp: 104–105 °C. ¹H NMR (300 MHz, CDCl₃, δ): 8.28–8.26 (d, J = 7.9 Hz, 1H), 8.06–8.03 (d, J = 7.8 Hz, 1H), 7.77–7.75 (d, J = 7.6 Hz, 1H), 7.67–7.64 (d, J = 7.2 Hz, 2H), 7.54–7.25 (m, 5H), 7.15–7.10 (m, 2H), 5.31 (s, 2H), 4.01 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 165.9, 163.6, 157.7, 151.2, 141.3, 136.6, 133.2, 131.9, 128.4, 127.7, 126.87, 126.83, 124.1, 121.9, 121.0, 116.2, 114.7, 113.6, 70.6, 52.4. IR (KBr): 1703, 1601, 1450, 1240, 752 cm⁻¹. MS *m/z*: 360 (M⁺+1, 19), 359 (M⁺, 80), 342 (15), 327 (16), 91 (100). HRMS-EI (*m/z*): [M]⁺ calcd for C₂₂H₁₇NO₄, 359.1158; found, 359.1165.

4.1.1.22. 2-(2-Benzyloxy-phenyl)-benzoxazole-4-carboxylic acid (24). To a solution of compound 23 (0.09 g, 0.3 mmol) in tetrahydrofuran (2 mL) was added 5 N sodium hydroxide solution (1 mL), and refluxed for 2 h. After cooled to room temperature, the reaction solution was acidified by concd hydrochloric acid and extracted with ethyl acetate. The extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was dried under vacuum to give product 24 (0.07 g, 0.2 mmol). Yield: 84%. mp: 99-102 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.75 (s, 1H), 8.25-8.22 (d, J = 7.9 Hz, 1H), 8.15-8.12 (d, J = 7.8 Hz, 1H), 7.80–7.77 (d, J = 8.1 Hz, 1H), 7.61–7.34 (m, 7H), 7.21–7.14 (m, 2H), 5.34 (s, 2H). ¹³C NMR $(125.7 \text{ MHz}, \text{ CDCl}_3, \delta)$: 165.1, 163.0, 158.1, 149.8, 141.0, 136.0, 134.3, 131.6, 128.8, 128.2, 127.2, 126.9, 126.2, 125.3, 121.1, 119.9, 115.2, 114.4, 113.6, 70.8. IR (KBr): 1745, 1604, 1430, 1251, 1020, 746 cm⁻¹. MS *m*/*z*: 346 (M⁺+1, 13), 345 (M⁺, 59), 328 (9), 237 (10), 91 (100). HRMS-EI (*m*/*z*): [M]⁺ calcd for $C_{21}H_{15}NO_4$, 345.1001; found, 345.0995.

4.1.1.23. 2-(2-Hvdroxy-phenyl)-benzoxazole-4-carboxylic acid methyl ester (25). A solution of compound 23 (0.05 g, 0.1 mmol) and *p*-toluenesulfonic acid (0.06 g, 0.1 mmol)0.3 mmol) in anhydrous toluene (2 mL) was refluxed for 1.5 h, and the reaction was guenched by adding saturated sodium bicarbonate solution. The aqueous phase was extracted with ethyl acetate, and the extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49, 1:19) as the mobile phase producing compound 25 (0.02 g, 0.07 mmol). Yield: 51%. mp: 131-135 °C. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3, \delta)$: 11.9 (s, 1H), 8.09–8.02 (m, 2H), 7.82-7.79 (d. J = 8.0 Hz. 1H), 7.51-7.43 (m. 2H), 7.16-7.437.13 (d, J = 7.9 Hz, 1H), 7.05–7.0 (dd, J = 7.9, 7.6 Hz, 1H), 4.06 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 165.5, 164.3, 159.3, 149.7, 139.4, 134.2, 127.4, 127.2, 124.7, 121.2, 119.2, 117.6, 114.9, 109.9, 52.4. IR (KBr): 1717, 1633, 1548, 1301, 747 cm⁻¹. MS m/z: 269 (M⁺ 62), 238 (23), 237 (100), 211 (36), 209 (15), 133 (13). HRMS-EI (m/z): [M]⁺ calcd for C₁₅H₁₁NO₄, 269.0688; found, 269.0684.

