

ORIGINAL PAPER

An efficient method for the preparation of benzyl γ -ketohexanoates

Muhammad Iqbal*, Imam B. Baloch, Musa K. Baloch

Department of Chemistry, Gomal University, 29050 Dera Ismail Khan, Pakistan

Received 4 August 2012; Accepted 12 September 2012

Twenty acid chlorides, 4-(mono/di-benzyloxy)-4-ketobutanoyl chlorides (*Ia*-XX*a*) were synthesised by the reaction of monoesters of succinic acid with thionyl chloride. The product thus obtained (4-benzyloxy-4-ketobutanoyl chlorides) was treated with diethylcadmium to convert it into the corresponding keto-esters (*Ib*-XX*b*), the mono/di-benzyl- γ -ketohexanoates, with a good yield. All the compounds thus prepared were characterised by physical, spectroscopic (UV-VIS, IR, ^1H NMR, ^{13}C NMR), and mass measurements techniques.

© 2012 Institute of Chemistry, Slovak Academy of Sciences

Keywords: γ -ketoesters, mono/disubstituted-benzyl alcohols, diethylcadmium, monoesters of succinic acid

Introduction

The γ -ketohexanoates are bi-functional compounds having ketone and ester groups. They are considered to be the precursors of numerous important compounds used in the pharmaceutical and agrochemical industries (Dahl et al., 1999; Forni et al., 1994; Fujisawa et al., 1994; Nakamura et al., 2003; Shafiee et al., 1998). These are synthons of some biologically active compounds like sex hormones and pheromones, anti-asthma drugs, enzyme inhibitors, additives in food, and perfumes (Hayakawa et al., 1998; Heiss et al., 2001; Itoh et al., 2002; Kataoka et al., 1999; Kizaki et al., 2001; Larock, 1999; Sheldon & Arends, 2004; Tojo & Fernández, 2006; Yamamoto et al., 2002a, 2002b). Hence, a number of techniques for their synthesis have been proposed and some significant routes leading to the synthesis of γ -ketoesters have been described by Ballini et al. (2002), Izquierdo et al. (2011), Wehrli and Chu (1973, 1978), Ronsheim et al. (2002), Brockman & Fabio (1957), and Taylor (1958). A great deal of work has been reported in this area of organic chemistry over the last decade (Bandgar et al., 2005; Csende, 2002; Hilgenkamp & Zercher, 2001; Huang et al., 2005; Kashima et al., 2001; Wang et al., 2009; Williams et al., 2001, 2002).

These methods require very expensive transition

metal complexes, their ions or oxides as catalysts and toxic solvents such as ketoacids, strong inorganic acids, nitrogenous and halogenated solvents, the use of which can be harmful to the environment (Poliakoff et al., 2002; Larock, 1999; Sheldon & Arends, 2004; Tojo & Fernández, 2006; Bandgar et al., 2005; Csende, 2002; Hilgenkamp & Zercher, 2001; Kashima et al., 2001; Wang et al., 2009; Williams et al., 2001, 2002; Huang et al., 2005). The other common technique is anaerobic oxidation; this faces criticism on the grounds that it is difficult to control and to obtain a well-defined product (Csende et al., 1993; Stájer et al., 1994; Poliakoff et al., 2002; Hudlicky, 1990). However, the contribution made by Von Rudloff (1958), Cason (1942, 1946), and Cason and Prout (1944, 1948) to the synthesis of γ -ketoesters is essential as it provides basic and important information in this respect.

The above aspects prompted us to synthesise new mono/di-benzyl- γ -ketohexanoates (*Ib*-XX*b*) from the corresponding acid chlorides (*Ia*-XX*a*) using the methodology devised by Cason and Prout (1944, 1948) and Von Rudloff (1958). The compounds reported (*Ia*-XX*a* and *Ib*-XX*b*) were synthesised by following a two-step reaction using diethyl ether as solvent and diethyl cadmium as ethylating agent. In the first step, the benzyl hydrogen succinates (*I*-XX) were converted into the corresponding acid chlorides (*Ia*-XX*a*) by al-

*Corresponding author, e-mail: miqbal_44@yahoo.com

Table 1. Physical characteristics of *Ia*–*XXa*

Entry	Ar ^a	Time min	Yield %	Formula	M _r	Physical state	w _i (calc.)/% w _i (found)/%					
							C	H	N	Cl	Br	I
<i>Ia</i>	2-MeO-C ₆ H ₄	165	75	C ₁₂ H ₁₃ ClO ₄	256.68	Viscous oil	56.15 56.23	5.10 4.78	—	13.81 13.94	—	—
<i>IIa</i>	3-MeO-C ₆ H ₄	165	70	C ₁₂ H ₁₃ ClO ₄	256.68	Viscous oil	56.15 56.34	5.10 5.22	—	13.81 13.33	—	—
<i>IIIa</i>	4-MeO-C ₆ H ₄	165	74	C ₁₂ H ₁₃ ClO ₄	256.68	Viscous oil	56.15 56.33	5.10 5.23	—	13.81 13.86	—	—
<i>IVa</i>	2,3-(MeO) ₂ -C ₆ H ₃	150	79	C ₁₃ H ₁₅ ClO ₅	286.70	Viscous oil	54.46 54.37	5.27 5.31	—	12.37 12.66	—	—
<i>Va</i>	2,4-(MeO) ₂ -C ₆ H ₃	150	79	C ₁₃ H ₁₅ ClO ₅	286.70	Viscous oil	54.46 54.64	5.27 5.11	—	12.37 12.56	—	—
<i>VIa</i>	2,5-(MeO) ₂ -C ₆ H ₃	150	75	C ₁₃ H ₁₅ ClO ₅	286.70	Viscous oil	54.46 54.55	5.27 5.13	—	12.37 12.14	—	—
<i>VIIa</i>	3,4-(MeO) ₂ -C ₆ H ₃	150	72	C ₁₃ H ₁₅ ClO ₅	286.70	Viscous oil	54.46 54.61	5.27 5.16	—	12.37 12.48	—	—
<i>VIIIa</i>	3,5-(MeO) ₂ -C ₆ H ₃	150	74	C ₁₃ H ₁₅ ClO ₅	286.70	Viscous oil	54.46 54.59	5.27 5.16	—	12.37 12.22	—	—
<i>IXa</i>	2-NO ₂ -C ₆ H ₄	180	70	C ₁₁ H ₁₀ ClNO ₅	271.66	Viscous oil	48.63 48.86	3.71 3.65	5.16 5.31	13.05 12.84	—	—
<i>Xa</i>	3-NO ₂ -C ₆ H ₄	180	69	C ₁₁ H ₁₀ ClNO ₅	271.66	Viscous oil	48.63 48.33	3.71 3.58	5.16 5.10	13.05 12.97	—	—
<i>XIa</i>	4-NO ₂ -C ₆ H ₄	180	67	C ₁₁ H ₁₀ ClNO ₅	271.66	Viscous oil	48.63 48.42	3.71 3.88	5.16 5.23	13.05 12.82	—	—
<i>XIIa</i>	2-Cl-C ₆ H ₄	210	63	C ₁₁ H ₁₀ Cl ₂ O ₃	260.95	Viscous oil	50.60 50.76	3.86 3.65	—	27.16 27.33	—	—
<i>XIIIa</i>	3-Cl-C ₆ H ₄	210	65	C ₁₁ H ₁₀ Cl ₂ O ₃	260.95	Viscous oil	50.60 50.47	3.86 3.79	—	27.16 27.32	—	—
<i>XIVa</i>	4-Cl-C ₆ H ₄	210	66	C ₁₁ H ₁₀ Cl ₂ O ₃	260.95	Viscous oil	50.60 50.67	3.86 3.68	—	27.16 26.98	—	—
<i>VIa</i>	2-Br-C ₆ H ₄	225	68	C ₁₁ H ₁₀ BrClO ₃	305.76	Viscous oil	43.24 43.16	3.30 3.45	—	11.60 11.26	26.15 26.45	—
<i>XVIa</i>	3-Br-C ₆ H ₄	225	67	C ₁₁ H ₁₀ BrClO ₃	305.76	Viscous oil	43.24 43.35	3.30 3.21	—	11.60 11.53	26.15 26.24	—
<i>XVIIa</i>	4-Br-C ₆ H ₄	225	72	C ₁₁ H ₁₀ BrClO ₃	305.76	Viscous oil	43.24 43.17	3.30 3.26	—	11.60 11.76	26.15 26.08	—
<i>XVIIIa</i>	2-I-C ₆ H ₄	210	63	C ₁₁ H ₁₀ ClIO ₃	352.65	Viscous oil	37.47 37.42	2.86 2.77	—	10.06 10.32	—	36.00 35.98
<i>XIXa</i>	3-I-C ₆ H ₄	210	64	C ₁₁ H ₁₀ ClIO ₃	352.65	Viscous oil	37.47 37.56	2.86 2.67	—	10.06 9.97	—	36.00 36.23
<i>XXa</i>	4-I-C ₆ H ₄	210	66	C ₁₁ H ₁₀ ClIO ₃	352.65	Viscous oil	37.47 37.41	2.86 2.98	—	10.06 9.98	—	36.00 35.76

a) Aromatic substituent (Fig. 2).

lowing them to react with thionyl chloride and in the second step the acid chlorides thus obtained were converted into novel benzyl γ -ketohexanoates (*Ib*–*XXb*) by treating them with diethyl cadmium.

