Paper

TiCl₄-Mediated Synthesis of 3,4-Hetero-Disubstituted Isocoumarins by Means of Isocyanide Insertion Reactions

Α

Sudipta Ponra^{a,c} Aude Nyadanu^{b,c} Sylvie Maurin^c Laurent El Kaïm^{*b} Laurence Grimaud^{*c} Maxime R. Vitale^{*a}

^a Chimie ParisTech, PSL Research University, CNRS, Institut de Recherche de Chimie Paris (IRCP), 75005 Paris, France

S. Ponra et al.

maxime.vitale@chimie-paristech.fr ^b LSO (UMR 7652), École Polytechnique - ENSTA ParisTech, Université

Paris-Saclay, 91120, Palaiseau, France

laurent.elkaim@ensta-paristech.fr

^c PASTEUR, Département de Chimie, École Normale Supérieure, PSL Research University, Sorbonne Universités, UPMC Univ. Paris 06, CNRS, 75005 Paris, France laurence.grimaud@ens.fr

Received: 11.10.2017 Accepted after revision: 07.11.2017 Published online: 13.12.2017 DOI: 10.1055/s-0036-1591733; Art ID: ss-2017-t0657-op

Abstract The isocyanide insertion into sulfanyl-phthalides under Lewis acid conditions is reported with full details. This sequential three-component reaction affords a new straightforward and highly convenient method for the preparation of 3-amino-4-sulfanyl isocoumarins, the potential of which is still unexplored.

Key words isocyanides, isocoumarins, thio-Passerini reaction, thiocarbonyl surrogates

Isocoumarins constitute a family of synthetic or naturally occurring molecules that demonstrate an exceptionally wide array of biological activities and, for more than half a century, have been the cornerstone of many organic and pharmaceutical chemistry research programs.^{1,2} Among this class, several 3,4-disubstituted isocoumarins including oospolactone, polygolide and cephalosol have exhibited interesting antifungal, anti-inflammatory and antimicrobial activities.³ Furthermore, some 4-chloro-substituted analogues are very potent protease inhibitors with potential applications in Alzheimer's disease, while various 3-amino-4-aryl derivatives have demonstrated encouraging anticholesteremic and anti-arteriosclerotic activities (Figure 1).⁴

As a consequence, the development of new synthetic routes to 3,4-disubstituted isocoumarin scaffolds has become a very active field of research.²⁻⁷ Whereas the vast majority of the reported methods rely on the use of internal alkynes by means of transition-metal-catalyzed or electrophilic annulation reactions, ^{1d,5,6} the regioselective incorporation of heteroatoms on position 3 and/or 4 still remains a significant challenge.





Figure 1 Representative biologically active 3,4-disubstituted isocoumarins

In this area, isocyanide-based insertion reactions have emerged as particularly promising strategies.^{8,9} In 2010, Marcaccini et al. reported that, starting from 2-formylbenzoic acid, an isonitrile and an amine, various 3,4-diaminoisocoumarins could be obtained (Scheme 1, eq. 1).^{8a} Later, by replacing the amine partner with malononitrile, Soleimani et al. were able to extend this work to the synthesis of 3,4-pyrazolo-isocoumarins (Scheme 1, eq. 2).^{8b}

In continuation of our interest in isocyanide chemistry,¹⁰ we recently reported the preparation of various thiophthalides by a formal thio-Passerini reaction.¹¹ During this work, we incidentally discovered that, in specific cases, valuable 3-amino-4-sulfanyl isocoumarins could be obtained (Scheme 1, eq. 3). We report herein with full details

this regioselective method for access to 3,4-hetero-disubstituted isocoumarins, starting from sulfanyl-phthalides or directly from 2-formylbenzoic acids.



We previously reported that, upon reaction with isonitriles and TiCl₄ at room temperature, 3-(*tert*-butylsulfanyl)phthalides allowed the formation of a variety of thiophthalides in respectable yields (Scheme 2, eq. 1).¹¹ During the related optimization studies, we alternatively witnessed an interesting reactivity dichotomy when 3-phenylsulfanylphthalide (**1a**) was employed. Indeed, with *tert*-butyl isocyanide (**2a**), this particular substrate in which solely the *S*substitution changes led cleanly to the 3-amino-4-sulfanyl isocoumarin **3aa** in a good 76% yield (Scheme 2, eq. 2). While, to the best of our knowledge, the preparation of such heterocyclic scaffolds has been scarcely studied,^{7g,h} the potential of an original synthetic access to these 3,4-heterodisubstituted isocoumarins encouraged us to study the scope and limitations of this novel transformation.



Scheme 2 Divergent synthesis of thiophthalides and 3,4-disubstituted isocoumarin 3aa

-	0	

Downloaded by: University of Michigan. Copyrighted material.

For this purpose, in dichloromethane at room temperature, 3-phenylsulfanyl-phthalide (**1a**) was initially submitted to a variety of isocyanides **2b-h** (1.2 equiv) and TiCl₄ (1.0 equiv) (Scheme 3). To our delight, the desired isocyanide insertion reaction took place regardless of the aliphatic or aromatic nature of the isocyanide engaged in the process. Accordingly, secondary alkyl as well as primary alkyl isocyanides **2b-e** afforded the desired 3,4-disubstituted isocoumarins **3ab-ae** with up to 94% yield, while aromatic isocyanides **2f** and **2g** afforded **3af** and **3ag** in 86% and 79% yield, respectively. As a probable consequence of competitive polymerization processes, the use of electron-rich 2-methoxyphenyl isocyanide **2h** only afforded a 14% yield of **3ah**.



Scheme 3 Scope of various isocyanides in the TiCl_4-promoted isocyanide insertion reaction into sulfanyl-phthalide 1a

We next studied the impact of the sulfur atom substitution on the overall efficiency of the isocyanide insertion process. A large panel of diversely functionalized sulfanylphthalides was prepared according to a straightforward protocol in which 2-formylbenzoic acids **4a–c** and various thiols **5a–r** were refluxed in toluene in the presence of Mg-SO₄ (Scheme 4). Pleasingly, this method proved to be quite robust and allowed the required sulfanyl-phthalides **1b–r**, **1a'** and **11"** to be obtained in good to excellent yields (42– 98%), apart from in the case of 1-naphthyl-substituted compound **1k** (18% yield).

Submitting S-aryl substituted sulfanyl-phthalides to the isocyanide-insertion conditions afforded the desired isocoumarin derivatives in good to excellent yields (Scheme 5). The substitution on the aromatic core poorly influences the efficiency of the reaction as electron-donating groups such as methoxy or alkyl give yields similar to those obtained

Syn thesis

S. Ponra et al.

۸

С



Scheme 4 Preparation of a wide variety of sulfanyl-phthalides 1

with electron-withdrawing groups such as halogen atoms. The *ortho*, *meta* or *para* position of the substituent seemed to have no effect on the reaction as isocoumarins **3fb–hb** were obtained in almost identical yields.

The scope was next examined for *S*-alkyl-substituted phthalides. Depending on the nature of the starting thiol, different results were observed. Primary thiols with long aliphatic chains gave the expected isocoumarins in good yields (Table 1, entries 1–3). However, secondary or benzylic thiols afforded the desired products along with substantial amounts (16–32%) of the corresponding thiophthalides **6** (Table 1, entries 5–7). We surmised that such outcomes could be rationalized by the ability of the *S*-substituent to stabilize the formed carbocation to allow the final 1,5-acyl transfer (Scheme 6). This hypothesis was further confirmed by the formation of **6b** as the major product in 70% isolated yield when using *tert*-butyl thiol (Table 1, entry 8).

These experimental results led us to propose a plausible mechanistic path for the isocoumarin synthesis (Scheme 6). The complexation of the Lewis acid with the starting sulfanyl-phthalide **1** induces the formation of the thiocarbenium intermediate **A**, which is electrophilic enough to promote nucleophilic addition of the isocyanide. The resulting nitrilium species **B** is then trapped intramolecularly to give **C**. After tautomerization, the latter evolves toward the pyrylium



Scheme 5 Evaluation of the scope of the reaction starting from various S-aryl-substituted sulfanyl-phthalides and cyclohexylisocyanide (CyNC)

Table 1 Evaluation of S-Alkyl-Substituted Sulfanyl-phthalides



Entry	Phthalide 1 (R ¹)	Isocyanide 2 (R²)	Isocoumarin 3 (yield)ª	Thiophthalide 6 (yield)ª
1	1m ("Dodecyl)	2b (Cy)	3mb (93%)	_b
2	1m ("Dodecyl)	2a (^t Bu)	3ma (50%) ^c	_b
3	1n ("Hex)	2b (Cy)	3nb (93%)	_b
4	1r (CH ₂ CO ₂ Me)	2b (Cy)	3rb (24%)	_b
5	1o (Cy)	2b (Cy)	3ob (68%)	6b (32%)
6	1o (Cy)	2d (Bn)	3od (55%)	6d (29%)
7	1q (Bn)	2b (Cy)	3qb (70%)	6b (16%)
8	1p (^{<i>t</i>} Bu)	2b (Cy)	3pb (16%)	6b (70%)

^a Yield of isolated product.

^b Not detected by ¹H NMR spectroscopy of the crude reaction mixture.

 $^{\rm c}$ Partial deprotection of isocoumarin **3ma** was additionally observed (**3ma'**, R² = H, 12%).

D

Syn<mark>thesis</mark>

S. Ponra et al.

D, which is most probably responsible for the intense darkblue color of the reaction mixture. Indeed, when adding water at the end of the reaction, the color vanishes and the expected isocoumarin **3** is formed. This hypothesis is rather consistent with the need for a stoichiometric amount of Lewis acid as the pyrylium species **D** is still linked to one molecule of TiCl₄. In the case of more labile R¹ substituents, intermediate **C** could evolve through the loss of this group and competitive 1,5-acyl transfer from *O* to *S*, which would give the corresponding thiophthalide **6**.