4.1.1.24. 2-{[2-(2-Benzyloxy-phenyl)-benzoxazole-4carbonyl]-amino}-3-hydroxy-benzoic acid methyl ester (26). A solution of compound 24 (0.07 g, 0.2 mmol) in anhydrous dichloromethane (10 mL) was added oxalvl chloride (0.15 mL, 1.7 mmol), and stirred for 10 minutes at room temperature; following added 5 drops of N,N-dimethylformamide, and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (15 mL). To this solution was added a solution of compound 20 (0.04 g, 0.2 mmol) and pyridine (0.5 mL) in anhydrous dichloromethane (6 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:9) as the mobile phase producing compound 26 (0.08 g, 0.2 mmol). Yield: 76%. mp: 155–158 °C. ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3, \delta)$: 12.57 (s, 1H), 8.80 (s, 1H), 8.49-8.46 (d, J = 7.7 Hz, 1H), 8.30–8.27 (d. J = 7.8 Hz, 1H), 7.80–7.77 (d, J = 8.1 Hz, 1H), 7.64– 7.61 (d, J = 7.7 Hz, 1H), 7.53–7.50 (m, 4H), 7.37– 7.27 (m, 4H), 7.23–7.21 (m, 1H), 7.19–7.10 (m, 1H), 5.24 (s, 2H), 3.77 (s, 3H). ¹³C NMR (125.7 MHz, $CDCl_3$, δ): 167.1, 164.4, 163.4, 157.9, 151.2, 150.8, 139.9, 136.2, 133.6, 132.3, 128.3, 127.8, 127.6, 126.7, 126.5, 126.3, 125.6, 124.7, 123.6, 123.0, 122.5, 121.2, 115.7, 114.7, 113.7, 70.7, 52.2. IR (KBr): 3208, 1713, 1527, 1458, 1291, 750 cm⁻¹. MS m/z: 494 (M⁺, 39), 328 (100), 238 (48), 237 (31), 91 (25). HRMS-EI (m/z): $[M]^+$ calcd for C₂₉H₂₂N₂O₆, 494.1478; found, 494.1482.

4.1.1.25. 2'-(2-Hydroxy-phenyl)-[2,4']bibenzoxazolyl-4-carboxylic acid methyl ester (UK-1). A solution of compound **26** (0.06 g, 0.1 mmol) and *p*-toluenesulfonic acid (0.06 g, 0.3 mmol) in anhydrous p-xylene (2 mL) was refluxed for 2 h, and the reaction was guenched by adding saturated sodium bicarbonate solution. The aqueous phase was extracted with ethyl acetate, and the extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:19, 1:9) as the mobile phase producing UK-1 (0.02 g, 0.1 mmol). Yield: 43%. mp: 209–210 °C. ¹H NMR (300 MHz, CDCl₃, δ): 12.06 (s, 1H), 8.40-8.37 (d, J = 7.7 Hz, 1H), 8.13–8.11 (d, J = 7.3 Hz, 1H), 8.10-8.07 (d, J = 7.9 Hz, 1H), 7.90-7.88 (d, J = 8.0 Hz, 1H), 7.83-7.80 (d, J = 8.2 Hz, 1H), 7.58-7.46 (m, 3H), 7.21–7.18 (d, J = 8.3 Hz, 1H), 7.05–7.0 (t, J = 7.7, 7.2 Hz, 1H), 4.13 (s, 3H). ¹³C NMR (125.7 MHz, $CDCl_3$, δ): 166.1, 164.5, 161.8, 159.5, 151.2, 149.8, 141.4, 138.6, 134.2, 127.4, 127.1, 125.28, 125.25, 124.8, 122.3, 119.5, 117.7, 117.5, 115.0, 113.8, 109.9, 52.6. IR (KBr): 2923, 1724, 1546, 1427, 1259, 744 cm⁻¹. MS m/ z: 387 (M⁺+1, 24), 386 (M⁺, 100), 355 (26), 354 (83), 328 (28). HRMS-EI (m/z): [M]⁺ calcd for C₂₂H₁₄N₂O₅, 386.0903; found, 386.0904.