Experimental

All the required chemicals were of analytical grade, purchased from Merck (Germany) and used as received. The UV-VIS spectrum was recorded in absolute methanol on an Irmeco UV/VIS Model U-2020 spectrophotometer (Germany). IR spectra were obtained on a Tensor 27 FT-IR spectrophotometer supplied by Bruker (Germany). ¹H NMR (300 MHz) and

¹³C NMR (75 MHz) (1D, 2D, homo/hetero-nuclear) spectra were procured in CDCl₃ on a Bruker Biospin, AMX 300 MHz FT NMR spectrometer (Bruker, Germany). EI-MS were acquired with a direct insertion probe on a double-focusing Finnigan MAT 112 at 70 eV. HR-EI-MS measurements were made on a JEOL HX 110 spectrometer (Jeol, Japan). Elemental analyses were performed by Midwest Microlab (USA). Column chromatography was carried out using silica gel (PF₂₅₄, mesh size 60–70; Merck, Germany), and thin layer chromatography was performed on pre-coated silica gel plates (20 cm × 20 cm, 0.2 mm thickness) with UV fluorescence (PF₂₅₄) (Merck, Germany).

Table 2. Spectroscopic data of acid halides (*Ia*–*XXa*)

Compound	Spectral data
<i>Ia</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3056 (ArH), 1792 (COCl), 1737 (C=O), 1595, 1546 (C=C, ArH), 1026 (C—O), 723 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.60 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.74 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.89 (s, 3H, OMe), 5.14 (s, 2H, C-1''), 6.96 (m, 2H, ArH), 7.46 (m, 1H, ArH), 7.80 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 31.5 (C-2), 36.6 (C-3), 56.6 (OMe), 67.8 (C-1''), 113.8 (C-3'), 120.1 (C-5'), 121.2 (C-1'), 128.1 (C-4'), 128.9 (C-6'), 159.1 (C-2'), 169.9 (C-1), 175.6 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 256.68 (11) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 278 (3.7)
<i>IIa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3057 (ArH), 1793 (COCl), 1738 (C=O), 1592, 1541 (C=C, ArH), 1021 (C—O), 721 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.60 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.74 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.89 (s, 3H, OMe), 5.14 (s, 2H, C-1''), 6.96 (m, 2H, ArH), 7.46 (m, 1H, ArH), 7.80 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 30.9 (C-2), 36.2 (C-3), 56.7 (OMe), 67.5 (C-1''), 112.5 (C-2'), 115.6 (C-3'), 112.2 (C-4'), 118.4 (C-6'), 128.3 (C-5'), 137.6 (C-1'), 169.1 (C-1), 175.3 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 256.68 (13) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 278 (3.7)
<i>IIIa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3057 (ArH), 1793 (COCl), 1738 (C=O), 1593, 1542 (C=C, ArH), 1023 (C—O), 722 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.61 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.76 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.82 (s, 3H, OMe), 5.18 (s, 2H, C-1''), 6.97 (m, 2H, ArH), 7.25 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 30.1 (C-2), 37.6 (C-3), 55.5 (OMe), 67.8 (C-1''), 112.4 (C-5'), 112.5 (C-3'), 126.4 (C-6'), 127.2 (C-1'), 127.3 (C-2'), 157.5 (C-4'), 168.9 (C-1), 176.5 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 256.68 (16) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 278 (3.7)
<i>IVa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3065 (ArH), 1797 (COCl), 1741 (C=O), 1598, 1547 (C=C, 1027 (C—O), 725 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.64 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.79 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.87 (s, 6H, 2 × OMe), 5.23 (s, 2H, C-1''), 6.96–6.98 (m, 2H, ArH), 7.12–7.31 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 30.5 (C-2), 36.7 (C-3), 56.1 (OMe), 58.6 (OMe), 67.7 (C-1''), 112.3 (C-4'), 120.1 (C-6'), 120.7 (C-5'), 127.2 (C-1'), 143.8 (C-3'), 156.5 (C-2'), 169.9 (C-1), 177.4 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 286.70 (17) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 282 (3.7)
<i>Va</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3067 (ArH), 1799 (COCl), 1743 (C=O), 1591, 1544, 1431 (C=C, ArH), 1245, 1135, 1029 (C—O), 727 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.66 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.78 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.82 (s, 6H, 2 × OMe), 5.23 (s, 2H, C-1''), 6.73 (m, 1H, ArH), 6.92 (m, 1H, ArH), 7.18 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.4 (C-2), 36.3 (C-3), 56.3 (2 × OMe), 67.2 (C-1''), 99.8 (C-3'), 105.9 (C-5'), 115.8 (C-1'), 128.2 (C-6'), 158.6 (C-2'), 159.6 (C-4'), 169.2 (C-1), 176.7 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 286.70 (18) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 282 (3.7)
<i>VIa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3069 (ArH), 1790 (COCl), 1747 (C=O), 1594, 1541, 1433 (C=C, ArH), 1249, 1131, 1022 (C—O), 724 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.60 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.74 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.89 (s, 6H, 2 × OMe), 5.14 (s, 2H, C-1''), 6.96 (m, 2H, ArH), 7.46 (m, 1H, ArH), 7.80 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.6 (C-2), 36.1 (C-3), 56.6 (2 × OMe), 64.8 (C-1''), 113.3 (C-3'), 120.1 (C-6'), 121.1 (C-1'), 128.8 (C-4'), 128.8 (C-6'), 159.2 (C-2'), 169.9 (C-1), 179.7 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 286.70 (14) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 282 (3.7)
<i>VIIa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3077 (ArH), 1799 (COCl), 1738 (C=O), 1598, 1547, 1431 (C=C, ArH), 1241, 1129, 1021 (C—O), 725 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.79 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.93 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.84 (s, 6H, 2 × OMe), 5.19 (s, 2H, C-1''), 6.66 (m, 1H, ArH), 6.91 (m, 1H, ArH), 7.40 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.5 (C-2), 36.3 (C-3), 55.2 (2 × OMe), 68.4 (C-1''), 111.7 (C-5'), 113.3 (C-2'), 120.7 (C-6'), 123.5 (C-1'), 147.8 (C-4'), 149.1 (C-3'), 167.4 (C-1), 176.6 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 286.70 (19) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 282 (3.7)
<i>VIIIa</i>	IR, $\tilde{\nu}$ /cm ⁻¹ : 3046 (ArH), 1789 (COCl), 1733 (C=O), 1601, 1548, 1432 (C=C, ArH), 1243, 1127, 1019 (C—O), 726 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.67 (t, 2H, <i>J</i> = 6.5 Hz, C-2), 2.87 (t, 2H, <i>J</i> = 6.5 Hz, C-3), 3.83, 3.84 (s, 6H, 2 × OMe), 5.16 (s, 2H, C-1''), 6.92–6.53 (m, 3H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.2 (C-2), 36.1 (C-3), 57.4 (2 × OMe), 68.9 (C-1''), 98.2 (C-4'), 104.6 (C-2', C-6'), 136.7 (C-1'), 160.7 (C-3', C-5'), 168.5 (C-1), 173.3 (C-4) MS, <i>m/z</i> (<i>I_r</i> /%): 286.70 (11) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ϵ): 278 (3.4)

Table 2. (continued)