Scheme 6 A plausible mechanistic pathway

We envisioned developing a one-pot procedure for the formation of 3-amino-4-sulfanyl isocoumarins 3 starting directly from 2-formylbenzoic acids 4 and thiols 5. Unfortunately, all our attempts at a multicomponent version of the reaction failed. The sulfanyl-phthalides 1 were thus preformed at room temperature by mixing the 2-formylbenzoic acid **4** with the thiol **5** in the presence of catalytic amounts of the Lewis acid and 4 Å molecular sieves. After one hour, a slight excess of isocyanide 2 was added to the reaction mixture along with one equivalent of TiCl₄. Under these optimized conditions, the desired 3.4-disubstituted isocoumarins 3 were isolated in good yields in all the cases examined (Scheme 7). Albeit sequential, this method allows a convenient and efficient synthesis of 4-amino-3-thiosubstituted isocoumarins 3 avoiding the tedious isolation of sulfanyl-phthalides 1.

To conclude, we have developed an original access to a large variety of interesting 3,4-hetero-disubstituted isocoumarin derivatives starting from easily accessible 2-formylbenzoic acids, thiols and isocyanides. This straightforward and operationally simple process enables the synthesis of isocoumarins with different substitution patterns. In addition, this method constitutes a new tool to explore the un-



Scheme 7 One-pot synthesis of 3,4-disubstituted isocoumarins starting from 2-formylbenzoic acids

tapped potential of 3-amino-4-sulfanyl isocoumarins which, in addition to potentially possessing biological activities, display notable fluorescence properties (see the Supporting Information for further details).

All reactions were performed under an argon atmosphere and the reagents were ordered from commercial suppliers and used without further purification. Solvents were distilled or purified on activated alumina columns before use. Isocyanides **2f-h** and substituted 2formylbenzoic acids **4b** and **4c** were prepared according to literature procedures.^{8d,12} The preparation and characterization of compounds **1a, 1m, 1o, 1p, 3aa, 3ab, 3ad, 3af, 3ma, 3mb** and **3ob** have previously been reported.¹¹ Column chromatography was accomplished using Acros Silica gel 60 (40–63 µm). Melting points were obtained using a Kofler bench apparatus. ¹H, ¹³C and ¹⁹F NMR spectra were obtained using Bruker AVANCE 300 or 400 spectrometers with CDCl₃ as the solvent. High-resolution mass spectrometry was performed using a Jed JMS-GCmate II instrument.

General Procedure A

To a stirred solution of 2-formylbenzoic acid **4a**–**c** (1.1 mmol), *p*-toluenesulfonic acid (8 mg) and MgSO₄ (2.0 g) in anhyd toluene (5 mL) was added the appropriate thiol **5** (1.0 mmol). The mixture was heated at reflux for 3 h before being cooled to room temperature and filtered through a small pad of silica. After washing with toluene (3 × 20 mL), the solvent was removed under reduced pressure to afford essentially pure racemic 3-sulfanyl-phthalide **1** or, if needed, the residue was submitted to purification by silica gel flash column chromatography.

General Procedure B

To a screw-cap vial under an argon atmosphere were sequentially added thiophthalide **1** (100 mg, 1 equiv), isocyanide **2** (1.2 equiv), CH_2Cl_2 (5 mL) and $TiCl_4$ (1 M in CH_2Cl_2 , 1 equiv). The resulting mixture was stirred at room temperature for 1 h before dropwise addition of saturated aqueous NaHCO₃ (10 mL). After separation of the organic phase, the aqueous layer was extracted with CH_2Cl_2 (4 × 15 mL) and the combined organic extracts washed with H_2O (40 mL) and brine (40 mL) before being dried over MgSO₄. After filtration and evaporation of the solvents under reduced pressure, the crude material was purified by flash column chromatography on silica gel to afford the desired isocoumarin derivative **3**.

General Procedure C

To a screw-cap vial under an argon atmosphere were sequentially added the 2-carboxybenzaldehyde **4** (1.10 mmol, 1.1 equiv), the appropriate thiol **5** (1.00 mmol, 1.0 equiv), CH₂Cl₂ (5 mL), activated 4 Å MS (2.00 g) and TiCl₄ (200 μ L, 1 M in CH₂Cl₂, 0.2 equiv). After stirring the resulting mixture at room temperature for 1 h, isocyanide **2** (1.50 mmol, 1.5 equiv) and TiCl₄ (1.00 mL, 1 M in CH₂Cl₂, 1 equiv) were sequentially added. After 1 h, the reaction mixture was poured into saturated aqueous NaHCO₃ (20 mL) and the organic layer was separated. The aqueous phase was then extracted with CH₂Cl₂ (4 × 20 mL) and the combined organic extracts washed with H₂O (50 mL) and brine (50 mL) before being dried over MgSO₄. After filtration and evaporation of the solvents under reduced pressure, the crude material was purified by flash column chromatography on silica gel to afford the desired isocoumarin derivative **3**.

(R/S)-3-(p-Tolylthio)isobenzofuran-1(3H)-one (1b)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 4methylbenzenethiol (**5b**) (124 mg, 1 mmol) and following the general procedure A, **1b** was obtained as a white solid (251 mg, 98% yield); mp 126–128 °C.

¹H NMR (400 MHz, CDCl₃): δ = 7.81–7.62 (m, 3 H), 7.49 (t, J = 7.4 Hz, 1 H), 7.39–7.33 (m, 2 H), 7.05 (d, J = 7.9 Hz, 2 H), 6.66 (s, 1 H), 2.29 (s, 3 H).

 ^{13}C NMR (101 MHz, CDCl_3): δ = 169.3, 146.4, 139.4, 134.3 (2 C), 130.0, 129.9, 126.4, 125.5, 123.5, 86.8, 21.3.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₅H₁₂O₂S: 256.0558; found: 256.0556.

(R/S)-3-[(4-Methoxyphenyl)thio]isobenzofuran-1(3H)-one (1c)

Starting from phthalaldehydic acid (**4a**) (400 mg, 2.7 mmol, 1.1 equiv) and 4-methoxythiophenol (**5c**) (300 μ L, 2.4 mmol, 1 equiv) and following the general procedure A, **1c** was obtained as a white solid (498 mg, 76% yield); mp 112–114 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.73 (d, J = 7.6 Hz, 1 H), 7.71–7.66 (m, 1 H), 7.63 (dd, J = 7.5, 0.7 Hz, 1 H), 7.49–7.44 (m, 1 H), 7.39–7.32 (m, 2 H), 6.75–6.71 (m, 2 H), 6.60 (s, 1 H), 3.74 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 169.1, 160.6, 146.3, 136.7, 134.1, 129.8, 126.2, 125.3, 123.3, 119.5, 114.4, 86.7, 55.2.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₅H₁₂O₃S: 272.0507; found: 272.0508.

(R/S)-3-(4-tert-Butylphenylthio)isobenzofuran-1(3H)-one (1d)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 4-*tert*-butylbenzenethiol (**5d**) (168 μ L, 1 mmol) and following the general procedure A, **1d** was obtained as a white solid (284 mg, 95% yield); mp 102–104 °C.

¹H NMR (400 MHz, CDCl₃): δ = 7.71 (d, *J* = 7.7 Hz, 1 H), 7.65–7.40 (m, 2 H), 7.46–7.34 (m, 3 H), 7.24–7.19 (m, 2 H), 6.61 (s, 1 H), 1.19 (s, 9 H).

 ^{13}C NMR (101 MHz, CDCl₃): δ = 169.3, 152.5, 146.3, 134.3, 133.7, 130.0, 127.1, 126.4, 126.2, 125.5, 123.5, 87.0, 34.7, 31.3.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₈H₁₈O₂S: 298.1028; found: 298.1033.

(R/S)-3-[(4-Fluorophenyl)thio]isobenzofuran-1(3H)-one (1e)

Starting from phthalaldehydic acid (**4a**) (500 mg, 3.3 mmol, 1.1 equiv) and 4-fluorothiophenol (**5e**) (320 μ L, 3 mmol, 1 equiv) and following the general procedure A, **1e** was obtained as a white solid (443 mg, 57% yield); mp 112–114 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.77 (d, *J* = 7.6 Hz, 1 H), 7.71 (td, *J* = 7.5, 1.1 Hz, 1 H), 7.64 (dd, *J* = 7.7, 1.0 Hz, 1 H), 7.50 (t, *J* = 7.4 Hz, 1 H), 7.48–7.42 (m, 2 H), 6.96–6.91 (m, 2 H), 6.65 (s, 1 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 169.0, 163.4 (d, $J_{\text{C-F}}$ =250.2 Hz), 146.0, 136.7 (d, $J_{\text{C-F}}$ = 8.7 Hz), 134.3, 130.0, 126.2, 125.5, 124.6 (d, $J_{\text{C-F}}$ = 3.4 Hz), 123.3, 116.2 (d, $J_{\text{C-F}}$ = 22.0 Hz), 86.3.

¹⁹F NMR (300 MHz, CDCl₃): δ = -111.2.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₄H₉FO₂S: 260.0307; found: 260.0315.

(R/S)-3-(4-Bromophenylthio)isobenzofuran-1(3H)-one (1f)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 4bromobenzenethiol (**5f**) (189 mg, 1 mmol) and following the general procedure A, **1f** was obtained as a white solid (313 mg, 98% yield); mp 142–144 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.80 (d, J = 7.6 Hz, 1 H), 7.75–7.67 (m, 1 H), 7.64 (dd, J = 7.6, 0.8 Hz, 1 H), 7.52 (t, J = 7.4 Hz, 1 H), 7.42–7.32 (m, 4 H), 6.68 (s, 1 H).

¹³C NMR (75 MHz, CDCl₃): δ = 169.0, 146.0, 135.4, 134.5, 132.4, 130.3, 129.4, 126.4, 125.7, 123.8, 123.4, 86.1.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₁₄H₉BrO₂S: 319.9507; found: 319.9507.

(R/S)-3-(3-Bromophenylthio)isobenzofuran-1(3H)-one (1g)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 3-bromobenzenethiol (**5g**) (118 μ L, 1 mmol) and following the general procedure A, **1g** was obtained as a white solid (308 mg, 96% yield); mp 114–116 °C.

¹H NMR (400 MHz, CDCl₃): δ = 7.81 (d, J = 7.7 Hz, 1 H), 7.72 (td, J = 7.6, 1.0 Hz, 1 H), 7.67–7.60 (m, 2 H), 7.53 (t, J = 7.5 Hz, 1 H), 7.49–7.36 (m, 2 H), 7.14 (t, J = 7.9 Hz, 1 H), 6.72 (s, 1 H).