4.1.1.26. 2-Methoxy-benzoic acid methyl ester (27). To a solution of salicylic acid 7 (2.20 g, 16.0 mmol) in anhydrous acetone (30 mL) were added potassium carbonate (5.53 g, 40.0 mmol) and iodomethane (2.18 mL, 35.0 mmol), and refluxed overnight. After cooled to room temperature, the reaction mixture was filtered through Celite, and the filtrate was concentrated. The residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:19, 1:9) as the mobile phase producing compound 27 (2.40 g, 14.0 mmol). Yield: 91%. ¹H NMR (300 MHz, $CDCl_3$, δ): 7.81– 7.78 (d, J = 7.9 Hz, 1H), 7.50–7.44 (m, 1H), 7.0–6.95 (m, 2H), 3.91 (s, 3H), 3.89 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 166.6, 159.0, 133.5, 131.6, 120.0, 119.9, 111.9, 55.9, 51.9. IR (KBr): 1728, 1492, 1254, 1085, 756 cm⁻¹. MS m/z: 166 (M⁺, 38), 135 (100), 133 (33), 105 (14), 92 (16), 77 (41). HRMS-EI (m/z): $[M]^+$ calcd for C₉H₁₀O₃, 166.0630; found, 166.0632.

4.1.1.27. 2-Methoxy-benzoic acid (28). To a solution of compound **27** (2.40 g, 14.0 mmol) in methanol (15 mL) was added 6 M sodium hydroxide solution (10 mL), and refluxed for 3 h. After concentration, the remained concentrated solution was acidified by adding 6 N hydrochloric acid to give a precipitate. The precipitate was filtered and dried under vacuum to produce compound **28** (1.71 g, 11.0 mmol). Yield: 56%. mp: 108–111 °C. ¹H NMR (300 MHz, CDCl₃, δ): 10.97 (s, 1H), 8.21–8.17 (d, J = 7.8 Hz, 1H), 7.61–7.55 (dd, J = 8.6, 7.2 Hz, 1H), 7.17–7.12 (dd, J = 7.4, 7.1 Hz, 1H), 7.08–7.05 (d, J = 8.4 Hz, 1H), 4.08 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 165.3, 158.0, 135.0, 133.8, 122.2, 117.5, 111.5, 56.6. IR (KBr): 1692, 1492, 1255, 1087, 1019, 761 cm⁻¹. MS *m/z*: 152 (M⁺, 100),

135 (29), 123 (51), 105 (78), 79 (19), 77 (33). HRMS-EI (m/z): $[M]^+$ calcd for $C_8H_8O_3$, 152.0473; found, 152.0471.

4.1.1.28. 2-Hydroxy-3-(2-methoxy-benzoylamino)benzoic acid methyl ester (30). A solution of compound 28 (0.15 g, 1.0 mmol) in anhydrous dichloromethane (15 mL) was added oxalyl chloride (0.26 mL, 3.0 mmol), and stirred for 10 minutes at room temperature; following added 5 drops of N,N-dimethylformamide, and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (7 mL). To this solution was added a solution of compound 29 (0.15 g, 0.9 mmol) and pyridine (1 mL) in anhydrous dichloromethane (10 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (3:17, 3:7) as the mobile phase producing compound **30** (0.20 g, 0.7 mmol). Yield: 74%. mp: 166–168 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.35 (s, 1H), 10.77 (s, 1H), 8.86-8.83 (d, J = 8.0 Hz, 1H), 8.32–8.29 (d, J = 7.9 Hz, 1H), 7.58–7.54 (d, J = 8.3 Hz, 1H), 7.53–7.47 (m, 1H), 7.16–7.10 (dd, J = 7.5, 7.5 Hz, 1H), 7.06–7.03 (d, J = 8.3 Hz, 1H), 6.96–6.91 (dd, J = 8.3, 8.1 Hz, 1H), 4.09 (s, 3H), 3.98 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 171.0, 163.2, 157.4, 150.9, 133.2, 132.3, 128.3, 125.5, 123.4, 121.7, 121.4, 119.2, 111.5, 56.1, 52.4. IR (KBr): 1665, 1536, 1344, 1286, 756 cm⁻¹. MS *m*/*z*: 302 (M⁺+1, 5), 301 (M⁺, 31), 269 (6), 136 (8), 135 (100). HRMS-EI (m/z): $[M]^+$ calcd for C₁₆H₁₅NO₅, 301.0950; found, 301.0946.

