

How we drifted into peptide chemistry and where we have arrived at

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Abstract—The history of peptide chemistry in our group is described. It all started with the cyclic undecapeptide cyclosporin, the immunosuppressive compound, which is commercialised as Sandimmune®/Neoral® by Sandoz/Novartis, and which has revolutionized transplant medicine. The discovery that cyclosporin can be deprotonated to a hexalithio derivative, and thus *C*-alkylated on a sarcosine moiety, led us into a research project on *peptide modifications*. We defined structural prerequisites for the use of peptide enolates and for electrolytic decarboxylation of peptides. Parallel to these activities, the group was engaged in developing synthetic methodologies aimed at stereoselective preparations of α -, β -, and γ -amino acid derivatives (cf. diastereoselective alkylations, self regeneration of stereogenic centers, axially chiral enolates). A third avenue into peptide chemistry originated from our investigations on the biopolymer PHB (poly-3-hydroxybutanoic acid); the question arose ‘what happens upon replacement of chain-bound O by NH in the polyester?’ A brief summary is given of the results obtained in our ensuing discovery tour of β -peptides built of homologated proteinogenic amino acids. They form secondary structures with short chain lengths and they have unexpected physiological properties, rendering them candidates for peptidic drugs. The synthesis of β^3 -peptides is straightforward, and in the meantime most of the Fmoc-protected building blocks are commercial. The β^2 -homoamino acids are less readily available. Their preparation and the assembly of a β^2 -eicosapeptide with the twenty proteinogenic side chains are discussed herein. The reasons for the chosen sequence and the strategy of what turned out to be a 159-step synthesis are described. Full experimental details are given for the preparation of the dimeric Fmoc- β^2 hXaa(PG)- β^2 hXaa(PG)-OH building blocks used, for their solid-phase coupling to two β^2 -decapeptide segments, for the thioligation, and for the purification, isolation and spectroscopic characterization of the resulting 20mer. An outlook to future projects in the exciting field of β - and γ -peptide chemistry and biology is given.

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1. Introduction

At the beginning of the senior author’s (D. S.) journey through organic chemistry¹ he would never have expected to become a peptide chemist towards the end of his career. The lectures of his mentor in Karlsruhe, Rudolf Criegee, did not cover this subject at all. If asked, he would probably have answered, like most organic chemists at the time, that peptide chemistry is a highly specialized field, and that it is chemically boring to do nothing but create amide bonds. Thus, D. S. became a physical organic chemist studying the mechanism of peroxide decomposition and of cyclobutene ring-opening reactions. In the postdoctoral work with E. J. Corey in Cambridge and in the first steps into independent research back in Karlsruhe and later in Giessen with a growing

group he was engaged in sulfur-, lithium-, nitroso-, and nitro-organic chemistry as a synthetic methodologist (umpolung of reactivity, pool of chiral building blocks). Later, in Zürich, the group moved into the areas of stereoselective transformations, self-regeneration of stereocenters, total synthesis of natural products (such as elaiophyllidin and myxovirescin), structure and mechanisms of organolithium compounds, use of organotitanium reagents, TADDOL as a chiral auxiliary system of broad applicability, all the way to novel crosslinkers for polymerization and catalysts immobilized on controlled-pore glass.

The first encounter with peptide chemistry^{2,3} occurred in one of the senior author’s regular consulting visits at Sandoz in Basel. The issue was to find a more sensitive method of detection of their immunosuppressive drug cyclosporin A in plasma. With the experience of our group in organosulfur and -selenium chemistry⁴ we treated cyclosporin with naphthylselenenylchloride to induce a selenocyclization of the side chain in the unique C₉-amino acid of the peptide; the product has a much higher extinction coefficient than

Keywords: Cyclosporin; Chiral enolates; Peptide chemistry; β -Peptides.

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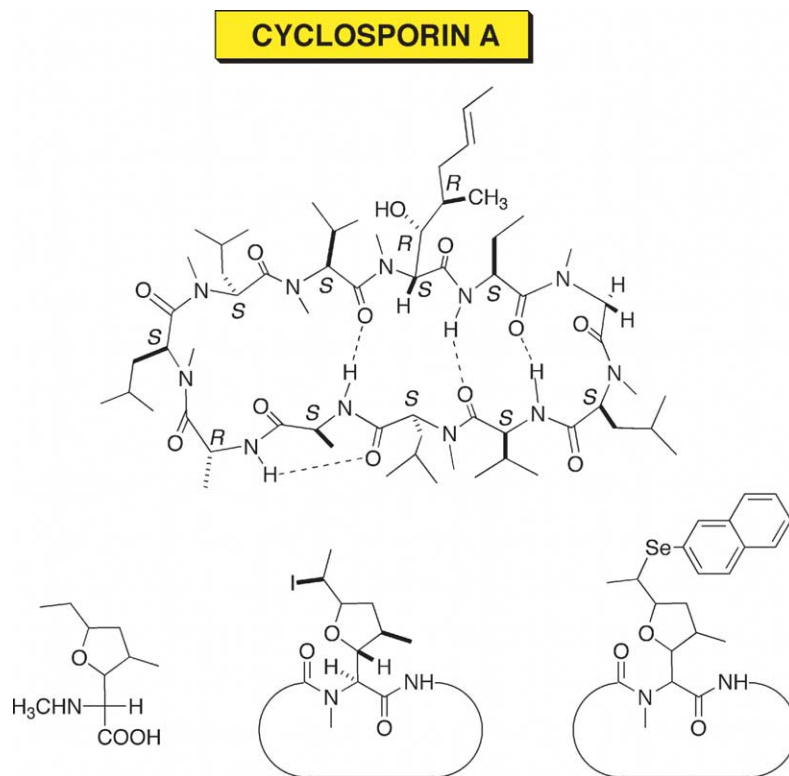


Figure 1. Formula of cyclosporin A and the THF derivatives formed with strong acid, iodine or $C_{10}H_7SeCl$.

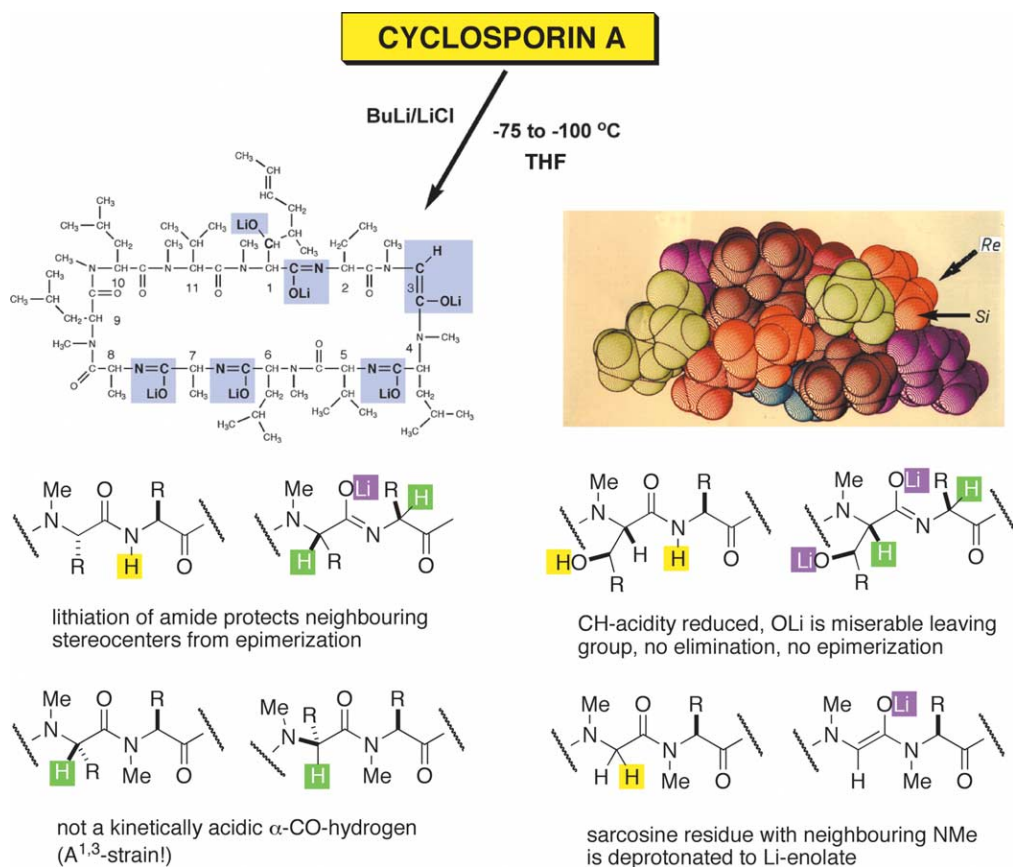


Figure 2. Hexalithiated cyclosporin A with a sarcosine enolate moiety that reacts selectively with electrophiles. Different types of amino-acid building blocks in cyclosporine and rationalization for lack of epimerization and elimination. The sarcosine CH_2 -protons are the least acidic ones!

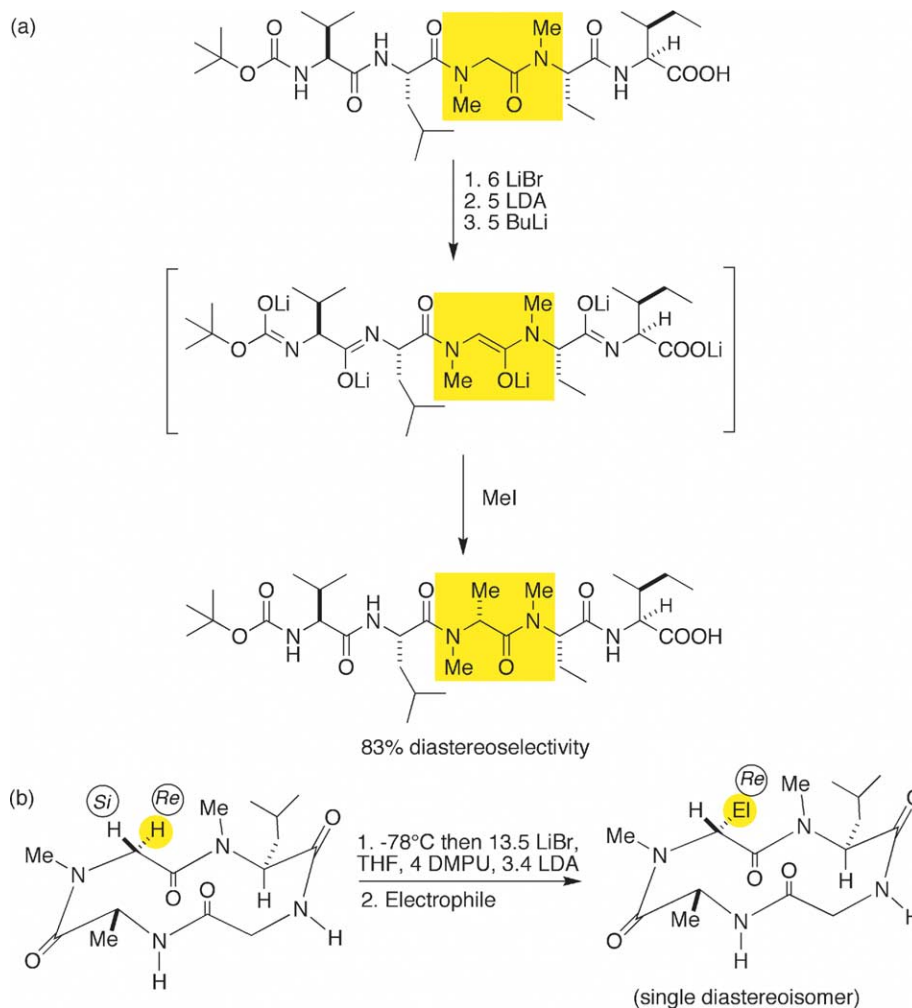


Figure 3. Li-Enolates of peptides and solubilization of peptides in THF; alkylation of open-chain (a) and of cyclic (b) peptides.

cyclosporin itself, which allowed for HPLC analysis with a detection limit of 5 ng (Fig. 1).⁵

The cyclic undecapeptide cyclosporin A containing one (*R*)-amino-acid and seven *N*-methyl-amino-acids has actually been seminal to our group's entry into the field of peptide chemistry. Returning from another visit to Sandoz in 1983, where there was a brain-storming session about possible chemical transformations of cyclosporin, D. S. carried a bottle of the peptide for an experiment he had proposed, causing shaking of heads among experts: why not generate a Li-enolate at the sarcosine residue and introduce side-chains by reactions with electrophiles? Indeed, treatment of cyclosporine with as strong bases/nucleophiles as butyllithium (in excess of six equivalents) and addition of typical electrophiles led to highly selective replacement of either the *Re*- or the *Si*-hydrogen in the sarcosine moiety, depending on the particular base and conditions employed;^{6–10} an analysis of how this was possible is given in Figure 2.

Thus, we have excised a single proton from a peptide of molecular mass 1200 and replaced it by a side-chain substituent. The derivatives, in which the diastereotopic *Re*-hydrogen had been replaced are immunosuppressive like

cyclosporin and have non-altered backbone structures. Those with H^{*Si*}-replacement have a different structure and exhibit different physiological activities.

The work on cyclosporin alkylations has triggered a series of investigations about peptide enolates,^{6,11,12} about solubilization of peptides in THF by addition of Li salts^{10,13,14} (Fig. 3), about direct thionations (C=O → C=S) of cyclosporin with Lawesson's reagent,¹⁵ and about cyclosporin as a Li- and Ca-specific ionophor.¹⁶

We started a program entitled 'chemical modifications of peptides' which led to a number of dissertations in the group. One line of work was dedicated to the use of peptides containing amino-malonic-acid derivatives, which require only weak bases for alkylations¹⁷ (Fig. 4). In a quite different investigation we used electrochemical oxidative decarboxylations of peptides containing up to ten amino-acid residues, a process leading to modifications of the C-termini¹⁸ (Fig. 5). Yet another project was the in situ generation of ketenes for certain peptide couplings which enabled us to incorporate a single β-homoamino-acid unit into a larger peptide: activation of the C-terminal CO₂H-group, reaction with diazomethane, and decomposition of the resulting diazoketone in the

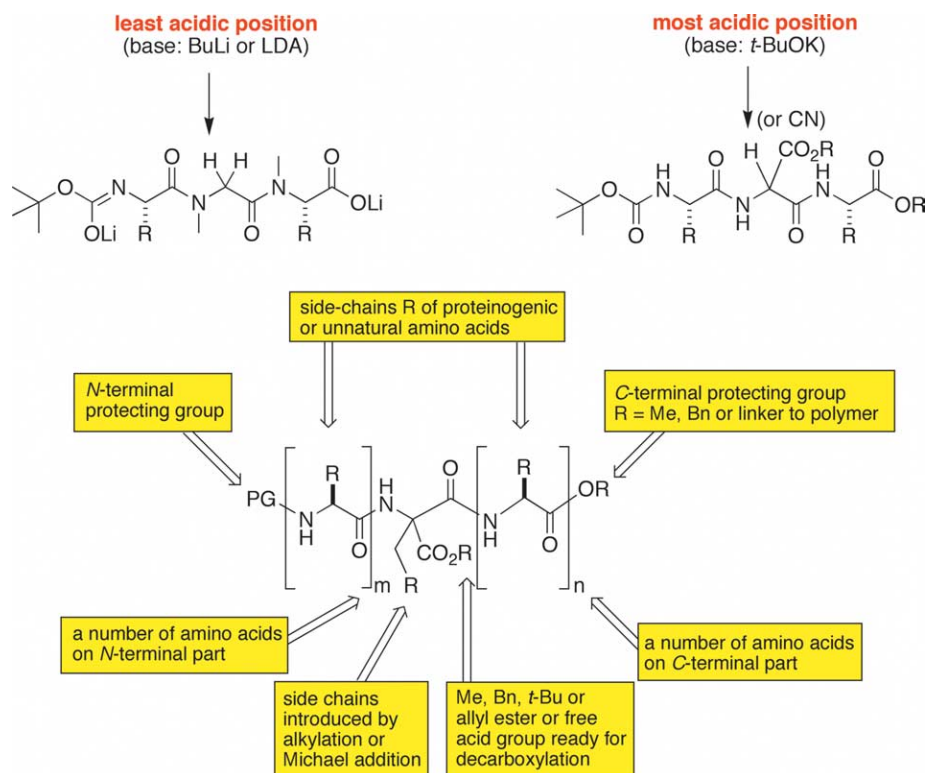


Figure 4. Peptide alkylation by going from the least to the most acidic CH group and application for peptide modification. The process is highly stereoselective in some cases and is amenable to combinatorial applications.

presence of a second peptide with unprotected *N*-terminus provides a homologative fragment coupling (Arndt–Eistert sequence of reactions, see Figure 6).¹⁹ The intermediate activate β -homoamino-acid derivative can also be trapped with carbohydrates or nucleosides to give chimeric products.^{20,21}

β -Homoamino acids had been part of our projects on synthetic methods for a long time. Following the work on dilithiated β -hydroxyesters, for instance malates,^{22,23} we had generated aspartate-enolates^{24,25} as early as 1981. A general method for the diastereoselective preparation of

enantiopure $\beta^{2,3}$ -homoamino acids involves formation of the Li_2 -derivative and alkylation of the corresponding β^3 -homoamino acids^{26,27} (for nomenclature see an extensive review article on β -peptides²⁸). A route to α -branched aspartates employs the principle of self-regeneration of stereocenters (SRS),^{25,29,30} and chiral enolates of the achiral 3-amino-propanoic acid can be generated from suitable hydropyrimidines.³¹ Formulae of some of these nucleophilic reagents are shown in Figure 7.

We actually conceived the idea of studying β -peptides in the course of our work on the biopolymer PHB (= poly(3-

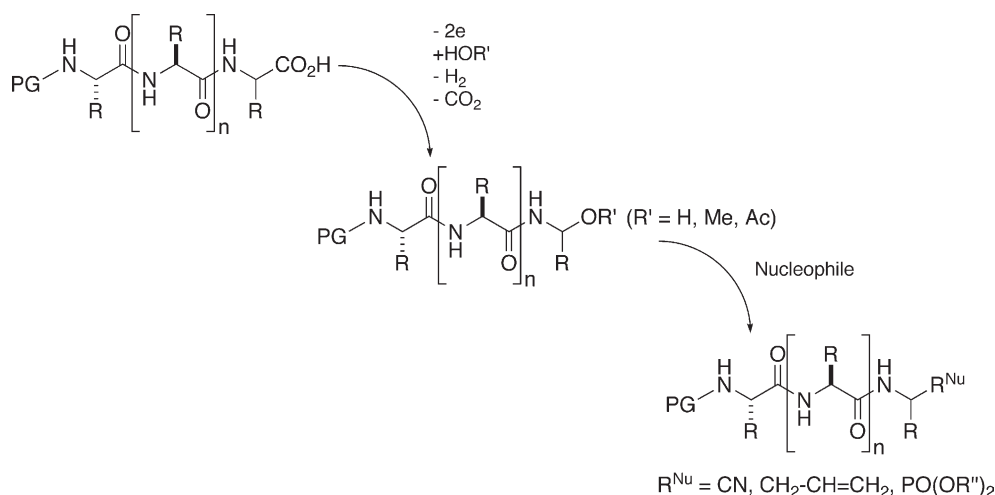


Figure 5. Hofer-Moest Electrolysis of peptides in a protic solvent (H_2O , MeOH , AcOH) and subsequent Lewis-acid-mediated nucleophilic substitution to *N*-terminally modified peptides.

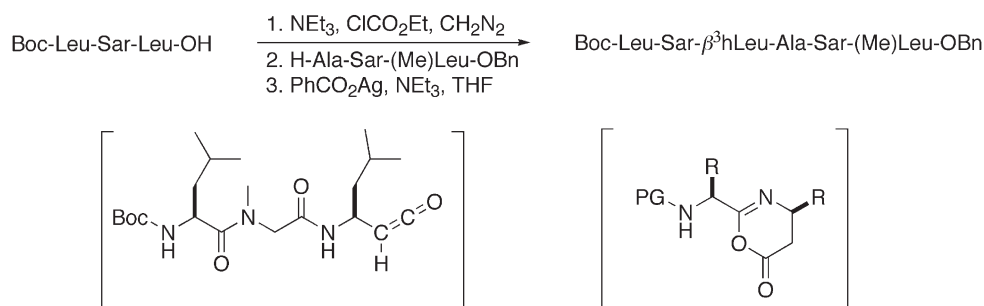


Figure 6. Homologative peptide-fragment coupling through a peptidic ketene intermediate. In the absence of an *N*-methyl-group the actual acylating reagent may be a di-dehydro-oxazinone. H- $\beta^3\text{hLeu-OH}$: (*S*)-3-amino-5-methyl-hexanoic acid.

hydroxybutyrate)): the backbones of cyclic oligo-((*R*)-3-hydroxybutanoates) were found to have a propensity to form (*P*)-helical conformations from which a helix could be modelled, containing chain-bound and carbonyl oxygens at such a distance and relative position that replacement of the former by an NH was expected to lead to hydrogen bonding, and thus stabilization of the helix^{28,32} (Fig. 8, top). Exchange of O by NH in the backbone of a poly(3-hydroxyalkanoate) renders the same backbone structure as insertion of a CH_2 -group in each and every amino-acid residue of a peptide (Fig. 8, bottom). The experimental test of these ideas led to many chemical, structural, and biological surprises, a full account of which is given in a review article²⁸ and in ca. 100 papers of our group since 1996 (see the attached complete list of publications of D. S.).