 ^{13}C NMR (101 MHz, CDCl₃): δ = 168.9, 145.7, 135.6, 134.5, 132.9, 131.9, 131.8, 130.5, 130.4, 126.2, 125.6, 123.5, 122.7, 86.1.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₄H₉BrO₂S: 319.9507; found: 319.9508.

(*R/S*)-3-(2-Bromophenylthio)isobenzofuran-1(3*H*)-one (1h)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 2-bromobenzenethiol (**5h**) (120 μ L, 1 mmol) and following the general procedure A, **1h** was obtained after crystallization from hexane/toluene (5:1) as a white solid (134 mg, 42% yield); mp 132–134 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.90 (d, *J* = 7.6 Hz, 1 H), 7.83–7.69 (m, 3 H), 7.64–7.53 (m, 2 H), 7.33 (td, *J* = 7.7, 1.4 Hz, 1 H), 7.25–7.15 (m, 1 H), 6.83 (s, 1 H).

¹³C NMR (75 MHz, CDCl₃): δ = 169.1, 145.6, 134.6, 133.7, 133.52, 133.46, 130.5, 129.8, 128.4, 126.4, 126.3, 125.8, 123.8, 85.7.

HRMS (ESI): m/z [M⁺]⁺ (⁷⁹Br) and [M⁺]⁺ (⁸¹Br) calcd for C₁₄H₉BrO₂S: 319.9507 and 321.9507; found: 319.9509 and 321.9599.

(R/S)-3-(3,4-Dimethylphenylthio)isobenzofuran-1(3H)-one (1i)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and 3,4dimethylbenzenethiol (**5i**) (135 μ L, 1.0 mmol) and following the general procedure A, **1i** was obtained as a white solid (234 mg, 87% yield); mp 130–132 °C.

¹H NMR (400 MHz, CDCl₃): δ = 7.71–7.54 (m, 3 H), 7.41 (t, J = 7.4 Hz, 1 H), 7.21–7.11 (m, 2 H), 6.92 (d, J = 7.8 Hz, 1 H), 6.57 (s, 1 H), 2.11 (t, J = 4.5 Hz, 6 H).

¹³C NMR (101 MHz, CDCl₃): δ = 169.3, 146.3, 137.9, 137.6, 135.1, 134.3, 131.5, 130.3, 130.0, 126.9, 126.3, 125.4, 123.5, 87.0, 19.6 (2 C). HRMS (ESI): m/z [M[·]]⁺ calcd for C₁₆H₁₄O₂S: 270.0715; found: 270.0719.

(R/S)-3-[(3,4-Dimethoxyphenyl)thio]isobenzofuran-1(3H)-one(1j)

Starting from phthalaldehydic acid (**4a**) (200 mg, 1.3 mmol) and 3,4dimethoxythiophenol (**5j**) (173 μ L, 1.2 mmol) and following the general procedure A, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (7:3) as eluent gave **1j** as a white solid (351 mg, 97% yield); mp 119– 121 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.73 (d, *J* = 7.7 Hz, 1 H), 7.72–7.62 (m, 2 H), 7.50–7.44 (m, 1 H), 7.03 (dd, *J* = 8.3, 2.1 Hz, 1 H), 6.89 (d, *J* = 2.1 Hz, 1 H), 6.70 (d, *J* = 8.3 Hz, 1 H), 6.65 (s, 1 H), 3.82 (s, 3 H), 3.79 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 169.1, 150.2, 148.7, 146.4, 134.1, 129.8, 128.4, 126.3, 125.4, 123.3, 119.6, 117.7, 111.1, 86.6, 56.0, 55.8.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₆H₁₄O₄S: 302.0613; found: 302.0618.

(R/S)-3-(Naphthalen-1-ylthio)isobenzofuran-1(3H)-one (1k)

Starting from phthalaldehydic acid (**4a**) (500 mg, 3.3 mmol) and naphthalene-1-thiol (**5k**) (420 μ L, 3.0 mmol) and following the general procedure A, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (9:1) as eluent gave **1k** as a white solid (161 mg, 18% yield); mp 103–105 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.57 (d, *J* = 8.4 Hz, 1 H), 7.92 (d, *J* = 7.2 Hz, 1 H), 7.86–7.77 (m, 3 H), 7.74–7.71 (m, 1 H), 7.69–7.59 (m, 2 H), 7.57–7.38 (m, 3 H), 6.74 (s, 1 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 169.0, 146.0, 134.1, 134.0, 133.9, 130.1, 130.0, 128.5, 128.0, 127.0, 126.4, 126.1, 125.5, 125.4, 125.3, 123.4, 86.8.

Paper

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₈H₁₂O₂S: 292.0558; found: 292.0560.

(R/S)-3-(Naphthalen-2-ylthio)isobenzofuran-1(3H)-one (11)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and naphthalene-2-thiol (**5l**) (160.2 mg, 1.0 mmol) and following the general procedure A, **1l** was obtained as a white solid (208 mg, 71% yield); mp 120–122 °C.

 ^1H NMR (400 MHz, CDCl_3): δ = 8.04 (d, J = 1.3 Hz, 1 H), 7.80–7.66 (m, 6 H), 7.57–7.44 (m, 4 H), 6.77 (s, 1 H).

 ^{13}C NMR (101 MHz, CDCl₃): δ = 169.2, 146.0, 134.4, 133.4, 133.0, 132.9, 130.1, 130.0, 128.8, 128.0, 127.8, 127.7, 126.9, 126.8, 126.2, 125.5, 123.5, 86.7.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₈H₁₂O₂S: 292.0558; found: 292.0568.

(R/S)-3-(Hexylthio)isobenzofuran-1(3H)-one (1n)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and hexane-1-thiol (**5n**) (141 μ L, 1.0 mmol) and following the general procedure A, **1n** was obtained as a colorless oil (233 mg, 93% yield).

¹H NMR (300 MHz, CDCl₃): δ = 7.82 (d, *J* = 7.5 Hz, 1 H), 7.65 (dd, *J* = 10.9, 4.1 Hz, 1 H), 7.56–7.43 (m, 2 H), 6.53 (s, 1 H), 2.69–2.44 (m, 2 H), 1.64–1.47 (m, 2 H), 1.40–1.10 (m, 6 H), 0.81 (dd, *J* = 6.8, 5.4 Hz, 3 H). ¹³C NMR (75 MHz, CDCl₃): δ = 169.3, 146.8, 134.3, 129.9, 126.2, 125.4, 123.1, 85.0, 31.2, 30.7, 29.5, 28.3, 22.4, 13.9.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₄H₁₈O₂S: 250.1028; found: 250.1023.

(R/S)-3-(Benzylthio)isobenzofuran-1(3H)-one (1q)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and phenylmethanethiol (**5q**) (118 μ L, 1.0 mmol) and following the general procedure A, **1q** was obtained as a white solid (252 mg, 98% yield); mp 83–85 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.80 (d, J = 7.6 Hz, 1 H), 7.57 (td, J = 7.5, 1.1 Hz, 1 H), 7.46 (dd, J = 10.9, 4.0 Hz, 1 H), 7.38 (dd, J = 7.6, 0.8 Hz, 1 H), 7.29–7.14 (m, 5 H), 6.32 (s, 1 H), 3.93 (d, J = 13.2 Hz, 1 H), 3.74 (d, J = 13.2 Hz, 1 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 169.3, 146.5, 136.5, 134.3, 130.0, 129.2, 128.7, 127.5, 126.3, 125.6, 123.0, 83.3, 35.0.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₅H₁₂O₂S: 256.0558; found: 256.0570.

(*R/S*)-Methyl 2-(3-Oxo-1,3-dihydroisobenzofuran-1-ylthio)ace-tate (1r)

Starting from phthalaldehydic acid (**4a**) (165 mg, 1.1 mmol) and methyl 2-mercaptoacetate (**5r**) (91 μ L, 1.0 mmol) and following the general procedure A, **1q** was obtained as colorless oil (186 mg, 78% yield).

¹H NMR (300 MHz, $CDCI_3$): δ = 7.85 (d, *J* = 7.0 Hz, 1 H), 7.69 (t, *J* = 7.5 Hz, 1 H), 7.54 (t, *J* = 7.2 Hz, 2 H), 6.71 (s, 1 H), 3.67 (d, *J* = 1.9 Hz, 3 H), 3.46 (dd, *J* = 15.4, 1.9 Hz, 1 H), 3.25 (d, *J* = 15.3 Hz, 1 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 169.8, 168.9, 145.8, 134.5, 130.3, 126.3, 125.7, 123.2, 83.5, 52.7, 31.6.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₁H₁₀O₄S: 238.0300; found: 238.0310.

(R/S)-6-Methyl-3-(phenylthio)isobenzofuran-1(3H)-one (1a')

Starting from 2-formyl-5-methylbenzoic acid (**4b**) (271 mg, 1.65 mmol) and thiophenol (**5a**) (154 μ L, 1.5 mmol) and following the general procedure A, **1a'** was obtained as a white solid (321 mg, 84% yield); mp 128–130 °C.

 ^1H NMR (300 MHz, CDCl_3): δ = 7.50–7.37 (m, 5 H), 7.21–7.13 (m, 3 H), 6.58 (s, 1 H), 2.33 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 169.3, 143.5, 140.6, 135.5, 133.5, 130.8, 129.1, 128.8, 126.5, 125.4, 123.1, 86.5, 21.4.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₅H₁₂O₂S: 256.0558; found: 256.0563.

(*R/S*)-6-Methoxy-3-(naphthalen-2-ylthio)isobenzofuran-1(3*H*)one (11")

Starting from 2-formyl-5-methoxybenzoic acid (**4c**) (297 mg, 1.65 mmol) and naphthalene-2-thiol (**5l**) (240 mg, 1.5 mmol) and following the general procedure A, **11**" was obtained as a white solid (466 mg, 96% yield); mp 130–132 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.06 (d, J = 1.2 Hz, 1 H), 7.84–7.70 (m, 3 H), 7.60–7.45 (m, 4 H), 7.27–7.18 (m, 2 H), 6.74 (s, 1 H), 3.80 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 169.2, 161.4, 138.2, 133.5, 132.9, 132.7, 129.9, 128.7, 128.5, 127.8, 127.7, 126.9, 126.7, 124.3, 123.1, 107.4, 86.8, 55.8.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₉H₁₄O₃S: 322.0664; found: 322.0662.

