

Cite this: *Chem. Commun.*, 2012, **48**, 4722–4724

www.rsc.org/chemcomm

COMMUNICATION

A triphenylene based zinc ensemble as an oxidation inhibitor†

Vandana Bhalla,* Harshveer Arora, Abhimanew Dhir and Manoj Kumar

Received 13th December 2011, Accepted 8th February 2012

DOI: 10.1039/c2cc17802h

The zinc complex of a new triphenylene based receptor is evaluated for its anti-oxidant activity which is better in comparison to that of commercially available anti-oxidants.

Zinc, an essential nutrient, plays an important role as an anti-oxidant in the human body.¹ Deficiency of zinc increases the production of reactive oxygen species (ROS),² which are a class of radical or non-radical oxygen-containing molecules that show high reactivity to bio-molecules.³ Excessive ROS generation is involved in the pathogenesis of many diseases, including cardiovascular disease, cancer, and neurological disorders.⁴ Thus, regulation of generation of these ROS is highly significant.

Our research involves the design, synthesis and evaluation of artificial receptors for metal ions and anions.⁵ In continuation of this work, in the present manuscript, we have synthesized a new triphenylene based receptor **3** (*vide infra*) (Scheme 1) which selectively senses Zn²⁺ ions.⁶ Further, the zinc ensemble of **3** (**Zn-3**) has been evaluated for its anti-oxidizing property and it was observed that the anti-oxidant activity of **Zn-3** is better than five commercially available anti-oxidants. Earlier, a number of artificial receptors for Zn²⁺ ions with bio-mimetic applications have been reported in the literature⁷ which includes complexes for understanding the intrinsic properties of substrate or inhibitor recognition by Zn²⁺ ions at the active centers of enzymes.⁸ Recently, Reinaud *et al.* reported the tripodal bio-mimetic macrocyclic Zn²⁺ complexes.⁹ However, the development of bio-mimetic synthetic zinc complexes showing anti-oxidant activity has not been explored. Thus, to the best of our knowledge, this is the first report where a triphenylene based receptor for Zn²⁺ ions shows anti-oxidant activity.

Suzuki–Miyaura coupling of 2,3,6,7,10,11-hexabromo-triphenylene **1a** with boronic ester **1b**¹⁰ yielded hexamine **1c** in 70% yield (Scheme 1) (for synthetic details see S3, S9–S11 of ESI†). Condensation of hexamine **1c** with quinoline-2-carboxaldehyde **2** in THF at room temperature furnished compound **3** in 75% yield (Scheme 1). The structure of compound **3** was confirmed from its spectroscopic data (for synthetic details see S4, S12–S14 of ESI†) and its purity was determined from HPLC (see ESI†, S26).

Department of Chemistry, UGC-Centre for Advanced Studies, Guru Nanak Dev University, Amritsar, Punjab, India.

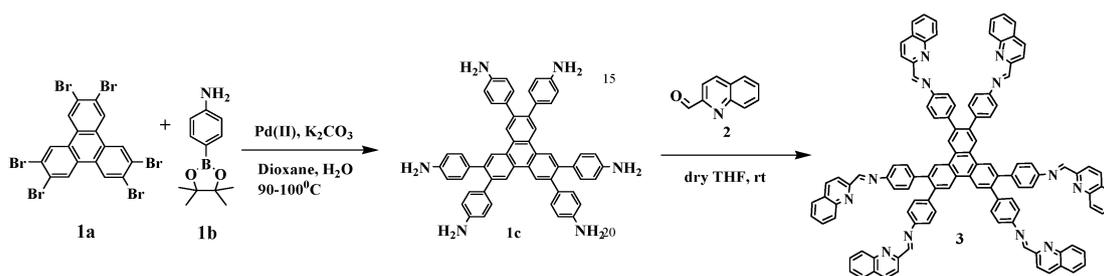
E-mail: vanmanan@yahoo.co.in; Fax: +91-(183)-2258820;

Tel: +91-(183)-2258802-9 Extn. 3205

† Electronic supplementary information (ESI) available: Experimental procedure and spectral and analytical data of compounds **1c** and **3**. See DOI: 10.1039/c2cc17802h