Compound	Spectral data
<i>IXa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3085 (ArH), 1788 (COCl), 1734 (C=O), 1602, 1549, 1433 (C=C, ArH), 1365 (NO ₂), 1244, 1128, 1018 (C—O), 724 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.81 (t, 2H, J = 6.5 Hz, C-2), 2.98 (t, 2H, J = 6.5 Hz, C-3), 5.14 (s, 2H, C-1''), 7.54–7.59 (2H, m, ArH), 7.75 (1H, m, ArH), 7.92 (1H, m, ArH) ¹³ C NMR (CDCl ₃), δ : 30.2 (C-2), 35.5 (C-3), 64.7 (C-1''), 113.7 (C-6''), 126.2 (C-3''), 129.5 (C-4''), 131.1 (C-5''), 135.3 (C-1''), 148.3 (C-2''), 169.7 (C-1), 172.3 (C-4) MS, m/z (Ir/%): 271.66 (16) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 238 (3.1)
<i>Xa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3087 (ArH), 1786 (COCl), 1735 (C=O), 1603, 1559, 1443 (C=C, ArH), 1371 (NO ₂), 1255, 1138, 1028 (C—O), 722 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.83 (t, 2H, J = 6.5 Hz, C-2), 2.99 (t, 2H, J = 6.5 Hz, C-3), 5.21 (s, 2H, C-1''), 7.45 (m, 1H, ArH), 7.76 (1H, m, ArH), 7.95–7.99 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.8 (C-2), 36.7 (C-3), 66.7 (C-1''), 121.9 (C-4''), 123.6 (C-2''), 128.9 (C-5''), 132.3 (C-6''), 141.2 (C-1''), 147.2 (C-3''), 167.8 (C-1), 173.6 (C-4) MS, m/z (Ir/%): 271.66 (14) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 238 (3.1)
<i>XIa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3083 (ArH), 1785 (COCl), 1734 (C=O), 1601, 1558, 1441 (C=C, ArH), 1375 (NO ₂), 1251, 1134, 1025 (C—O), 727 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.78 (t, 2H, J = 6.5 Hz, C-2), 2.98 (t, 2H, J = 6.5 Hz, C-3), 5.19 (s, 2H, C-1''), 7.57 (m, 2H, ArH), 7.98–8.05 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.3 (C-2), 35.8 (C-3), 66.9 (C-1''), 123.2 (C-3', C-5'), 127.1 (C-2', C-6'), 141.6 (C-1''), 145.7 (C-4''), 169.5 (C-1), 173.2 (C-4) MS, m/z (Ir/%): 271.66 (14) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 238 (3.1)
<i>XIIa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3053 (ArH), 1787 (COCl), 1738 (C=O), 1603, 1551, 1439 (C=C, ArH), 1257, 1138, 1021 (C—O), 721 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.67 (t, 2H, J = 6.5 Hz, C-2), 2.85 (t, 2H, J = 6.5 Hz, C-3), 5.11 (s, 2H, C-1''), 7.22 (m, 1H, ArH), 7.29–7.34 (m, 2H, ArH), 7.65 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.7 (C-2), 35.6 (C-3), 66.7 (C-1''), 126.2 (C-5'), 127.2 (C-6'), 128.1 (C-3'), 128.3 (C-4'), 129.8 (C-2'), 137.8 (C-1''), 169.9 (C-1), 174.3(C-4) MS, m/z (Ir/%): 260.95 (17) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 268 (4.1)
<i>XIIIa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3059 (ArH), 1791 (COCl), 1736 (C=O), 1605, 1548, 1432 (C=C, ArH), 1251, 1132, 1019 (C—O), 726 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.68 (t, 2H, J = 6.5 Hz, C-2), 2.87 (t, 2H, J = 6.5 Hz, C-3), 5.19 (s, 2H, C-1''), 7.31–7.36 (m, 2H, ArH), 7.43 (m, 1H, ArH), 7.59 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.7 (C-2), 35.5 (C-3), 64.4 (C-1''), 123.7 (C-6'), 125.4 (C-2'), 126.2 (C-4'), 128.8 (C-5'), 133.0 (C-3''), 137.6 (C-1''), 168.5 (C-1), 174.4 (C-4) MS, m/z (Ir/%): 260.95 (18) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 268 (4.1)
<i>XIVa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3061 (ArH), 1789 (COCl), 1737 (C=O), 1606, 1549, 1436 (C=C, ArH), 1252, 1131, 1018 (C—O), 724 (C—Cl) ¹ H NMR (CDCl ₃), δ : 2.72 (t, 2H, J = 6.5 Hz, C-2), 2.88 (t, 2H, J = 6.5 Hz, C-3), 5.21 (s, 2H, C-1''), 7.28 (m, 2H, ArH), 7.39 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.6 (C-2), 35.7 (C-3), 65.9 (C-1''), 128.5 (C-3', C-5'), 129.2 (C-2', C-6'), 132.7 (C-4'), 133.7 (C-1''), 168.9 (C-1), 174.5 (C-4) MS, m/z (Ir/%): 260.95 (15) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 268 (4.1)
<i>XVa</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3063 (ArH), 1788 (COCl), 1739 (C=O), 1602, 1575, 1431 (C=C, ArH), 1258, 1139, 1028 (C—O), 722 (C—Cl), 521 (C—Br) ¹ H NMR (CDCl ₃), δ : 2.75 (t, 2H, J = 6.5 Hz, C-2), 2.83 (t, 2H, J = 6.5 Hz, C-3), 5.12 (s, 2H, C-1''), 7.23–7.33 (m, 3H, ArH), 7.55 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 29.3 (C-2), 35.5 (C-3), 63.9 (C-1''), 125.2 (C-2'), 126.2 (C-5'), 128.0 (C-3'), 128.3 (C-4'), 130.2 (C-6'), 136.6 (C-1''), 168.8 (C-1), 174.4 (C-4) MS, m/z (Ir/%): 305.76 (12) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 268 (4.1)

Table 2. (continued)

Compound	Spectral data
XVIa	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3064 (ArH), 1786 (COCl), 1729 (C=O), 1599, 1567, 1443 (C=C, ArH), 1253, 1144, 1024, 723 (C—Cl), 523 (C—Br) ^1H NMR (CDCl_3), δ : 2.73 (t, 2H, $J = 6.5$ Hz, C-2), 2.84 (t, 2H, $J = 6.5$ Hz, C-3), 5.18 (s, 2H, C-1''), 7.25 (m, 1H, ArH), 7.43 (m, 1H, ArH), 7.50–7.54 (m, 2H, ArH) ^{13}C NMR (CDCl_3), δ : 29.8 (C-2), 36.6 (C-3), 66.8 (C-1''), 121.7 (C-3'), 124.5 (C-6'), 128.1 (C-5'), 128.9 (C-4') MS, m/z ($I_r/\%$): 305.76 (15) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 268 (4.1)
XVIIa	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3061 (ArH), 1787 (COCl), 1731 (C=O), 1598, 1569, 1441 (C=C, ArH), 1257, 1142, 1021, 725 (C—Cl), 525 (C—Br) ^1H NMR (CDCl_3), δ : 2.73 (t, 2H, $J = 6.5$ Hz, C-2), 2.82 (t, 2H, $J = 6.5$ Hz, C-3), 5.22 (s, 2H, C-1''), 7.21 (m, 2H, ArH), 7.86 (m, 2H, ArH) ^{13}C NMR (CDCl_3), δ : 29.4 (C-2), 36.5 (C-3), 66.2 (C-1''), 121.4 (C-4'), 128.7 (C-2', C-6'), 130.6 (C-3', C-5'), 136.3 (C-1'), 167.8 (C-1), 173.6 (C-4) MS, m/z ($I_r/\%$): 305.76 (13) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 268 (4.1)
XVIIIa	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3075 (ArH), 1791 (COCl), 1736 (C=O), 1593, 1561, 1446 (C=C, ArH), 1261, 1147, 1025, 723 (C—Cl), 487 (C—I) ^1H NMR (CDCl_3), δ : 2.76 (t, 2H, $J = 6.9$ Hz, C-2), 2.85 (t, 2H, $J = 7.3$ Hz, C-3), 5.20 (s, 2H, C-1''), 7.11–7.14 (m, 2H, ArH), 7.34 (m, H, ArH), 7.81 (m, H, ArH) ^{13}C NMR (CDCl_3), δ : 29.1 (C-2), 36.6 (C-3), 66.9 (C-1''), 94.5 (C-2'), 126.7 (C-5'), 127.5 (C-6'), 128.6 (C-4'), 138.5 (C-3'), 139.3 (C-1'), 167.7 (C-1), 173.3 (C-4) MS, m/z ($I_r/\%$): 352.65 (12) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 278 (3.7)
XIXa	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3077 (ArH), 1793 (COCl), 1738 (C=O), 1595, 1562, 1448 (C=C, ArH), 1263, 1148, 1026, 723 (C—Cl), 485 (C—I) ^1H NMR (CDCl_3), δ : 2.77 (t, 2H, $J = 6.9$ Hz, C-2), 2.88 (t, 2H, $J = 6.9$ Hz, C-3), 5.25 (s, 2H, C-1''), 7.16 (t, 1H, $J = 7.8$ Hz, ArH), 7.31 (d, 1H, $J = 7.8$ Hz, ArH), 7.67 (d, $J = 7.8$ Hz, 1H, ArH), 7.72 (s, 1H, ArH) ^{13}C NMR (CDCl_3), δ : 29.3 (C-2), 36.2 (C-3), 66.5 (C-1''), 94.6 (C-3'), 126.7 (C-6'), 129.6 (C-5'), 135.8 (C-4'), 138.5 (C-2'), 139.4 (C-1'), 167.4 (C-1), 173.5 (C-4) MS, m/z ($I_r/\%$): 352.65 (11) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 278 (3.7)
XXa	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3064 (ArH), 1786 (COCl), 1729 (C=O), 1599, 1567, 1443 (C=C, ArH), 1253, 1144, 1024, 723 (C—Cl), 523 (C—Br) ^1H NMR (CDCl_3), δ : 2.73 (t, 2H, $J = 6.5$ Hz, C-2), 2.84 (t, 2H, $J = 6.5$ Hz, C-3), 5.18 (s, 2H, C-1''), 7.25 (m, 1H, ArH), 7.43 (m, 1H, ArH), 7.50–7.54 (m, 2H, ArH) ^{13}C NMR (CDCl_3), δ : 29.8 (C-2), 36.6 (C-3), 66.8 (C-1''), 121.7 (C-3'), 124.5 (C-6'), 128.1 (C-5'), 128.9 (C-4') MS, m/z ($I_r/\%$): 352.65 (15) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 268 (4.1)

Twenty (*I*–*XX*) benzyl hydrogen succinates were prepared by following the procedure described previously (Iqbal et al., 2012; Cason & Prout, 1944, 1948; Von Rudloff, 1958). In brief, the corresponding benzyl alcohols (15 mmol) were added to succinic anhydride (15 mmol), *p*-toluenesulfonic acid (12.5 g, 0.06 mmol), and toluene (15 mL) in a single-necked round-bottom flask (100 mL) equipped with a magnetic stirrer, Dean–Stark trap, and a reflux condenser. After refluxing the mixture for 14 h under nitrogen, it was cooled to ambient temperature, treated with a saturated aqueous solution of sodium bicarbonate and extracted with hexane (3×50 mL). The extract was dried over MgSO_4 , the solvent was removed, and the residue was subjected to open column chromatography on silica gel using a mixture of hexane/ethyl acetate ($\varphi_r = 4 : 1 \rightarrow 1 : 4$) as the eluent to obtain the pure monoesters *I*–*XX*.

General procedure for preparation of 4-mono-/di-benzylxy-4-ketobutanoyl chlorides (Ia–XXa)

Acid chlorides (*Ia*–*XXa*) were prepared by a published procedure (Cason & Prout, 1944, 1948; Von Rudloff, 1958). In brief, the required monoester (*I*–*XX*; 8 mmol) and thionyl chloride (1.64 mL, 22.5 mmol) were mixed in a round-bottom reaction flask (100 mL) fitted with a reflux condenser. The mixture was heated to 30–40°C for different lengths of time (Table 1) and the excess thionyl chloride was removed by heating at reduced pressure. The residue was subjected to open column chromatography on silica gel using a mixture of hexane/ethyl acetate ($\varphi_r = 4 : 1 \rightarrow 1 : 4$) as the eluent to obtain pure *Ia*–*XXa*. The product was characterised by the physical characteristics (Table 1) and spectroscopic data (Table 2).