3-(Pentylamino)-4-(phenylthio)-1H-isochromen-1-one (3ac)

Starting from 3-(phenylthio)isobenzofuran-1(3*H*)-one (**1a**) (100 mg, 0.41 mmol), 1-isocyanopentane (**2c**) (63 μ L, 0.50 mmol), TiCl₄ (410 μ L, 1 M in CH₂Cl₂, 0.41 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3ac** as a yellow solid (121 mg, 87% yield); mp 102–104 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.08–7.96 (m, 1 H), 7.50 (dd, *J* = 8.2, 0.5 Hz, 1 H), 7.45–7.35 (m, 1 H), 7.16–6.96 (m, 6 H), 5.73 (t, *J* = 5.9 Hz, 1 H), 3.31 (m, 2 H), 1.52–1.40 (m, 2 H), 1.25–1.11 (m, 4 H), 0.76 (t, *J* = 6.9 Hz, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.5, 159.7, 142.3, 136.1, 135.6, 130.0, 129.2, 125.6, 125.5, 123.2, 121.9, 114.9, 75.5, 42.0, 29.9, 28.8, 22.4, 14.0.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₀H₂₁NO₂S: 339.1293; found: 339.1306.

Methyl 2-[1-Oxo-4-(phenylthio)-1*H*-isochromen-3-ylamino]acetate (3ae)

Starting from 3-(phenylthio)isobenzofuran-1(3*H*)-one (**1a**) (50 mg, 0.21 mmol), methyl 2-isocyanoacetate (**2e**) (23 μ L, 0.25 mmol), TiCl₄ (210 μ L, 1 M in CH₂Cl₂, 0.21 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3ae** as a pale yellow solid (49.3 mg, 70% yield); mp 136–138 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.17–8.11 (m, 1 H), 7.68–7.63 (m, 1 H), 7.60–7.50 (m, 1 H), 7.25–7.09 (m, 6 H), 6.23 (t, J = 5.8 Hz, 1 H), 4.20 (d, J = 6.0 Hz, 2 H), 3.76 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 170.1, 160.1, 158.6, 141.7, 135.8, 130.1, 129.3, 125.8 (2 C), 124.0, 122.4, 115.6, 78.0, 52.6, 43.3.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₈H₁₅NO₄S: 341.0722; found: 341.0710.

3-(4-Chlorophenylamino)-4-(phenylthio)-1*H*-isochromen-1-one (3ag)

Starting from 3-(phenylthio)isobenzofuran-1(3*H*)-one (**1a**) (100 mg, 0.41 mmol), 4-chlorophenylisocyanide (**2g**) (68.5 mg, 0.50 mmol), Ti-Cl₄ (410 μ L, 1 M in CH₂Cl₂, 0.41 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3ag** as a yellow solid (123 mg, 79% yield); mp 130–132 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.09 (dd, *J* = 8.0, 0.9 Hz, 1 H), 7.62 (d, *J* = 8.3 Hz, 2 H), 7.55–7.40 (m, 1 H), 7.24–7.02 (m, 10 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 159.5, 156.1, 141.0, 136.0, 135.9, 135.2, 130.3, 129.58, 129.53, 129.48, 126.2, 125.8, 124.8, 123.0, 122.1, 116.4, 80.0.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₁H₁₄ClNO₂S: 379.0434; found: 379.0452.

3-(2-Methoxyphenylamino)-4-(phenylthio)-1*H*-isochromen-1one (3ah)

Starting from 3-(phenylthio)isobenzofuran-1(3*H*)-one (**1a**) (100 mg, 0.41 mmol), 2-methoxyphenylisocyanide (**2h**) (66.3 mg, 0.50 mmol), TiCl₄ (410 μ L, 1 M in CH₂Cl₂, 0.41 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3ah** as a yellow solid (21.9 mg, 14% yield); mp 100–102 °C.

 ^1H NMR (300 MHz, CDCl_3): δ = 8.46 (s, 1 H), 8.24–8.19 (m, 1 H), 7.97–7.92 (m, 1 H), 7.81–7.76 (m, 1 H), 7.65–7.60 (m, 1 H), 7.31–7.14 (m, 6 H), 7.10–7.00 (m, 2 H), 6.92–6.87 (m, 1 H), 3.83 (s, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 159.9, 156.7, 149.0, 141.7, 135.8, 130.2, 129.3, 127.4, 126.4, 126.0, 124.3, 123.7, 122.9, 121.5, 119.7, 116.0, 110.7, 80.4, 56.1.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₂H₁₇NO₃S: 375.0929; found: 375.0929.

3-(Cyclohexylamino)-4-(p-tolylthio)-1H-isochromen-1-one (3bb)

Starting from 3-(*p*-tolylthio)isobenzofuran-1(3*H*)-one (**1b**) (100 mg, 0.39 mmol), cyclohexylisocyanide (**2b**) (58 μ L, 0.47 mmol), TiCl₄ (390 μ L, 1 M in CH₂Cl₂, 0.39 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (9:1) as eluent gave **3bb** as a yellow solid (98 mg, 69% yield). Starting from 4-methylbenzenethiol (**5b**) (1.0 mmol) and following general procedure C, **3bb** was obtained as a white solid (204 mg, 56% yield); mp 142–144 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.11 (d, *J* = 7.9 Hz, 1 H), 7.60 (d, *J* = 8.1 Hz, 1 H), 7.50 (t, *J* = 7.6 Hz, 1 H), 7.11 (t, *J* = 7.5 Hz, 1 H), 7.07–6.98 (m, 4 H), 5.72 (d, *J* = 8.4 Hz, 1 H), 3.92–3.69 (m, 1 H), 2.27 (s, 3 H), 1.99–1.96 (m, 2 H), 1.77–1.53 (m, 3 H), 1.38–1.34 (m, 2 H), 1.27–1.14 (m, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.6, 159.0, 142.2, 135.4, 135.3, 132.3, 129.9, 125.5, 123.0, 121.9, 114.8, 76.2, 50.6, 33.7, 25.3, 24.6, 20.8.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₂H₂₃NO₂S: 365.1449; found: 365.1442.

Paper

369.1207.

S. Ponra et al.

3-(Cyclohexylamino)-4-[(4-methoxyphenyl)thio]-1*H*-isochromen-1-one (3cb)

Starting from 3-[(4-methoxyphenyl)thio]isobenzofuran-1(3*H*)-one (**1c**) (100 mg, 0.37 mmol), cyclohexylisocyanide (**2b**) (55 μ L, 0.44 mmol), TiCl₄ (370 μ L, 1 M in CH₂Cl₂, 0.37 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (95:5) as eluent gave **3cb** as a yellow solid (91 mg, 65% yield); mp 135–137 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.11 (dd, *J* = 8.0, 1.4 Hz, 1 H), 7.70–7.59 (m, 1 H), 7.60–7.47 (m, 1 H), 7.46–7.33 (m, 1 H), 7.16–7.06 (m, 2 H), 6.87–6.77 (m, 2 H), 5.75 (d, *J* = 8.5 Hz, 1 H), 3.81–3.80 (m, 1 H), 3.75 (s, 3 H), 1.98 (d, *J* = 12.1 Hz, 2 H), 1.74–1.60 (m, 3 H), 1.43–1.35 (m, 2 H), 1.28–1.16 (m, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 159.0, 158.2, 142.3, 135.5, 132.7, 129.9, 127.5, 126.6, 123.0, 122.0, 114.9, 114.6, 55.4, 50.6, 33.8, 25.4, 24.6.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₂H₂₃NO₃S: 381.1399; found: 381.1393.

4-{[4-(*tert*-Butyl)phenyl]thio}-3-(cyclohexylamino)-1*H*-isochromen-1-one (3db)

Starting from 3-(4-*tert*-butylphenylthio)isobenzofuran-1(3*H*)-one (**1d**) (100 mg, 0.34 mmol), cyclohexylisocyanide (**2b**) (50 μ L, 0.40 mmol), TiCl₄ (340 μ L, 1 M in CH₂Cl₂, 0.34 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (95:5) as eluent gave **3db** as a yellow solid (105 mg, 76% yield); mp 109–111 °C.

¹H NMR (300 MHz, $CDCl_3$): $\delta = 8.12$ (dd, J = 8.0, 1.0 Hz, 1 H), 7.63 (dd, J = 8.2, 0.5 Hz, 1 H), 7.55–7.45 (m, 1 H), 7.27–7.23 (m, 2 H), 7.13 (t, J = 7.5 Hz, 1 H), 7.06 (d, J = 8.4 Hz, 2 H), 5.72 (d, J = 8.5 Hz, 1 H), 3.91–3.74 (m, 1 H), 2.02–1.92 (m, 2 H), 1.74–1.68 (m, 2 H), 1.64–1.57 (m, 1 H), 1.45–1.34 (m, 2 H), 1.27 (s, 9 H), 1.24–1.15 (m, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.6, 159.0, 148.7, 142.3, 135.5, 132.4, 129.9, 126.2, 125.3, 123.0, 122.1, 114.8, 76.3, 50.6, 34.4, 33.7, 31.2, 25.3, 24.6.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₅H₂₉NO₂S: 407.1919; found: 407.1933.

3-(Cyclohexylamino)-4-[(4-fluorophenyl)thio]-1*H*-isochromen-1-one (3eb)

Starting from 3-[(4-fluorophenyl)thio]isobenzofuran-1(3*H*)-one (**1e**) (100 mg, 0.38 mmol), cyclohexylisocyanide (**2b**) (57 μ l, 0.46 mmol), TiCl₄ (380 μ L, 1 M in CH₂Cl₂, 0.38 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (9:1) as eluent gave **3eb** as a yellow solid (96 mg, 68% yield); mp 140–142 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.18–8.04 (m, 1 H), 7.61–7.56 (m, 1 H), 7.55–7.51 (m, 1 H), 7.15–7.12 (m, 1 H), 7.11–7.05 (m, 2 H), 6.98–6.90 (m, 2 H), 5.69 (d, *J* = 8.4 Hz, 1 H), 3.87–3.74 (m, 1 H), 1.97 (dd, *J* = 12.6, 3.5 Hz, 2 H), 1.74–1.66 (m, 2 H), 1.65–1.58 (m, 1 H), 1.46–1.35 (m, 2 H), 1.26–1.15 (m, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 161.2 (d, J_{C-F} = 244.0 Hz), 160.4, 159.1, 142.0, 135.6, 130.9 (d, J_{C-F} = 3.1 Hz), 130.0, 127.1 (d, J_{C-F} = 7.8 Hz), 123.2, 121.7, 116.3 (d, J_{C-F} = 22.1 Hz), 114.9, 76.1, 50.6, 33.7, 25.3, 24.6.

