# Acylshikonin Analogues: Synthesis and Inhibition of DNA Topoisomerase-I

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Compounds bearing an acyl group of a various size at 1'-OH of shikonin were synthesized as acyl analogues of shikonin, which was isolated from the root of  $Lithospermum\ erythrorhizon$ , and evaluated for inhibitory effect on topoisomerase-I activity. A selective acylation at 1'-OH of shikonin in the presence of dicyclohexylcarbodiimide and 4-(dimethylamino)pyridine gave rise to a good yield of corresponding acylshikonin derivatives. In general, analogues with an acyl group of shorter chain lengths  $(C_2-C_6)$  exerted a stronger inhibitory action than those with longer chain lengths  $(C_7-C_{20})$ . While the halogen substitution at C-2 of the acetyl moiety failed to increase the inhibitory potency, the placement of double bonds in the acyl group  $(C_5-C_7)$  augmented the potency remarkably. Of the 32 derivatives evaluated, 15 compounds exhibited a higher inhibitory effect than shikonin. Noteworthy, the inhibitory potency of acetylshikonin, propanoylshikonin, and 4-pentenoylshikonin was approximately 4-fold greater than that of camptothecin. All these data suggest that the size of acyl moiety is important for the enhancement of potency, and the presence of olefinic double bonds is also beneficial.

#### Introduction

There are two major types of antitumor agents which are known to inhibit DNA topoisomerases. The wellstudied class of DNA topoisomerase-I inhibitors are camptothecin analogues, some of which are being tested clinically as anticancer drugs against colon and other cancers. 1-5 However, some limitations such as poor water solubility and toxicity still exist. Meanwhile, the anthracycline compounds, highly effective in the treatment of human neoplastic diseases<sup>6</sup> such as leukemias and sarcomas, seem to inhibit topoisomerase-II as a primary target.7 These compounds, however, also inhibit topoisomerase-I nonspecifically,8 and moreover, the myocardial toxicity of these drugs limits their wider use in chemotherapy for prolonged periods.9 For these reasons, efforts have been made to develop compounds showing a selective inhibition of the respective DNA topoisomerase.

Previously, 10 it was reported that shikonin, one of antineoplastic components isolated from Lithospermi radix (Lithospermum erythrorhizon, Boraginaceae), exhibited a strong cytotoxicity against L1210 cells. The following study<sup>11</sup> showed that acetylshikonin, another constituent of Lithospermi radix, possessed a good antitumor action (T/C value, 182%) in ICR mice bearing Sarcoma 180 cells. Since shikonin and acetylshikonin contain a substituted naphthazarine skeleton, which is present in the structure of anthracyclinones<sup>12</sup> and mitoxanthrone, 13 it was assumed that some part of antineoplastic action of these compounds might be related to the inhibition of DNA topoisomerases. Furthermore, the higher antitumor action of acetylshikonin, compared to shikonin, encouraged us to synthesize shikonin analogues with acyl moieties of various sizes.

Here, we report that shikonin and its analogues possessing short acyl chain lengths at 1'-OH are potent

inhibitors of DNA topoisomerase-I as assayed by *in vitro* relaxation of supercoiled DNA.

### Chemistry

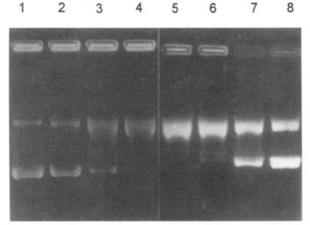
The synthesis of acylshikonins has been carried out by a selective acylation at 1'-OH of shikonin in the presence of dicyclohexylcarbodiimide (DCC) and 4-(dimethylamino)pyridine (DMAP) and is outlined in Scheme 1. The reaction in a molar ratio of shikonin/DMAP (1: 0.4) resulted in relatively good yields (30-92%) of 1'acylshikonins [2-[1-(acyloxy)-4-methylpent-3-enyl]-5,8dihydroxy-1,4-naphthoquinones]. The hydrogen bonding between phenolic OH and quinoid carbonyl seems to be a reason for hindered acylation at the phenolic OH's. Evidence for the acylation was mainly brought from the <sup>1</sup>H-NMR spectra; the signals at  $\delta$  12.30–12.60 ppm, which were attributed to protons at two phenolic OH's. was still present, but the signal at  $\delta$  2.38 ppm, due to the 1'-OH, disappeared. Instead, the signals at  $\delta$  2.15-7.70 ppm, corresponding to protons at C-2 in acyl moiety, appeared. The carbonyl group of the newlyformed acyl moiety could be also recognized from signals ( $\delta$  160-176 ppm) in the <sup>13</sup>C-NMR spectra.