As expected, the additional tetrahedral carbon atom in each amino-acid residue of β -peptides leads to greater structural variety. There are not just two enantiomeric forms but also positional isomers (β^2 - and β^3 -homoamino acids), diastereoisomers (*L*- and *u*- $\beta^{2,3}$ -homoamino acids with two side chains), and there can be heteroatoms on the backbone (cf. 2-halo- or 2-hydroxy-3-amino acids). As a consequence, five β -peptidic helices have been identified: an 8-,^{28,33} 10-,³⁴ 10/12-,³⁵ 12-,³⁶ and 14-helix^{37,38} (the numbers refer to the size of the hydrogen-bonded rings within the helix structures). Also, the β -peptidic backbone can be forced to adopt a pleated-sheet structure or to form a hairpin turn.³⁹ With two exceptions,^{34,39} these structures are seen in solution with as few as six residues, they can be designed and found by molecular-modelling programs.⁴⁰ Thus, there are more secondary structures than in the ' α -world', but they can be predicted and constructed from a small number

of β -homoamino acid residues. Some of the structures are shown in Figure 9.

Due to the different dimensions, geometries, and polarities of the β -peptidic structures the biological properties of β -peptides differ from those of α -peptides in those cases where exact fitting is mandatory: they do not bind to the active sites of peptidases and are proteolytically stable.⁴¹ More surprisingly, they are even metabolically most stable in mammals, such as rats,^{42,43} in insects, and in plant-cell cultures,⁴⁴ and very slow biodegradation by environmental microorganisms has been demonstrated in one case.⁴⁵ On the other hand, β -peptides can be used to mimic α -peptidic hairpin turns, motifs which are often decisive for so-called ligand–receptor recognitions. This was demonstrated by the design of *N*-acyl- β -tetrapeptide amides with specific nanomolar binding as agonists at one of the human receptors for the peptidic hormone somatostatin (Fig. 10).^{46,47}

One of these peptides was recently shown to be orally bioavailable, to pass the blood-brain barrier and to regulate numerous genes in brain tissues.⁴³ Many other biological tests have been performed with β -peptides (for instance inhibition of an intestinal transport protein, antibiotic and hemolytic activities, binding to DNA and RNA; see references in a review article²⁸). In recent investigations of β -oligoarginine derivatives,⁴⁸ it was shown that these polyelectrolytes enter mammalian cell in vitro and in vivo to end up in the nucleoli of cell nuclei (Fig. 11), where they remain located for longer periods of time (in contrast to corresponding α -oligo-arginines, which are proteolytically degraded once having entered cells). There seem to be no toxic effects; the cell

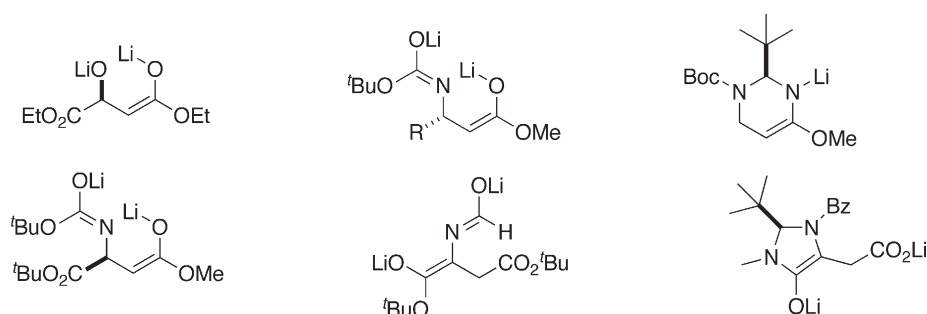


Figure 7. Chiral Li-enolates of β -heterosubstituted carboxylic acid esters. Malic acid, β -amino acids, aspartic acid.

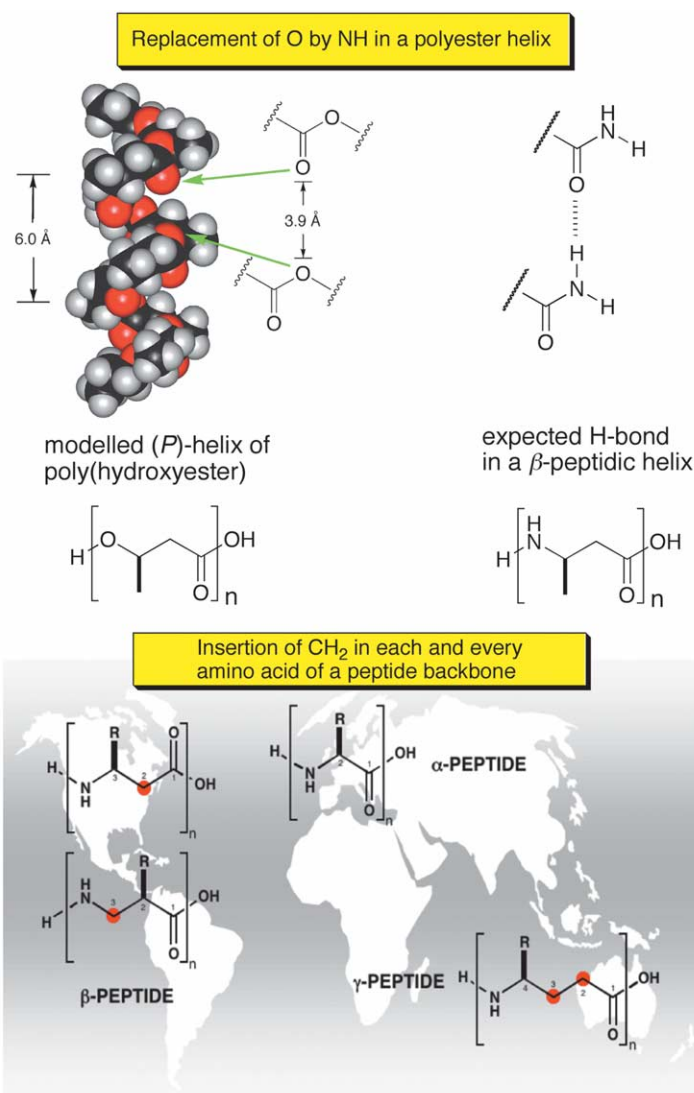


Figure 8. The question ‘what happens upon replacement of O by NH in the modeled helix of PHB?’ (top) is structurally equivalent to the question ‘what happens upon CH₂-insertion in the amino acids of a peptide backbone?’ (bottom).

culture of human keratinocytes keeps growing in the presence of the β -oligoarginines.

Thus we have gone a long way from the first experiments with cyclosporin A in 1980 to animal experiments with ¹⁴C-labelled β -peptides in 2003.

So far, there was no mention made about synthesis of the β -peptides, which was actually the main occupation of everybody in the group! At the beginning, we prepared the β^3 -homoamino acids from the corresponding α -amino acids by Arndt–Eistert homologation ourselves. In the meantime, 18 of the 20 β^3 -homoamino acids with the side chains of the proteinogenic α -amino acid analogs are commercial (*N*-Fmoc- and acid labile side-chain protection); the exceptions are β^3 hCys and β^3 hHis.⁴⁹ The β^2 -homoamino acids, on the other hand, have to be prepared enantioselectively. Since our research on β -peptides was focused on oligomers of homologs of the natural α -amino acids, we prepared the whole set of the 19 β^2 -homoamino acids (β^2 hGly = β^3 hGly!). We use the chiral-auxiliary approach

applying the modified Evans oxazolidinone DIOZ,^{50,51} as outlined in Figure 12; for details see the original publications.^{46,48,52–57}

Notably, it takes up to 13 steps to prepare some of the acids Fmoc- β^2 hXaa(PG)-OH! Having the β -homoamino acids available for solid-phase synthesis by the Fmoc strategy (manual or in a synthesizer), we could make use of all the methods common in α -peptide synthesis, purification, analysis, structure determination, and modelling (Fig. 13), with certain adjustments (see discussion in a review article²⁸).

For longer-chain β -peptides the thioligation method,⁵⁸ which works equally well for α -, β^2 -, and β^3 -peptide couplings,⁵⁹ turned out advantageous. We have recently addressed the issue, to which chain lengths a β -peptide might form the 3_{14} -helix: α -peptidic helices in proteins are typically only 15–20 residues long, one reason being a destabilization by the resulting macrodipole which increases with chain-length.⁶⁰ Thus, we have prepared a

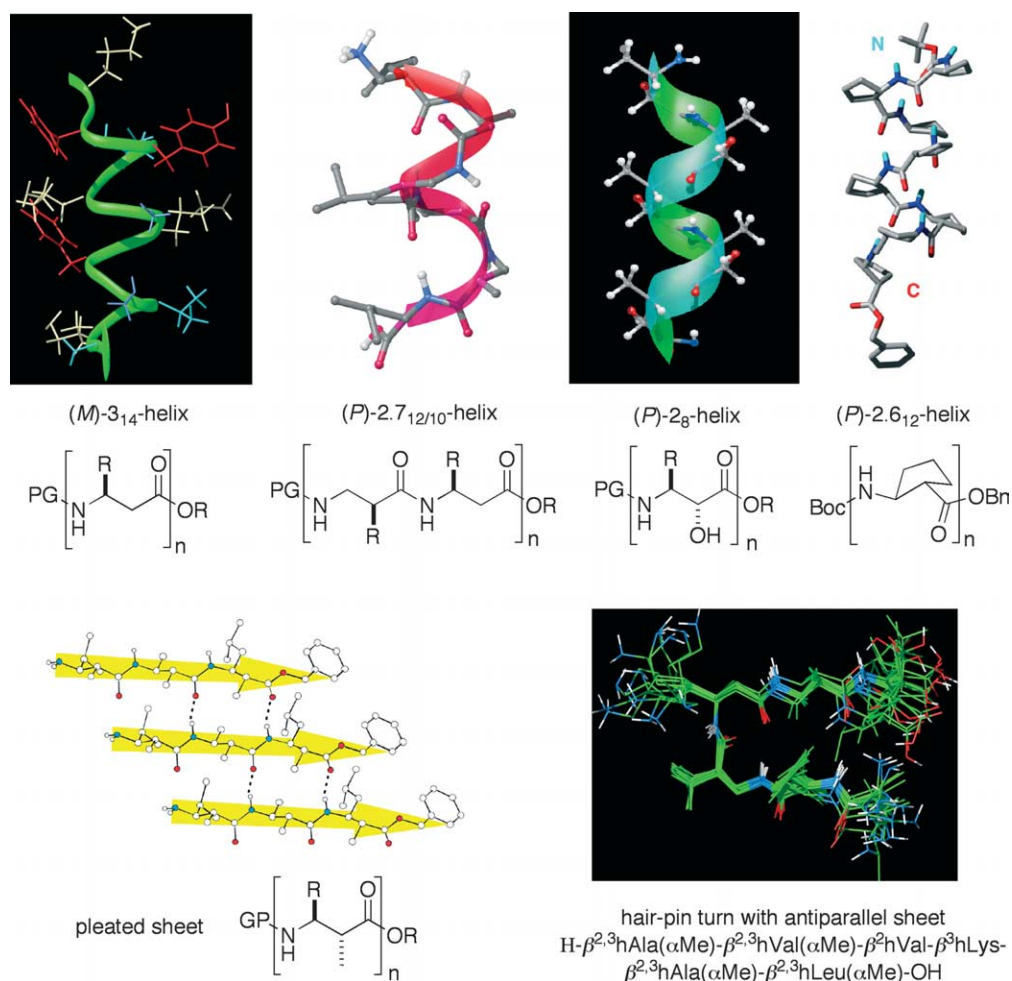


Figure 9. Helix, sheet, and turn structures of β -peptides. Except for the parallel pleated-sheet structure all secondary structures shown can be observed by NMR spectroscopy of solutions. This Figure has, in part, been reproduced by permission of the Verlag Helvetica Chimica Acta. [Rossi, F.; Lelais, G.; Seebach, D. *Helv. Chim. Acta*, **2003**, 86, 2653. Etezady-Esfarjani, T.; Hilty, C.; Wüthrich, K.; Rueping, M.; Schreiber, J.; Seebach, D. *Helv. Chim. Acta*, **2002**, 85, 1197.]

β^3 -eicosapeptide consisting of the 20 different β^3 -hom amino acids;⁶¹ the sequence was chosen such that an amphiphatic helix would result, and stabilization by salt bridges between (*i*) and (*i*+3)-positions was part of its design, see the helical-wheel-type presentation in Figure 14. The CD spectra in methanol and water exhibit an intensive negative Cotton effect between 210 and 220 nm which we may consider typical of a 3₁₄-helical secondary structure, however without the usually more intensive short-wave-length maximum seen with short β^3 -peptides. The NMR-solution structure determination of this 20mer is underway, and it looks like there is a helix in methanol over the full length of the 20 residues.⁶²

As the last major project of our group before retirement of D. S. (with the concomitant necessary reduction of the research-group size) we joined forces and made essentially everybody (from advanced lab-course students, through master-thesis candidates, the last PhD students all the way to the post-doctoral co-workers) part of a team to synthesize the *all*- β^2 -eicosapeptide **1** with the 20 proteinogenic amino-acid side chains (see below, Fig. 16).

The reason for embarking on this adventure, which eventually turned out to be a 159-step synthesis, was

manifold. First of all, we wanted to demonstrate that we actually can synthesize all the necessary β^2 -hom amino-acid building blocks with the chiral auxiliary DIOZ. Then, we decided to find ways of avoiding racemization/epimerization in β^2 -hom amino-acid coupling, a problem we had noticed some time ago.⁶³ Also, the 3₁₄-helix of short-chain β^2 -peptides has turned out to be less stable than that of isomeric β^3 -peptides,^{64,65} so that a comparison of larger β^2 - and β^3 -peptides was important, to find out, whether the former ones fold to other secondary structures. Finally, there was an atmosphere of sportive ambition in the group about getting it done!

The synthesis of the β^2 -eicosapeptide **1** was designed to be as safe as possible. To make sure that there would not be insurmountable problems in the purification of the final product, we used a convergent synthesis for the β^2 -peptide (Fig. 15), aware of the fact that there would be more danger of epimerization/racemization than with β^3 -hom amino acids, and remembering that the isolation of the pure β^3 -eicosapeptide (Fig. 14), assembled in one stroke, had been quite cumbersome.^{28,61}

For the choice of the sequence (there are more than 10¹⁸ possibilities) we applied several different criteria:

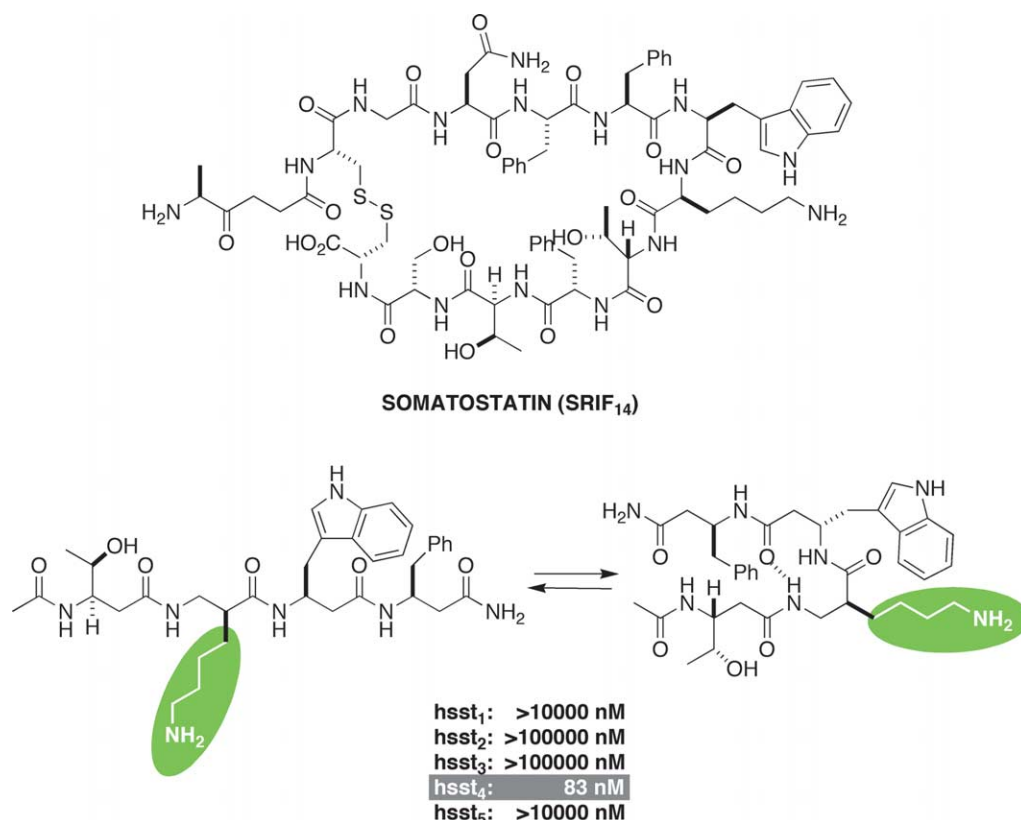


Figure 10. Formula of somatostatin and of a β -tetrapeptide derivative binding to one of the five human somatostatin receptors.

- The 3_{14} -helix of the eicosapeptide, should it be formed, was supposed to be amphipathic, with stripes of polar and non-polar side chains on its surface (Fig. 16).
- Also, the helix should experience salt-bridge stabilization, by putting the two pairs of positively and negatively charged side chains of Arg/Glu and Lys/Asp in (*i*)- and (*i*+3)-positions, that is, in juxtaposition on the helix at a distance of approximately 5 Å (cf. Fig. 9, 3_{14} -helix, top left).
- Next, we considered the well known ‘capping effect’,⁶⁰ according to which negative side chains near the positive and positive side chains near the negative end of a peptidic helix dipole⁶⁶ (Fig. 17) have a stabilizing effect, also in β -peptides,⁶⁷ thus we placed the β^2 hArg in position 3 and the β^2 hAsp in position 17 of the β^2 -eicosapeptide **1**.
- The choice of β^2 hCys in position 11 is dictated by the thioligation, and we put β^2 hAla next to it (position 10) to reduce steric hindrance in the course of this coupling process.
- Also, the amino acids bearing the side chains of His and Met have been shown to be incompatible with the types of reactions (cf. treatment with CH_2N_2 or with $\text{I}-\text{CH}_2\text{CN}$) used for the solid-phase synthesis of peptide thioesters;⁶⁸ therefore the corresponding β^2 -homoamino acids had to be incorporated in the decapeptide **3** with the terminal β^2 hCys residue (Figs. 15 and 16).
- The β^2 hPro residue was necessary to be placed in position 20 (i.e., first on the Wang resin), because this amino acid with its secondary piperidine-amino group does not fit into a 3_{14} -helix; rather it is a hairpin-turn structural element.⁶⁹

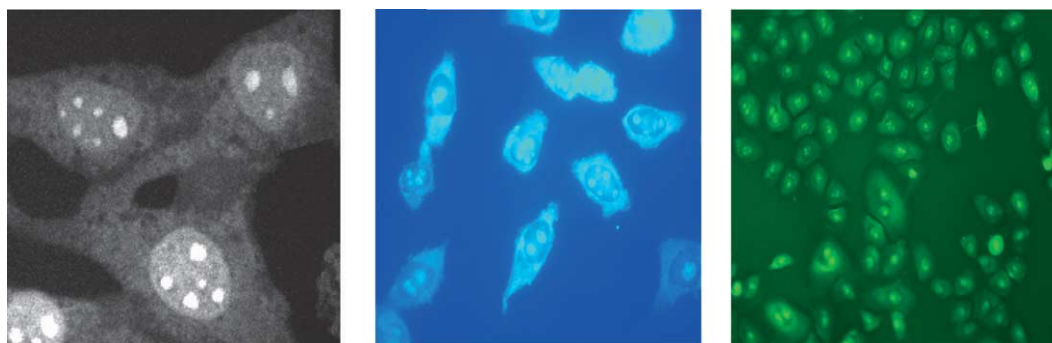
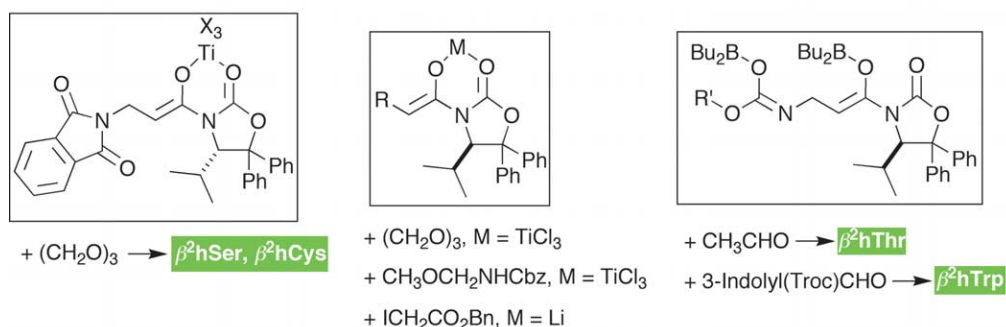


Figure 11. Fluorescence microscopy of mouse fibroblast (left), HeLa (center), and human keratinocyte cells (right) after treatment with fluoresceinylated β^3 -oligoarginines consisting of 7, 8 or 10 β^3 hArg residues. This Figure has, in part, been reproduced by permission of the Verlag Helvetica Chimica Acta. [Seebach, D.; Namoto, K.; Mahajan, Y. R.; Bindshädler, P.; Sustmann, R.; Kirsch, M.; Ryder, N. S.; Weiss, M.; Sauer, M.; Roth, C.; Werner, S.; Beer, H.-D.; Munding, C. *Chem. Biodiversity*, **2004**, *1*, 65.]