The binding behaviour of compound **3** toward different metal ions was evaluated with the help of UV-vis and fluorescence spectroscopy. The absorption spectrum of compound **3** (10 μM) shows three absorption bands at 246, 296 and 355 nm in THF : H₂O (95 : 5). Among all the metal ions tested (Zn²⁺, Fe³⁺, Fe²⁺, Cu²⁺, Hg²⁺, Ni²⁺, Cd²⁺, Pb²⁺, Mn²⁺, Co²⁺, Li⁺, Na⁺, Mg²⁺, K⁺, Ca²⁺, Ba²⁺) two new bands are formed at 290 and 325 nm upon addition of only Zn²⁺ ions (200 equiv.) with two isosbestic points at 285 and 335 nm (see ESI†, S6). These changes in the UV-vis spectrum of **3** upon addition of Zn²⁺ ions are ascribed to the formation of a complex between **3** and Zn²⁺ ions.

Compound **3** showed no fluorescence emission in THF : H₂O (95 : 5) (Fig. 1) when excited at 355 nm, which may be attributed to photo-induced electron transfer (PET)¹¹ from imino nitrogen to the photo-excited quinoline moiety. Upon addition of only Zn²⁺ ions (20 equiv.), an enhancement in the fluorescence spectrum was observed at 438 nm. The fluorescence enhancement of receptor **3** in the presence Zn²⁺ ions is attributed to the co-ordination of imino and quinolinyl nitrogens of **3** with Zn²⁺ leading to the formation of a supramolecular complex, as a result of which the PET from imino nitrogen to the quinoline moiety is suppressed. Under the same conditions, the fluorescence behaviour of **3** was tested with other metal ions (Fe³⁺, Fe²⁺, Cu²⁺, Hg²⁺, Ni²⁺, Cd²⁺, Pb²⁺, Mn²⁺, Co²⁺, Li⁺, Na⁺, Mg²⁺, K⁺, Ca²⁺, Ba²⁺), however no change in emission spectrum was observed with any other metal ion (see ESI†, S7). The fluorescence spectra of **3** (5 μM) at various concentrations of Zn²⁺ ions are shown in Fig. 1. Fitting the changes in the fluorescence spectra of compound **3** with Zn²⁺ ions, using the nonlinear regression analysis program SPECFIT,¹² gave a good fit and demonstrated that a 1 : 3 stoichiometry (host : guest) was the most stable species in the solution with a binding constant log β_{1,3} = 13.23 (see ESI†, S15 and S16). The complex formation was further confirmed by mass and IR spectroscopy (for mass spectrum see ESI†, S25). The IR absorption band at 1616 cm⁻¹ due to the imine moiety of **3** shifted to a lower frequency at 1590 cm⁻¹ ($n = 26 \text{ cm}^{-1}$) upon complexation with zinc, which confirms the coordination of imino nitrogen atoms with Zn²⁺ ions (see ESI†, S23 and S24). To test the practical applicability of compound **3** as a Zn²⁺ selective fluorescence sensor, competitive experiments were carried out in the presence of Zn²⁺ ions at 20 equiv. mixed with other cations (Fe³⁺, Fe²⁺, Cu²⁺, Hg²⁺, Ni²⁺, Cd²⁺, Pb²⁺, Mn²⁺, Co²⁺, Li⁺, Na⁺, Mg²⁺, K⁺, Ca²⁺, Ba²⁺) at 200 equiv., no significant variation was found by comparison with and without the other metal ions



Scheme 1

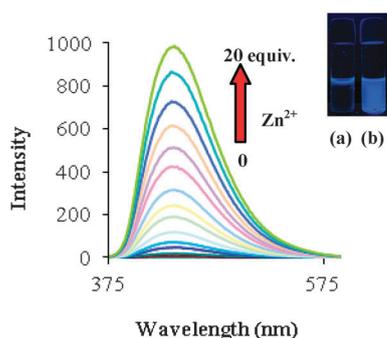


Fig. 1 Fluorescence emission spectrum of **3** (5 μM) upon addition of Zn^{2+} (20 equiv.) ions in THF : H_2O (95 : 5). Inset shows the fluorescence intensity changes upon the addition of Zn^{2+} ions from (a) 0 to (b) 20 equiv.