#### **Results and Discussion**

Figure 1 shows that the DNA relaxation activity of topoisomerase-I, based on the densitometry measurement, was reduced by n-hexane extract of L. erythrorhizon in a concentration-dependent manner. In further studies, two active components were isolated from n-hexane extract and identified to be shikonin and acetylshikonin, which showed IC<sub>50</sub> values of 208 and 45  $\mu$ M (Figure 2), respectively. In a separate experiment, shikonin and acetylshikonin were tested for the inhibition of topoisomerase-II. However, these compounds exerted no significant inhibition of topoisomerase-II activity even at 500  $\mu$ M. Thus, it appeared that shikonin and acetylshikonin exerted a preferential inhibition of topoisomerase-I. The higher potency of acetylshikonin, compared to shikonin, in the inhibition of DNA topoisomerase-I led us to examine analogues of acylshikonin bearing various acyl moieties; they included short

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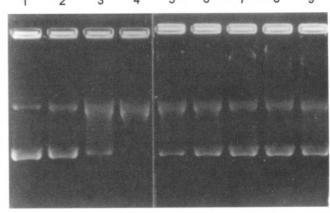
<sup>\*</sup> Abstract published in Advance ACS Abstracts, February 15, 1995.



**Figure 1.** Effect of *n*-hexane extract of Lithospermi root on DNA topoisomerase-I from HeLa cells. The reaction mixture  $(20~\mu\text{L})$  contained 50 mM HEPES (pH 7.0), 10 mM MgCl<sub>2</sub>, 150 mM KCl, 0.75  $\mu\text{g}$  of DNA, 1 mM DTT, 5  $\mu\text{g}$  of BSA, and the enzyme. Where indicated, the *n*-hexane extract of various concentrations was included. Lane 1: no enzyme, no drug. Lanes 2–4: 0.5, 1, and 2 units of the enzyme. Lanes 5–8: 2 units of the enzyme, and 25, 50, 100, and 200  $\mu\text{g/mL}$  of *n*-hexane extract.

or long fatty acyl groups and benzoyl or phenylacetyl groups, as well as retinovl moiety. When the synthetic analogues were evaluated for the inhibitory effect on topoisomerase-I activity, it was found that the inhibitory potency (IC<sub>50</sub> value) was strongly dependent on the size of acyl substituent (Table 1). Of the compounds tested, acetylshikonin (1), n-propanoylshikonin (4), and 4-pentenoylshikonin (8) were the most potent inhibitors with similar IC<sub>50</sub> values (40–45  $\mu$ M). Moreover, these analogues (1, 4, and 8) were approximately 4-fold more potent than camptothecin (IC50, 172 µM). Among the analogues (4, 5, 7, 10, and 14) with stretched and saturated acyl chains (C<sub>3</sub>-C<sub>7</sub>), the inhibitory potency seemed to decrease gradually with increasing size (n)of acyl moiety. And, the analogues (17-20 and 26-30) with chain lengths longer than n-heptanoyl group showed a negligible inhibition (IC<sub>50</sub>, >625  $\mu$ M). In addition, undecenyl- and retinoylshikonins had no remarkable activity.

Substitution on the acyl group was attempted in order to find a more potent inhibitor; the halogen substitution at C-2 of the acetyl moiety with an electron-withdrawing



**Figure 2.** Effect of acetylshikonin on DNA topoisomerase-I from HeLa cells. The reaction mixture (20  $\mu$ L) contained 50 mM HEPES (pH 7.0), 10 mM MgCl<sub>2</sub>, 150 mM KCl, 0.75  $\mu$ g of DNA, 1 mM DTT, 5  $\mu$ g of BSA, and the enzyme. Where indicated, acetylshikonin of various concentrations was included. Lane 1: no enzyme, no drug. Lanes 2–4: 0.5, 1, and 2 units of the enzyme. Lanes 5–9: 2 units of the enzyme and 15, 30, 60, 120, and 240  $\mu$ g/mL of acetylshikonin.

group such as chloro resulted in about a 3-fold decrease in inhibitory potency as seen in compounds 2 and 3. Branching the n-propanoyl moiety of compound 4 by adding an α-methyl (6) also led to the diminishment of potency with respect to compound 4, but a modest enhancement with respect to compound 5. Noteworthy, the placement of double bonds in acyl group (C<sub>5</sub>-C<sub>7</sub>) contributed remarkably to augment the inhibitory action; IC50 values for n-pentanoyl-, n-hexanoyl-, and n-heptanoylshikonins were 161, 207, and 625  $\mu$ M, respectively, and those for 4-pentencyl-, trans-2-hexenoyl-, and 6-heptenoylshikonins were 40, 153, and 250 uM, respectively. The improvement of potency was more remarkable with dienes; compound 15 is more active than compound 16. The introduction of a benzoyl group (22) on 1'-OH of shikonin resulted in a slight increase in potency as compared to shikonin, but the potency decreases with a phenylacetyl (23) or a diphenylacetyl moiety (24). Our present data show that among 32 synthetic analogues, 15 compounds are more potent inhibitors than shikonin. Moreover, 11 compounds expressed a greater inhibitory action than camptothecin as a prototype DNA topoisomerase-I inhibitor. There may be two important factors for the