Preparation of β^2 -Homoamino Acid Derivatives

R	$\beta^2\text{hXaa}$
 CH ₃ PhCH ₂ BnOC ₆ H ₄ CH ₂ (CH ₃) ₂ CH (CH ₃) ₂ CHCH ₂ C ₂ H ₅ (CH ₃)CH CH ₃ SCH ₂ CH ₂ Br(CH ₂) _{2,3} CH ₂ (4-Imidazolyl-1-yl)Trt ^t BuO ₂ C(CH ₂) _{1,2} TrtNHCO(CH ₂) _{1,2}	 Ala Phe Tyr Val Leu Ile Met Arg, Pro, Lys His Asp, Glu Asn, Gln

Figure 12. Preparation of 19 β^2 -homoamino acid derivatives with the proteinogenic side chains, using enolates derived from the chiral auxiliary DIOZ. For the preparation of Fmoc-(S) β^2 hTrp(Boc)-OH, the classical Evans auxiliary (without the geminal Ph groups) gives better results.

Methods

Synthesis of β amino acids:	by known methods
Coupling of β amino acids:	Fmoc-, Boc-, Cbz-protection; EDC/HOBt or HATU activation
Solubilization:	with LiCl or LiBr in solution and on the resin
Solid phase synthesis:	Rink-, Wang-, Sulfonamide or <i>o</i> -chlorotrityl polystyrene (Merrifield) resins
Machine synthesis:	Applied Biosystems 433A Synthesizer
Fragment coupling:	Kemp-Kent ligation
Purification:	HPLC on RP columns
Sequencing:	Low-energy ESI MS/MS coupling
Molecular dynamics:	GROMOS 96
Structure determination:	Routine use of NMR and CD measurement
Characterization:	HiRes Maldi and HiRes ESI mass spectrometry

Figure 13. The synthesis and analysis of β -peptides from the Fmoc- β^2 - or - β^3 hXaa(PG)-OH is accomplished by the well established methods of α -peptide chemistry (see text books and monographs).

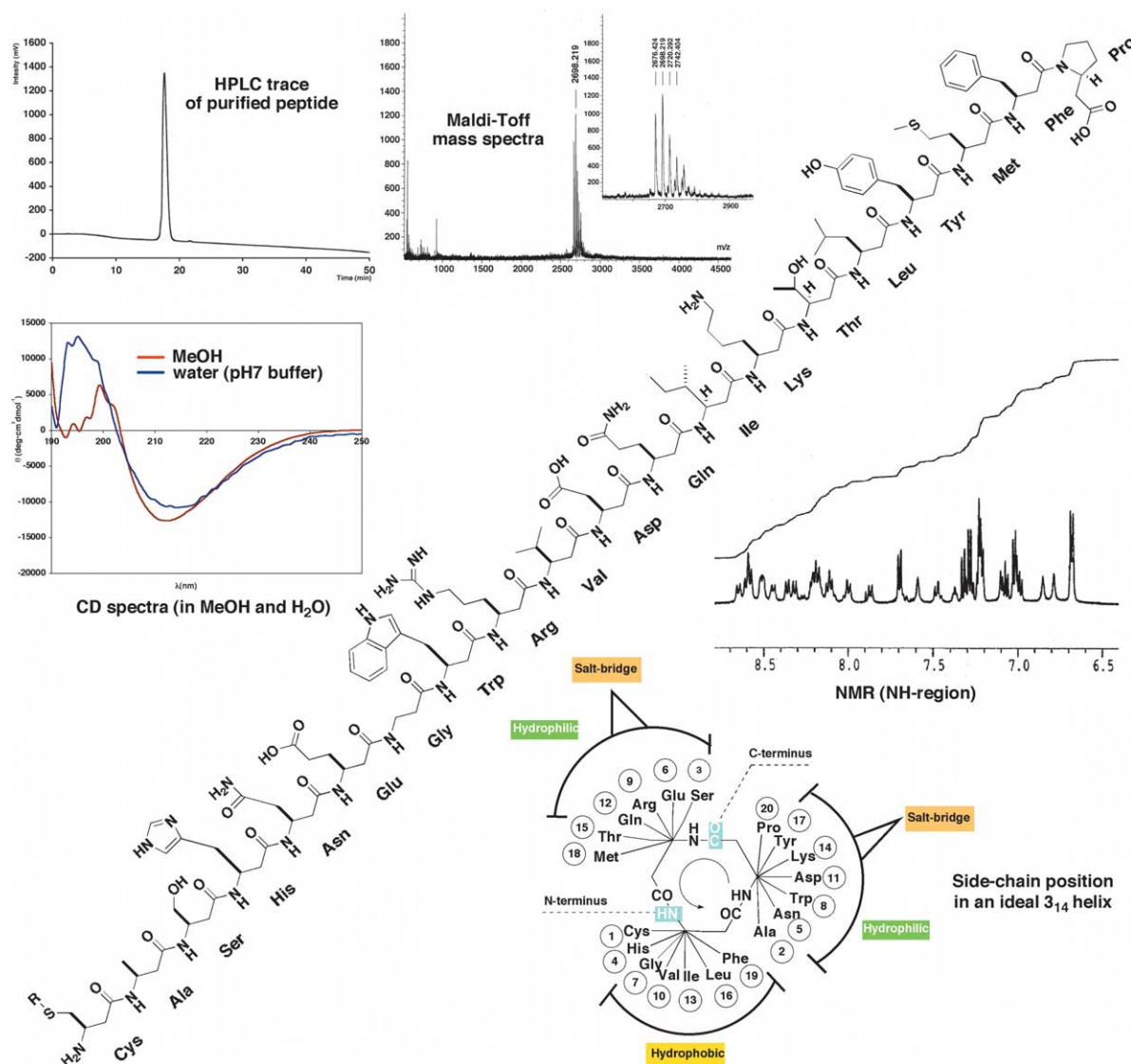


Figure 14. A β^3 -eicosapeptide containing the 20 homologated proteinogenic amino acids. The compound was prepared on solid support and purified by preparative HPLC. MS, HPLC of purified sample, CD (normalized) and NMR spectra, and helical-wheel presentation of an (*M*)- 3_{14} -helix, which might be formed by the β^3 -eicosapeptide.

- The β^2 hGly in position 1 was chosen in order to have a sterically unbiased *N*-terminus for derivatization.
- Furthermore, we employed β^2 -di-peptide-fragment coupling, to make sure that the two β^2 -decapeptide fragments **2** and **3** would be easy to purify in case of failure of a coupling step (a decamer is expected to be separated more easily from an octamer than from a nonamer); the β^2 -di-peptide building blocks **4**–**12** could readily be isolated in diastereomerically pure form.
- The sequence of the dimer segments was, wherever possible, chosen such that the less epimerization-prone β^2 -homoamino acid was at the *C*-, and the more ‘dangerous’ one (Phe, Asp, His, Cys, Tyr, Asn side chains) at the *N*-end; in this way, activation of the carboxylic acid group as active ester would involve less risk of epimerization.

The formulae of the suitably protected dipeptide derivatives are shown in Figure 18, and their preparations are

outlined in the experimental part, where the not previously described intermediates **13**–**25**, are fully characterized, including specific references to their β^2 hXaa-precursors. The dipeptide-coupling steps are preceded by numerous protection, deprotection and protective-group interchange operations. The enantiomer purities of all β^2 hXaa starting materials were checked by HPLC analysis on chiral columns and/or by NMR spectroscopy of diastereomeric Pd-complexes.⁷⁰ Likewise, the diastereomer purity of the Fmoc- β^2 -dipeptide acids was confirmed by NMR and RP-HPLC analysis before use in the solid-phase coupling steps, to make sure that no epimerization has occurred during dipeptide coupling or, else, that any epimer, which might have been formed, had actually been removed in the chromatographic purification procedure.

For the synthesis of β^2 -decapeptide **3** bearing an *N*-terminal β^2 hCys, by the Fmoc/*t*Bu solid-phase strategy on Wang resin, the first dipeptide **4** was attached to the resin using the

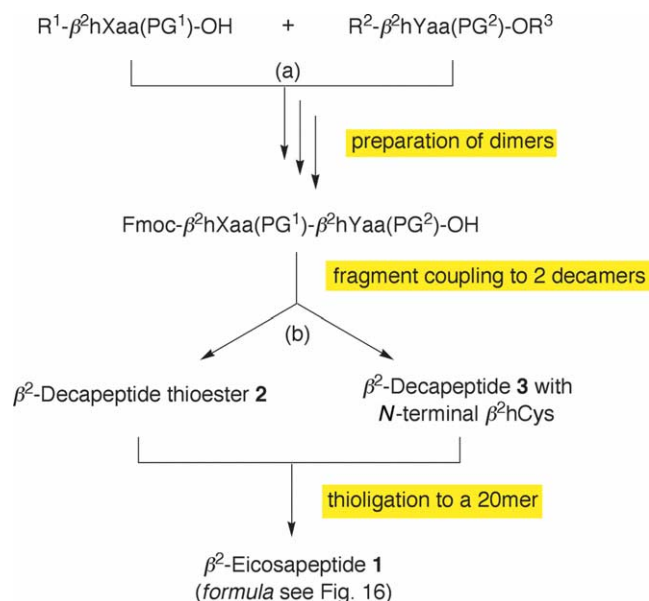


Figure 15. Strategy for the synthesis of a β^2 -eicosapeptide through two β^2 -decapeptides by dimer-fragment solid-phase coupling on sulfonamide (2) and Wang (3) resin, with subsequent thioligation.

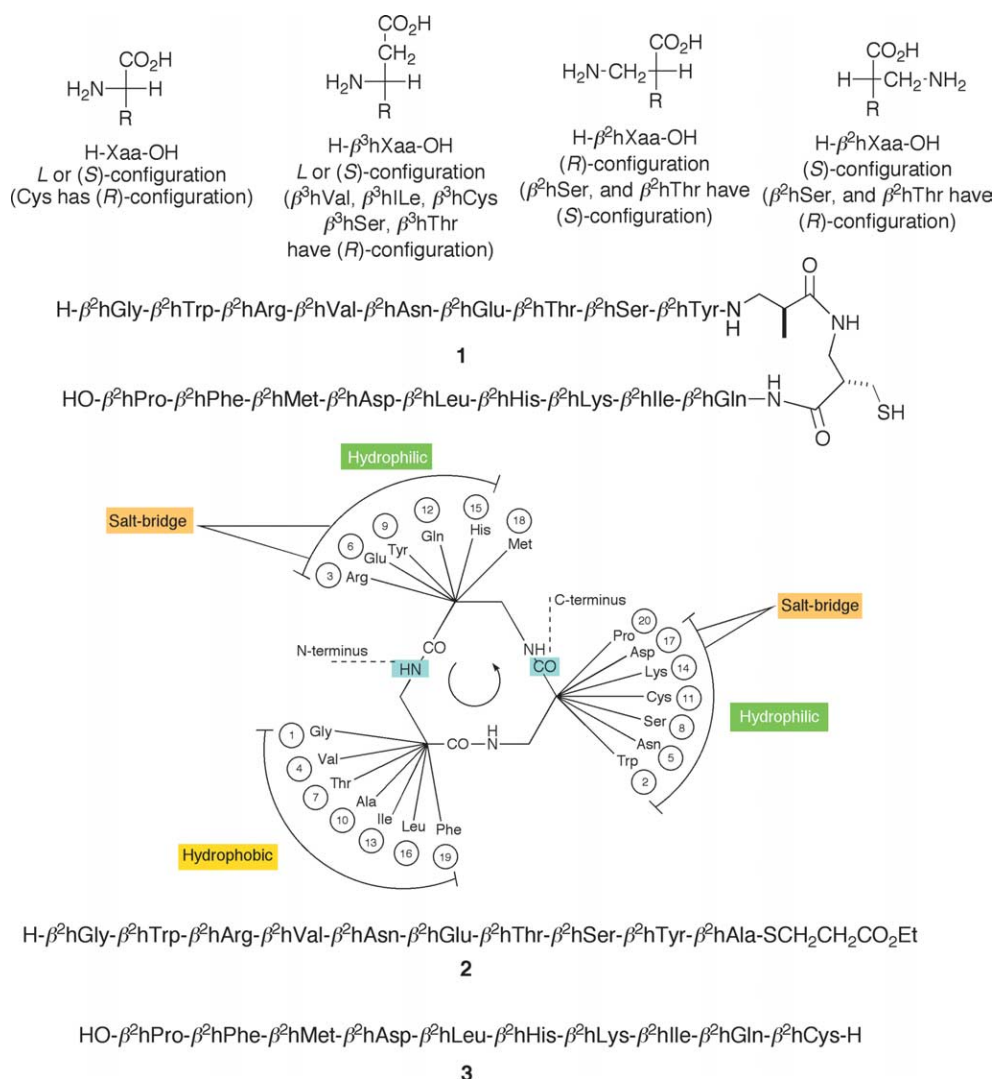


Figure 16. Configurational nomenclature of β -homoamino acids (top), and β^2 -eicosapeptide 1 with central $\beta^2\text{hAla}$ and $\beta^3\text{hCys}$ segment, idealized helical-wheel-type presentation of its supposed (M)- 3_{14} -helix secondary structure (middle), and the two β^2 -decapeptide precursors 2 and 3 for thioligation (bottom).

MSNT/MeIm method⁷¹ and the resin loading determined (after treatment with piperidine, 20% in DMF), by measuring the absorbance of the dibenzofulvene-piperidine adduct at 290 nm (we use the common abbreviations of peptide chemistry⁷²). The unreacted OH groups were then 'capped' by acetylation (Ac_2O and DMAP). Chain elongation on solid support was performed with HATU and 3 equiv. of the Fmoc-protected β^2 -dipeptides 5–8, and with piperidine for Fmoc deprotection. After the last coupling, the peptide was cleaved from the resin and the side chains deprotected by treatment with $\text{CF}_3\text{COOH/EDT/TIS/H}_2\text{O}$. Finally purification by reverse-phase HPLC yielded the β^2 -peptide 3, which was analysed by high-resolution mass spectrometry (ESI HRMS).

The β^2 -peptide 2 bearing a C-terminal thioester was prepared using the methodology developed by Ingenito et al.⁶⁸ and based on Kenner's acylsulfonamide safety-catch linker.^{73,74} The loading of the resin was achieved with 4 equiv. of the Fmoc-protected β^2 -dipeptide 9, and DIPCDI/MeIm in DCM/DMF. The peptide was then assembled by the standard Fmoc protocol (HATU as coupling reagent and

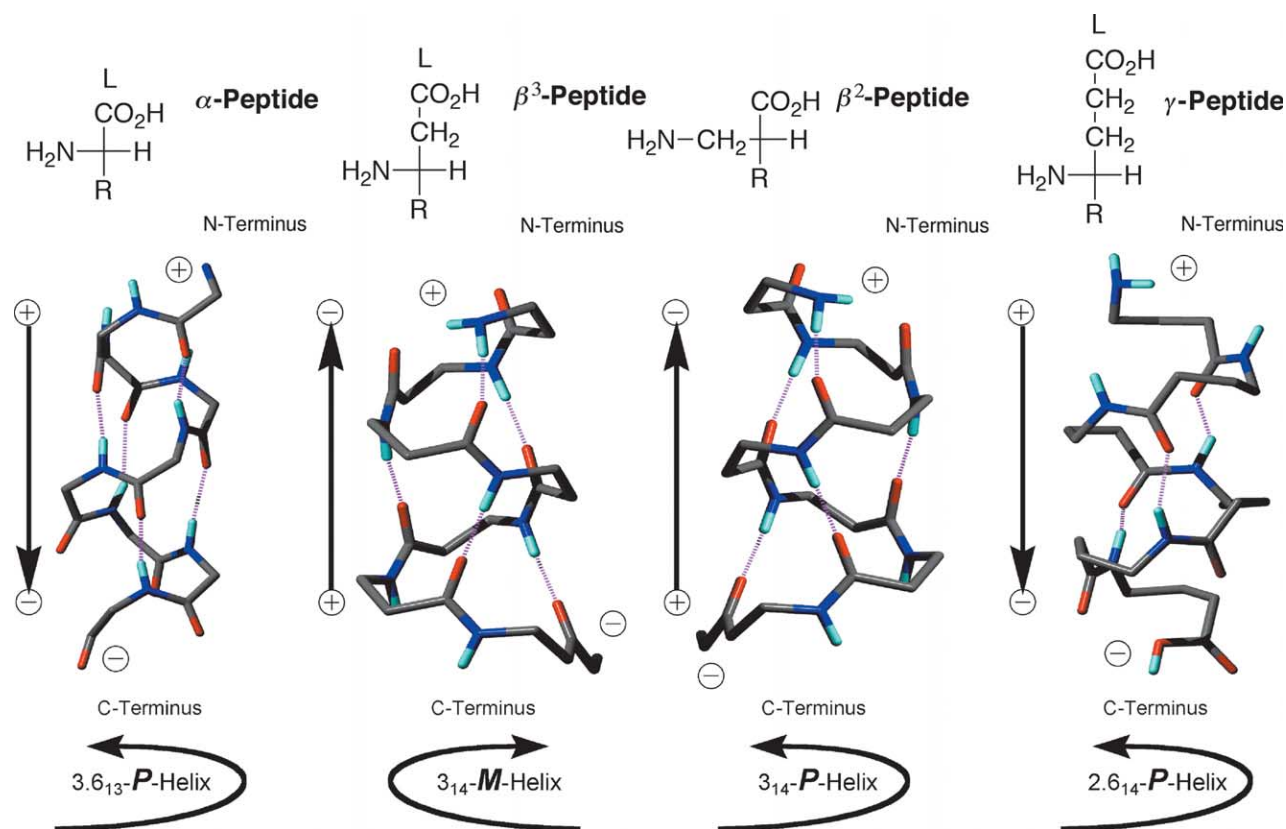


Figure 17. Helicity and direction of macrodipole reverse, as we go from α - to β^3 - or β^2 - to γ^4 -peptides built of homochiral amino acids. The α - and γ -peptidic helices suffer from destabilizing pole-charge interaction, which is a stabilizing effect in the β -peptidic 3_{14} -helix. Furthermore, the resulting macrodipoles (increasing with chain lengths) destabilize the helices, which is counteracted by side chains with opposite charge ($\oplus$ near $\ominus$ pole and vice versa). Note that helices built of β^2 hXaa (shown here) and of the enantiomeric building blocks (Figure 12) have opposite helicity. This Figure has, in part, been reproduced by permission of the Verlag Helvetica Chimica Acta. [Seebach, D.; Schreiber, J. V.; Abele, S.; Daura, X.; van Gunsteren, W. F. *Helv. Chim. Acta*, **2000**, 83, 34.]

piperidine for Fmoc deprotection). After the last coupling step, activation of the safety-catch linker, treatment with diazomethane followed by a displacement reaction involving NaSPH/HS(CH₂)₂COOC₂H₅ led to the still protected decapeptide. Finally, the side chain-protecting groups were removed in solution by treatment with TFA in the presence of an appropriate scavenger. In this way we obtained the β^2 -peptide **2**. The displacement reaction did first not work, even in the presence of LiBr.^{10,13,14,75,76} However, heating the reaction mixture at 80 °C overnight, and deprotection resulted in the formation of the desired thioester, which was purified by preparative reverse-phase HPLC and identified by high-resolution mass spectrometry. CD Spectra of the two β^2 -decapeptides **2** and **3** are shown in Figure 19.