4.1.1.29. 2-(2-Methoxy-phenyl)-benzoxazole-7-carboxylic acid methyl ester (31) and 2-(2-hydroxy-phenyl)benzoxazole-7-carboxylic acid methyl ester (32). A solution of compound 30 (0.24 g, 0.8 mmol) and *p*-toluenesulfonic acid (0.38 g, 2.0 mmol) in anhydrous *p*-xylene (9.7 mL) was refluxed for 4 days, and the reaction was quenched by adding saturated sodium bicarbonate solution. The aqueous phase was extracted with ethyl acetate, and the extract was washed with brine and dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was dried under vacuum to produce compound 31 (0.11 g, 0.07 mmol) and compound 32 (20 mg, 0.07 mmol).

4.1.1.30. Compound 31. Yield: 32%. mp: 88–90 °C. ¹H NMR (300 MHz, CDCl₃, δ): 8.26–8.23 (d, J = 7.7 Hz, 1H), 8.05–7.99 (m, 2H), 7.57–7.52 (m, 1H), 7.44–7.39 (dd, J = 7.9, 7.9 Hz, 1H), 7.19–7.10 (m, 1H), 4.07 (s, 3H), 4.05 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 164.7, 162.3, 158.7, 149.0, 143.3, 133.3, 131.4, 127.0, 124.9, 124.1, 120.8, 115.3, 114.6, 112.1, 56.2, 52.3. IR (KBr): 1716, 1621, 1428, 1301, 1037, 740 cm⁻¹. MS *m*/*z*: 283 (M⁺, 70), 252 (21), 251 (37), 224 (17), 223 (100), 167 (16). HRMS-EI (*m*/*z*): [M]⁺ calcd for C₁₆H₁₃NO₄, 283.0845; found, 283.0852.

4.1.1.31. Compound 32. Yield: 9%. mp: 137–139 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.28 (s, 1H), 8.16– 8.13 (d, J = 7.8 Hz, 1H), 8.04–8.01 (d, J = 7.8 Hz, 1H), 7.94–7.91 (d, J = 7.9 Hz, 1H), 7.50–7.43 (m, 2H), 7.15– 7.12 (d, J = 8.4 Hz, 1H), 7.07–7.02 (dd, J = 7.9, 7.6 Hz, 1H), 4.07 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 164.4, 163.7, 158.7, 148.1, 141.4, 134.0, 127.5, 127.3, 124.7, 123.8, 119.7, 117.4, 115.0, 110.0, 52.5. IR (KBr): 1728, 1631, 1290, 1060, 754 cm⁻¹. MS *m/z*: 270 (M⁺+1, 12), 269 (M⁺, 73), 238 (24), 237 (100), 209 (9). HRMS-EI (*m/z*): [M]⁺ calcd for C₁₅H₁₁NO₄, 269.0688; found, 269.0680.

2-(2-Methoxy-phenyl)-benzoxazole-7-car-4.1.1.32. boxylic acid (33). To a solution of compound 31 (52 mg, 0.18 mmol) in tetrahydrofuran (3 mL) was added 5 N sodium hydroxide solution (0.73 mL), and refluxed for 2 days. After cooled to room temperature, the reaction solution was acidified by concd hydrochloric acid to give a white precipitate. The precipitate was filtered and washed with water, and dried under vacuum to produce compound 33 (45 mg, 0.17 mmol). Yield: 91%. mp: 275 °C (decomposed). ^TH NMR (300 MHz, CD₃OD, δ): 8.32–8.28 (dd, J = 7.8, 7.8 Hz, 1H), 7.86– 7.83 (d, J = 7.7 Hz, 1H), 7.79–7.76 (d, J = 7.9 Hz, 1H), 7.60–7.54 (dd, J = 7.8, 7.7 Hz,1H), 7.40–7.35 (dd, J = 7.8, 7.8 Hz, 1H), 7.24–7.21 (d, J = 7.9 Hz, 1H), 7.16–7.10 (m, 1H), 4.0 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 170.7, 161.9, 158.7, 148.4, 142.2, 133.1, 131.0, 126.5, 123.7, 123.2, 120.6, 120.3, 115.1, 111.9, 54.9. IR (KBr): 3432, 1581, 1400, 1124, 763, 617 cm⁻¹ MS m/z: 269 (M⁺, 4) 175 (100), 154 (50), 137 (25), 136 (34), 57 (22). HRMS-FAB (m/z): $[M + H]^+$ calcd for C₁₅H₁₁NO₄, 270.0766; found, 270.0775.