¹⁹F NMR (300 MHz, CDCl₃): δ = -117.1.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₁H₂₀FNO₂S: 369.1199; found:

4-(4-Bromophenylthio)-3-(cyclohexylamino)-1*H*-isochromen-1-one (3fb)

Starting from 3-(4-bromophenylthio)isobenzofuran-1(3*H*)-one (**1f**) (100 mg, 0.31 mmol), cyclohexylisocyanide (**2b**) (47 µL, 0.37 mmol), TiCl₄ (310 µL, 1 M in CH₂Cl₂, 0.31 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3fb** as a yellow solid (113 mg, 84% yield); mp 150–152 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.16–8.05 (m, 1 H), 7.55–7.49 (m, 2 H), 7.35–7.28 (m, 2 H), 7.16–7.09 (m, 1 H), 7.00–6.92 (m, 2 H), 5.66 (d, J = 8.5 Hz, 1 H), 3.90–3.72 (m, 1 H), 2.03–1.91 (m, 2 H), 1.77–1.55 (m, 3 H), 1.48–1.11 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 160.4, 159.3, 141.9, 135.7, 135.4, 132.2, 130.1, 127.0, 123.3, 121.8, 119.2, 115.0, 75.1, 50.8, 33.8, 25.4, 24.7.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₁H₂₀BrNO₂S: 429.0398; found: 429.0411.

4-(3-Bromophenylthio)-3-(cyclohexylamino)-1*H*-isochromen-1-one (3gb)

Starting from 3-(3-bromophenylthio)isobenzofuran-1(3*H*)-one (**1g**) (100 mg, 0.31 mmol), cyclohexylisocyanide (**2b**) (46 μ L, 0.37 mmol), TiCl₄ (310 μ L, 1 M in CH₂Cl₂, 0.31 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3gb** as a yellow solid (123 mg, 92% yield); mp 108–110 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.13 (d, J = 7.9 Hz, 1 H), 7.64–7.50 (m, 2 H), 7.28–7.21 (m, 2 H), 7.20–7.04 (m, 2 H), 7.04–6.95 (m, 1 H), 5.62 (d, J = 8.6 Hz, 1 H), 3.91–3.75 (m, 1 H), 2.00–1.95 (m, 2 H), 1.78–1.57 (m, 3 H), 1.50–1.31 (m, 2 H), 1.31–1.13 (m, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.4, 159.2, 141.8, 138.5, 135.7, 130.5, 130.1, 128.6, 127.9, 123.7, 123.3, 123.2, 121.7, 114.9, 74.6, 50.7, 33.7, 25.3, 24.6.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₁H₂₀BrNO₂S: 429.0398; found: 429.0409.

4-(2-Bromophenylthio)-3-(cyclohexylamino)-1*H*-isochromen-1-one (3hb)

Starting from 3-(2-bromophenylthio)isobenzofuran-1(3*H*)-one (**1h**) (100 mg, 0.31 mmol), cyclohexylisocyanide (**2b**) (47 μ L, 0.37 mmol), TiCl₄ (310 μ L, 1 M in CH₂Cl₂, 0.31 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3hb** as a yellow crystalline solid (110 mg, 82% yield); mp 148–150 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.16–8.10 (m, 1 H), 7.53 (m, 3 H), 7.18–7.06 (m, 2 H), 7.03–6.95 (m, 1 H), 6.74 (dd, J = 7.9, 1.6 Hz, 1 H), 5.62 (d, J = 8.5 Hz, 1 H), 3.89–3.75 (m, 1 H), 1.96 (dd, J = 12.2, 3.1 Hz, 2 H), 1.78–1.56 (m, 3 H), 1.42–1.13 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.4, 159.3, 141.9, 136.8, 135.7, 133.2, 130.1, 128.0, 126.7, 125.5, 123.4, 121.9, 121.0, 115.1, 74.9, 50.9, 33.8, 25.4, 24.8.

HRMS (ESI): m/z [M⁻]⁺ (⁷⁹Br) and [M⁻]⁺ (⁸¹Br) calcd for C₂₁H₂₀BrNO₂S: 429.0398 and 431.0398; found: 429.0407 and 431.0359.

Paper

3-(Cyclohexylamino)-4-[(3,4-dimethylphenyl)thio]-1*H*-isochromen-1-one (3ib)

Starting from 3-(3,4-dimethylphenylthio)isobenzofuran-1(3*H*)-one (**1i**) (100 mg, 0.37 mmol), cyclohexylisocyanide (**2b**) (55 μ L, 0.44 mmol), TiCl₄ (370 μ L, 1 M in CH₂Cl₂, 0.37 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (95:5) as eluent gave **3ib** as a yellow solid (125 mg, 89% yield). Starting from 3,4-dimethylbenzenethiol (**5i**) (1.0 mmol) and following general procedure C, **3ib** was obtained as a yellow solid (169 mg, 45% yield); mp 128–130 °C.

¹H NMR (300 MHz, CDCl₃): $\delta = 8.19-8.06$ (m, 1 H), 7.63 (d, J = 8.1 Hz, 1 H), 7.55–7.46 (m, 1 H), 7.16–7.07 (m, 1 H), 6.97 (dd, J = 11.9, 4.7 Hz, 2 H), 6.85 (dd, J = 7.9, 1.9 Hz, 1 H), 5.76 (d, J = 8.5 Hz, 1 H), 3.91–3.77 (m, 1 H), 2.19 (s, 6 H), 2.00 (dd, J = 12.2, 3.0 Hz, 2 H), 1.79–1.57 (m, 3 H), 1.51–1.14 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.6, 159.1, 142.4, 137.6, 135.5, 134.2, 132.6, 130.4, 129.9, 126.9, 123.0 (2 C), 122.1, 115.0, 76.5, 50.7, 33.8, 25.4, 24.7, 19.8, 19.2.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₃H₂₅NO₂S: 379.1606; found: 379.1601.

3-(Cyclohexylamino)-4-[(3,4-dimethoxyphenyl)thio]-1*H*-isochromen-1-one (3jb)

Starting from 3-(3,4-dimethoxyphenylthio)isobenzofuran-1(3*H*)-one (**1j**) (100 mg, 0.33 mmol), cyclohexylisocyanide (**2b**) (49 μ L, 0.40 mmol), TiCl₄ (330 μ L, 1 M in CH₂Cl₂, 0.33 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (8:2) as eluent gave **3jb** as a yellow pasty residue (75 mg, 55% yield).

¹H NMR (300 MHz, CDCl₃): δ = 8.11 (dd, *J* = 7.9, 1.0 Hz, 1 H), 7.63 (d, *J* = 7.9 Hz, 1 H), 7.53 (m, 1 H), 7.12 (m, 1 H), 6.73 (d, *J* = 8.4 Hz, 1 H), 6.72 (d, *J* = 2.1 Hz, 1 H), 6.64 (dd, *J* = 8.4, 2.1 Hz, 1 H), 5.75 (d, *J* = 8.6 Hz, 1 H), 3.86–3.81 (m, 1 H), 3.81 (s, 3 H), 3.78 (s, 3 H), 2.02–1.94 (m, 2 H), 1.76–1.67 (m, 2 H), 1.66–1.58 (m, 1 H), 1.49–1.32 (m, 2 H), 1.30–1.12 (m, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.6, 159.0, 149.5, 147.6, 142.2, 135.5, 129.9, 127.0, 123.1, 121.9, 118.0, 114.8, 112.0, 109.6, 77.0, 56.0, 55.9, 50.6, 33.7, 25.3, 24.6.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₃H₂₅NO₄S: 411.1504; found: 411.1515.

3-(Cyclohexylamino)-7-methyl-4-(phenylthio)-1H-isochromen-1one (3a'b)

Starting from 6-methyl-3-(phenylthio)isobenzofuran-1(3*H*)-one (**1a**') (100 mg, 0.39 mmol), cyclohexylisocyanide (**2b**) (58 µL, 0.47 mmol), TiCl₄ (390 µL, 1 M in CH₂Cl₂, 0.39 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **3a'b** as a yellow crystalline solid (132 mg, 93% yield); mp 126–128 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.93 (s, 1 H), 7.50 (d, *J* = 8.3 Hz, 1 H), 7.35 (dd, *J* = 8.3, 1.7 Hz, 1 H), 7.28–7.17 (m, 2 H), 7.11 (dd, *J* = 7.2, 5.4 Hz, 3 H), 5.60 (d, *J* = 8.5 Hz, 1 H), 3.81 (qd, *J* = 10.2, 5.1 Hz, 1 H), 2.35 (s, 3 H), 1.97 (dd, *J* = 12.4, 3.1 Hz, 2 H), 1.76–1.55 (m, 3 H), 1.48–1.08 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.9, 158.8, 139.9, 137.2, 136.3, 133.1, 129.6, 129.3, 125.6, 125.5, 122.2, 115.0, 75.9, 50.8, 33.9, 25.5, 24.8, 20.9.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₂H₂₃NO₂S: 365.1449; found: 365.1461.

3-(Cyclohexylamino)-7-methoxy-4-(naphthalen-2-ylthio)-1*H*-isochromen-1-one (31"b)

Starting from 6-methoxy-3-(naphthalen-2-ylthio)isobenzofuran-1(3*H*)-one (**1***I''*) (100 mg, 0.31 mmol), cyclohexylisocyanide (**2a**) (46 μ L, 0.37 mmol), TiCl₄ (310 μ L, 1 M in CH₂Cl₂, 0.31 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **31''b** as a yellow solid (114 mg, 86% yield); mp 132–134 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.65 (dd, J = 11.8, 8.0 Hz, 2 H), 7.58–7.44 (m, 3 H), 7.41–7.26 (m, 3 H), 7.17 (dd, J = 8.2, 2.2 Hz, 1 H), 7.06 (dd, J = 9.0, 2.7 Hz, 1 H), 5.49 (d, J = 8.6 Hz, 1 H), 3.83–3.63 (m, 4 H), 1.89 (dd, J = 12.3, 3.2 Hz, 2 H), 1.68–1.43 (m, 3 H), 1.40–0.96 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.8, 158.3, 156.3, 136.4, 134.0, 133.7, 131.8, 129.0, 127.9, 127.0, 126.8, 126.0, 125.6, 124.1, 124.0, 123.2, 115.7, 110.0, 76.0, 55.8, 50.9, 33.9, 25.5, 24.8.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₆H₂₅NO₃S: 431.1555; found: 431.1572.