The chemical ligation-methodology, which allows the coupling of unprotected peptide fragment in aqueous solution, has made considerable advance in recent years. It offers a new route for the synthesis of larger peptides and proteins.^{77,78} In the thioligation reaction the coupling process starts with a trans thioesterification reaction involving a peptide already bearing a C-terminal thioester and the sulfhydryl group of a second peptide bearing an N-terminal Cys. The thioester-linked intermediate undergoes a subsequent rapid intramolecular S $\rightarrow$ N acyl shift, forming the amide bond at the ligation site. In the case of peptides containing a β^2 -homocysteine, the intramolecular S $\rightarrow$ N acyl shift in the ligation reaction proceeds through a 6-membered, rather than a 5-membered

heterocycle, which is involved with Cys and β^3 hCys as coupling components (Fig. 20).

We applied this method for the final step in the synthesis of the β^2 -peptide **1** containing all the β^2 -homocysteine amino acids with proteinogenic side chains. The ligation was performed under standard conditions⁷⁹ (aqueous solution, pH 7.5 phosphate buffer and 4% (v/v) PhSH). After 4 h, more than 70% conversion had occurred and the reaction was essentially complete after 12 h as evident from the analytical RP-HPLC traces shown in Figure 21.

The crude product **1** was then purified by preparative reverse-phase HPLC and characterised by high-resolution mass spectrometry. The normalized CD spectrum of the β^2 -eicosapeptide **1** in methanol shows the familiar negative Cotton effect between 210 and 220 nm with an intensity similar to that observed with the isomeric β^3 -eicosapeptide (Fig. 14). However in water the negative Cotton effect (trough) almost vanishes, and an intensive positive Cotton effect (peak) appears at shorter wavelengths. Similar changes of β -peptidic CD patterns upon replacement of MeOH by H₂O as solvent had been observed previously,^{58,80–82} and commented with awe ('miraculous');⁸¹ they may suggest an alteration of the secondary structures, or unfolding to a 'totally disordered' backbone conformation. However, only a full NMR investigation will be able to elucidate what is going on (Fig. 22).^{62,82}

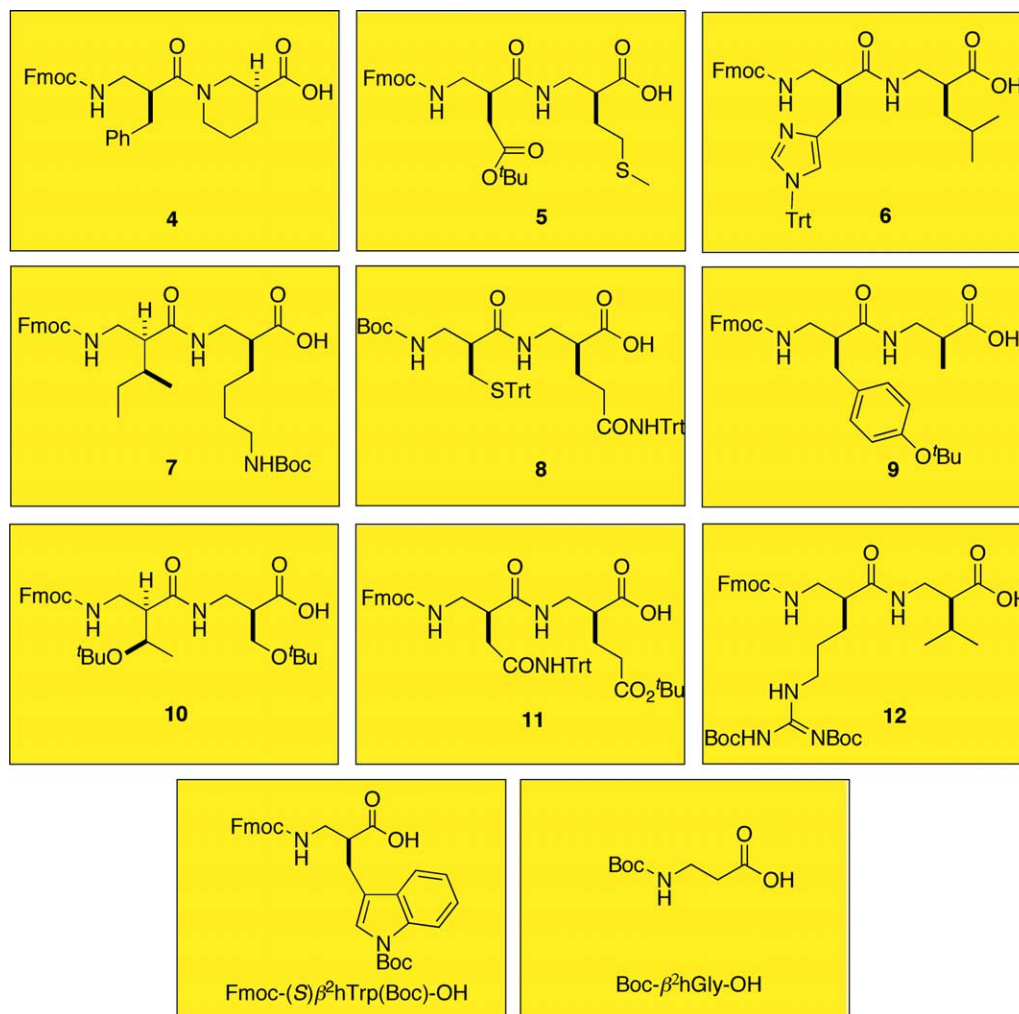


Figure 18. Nine β^2 -deptide derivatives **4–12** and the Fmoc-(*S*) β^2 hTrp(Boc) and β^2 hGly components (*N*-terminal in **1** and **2**) for the solid-phase synthesis of the β^2 -decapeptides **2** and **3** (Figure 16).

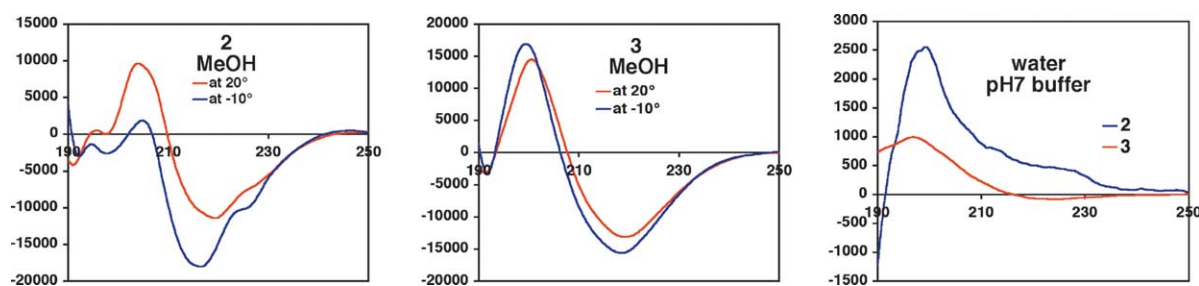


Figure 19. CD Spectra (normalized) of the two β^2 -decapeptides **2** and **3** in MeOH at +20 and -10°C , and in H_2O . The Cotton effect observed between 215 and 220 nm increases with decreasing temperature.^{64,65} The pattern obtained with **3** we would consider typical of a 3_{14} -helix. The shoulder near 225 nm and the drastic reduction of intensity of the positive Cotton effect near 205 nm seen with the β^2 -decapeptide **2**, however, is totally surprising and can not be interpreted at present. In aqueous pH 7 buffer the β^2 -decapeptide **2** shows a positive Cotton effect at 200 nm albeit with low intensity; the β^2 -decapeptide **3** does not show any significant Cotton effect. Compare the CD spectra of the β^2 -eicosapeptide **1** in H_2O and MeOH in Figure 22 below.

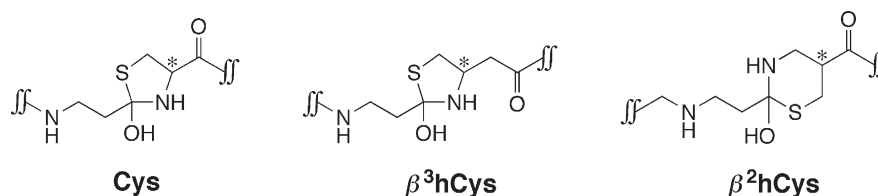


Figure 20. Formulae of the cyclic intermediates formed during the $\text{S} \rightarrow \text{N}$ acyl shift in the course of the thioligation reaction involving Cys, β^3 hCys and β^2 hCys.

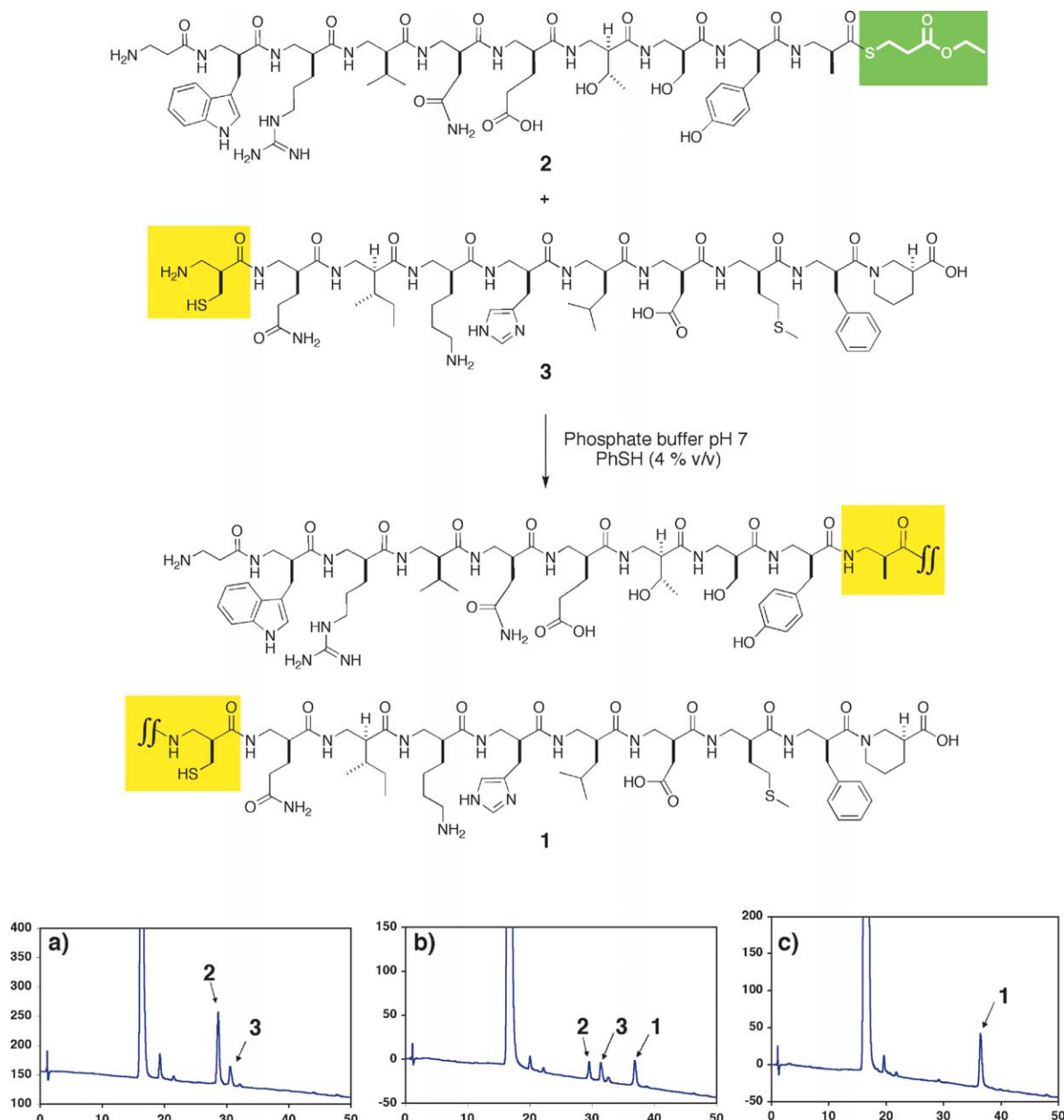


Figure 21. Analytical-HPLC traces of the ligation reaction between β^2 -decapeptide **2** with a C-terminal thioester group and β^2 -decapeptide **3** with an N-terminal β^2 -Cys residue. Samples taken from the reaction mixture at: (a) $t = 10$ min, (b) $t = 2$ h, and (c) $t = 12$ h (chromatographic conditions see Section 2).

The syntheses of the two β -eicosamers may be taken as a demonstration, that any sequence of β^2 - or β^3 -homoamino acid residues with the proteinogenic side chains can be assembled. This makes us confident that we will be able to construct—by design— β -peptides with tertiary and quaternary structures and, possibly, with catalytic activities. Some evidence for aggregation of long-chain β^3 -peptides (consisting of homologated ‘natural’ α -amino acid residues) has already emerged from concentration-dependent CD spectra.⁵⁸ Also, intramolecular helix–helix interaction has been deduced from CD spectra of a β^3 -peptide with proteinogenic side chains.⁸³

We should, however, not be too sure of our ability to synthesize any β -peptidic sequence: with the sheet-forming

α -branched $\beta^{2,3}$ -homoamino acid residues we^{39,84} and others⁸⁵ have observed difficulties in the solid-phase synthesis of corresponding β -peptides. On the other hand, we are optimistic, as synthetic organic chemists must be ‘by definition’, that there will be a solution to any synthetic problem, if we just try hard enough.

Besides construction of more complex architectures with function, the major goal in the field of β -peptides, and also γ -peptides,⁸⁶ is the exploitation of their biological, pharmacological, and biomedical potential.²⁸ Recent experiments with short-chain β -peptides (proteolytically and metabolically stable!) have involved structure-dependent tissue-specific distributions, gene profiling in brain and lung tissues, affinity to MHC-type-I proteins,

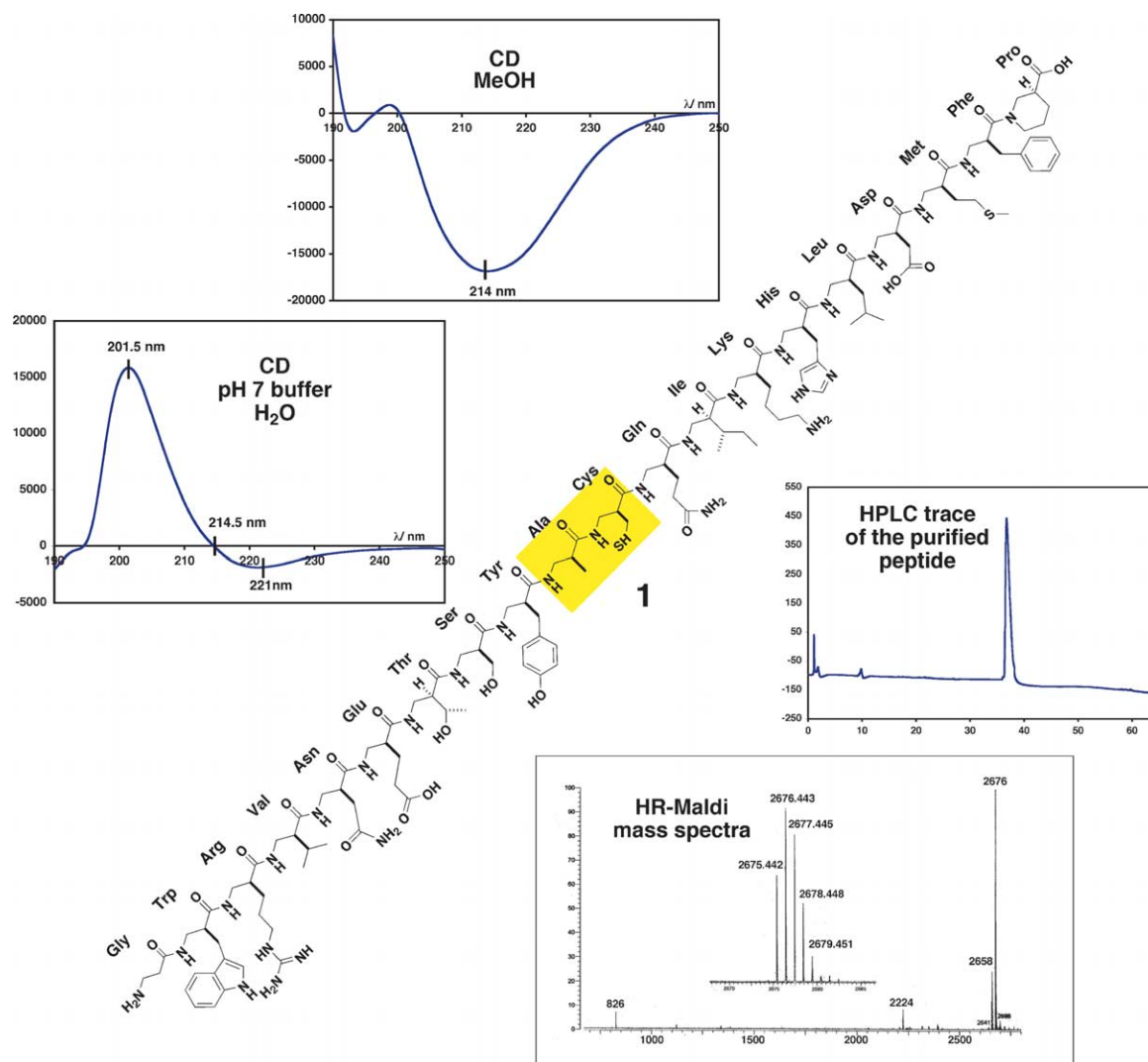


Figure 22. Formula, mass spectra, HPLC of a purified sample, CD spectrum (normalized) in MeOH and in H₂O of β^2 -eicosapeptide **1**.

and human-leukocyte-antigen-mediated protection of pig cells against human natural-killer-cell cytotoxicity.⁴³

2. Experimental

2.1. General

Abbreviations: The official abbreviations of Peptide Science⁷² are used throughout this paper. DMAP (4-(dimethylamino)pyridine), DIPCDI (diisopropylcarbodiimide), DIPEA (diisopropylethylamine), EDC (*N*-(3-dimethylaminopropyl)-*N'*-ethyl-carbodiimide hydrochloride), EDT (ethanedithiol), FC (flash chromatography), FmocOSu (*N*-(9-Fluorenylmethoxycarbonyloxy)succinimide), HATU (*O*-(7-azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate), h.v. (high vacuum, 0.01–0.1 Torr), 1-MeIm (1-methylimidazole), NMM (*N*-methylmorpholine), MSNT (1-(mesitylene-2-sulphonyl)-3-nitro-1*H*-1,2,4-triazole), TBAF (tetra-*n*-butylammonium fluoride), TFA (trifluoroacetic acid), TIS (triisopropylsilane), TNBS (2,4,6-trinitrobenzenesulfonic acid). Solvents for chromatography were distilled from Sikkon (anh. CaSO₄; Fluka), THF was distilled from Na, CH₂Cl₂ and NEt₃ from CaH₂. LiCl was

dried in h.v. at 100 °C for 1 h. All other reagents were used as received from Fluka. TLC: Merck silica gel 60 F₂₅₄ plates; detection with UV or 'Mo-stain' solution (25 g phosphormolybdic acid, 10 g Ce(SO₄)₂·H₂O, 60 mL conc. H₂SO₄, 940 mL H₂O), FC: Fluka silica gel 60 (40–63 μ m); at ca. 0.2 bar. Anal HPLC: Merck HPLC system (LaChrom, pump type L-7150, UV detector L-7400, Interface D-7000, HPLC Manager D-7000). Macherey-Nagel C₈-column (Nucleosil 100-5 C₈ 250×4 mm); Waters HPLC system (pump type 515, data module type 746, tunable absorbance detector type 484). Chiralcel OD-H column. Prep. HPLC: Merck HPLC system (LaChrom, pump type L-7150, UV detector L-7400, Interface D-7000, HPLC Manager D-7000) Macherey-Nagel C₈ column (Nucleosil 100-7 C₈ (250×21 mm)). Circular dichroism (CD): CD spectra were recorded on a Jasco J-710 spectropolarimeter from 190 to 250 nm with a Jasco PTC-348 WI Peltier System at 20 °C or –10 °C in 1 mm rectangular cells. The optical system was flushed with N₂ at a flow rate of ca. 10 L/min. Parameters: band width 1.0 nm, resolution 0.2–1 nm, sensitivity 100 mdeg, response 0.5 s, speed 50 nm/min, 5 accumulations. All spectra were corrected for the corresponding solvent spectrum and normalized. Peptide concentrations were typically 0.2 mM. The molar ellipticity [θ] in

deg·cm²·mol⁻¹ (λ in nm). Smoothing was done by Jasco software. Solvents: MeOH (HPLC grade), aq. Buffer pH 7.0: 0.1 M KH₂PO₄/0.1 M NaOH. NMR: Bruker AMX 500 (¹H 500 MHz, ¹³C 125 MHz), AMX-400 (¹H 400 MHz, ¹³C 100 MHz) and Varian Gemini (¹H 300 MHz, ¹³C 75 MHz) chemical shifts δ in ppm downfield from internal SiMe₄ (0 ppm). Mass Spectra: IonSpec Ultima 4.7 T FT Ion Cyclotron Resonance (ICR, HR-MALDI, in a 2,5-dihydroxybenzoic acid matrix), or Finnigan MAT TSQ 700 (ESI) mass spectrometer; in m/z (% of basis peak). Melting points: Büchi-510 apparatus; uncorrected. Optical rotations: Perkin–Elmer 241 polarimeter (10 cm, 1 mL cell, room temperature). IR: Perkin–Elmer 1600 FT-IR spectrophotometer. Elemental analyses were performed by the Microanalytical Laboratory of the Laboratorium für Organische Chemie, ETH Zürich.

