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Improved synthetic method of Benzo[*a*]pheno-selenazinium phototherapeutic agents

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ABSTRACT

Benzo[*a*]phenoselenazinium dyes are excellent photodynamic therapeutic agents that have a red absorption (660 nm), high singlet oxygen quantum yield (>0.8), and good water-solubility and liposolubility. Bis-(3-diethylaminophenyl) diselenide, the most important intermediate for the synthesis of benzo[*a*]phenoselenazinium dyes, is previously prepared in one pot through 3 steps, during which special care is needed for the preparation of Grinard reagent and toxic selenium vapor. In this work, we employed CuO nanopowder to directly couple haloaniline with selenium to obtain pure bis-(3-diethylaminophenyl) diselenide (one step) under a mild condition with a high reaction yield without the need of column chromatography. Four benzo[*a*]phenoselenazinium dyes, **5a-d**, were easily prepared using the improved synthesis method; and their phototoxicity in living cells were investigated. The improved method could be employed in large-scale synthesis of benzo[*a*]phenoselenazinium dyes, thereby may stimulate the photodynamic study of this type of photosensitizer.

1. Introduction

As a treatment that has low toxicity, low cost and minimal invasiveness, but high effectiveness and dual selectivity, photodynamic therapy (PDT) is a promising alternative therapy for ophthalmologic, dermatologic, and cancerous oral lesions, head and neck cancers, bladder carcinoma, and a variety of intraperitoneal carcinomatosis and sarcomatous transformation [1-11]. In PDT, photosensitizers (PSs) are the key components which can be excited by absorbing photons so that they can subsequently react with oxygen to generate reactive oxygen species (ROS) such as singlet oxygen, superoxide anion radicals, hydroxyl radicals, and so on. Singlet oxygen (¹O₂) is the main ROS that can kill cancer cells through multifactorial mechanisms, such as stimulation of the inflammatory and immune responses [12-21]; therefore, photosensitizers with high singlet oxygen quantum yield are desirable. We first developed a new photodynamic therapeutic agent, benzo[a]phenoselenazinium dye EtNBSe, an effective photosensitizer that treats cutaneous leishmaniasis, tuberculosis and Mycobacterium bovis by oxidative damage [22-25]. Following the development, more studies on the derivative of EtNBSe have also been conducted [26-28]. All these studies have shown that benzo[a]phenoselenazinium photosensitizers

are highly suitable for photodynamic therapy *in vivo* because they have a red absorption in the optical window (600–900 nm), a high single oxygen quantum yield, good water-solubility and liposolubility, and excellent photo-stability [29–32].

We have previously synthesized benzo[a]phenoselenazinium dyes in six steps [25], as illustrated in Scheme 1. First, iodoaniline is converted into the Grignard reagent (3-diethylamino) phenylmagnesium iodide that subsequently reacts with selenium powder, is oxidized with air, and is finally nitrosated to produce bis-(3-diethylamino-6-nitrosophenyl) diselenide. Bis-(3-diethylamino-6-nitrosophenyl) diselenide is then condensed with 1-naphthylamine derivatives to afford dye EtNBSe. Although it was a successful approach, the synthesis of the key intermediate bis-(3-diethylaminophenyl) diselenide requires harsh conditions during the following two steps: i) the preparation and handling of the Grignard reagent (3-diethylamino) phenylmagnesium iodide requires a very dry environment; and ii) during the air oxidation process, toxic selenium vapor is produced, and aqueous sodium hypochlorite is needed to trap and bleach the vapor. In order to prepare more EtNBSe analogue dyes at a larger scale for use in intensive medical studies, there is a high demand for the neat synthesis under mild reaction conditions.

In this work, we improved the preparative process of bis-(3-

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diethylaminophenyl) diselenide with an aim to increase the level of the synthesis of **EtNBSe** derivatives to a larger scale (Scheme 2).