## Scheme 1. Synthesis of Acylshikonins

DCC; Dicyclohexylcarbodiimide DMAP; 4-Dimethylaminopyridine

R-COOH: Alkyl & Alkenyl carboxylic acid (C<sub>2</sub>-C<sub>12</sub>)
Fatty acid (C<sub>15</sub>-C<sub>18</sub>)
Retinoic acid (cis & trans)

Table 1. IC<sub>50</sub> Values<sup>a</sup> of Acylshikonin Analogues in the Inhibition of DNA Topoisomerase-I

compd. no	RCO-	$IC_{50}^b (\mu M)$	compd no.	RCO-	$IC_{50} (\mu M)$
1	acetyl	$45\pm3^c$	17	n-octanoyl	>625
2	monochloroacetyl	$137 \pm 8^c$	18	n-nonanoyl	>625
3	trichloroacetyl	$115\pm6^c$	19	n-decanoyl	>625
4	n-propanoyl	$44\pm3^c$	20	n-dodecanovl	>625
5	n-butanoyl	$144\pm10^{c}$	21	3-methyl-2-butenoyl	$77 \pm 6^{\circ}$
6	isobutanoyl	$84 \pm 5^c$		,	
7	n-pentanoyl	$161 \pm 10$	22	benzoyl	$153 \pm 11$
8	4-pentenoyl	$40\pm3^c$	23	phenylacetyl	$222 \pm 18$
9	trans-2-pentenoyl	$80 \pm 5^{c}$	24	diphenylacetyl	>625
10	n-hexanoyl	$207 \pm 15$	25	undecylenyl	>625
11	trans-2-hexenoyl	$153 \pm 11$	26	stearovl	>625
12	trans-3-hexenovl	$153\pm15$	27	palmitoyl	>625
13	2.4-hexadienovĺ	$130 \pm 10^{c}$	28	oleovl	>625
14	<i>n</i> -heptanoyl	$625\pm30$	29	linoleoyl	>625
15	2.6-heptadienovl	$126\pm6^c$	30	linolenovl	>625
16	6-heptenoyl	$250 \pm 13$	31	trans-retinovl	>625
			32	cis-retinoyl	>625
shikonin		$208\pm10$	camptothecin		$127 \pm 9$

<sup>&</sup>lt;sup>a</sup> See the Experimental Section for procedures. <sup>b</sup> IC<sub>50</sub> ( $\pm$ SEM) is the concentration required to inhibit 50% of topoisomerase-I activity. Statistical analysis was done by Student's t test. <sup>c</sup> IC<sub>50</sub> is significantly different from that of camptothecin; p < 0.05 (n = 3).

contribution of acyl moiety to the enhanced potency. One is the length of acyl moiety; acetyl- or n-propanoylshikonin is about 3 times more potent than n-butanoylshikonin. Another is the existence of olefinic double bonds in acyl chains; a similarity of  $IC_{50}$  values between 2,4-hexadienoylshikonin (13) and 2,6-heptadienoylshikonin (15) in comparison with the 3-fold difference of  $IC_{50}$  values between n-hexanoylshikonin (10) and n-heptanoylshikonin (14) reflects a considerable contribution of dienes. Although analogues bearing chain lengths longer than the heptanoyl moiety demonstrated a negligible effect, a role of these compounds as potential prodrugs may be possible in the cells containing esterases.

The chain length of acyl group for the maximal inhibition of topoisomerase-I was found to be two or three carbon atoms. These data suggest that there may be a size limitation of acyl moiety for the effective interaction of acylshikonins with topoisomerase-I and/or DNA. Although the mechanism for antitumor action of acetylshikonin was not examined here, it is assumed that the inhibition of DNA topoisomerase-I could be one of the mechanisms for its antitumor action. The support for the assumption may come from further studies on the relationship between the topoisomerase-I—inhibiting activity and the antitumor action of acylshikonins.