2.2. General procedures

2.2.1. Peptide coupling to give dimers 4–12: general procedure 1 (GP1). The appropriate *N*-deprotected amino acid (1 equiv.) was dissolved in CH₂Cl₂ (0.1 M) and cooled in an ice-bath. To the mixture was successively added NMM (3 equiv.) and the *N*-protected amino acid (1 equiv.). To this solution either HATU (1.2 equiv.) (GP1a) or EDC (1.2 equiv.) and HOBt (1.2 equiv.) (GP1b) was added and the mixture allowed to warm up to 25 °C and stirred overnight. The mixture was then diluted with CH₂Cl₂ and washed with 1 M HCl, 10% aq. K₂CO₃ and brine solutions. The organic phase was dried (MgSO₄) and the solvent removed under reduced pressure. The crude dipeptide was purified by FC.

2.2.2. Hydrogenolysis of Cbz and Bn-ester groups: general procedure 2 (GP2). The corresponding substrate was dissolved in either MeOH or THF (0.02 M) and ca. 10% (w/w) Pd/C (10%) was added. The apparatus was evacuated and flushed with H₂ (3×), and the solution was stirred under an atmosphere of H₂ for the indicated amount of time (monitoring by TLC). Subsequent filtration through Celite and removal of solvent under reduced pressure yielded the product, which was used in the next step without further purification.

2.2.3. Saponifications: general procedure 3 (GP3). The appropriate ester (1 equiv.) was dissolved in MeOH/H₂O 3:1 (0.1 M) at 25 °C. To the resulting solution, LiOH·H₂O (2.5 equiv.) was added and the reaction mixture was stirred 3 h. The mixture was diluted with H₂O and extracted with

Et₂O. The aqueous phase was acidified with 1 N HCl to pH~1 and extracted with AcOEt (3×). The combined organic extracts were dried (MgSO₄) and the solvent removed under reduced pressure. The crude product was purified by FC.

2.2.4. Fmoc-protection: general procedure 4 (GP4). To a solution of the *N*-deprotected dipeptide in 0.15 M Na₂CO₃ (2 equiv.) was added FmocOSu (1.2 equiv.) in acetone (0.1 M). If necessary, the pH was adjusted to 9–10 with additional aq. Na₂CO₃ solution and the mixture stirred at 25 °C for 4 h. The acetone was carefully removed under reduced pressure at 30 °C and the resulting mixture diluted with H₂O. At this point the pH of the solution was adjusted to 9–10 using 0.6 M aq. Na₂CO₃. The aq. mixture was then extracted with Et₂O (2×). The aq. phase was separated, cooled to 0 °C and AcOEt added. With continuous stirring of the biphasic system at 0 °C, the pH of the aq. phase was adjusted to 4–5 by slow addition of 10% aq. citric acid. The org. layer was separated and the aq. layer extracted with AcOEt (2×). The combined org. layers were washed with brine, then dried (Na₂SO₄), and concentrated under reduced pressure. The crude product was purified by FC.

2.2.5. Preparation of trimethylsilylethyl esters: general procedure 5 (GP5). To a solution of the *C*-unprotected amino acid (1 equiv.) in CH₂Cl₂ (0.1 M) at 0 °C under Ar, was added trimethylsilylethanol (1.5 equiv.), DMAP (0.2 equiv.), and EDC (1.2 equiv.). The resulting mixture was stirred at 0 °C for 16 h then diluted with AcOEt and the org. phase washed with 10% Na₂CO₃ (2×), and brine, then dried (MgSO₄), and concentrated under reduced pressure. The crude product was purified by FC.

2.2.6. Trimethylsilylethyl ester deprotection: general procedure 6 (GP6). To a solution of the corresponding Si-ester (1.0 equiv.) in THF (0.1 M) was added TBAF·3H₂O (4 equiv.) and the mixture stirred at 25 °C for 1 d. The reaction mixture was then diluted with AcOEt and the organic phase washed with sat. aq. NH₄Cl, and brine, then dried (MgSO₄), and concentrated under reduced pressure. The crude product was purified by FC.

2.2.7. Reversed-phase (RP) HPLC analysis and purification. RP-HPLC analysis was performed on a Macherey-Nagel C₈ column (Nucleosil 100-5 C₈ (250×4 mm)) by using a linear gradient of A (0.1% TFA in H₂O) and B (MeCN) at a flow rate of 1.2 mL/min with UV detection at 220 nm; t_R in min. RP-HPLC purification was performed on

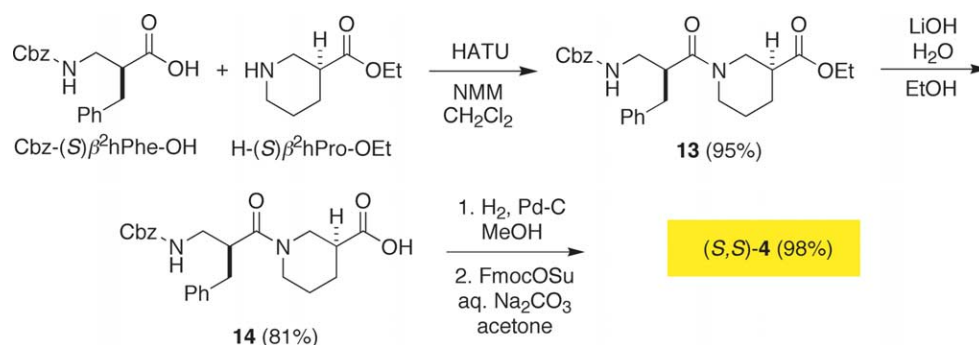


Figure 23. Preparation of Fmoc-(S)-β²hPhe-(S)-β²hPro-OH (**4**).

a Macherey-Nagel C₈ column (Nucleosil 100-5 C₈ (250×21 mm)) by using a linear gradient of A and B at a flow rate of 18 mL/min (Merck HPLC system).

2.3. Preparation of Fmoc-protected dipeptides 4–12

2.3.1. Cbz-(S)β²hPhe-(S)β²hPro-OEt (13). Amino acids Cbz-(S)β²hPhe-OH⁸⁷ (1.05 g, 3.35 mmol) and H-(S)β²hPro-OEt⁸⁸ (0.53 g, 3.35 mmol) were coupled according to GP1a. FC (AcOEt/hexane 1:1) gave **13** (1.44 g, 95%) as colorless oil; *R*_f=0.23 (AcOEt/hexane 1:1); [α]_D²⁰=+27.5 (*c*=0.80, CHCl₃); IR (CHCl₃) ν_{max} 3450 (w), 3007 (m), 2944 (w), 1720 (s), 1626 (s), 1511 (s), 1454 (m), 1139 (m), 1085 (m), 1030 (m), 856 (w) cm⁻¹. ¹H NMR (500 MHz, DMSO-*d*₆) **13**+rotamers δ 0.92–1.14 (m, 2H, CH₂), 1.17 (t, *J*=6.3 Hz, 3H, CH₃), 1.26–1.50 (m, 2H, CH₂), 1.66–1.84 (m, 2H, CH₂), 2.29 (m, 1H, CH), 2.51–2.90 (m, 4H, CH, CH₂, CHH), 3.06–3.19 (m, 2H, CH₂N), 3.67 (t, *J*=14.4 Hz, 1H, NH), 4.03 (m, 2H, OCH₂), 4.27 (d, *J*=11 Hz, 1H, CHH), 5.01 (m, 2H, OCH₂Ph), 7.07–7.49 (m, 10H, arom.); ¹³C NMR (125 MHz, DMSO-*d*₆) **13**+rotamers δ: 13.9, 23.7, 24.4, 26.5, 26.7, 35.8, 36.3, 40.0, 40.5, 40.6, 41.4, 41.9, 42.2, 42.9, 43.1, 43.2, 45.0, 46.8, 59.8, 59.9, 65.1, 126.1, 127.6, 128.1, 128.2, 128.8, 137.2, 139.3, 156.1, 171.0, 172.2, 172.4; MALDI HRMS calcd for C₂₆H₃₂N₂O₅Na (M+Na)⁺: 475.2203, found: 475.2207. Anal. calcd for C₂₆H₃₂N₂O₅: C 69.01, H 7.13, N 6.19; found: C 69.01, H 7.16, N 5.93 (Fig. 23).

2.3.2. Cbz-(S)β²hPhe-(S)β²hPro-OH (14). Dipeptide ester **13** (1.34 g, 2.96 mmol) was hydrolyzed according to GP3. The resulting crude material was crystallized from CHCl₃/hexane to give **14** (1.02 g, 81%) as white crystals; mp 189–190 °C; *R*_f=0.32 (AcOEt/hexane/AcOH 10:10:1); [α]_D²⁰=+2.4 (*c*=0.15, CHCl₃); IR (CHCl₃) ν_{max} 3446 (w), 3008 (m), 1715 (s), 1627 (s), 1513 (m), 1454 (m), 1082 (m), 1046 (m), 1005 (w), 877 (w) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) **14**+rotamers δ 0.91–1.50 (m, 5H, 2×CH₂ and CHH), 1.68–1.91 (m, 2H, CH₂), 2.21–2.86 (m, 3H, NH, CH, CHH), 3.06–3.19 (m, 2H, CH₂), 3.54 (m, 1H, CH), 4.20 (d, *J*=12.9 Hz, 1H, CHH), 4.43 (d, *J*=10.7 Hz, 1H, CHH), 5.02 (m, 2H, OCH₂Ph), 7.06–7.39 (m, 10H, arom.), 12.32 (br s, 1H, COOH); ¹³C NMR (100 MHz, DMSO-*d*₆) **14**+rotamers δ: 23.9, 24.6, 26.8, 26.9, 36.3, 40.6, 40.7, 41.5,

42.0, 42.1, 42.8, 43.1, 43.2, 45.0, 47.0, 65.1, 126.0, 126.1, 127.5, 126.6, 126.7, 128.1, 128.15, 128.22, 128.24, 128.6, 128.8, 137.1, 137.2, 139.2, 139.3, 156.1, 170.9, 171.1, 174.0, 174.2; MALDI HRMS calcd for C₂₄H₂₈N₂O₅Na (M+Na)⁺: 447.1890, found: 447.1887. Anal. calcd for C₂₄H₂₈N₂O₅: C 67.91, H 6.65, N 6.60; found: C 67.89, H 6.45, N 6.54.

2.3.3. Fmoc-(S)β²hPhe-(S)β²hPro-OH (4). Cbz-dipeptide **14** was hydrogenolyzed according to GP2 then Fmoc-protected according to GP4. The crude peptide was purified by FC (AcOEt/hexane/AcOH 10:10:0.1) to give **4** (1.13 g, 98%) as a white foam; mp 84–87 °C; *R*_f=0.35 (AcOEt/hexane/AcOH 10:10:1); [α]_D²⁰=−1.6 (*c*=0.73, CHCl₃); IR (CHCl₃) ν_{max} 3450 (w), 3008 (m), 2949 (m), 2862 (w), 1713 (s), 1625 (s), 1514 (s), 1467 (m), 1450 (s), 1181 (m), 1144 (m), 1084 (m), 1008 (m), 990 (w), 856 (w) cm⁻¹; ¹H NMR (400 MHz, CD₃OD) **4**+rotamers δ 1.02–1.99 (m, 4H, 2×CH₂), 2.27–2.93 (m, 4H, 2×CH and CH₂), 3.21–3.79 (m, 4H, 2×CH₂), 4.21 (m, 2H, CH₂), 4.30–4.53 (m, 3H, OCH₂CH), 7.08–7.79 (m, 13H, arom.); ¹³C NMR (100 MHz, CD₃OD) **4**+rotamers δ 21.5, 25.35, 25.41, 25.7, 26.4, 28.4, 28.6, 37.8, 38.1, 42.39, 42.48, 42.54, 43.3, 43.5, 43.6, 44.1, 44.5, 44.6, 44.8, 44.83, 44.86, 44.93, 45.1, 45.3, 47.4, 67.6, 67.7, 67.8, 67.9, 121.0, 126.0, 126.1, 126.2, 126.3, 127.5, 127.6, 127.7, 128.2, 128.8, 129.3, 129.5, 129.6, 129.7, 130.0, 130.3, 140.3, 140.7, 142.7, 145.4, 158.9, 174.2, 174.3, 174.4, 176.2, 176.6, 176.9; MALDI HRMS calcd for C₃₁H₃₂N₂O₅Na (M+Na)⁺: 535.2203; found: 535.2199. Anal. calcd for C₃₁H₃₂N₂O₅: C 72.64, H 6.29, N 5.46; found: C 72.71, H 6.43, N 5.24 (Fig. 23).

2.3.4. Boc-(S)β²hMet-OCH₂CCl₃ (15). The crude H-(S)β²hMet-OH⁵⁵ (2.42 mmol) was dissolved in H₂O (2.5 mL) and aq. 1 M NaOH (5 mL) at 5 °C (ice bath). To the resulting solution Boc₂O (0.63 g, 2.90 mmol, 1.2 equiv.) in dioxane (5 mL), was added and the mixture stirred for 30 min at 25 °C. The solution was concentrated to half of its original volume, cooled again in an ice bath, covered with a layer of AcOEt and acidified with a dilute solution of KHSO₄ to pH 2–3. The aqueous phase was extracted with AcOEt (2×). The combined organic extracts were washed with H₂O, dried (MgSO₄) and the solvent removed under reduced pressure. The crude product (0.45 g, 1.71 mmol)

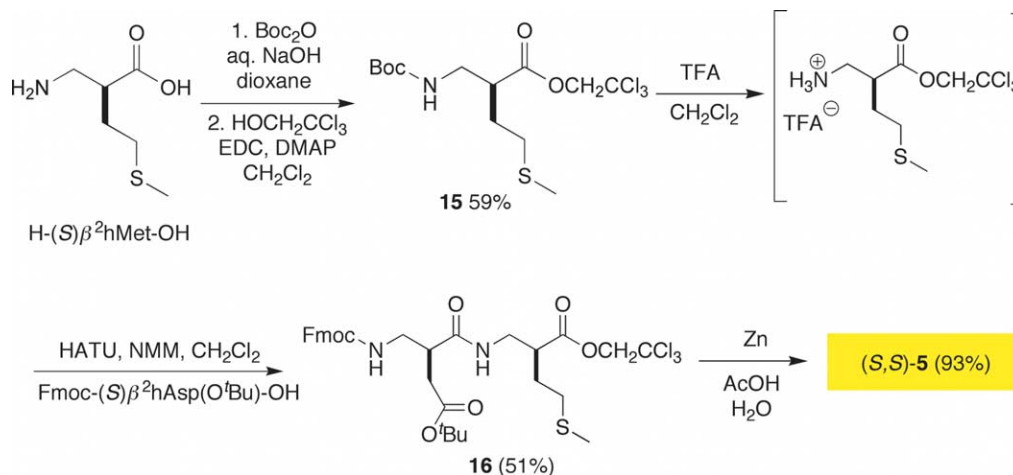


Figure 24. Preparation of Fmoc-(S)β²hAsp(O^tBu)-(S)β²hMet-OH (5).

and DMAP (42 mg, 0.34 mmol, 20 mol%) were dissolved in CH_2Cl_2 (20 mL) and $\text{Cl}_3\text{CCH}_2\text{OH}$ (0.19 mL, 0.28 g, 1.88 mmol, 1.1 equiv.) was added. The resulting solution was cooled in an ice bath then EDC (0.39 g, 2.05 mmol, 1.2 equiv.) added and the reaction mixture stirred for 18 h at 25 °C. Sat. aq. NH_4Cl was added and the organic phase washed with 0.1 N HCl, 0.1 M K_2CO_3 and brine then dried (MgSO_4) and solvent removed under reduced pressure. The crude product was purified by FC (AcOEt/hexane 3:7) to give **15** (0.56 g, 59%) as colorless oil; $R_f=0.61$ (AcOEt/hexane 1:1); $[\alpha]_D^{25}=+3.1$ ($c=0.26$, CHCl_3); IR (neat) ν_{max} 3344 (w), 3054 (w), 2976 (w), 2140 (m), 1752 (s), 1710 (s), 1513 (m), 1445 (m), 1366 (m), 1272 (m), 1250 (m), 1167 (s), 789 (m) cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 1.43 (s, 9H, ^tBu), 1.86 (m, 1H, CHHS), 2.03 (m, 1H, CHHS), 2.10 (s, 3H, SCH_3), 2.60 (t, $J=7.5$ Hz, 2H, CH_2), 2.95 (m, 1H, CHCO), 3.39 (m, 2H, CH_2N), 4.78 (s, 2H, OCH_2), 4.89 (br s, 1H, NH); ^{13}C NMR (75 MHz, CDCl_3) δ 15.4, 28.5, 28.6, 31.6, 41.3, 44.7, 74.0, 79.7, 94.8, 155.7, 172.6; MALDI HRMS calcd for $\text{C}_{13}\text{H}_{22}\text{Cl}_3\text{NO}_4\text{SNa}$ ($\text{M}+\text{Na}$) $^+$: 416.0227; found: 416.0233.

2.3.5. Fmoc-(S)- $\beta^2\text{hAsp}(\text{O}^t\text{Bu})$ -(S)- $\beta^2\text{hMet-OCH}_2\text{CCl}_3$ (16). To a solution of compound **15** (0.63 g, 1.60 mmol) in CH_2Cl_2 (6 mL), was added slowly TFA (6 mL) and the mixture stirred for 2 h at 25 °C. After removal of solvent under reduced pressure, the amino ester was coupled with Fmoc-(S)- $\beta^2\text{hAsp}(\text{O}^t\text{Bu})\text{-OH}$ ⁵⁶ (0.68 g, 1.60 mmol) according to GP1a. FC (AcOEt/hexane 1:1) yielded **16** (0.57 g, 51%) as a white solid. $R_f=0.33$ (AcOEt/hexane 1:1); $[\alpha]_D^{25}=+12.8$ ($c=0.49$, CHCl_3); IR (CHCl_3) ν_{max} 3445 (w), 3008 (w), 2974 (w), 2923 (w), 1720 (s), 1667 (m), 1513 (m), 1450 (m), 1368 (m), 1153 (s), 1077 (w), 1046 (w), 841 (w) cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 1.44 (s, 9H, ^tBu), 1.80 (m, 1H, CHHS), 2.03 (m, 1H, CHHS), 2.08 (s, 3H, CH_3S), 2.37 (m, 1H, CH), 2.54 (m, 3H, CH and CH_2), 2.81–3.05 (m, 2H, CH_2), 3.29–3.57 (m, 4H, $2\times\text{CH}_2\text{N}$), 4.20 (t, $J=6.2$ Hz, 1H, CHCH_2O), 4.39 (d, $J=6.2$ Hz, 2H, CHCH_2O), 4.67–4.83 (m, 2H, CH_2CCl_3), 5.46 (br s, 1H, NH), 6.57 (br s, 1H, NH), 7.26–7.42 (m, 4H, arom.), 7.59 (d, $J=7.1$ Hz, 2H, arom.), 7.76 (d, $J=7.2$ Hz, 2H, arom.); ^{13}C NMR (75 MHz, CDCl_3) δ 15.4, 28.1, 28.7, 31.4, 35.7, 39.9, 42.2, 42.6, 44.1, 47.3, 66.7, 68.4, 74.1, 81.4, 119.9, 125.0, 126.9, 127.6, 141.2, 143.7, 156.4, 171.3, 172.2, 173.6; MALDI HRMS calcd for $\text{C}_{32}\text{H}_{39}\text{Cl}_3\text{N}_2\text{O}_7\text{SNa}$ ($\text{M}+\text{Na}$) $^+$: 723.1436; found: 723.1444.