2. Experimental

2.1. Instruments and materials

¹H NMR and ¹³C NMR spectra were on a Bruker 400 spectrometer; the chemical shifts were reported in ppm in relative to those of TMS (tetramethyl silicane), an internal standard. Mass spectra were acquired using a Bruker Daltonics micrOTOF-Q II mass spectrometer. Absorption and emission spectra were recorded on Hitachi F-7000 fluorometer and Shimadzu UV-2450 spectrophotometer, respectively. Unless otherwise stated, all reagents were used as received. Twice-distilled water was used in all experiments.

2.2. Synthesis of CuO nanopowder

Fifty milliliters of aqueous CuSO₄ solution (0.5 M) was mixed by stirring with 50.0 mL of aqueous NaOH solution (1.0 M) in a beaker; immediately after mixing, blue flocculent precipitate was formed. Next, add 130.0 mL of sodium carbonate aqueous solution (0.5 M) to the mixture, and continue to stir for 30 min. Collect the precipitate by filtration and then wash with distilled H_2O until SO_4^{2-} was no longer detected by BaCl₂. The obtained solid was dried in an oven at 90 °C for 3 h and was then ground into powder. Finally, the powder was calcined at 350 °C for 4 h from which black CuO nanopowder was obtained.

2.3. Synthesis of bis-(3-diethylaminophenyl) diselenide (2)

Under argon atmosphere and while being stirred, powdered Se metal (9.53 g, 120.0 mmol), CuO nanoparticles (0.46 g, 6.0 mmol) and KOH (6.74 g, 120.0 mmol) were added to a solution of 3-iodo-N, N-dieth-ylaniline (1) (16.5 g, 60.0 mmol) in dry DMSO (60.0 mL). The mixture was then heated to 110 °C overnight. After cooling down to room temperature, 10.0 mL of water was added. After that, the reaction mixture was extracted three times with ethyl acetate (3 x 40.0 mL). The organic phase was washed twice with brine and thereafter dried over anhydrous Na₂SO₄ overnight. After ethyl acetate was removed under a reduced pressure, light red oil (20.35 g, 74%) was obtained.

2.4. Synthesis of bis-(3-diethylamino-6-nitrosophenyl) diselenide (3)

In an ice-water bath, a solution of $NaNO_2$ (6.20 g, 98.2 mmol) in 80.0 mL water was added dropwise to a solution of bis-(3-dieth-ylaminophenyl) diselenide (2) (20.35 g, 44.8 mmol) in 100.0 mL of 1.0 M HCl for 10 min. After another 10 min stirring, extract with dichloromethane (4 x 75.0 mL) and wash the organic phase twice with

brine. After being dried over anhydrous Na_2SO_4 , the solvent was distilled *in vacuo* to obtain a crude solid, which was recrystallized in isopropanol to afford an orange powder (16.35 g, 71%).

2.5. Synthesis of 1-naphthylamine derivatives (4a-d)

The synthetic method for compounds **4a-d** have been described in the literature [33].

2.6. General procedure for the synthesis of benzo[a] phenoselenazinium dyes 5a-d

Benzo[a]phenoselenazinium dyes 5a-d were synthesized from bis-(3-diethylamino-6-nitrosophenyl) diselenide and the corresponding 1naphthylamine derivatives (4a-d). Compound 3 (4.62 g, 9.0 mmol) was mixed with the corresponding 1-naphthylamine derivatives 4ad (26.0 mmol) in 60.0 mL of trifluoroethanol, and the mixture was refluxed for 30 min. The solvent was then removed in vacuo to obtain a blue residue, which was subsequently washed with 60.0 mL of ethyl ether. The resultant residue was dissolved in 300.0 mL of a mixture containing 1.0 M aqueous sodium hydroxide solution and dichloromethane (1:1, v/v). The organic phase was washed twice with brine; after that, concentrated hydrochloric acid (0.1 mL) was added to the solution, which caused the solution color to change from magenta to dark blue. The solvent was removed in vacuo to obtain a blue residue, which was then purified by flash silica gel column using a solvent gradient of methanol/dichloromethane (0:100, 1:50, 1:30, 1:20, and 1:10, v/v) to yield the product.