2-Hydroxy-3-{[2-(2-methoxy-phenyl)-benz-4.1.1.33. oxazole-7-carbonyl]-amino}-benzoic acid methyl ester (34). A solution of compound 33 (23 mg, 0.09 mmol) in anhydrous dichloromethane (7 mL) was added oxalyl chloride (22 uL, 0.26 mmol), and stirred for 10 min at room temperature; following added 5 drops of N,N-dimethylformamide, and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (10 mL). To this solution was added a solution of compound 29 (18 mg, 0.1 mmol) and pyridine (0.1 mL) in anhydrous dichloromethane (5 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/ hexane (1:4, 3:7) as the mobile phase producing compound 34 (30 mg, 0.07 mmol). Yield: 67%. mp: 235-238 °C. ¹H NMR (300 MHz, CDCl₃, δ): 11.61 (s, 1H), 9.89 (s, 1H), 8.90–8.87 (d, J = 7.8 Hz, 1H), 8.43–8.40 (d, J = 7.9 Hz, 1H), 8.22–8.20 (d, J = 7.8 Hz, 1H), 8.03-8.01 (d, J = 7.8 Hz, 1H), 7.66-7.51 (m, 3H), 7.22-7.12 (m, 2H), 7.03–6.98 (dd, J = 7.8, 7.8 Hz, 1H), 4.05 (s, 3H), 4.01 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 171.0, 161.1, 158.7, 150.8, 146.6, 142.6, 133.5, 131.4, 127.7, 126.7, 125.7, 124.8, 124.3, 124.0, 120.7, 119.4, 117.4, 114.8, 112.0, 111.7, 56.1, 52.5. IR (KBr): 1670, 1548, 1349, 1253, 765 cm⁻¹. MS m/z: 419 (M⁺+1, 13), 418 (M⁺, 49), 386 (11), 253 (16), 252 (100). HRMS-EI (m/z): [M]⁺ calcd for C₂₃H₁₈N₂O₆, 418.1165; found, 418.1167.

4.1.1.34. 2'-(2-Methoxy-phenyl)-[2,7']bibenzoxazolyl-7-carboxylic acid methyl ester (35). A solution of compound 34 (0.14 g, 0.33 mmol) and p-toluenesulfonic acid (0.16 g, 0.84 mmol) in anhydrous p-xylene (7.5 mL) was refluxed for 32.5 h, and the reaction was guenched by adding saturated sodium bicarbonate solution. The aqueous phase was extracted with ethyl acetate, and the extract was washed with brine and dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:9, 3:7, 1:1) as the mobile phase producing compound 35 (40 mg, 0.1 mmol). Yield: 30%. mp: 164–166 °C. ¹H NMR (300 MHz. $CDCl_3$, δ): 8.51–8.49 (d, J = 6.1 Hz, 1H), 8.32–8.29 (d, J = 8.1 Hz, 1H), 8.09–8.06 (m, 3H), 7.61–7.46 (m, 3H), 7.22–7.13 (m, 2H), 4.12 (s, 3H), 4.09 (s, 3H). ¹³C NMR (75 MHz, CDCl₃, δ): 164.6, 162.4, 160.8, 158.8, 150.8, 149.6, 147.7, 143.8, 143.4, 133.2, 131.4, 127.5, 125.0, 124.9, 124.6, 123.8, 120.7, 115.5, 115.0, 112.1, 111.0, 56.3, 52.4. IR (KBr): 3397 (br), 3079, 3020, 2916, 2852, 1731, 1617, 1542, 1291, 1431, 746 cm⁻ MS m/z: 401 (M⁺+1, 26), 154 (35), 81 (39), 69 (60), 57 (74), 55 (100). HRMS-FAB (m/z): $[M + H]^+$ calcd for C₂₃H₁₆N₂O₅, 401.1137; found, 401.1142.