2.3.6. Fmoc-(S)- $\beta^2\text{hAsp}(\text{O}^t\text{Bu})$ -(S)- $\beta^2\text{hMet-OH}$ (5). Dipeptide ester **16** (0.86 g, 1.22 mmol) was dissolved in a mixture of AcOH (50 mL) and H_2O (5 mL). To the cooled solution

(ice bath), Zn powder (3.99 g, 61 mmol, 50 equiv.) was added in portions over 2 h. The reaction mixture was allowed to warm up to 25 °C and stirred for 3 h. Zn was removed by filtration and the filtrate diluted with H_2O and extracted with AcOEt (3 $\times$). The combined organic extracts were dried (MgSO_4) and the solvent removed under reduced pressure. FC (AcOEt/hexane/AcOH 10:10:0.1) of the crude product yielded **5** (0.65 g, 93%) as a white solid; mp 143–144 °C; $R_f=0.37$ (AcOEt/hexane/AcOH 10:10:1); $[\alpha]_D^{25}=+15.9$ ($c=0.43$, CHCl_3); IR (CHCl_3) ν_{max} 3436 (w), 3005 (m), 2974 (m), 1720 (s), 1667 (m), 1512 (m), 1450 (m), 1368 (m), 1154 (m), 1077 (w), 841 (w) cm^{-1} ; ^1H NMR (300 MHz, CD_3OD) δ 1.43 (s, 9H, ^tBu), 1.84 (m, 2H, CH_2S), 2.05 (s, 3H, CH_3S), 2.32 (m, 1H, CH), 2.47–2.58 (m, 3H, CH and CH_2), 2.77 (m, 2H, CH_2), 3.15–3.43 (m, 4H, $2\times\text{CH}_2$), 4.21 (t, $J=6.6$ Hz, 1H, CHCH_2O), 4.36 (d, $J=6.5$ Hz, 2H, CHCH_2O), 7.11 (m, 1H, NH), 7.28–7.40 (m, 4H, arom.), 7.64 (d, $J=7.2$ Hz, 2H, arom.), 7.79 (d, $J=7.5$ Hz, 2H, arom.), 8.05 (br s, 1H, NH); ^{13}C NMR (75 MHz, CD_3OD) δ 15.0, 28.1, 29.9, 32.1, 36.0, 41.2, 43.6, 44.0, 45.3, 47.8, 67.5, 81.6, 120.5, 125.8, 127.7, 128.3, 142.1, 144.8, 144.9, 172.0, 175.0, 176.8; MALDI HRMS calcd for $\text{C}_{30}\text{H}_{38}\text{N}_2\text{O}_7\text{SNa}$ ($\text{M}+\text{Na}$) $^+$: 593.2292, found 593.2299 (Fig. 24).

2.3.7. Fmoc-(S)- $\beta^2\text{hHis}(\text{Trt})$ -(S)- $\beta^2\text{hLeu-OBn}$ (17). Fmoc-(S)- $\beta^2\text{hHis}(\text{Trt})\text{-OH}$ ⁵⁷ (0.72 g, 1.34 mmol) and $p\text{TsOH-H}-(\text{S})\beta^2\text{hLeu-OBn}$ ⁶⁵ (0.49 g, 1.41 mmol) were coupled according to GP1a in the presence of 4 equiv. of NMM. The crude product was purified by FC (AcOEt/hexane 3:1) to yield 0.96 g (84%) of **17** as an amorphous solid. $R_f=0.12$ (AcOEt/hexane 3:1). $[\alpha]_D^{25}=-2.5$ ($c=0.75$, CHCl_3); IR (CHCl_3) ν_{max} 3734 (w), 3628 (w), 3063 (w), 2923 (w), 2853 (w), 1718 (s), 1652 (m), 1539 (m), 1495 (w), 1448 (s), 1169 (m), 1245 (m), 1139 (m), 1085 (w), 843 (m) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 0.76 (d, $J=6.3$ Hz, 3H, Me); 0.77 (d, $J=6.3$ Hz, 3H, Me) 1.18–1.23 (m, 1H, $\text{CHH}(i\text{-Pr})$), 1.37–1.47 (m, 2H, $\text{CHH}(i\text{-Pr})$, Me_2CH); 2.52–2.62 (m, 4H, $2\times\text{CHCO}$ and CH_2Ar), 2.66–2.73 (m, 1H, CHHN); 2.97–3.06 (m, 1H, CHHN), 3.08–3.15 (m, 1H, CHHN), 3.20–3.26 (m, 1H, CHHN), 4.16–4.25 (m, 3H, OCH_2CH), 5.00–5.10 (m, 2H, CH_2Ph), 6.60 (s, 1H, arom.), 7.05–7.07 (m, 6H, arom.), 7.21 (s, 1H, arom.), 7.27–7.41 (m, 19H, 18 arom., NH), 7.66–7.68 (m, 2H, arom.), 7.88–7.99 (m, 2H, arom.), 8.02 (s, 1H, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 21.7, 22.7, 25.5, 28.6, 38.3, 40.6, 42.1, 43.2, 45.7, 46.6, 65.5, 74.3, 118.3, 120.0, 125.1, 125.1, 126.9, 127.5, 127.8, 127.8, 128.0, 128.3, 129.1, 136.0, 137.2, 138.4, 140.6, 142.2, 143.7, 156.0, 173.0, 173.9. MALDI HRMS calcd for $\text{C}_{55}\text{H}_{54}\text{N}_4\text{O}_5\text{Na}$ ($\text{M}+\text{Na}$) $^+$: 873.3992; found: 873.3986.

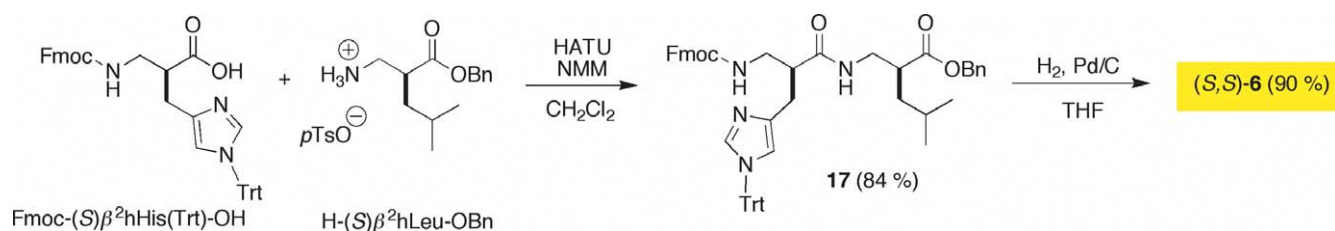


Figure 25. Preparation of Fmoc-(S)- $\beta^2\text{hHis}(\text{Trt})$ -(S)- $\beta^2\text{hLeu-OH}$ (**6**).

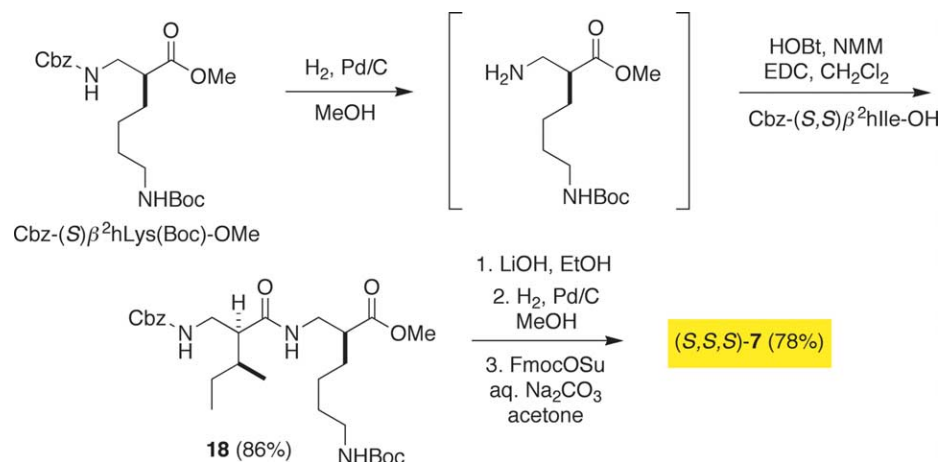


Figure 26. Preparation of Fmoc-(S,S)β²hIle-(S)β²hLys(Boc)-OH (7).

2.3.8. Fmoc-(S)β²hHis(Trt)-(S)β²hLeuOH (6). The C-terminus of the fully protected dipeptide **17** (0.96 g, 1.14 mmol) was deprotected according to GP2 in THF for 8 h. The crude product was purified by FC (CH₂Cl₂/MeOH 9:1) to yield 0.78 g (90%) of **6** as an amorphous solid. $R_f=0.18$ (CH₂Cl₂/MeOH 9:1); $[\alpha]_D^{25}=+10.6$ ($c=0.43$, CHCl₃); IR (CHCl₃) ν_{max} 3297 (w), 3061 (w), 2922 (w), 2854 (w), 1719 (s), 1649 (s), 1551 (s), 1449 (s), 1245 (m), 1132 (m), 1001 (w), 842 (m) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 0.77 (d, $J=6.4$ Hz, 3H, Me); 0.79 (d, $J=6.4$ Hz, 3H, Me) 1.12–1.14 (m, 1H, CHH(*i*-Pr)); 1.36–1.44 (m, 1H, CHH(*i*-Pr)), 1.52–1.53 (m, 1H, Me₂CH), 2.40–2.46 (m, 1H, CHCO), 2.52–2.69 (m, 3H, CH₂Ar, CHCO), 2.96–3.17 (m, 4H, 2×CH₂N), 4.16–4.25 (m, 3H, OCH₂CH), 6.64 (s, 1H, arom.), 7.05–7.07 (m, 6H, arom.), 7.21 (s, 1H, arom.), 7.28–7.44 (m, 14H, 13 arom., NH), 7.67–7.68 (m, 2H, arom.), 7.86–7.88 (m, 2H, arom.), 7.97 (s, NH), 12.35 (br s, 1H, OH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 21.8, 22.8, 25.5, 28.4, 38.5, 40.9, 41.8, 45.4, 46.6, 54.8, 65.3, 74.4, 119.9, 121.3, 127.0, 127.2, 127.5, 127.8, 128.0, 128.8, 129.2, 138.1, 140.6, 140.6, 142.2, 143.8, 156.0, 172.8; MALDI HRMS calcd for C₄₈H₄₈N₄O₅Na (M+Na)⁺: 783.3523; found: 783.3517 (Fig. 25).

2.3.9. Cbz-(S,S)β²hIle-(S)β²hLys(Boc)-OMe (18). The Cbz-protecting group of Cbz-(S)β²hLys(Boc)-OMe⁵⁴ (1.01 g, 2.47 mmol) was hydrogenated according to GP2 and the resulting crude H-(S)β²hLys(Boc)-OMe was coupled with Cbz-(S,S)β²hIle-OH⁵⁵ (0.69 g, 2.47 mmol) according to GP1b. The crude product was purified by FC

(hexane/AcOEt 8:2→5:5) to yield **18** (1.13 g, 86%) as a colorless solid; mp 93–96 °C; $R_f=0.18$ (AcOEt/hexane 1:1); $[\alpha]_D^{25}=+30.3$ ($c=1.0$, CHCl₃); IR (CHCl₃) ν_{max} 3450 (w), 3007 (w), 2968 (m), 2872 (w), 1713 (s), 1666 (m), 1509 (s), 1456 (m), 1367 (m), 1170 (m), 1064 (w) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.87 (t, $J=7.4$ Hz, 3H, Me), 0.91 (d, $J=6.9$ Hz, 3H, Me), 1.08–1.19 (m, 1H, CH), 1.30–1.37 (m, 2H, CH₂), 1.43 (s, 9H, ^tBu), 1.42–1.51 (m, 2H, CH₂), 1.52–1.58 (m, 2H, CH₂), 1.59–1.67 (m, 2H, CH₂), 2.22–2.26 (m, 1H, CH), 2.60–2.66 (m, 1H, CH), 3.07 (d, $J=6.0$ Hz, 2H, CH₂), 3.26–3.35 (m, 2H, CH₂), 3.42–3.54 (m, 2H, CH₂), 3.68 (s, 3H, MeO), 4.55 (br s, 1H, NH), 5.07 (q, $J=12.4$ Hz, 2H, CH₂Ph), 5.35 (br s, 1H, NH), 6.10 (br s, 1H, NH), 7.30–7.36 (m, 5H, arom.); ¹³C NMR (125 MHz, CDCl₃) δ 11.4, 15.9, 23.7, 27.2, 28.4, 28.9, 29.4, 29.7, 29.9, 35.2, 39.9, 40.2, 40.4, 44.8, 45.1, 51.9, 52.0, 58.5, 66.6, 72.3, 79.2, 128.0, 128.5, 136.6, 156.1, 156.5, 174.4, 175.4; MALDI HRMS calcd for C₂₈H₄₅N₃O₇Na (M+Na)⁺: 558.3150; found: 558.3157. Anal. calcd for C₂₈H₄₅N₃O₇: C 62.78, H 8.47, N 7.84; found: C 62.80, H 8.28, N 7.71.

2.3.10. Fmoc-(S,S)β²hIle-(S)β²hLys(Boc)-OH (7). The methyl ester- and the Cbz-protecting group of compound **18** (1.06 g, 1.98 mmol) were removed according to GP3 and GP2 respectively, and the resulting amino acid Fmoc-protected according to GP4. FC (CH₂Cl₂/MeOH 100:1→CH₂Cl₂/MeOH/AcOH 20:1:0.2) yielded **7** (0.94 g, 78%) colorless solid; mp 114–117 °C; $R_f=0.30$ (CH₂Cl₂/MeOH/AcOH 20:1:0.2); $[\alpha]_D^{25}=+29.4$ ($c=1.0$, CHCl₃); IR (CHCl₃) ν_{max} 3449 (w), 3344 (w), 3007 (w), 2969 (m), 2933

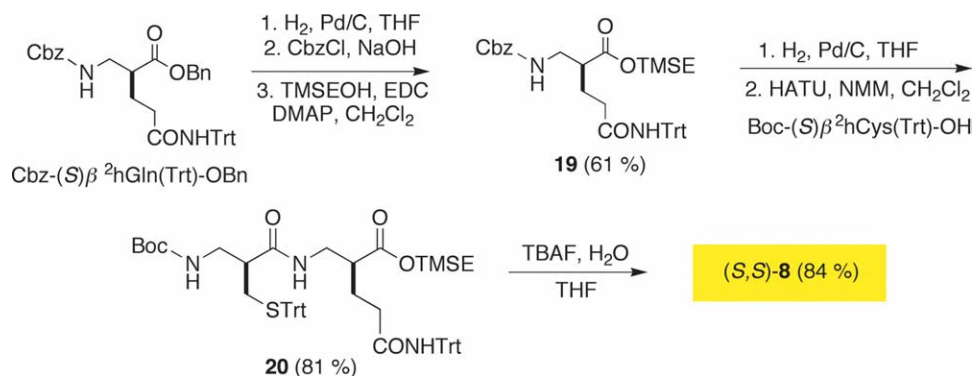


Figure 27. Preparation of Boc-(S)β²hCys(Trt)-(S)β²hGln(Trt)-OH (8).

(m), 2872 (w), 1710 (s), 1662 (m), 1510 (s), 1451 (m), 1368 (m), 1167 (m), 1078 (w), 990 (w) cm^{-1} ; ^1H NMR (500 MHz, CD_3OD) δ 0.89 (t, $J=7.4$ Hz, 3H, Me), 0.94 (t, $J=6.8$ Hz, 3H, Me), 1.10–1.57 (m, 2H, CH_2), 1.29–1.39 (m, 2H, CH_2), 1.41 (s, 9H, ^tBu), 1.42–1.49 (m, 2H, CH_2), 1.51–1.63 (m, 4H, $2\times\text{CH}_2$), 2.29–2.34 (m, 1H, CH), 2.58–2.62 (m, 1H, CH), 3.00 (d, $J=6.9$ Hz, 2H, CH_2), 3.20–3.28 (m, 2H, CH_2), 3.32–3.39 (m, 2H, CH_2), 4.19 (t, $J=6.9$ Hz, 2H, CH_2), 4.31 (d, $J=7.1$ Hz, 2H, CH_2), 6.89 (t, $J=5.9$ Hz, 1H, NH), 7.29–7.80 (m, 8H, arom.); ^{13}C NMR (125 MHz, CD_3OD) δ 11.7, 16.5, 25.3, 28.3, 28.8, 30.7, 30.9, 36.5, 41.1, 41.8, 42.0, 46.6, 48.5, 53.4, 67.9, 73.6, 79.8, 121.0, 126.2, 126.3, 128.2, 128.8, 142.6, 145.3, 145.4, 158.6, 158.7, 176.9, 178.2; MALDI HRMS calcd for $\text{C}_{34}\text{H}_{47}\text{N}_3\text{O}_7\text{Na}$ ($\text{M}+\text{Na}$) $^+$: 632.3306; found: 632.3313. Anal. calcd for $\text{C}_{34}\text{H}_{47}\text{N}_3\text{O}_7$: C 66.97, H 7.77, N 6.89; found: C 66.97, H 7.66, N 6.64 (Fig. 26).

2.3.11. Cbz-(S) β^2 hGln(Trt)-OTMSE (19). Cbz-(S) β^2 hGln(Trt)-OBn⁵⁶ (2.45 g, 3.9 mmol) was transformed to compound **19** in three steps. The first transformation involved hydrogenolysis in MeOH for 1.5 h according to GP2 to yield the amino acid (H- β^2 hGln(Trt)-OH), which without purification was *N*-Cbz protected using the following procedure: to an aqueous solution of the free amino acid in 0.5 M NaOH (1.1 equiv.) at 0 °C was added with continuous stirring BnO_2CCl (1.3 equiv.). The pH was kept basic by periodic addition of 1 M NaOH. After the addition was completed, the reaction mixture was stirred for one additional hour then extracted with AcOEt after adjusting the pH to 3 using 1 M HCl. The organic layer was washed with brine, dried (MgSO_4) and the solvent evaporated under reduced pressure to give a yellow oil. Unreacted BnO_2CCl and BnOH were removed from the mixture by passing the mixture through a short column, and the crude product treated according to GP5. The purification by FC (Et_2O /pentane 1:3) yielded **19** (1.52 g, 61%) as a white solid; mp 135–136 °C; $R_f=0.42$ (Et_2O /pentane 1:3); $[\alpha]_D=-6.8$ ($c=1.0$, CHCl_3); IR (CHCl_3) ν_{max} 3439 (w), 3007 (w), 2956 (w), 1716 (s), 1516 (m), 1490 (m), 1448 (w), 860 (w), 839 (w) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 0.01 (s, 9H, SiMe_3), 0.90–0.95 (m, 2H, CH_2Si); 1.56–1.65 (m, 2H, $\text{CH}_2\text{CH}_2\text{CON}$), 2.24–2.27 (m, 2H, CH_2CON), 2.44–2.47 (m, 1H, CHCO), 3.06–3.19 (m, 2H, CH_2N), 4.06–4.11 (m, 2H, OCH_2), 4.95–5.02 (m, 2H, CH_2Ph), 7.14–7.35 (m, 21H, 20 arom., NH), 8.53 (s, 1H, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ -1.5, 16.8, 25.0, 33.2, 42.0, 44.9, 62.0, 65.2, 69.2, 126.2, 127.4, 127.6, 127.7, 128.3, 128.5, 137.1, 144.9, 156.1, 171.2, 173.5; MALDI HRMS calcd for $\text{C}_{38}\text{H}_{44}\text{N}_2\text{O}_5\text{SiNa}$ ($\text{M}+\text{Na}$) $^+$: 659.2917; found: 659.2912.