5a. Blue solid, yield: 41%. HRMS (ESI) *m/z*: calcd for $C_{22}H_{24}N_3Se$ [M]⁺, 410.1130; found, 410.1123. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.48 (s, 1H), 8.73 (d, *J* = 5.2 Hz, 1H), 8.62 (s, 1H), 7.89–7.76 (m, 1H), 7.65 (d, *J* = 4.5 Hz, 2H), 7.19 (d, *J* = 8.3 Hz, 1H), 7.13 (s, 1H), 7.01 (s, 1H), 3.70 (s, 6H), 1.48 (d, *J* = 5.8 Hz, 2H), 1.42–1.30 (m, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 152.9, 150.3, 143.5, 138.7, 134.4, 133.7, 132.7, 131.96, 130.8, 129.4, 125.3, 124.4, 123.8, 115.8, 107.6, 105.9, 45.7, 39.3, 14.1, 12.7.

5b. Blue solid, yield: 40%. HRMS (ESI) *m/z*: calcd for C₂₃H₂₆N₃O₃SSe [M]⁺, 504.0855; found, 504.0354. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.20 (s, 1H), 8.99 (dd, *J* = 8.2, 1.1 Hz, 1H), 8.62 (d, *J* = 8.1 Hz, 1H), 8.02 (d, *J* = 9.4 Hz, 1H), 7.95 (s, 1H), 7.91 (t, *J* = 7.6 Hz, 1H), 7.85–7.79 (m, 1H), 7.73 (d, *J* = 2.7 Hz, 1H), 7.31 (dd, *J* = 9.5, 2.7 Hz, 1H), 3.73 (dd, *J* = 14.3, 7.1 Hz, 2H), 3.64 (q, *J* = 6.9 Hz, 4H), 1.37 (t, *J* = 7.2 Hz, 4H), 1.24 (t, *J* = 7.0 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 153.2 150.9, 144.0, 139.3, 134.9, 133.9, 132.6, 131.2, 129.7, 125. 8, 124.7, 123.3, 116.5, 108.0, 106.0, 46.0, 39.6, 29.9, 14.1, 12.9.

5c. Blue solid, yield: 38%. HRMS (ESI) m/z: calcd for C₂₃H₂₄N₃O₂Se [M]⁺, 454.1028; found, 454.1020. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.63 (d, J = 7.7 Hz, 1H), 8.34 (d, J = 7.3 Hz, 1H), 7.62 (dd, J = 17.0, 7.3 Hz,



Scheme 1. Schematic diagram showing the previous synthetic route of photosensitizer EtNBSe (5a).



Scheme 2. The improved synthetic route of the photosensitizers 5a-d presented in this work.

4H), 7.11 (s, 1H), 6.85 (d, J = 8.3 Hz, 1H), 3.80 (t, J = 6.6 Hz, 2H), 3.53–3.44 (m, 4H), 2.70 (t, J = 6.5 Hz, 2H), 1.16 (t, J = 6.9 Hz, 6H). ¹³C NMR (100 MHz, DMSO- d_6) δ 175.0, 156.2, 151.0, 145.2, 139.1, 134.6, 132.2, 132.1, 129.7, 125.9, 124.2, 124.1, 117.0, 115.8, 111.1, 109.3, 107.9, 55.4, 45.6, 29.5, 22.6, 19.0, 13.2.