4.1.1.35. 2-(2-Benzyloxy-benzoylamino)-benzoic acid methyl ester (36). A solution of compound 10 (2.00 g. 8.8 mmol) in anhydrous dichloromethane (60 mL) was added oxalyl chloride (3.8 mL, 43.6 mmol), and stirred for 10 min at room temperature; following added N,Ndimethylformamide (0.1 mL), and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (50 mL). To this solution was added a solution of compound 2 (1.32 g, 8.8 mmol) and pyridine (3 mL) in anhydrous dichloromethane (40 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/ hexane (1:49) as the mobile phase producing compound **36** (2.55 g, 7.0 mmol). Yield: 81%. mp: 116-119 °C. ¹H NMR (300 MHz, CDCl₃, δ) : 12.7 (s, 1H), 8.95–8.92 (d, J = 8.6 Hz, 1H), 8.13-8.11 (d, J = 7.8 Hz, 1H), 8.02-7.99 (d, J = 8 Hz, 1H), 7.45-7.42 (dd, J = 7.3, 6.4 Hz, 1H), 7.45–7.28 (m, 6H), 7.13–7.03 (m, 2H), 7.0–6.97 (d, J = 8.3 Hz, 1H), 5.43 (s, 2H), 3.73 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 167.6, 164.7, 156.3, 141.2, 136.7, 134.1, 132.8, 132.1, 130.7, 128.5, 127.7, 126.8, 123.7, 122.5, 121.8, 121.2, 116.5, 113.3, 70.4, 51.9. IR (KBr): 1710, 1586, 1450, 1265, 757 cm⁻¹. MS m/z: 361 (M⁺, 5), 241 (16), 240 (19), 151 (29), 92 (14), 91 (100). HRMS-EI (m/z): [M]⁺ calcd for C₂₂H₁₉NO₄, 361.1314; found, 361.1308.

4.1.1.36. 2-(2-Benzyloxy-thiobenzoylamino)-benzoic acid methyl ester (37). To a solution of compound 36 (0.36 g, 1.0 mmol) in anhydrous toluene (4 mL) was added the Lawesson's reagent (0.40 g, 1.0 mmol) in glove box. The resulting solution was refluxed for 7.5 h under argon atmosphere, and the reaction was quenched by adding water. The aqueous solution was extracted with ethyl acetate, and the extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49) as the mobile phase producing compound 37 (0.19 g, 0.5 mmol). Yield: 51%. mp: 74–76 °C. ¹H NMR (300 MHz, CDCl₃, δ): 12.71 (b, 1H), 9.59–9.56 (d, J = 8.2 Hz, 1H), 8.04– 8.01 (d, J = 7.6 Hz, 1H), 7.96–7.94 (d, J = 7.2 Hz, 1H), 7.65-7.60 (dd, J = 7.6, 6.8 Hz, 1 H), 7.37-7.32 (m, 3H),7.23-7.21 (m, 4H), 7.06-6.98 (m, 2H), 5.20 (s, 2H), 3.79 (s. 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 134.0, 133.7, 133.6, 132.3, 131.5, 130.9, 128.6, 128.4, 127.7, 126.6, 124.9, 121.9, 121.3, 118.4, 113.3, 70.6, 52.3. IR (KBr): 1703, 1588, 1449, 1349, 1267, 759 cm⁻¹. MS m/ z: 377 (M⁺, 0.61), 345 (25), 344 (100), 318 (20), 312 (27). HRMS-EI (m/z): [M]⁺ calcd for C₂₂H₁₉NO₃S, 377.1086; found, 377.1091.

4.1.1.37. 2-(2-Benzyloxy-phenyl)-benzothiazole-4-carboxylic acid methyl ester (38). To a solution of potassium ferricyanide(III) (2.87 g, 8.7 mmol) in water (2 mL) at 80-90 °C was added a solution of compound 37 (0.82 g, 2.2 mmol) in ethanol (1 mL), 30 % sodium hydroxide solution (1.68 mL), and water (0.72 mL) at a rate of 5 mL per minute. After stirred for 1 h, the reaction solution was cooled to room temperature and poured into water. The resulting solution was extracted with ethyl acetate, and the extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was dissolved in a mixture of diethyl ether and methanol, and added diazomethane at 0 °C until ceased to generate nitrogen. After adding a few drops of acetic acid, the reaction solution was concentrated, and the residue was purified by flash column chromatography using silica gel as the stationary phase and using hexane and then ethyl acetate/hexane (1:49) as the mobile phase producing compound 38 (0.17 g, 0.5 mmol). Yield: 21%. mp: 112-116 °C. ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3, \delta)$: 8.73–8.70 (d, J = 7.9 Hz, 1 H), 8.11-8.04 (m, 2H), 7.54-7.52 (d, J = 7.6 Hz, 2H), 7.45-7.32 (m, 5H), 7.17-7.09 (m, 2H), 5.36 (s, 2H), 4.07 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 166.9, 164.8, 156.4, 149.9, 137.8, 135.9, 132.2, 130.3, 128.68, 128.66, 128.3, 127.8, 125.7, 124.3, 123.7, 122.3, 121.4, 112.7, 71.0, 52.4. IR (KBr): 1696, 1452, 1292, 1241, 1113, 751 cm⁻¹. MS m/z: 376 (M⁺+1, 14), 375 (M⁺, 54), 358 (20), 269 (12), 211 (11). HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₁₇NO₃S, 375.0929; found, 375.0925.