Table 3. Physical characteristics of *Ib*–*XXb*

Entry	Ar	Time		Yield %	Formula	<i>M</i> _r	Physical state	<i>w_i</i> (calc.)/% <i>w_i</i> (found)/%					
		h	%					C	H	N	Cl	Br	I
<i>Ib</i>	2-MeO-C ₆ H ₄	13	69	C ₁₄ H ₁₈ O ₄	250.29	Viscous oil	67.18 67.43	7.25 7.16	—	—	—	—	—
<i>IIb</i>	3-MeO-C ₆ H ₄	12	73	C ₁₄ H ₁₈ O ₄	250.29	Viscous oil	67.18 67.32	7.25 7.13	—	—	—	—	—
<i>IIIb</i>	4-MeO-C ₆ H ₄	13	71	C ₁₄ H ₁₈ O ₄	250.29	Viscous oil	67.18 67.13	7.25 7.36	—	—	—	—	—
<i>IVb</i>	2,3-(MeO) ₂ -C ₆ H ₃	11	74	C ₁₅ H ₂₀ O ₅	280.11	Viscous oil	64.27 64.12	7.19 7.14	—	—	—	—	—
<i>Vb</i>	2,4-(MeO) ₂ -C ₆ H ₃	11	79	C ₁₅ H ₂₀ O ₅	280.11	Viscous oil	64.27 64.18	7.19 7.23	—	—	—	—	—
<i>VIb</i>	2,5-(MeO) ₂ -C ₆ H ₃	13	75	C ₁₅ H ₂₀ O ₅	280.11	Viscous oil	64.27 64.23	7.19 7.11	—	—	—	—	—
<i>VIIb</i>	3,4-(MeO) ₂ -C ₆ H ₃	14	74	C ₁₅ H ₂₀ O ₅	280.11	Viscous oil	64.27 64.34	7.19 7.11	—	—	—	—	—
<i>VIIIb</i>	3,5-(MeO) ₂ -C ₆ H ₃	12	71	C ₁₅ H ₂₀ O ₅	280.11	Viscous oil	64.27 64.19	7.19 7.35	—	—	—	—	—
<i>IXb</i>	2-NO ₂ -C ₆ H ₄	13	68	C ₁₃ H ₁₅ NO ₅	265.12	Viscous oil	58.86 58.95	5.70 5.56	5.28 5.32	—	—	—	—
<i>Xb</i>	3-NO ₂ -C ₆ H ₄	13	66	C ₁₃ H ₁₅ NO ₅	265.12	Viscous oil	58.86 56.73	5.70 5.67	5.28 5.38	—	—	—	—
<i>XIb</i>	4-NO ₂ -C ₆ H ₄	13	59	C ₁₃ H ₁₅ NO ₅	265.12	Viscous oil	58.86 56.95	5.70 5.66	5.28 5.45	—	—	—	—
<i>XIIb</i>	2-Cl-C ₆ H ₄	12	59	C ₁₃ H ₁₅ ClO ₃	254.71	Viscous oil	61.30 61.28	5.94 5.77	—	13.92 13.99	—	—	—
<i>XIIIb</i>	3-Cl-C ₆ H ₄	13	64	C ₁₃ H ₁₅ ClO ₃	254.71	Viscous oil	61.30 61.19	5.94 5.83	—	13.92 13.87	—	—	—
<i>XIVb</i>	4-Cl-C ₆ H ₄	14	64	C ₁₃ H ₁₅ ClO ₃	254.71	Viscous oil	61.30 61.55	5.94 5.45	—	13.92 13.88	—	—	—
<i>XVb</i>	2-Br-C ₆ H ₄	12	63	C ₁₃ H ₁₅ BrO ₃	299.11	Viscous oil	52.19 52.23	5.05 5.12	—	—	26.71 25.72	—	—
<i>XVIb</i>	3-Br-C ₆ H ₄	14	59	C ₁₃ H ₁₅ BrO ₃	299.11	Viscous oil	52.19 52.33	5.05 4.87	—	—	26.71 26.68	—	—
<i>XVIIb</i>	4-Br-C ₆ H ₄	14	60	C ₁₃ H ₁₅ BrO ₃	299.11	Viscous oil	52.19 52.21	5.05 5.02	—	—	26.71 26.85	—	—
<i>XVIIIb</i>	2-I-C ₆ H ₄	15	59	C ₁₃ H ₁₅ IO ₃	346.09	Viscous oil	45.11 45.31	4.37 4.16	—	—	—	36.66 36.55	—
<i>XIXb</i>	3-I-C ₆ H ₄	16	67	C ₁₃ H ₁₅ IO ₃	346.09	Viscous oil	45.11 44.87	4.37 4.46	—	—	—	36.66 36.45	—
<i>XXb</i>	4-I-C ₆ H ₄	14	66	C ₁₃ H ₁₅ IO ₃	346.09	Viscous oil	45.11 45.24	4.37 4.15	—	—	—	36.66 36.67	—

Preparation of diethylcadmium reagent

The diethylcadmium reagent was prepared from a freshly prepared Grignard solution (Cason & Prout, 1944, 1948; Von Rudloff, 1958). Dry CdCl₂ (2.15 g, 11.8 mmol) was added to the Grignard reagent (prepared from ethyl bromide (1.06 g, 11.8 mmol) and magnesium metal (0.26 g, 11.8 mmol) and cooled to 0°C) and dry diethylether (60 mL) over a period of 10 min with vigorous stirring. Stirring was continued for a further hour and the diethylcadmium reagent was used as prepared.

General procedure for preparation of benzyl-γ-ketohexanoates (*Ib*–*XXb*)

All γ-ketohexanoates (*Ib*–*XXb*) were prepared from acid chlorides (*Ia*–*XXa*), by following the methodologies described by Cason and Prout (1944, 1948) and Von Rudloff (1958). For the preparation of compounds *Ib*–*XXb*, the corresponding 4-benzylxylo-4-ketobutanoyl chloride (5 mmol) was slowly added to the freshly prepared solution of diethyl cadmium. After maintaining reflux for different lengths of time (Table 3), the mixture was allowed to stand overnight. The material was then poured into a beaker contain-

Table 4. Spectroscopic data of synthesised γ -ketohexanoates (*Ib*–*VIIb*) from acid halides