3-(Cyclohexylamino)-4-(naphthalen-1-ylthio)-1*H*-isochromen-1-one (3kb)

Starting from 3-(naphthalen-2-ylthio)isobenzofuran-1(3*H*)-one (**1k**) (90 mg, 0.31 mmol), cyclohexylisocyanide (**2b**) (46 µl, 0.37 mmol), TiCl₄ (310 µL, 1 M in CH₂Cl₂, 0.31 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (95:5) as eluent gave **3kb** as a yellow solid (80 mg, 68% yield); mp 135–137 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.38 (d, *J* = 8.4 Hz, 1 H), 8.16 (d, *J* = 8.0 Hz, 1 H), 7.89 (d, *J* = 7.6 Hz, 1 H), 7.66–7.60 (m, 2 H), 7.59–7.53 (m, 2 H), 7.51–7.44 (m, 1 H), 7.28–7.24 (m, 1 H), 7.16–7.11 (m, 1 H), 6.96 (d, *J* = 7.4 Hz, 1 H), 5.67 (d, *J* = 8.5 Hz, 1 H), 3.89–3.78 (m, 1 H), 2.01–1.91 (m, 2 H), 1.73–1.64 (m, 2 H), 1.63–1.56 (m, 1 H), 1.43–1.32 (m, 2 H), 1.22–1.08 (m, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 160.6, 159.2, 142.1, 135.6, 134.0, 132.3, 130.9, 130.0, 128.7, 126.3, 126.2, 125.9, 125.6, 123.5, 123.2, 122.0, 121.0, 115.1, 74.0, 50.7, 33.7, 25.3, 24.6.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₅H₂₃NO₂S: 401.1449; found: 401.1450.

3-(Cyclohexylamino)-4-(naphthalen-2-ylthio)-1*H*-isochromen-1-one (3lb)

Starting from 3-(naphthalen-2-ylthio)isobenzofuran-1(3*H*)-one (**1l**) (100 mg, 0.34 mmol), cyclohexylisocyanide (**2b**) (51 μ L, 0.41 mmol), TiCl₄ (340 μ L, 1 M in CH₂Cl₂, 0.34 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3lb** as a yellow solid (115 mg, 84% yield). Starting from naphthalene-2-thiol (**5l**) (1.0 mmol) and following general procedure C, **3lb** was obtained as a yellow solid (137 mg, 34% yield); mp 134–136 °C.

¹H NMR (400 MHz, CDCl₃): δ = 8.02 (dd, *J* = 8.0, 1.0 Hz, 1 H), 7.61 (dd, *J* = 13.4, 8.2 Hz, 2 H), 7.51 (d, *J* = 8.6 Hz, 2 H), 7.39–7.22 (m, 4 H), 7.15 (dd, *J* = 8.6, 1.9 Hz, 1 H), 7.02–6.95 (m, 1 H), 5.64 (d, *J* = 8.5 Hz, 1 H), 3.77–3.65 (m, 1 H), 1.90–1.80 (m, 2 H), 1.61–1.41 (m, 3 H), 1.33–0.92 (m, 5 H).

¹³C NMR (101 MHz, CDCl₃): δ = 160.7, 159.2, 142.2, 135.6, 133.9, 133.4, 131.7, 130.0, 128.9, 127.8, 126.9, 126.7, 125.5, 124.0, 123.2, 123.0, 122.0, 115.0, 75.7, 50.7, 33.7, 25.3, 24.7.

HRMS (ESI): m/z [M⁺]⁺ calcd for C₂₅H₂₃NO₂S: 401.1449; found: 401.1463.

3-(Cyclohexylamino)-4-(hexylthio)-1H-isochromen-1-one (3nb)

Starting from 3-(hexylthio)isobenzofuran-1(3*H*)-one (**1n**) (100 mg, 0.40 mmol), cyclohexylisocyanide (**2b**) (60 µL, 0.48 mmol), TiCl₄ (400 µL, 1 M in CH₂Cl₂, 0.4 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3nb** as a yellow solid (133.8 mg, 93% yield). Starting from hexane-1-thiol (**5n**) (1.0 mmol) and following general procedure C, **3nb** was obtained as a yellow solid (187 mg, 52% yield); mp 58–60 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.09–8.05 (m, 1 H), 7.79–7.71 (m, 1 H), 7.60–7.55 (m, 1 H), 7.12–7.07 (m, 1 H), 5.81 (d, *J* = 8.6 Hz, 1 H), 3.86–3.69 (m, 1 H), 2.57–2.48 (m, 2 H), 2.07–1.95 (m, 2 H), 1.82–1.15 (m, 16 H), 0.86 (t, *J* = 6.8 Hz, 3 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 160.9, 158.6, 142.8, 135.3, 130.1, 122.7, 122.0, 115.0, 78.9, 50.6, 34.8, 34.0, 31.5, 29.8, 28.7, 25.6, 24.8, 22.6, 14.1.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₁H₂₉NO₂S: 359.1919; found: 359.1929.

4-(*tert*-Butylthio)-3-(cyclohexylamino)-1*H*-isochromen-1-one (3pb)

Starting from 3-(*tert*-butylthio)isobenzofuran-1(3*H*)-one (**1p**) (100 mg, 0.45 mmol), cyclohexylisocyanide (**2b**) (67 μ L, 0.54 mmol), TiCl₄ (450 μ L, 1 M in CH₂Cl₂, 0.45 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3pb** as a yellow sticky gum (23.7 mg, 16% yield).

¹H NMR (300 MHz, CDCl₃): δ = 8.08–8.03 (m, 1 H), 7.87–7.82 (m, 1 H), 7.60–7.40 (m, 1 H), 7.10–7.04 (m, 1 H), 5.98 (d, *J* = 8.5 Hz, 1 H), 3.87–3.71 (m, 1 H), 2.00 (d, *J* = 4.1 Hz, 2 H), 1.77–1.61 (m, 4 H), 1.54–1.18 (m, 13 H).

 ^{13}C NMR (75 MHz, CDCl_3): δ = 160.9, 159.9, 144.3, 134.9, 129.8, 123.2, 122.6, 114.6, 50.6, 49.9, 34.2, 33.9, 31.5, 25.6, 24.9.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₉H₂₅NO₂S: 331.1606; found: 331.1617.

Methyl 2-[3-(Cyclohexylamino)-1-oxo-1*H*-isochromen-4-ylthio]acetate (3rb)

Starting from methyl 2-(3-oxo-1,3-dihydroisobenzofuran-1ylthio)acetate (**1r**) (100 mg, 0.42 mmol), cyclohexylisocyanide (**2b**) (63 μ L, 0.50 mmol), TiCl₄ (420 μ L, 1 M in CH₂Cl₂, 0.42 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3rb** as an orange viscous liquid (34.6 mg, 24% yield).

 ^1H NMR (300 MHz, CDCl₃): δ = 8.10–8.04 (m, 1 H), 7.72–7.69 (m, 1 H), 7.65–7.50 (m, 1 H), 7.14–7.07 (m, 1 H), 6.28 (d, J = 8.3 Hz, 1 H), 3.85–3.70 (m, 1 H), 3.64 (s, 3 H), 3.29 (s, 2 H), 2.06–1.95 (m, 2 H), 1.84–1.72 (m, 2 H), 1.65 (dd, J = 9.1, 3.4 Hz, 1 H), 1.52–1.17 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 171.3, 160.7, 159.3, 142.4, 135.5, 130.3, 122.9, 121.5, 114.9, 52.7, 50.8, 36.8, 33.8, 25.6, 24.9.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₁₈H₂₁NO₄S: 347.1191; found: 347.1176.

3-(Benzylamino)-4-(cyclohexylthio)-1H-isochromen-1-one (3od)

Starting from 3-(cyclohexylthio)isobenzofuran-1(3*H*)-one (**1o**) (100 mg, 0.40 mmol), benzylisocyanide (**2d**) (59 μ L, 0.48 mmol), TiCl₄ (400 μ L, 1 M in CH₂Cl₂, 0.40 mmol) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3od** as a yellow solid (81 mg, 55% yield); mp 128–130 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.00 (dd, *J* = 8.0, 0.9 Hz, 1 H), 7.71 (d, *J* = 8.2 Hz, 1 H), 7.53–7.40 (m, 1 H), 7.30–7.15 (m, 5 H), 7.10–7.02 (m, 1 H), 6.14 (t, *J* = 6.2 Hz, 1 H), 4.50 (d, *J* = 6.3 Hz, 2 H), 2.69–2.57 (m, 1 H), 1.83–1.72 (m, 2 H), 1.67–1.38 (m, 3 H), 1.29–1.00 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.5, 158.9, 143.0, 138.3, 135.2, 130.0, 128.9, 127.8, 127.7, 123.1, 122.6, 115.3, 79.2, 47.1, 46.0, 33.6, 26.2, 25.7.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₂H₂₃NO₂S: 365.1449; found: 365.1450.

4-(Benzylthio)-3-(cyclohexylamino)-1H-isochromen-1-one (3qb)

Starting from 3-(benzylthio)isobenzofuran-1(3*H*)-one (**1q**) (100 mg, 0.39 mmol), cyclohexylisocyanide (**2b**) (58 μ L, 0.47 mmol), TiCl₄ (390 μ L, 1 M in CH₂Cl₂, 1.0 equiv) and following the general procedure B, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (98:2) as eluent gave **3qb** as a yellow sticky gum (100 mg, 70% yield).

 ^1H NMR (300 MHz, CDCl₃): δ = 8.13–8.05 (m, 1 H), 7.75–7.69 (m, 1 H), 7.52–7.65 (m, 1 H), 7.25–7.17 (m, 3 H), 7.14–7.04 (m, 3 H), 5.41 (d, J=8.3 Hz, 1 H), 3.67 (s, 2 H), 3.57–3.44 (m, 1 H), 1.75–1.50 (m, 5 H), 1.39–0.84 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.7, 159.2, 142.6, 138.5, 135.4, 130.1, 128.9, 128.6, 127.2, 122.7, 121.7, 114.8, 77.1, 50.3, 38.6, 33.6, 25.5, 24.7.