4.1.1.38. 2-(2-Hydroxy-phenyl)-benzothiazole-4-carboxylic acid methyl ester (39). To a solution of compound **38** (91.0 mg, 0.2 mmol) in anhydrous dichloromethane (2 mL) were added ethanethiol (1.5 mL) and boron trifluoride diethyl etherate (1.0 mL), and stirred at room temperature for 5 days. The reaction was quenched by

adding saturated sodium bicarbonate solution and extracted with diethyl ether. The extract was dried over anhydrous magnesium sulfate. After filtration and concentration, the residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49, 1:19) as the mobile phase producing compound **39** (56.0 mg, 0.2 mmol). Yield: 81%. mp: 163–165 °C. ¹H NMR (300 MHz, CDCl₃, δ): 8.24-8.21 (d, J = 7.7 Hz, 1H), 8.12-8.09 (d, J = 8.0 Hz, 1 H), 7.71-7.68 (d, J = 7.9 Hz, 1H), 7.52-7.47 (dd, J = 7.9, 7.8 Hz, 1H), 7.45–7.40 (dd, J = 8.0, 7.9 Hz, 1H), 7.18–7.16 (d, J = 8.4 Hz, 1H), 7.00-6.94 (t, J = 8.0, 6.4 Hz, 1H), 4.09 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 170.7, 165.9, 158.7, 149.4, 134.1, 133.5, 129.7, 128.4, 126.0, 125.0, 123.1, 119.4, 118.2, 116.3, 52.5. IR (KBr): 1717, 1487, 1258, 1114, 741 cm⁻¹. MS *m*/*z*: 286 (M⁺+1, 9), 285 (M⁺, 59), 254 (18), 253 (100), 225 (10). HRMS-EI (m/z): [M]⁺ calcd for C₁₅H₁₁NO₃S, 285.0460; found, 285.0466.

4.1.1.39. 2-(2-Benzyloxy-phenyl)-benzothiazole-4-carboxylic acid (40). To a solution of compound 38 (0.21 g, 0.6 mmol) in tetrahydrofuran (10.5 mL) was added 1 N sodium hydroxide solution (16.1 mL), and refluxed overnight. After cooled to room temperature, the reaction solution was acidified by concd hydrochloric acid to give a white precipitate. The precipitate was filtered and washed with water, and dried under vacuum to produce compound 40 (0.19 g, 0.5 mmol). Yield: 95%. mp: 223-225 °C. ¹H NMR (300 MHz, CDCl₃, δ)g 8.47–8.43 (d, J = 7.9 Hz, 1H), 8.38–8.35 (d, J = 8.0 Hz, 1H), 8.09– 8.06 (d, J = 8.0 Hz, 1H), 7.55–7.50 (m, 4H), 7.47–7.41 (m, 3H), 7.22–7.16 (m, 2H), 5.39 (s, 2H). ¹³C NMR $(125.7 \text{ MHz}, \text{ CDCl}_3, \delta)$: 166.0, 156.7, 149.6, 135.8, 135.4, 133.4, 129.6, 129.5, 128.7, 128.6, 128.0, 126.3, 125.1, 121.7, 121.6, 120.5, 113.0, 71.2. IR (KBr): 1731, 1454, 1248, 994, 747 cm⁻¹. MS *m*/*z*: 362 (M⁺+1, 12), 361 (M⁺, 53), 317 (15), 253 (16), 211 (10). HRMS-EI (*m/z*): $[M]^+$ calcd for $C_{21}H_{15}NO_3S$, 361.0773; found, 361.0780.