Compound	Spectral data
<i>Ib</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3057 (ArH), 1749 (C=O), 1717 (RO—C=O), 1595, 1546, 1429 (C=C, ArH), 1239, 1127, 1026 ¹ H NMR (CDCl ₃), δ : 1.05 (t, 3H, J = 4 Hz, C-6), 2.55 (q, J = 7.4 Hz, 2H, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.82 (t, 2H, J = 6.8 Hz, C-3), 3.86 (s, 3H, OMe), 5.22 (s, 1H, C-1''), 6.81–6.98 (m, 3H, ArH), 7.26 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 28.3 (C-2), 34.2 (C-3), 36.1 (C-5), 56.3 (OMe), 62.8 (C-1''), 123.7 (C-5'), 124.7 (C-1'), 127.5 (C-6''), 128.9 (C-4'), 129.3 (C-3'), 155.6 (C-2'), 172.2 (C-1), 212.1 (C-4) MS, m/z (I_r /%): 250.29 (23) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 271.3 (3.3)
<i>IIb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3065 (ArH), 1725, 1714 (C=O), 1592, 1545, 1431 (C=C), 1230, 1128, 1025 ¹ H NMR (CDCl ₃), δ : 1.05 (t, 3H, J = 7.4 Hz, C-6), 2.57 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.83 (t, 2H, J = 6.8 Hz, C-3), 3.82 (s, 3H, OMe), 5.21 (s, 2H, C-1''), 6.96–6.99 (m, 2H, ArH), 7.11 (m, 1H, ArH), 7.32 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.3 (C-2), 31.3 (C-3), 36.1 (C-5), 54.9 (OMe), 65.4 (C-1''), 112.2 (C-2'), 113.3 (C-4'), 118.3 (C-6''), 129.3 (C-5'), 138.8 (C-1'), 159.4 (C-3'), 172.2 (C-1), 212.1 (C-4) MS, m/z (I_r /%): 250.29 (29) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 271.6 (3.5)
<i>IIIb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3069 (ArH), 1747 (C=O), 1722 (CO ₂ Ar), 1596, 1543, 1430 (C=C), 1243, 1129, 1023 ¹ H NMR (CDCl ₃), δ : 1.04 (t, 3H, J = 7.4 Hz, C-6), 2.56 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.82 (t, 2H, J = 6.8 Hz, C-3), 3.76 (s, 3H, OMe), 5.21 (s, 2H, C-1''), 6.94–6.96 (m, 2H, ArH), 7.17–7.19 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.8 (C-2), 33.4 (C-3), 34.2 (C-5), 55.4 (OMe), 66.2, (C-1''), 113.6 (C-3', C-5'), 129.2 (C-1'), 129.3 (C-2', C-6''), 159.1 (C-4'), 172.2 (C-1), 212.1 (C-4) MS, m/z (I_r /%): 250.29 (25) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 271.1 (3.6)
<i>IVb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3085 (ArH), 2850, 1757 (C=O), 1723 (CO ₂ Ar), 1592, 1538, 1435 (C=C), 1237, 1133, 1041 ¹ H NMR (CDCl ₃), δ : 1.02 (t, 3H, J = 7.4 Hz, C-6), 2.61 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.85 (t, 2H, J = 6.8 Hz, C-3), 3.79 (s, 3H, OMe), 3.82 (s, 3H, OMe), 5.13 (s, 2H, C-1''), 6.67–6.99 (m, 3H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.3 (C-2), 32.4 (C-3), 34.9 (C-5), 55.6 (OMe), 59.7 (OMe), 62.0 (C-1''), 115.5 (C-4'), 119.1 (C-6''), 124.2 (C-5'), 127.6 (C-1'), 138.9 (C-2'), 147.2 (C-3'), 174.2 (C-1), 213.1 (C-4) MS, m/z (I_r /%): 280.11 (27) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 278.8 (3.7)
<i>Vb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3054 (ArH), 1754 (C=O), 1726 (CO ₂ Ar), 1600, 1540, 1436 (C=C), 1290, 1135 ¹ H NMR (CDCl ₃), δ : 1.05 (t, 3H, J = 7.4 Hz, C-6), 2.62 (q, 2H, J = 7.4 Hz, C-5), 2.79 (t, 2H, J = 6.8 Hz, C-2), 2.85 (t, 2H, J = 6.8 Hz, C-3), 3.78 (s, 3H, OMe), 3.85 (s, 6H, 2 × OMe), 5.21 (s, 2H, C-1''), 6.63 (s, 1H, ArH), 6.63 (m, 1H, ArH), 6.84 (m, 1H, ArH), 7.41 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.3 (C-2), 33.2 (C-3), 36.1 (C-5), 55.4 (OMe), 56.5 (OMe), 62.8 (C-1''), 100.4 (C-3'), 108.5 (C-5'), 116.1 (C-1'), 128.8 (C-6''), 158.3 (C-2'), 162.6 (C-4'), 171.2 (C-1), 214.1 (C-4) MS, m/z (I_r /%): 280.11 (24) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 278.6 (3.87)
<i>VIb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3092 (ArH), 1753 (C=O), 1722 (ArO—C=O), 1605, 1595, 1435 (C=C), 1245 (C=O), 1125 ¹ H NMR (CDCl ₃), δ : 1.04 (t, 3H, J = 7.4 Hz, C-6), 2.49 (q, 2H, J = 7.4 Hz, C-4), 2.73 (t, 2H, J = 6.8 Hz, C-2), 2.86 (t, 2H, J = 6.8 Hz, C-3), 3.82 (s, 6H, 2 × OMe), 5.21 (s, 2H, C-1''), 6.82–7.33 (m, 3H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.6 (C-2), 32.6 (C-3), 35.3 (C-5), 56.3 (2 × OMe), 63.4 (C-1''), 114.8 (C-6''), 115.8 (C-4'), 124.6 (C-3'), 128.7 (C-1'), 148.5 (C-2'), 155.5 (C-5'), 171.4 (C-1), 214.7 (C-4) MS, m/z (I_r /%): 280.11 (18) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 282.5 (3.6)
<i>VIIb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 3086 (ArH), 1755 (C=O), 1721 (CO ₂ Ar), 1603, 1592, 1433 (C=C), 1247, 1128 ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.48 (q, 2H, J = 7.4 Hz, C-5), 2.75 (t, 2H, J = 6.8 Hz, C-2), 2.85 (t, 2H, J = 6.8 Hz, C-3), 3.81 (s, 6H, 2 × OMe), 5.17 (s, 2H, C-1''), 6.43–7.21 (m, 3H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 31.2 (C-2), 35.8 (C-3), 36.1 (C-5), 55.7 (2 × OMe), 66.3 (C-1''), 111.4 (C-5'), 121.2 (C-6'), 124.7 (C-1'), 146.3 (C-4'), 147.3 (C-2'), 148.7 (C-3'), 171.8 (C-1), 214.4 (C-4) MS, m/z (I_r /%): 280.11 (18) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 279.7 (3.8)
<i>VIIIb</i>	IR, $\tilde{\nu}$ /cm ^{−1} : 1751 (C=O), 1726 (CO ₂ Ar), 1592, 1538, 1515, 1435 (C=C), 1235, 1155, 1141, 1133, 1041, 1022 ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.44 (q, 2H, J = 7.4 Hz, C-5), 2.71 (t, 2H, J = 6.8 Hz, C-2), 2.81 (t, 2H, J = 6.8 Hz, C-3), 3.84 (s, 6H, 2 × OMe), 5.19 (s, 2H, C-1''), 6.45–6.99 (m, 3H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.2 (C-2), 32.7 (C-3), 36.4 (C-5), 55.6 (s, 2 × OMe), 66.7 (C-1''), 98.7 (C-4'), 105.1 (C-2', C-6'), 137.2 (C-1'), 158.4 (C-3', C-5'), 171.1 (C-1), 215.4 (C-4) MS, m/z (I_r /%): 280.11 (22) (M ⁺) UV-VIS (MeOH), λ_{max} /nm (log ε): 279.9 (3.8)

Table 4. (continued)

Compound	Spectral data
<i>IXb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3092 (ArH), 1746 (C=O), 1723 (CO ₂ R), 1596, 1530, 1441 (C=C, ArH), 1375 (NO ₂), 1244, 1139, 1045 ¹ H NMR (CDCl ₃), δ : 1.06 (t, J = 7.4 Hz, 3H, CH ₃ , C-6), 2.57 (q, J = 7.4 Hz, 2H, CH ₂ , C-5), 2.72 (t, J = 6.8 Hz, 2H, C-2), 2.85 (t, J = 6.8 Hz, 2H, C-3), 5.19 (s, 2H, C-1''), 7.62 (m, H, ArH), 7.83 (m, 1H, ArH), 8.19–8.26 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.1 (C-2), 32.6 (C-3), 36.7 (C-5), 65.1 (C-1''), 124.7 (C-3'), 129.2 (C-4'), 129.9 (C-5'), 134.3 (C-1'), 134.3 (C-6'), 147.4 (C-2'), 171.2 (C-1), 213.2 (C-4) MS, m/z (I _r /%): 265.12 (16) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 267 (4.0)
<i>Xb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3086 (ArH), 1757, 1721 (C=O), 1597, 1539, 1440 (C=C), 1374 (NO ₂), 1249, 1137, 1044 ¹ H NMR (CDCl ₃), δ : 1.06 (t, 3H, J = 7.4 Hz, C-6), 2.57 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.80 (t, 2H, J = 6.8 Hz, C-3), 5.19 (s, 2H, C-1''), 7.57 (m, 1H, ArH), 7.79 (m, 1H, ArH), 8.18–8.24 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.3 (C-2), 36.1 (C-3), 36.2 (C-5), 65.6 (C-1'), 123.3 (C-2'), 123.8 (C-4'), 130.6 (C-5'), 132.4 (C-6'), 143.2 (C-1'), 147.8 (C-3'), 171.4 (C-1), 213.8 (C-4) MS, m/z (I _r /%): 265.12 (19) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 267.5 (4.0)
<i>XIb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3071 (ArH), 1753, 1722 (C=O), 1595, 1535, 1440 (C=C), 1372 (NO ₂), 1248, 1136, 1045 ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.57 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.82 (t, 2H, J = 6.8 Hz, C-3), 5.18 (s, 2H, C-1''), 7.59 (m, 2H, ArH), 8.20 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.8 (C-2), 31.4 (C-3), 36.6 (C-5), 66.3 (C-1''), 123.7 (C-3', C-5'), 129.9 (C-2', C-6'), 143.4 (C-1'), 147.4 (C-4'), 171.6 (C-1), 214.2 (C-4) MS, m/z (I _r /%): 265.12 (14) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 267.6 (4.2)
<i>XIIb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3077 (ArH), 1748 (C=O), 1721 (CO ₂ R), 1597, 1529, 1497, 1456 (C=C), 1387, 1251, 1237, 1134, 1208, 1094, 1047, 1015, 723 (C—Cl) ¹ H NMR (CDCl ₃), δ : 1.04 (t, 3H, J = 7.4 Hz, C-6), 2.59 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.82 (t, 2H, J = 6.8 Hz, C-3), 5.10 (s, 2H, C-1''), 7.28 (m, 1H, ArH), 7.31–7.33 (m, 2H, ArH), 7.65 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 28.5 (C-2), 30.6 (C-3), 36.1 (C-5), 62.2 (C-1''), 127.1 (C-6'), 127.4 (C-5'), 129.2 (C-3'), 129.6 (C-2'), 131.2 (C-4'), 138.4 (C-1'), 174.2 (C-1), 213.2 (C-4) MS, m/z (I _r /%): 254.71 (19) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 266.5 (4.1)
<i>XIIIb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3079 (ArH), 1739, 1722 (C=O), 1599, 1525, 1452 (C=C), 1248, 1133, 727 (C—Cl) ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.42 (q, 2H, J = 7.4 Hz, C-5), 2.72 (t, 2H, J = 6.8 Hz, C-2), 2.81 (t, 2H, J = 6.8 Hz, C-3), 5.19 (s, 2H, C-1''), 7.33–7.36 (m, 2H, ArH), 7.46 (m, 1H, ArH), 7.59 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 29.3 (C-2), 36.1 (C-3), 36.2 (C-5), 65.7 (C-1''), 124.1 (C-6'), 125.8 (C-2'), 128.1 (C-4'), 130.1 (C-5'), 133.5 (C-3'), 138.6 (C-1'), 172.0 (C-1), 212.1 (C-4) MS, m/z (I _r /%): 254.71 (16) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 266.8 (3.7)
<i>XIVb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3093 (ArH), 1754, 1720 (C=O), 1597, 1529 (C=C), 1383, 1251, 1047, 721 (C—Cl) ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.42 (q, 2H, J = 7.4 Hz, C-5), 2.73 (t, J = 6.8 Hz, 2H, C-2), 2.83 (t, J = 6.8 Hz, 2H, C-3), 5.19 (s, 2H, C-1''), 7.38 (m, 2H, ArH), 7.45 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 28.7 (C-2), 32.3 (C-3), 35.7 (C-5), 66.5 (C-1''), 128.6 (C-3', C-5'), 128.8 (C-2', C-6'), 133.8 (C-4'), 133.9 (C-1'), 171.7 (C-1), 213.0 (C-4) MS, m/z (I _r /%): 254.71 (15) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 266.1 (3.7)
<i>XVb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3085 (ArH), 1736, 1715 (C=O), 1596, 1527 (C=C), 1413, 1254, 1054, 535 (C—Br) ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.42 (q, 2H, J = 7.4 Hz, C-5), 2.73 (t, 2H, J = 6.8 Hz, C-2), 2.82 (t, 2H, J = 6.8 Hz, C-3), 5.10 (s, 2H, C-1''), 7.24–7.28 (m, 2H, ArH), 7.37 (m, 1H, ArH), 7.51 (m, 1H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.7 (C-2), 32.2 (C-3), 37.1 (C-5), 66.3 (C-1''), 123.7 (C-2'), 127.5 (C-5'), 128.6 (C-3'), 128.7 (C-4'), 131.2 (C-6'), 139.7 (C-1'), 173.2 (C-1), 214.2 (C-4) MS, m/z (I _r /%): 299.11 (18) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 263.6 (3.9)
<i>XVIIb</i>	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3085 (ArH), 1739, 1718 (C=O), 1591, 1522, 1451 (C=C), 1417, 1271, 1252, 1140, 1051, 528 (C—Br) ¹ H NMR (CDCl ₃), δ : 1.03 (t, 3H, J = 7.4 Hz, C-6), 2.43 (q, 2H, J = 7.4 Hz, C-5), 2.75 (t, 2H, J = 6.8 Hz, C-2), 2.83 (t, 2H, J = 6.8 Hz, C-3), 5.20 (s, 2H, C-1''), 7.29 (m, 1H, ArH), 7.44 (m, 1H, ArH), 7.50–7.55 (m, 2H, ArH) ¹³ C NMR (CDCl ₃), δ : 7.7 (C-6), 27.8 (C-2), 32.3 (C-3), 37.2 (C-5), 66.3 (C-1''), 123.5 (C-3'), 125.8 (C-6'), 129.1 (C-5'), 130.3 (C-4'), 131.7 (C-2'), 143.6 (C-1'), 173.4 (C-1), 214.4 (C-4) MS, m/z (I _r /%): 299.11 (21) (M ⁺) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ (log ε): 263.2 (3.8)