(see ESI†, S7). This means that compound **3** has a high affinity for Zn^{2+} ions. The detection limit of **3** as a fluorescent sensor for Zn^{2+} ions was found to be $1 \times 10^{-6} \text{ mol l}^{-1}$, which is sufficiently low for the detection of submillimolar concentrations of Zn^{2+} ions found in many chemical systems.¹³ The fluorescence quantum yield¹⁴ (ϕ_f) of compound **3** in the free and **3**· Zn^{2+} bound state was found to be 0.001 and 0.65, respectively (for calculations see ESI†, S4). This substantial increase in the quantum yield of compound **3** in the presence of Zn^{2+} ions showed its credibility as a good Zn^{2+} sensor.

Further, we evaluated the zinc ensemble of **3** (**Zn-3**) for its anti-oxidant activity using fluorescence spectroscopy and cyclic voltammetric studies (*vide infra*). For this, we utilized β -hydroxy naphthaldehyde¹⁵ as a template, carried out its Dakin oxidation¹⁶ and observed its fluorescence behaviour in the presence and absence of **Zn-3** as fluorescent probes for detection of ROS, which has been reported recently.¹⁷ However, such an evaluation of anti-oxidant activity of a metal coordinated ligand using an organic template is unprecedented.

β -Hydroxy naphthaldehyde **4** (3 μM) shows an emission at 438 nm in THF : H_2O (95 : 5) when excited at 318 nm (Fig. 2). The fluorescence emission of compound **4** is attributed to fast keto–enol tautomerism (representing the enol form **4** and keto form **5**, Scheme 2a) involving the phenomenon of excited state intramolecular proton transfer (ESIPT).¹⁸ To carry out Dakin oxidation we added alkaline $\text{H}_2\text{O}_2/\text{NaOH}$ (pH = 7.4) to the solution of **4** (3 μM) at 25 °C. Upon addition of H_2O_2 (225 μl) the emission band at 438 nm is blue shifted to 383 nm. We propose that this fluorescence response of compound **4** is due to the modulation of the existing ESIPT state (**5**) by the interaction of HOO^- ions with the hydroxyl group of **4**. Upon further addition of H_2O_2 (525 μl) quenching of the fluorescence

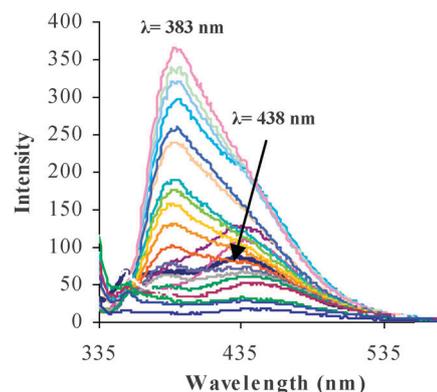
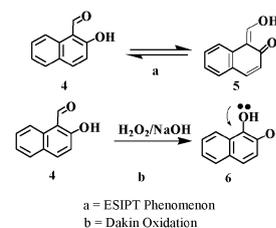


Fig. 2 Fluorescence emission spectra of **4** (3 μM) upon addition of H_2O_2 (750 μl) in THF : H_2O (95 : 5). Inset shows the (a) ESIPT phenomenon in β -OH naphthaldehyde and (b) its Dakin's oxidation.



Scheme 2

emission was observed. This quenching is ascribed to the oxidative conversion of **4** to species **6** (Scheme 2b). In species **6**, the inhibition of ESIPT makes photoinduced electron transfer (PET) operational (Scheme 2b) and as a result fluorescence emission gets quenched. Further, to observe the anti-oxidation effect of **Zn-3**, we performed the fluorescence titration with H_2O_2 under similar conditions in the presence of 250 μl of 100 μM of **Zn-3**. We observed that in the presence of **Zn-3** the rate of oxidation of **4** decreases and the amount of H_2O_2 required to quench the fluorescence is larger (1300 μl) (see ESI†, S17) than in the absence of **Zn-3**. This may be ascribed to the fact that hydroperoxide anions (HOO^-) generated during the reaction¹⁶ (which are responsible for the oxidation) show preferential binding to the charged zinc centre of **Zn-3**. The rate of oxidation slows down, indicating that **Zn-3** has an anti-oxidant effect.