4.1.1.40. 2-{[2-(2-Benzyloxy-phenyl)-benzothiazole-4carbonyl]-amino}-benzoic acid methyl ester (41). A solution of compound 40 (0.10 g, 0.3 mmol) in anhydrous dichloromethane (4 mL) was added oxalyl chloride (1.0 mL, 11.5 mmol), and stirred for 10 minutes at room temperature; following added 5 drops of N,N-dimethylformamide, and stirred for 2 h. Then the solvent was removed and dried under vacuum for 3 h. The residue was dissolved in anhydrous dichloromethane (2 mL). To this solution was added a solution of compound 2 (0.05 g, 0.3 mmol) and pyridine (0.5 mL) in anhydrous dichloromethane (3 mL) via canula, and stirred overnight. The reaction was quenched by adding water and extracted with dichloromethane. The organic phase was dried over anhydrous magnesium sulfate. After filtration and concentration, the resulting residue was purified by flash column chromatography using silica gel as the stationary phase and using ethyl acetate/hexane (1:49) as the mobile phase producing compound 41 (0.08 g, 0.2 mmol). Yield: 54%. mp: 233–235 °C. ¹H NMR (300 MHz, CDCl₃, δ): 12.9 (s, 1H), 8.73-8.70 (d, J = 7.6 Hz, 1H), 8.65-8.62 (d, J = 7.6 Hz, 1H), 8.47–8.45 (d, J = 7.6 Hz, 1H), 8.12– 8.09 (d, J = 8.0 Hz, 1H), 8.06-8.03 (d, J = 7.9 Hz, 1H),

7.57–7.42 (m, 8H), 7.24–7.18 (dd, J = 7.6, 7.6 Hz, 1H), 7.14–7.05 (m, 2H), 5.38 (s, 2H), 3.74 (s, 3H). ¹³C NMR (125.7 MHz, CDCl₃, δ): 167.2, 164.4, 164.1, 156.6, 148.8, 140.1, 136.9, 135.8, 133.6, 132.3, 131.0, 130.5, 128.8, 128.7, 128.4, 127.9, 126.7, 125.0, 124.5, 124.1, 123.4, 121.9, 121.1, 119.2, 112.7, 71.1, 52.1. IR (KBr): 1718, 1662, 1535, 1449, 1244, 752 cm⁻¹. MS *m*/*z*: 495 (M⁺+1, 33), 494 (M⁺, 100), 344 (30), 254 (37), 253 (94). HRMS-EI (*m*/*z*): [M]⁺ calcd for C₂₉H₂₂N₂O₄S, 494.1300; found, 494.1304.

4.2. Biological evaluation

4.2.1. Cell culture. The human bladder papillary transitional cell carcinoma cell line BFTC-905, the human lung carcinoma cell line A-549, the human uterus sarcoma cell line MES-SA, and the human embryonal rhabdomyosarcoma cell line RD were obtained from the Bioresources Collection & Research Center (BCRC, Taiwan). All cancer cell lines were maintained in the BCRC recommended medium supplemented with 10% fetal bovine serum and penicillin–streptomycin. After 24 h incubation at 37 °C in humidified 5% carbon dioxide to allow cell attachment, the cells were treated with various concentrations of UK-1 derivatives and incubated for 72 h under the same conditions. Even though the UK-1 derivatives were dissolved in DMSO, in the treatment stage the final DMSO concentration never exceeded 0.5%.

4.2.2. Cytotoxicity assay. Cell viability was determined by measuring the metabolism of a tetrazolium substrate, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT).¹⁶ Briefly, the cells $(1 \times 10^4 \text{ cells per})$ well) were incubated in 96-well plates in which each cell contained 200 µl of the growth medium at 37 °C. Following overnight incubation, the growth medium was removed from cultures and replaced with 200 µl of the test article solution. The MTT test was conducted using six concentrations, $100 \,\mu\text{M}$, $75 \,\mu\text{M}$, $50 \,\mu\text{M}$, $25 \,\mu\text{M}$, 5 uM, and 0 uM, in four cell lines. Tests at each concentration were conducted in triplicate. Test article solutions were incubated with target cells for approximately 72 h at 37 \pm 1 °C with 5 \pm 1% CO₂. Following exposure, the test solutions were removed and replaced with 500 µg/ml MTT solution. After around 6 h of incubation, the MTT solution was removed. Sodium dodecyl sulfate (SDS) solution (10%) was added into cell cultures and the optical density (OD) values were read using an ELISA reader (SUNRISE, TECAN, Switzerland) at a wavelength of 570 nm. The mean and standard deviation of each group were calculated. The OD reading of every group was first substrate with blank control (background). The relative survival rate was the OD reading of the treated group divided by that of the negative control group, multiplied by 100. The percentage of toxicity was calculated as (1 - survival rate) \times 100%. Therefore, the negative control was set to 100% survival or 0% toxicity.

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