2.3.12. Boc-(S) β^2 hCys(Trt)-(S) β^2 hGln(Trt)-OTMSE (20). Compound **19** (1.00 g, 1.57 mmol) was hydrogenated in THF to give H-(S) β^2 hGln(Trt)-OTMSE according to GP2, then coupled with Boc-(S) β^2 hCys(Trt)-OH⁵⁷ (0.75 g, 1.57 mmol) according to GP1a. The crude product was purified by FC (CH_2Cl_2 /*i*-PrOH 96:4) to yield **20** (1.22 g, 81%) as an amorphous solid; $R_f=0.32$ (CH_2Cl_2 /*i*-PrOH 96:4); $[\alpha]_D=+11.4$ ($c=1.0$, CHCl_3); IR (CHCl_3) ν_{max} 3436 (w), 3026 (w), 1708 (s), 1681 (m), 1492 (s), 1446 (w), 1251 (m), 1164 (m), 1041 (w) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 0.00 (s, 9H, SiMe_3) 0.93 (t, $J=8.5$ Hz, 2H,

CH_2Si), 1.31 (s, 9H, ^tBu), 1.61–1.64 (m, 2H, $\text{CH}_2\text{CH}_2\text{CON}$), 2.01–2.03 (m, 1H, CHHS), 2.25–2.28 (m, 2H, CH_2CON), 2.30–2.33 (m, 2H, CHHS and CHCO), 2.47–2.50 (m, 1H, CHCO), 2.86–2.90 (m, 2H, CH_2N), 3.11–3.25 (m, 2H, CH_2N), 4.00–4.14 (m, 2H, OCH_2), 6.49 (s, 1H, NH), 7.15–7.30 (m, 30H, arom.), 7.93 (s, 1H, NH), 8.54 (s, 1H, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ -1.6, 16.8, 25.1, 28.1, 31.1, 33.1, 39.9, 42.0, 44.3, 45.4; 61.9, 65.8, 69.1, 77.5, 126.2, 126.5, 127.3, 127.8, 128.4, 129.0, 144.3, 144.8, 155.2, 171.1, 171.6, 173.5; MALDI HRMS calcd for $\text{C}_{58}\text{H}_{67}\text{N}_3\text{O}_6\text{SSiNa}$ ($\text{M}+\text{Na}$) $^+$: 984.4418; found: 984.4400.

2.3.13. Boc-(S) β^2 hCys(Trt)-(S) β^2 hGln(Trt)-OH (8). The C-terminus of the fully protected dipeptide **20** (1.32 g, 1.37 mmol) was deprotected according to GP2. The crude product was purified by FC (AcOEt/Hexane/AcOH 20:10:0.1) to yield **8** (1.00 g, 85%) as an amorphous solid. $R_f=0.10$ (AcOEt/hexane/AcOH 20:10:0.1); $[\alpha]_D=-6.3$ ($c=0.9$, CHCl_3); IR (CHCl_3) ν_{max} 3432 (w), 3058 (w), 3009 (w), 2980 (w), 1706 (s), 1667 (m), 1494 (s), 1445 (m), 1366 (w), 1228 (m), 1164 (m) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 1.31 (s, 9H, ^tBu), 1.62–1.66 (m, 2H, $\text{CH}_2\text{CH}_2\text{CON}$), 2.02–2.05 (m, 1H, CHHS), 2.28–2.33 (m, 4H, CHHS, CH_2CON , and CHCO), 2.37–2.43 (m, 1H, CHCO), 2.84–2.88 (m, 2H, CH_2N), 3.06–3.10 (m, 1H, CHHN), 3.25–3.31 (m, 1H, CHHN), 6.50 (t, $J=5.8$ Hz, 1H, NH), 7.15–7.32 (m, 30H, arom.), 7.89 (t, $J=5.8$ Hz, 1H, NH), 8.57 (s, 1H, NH), 12.30 (br s, 1H, OH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 25.1, 28.1, 31.1, 33.4, 41.3, 42.1, 44.3, 45.4, 65.8, 69.1, 77.5, 126.2, 126.5, 127.3, 127.8, 128.4, 129.0, 144.3, 144.8, 155.3, 171.3, 171.6, 175.2; MALDI HRMS calcd for $\text{C}_{53}\text{H}_{55}\text{N}_3\text{O}_6\text{SNa}$ ($\text{M}+\text{Na}$) $^+$: 884.3708; found: 884.3704 (Fig. 27).

2.3.14. Cbz-(S) β^2 hTyr(^tBu)-(S) β^2 hAla-OBn (21). H-(S) β^2 hAla-OH⁸⁹ was dissolved in toluene, BnOH and *p*TsOH were added and the mixture was heated at reflux for 18 h using a Dean–Stark trap to azeotropically remove H_2O . The reaction mixture was cooled to 25 °C, toluene was evaporated and the residue washed several times with Et_2O . The resulting benzyl ester (0.92 g, 2.52 mmol) was coupled with Cbz-(S) β^2 hTyr(^tBu)-OH⁵⁵ (0.97 g, 2.52 mmol) according to GP1a. FC (AcOEt/hexane 1:1) yielded **21** (1.17 g, 83%) as a white solid; mp 74–76 °C; $R_f=0.18$ (AcOEt/hexane 1:1); $[\alpha]_D=+14.8$ ($c=0.54$, CHCl_3); IR (CHCl_3) ν_{max} 3443 (w), 2980 (m), 1719 (s), 1667 (m), 1507 (s), 1456 (m), 1367 (m), 1160 (m), 894 (m) cm^{-1} ; ^1H NMR (400 MHz, CD_3OD) δ 0.96 (d, $J=7.1$ Hz, 3H, Me), 1.28 (s, 9H, ^tBu), 2.54 (q, $J=7.0$ Hz, 1H, CHCO), 2.66–2.93 (m, 3H, CHCH_2Ph), 3.11–3.29 (m, 4H, $2\times\text{CH}_2\text{N}$), 5.05 (s, 2H, OCH_2Ph), 5.07 (AB, $J=12.6$ Hz, 2H, OCH_2Ph), 6.85 (d, $J=8.4$ Hz, 2H, arom.), 7.04 (d, $J=8.4$ Hz, 2H, arom.), 7.26–7.34 (m, 10H, arom.); ^{13}C NMR (100 MHz, CD_3OD) δ 15.2, 29.2, 36.5, 40.8, 42.9, 44.1, 50.4, 67.4, 67.5, 79.4, 125.2, 128.8, 129.0, 129.1, 129.2, 129.5, 129.6, 130.4, 135.5, 137.6, 138.3, 155.0, 158.8, 176.0; MALDI HRMS calcd for $\text{C}_{33}\text{H}_{40}\text{N}_2\text{O}_6\text{Na}$ ($\text{M}+\text{Na}$) $^+$ 583.2779, found 583.2785. Anal. calcd for $\text{C}_{33}\text{H}_{40}\text{N}_2\text{O}_6$: C 70.69, H 7.19, N 5.00; found: C 70.52, H 7.34, N 5.14.

2.3.15. Fmoc-(S) β^2 hTyr(^tBu)-(S) β^2 hAla-OH (9). The dipeptide derivative **21** was hydrogenated according to

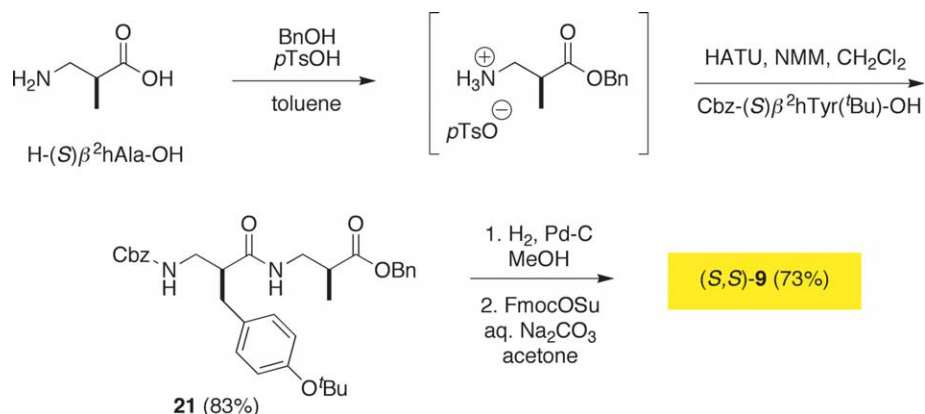


Figure 28. Preparation of Fmoc-(S,S)-β²hTyr(ᵀBu)-(S)-β²hAla-OH (**8**).

GP2 and Fmoc-protected according to GP4. The crude peptide was purified by FC (AcOEt/hexane/AcOH 10:10:0.1) to give **9** (0.72 g, 73%) as a white solid; mp 143–144 °C; R_f =0.16 (AcOEt/hexane/AcOH 10:10:1); $[\alpha]_D^{25}$ =+7.3 (c =0.55, CHCl₃); IR (CHCl₃) $\nu_{\max}$ 3442 (w), 3005 (m), 2980 (m), 1717 (s), 1683 (m), 1508 (s), 1467 (m), 1450 (m), 1368 (m), 1157 (m), 1082 (w), 891 (m) cm⁻¹; ¹H NMR (400 MHz, CD₃OD) δ 0.98 (d, J =7.1 Hz, 3H, Me), 1.29 (s, 9H, ^tBu), 2.45 (q, J =7.1 Hz, 1H, CHMe), 2.67–2.80 (m, 3H, CHHN, CH₂Ph), 3.15 (dd, J =7.0, 13.4 Hz, 1H, CH), 3.26 (m, 3H, CH₂N, CHHN), 4.19 (t, J =6.8 Hz, 1H, CHCH₂O), 4.34 (d, J =6.9 Hz, 2H, CHCH₂O), 6.87 (d, J =8.5 Hz, 2H, arom.), 7.07 (d, J =8.4 Hz, 2H, arom.), 7.29 (m, 2H, arom.), 7.37 (m, 2H, arom.), 7.63 (d, J =7.5 Hz, 2H, arom.), 7.77 (m, 2H, arom.); ¹³C NMR (100 MHz, CD₃OD) δ 15.4, 29.2, 36.5, 40.6, 43.0, 44.2, 48.5, 50.4, 67.8, 79.5, 121.0, 125.3, 126.2, 128.2, 128.8, 130.5, 135.7, 142.7, 145.4, 155.0, 158.9, 176.1, 178.4; MALDI HRMS calcd for C₃₃H₃₈N₂O₆Na (M+Na)⁺: 581.2622, found: 581.2629. Anal. calcd for C₃₃H₃₈N₂O₆: C 70.95, H 6.86, N 5.01; found: C 70.99, H 6.87, N 4.89 (Fig. 28).

2.3.16. Cbz-(R,S)-β²hThr(ᵀBu)-(R)-β²hSer(ᵀBu)-OBn (**22**).

Cbz-(R,S)-β²hThr(ᵀBu)-OH⁵⁷ (0.70 g, 2.16 mmol) and H-(R)-β²hSer(ᵀBu)-OBn⁵⁷ (0.6 g, 2.27 mmol) were coupled according to GP1a. The crude product was purified by FC (AcOEt/hexane 3:4) to obtain 0.97 g (79%) of **22** as an amorphous solid. R_f =0.35 (AcOEt/hexane 3:4); $[\alpha]_D^{25}$ =+27.6 (c =0.41, CHCl₃); IR (CHCl₃) $\nu_{\max}$ 3323 (w), 2972 (m), 2925 (w), 1726 (s), 1653 (m), 1534 (m), 1457 (m), 1233 (m), 1193 (m), 1091 (w) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 0.97 (d, J =4.8 Hz, 3H, Me); 1.05 (s, 9H, ^tBu), 1.10 (s, 9H, ^tBu), 2.36–2.40 (m, 1H, CHCO), 2.74–2.79 (m, 1H, CHCO), 3.10–3.27 (m, 3H, CH₂N, OCHH), 3.33–3.37 (m, 1H, OCHH), 3.44–3.51 (m, 2H, CH₂N), 3.68–3.70 (m, 1H, OCH), 4.95–5.17 (m, 4H, 2×CH₂Ph), 7.03 (t, J =4.6 Hz, 1H, NH) 7.27–7.37 (m, 10H, arom.); 7.72 (t,

J =4.6 Hz, 1H, NH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 20.1, 27.0, 28.2, 37.3, 39.7, 46.1, 52.6, 60.3, 65.0, 65.2, 66.2, 72.0, 73.2, 127.4, 127.6, 127.6, 127.8, 128.2, 128.2, 136.1, 137.2, 155.9, 171.7, 172.2; MALDI HRMS calcd for C₃₂H₄₆N₂O₇Na: 593.3203; found: 593.3197.

2.3.17. Fmoc-(R,S)-β²hThr(ᵀBu)-(R)-β²hSer(ᵀBu)-OH (**10**).

The fully protected dipeptide **22** (0.9 g, 1.58 mmol) was hydrogenolyzed in MeOH for 16 h according to GP2. The resulting amino acid was Fmoc-protected according to GP4. The crude product was purified by FC (AcOEt/Hexane/AcOH 10:10:0.1) to obtain 0.73 g (81%) of **22** as colorless oil. R_f =0.20 (AcOEt/hexane/AcOH 10:10:0.1). $[\alpha]_D^{25}$ =+12.7 (c =0.52, CHCl₃); IR (CHCl₃) $\nu_{\max}$ 3312 (w), 2971 (m), 2925 (w), 1718 (s), 1653 (m), 1539 (m), 1450 (m), 1363 (m), 1249 (m), 1193 (m), 1091 (w) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.00 (d, J =4.8 Hz, 3H, Me), 1.08 (s, 9H, ^tBu), 1.12 (s, 9H, ^tBu), 2.36–2.43 (m, 1H, CHCO), 2.53–2.60 (m, 1H, CHCO), 3.12–3.22 (m, 3H, CH₂N, OCHH), 3.35–3.41 (m, 1H, OCHH), 3.45 (d, J =4.6 Hz, 2H, CH₂N); 3.69–3.74 (m, 1H, OCH), 4.18–4.27 (m, 3H, OCH₂CH), 7.12 (t, J =4.5 Hz, 1H, NH), 7.30–7.34 (m, 2H, arom.), 7.39–7.43 (m, 2H, arom.), 7.62 (t, J =4.5 Hz, 1H, NH), 7.67 (d, J =5.8 Hz, 2H, arom.), 7.88 (d, J =6.1 Hz, 2H, arom.), 12.22 (br s, 1H, OH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 20.1, 27.1, 28.1, 37.4, 39.9, 46.0, 46.6, 52.4, 60.4, 65.4, 66.3, 72.2, 73.3, 120.0, 125.1, 126.9, 127.5, 140.6, 143.8, 155.9, 171.6, 173.8; MALDI HRMS calcd for C₃₂H₄₄N₂O₇Na: 591.3047; found: 591.3041 (Fig. 29).

2.3.18. Cbz-(S)-β²hGlu(O^tBu)-OTMSE (**23**).

Cbz-(S)-β²hGlu(O^tBu)-OH⁵⁶ (0.9 g, 2.56 mmol) was treated according to GP5. Purification of the crude product by FC (Et₂O/pentane 1:1) gave **23** (1.06 g, 92%) as a colorless oil. R_f =0.43 (Et₂O/pentane 1:1). $[\alpha]_D^{25}$ =+4.7 (c =0.9, CHCl₃); IR (CHCl₃) $\nu_{\max}$ 3446 (w), 3025 (w), 2954 (w), 1718 (s), 1508 (m), 1451 (w), 1364 (w), 1246 (m), 1148 (m), 1056

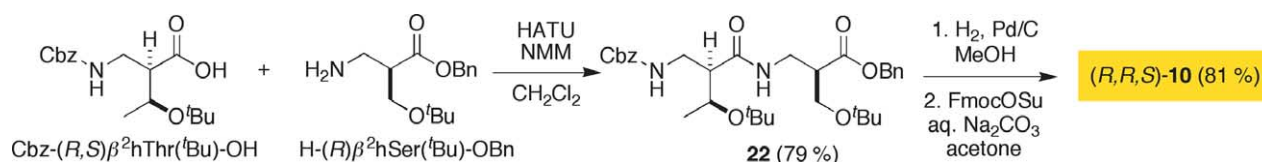


Figure 29. Preparation of Fmoc-(R,R,S)-β²hThr(ᵀBu)-(R)-β²hSer(ᵀBu)-OH (**10**).

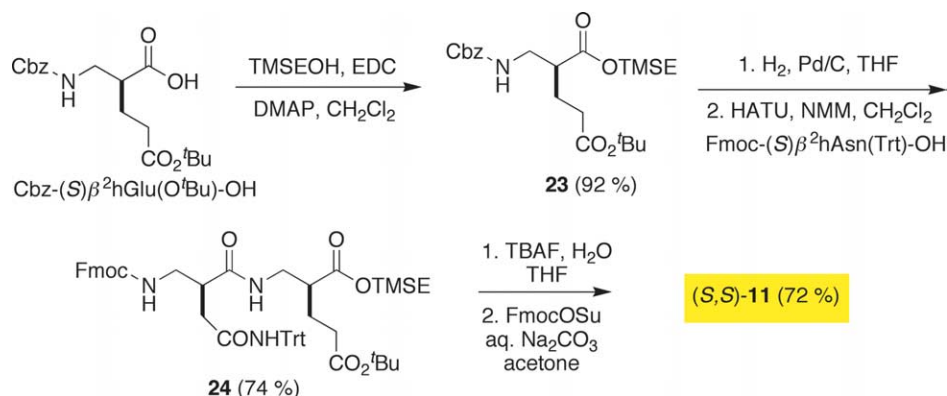


Figure 30. Preparation of Fmoc-(S)β²hAsn(Trt)-(S)β²hGlu(OᵀBu)-OH (**11**).

(w), 856 (w) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 0.02 (s, 9H, SiMe_3), 0.91–0.95 (m, 2H, CH_2Si), 1.39 (s, 9H, ^tBu), 1.62–1.73 (m, 2H, $\text{CH}_2\text{CH}_2\text{CO}$), 2.11–2.26 (m, 2H, CH_2CO), 2.24–2.27 (m, 2H, CH_2CON), 2.51–2.55 (m, 1H, CHCO), 3.07–3.23 (m, 2H, CH_2N), 4.07–4.11 (m, 2H, OCH_2), 5.00 (s, 2H, CH_2Ph), 7.28–7.40 (m, 6H, 5 arom., NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ -1.6, 16.7, 24.2, 27.6, 32.1, 41.7, 44.4, 62.0, 65.1, 79.6, 127.5, 127.7, 128.2, 137.0, 156.0, 171.4, 173.2; MALDI HRMS calcd for $\text{C}_{23}\text{H}_{37}\text{NO}_6\text{SiNa}$ ($\text{M}+\text{Na}$) $^+$: 474.2388; found: 474.2382.

2.3.19. Fmoc-(S)β²hAsn(Trt)-(S)β²hGlu(OᵀBu)-OTMSE (24). Compound **23** (0.93 g, 2.06 mmol) was hydrogenated according to GP2 in THF. Coupling with Fmoc-(S)β²hAsn(Trt)-OH⁵⁶ (1.26 g, 2.06 mmol) according to GP1a give a crude product, which was purified by FC ($\text{AcOEt}/\text{hexane}$ 1:1) to yield **24** (1.39 g, 74%) as an amorphous solid. $R_f=0.23$ ($\text{AcOEt}/\text{hexane}$ 1:1). $[\alpha]_D=-16.0$ ($c=0.43$, CHCl_3); IR (CHCl_3) ν_{max} 3436 (w), 3019 (w), 1716 (s), 1677 (m), 1504 (m), 1445 (w), 1248 (w), 1154 (m) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ -0.01 (s, 9H, SiMe_3), 0.89–0.94 (m, 2H, CH_2Si), 1.34 (s, 9H, ^tBu), 1.64–1.71 (m, 2H, $\text{CH}_2\text{CH}_2\text{CO}$), 2.07–2.21 (m, 2H, CH_2CO), 2.27–2.33 (m, 1H, CHCO), 2.52–2.59 (m, 2H, CHHCON and CHCO), 2.71–2.75 (m, 1H, CHHCON), 2.99–3.05 (m, 1H, CHHN), 3.06–3.08 (m, 2H, CH_2N), 3.33–3.39 (m, 1H, CHHN), 4.03–4.13 (m, 2H, OCH_2), 4.18–4.29 (m, 3H, OCH_2CH), 7.13–7.41 (m, 20H, 19

arom., NH), 7.66 (d, $J=7.5$ Hz, 2H, arom.), 7.87 (d, $J=7.6$ Hz, 2H, arom.), 7.89 (s, 1H, NH), 8.52 (s, 1H, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ -1.6, 16.7, 24.1, 27.6, 32.0, 33.1, 36.0, 40.1, 42.3, 42.4, 43.9, 46.6, 62.0, 65.4, 69.2, 79.5, 119.9, 121.3, 125.1, 126.9, 127.2, 127.3, 128.4, 140.6, 143.8, 144.8, 155.9, 170.1, 171.5, 172.7, 173.4; MALDI HRMS calcd for $\text{C}_{54}\text{H}_{63}\text{N}_3\text{O}_8\text{SiNa}$ ($\text{M}+\text{Na}$) $^+$: 932.4282; found: 932.4277.