HRMS (APPI): m/z [M⁺]⁺ calcd for C₂₂H₂₃NO₂S: 365.1449; found: 365.1436.

3-(Pentylamino)-4-(p-tolylthio)-1H-isochromen-1-one (3bc)

Starting from 2-carboxybenzaldehyde (**4a**) (165 mg, 1.1 mmol), 4methylbenzenethiol (**5b**) (124 mg, 1.0 mmol), 1-pentylisocyanide (**2c**) (189 mg, 1.5 mmol), TiCl₄ (1.2 mL, 1 M in CH_2Cl_2 , 1.2 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using petroleum ether/ethyl acetate (95:5) as eluent gave **3bc** as a yellow pasty residue (257 mg, 73% yield).

¹H NMR (400 MHz, $CDCI_3$): $\delta = 8.12$ (d, J = 8.0 Hz, 1 H), 7.61 (d, J = 8.2 Hz, 1 H), 7.53 (dd, J = 11.4, 3.9 Hz, 1 H), 7.13 (dd, J = 11.0, 4.0 Hz, 1 H), 7.03 (q, J = 8.0 Hz, 4 H), 5.81 (t, J = 5.8 Hz, 1 H), 3.41 (dd, J = 13.5, 6.7 Hz, 2 H), 2.27 (s, 3 H), 1.61–1.51 (m, 2 H), 1.33–1.24 (m, 4 H), 0.87 (t, J = 6.8 Hz, 3 H).

 ^{13}C NMR (101 MHz, CDCl₃): δ = 160.5, 159.5, 142.3, 135.6, 135.4, 132.4, 130.0, 129.9 (2 C), 125.4 (2 C), 123.1, 121.9, 114.8, 75.9, 41.9, 29.9, 28.8, 22.3, 20.9, 13.9.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₁H₂₃NO₂S: 353.1449; found: 353.1448.

к

S. Ponra et al.

3-[(2,6-Dimethylphenyl)amino]-4-(*p*-tolylthio)-1*H*-isochromen-1-one (3bf)

Starting from 2-carboxybenzaldehyde (**4a**) (165 mg, 1.1 mmol), 4methylbenzenethiol (**5b**) (124 mg, 1.0 mmol), 2,6-dimethylphenylisocyanide (**2f**) (197 mg, 1.5 mmol), TiCl₄ (1.2 mL, 1 M in CH₂Cl₂, 1.2 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (8:2) as eluent gave **3bf** as a yellow solid (190 mg, 49% yield); mp 135–141 °C.

¹H NMR (300 MHz, CDCl₃): δ = 8.13 (dd, *J* = 8.0, 1.4 Hz, 1 H), 7.78–7.72 (m, 1 H), 7.65–7.55 (m, 1 H), 7.24–7.15 (m, 2 H), 7.13–7.06 (m, 7 H), 2.30 (s, 3 H), 2.20 (s, 6 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.1, 158.0, 141.9, 136.1, 135.69, 135.65, 133.6, 133.1, 132.0, 130.3, 130.1, 128.5, 128.2, 127.5, 125.7, 123.7, 122.2, 115.6, 20.9, 18.4.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₄H₂₁NO₂S: 387.1293; found: 387.1286.

4-(Dodecylthio)-3-(pentylamino)-1H-isochromen-1-one (3mc)

Starting from 2-carboxybenzaldehyde (**4a**) (165 mg, 1.1 mmol), dodecane-1-thiol (**5m**) (240 μ L, 1.0 mmol), pentylisocyanide (**2c**) (189 μ L, 1.5 mmol), TiCl₄ (1.2 mL, 1 M in CH₂Cl₂, 1.2 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **3mc** as a yellow sticky gum (163 mg, 38% yield).

¹H NMR (300 MHz, CDCl₃): δ = 8.11–8.01 (m, 1 H), 7.75 (d, *J* = 7.9 Hz, 1 H), 7.60–7.50 (m, 1 H), 7.12–7.05 (m, 1 H), 5.90 (t, *J* = 6.0 Hz, 1 H), 3.40 (dd, *J* = 13.3, 6.9 Hz, 2 H), 2.55–2.47 (m, 2 H), 1.65–1.40 (m, 4 H), 1.40–1.17 (m, 22 H), 0.95–0.80 (m, 6 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.7, 159.1, 142.7, 135.2, 130.0, 122.7, 122.0, 115.0, 78.8, 41.9, 34.7, 32.0, 30.1, 29.8, 29.7 (3 C), 29.6, 29.4, 29.3, 29.0 (2 C), 22.7, 22.4, 14.1, 14.0.

HRMS (ESI): m/z [M[•]]⁺ calcd for C₂₆H₄₁NO₂S: 431.2858; found: 431.2849.

3-(Benzylamino)-4-(dodecylthio)-1H-isochromen-1-one (3md)

Starting from 2-carboxybenzaldehyde (**4a**) (165 mg, 1.1 mmol), 1-dodecanethiol (**5m**) (241 μ L, 1.0 mmol), benzylisocyanide (**2d**) (183 μ L, 1.5 mmol), TiCl₄ (1.2 mL, 1 M in CH₂Cl₂, 1.2 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (9:1) as eluent gave **3md** as a yellow pasty residue (277 mg, 61% yield).

¹H NMR (300 MHz, $CDCl_3$): $\delta = 8.11$ (dd, J = 8.0, 0.9 Hz, 1 H), 7.81–7.76 (m, 1 H), 7.65–7.55 (m, 1 H), 7.38–7.33 (m, 4 H), 7.34–7.28 (m, 1 H), 7.20–7.10 (m, 1 H), 6.18 (t, J = 6.2 Hz, 1 H), 4.61 (d, J = 6.2 Hz, 2 H), 2.51 (t, J = 7.4 Hz, 2 H), 1.55–1.42 (m, 2 H), 1.24 (d, J = 8.4 Hz, 18 H), 0.88 (t, J = 6.7 Hz, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.5, 158.4, 142.4, 138.2, 135.3, 130.0, 128.8 (2 C), 127.7, 127.6 (2 C), 123.1, 122.1, 115.3, 79.8, 45.9, 34.7, 31.9, 29.7, 29.6 (2 C), 29.6, 29.5, 29.3, 29.2, 29.0, 22.7, 14.1.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₈H₃₇NO₂S: 451.2545; found: 451.2544.

3-(2,6-Dimethylphenylamino)-4-(dodecylthio)-1*H*-isochromen-1-one (3mf)

Starting from 2-carboxybenzaldehyde (**4a**) (165 mg, 1.1 mmol), dodecane-1-thiol (**5m**) (240 μ L, 1.0 mmol), 2,6-dimethylphenylisocyanide (**2f**) (197 mg, 1.5 mmol), TiCl₄ (1.2 mL, 1 M in CH₂Cl₂, 1.2 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **3mf** as a pasty orange solid (236 mg, 51% yield).

¹H NMR (300 MHz, CDCl₃): δ = 8.17–8.06 (m, 1 H), 7.91 (d, *J* = 8.2 Hz, 1 H), 7.67 (s, 1 H), 7.34 (s, 1 H), 7.22–7.13 (m, 4 H), 2.77–2.67 (m, 2 H), 2.33 (s, 6 H), 1.77–1.65 (m, 2 H), 1.51–1.23 (m, 18 H), 0.93 (t, *J* = 6.6 Hz, 3 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.1, 157.4, 142.2, 136.1, 135.3, 134.2, 130.1, 128.5, 127.3, 123.3, 122.2, 115.8, 79.9, 34.8, 32.0, 30.1, 29.7 (3 C), 29.5, 29.4, 29.3, 29.1, 22.7, 18.6, 14.2.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₉H₃₉NO₂S: 465.2702; found: 465.2710.

3-(Cyclohexylamino)-7-methyl-4-(*p*-tolylthio)-1*H*-isochromen-1one (3b'b)

Starting from 2-formyl-5-methylbenzoic acid (**4b**) (100 mg, 0.61 mmol), 4-methylbenzenethiol (**5b**) (69.0 mg, 0.55 mmol), cyclohexylisocyanide (**2b**) (103 μ L, 0.83 mmol), TiCl₄ (665 μ L, 1 M in CH₂Cl₂, 0.665 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **3b'b** as a yellow crystalline solid (109.1 mg, 52% yield); mp 138–140 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.93 (s, 1 H), 7.51 (d, J = 8.3 Hz, 1 H), 7.34 (dd, J = 8.3, 1.6 Hz, 1 H), 7.07–6.97 (m, 4 H), 5.63 (d, J = 8.5 Hz, 1 H), 3.87–3.72 (m, 1 H), 2.31 (d, J = 23.5 Hz, 6 H), 1.97 (dd, J = 12.3, 3.0 Hz, 2 H), 1.77–1.55 (m, 3 H), 1.48–1.12 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.9, 158.7, 140.0, 137.1, 135.4, 133.0, 132.7, 130.0, 129.5, 125.7, 122.2, 115.0, 76.5, 50.8, 33.9, 25.5, 24.8, 21.0, 20.9.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₃H₂₅NO₂S: 379.1606; found: 379.1595.

3-(Cyclohexylamino)-7-methoxy-4-(*p*-tolylthio)-1*H*-isochromen-1-one (3b"b)

Starting from 2-formyl-5-methoxybenzoic acid (**4c**) (100 mg, 0.55 mmol), 4-methylbenzenethiol (**5b**) (63.0 mg, 0.50 mmol), cyclohexylisocyanide (**2b**) (94 μ L, 0.75 mmol), TiCl₄ (600 μ L, 1 M in CH₂Cl₂, 0.6 mmol) and following the general procedure C, then purification of the crude material by flash column chromatography on silica gel using cyclohexane/ethyl acetate (99:1) as eluent gave **3b"b** as a yellow solid (112.3 mg, 57% yield); mp 118–120 °C.

¹H NMR (300 MHz, CDCl₃): δ = 7.61–7.45 (m, 2 H), 7.15 (dd, J = 8.9, 2.8 Hz, 1 H), 7.08–6.95 (m, 4 H), 5.57 (d, J = 8.6 Hz, 1 H), 3.87–3.71 (m, 4 H), 2.26 (s, 3 H), 2.04–1.91 (m, 2 H), 1.75–1.55 (m, 3 H), 1.48–1.09 (m, 5 H).