### **Experimental Section**

General Experimental Procedures. Melting points were measured with an Electrothermal melting point apparatus and were uncorrected. IR spectra were recorded on a JASCO Report-100 spectrophotometer, and <sup>1</sup>H-NMR spectra were obtained using a Varian-Gemini 200 NMR spectrometer; the chemical shift values were expressed as ppm relative to TMS as an internal standard. High-resolution mass spectra were obtained on VG70-VSEQ (VG analytical) under standard conditions. Elemental analyses were determined by CE-2400

(Perkin-Elmer). Analytical thin-layer chromatography (TLC) was carried out on Merck precoated silica gel 60 F-254 plates. EM Kieselgel 60 (230-400 mesh ASTM) was used for column chromatography.

Isolation of Topoisomerase-I Inhibiting Substances from the Roots of Lithospermum erythrorhizon. The airdried root (1 kg) was extracted twice with n-hexane (2 L) in a Soxhlet apparatus for 16 h at room temperature. The extract was concentrated under vacuum at 35 °C to give a dark viscous gum (9 g) after removal of the solvent. The gum was fractionated by silica gel column chromatography with a solvent system of n-hexane:dichloromethane (5:1 to 1:5), and five fractions exhibiting a significant inhibition of topoisomerase-I were obtained. Recrystallization of the second and third fractions gave crystals of acetylshikonin (250 mg) and shikonin (220 mg), respectively. The structures of those compounds were established by spectroscopic comparison with those of authentic substances. 14

Synthesis of Acylshikonins. Shikonin isolated as mentioned above was used as starting material. Shikonin (1 mmol) was dissolved in 3 mL of anhydrous dichloromethane, and to the solution were added 1.05 mmol of DCC (dicyclohexylcarbodiimide), 0.25 mmol of DMAP (4-(dimethylamino)pyridine), and 1 mmol of organic carboxylic acid (99%, Aldrich, Milwaukee, WI) under nitrogen atmosphere in the ice bath. The resulting mixture was stirred for 3 h in the ice bath and then resulting mixture was stirred for 3 h in the ice bath and then mixture do additional 1 h at room temperature. The reaction mixture was diluted with 20 mL of n-hexane and filtered. Then, the filtrate was concentrated to 4 mL in vacuo. The crude product was purified by silica gel column chromatography as described in the preceding part.

Determination of DNA Topoisomerase-I or -II Activity. DNA topoisomerase-I activity was measured by the relaxation of superhelical DNA.  $^{15,16}$  The assay mixture (20  $\mu L$ ) containing 50 mM HEPES (pH 7.0), 150 mM KCl, 10 mM MgCl<sub>2</sub>, 1.0 mM dithiothreitol, 0.05 mg/mL bovine serum albumin, 0.75  $\mu g$  of supercoiled pBR322 DNA, and 5  $\mu L$  of enzyme was used as control. The assay mixture containing test substance was the test group. Both control and test groups were incubated at 37 °C, and after 30 min of incubation, the reaction was terminated by the addition of 5  $\mu L$  of a

solution consisting of 2% SDS, 20% glycerol, and 0.05% bromophenol blue. Electrophoresis was carried out over 1% agarose gel plates, equilibrated with TBE buffer (50 mM Tris base, 50 mM boric acid and 2.5 mM EDTA) and stained with  $0.5~\mu\text{g/mL}$  ethidium bromide solution. The plates were photographed under UV light. The IC<sub>50</sub> value is expressed as the concentration which caused 50% inhibition of the enzyme activity, corresponding to 50% inhibition of relaxation of supercoiled pBR322 DNA under the conditions used. In each test, camptothecin was used as a positive control, and the IC<sub>50</sub> value of each acylshikonin analogue was compared with that of camptothecin. Each value is the mean (±SEM) of triplicate experiments. In a separate experiment, the activity of DNA topoisomerase-II from Hella cells was measured by the relaxation of supercoiled plasmid DNA. The inhibitory effect of shikonin or acetylshikonin (500  $\mu$ M) was determined using the topoisomerase-II and supercoiled pUC19 DNA as substrate under conditions established earlier.17

Purification of Topoisomerase-I. 15,16 HeLa cells, purchased from ATCC, were grown in RPMI 1640 medium containing 5% fetal bovine serum. The cells were harvested and suspended in a solution consisting of 1 mM EDTA, 1 mM mercaptoethanol, 0.5 mM PMSF, 10% glycerol, and 50 mM phosphate buffer (pH 7.0) and then homogenized with Polytron homogenizer in the ice bath for 30 s. Phosphate buffer (0.2 M) was added to the homogenized solution to a final concentration of 0.05 M. After rehomogenization of this final solution for 30 s, the solution was kept for 1 h in the ice bath. The cooled solution was centrifuged (30000g, 30 min), and then the supernatant was subjected to phosphocellulose chromatography, which gave a partially purified enzyme preparation.

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Supplementary Material Available: Physical and chemical data for synthesized acylshikonin analogues (10 pages). Ordering information is given on any current masthead page.

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