Table 4. (continued)

Compound	Spectral data
XVIIb	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3083 (ArH), 1743, 1717 (C=O), 1598, 1532, 1455 (C=C, 1419, 1266, 531 (C—Br) ^1H NMR (CDCl_3), δ : 1.03 (t, 3H, $J = 7.4$ Hz, C-6), 2.44 (q, 2H, $J = 7.4$ Hz, C-5), 2.76 (t, 2H, $J = 6.8$ Hz, C-2), 2.85 (t, 2H, $J = 6.8$ Hz, C-3), 5.21 (s, 2H, C-1''), 7.22 (m, 2H, ArH), 7.87 (m, 2H, ArH) ^{13}C NMR (CDCl_3), δ : 7.7 (C-6), 27.3 (C-2), 32.7 (C-3), 37.4 (C-5), 66.5 (C-1''), 121.6 (C-4'), 132.3 (C-3', C-5'), 134.6 (C-1'), 139.7 (C-2', C-6'), 173.8 (C-1), 214.7 (C-4) MS, m/z (Ir/%): 299.11 (22) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 263.9 (3.9)
XVIIIb	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3046 (ArH), 1735, 1718 (C=O), 1592, 1524, 1456 (C=C, 1258, 1156, 488 (C—I) ^1H NMR (CDCl_3), δ : 1.04 (t, 3H, $J = 7.4$ Hz, C-6), 2.41 (q, 2H, $J = 7.4$ Hz, C-5), 2.66 (t, 2H, $J = 6.8$ Hz, C-2), 2.79 (t, 2H, $J = 6.8$ Hz, C-3), 5.10 (s, 2H, C-1''), 7.11–7.15 (m, 2H, ArH), 7.39 (m, 1H, ArH), 7.83 (m, 1H, ArH) ^{13}C NMR (CDCl_3), δ : 7.7 (C-6), 29.2 (C-2), 31.7 (C-3), 35.4 (C-5), 67.6 (C-1''), 92.5 (C-2'), 126.8 (C-6'), 127.7 (C-5'), 129.3 (C-4'), 138.2 (C-3'), 142.7 (C-1'), 169.2 (C-1), 208.5 (C=O) MS, m/z (Ir/%): 346.09 (11) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 260.3 (3.6)
XIXb	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3042 (ArH), 1732, 1719 (C=O), 1591, 1529, 1467 (C=C, 1251, 1161, 1058, 492 (C—I) ^1H NMR (CDCl_3), δ : 1.03 (t, 3H, $J = 7.4$ Hz, C-6), 2.40 (q, 2H, $J = 7.4$ Hz, C-5), 2.64 (t, 2H, $J = 6.8$ Hz, C-2), 2.77 (t, 2H, $J = 6.8$ Hz, C-3), 5.18 (s, 2H, C-1''), 7.15 (m, 1H, ArH), 7.43 (m, 1H, ArH), 7.75 (m, 1H, ArH), 7.84 (m, H, ArH) ^{13}C NMR (CDCl_3), δ : 7.7 (C-6), 29.4 (C-2), 31.3 (C-3), 35.6 (C-5), 65.1 (C-1''), 94.2 (C-3'), 127.3 (C-6'), 129.7 (C-5'), 136.1 (C-2'), 137.1 (C-4'), 142.2 (C-1'), 169.7 (C-1), 208.1 (C-4) MS, m/z (Ir/%): 346.09 (8) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 263.2 (3.9)
XXb	IR, $\tilde{\nu}/\text{cm}^{-1}$: 3048 (ArH), 1733, 1715 (C=O), 1599, 1521, 1460 (C=C, 1255, 1151, 1054, 483 (C—I) ^1H NMR (CDCl_3), δ : 1.03 (t, 3H, $J = 7.4$ Hz, C-6), 2.38 (q, 2H, $J = 7.4$ Hz, C-5), 2.65 (t, 2H, $J = 6.8$ Hz, C-2), 2.76 (t, 2H, $J = 6.8$ Hz, C-3), 5.19 (s, 2H, C-1''), 7.15 (m, 2H, ArH), 7.65 (m, 2H, ArH) ^{13}C NMR (CDCl_3), δ : 7.7 (C-6), 29.0 (C-2), 31.7 (C-3), 35.8 (C-5), 66.1 (C-1''), 92.3 (C-4'), 129.6 (C-2', C-6'), 134.2 (C-1'), 136.9 (C-3', C-5'), 169.3 (C-1'), 208.9 (C-4) MS, m/z (Ir/%): 346.09 (9) (M^+) UV-VIS (MeOH), $\lambda_{\text{max}}/\text{nm}$ ($\log \varepsilon$): 263.8 (3.9)

ing ice and aq. H_2SO_4 (1 M, 30 mL). The organic layer was extracted with diethyl ether (3×20 mL), washed with an aq. NaHCO_3 solution (20 mL), and dried over anhydrous Na_2SO_4 . After removal of the solvent, the material thus obtained was subjected to open column chromatography on silica gel eluting it with the mixture of hexane/ethyl acetate ($\varphi_r = 4 : 1 \rightarrow 1 : 4$) to gain pure compounds (*Ib*–*XXb*) which were characterised by their physical characteristics (Table 3) and spectroscopic data (Table 4). The reactions were carried out in an efficient fuming hood and the wastes were disposed of safely.

Results and discussion

In the present work, twenty new compounds (*Ib*–*XXb*) were synthesised from acid chlorides (*Ia*–*XXa*).

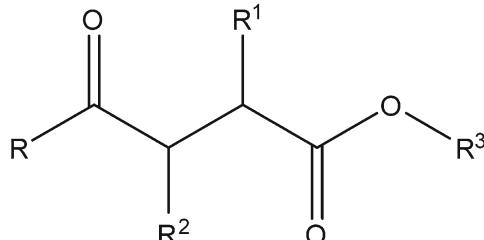


Fig. 1. General structure of γ -ketoesters.

Our γ -ketoesters differ from those previously reported, mainly in the nature of R, R¹, and R² (Fig. 1). In almost all reported examples, R³ is an alkyl or an un-substituted benzyl (Cason et al., 1951) (Fig. 1), whereas in our case R is ethyl, R¹ and R² are H, and

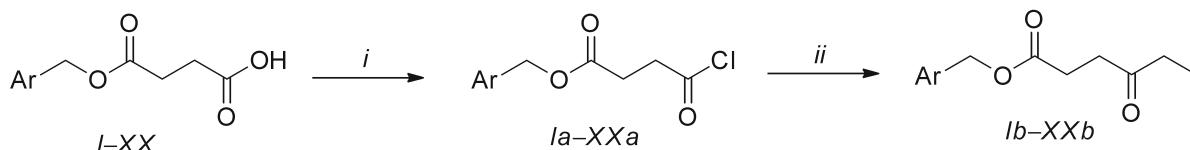


Fig. 2. Preparation of compounds *Ia*–*XXa* and *Ib*–*XXb*: *i*) SOCl_2 , 3 h, 30–40 °C, 63–79 %; *ii*) diethylcadmium, diethylether, 11–16 h, reflux, 59–79 %.