 ^{13}C NMR (75 MHz, CDCl₃): δ = 160.7, 158.1, 156.1, 136.4, 135.4, 132.6, 129.9, 125.9, 125.6, 123.9, 115.5, 109.8, 76.6, 55.7, 50.8, 33.9, 25.5, 24.7, 20.9.

HRMS (ESI): m/z [M[·]]⁺ calcd for C₂₃H₂₅NO₃S: 395.1555; found: 395.1557.

Funding Information

This work has been supported by IDEX, referenced as «ANR-10-IDEX-0001-02 PSL \star .

Acknowledgment

A.N. thanks the Ecole Polytechnique for a fellowship and the authors thank PSL \star , Chimie ParisTech, ENS and ENSTA-ParisTech for financial support.

Supporting Information

Supporting information for this article is available online at https://doi.org/10.1055/s-0036-1591733.

References

- (a) Barry, R. D. Chem. Rev. **1964**, 64, 229. (b) Hill, R. A. In Progress in the Chemistry of Organic Natural Products; Herz, W., Ed.; Springer-Verlag: New York, **1986**, 1. (c) Mohammad, A.; Ur, R. I.; Akhtar, K. G. J. Chem. Soc. Pak. **1998**, 20, 76. (d) Pochet, L.; Frederick, R.; Masereel, B. Curr. Pharm. Des. **2004**, 10, 3781. (e) Saeed, A. Eur. J. Med. Chem. **2016**, 116, 290.
- (2) (a) Napolitano, E. Org. Prep. Proced. Int. 1997, 29, 631. (b) Pal, S.; Chatare, V.; Pal, M. Curr. Org. Chem. 2011, 15, 782. (c) Ashraf, Z. Chem. Heterocycl. Compd. 2016, 52, 149. (d) Saeed, A.; Larik, F. A. Chem. Heterocycl. Compd. 2016, 52, 450.
- (3) (a) Nozawa, K.; Yamada, M.; Tsuda, Y.; Kawai, K.; Nakajima, S. Chem. Pharm. Bull. 1981, 29, 2689. (b) Furuta, T.; Fukuyama, Y.; Asakawa, Y. Phytochemistry 1986, 25, 517. (c) Yadav, P.; Purohit, N. V. J. Chem. Sci. 2013, 125, 165. (d) Purohit, N.; Yadav, P. Indian J. Pharm. Sci. 2011, 73, 171.
- (4) (a) Natsugari, H.; Tawada, H.; Ikeda, H. Eur. Patent Appl. EP0481383A1, **1992**. (b) Kerrigan, J. E.; Oleksyszyn, J.; Kam, C. M.; Selzler, J.; Powers, J. C. *J. Med. Chem.* **1995**, *38*, 544. (c) Powers, H. J. C.; Asgian, J. L.; Ekici, Ö. D.; James, K. E. Chem. Rev. **2002**, *102*, 4639. (d) Heynekamp, J. J.; Hunsaker, L. A.; Vander Jagt, T. A.; Royer, R. E.; Deck, L. M.; Vander Jagt, D. L. Bioorg. Med. Chem. **2008**, *16*, 5285. (e) Child, M. A.; Hall, C. I.; Beck, J. R.; Ofori, L. O.; Albrow, V. E.; Garland, M.; Bowyer, P. W.; Bradley, P. J.; Powers, J. C.; Boothroyd, J. C.; Weerapana, E.; Bogyo, M. Nat. Chem. Biol. **2013**, *9*, 651. (f) Peuchmaur, M.; Lacour, M. A.; Sévalle, J.; Lisowski, V.; Touati-Jallabe, Y.; Rodier, F.; Martinez, J.; Checler, F.; Hernandez, J. F. Bioorg. Med. Chem. **2013**, *21*, 1018.
- (5) (a) Larock, R. C.; Yum, E. K.; Doty, M. J.; Sham, K. K. C. J. Org. Chem. 1995, 60, 3270. (b) Larock, R. C.; Doty, M. J.; Han, X. J. Org. Chem. 1999, 64, 8770. (c) Ueura, K.; Satoh, T.; Miura, M. J. Org. Chem. 2007, 72, 5362. (d) Ueura, K.; Satoh, T.; Miura, M. Org. Lett. 2007, 9, 1407. (e) Kajita, Y.; Kurahashi, T.; Matsubara, S. J. Am. Chem. Soc. 2008, 130, 17226. (f) Shi, Y.; Roth, K. E.; Ramgren, S. D.; Blum, S. A. J. Am. Chem. Soc. 2009, 131, 18022. (g) Ackermann, L.; Pospech, J.; Graczyk, K.; Rauch, K. Org. Lett. 2012, 14, 930. (h) Zhao, P.; Chen, D.; Song, G.; Han, K.; Li, X.

J. Org. Chem. 2012, 77, 1579. (i) Chinnagolla, R. K.; Jeganmohan, M. Chem. Commun. 2012, 48, 2030. (j) Guo, X. X. J. Org. Chem. 2013, 78, 1660. (k) Frasco, D. A.; Lilly, C. P.; Boyle, P. D.; Ison, E. A. ACS Catal. 2013, 3, 2421. (l) Unoh, Y.; Hirano, K.; Satoh, T.; Miura, M. Tetrahedron 2013, 69, 4454. (m) Wang, H.; Han, X.; Lu, X. Tetrahedron 2013, 69, 8626. (n) Li, X. G.; Liu, K.; Zou, G.; Liu, P. N. Adv. Synth. Catal. 2014, 356, 1496. (o) Mo, J.; Wang, L.; Cui, X. Org. Lett. 2015, 17, 4960. (p) Warratz, S.; Kornhaaß, C.; Cajaraville, A.; Niepötter, B.; Stalke, D.; Ackermann, L. Angew. Chem. Int. Ed. 2015, 54, 5513. (q) Prakash, R.; Shekarrao, K.; Gogoi, S.; Boruah, R. C. Chem. Commun. 2015, 51, 9972. (r) Zhang, J.; Han, X.; Lu, X. J. Org. Chem. 2016, 81, 3423. (s) Liu, H.; Yang, Y.; Wu, J.; Wang, X.-N.; Chang, J. Chem. Commun. 2016, 52, 6801. (t) Zheng, M.; Huang, L.; Tong, Q.; Wu, W.; Jiang, H. Eur. J. Org. Chem. 2016, 663.

- (6) (a) Yao, T.; Larock, R. C. J. Org. Chem. 2003, 68, 5936. (b) Rossi, R.; Carpita, A.; Bellina, F.; Stabile, P.; Mannina, L. Tetrahedron 2003, 59, 2067. (c) Peuchmaur, M.; Lisowski, V.; Gandreuil, C.; Maillard, L. T.; Martinez, J.; Hernandez, J. F. J. Org. Chem. 2009, 74, 4158. (d) Roy, S.; Roy, S.; Neuenswander, B.; Hill, D.; Larock, R. C. J. Comb. Chem. 2009, 11, 1128. (e) Verma, A. K.; Rustagi, V.; Aggarwal, T.; Singh, A. P. J. Org. Chem. 2010, 75, 7691. (f) Li, Z.; Hong, J.; Weng, L.; Zhou, X. Tetrahedron 2012, 68, 1552. (g) Faizi, D. J.; Issaian, A.; Davis, A. J.; Blum, S. A. J. Am. Chem. Soc. 2016, 138, 2126.
- (7) (a) Nakamura, Y.; Ukita, T. Org. Lett. 2002, 4, 2317. (b) Tadd, A. C.; Fielding, M. R.; Willis, M. C. Chem. Commun. 2009, 3, 6744. (c) Chen, B.; Ma, S. Org. Lett. 2013, 15, 3884. (d) Waseem, M. A.; Abumahdi, S. A. A.; Srivastava, A.; Srivastava, A.; Siddiqui, R. I. R. Catal. Commun. 2014, 55, 70. (e) Panda, N.; Mishra, P.; Mattan, I. J. Org. Chem. 2016, 81, 1047. (f) Liu, P. N.; Li, X. G.; Sun, M.; Liu, K.; Jin, Q. Chem. Commun. 2014, 51, 2380. (g) Calderón Ortiz, L. K.; Würfel, H.; Täuscher, E.; Weiβ, D.; Birckner, E.; Görls, H.; Beckert, R. Synthesis 2014, 46, 126. (h) Knott, E. B. J. Chem. Soc. 1963, 402.
- (8) (a) Faggi, C.; Garcia-Valverde, M.; Marcaccini, S.; Menchi, G. Org. Lett. 2010, 12, 788. (b) Soleimani, E.; Zainali, M. J. Org. Chem.
 2011, 76, 10306. See also: (c) Ramazani, A.; Mahyari, A. Helv. Chim. Acta 2010, 93, 2203. (d) Zhang, Y.; Ao, Y.-F.; Huang, Z.-T.; Wang, D.-X.; Wang, M.-X.; Zhu, J. Angew. Chem. Int. Ed. 2016, 55, 5282.
- (9) For reviews on isocyanide insertion reactions, see: (a) Qiu, G.; Ding, Q.; Wu, J. Chem. Soc. Rev. 2013, 42, 5257. (b) Isocyanide Chemistry: Applications in Synthesis and Materials Science; Nenajdenko, V., Ed.; Wiley-VCH: Weinheim, 2012.
- (10) (a) El Kaïm, L.; Grimaud, L.; Oble, J. Angew. Chem. Int. Ed. 2005, 44, 7961. (b) El Kaïm, L.; Grimaud, L. Eur. J. Org. Chem. 2014, 7749; and references cited therein. (c) Basavanag, U. M.; Dos Santos, A.; El Kaïm, L.; Gamez-Montano, R.; Grimaud, L. Angew. Chem. Int. Ed. 2013, 52, 7194. (d) Martinand-Lurin, E.; Dos Santos, A.; El Kaïm, L.; Grimaud, L.; Retailleau, P. Chem. Commun. 2014, 50, 2214.
- (11) Ponra, S.; Nyadanu, A.; El Kaim, L.; Grimaud, L.; Vitale, M. R. Org. *Lett.* **2016**, *18*, 4060.
- (12) Dewanji, A.; Mìck-Lichtenfeld, C.; Bergander, K.; Daniliuc, C. G.; Studer, A. *Chem. Eur. J.* **2015**, *21*, 12295.