5d. Blue solid, yield: 45%. HRMS (ESI) m/z: calcd for C₂₂H₂₄N₃OSe [M]⁺, 426.1079; found, 426.1072. ¹H NMR (400 MHz, DMSO- d_6) δ 10.08 (s, 1H), 9.00 (d, J = 8.0 Hz, 1H), 8.60 (d, J = 8.1 Hz, 1H), 8.08–7.98 (m, 2H), 7.91 (t, J = 7.6 Hz, 1H), 7.83 (t, J = 7.2 Hz, 1H), 7.71 (s, 1H), 7.30 (d, J = 9.3 Hz, 1H), 5.14 (s, 1H), 3.79 (d, J = 2.4 Hz, 4H), 3.64 (q, J = 6.9 Hz, 4H), 1.24 (t, J = 7.0 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 157.3, 154.6, 147.5, 142.9, 138.5, 137.7, 137.4, 136.4, 134.7, 133.1, 129.3, 128.2, 126. 5, 120.3, 111.7, 110.2, 63.6, 50.5, 49.6, 16.3.

2.7. Measurement of singlet oxygen quantum yield (Φ_{Δ})

The ${}^{1}O_{2}$ quantum yields (Φ_{Δ}) of the photosensitizers were determined with methylene blue (MB) as a reference ($\Phi_{\Delta} = 0.5$ in methanol) [34]. 1,3-Diphenylisobenzofuran (DPBF) in air-saturated dichloromethane with an optical density (OD) at 1.0 was used to scavenge ${}^{1}O_{2}$. The concentration of the photosensitizer was adjusted to an OD between 0.2 and 0.3. The cuvette was irradiated for 10 s with monochromatic light at the maximum absorption wavelength of photosensitizers, and the absorbance was measured several times after each irradiation. The slope of the plot between the absorbance of DPBF at 411 nm versus time for each photosensitizer was calculated. Singlet oxygen quantum yield (Φ_{Δ}) was calculated according to a modified equation: $\Phi_{\Delta}^{\text{bod}} = \Phi_{\Delta}^{\text{ref}} *$ $(k_{\rm bod}/k_{\rm ref})^*(F_{\rm ref}/F_{\rm bod})$, where bod and ref refer to the photosensitizers and MB, respectively, k is the slope of the plot between the absorbance of DPBF (411 nm) versus irradiation time, and F is the absorption correction factor (which was calculated by $F = 1-10^{-0.D}$, where O.D. is the optical density of the solution at the irradiation wavelength).

2.8. In vitro toxicity assay

MTT assay was used to assess the toxicity to cancer cells of the photosensitizers [32]. HepG2 cells (100.0 μ L; density = 5000 cells/mL) in the logarithmic phase were added to 96-well plates and incubated at 37 °C in 5% CO₂ for 24 h. After that, 100.0 μ L of each of the photosensitizers **5a-d** in DMSO solution was added to each well, and cells were further incubated for 1 h. After being illuminated with 660 nm LED light (20 mW/cm²), cells were incubated for 24 h at 37 °C in 5% CO₂ atmosphere. Subsequently, the medium in each well was replaced with 100.0 μ L of fresh medium, and 20.0 μ L of MTT solution was added thereafter. After the culture plates were then incubated at 37 °C in 5% CO₂ for 4 h, the culture medium was discarded, and 100.0 μ L of DMSO was added. The absorbance at 570 nm of the samples was measured using a microplate reader. Cells incubated with photosensitizers without light irradiation were used as a control.

3. Results and discussion

3.1. Chemistry of the synthesis

In this study, bis-(3-diethylamino-6-nitrosophenyl) diselenide was synthesized in one step using CuO nanopowder as a catalyst. This new synthetic method had several advantages as follows: (i) the special care needed for the Grignard reaction was avoided; (ii) the toxic selenium vapor was not generated because air bubbles were omitted; and (iii) the reaction procedure was simple (as it could be accomplished in only one step), and the product yield was improved from 56% to 74.5% without the need of column chromatography. Thus, this new synthetic method was able to readily prepare four pure benzo[*a*]phenoselenazinium dyes **5a-d** with high yields.