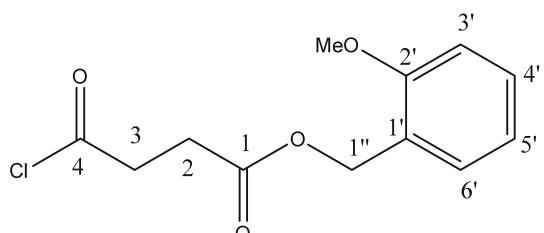
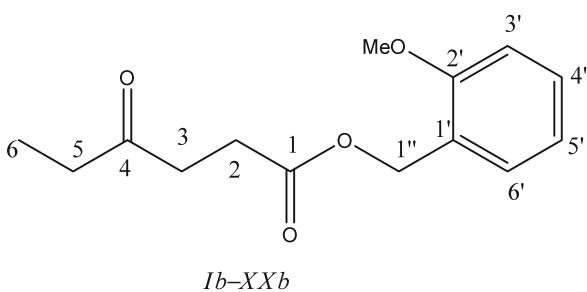
*Ia-XXa**Ib-XXb*

Fig. 3. Atom numbering in compounds for assignment of spectral data.

R^3 is a mono- or di-substituted benzyl group (*Ia-XXa* and *Ib-XXb*) (Figs. 1 and 2). The carbon atoms in acid chlorides (*Ia-XXa*) and ketoesters (*Ib-XXb*) are numbered for spectral data assignment as shown in Fig. 3.

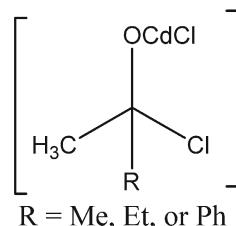


Fig. 4. Tetrahedral intermediate from reaction of $RCdCl$ with CH_3COCl .

Bansal (1996) proposed a mechanism for the reaction of dimethylcadmium with acid halides. The same can apply to diethylcadmium because of the similarity in structure and reactivity of the two. This mechanism is further supported by Roberts and Caserio (1964), who reported on the formation of a tetrahedral intermediate from the reaction of $RCdCl$ and CH_3COCl (Fig. 4).

The proposed mechanism for the reaction of diethylcadmium with acid halide is given below (Fig. 5). The structures of the synthesised compounds (*Ia-XXa* and *Ib-XXb*) were established by various spectroscopic techniques.

Compounds *Ia* and *Ib* were identified as representative of the respective series *Ia-XXa* and *Ib-XXb*, and discussed in detail. The IR spectrum of *Ia* showed no OH peak for the initial monoester *I*. The two new

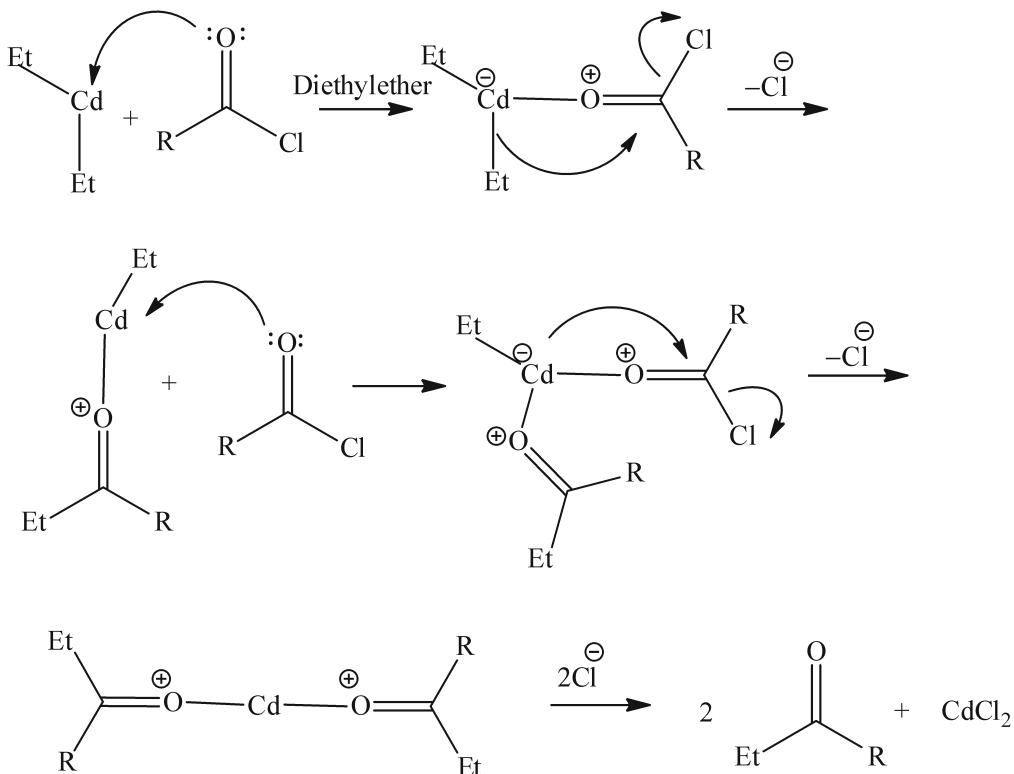


Fig. 5. Proposed mechanism for ketone formation from acid chlorides and diethylcadmium.

peaks observed (723 cm^{-1} and 1792 cm^{-1}) were assigned to C—Cl and Cl—C=O, respectively, confirming the formation of *Ia*. In the ^{13}C NMR spectra, compound *Ia* displaying a peak at δ 175.6 supported the conclusion; this was also verified by ^1H NMR and ^{13}C NMR (Table 2). The assignment was also verified by HMBC spectrum, in which the proton resonating at δ 2.74, which displayed interactions with carbon resonating at δ 175.6, was attributed to COCl (Fig. 5). In the high resolution electron impact MS (HRMS) spectrum, *Ia* displayed a molecular ion peak at m/z 256.6823 (calc. 256.6813) corresponding to $\text{C}_{12}\text{H}_{13}\text{ClO}_4$. Chemical analysis data of *Ia* supported the molecular formula $\text{C}_{12}\text{H}_{13}\text{ClO}_4$.

The structure of the remaining compounds (*IIa*–*XXa*) was established in the same way. Conversion of *Ia* into *Ib* was established by mass spectrum displaying a molecular ion peak in HRMS at m/z 250.2905 for $\text{C}_{14}\text{H}_{18}\text{O}_4$ (calc. 250.2903). 1D (^1H NMR and ^{13}C NMR) spectra similarly supported the predicted formation of *Ib* (Fig. 3, Table 4). Its formation was further verified by the presence of an ethyl moiety by displaying peaks at δ 1.05 (t, 3H, $J = 7.4\text{ Hz}$, C-6) and 2.55 (q, 2H, $J = 7.4\text{ Hz}$, C-5) and ^{13}C NMR peaks at 7.7 (C-6) and 36.1 (C-5).

Conclusions

A highly convenient synthesis of γ -ketoesters is described, using diethyl ether as solvent and diethylcadmium as ethylating agent. Twenty novel γ -ketoesters (mono-/dibenzyl γ -ketohexanoates), denoted as *Ib*–*XXb*, were successfully prepared employing a two-step reaction strategy. In the first step, reaction of the monoesters (*Ia*–*XXa*) of succinic acid with thionyl chloride delivered new mono-/di-benzyl 4-ketobutanoyl chlorides with good yields. In the second step, the products obtained in the reaction with diethylcadmium led to formation of the desired γ -ketoesters.