We also studied the fluorescence behaviour of **Zn-3** toward other anions (F^- , CN^- , OAc^- , Cl^- , Br^- , I^- , EDTA^{4-} , HSO_4^- , NO_3^- , H_2PO_4^- , ClO_4^-) and observed that no change in fluorescence emission of **Zn-3** was observed with any other anions (see ESI†, S8). Thus, **Zn-3** shows a selective fluorescence response towards hydroperoxide anions.

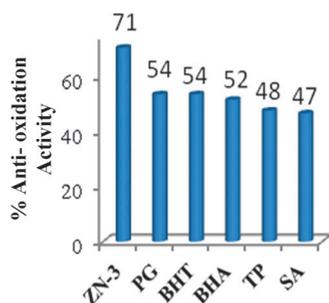


Fig. 3 Comparison of percentage anti-oxidation activity of **Zn-3** with other commercial anti-oxidants.

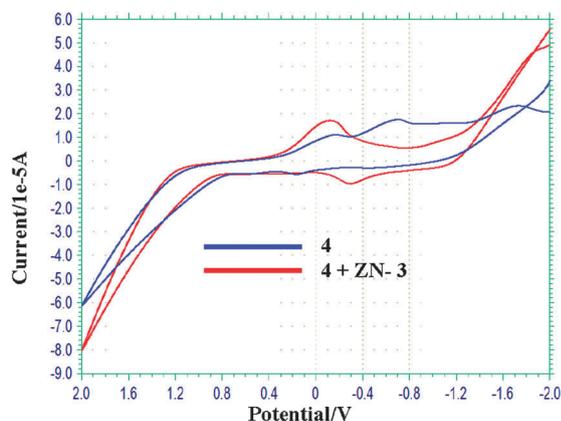


Fig. 4 Cyclic voltammogram of (a) **4** and (b) **4 + Zn-3**.

We also compared the anti-oxidant activity of **Zn-3** with five different commercially available anti-oxidants *viz.* propyl gallate (PG), butylated hydroxyl toluene (BHT), butylated hydroxyl anisole (BHA), α -tocopherol (TP) and sodium ascorbate (SA).¹⁹ It was observed that in the presence of 250 μ l of 100 μ M PG (see ESI[†], S18), BHT (see ESI[†], S19), BHA (see ESI[†], S20), TP (see ESI[†], S21) and SA (see ESI[†], S22) 1025 μ l, 1000 μ l, 1000 μ l, 975 μ l and 950 μ l of H₂O₂ were used, respectively, to quench the fluorescence of β -hydroxy naphthaldehyde **4**. The lower amount of hydrogen peroxide used in the presence of all these commercially available anti-oxidants in comparison to **Zn-3** shows that the **Zn-3** has a better anti-oxidant property. The percentage oxidation inhibition calculated for **Zn-3**, PG, BHT, BHA, TP, SA is 71%, 54%, 54%, 52%, 48% and 47% respectively (Fig. 3) (see ESI[†], S5).

The anti-oxidative nature of **Zn-3** observed from the above spectroscopic studies was further confirmed from electrochemical studies. The cyclic voltammogram of **4** [THF, $c = 1 \times 10^{-3}$, [(*n*Bu)₄N]ClO₄ as supporting electrolyte, using a glassy carbon working electrode, a (Ag/Ag⁺) reference electrode, and a Pt wire counter electrode] exhibits two electrochemical oxidation waves at $E_{1/2} = -0.175$ and $E_{1/2} = -0.70$ (Fig. 4). The cyclic voltammogram of **4 + Zn-3** under the same conditions showed one oxidation wave at $E_{1/2} = -0.13$ and a reduction wave

at $E_{1/2} = -0.30$ (Fig. 4b). Such a dramatic shift in the oxidation waves of **4** in the presence of **Zn-3** is attributed to the slow oxidation of **4** in the presence of **Zn-3**. Thus, the electrochemical studies firmly support the spectroscopic studies and prove the anti-oxidant behaviour of **Zn-3**.

To conclude, we synthesized a new triphenylene based receptor **3** incorporating quinoline moieties which behaves as a Zn²⁺ selective chemosensor. The Zn²⁺ ensemble of **3** shows anti-oxidant properties and may be used in the real world by diluting with THF prior to a hydroperoxide assay. This is proved by different spectral and electrochemical studies using β -hydroxy naphthaldehyde as an organic template and performing its Dakin oxidation in the absence and presence of **Zn-3**.