3.2. Optical properties of photosensitizers

We determined the optical spectra of the four photosensitizers **5a-d** in methanol and H₂O. All the dyes contain a delocalized positive charge and are soluble in methanol and H₂O. The four dyes exhibited an intensive absorption band within the optical window ($\lambda_{max} \approx 660$ nm; Fig. 1). In H₂O, the dyes exhibited an absorption band at about 627 nm, which might be caused by H-aggregation of the photosensitizers [25]. As expected, the dyes also exhibited weak fluorescence due to the effect of the heavy metal Se [35,36] (Fig. 1).

3.3. Singlet oxygen quantum yield

The singlet oxygen quantum yields of the photosensitizers **5a-d** in methanol were measured using the ${}^{1}O_{2}$ trapping reagent 1,3-diphenylisobenzofuran (DPBF). As shown in Fig. 2 and S2, illuminating the solutions of the photosensitizers **5a-d** in DPBF with 660 nm LED light caused their absorption intensity at 411 nm to decrease due to the irreversible 1,4-cycloaddition reaction between DPBF and ${}^{1}O_{2}$. Moreover, with increasing illumination time, the decrease was intensified. The singlet oxygen yields of the photosensitizers **5a-d** were determined to be over 0.8 (Table 1).

3.4. In vitro photodynamic therapy and imaging

HepG2 cells were cultured with photosensitizers **5a-d** for 30 min; after that, they were irradiated under a 660 nm light (20 mW/cm²) for 0, 1, 5, and 10 min, respectively; non-irradiated samples were also prepared for comparison. As shown in Fig. 3, the non-irradiated photosensitizers **5a-d** were non-toxic, suggesting they have good biocompatibility *in vitro*. In contrast, as expected, the light-irradiated photosensitizers **5a-d** were highly cytotoxic, and the cytotoxicity increased with increasing the dose of photosensitizer as well as with the irradiation time. Interestingly, photosensitizers **5a**, **5b** and **5d** at a low



Fig. 1. Absorption (top row) and fluorescence (bottom row) spectra of photosensitizers 5a-d in methanol (a, c) and in H₂O (b, d).



Fig. 2. The decay curves of the absorbance at 411 nm of DPBF in methanol as a function of irradiation time without and with photosensitizers **5a-d** and methylene blue (MB). Light source 660 nm LED (20 mW/cm^2).

concentration of 0.04 μ M displayed high cytotoxicity when the irradiation time was 10 min; however, this is not the case for photosensitizer **5c**. As shown in Fig. S4, these four dyes all exhibited bright red fluorescence in HepG2 cells. These results indicate that photosensitizers **5a**-**d** had good photodynamic therapeutic effect and could also be applied for cell imaging.

 Table 1

 Photophysical properties of photosensitizers 5a-d in methanol.

Φ_{Δ} (¹ O ₂)
0.80
0.83
0.86
0.86

4. Conclusion

In conclusion, CuO nanopowder was used as the catalyst to optimize the synthesis of the key intermediate of benzo[*a*]phenoselenazinium PDT agents, bis-(3-diethylaminophenyl) diselenide. This new synthetic method required a mild reaction condition, but resulted in the product with high purity without the need of purification by column chromatography; it also had a simple and straightforward operation procedure. This work presents the synthesis method for benzo[*a*]phenoselenazinium PDT agents that can be potentially up-scaled. With this method, the clinical studies and practical applications of the agents can potentially be boosted.



Fig. 3. Cytotoxicity against HepG2 cells of photosensitizers 5a-d at various concentrations under a 660 nm light (20 mW/cm²) irradiation for (a) 0 min, (b) 1 min, (c) 5 min, and (d) 10 min.

Credit author statement

Xiuxiu Yue prepared the photosensitizers **5a-d**, performed the measurements, analyzed the data, and wrote the paper. Jing Xu prepared the performed the experiments. Xiangzhi Song processed the data and writing-review & editing supervision with Xiaozhong Liu and James W. Foley. All authors have given approval to the final version of the paper.

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.dyepig.2021.109154.

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