2.3.20. Fmoc-(S)β²hAsn(Trt)-(S)β²hGlu(OᵀBu)-OH (**11**).

The C-terminus of dipeptide derivative **24** (1.15 g, 1.27 mmol) was deprotected according to GP6; however, considerable amounts of Fmoc-deprotection took place during the reaction. For this reason, the crude reaction mixture was evaporated under reduced pressure and treated in situ with FmocOSu (1.2 equiv.) according to GP4. The crude product was purified by FC ($\text{AcOEt}/\text{CH}_2\text{Cl}_2$ 1:7→1:3) to give **11** (0.74 g, 72%) as an amorphous solid. $R_f=0.23$ ($\text{AcOEt}/\text{hexane}$ 1:1); $[\alpha]_D=-11.2$ ($c=0.75$, CHCl_3); IR (CHCl_3) ν_{max} 3426 (w), 3343 (w), 3015 (w), 2985 (w), 1718 (s), 1677 (m), 1508 (m), 1492 (m), 1446 (w), 1369 (w), 1246 (m), 1154 (m) cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 1.35 (s, 9H, ^tBu), 1.62–1.74 (m, 2H, $\text{CH}_2\text{CH}_2\text{CO}$), 2.11–2.25 (m, 2H, CH_2CO), 2.28–2.32 (m, 1H, CHCO), 2.43–2.48 (m, 1H, CHCO), 2.55–2.60 (m, 1H, CHHCON), 2.72–2.78 (m, 1H, CHHCON), 2.99–3.06 (m, 1H, CHHN), 3.07–3.14 (m, 2H, CH_2N), 3.35–3.41 (m, 1H, CHHN), 4.19–4.29 (m, 3H, OCH_2CH), 7.13–7.42 (m, 21H, 19 arom., 2×NH), 7.68 (d, $J=6.0$ Hz, 2H, arom.), 7.88 (d, $J=5.8$ Hz, 2H, arom.),

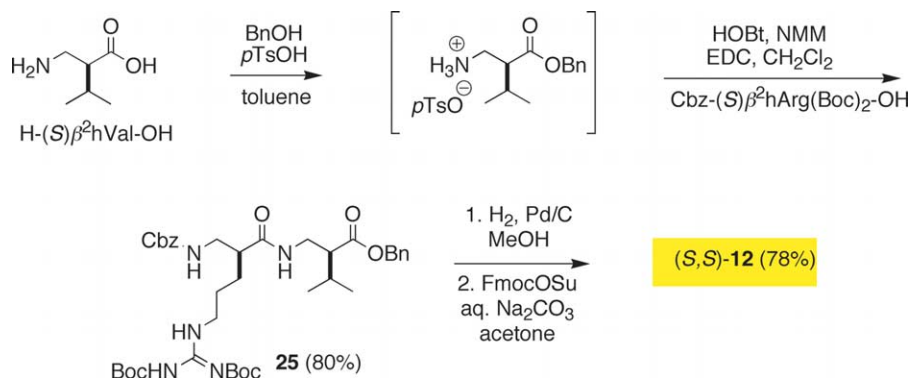


Figure 31. Preparation of Fmoc-(S)β²hArg(Boc)₂-(S)β²hVal-OH (**8**).

8.53 (s, 1H, NH), 12.30 (br s, 1H, OH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 24.2, 27.6, 32.2, 36.1, 40.0, 42.4, 42.4, 43.8, 46.6, 65.4, 69.2, 79.5, 120.0, 125.1, 126.2, 127.0, 127.3, 127.5, 128.4, 140.6, 143.8, 144.8, 156.0, 170.1, 171.6, 172.7, 175.0; MALDI HRMS calcd for $\text{C}_{49}\text{H}_{51}\text{N}_3\text{O}_8\text{Na}$ ($\text{M}+\text{Na}$) $^+$: 832.3598; found: 832.3568 (Fig. 30).

2.3.21. Cbz-(S) β^2 hArg(Boc) $_2$ -(S) β^2 hVal-OBn (25). H-(S) β^2 hVal-OH⁸⁹ was dissolved in toluene, BnOH and *p*TsOH were added and mixture was refluxed for 18 h using a Dean–Stark trap to azeotropically remove H_2O . The reaction mixture was cooled to 25 °C, and the toluene removed under reduced pressure. The crude product was purified by recrystallization from toluene/hexane to give the *p*TsOH-salt of H-(S) β^2 hVal-OBn (0.72 g, 1.82 mmol) which was coupled with Cbz-(S) β^2 hArg(Boc) $_2$ -OH⁴⁸ (0.95 g, 1.82 mmol) according to GP1b. FC (AcOEt/hexane 3:7) yielded **25** (1.06 g, 80%) as white solid; mp 45–47 °C; R_f =0.22 (AcOEt/hexane 1:1); $[\alpha]_D^{25}$ =+16.8 (c =0.37, CHCl_3); IR (CHCl_3) ν_{max} 3446 (w), 3328 (w), 3007 (m), 2974 (m), 2933 (w), 1719 (s), 1636 (s), 1615 (s), 1574 (m), 1509 (m), 1418 (m), 1369 (m), 1332 (m), 1282 (m), 1136 (s), 1051 (m), 1027 (m), 872(w), 648(w) cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 0.90 (d, J =6.8 Hz, 3H, Me), 0.94 (d, J =6.8 Hz, 3H, Me), 1.41 (m, 2H, CH_2), 1.47 (s, 9H, 'Bu), 1.49 (s, 9H, 'Bu), 1.55 (m, 1H, CHH), 1.78 (m, 1H, CHH), 1.93 (m, 1H, CH(Me) $_2$), 2.47 (m, 1H, CHCO), 2.50 (m, 1H, CHCO), 3.21–3.38 (m, 5H, 2 $\times$ CH $_2$ and CHH), 3.60 (m, 1H, CHH), 5.04 (d, J =12.2 Hz, 1H, OCHHPh), 5.08 (s, 2H, OCH $_2$ Ph), 5.15 (d, J =12.2 Hz, 1H, OCHHPh), 5.32 (br s, 1H, NH), 6.30 (br s, 1H, NH), 7.28–7.52 (m, 10H, arom.), 8.32 (br s, 1H, NH), 11.48 (br s, 1H, NH); ^{13}C NMR (100 MHz, CDCl_3) δ 20.0, 20.3, 26.6, 27.1, 28.1, 28.3, 28.9, 31.3, 38.8, 40.1, 42.8, 45.8, 51.9, 66.3, 66.6, 79.3, 83.2, 128.0, 128.1, 128.4, 128.5, 128.6, 135.8, 136.6, 153.3, 156.3, 156.6, 163.4, 174.2, 174.4; MALDI HRMS calcd for $\text{C}_{38}\text{H}_{55}\text{N}_5\text{O}_9\text{Na}$ ($\text{M}+\text{Na}$) $^+$: 748.3892; found: 748.3901. Anal. calcd for $\text{C}_{38}\text{H}_{55}\text{N}_5\text{O}_9$: C 62.88, H 7.64, N 9.65; found: C 62.92, H 7.68, N 9.51.

2.3.22. Fmoc-(S) β^2 hArg(Boc) $_2$ -(S) β^2 hVal-OH (12). Dipeptide derivative **25** (1.06 g) was hydrogenated according to GP2. Subsequent Fmoc-protection according to GP4 and FC (AcOEt/hexane 1:1 $\rightarrow$ 1:0) yielded **12** (0.99 g, 78%) as white solid; mp 93–96 °C; R_f =0.46 (AcOEt/hexane/AcOH 10:10:1); $[\alpha]_D^{25}$ =+29.3 (c =0.91, CHCl_3); IR (CHCl_3) ν_{max} 3446 (w), 3326 (w), 3007 (m), 2972 (m), 2934 (m), 1719 (s), 1643 (s), 1615 (s), 1513 (m), 1450 (m), 1416 (m), 1369 (m), 1333 (m), 1282 (m), 1138 (s), 1053 (m), 872 (w), 650 (w) cm^{-1} ; ^1H NMR (400 MHz, DMSO- d_6) δ 0.87 (d, J =6.8 Hz, 3H, Me), 0.91 (d, J =6.8 Hz, 3H, Me), 1.35–1.61 (m, 4H, CH $_2$ CH $_2$), 1.39 (s, 9H, 'Bu), 1.46 (s, 9H, 'Bu), 1.79 (m, 1H, CH(Me) $_2$), 2.30 (m, 1H, CHCO), 2.38 (br s, 1H, NH), 2.49 (m, 1H, CH), 2.99–3.10 (m, 3H, CHH, CH $_2$), 3.23 (m, 2H, CH $_2$), 3.35 (m, 1H, CHH), 4.17–4.28 (m, 3H, CHCH $_2$ O), 7.13–7.36 (m, 3H, 2 arom., NH), 7.41 (t, J =7.4 Hz, 2H, arom.), 7.68 (d, J =7.4 Hz, 2H, arom.), 7.88 (d, J =7.5 Hz, 2H, arom.), 7.94 (br s, 1H, NH), 8.24 (br s, 1H, NH), 11.48 (br s, 1H, COOH); ^{13}C NMR (100 MHz, DMSO- d_6) δ 19.5, 19.7, 20.2, 20.9, 26.1, 26.2, 26.8, 27.5, 27.9, 28.0, 31.2, 38.7, 42.6, 78.0, 82.8, 109.6, 119.9, 121.3, 127.2, 128.8, 137.3, 139.3, 142.5, 152.0, 155.1, 163.0,

173.1, 173.2; MALDI HRMS calcd for $\text{C}_{38}\text{H}_{53}\text{N}_5\text{O}_9\text{Na}$ ($\text{M}+\text{Na}$) $^+$: 746.3736, found: 746.3728. Anal. calcd for $\text{C}_{38}\text{H}_{53}\text{N}_5\text{O}_9$: C 63.05, H 7.38, N 9.67; found: C 63.17, H 7.40, N 9.44 (Fig. 31).

2.4. Solid-phase synthesis of **2** and **3** and their ligation to eicosapeptide **1**

2.4.1. H- β^2 hGly-(S) β^2 hTrp-(S) β^2 hArg-(S) β^2 hVal-(S) β^2 hAsn-(S) β^2 hGlu-(R,S) β^2 hThr-(R) β^2 hSer-(S) β^2 hTyr-(S) β^2 hAla-S-Ethylpropionate (2**).** The loading of the sulfamylbutyryl resin was performed according to a procedure found in the literature.⁹⁰ A solution of Fmoc-(S) β^2 hTyr('Bu)-(S) β^2 hAla-OH (**9**) (502 mg, 0.9 mmol), DIPCDI (0.5 mL, 3.6 mmol) and 1-MeIm (0.2 mL, 3.6 mmol) in CH_2Cl_2 /DMF (4:1) was added to the resin (205 mg, 0.225 mmol) that had been preswelled in CH_2Cl_2 for 1 h. The suspension was gently stirred using Ar bubbling for 18 h at 25 °C. Consequently, the resin was filtered off, washed with DMF (4 mL, 4 $\times$ 1 min), CH_2Cl_2 (4 mL, 4 $\times$ 1 min), and dried under h.v. overnight. Resin loading was measured using the absorbance of the benzofulvene–piperidine adduct according to Schreiber and Seebach⁸⁰ and was determined to be 0.66 mmol/g, which corresponds to 0.133 mmol of **9**. The Fmoc group was removed using 20% piperidine in DMF (4 mL, 4 $\times$ 10 min) under Ar bubbling. After filtration, the resin was washed with DMF (4 mL, 4 $\times$ 1 min). Solid phase synthesis was continued by sequential incorporation of *N*-Fmoc-protected dipeptides or *N*-Fmoc protected β^2 -homoamino acids building blocks (**10**, **11**, **12**, Fmoc-(S) β^2 hTrp(Boc)-OH, Boc- β^2 hGly-OH). For each coupling step, the resin was treated with a solution of Fmoc-protected building block (3 equiv.), HATU (2.9 equiv.) and DIPEA (6 equiv.) in DMF. The suspension was then gently stirred using Ar bubbling for 45–60 min. Monitoring of the coupling reaction was performed with TNBS.⁹¹ In the case of a positive TNBS test (indicating incomplete coupling), the suspension was filtrated, and treated again with a freshly prepared solution of the same *N*-Fmoc protected building block (2 equiv.) and coupling reagents. The resin was then filtered off and washed with DMF (4 mL, 4 $\times$ 1 min) prior to the subsequent Fmoc deprotection step using 20% piperidine in DMF (4 mL, 4 $\times$ 10 min). After filtration, the resin was washed with DMF (4 mL, 3 $\times$ 1 min) and solid-phase synthesis was continued by sequential incorporation of *N*-Fmoc protected building blocks. For each coupling step, the resin was treated as described above. After the last coupling the resin was filtered off, washed with DMF (4 mL, 4 $\times$ 1 min), CH_2Cl_2 (4 mL, 4 $\times$ 1 min), and activated for the cleavage according to Ingenito et al.⁶⁸ After swelling the resin in THF (4 mL), a solution of TMS–CHN $_2$ (2 M in hexane) was added, and the suspension was gently stirred using Ar bubbling for 2 h. Subsequently, the resin was filtered off, washed with THF (4 mL, 4 $\times$ 1 min) and DMF (4 mL, 4 $\times$ 1 min), to prepare it for the displacement reaction. The activated *N*-acylsulfonamide resin was swollen in DMF and filtered off. A solution of ethyl-3-mercaptopropionate (0.86 mL, 6.65 mmol) and sodium thiophenolate (9 mg, 0.066 mmol) in DMF (5 mL) was added and the resulting mixture heated at 80 °C for 24 h. Removal of side-chain protecting groups was accomplished in solution by treating the protected β -peptide thioester with a solution of TFA/ H_2O /TIS

(95/2.5/2.5). The solvents were removed under reduced pressure and the precipitate, which formed upon addition of cold Et₂O to the oily residue, was collected by centrifugation. Purification by RP-HPLC (10–50% B in 50 min, C₈) yielded **2** (10 mg, 10%) as a colorless fluffy solid. Anal. RP-HPLC: *t*_R 28.34 (10–40% B in 50 min, 40–95% B in 10 min C₈). CD (0.2 mM in MeOH, 20 °C): –11387.9 (219.5 nm); 0 (210 nm); +9643.96 (204 nm); CD (0.2 mM in MeOH, –10 °C): –17955.4 (216 nm); 0 (206.5 nm); +1895.45 (205 nm). ESI MS (positive mode): 1437.9 (15, (M+H)⁺), 730.5 (40, (M+Na+H)²⁺), 719.7 (100, (M+2H)²⁺), 480.0 (32, (M+3H)³⁺).

2.4.2. H-(S)β²hCys-(S)β²hGln-(S,S)β²hIle-(S)β²hLys-(S)β²hHis-(S)β²hLeu-(S)β²hAsp-(S)β²hMet-(S)β²hPhe-(S)β²hPro-OH (3**).** Esterification of the Wang resin was performed according to Chan and White.⁷¹ To a soln. of Fmoc-(S)β²hPhe-(S)β²hPro-OH (**4**) (461 mg, 0.9 mmol) in dry CH₂Cl₂ (4 mL) was added 1-MeIm (0.05 mL, 0.675 mmol) followed by MSNT (267 mg, 0.90 mmol). The mixture was stirred until all MSNT had dissolved. The solution was then transferred to a vessel containing the preswelled resin (200 mg, 0.18 mmol), and mixed under Ar bubbling for 1 h at 25 °C. The resin was then filtered off, washed with DMF (4 mL, 4×1 min), CH₂Cl₂ (4 mL, 4×1 min), and dried under h.v. overnight. The loading of the resin was determined by measuring the absorbance of the benzofulvene–piperidine adduct according to Schreiber and Seebach⁸⁰ and was found to be 0.61 mmol/g (68%), corresponding to 0.122 mmol of anchored **4**. The unreacted hydroxy groups were capped using Ac₂O (0.12 mL, 1.22 mmol) in DMF (4 mL) and DMAP (5 mg, 0.04 mmol, added in 0.5 mL DMF) for 30 min. The Fmoc group was removed using 20% piperidine in DMF (4 mL, 4×10 min) under Ar bubbling. After filtration, the resin was washed with DMF (4 mL, 4×1 min). Solid-phase synthesis was continued by sequential incorporation of *N*-Fmoc-protected dipeptides building blocks (**5**, **6**, **7**, **8**). For each coupling step, the resin was treated with a solution of Fmoc building block (3 equiv.), HATU (2.9 equiv.) and DIPEA (6 equiv.) in DMF. The suspension was then gently stirred using Ar bubbling for 45–60 min. Monitoring of the coupling reaction was performed with TNBS.⁹¹ In the case of a positive TNBS test (indicating incomplete coupling), the suspension was filtered, and treated again with a freshly prepared solution of the same *N*-Fmoc protected building blocks (2 equiv.) and coupling reagents. The resin was then filtered off and washed with DMF (4 mL, 4×1 min) prior to the subsequent Fmoc deprotection step using 20% piperidine in DMF (4 mL, 4×10 min). After filtration, the resin was washed with DMF (4 mL, 3×1 min) and solid-phase synthesis was continued by sequential incorporation of *N*-Fmoc protected building block. For each coupling step, the resin was treated as described above. After the last coupling the resin was filtered off, washed with DMF (4 mL, 4×1 min), CH₂Cl₂ (4 mL, 4×1 min), MeOH (4 mL, 4×1 min) and dried under h.v. for 24 h. The dry peptide resin was treated for 2 h with a TFA/H₂O/EDT/TIS (94:2.5:2.5:1) solution (10 mL). The resin was removed by filtration, washed with TFA, and the organic phase containing the peptide was concentrated under reduced pressure. The precipitate, which formed upon addition of cold Et₂O to the oily residue, was collected by

centrifugation. The crude peptide was purified by RP-HPLC (15–50% B in 40 min, 50–95% B in 20 min, C₈) to yield the TFA salt of **3** (65 mg, 38%) as a colorless fluffy solid. Homogeneity >95% (RP-HPLC). Anal. RP-HPLC: *t*_R 28.44 (10–40% B in 40 min, 40–95% B in 5 min, C₈). CD (0.2 mM in MeOH, 20 °C): –13128.5 (219 nm); 0 (207.5 nm); +14495.7 (200.5 nm); CD (0.2 mM in MeOH, –10 °C): –15643.1 (218.5 nm); 0 (206 nm); +16894.2 (199.5 nm). ESI HRMS (positive mode): 708.3576 (100, (M+2Na)²⁺, C₆₅H₁₀₆N₁₄Na₂O₁₄S₂²⁺; calcd 708.3625), 697.3718 (70, (M+H+Na)²⁺, C₆₅H₁₀₇N₁₄NaO₁₄S₂²⁺; calcd 697.3715), 686.3795 (10, (M+2H)²⁺, C₆₅H₁₀₈N₁₄O₁₄S₂²⁺; calcd 686.3805).

2.4.3. H-β²hGly-(S)β²hTrp-(S)β²hArg-(S)β²hVal-(S)β²hAsn-(S)β²hGlu-(R,S)β²hThr-(R)β²hSer-(S)β²hTyr-(S)β²hAla-(S)β²hCys-(S)β²hGln-(S,S)β²hIle-(S)β²hLys-(S)β²hHis-(S)β²hLeu-(S)β²hAsp-(S)β²hMet-(S)β²hPhe-(S)β²hPro-OH (1**).** β²-Peptide fragment **3** (5.2 mg, 3.1 μmol) and the C-terminal thioester β²-peptide **2** (4.8 mg, 3.1 μmol) were ligated in an aqueous buffer (100 mM phosphate, pH 7.5) (3 mL) containing thiophenol (4% v/v). The ligation reaction was performed at 25 °C and monitored using anal. RP-HPLC (see Fig. 21 above). Following completion of the ligation, the reaction mixture was diluted with H₂O (1 mL) containing 0.1% TFA and purified by preparative RP-HPLC (10–50% B in 40 min, 50–99% B in 10 min, C₈) to yield the TFA salt of **1** (5.12 mg, 54%) as a colorless fluffy solid. Anal. RP-HPLC: *t*_R 36.82 (10–50% B in 40 min, 50–99% B in 10 min, C₈). CD (0.2 mM in MeOH): –16849.8 (214 nm); 0 (200 nm); CD (0.2 mM in H₂O, pH 7): –1863.85 (220 nm); 0 (214.5 nm); +15824.65 (201.5 nm). MALDI HRMS: 2697.428 (10, (M+Na)⁺, C₁₂₇H₁₉₉N₂₉NaO₃₀S₂⁺; calcd 2697.428), 2675.442 (60, (M+H)⁺, C₁₂₇H₁₉₉N₂₉O₃₀S₂⁺; calcd 2675.445).

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