References

- Arends, I. W. C. E., & Sheldon R. A. (2004). Modern oxidation of alcohols using environmentally benign oxidants. In J. E. Bäckvall (Ed.), *Modern oxidation methods* (pp. 83–118). Weinheim, Germany: Wiley–VCH.
- Ballini, R., Barboni, L., Bosica, G., & Fiorini, D. (2002). One-pot synthesis of γ -diketones, γ -keto esters, and conjugated cyclopentenones from nitroalkanes. *Synthesis*, 18, 2725–2728. DOI: 10.1055/s-2002-35993.
- Bandgar, B. P., Hashmi, A. M., & Pandit, S. S. (2005). Facile and selective transesterification of β -keto esters using NaIO_4 , KIO_4 , and anhydrous CaCl_2 as inexpensive catalysts under neutral conditions. *Journal of the Chinese Chemical Society*, 52, 1101–1104.
- Bansal, R. K. (1996). *Synthetic approaches in organic chemistry*. Sudbury, MA, USA: Jones and Bartlett.
- Brockman, J. A., Jr., & Fabio, P. F. (1957). Syntheses of 6-ethyl-8-mercaptopoctanoic acid and its homologs. *Journal of the American Chemical Society*, 79, 5027–5029. DOI: 10.1021/ja01575a053.
- Cason, J. (1942). Branched-chain fatty acids. I. Synthesis of 17-methyloctadecanoic acid. *Journal of the American Chemical Society*, 64, 1106–1110. DOI: 10.1021/ja01257a029.
- Cason, J. (1946). Branched-chain fatty acids. IV. A further study of the preparation of ketones and keto esters by means of orgaocadmium reagents. *Journal of the American Chemical Society*, 68, 2078–2081. DOI: 10.1021/ja01214a061.
- Cason, J., & Prout, F. S. (1944). Branched-chain fatty acids. II. Syntheses in the C_{19} and C_{25} series. Preparation of keto esters. *Journal of the American Chemical Society*, 66, 46–50. DOI: 10.1021/ja01229a015.
- Cason, J., & Prout, F. S. (1948). Methyl 4-keto-7-methyloctanoate. *Organic Syntheses*, 28, 75.
- Cason, J., Taylor, P. B., & Williams, D. A. (1951). Branched-chain fatty acids. XX. Synthesis of compounds useful for relating melting point to structure. *Journal of Organic Chemistry*, 16, 1187–1192. DOI: 10.1021/jo50002a002.
- Csende, F. (2002). Some alternative synthetic routes to γ - and δ -oxo acid derivatives. *Acta Chimica Slovenica*, 49, 663–676.
- Csende, F., Szabó, Z., & Stájer, G. (1993). Synthesis and structural study of new saturated isoindol-1-one derivatives. *Heterocycles*, 36, 1809–1821. DOI: 10.3987/COM-93-6366.
- Dahl, A. C., Fjeldberg, M., & Madsen, J. O. (1999). Baker's yeast: improving the D-stereoselectivity in reduction of 3-oxo esters. *Tetrahedron: Asymmetry*, 10, 551–559. DOI: 10.1016/s0957-4166(99)00025-7.
- Forni, A., Moretti, I., Prati, F., & Torre, G. (1994). Stereochemical control in yeast reduction of fluorinated β -diketones. *Tetrahedron*, 50, 11995–12000. DOI: 10.1016/s0040-4020(01)89310-8.
- Fujisawa, T., Sugimoto, T., & Shimizu, M. (1994). Highly stereocontrolled access to trifluoromethylbenzylic alcohols possessing *p*-substituents by the bakers' yeast reduction. *Tetrahedron: Asymmetry*, 5, 1095–1098. DOI: 10.1016/0957-4166(94)80060-x.
- Hayakawa, R., Nozawa, K., Shimizu, M., & Fujisawa, T. (1998). Control of enantioselectivity in the bakers' yeast reduction of β -keto ester derivatives in the presence of a sulfur compound. *Tetrahedron Letters*, 39, 67–70. DOI: 10.1016/s0040-4039(97)10490-7.
- Heiss, C., Laivenieks, M., Zeikus, J. G., & Phillips, R. S. (2001). The stereospecificity of secondary alcohol dehydrogenase from *Thermoanaerobacter ethanolicus* is partially determined by active site water. *Journal of the American Chemical Society*, 123, 345–346. DOI: 10.1021/ja005575a.
- Hilgenkamp, R., & Zercher, C. K. (2001). Tandem chain extension-homoenolate formation: the formation of α -methylated- γ -keto esters. *Organic Letters*, 3, 3037–3040. DOI: 10.1021/ol016485t.
- Huang, D., Yan, M., Zhao, W. J., & Shen, Q. (2005). Efficient synthesis of γ -keto esters from enamines and EDA. *Synthetic Communications*, 35, 745–750. DOI: 10.1081/scc-200050387.
- Hudlicky, M. (1990). *Oxidation in organic chemistry*. Washington, DC, USA: American Chemical Society.
- Iqbal, M., Baloch, I. B., & Baloch, M. K. (2012). Synthesis and structural characterization of novel monoesters of succinic anhydride with aryl alcohols. *Chemistry Journal*, 2, 12–19.
- Itoh, N., Matsuda, M., Mabuchi, M., Dairi, T., & Wang, J. (2002). Chiral alcohol production by NADH-dependent phenylacetaldehyde reductase coupled with *in situ* regeneration of NADH. *European Journal of Biochemistry*, 269, 2394–2402. DOI: 10.1046/j.1432-1033.2002.02899.x.
- Izquierdo, J., Rodriguez, S., & Gonzalez, F. V. (2011). Regioselective ring opening and isomerization reactions of 3,4-epoxyesters catalyzed by boron trifluoride. *Organic Letters*, 13, 3856–3859. DOI: 10.1021/ol201378w.

- Kataoka, M., Yamamoto, K., Kawabata, H., Wada, M., Kita, K., Yanase, H., & Shimizu, S. (1999). Stereoselective reduction of ethyl 4-chloro-3-oxobutanoate by *Escherichia coli* transformant cells coexpressing the aldehyde reductase and glucose dehydrogenase genes. *Applied Microbiology and Biotechnology*, 51, 486–490. DOI: 10.1007/s002530051421.
- Kashima, C., Shirahata, Y., & Tsukamoto, Y. (2001). Preparation of β -substituted γ -keto esters by the Grignard reaction on *N*-acylpyrazoles. *Heterocycles*, 54, 309–317. DOI: 10.3987/com-00-s(I)37.
- Kizaki, N., Yasohara, Y., Hasegawa, J., Wada, M., Kataoka, M., & Shimizu, S. (2001). Synthesis of optically pure ethyl (*S*)-4-chloro-3-hydroxybutanoate by *Escherichia coli* transformant cells coexpressing the carbonyl reductase and glucose dehydrogenase genes. *Applied Microbiology and Biotechnology*, 55, 590–595. DOI: 10.1007/s002530100599.
- Larock, R. C. (1999). *Comprehensive organic transformations* (2nd ed.). New York, NY, USA: Wiley–VCH.
- Nakamura, K., Yamanaka, R., Matsuda, T., & Harada, T. (2003). Recent developments in asymmetric reduction of ketones with biocatalysts. *Tetrahedron: Asymmetry*, 14, 2659–2681. DOI: 10.1016/s0957-4166(03)00526-3.
- Poliakoff, M., Fitzpatrick, J. M., Farren, T. R., & Anastas, P. T. (2002). Green chemistry: science and politics of change. *Science*, 297, 807–810. DOI: 10.1126/science.297.5582.807.
- Roberts, J. D., & Caserio, M. C. (1964). *Basic principles of organic chemistry*. Menlo Park, CA, USA: W. A. Benjamin Inc.
- Ronsheim, M. D., Hilgenkamp, R. K., & Zercher, C. K. (2002). Formation of γ -keto esters from β -keto esters: Methyl 5,5-dimethyl-4-oxo-hexanoate. *Organic Syntheses*, 79, 146.
- Von Rudloff, E. (1958). Synthesis of some hexanediols. *Canadian Journal of Chemistry*, 36, 486–491. DOI: 10.1139/v58-069.
- Shafiee, A., Motamedi, H., & King, A. (1998). Purification, characterization and immobilization of an NADPH-dependent enzyme involved in the chiral specific reduction of the keto ester M, an intermediate in the synthesis of an anti-asthma drug, Montelukast, from *Microbacterium campomadoensis* (MB5614). *Applied Microbiology and Biotechnology*, 49, 709–717. DOI: 10.1007/s002530051236.
- Stájer, G., Csénde, F., Bernáth, G., Sohár, P., & Szúnyog, J. (1994). Preparation and steric structure of 3(2H)-pyridazinones and 1,2-oxazin-6-ones fused with three- to six-membered saturated carbocycles or norbornane skeleton. *Monatshefte für Chemie/Chemical Monthly*, 125, 933–944. DOI: 10.1007/bf00812708.
- Taylor, H. T. (1958). Preparation of unsaturated keto-acids from the interaction of ethylene and acid anhydrides. *Journal of the Chemical Society (Resumed)*, 1958, 3922–3924. DOI: 10.1039/jr9580003922.
- Tojo, G., & Fernández, M. (2006). *Oxidation of alcohols to aldehydes and ketones*. New York, NY, USA: Springer.
- Wang, W., Xu, B., & Hammond, G. B. (2009). Efficient synthesis of γ -keto esters through neighboring carbonyl group-assisted regioselective hydration of 3-alkynoates. *Journal of Organic Chemistry*, 74, 1640–1643. DOI: 10.1021/jo802450n.
- Wehrli, P. A., & Chu, V. (1973). Novel synthesis of γ -keto esters. *Journal of Organic Chemistry*, 38, 3436–3436. DOI: 10.1021/jo00959a053.
- Wehrli, P. A., & Chu, V. (1978). γ -Ketoesters from aldehydes via diethyl acylsuccinates: Ethyl 4-oxohexanoate. *Organic Syntheses*, 58, 79.
- Williams, D.B.G., Blann, K., & Holzapfel, C. W. (2001). Aryl γ -ketoesters as precursors for γ -butyrolactones in samarium(II) iodide-mediated reactions. *Synthetic Communications*, 31, 203–209. DOI: 10.1081/scc-100000200.
- Williams, D. B. G., Blann, K., Caddy, J., & Holzapfel, C. W. (2002). Aryl γ -ketoesters as precursors for γ -butyrolactone dimers in samarium(II) iodide-mediated reactions. *Synthetic Communications*, 32, 3755–3762. DOI: 10.1081/scc-120015393.
- Yamamoto, H., Kimoto, N., Matsuyama, A., & Kobayashi, Y. (2002a). Purification and properties of a carbonyl reductase useful for production of ethyl (*S*)-4-chloro-3-hydroxybutanoate from *Kluyveromyces lactis*. *Bioscience, Biotechnology, and Biochemistry*, 66, 1775–1778. DOI: 10.1271/bbb.66.1775.
- Yamamoto, H., Matsuyama, A., & Kobayashi, Y. (2002b). Synthesis of ethyl (*R*)-4-chloro-3-hydroxybutanoate with recombinant *Escherichia coli* cells expressing (*S*)-specific secondary alcohol dehydrogenase. *Bioscience, Biotechnology, and Biochemistry*, 66, 481–483. DOI: 10.1271/bbb.66.481.