V.B. is thankful to DST (New Delhi, India) (ref no. SR/S1/OC-63/2010) and DRDO (Ref. No. ERIP/ER/0703663/M/01/1105) for financial support.

References

- (a) S. R. Powell, *J. Nutr.*, 2000, **130**, 1447S; (b) T. M. Bray and W. J. Bettger, *Free Radical Biol. Med.*, 1990, **8**, 281.
- (a) B. Halliwell and M. Whiteman, *Br. J. Pharmacol.*, 2004, **142**, 231; (b) E. Ho and B. N. Ames, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 16770.
- B. D. Autreaux and M. B. Toledano, *Nat. Rev. Mol. Cell Biol.*, 2007, **8**, 813.
- (a) H. K. Seitz and F. Stickel, *Nat. Rev. Cancer*, 2007, **7**, 599; (b) L. Galluzzi, K. Blomgren and G. Kroemer, *Nat. Rev. Neurosci.*, 2009, **10**, 481.
- (a) M. Kumar, A. Dhir and V. Bhalla, *Chem. Commun.*, 2010, **46**, 6744; (b) M. Kumar, R. Kumar and V. Bhalla, *Chem. Commun.*, 2009, 7384 and references cited therein.
- The receptor **3** is better in comparison to the Zn²⁺ sensors reported in the literature (ESI[†], S27 and S28).
- (a) S. Aoki and E. Kimura, *Chem. Rev.*, 2004, **104**, 769; (b) L. Cronin and P. H. Walton, *Chem. Commun.*, 2003, 1572.
- E. Kimura, *Acc. Chem. Res.*, 2001, **34**, 171.
- (a) B. Colasson, N. Le Poul, Y. Le Mest and O. Reinaud, *Inorg. Chem.*, 2011, **50**, 10985; (b) K. Kano, M. Kondo, H. Inoue, H. Kitagishi, B. Colasson and O. Reinaud, *Inorg. Chem.*, 2011, **50**, 6353.
- V. Bhalla, R. Tejpal, M. Kumar, R. K. Puri and R. K. Mahajan, *Tetrahedron Lett.*, 2009, **50**, 2649.
- (a) I. Aoki, T. Sakaki and S. Shinkai, *J. Chem. Soc., Chem. Commun.*, 1992, 730; (b) J. H. Bu, Q. Y. Zheng, C. F. Chen and Z. T. Huang, *Org. Lett.*, 2004, **6**, 3301.
- H. Gampp, M. Maeder, C. J. Meyer and A. D. Zuberhuhler, *Talanta*, 1985, **32**, 95.
- G. L. Long and J. D. Winefordner, *Anal. Chem.*, 1983, **55**, 712A.
- J. N. Demas and G. A. Grosby, *J. Phys. Chem.*, 1971, **75**, 991.
- K. Reimer and F. Tiemann, *Ber. Dtsch. Chem. Ges.*, 1876, **9**, 824.
- (a) H. D. Dakin, *J. Am. Chem. Soc.*, 1909, **42**, 477; (b) H. D. Dakin, *Org. Synth.*, 1923, **3**, 28.
- (a) H. Maeda, Y. Fukuyasu, S. Yoshida, M. Fukuda, K. Saeki, H. Matsuno, Y. Yamauchi, K. Yoshida, K. Hirata and K. Miyamoto, *Angew. Chem., Int. Ed.*, 2004, **43**, 2389; (b) E. W. Miller, O. Tulyathan, E. Y. Isacoff and C. Chang, *J. Nat. Chem. Biol.*, 2007, **3**, 263.
- (a) P. T. Chou, J. H. Liao, C. Y. Wei, C. Y. Yang, W. S. Yu and Y. H. Chou, *J. Am. Chem. Soc.*, 2000, **112**, 986; (b) A. Kyrchenko, J. Herbich, M. Izydorzak, F. Wu, R. P. Thummel and J. Waluk, *J. Am. Chem. Soc.*, 1990, **112**, 11179.
- European Directive 95/2/EC on food additives, 1995, 1–61.