



Modified EDTA selectively recognized Cu^{2+} and its application in the disaggregation of β -amyloid-Cu (II)/Zn (II) aggregates

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ABSTRACT

The accumulation of the β -amyloid ($\text{A}\beta$) aggregates induced by $\text{Cu}^{2+}/\text{Zn}^{2+}$ in conjunction with toxicity is closely related to Alzheimer's disease (AD). Herein, we intended to improve the efficiency and selectivity of traditional chelator ethylenediaminetetraacetic acid (EDTA) combined with a fluorescent group 4-aminosalicylic acid (4-ASA) to acquire a novel potential chelator 4,4'-((2,2'-(ethane-1,2-diylbis((carboxymethyl)azanediyl))bis(acetyl))bis(azanediyl))bis(2-hydroxybenzoic acid) (**EDTA-ASA**) capable of disaggregating $\text{A}\beta$ -Cu(II)/Zn(II) aggregates. **EDTA-ASA** combines 4-ASA as fluorophore and multidentate amino nitrogen, hydroxyl and carboxyl groups to chelate Cu^{2+} from $\text{A}\beta$ -Cu (II) aggregates. The specific selectivity of **EDTA-ASA** towards Cu^{2+} in Tris-HCl buffer solution was investigated by fluorescence measurements. It exhibits high recognition towards Cu^{2+} with no significant interference of other competitive metal ions, which overcomes the deficiencies of EDTA. Importantly, the binding sites and binding mode for Cu^{2+} were clarified through DFT calculations. The thioflavin-T (ThT) fluorescence analyses and transmission electron microscopy (TEM) results have revealed **EDTA-ASA** exhibited an enhanced disaggregation capability on $\text{A}\beta$ -Cu (II)/Zn (II) aggregates in comparison to EDTA. The Cu^{2+} chelating affinity was sufficient for **EDTA-ASA** to sequester Cu^{2+} from $\text{A}\beta$ -Cu (II) aggregates.

1. Introduction

Alzheimer's disease (AD) is the most common form of dementia in the world and a devastating neurodegenerative disorder, it is incurable so far [1]. The predominant cause of AD is a heated discussion. Currently, though multiple factors mutually influence each other, the most prevailing hypothesis is still the amyloid cascade [2,3]. The amyloid cascade proposes amyloid- β ($\text{A}\beta$) aggregation as the leading cause of the disease. The misfolding of the extracellular $\text{A}\beta$ accumulated in senile plaques (SP) has been recognized as the dominant hallmark of AD [4]. $\text{A}\beta$ is a typical 39–43 residue polypeptide encompassing a C-terminal hydrophobic domain and an N-terminal hydrophilic sequence. Its interaction with Cu^{2+} , Zn^{2+} facilitates $\text{A}\beta$ aggregation, $\text{A}\beta$ misfolding and reactive oxygen species (ROS) production [5–7]. Especially,

high millimolar concentrations range of Cu^{2+} and Zn^{2+} are found in SP [8]. Up to now, the treatment of AD faces the greatest challenges in that there are few competent therapeutic approaches to modulate related metal ions and disaggregate $\text{A}\beta$ -Cu (II)/Zn (II) aggregates.

Metal chelators have been applied in AD therapy research due to their metal ion chelating ability and have been proven to be a critical approach [9–12]. In the past few decades, plenty of chelators for metal ions, especially for Cu^{2+} selective recognition, have been reported such as ethylenediaminetetraacetic acid (EDTA) [13], quinolines [14], clioquinol (CQ) [15a], 5,7-dichloro-2-((dimethylamino)methyl) 8-quinolinol (PBT-2) [16], calixarenes [17], curcumins [18], and rhodamines [19,20]. Potential regulation of Cu^{2+} -induced $\text{A}\beta$ aggregation has been shown in vitro and in vivo by using these metal chelators. Several chelators including CQ and PBT2 have been employed in murine AD

Abbreviations: AD, Alzheimer's disease; $\text{A}\beta$, Amyloid- β ; SP, Senile plaques; ROS, Reactive oxygen species; EDTA, Ethylenediaminetetraacetic acid; CQ, Clioquinol; PBT-2, 5,7-dichloro-2-((dimethylamino)methyl) 8-quinolinol; 4-ASA, 4-aminosalicylic acid; EDTA-ASA, 4,4'-((2,2'-(ethane-1,2-diylbis((carboxymethyl)azanediyl))bis(acetyl))bis(azanediyl))bis(2-hydroxybenzoic acid); PET, Photoinduced electron transfer; NMR, Nuclear magnetic resonance; ESI-MS, Electrospray ionization mass spectra; FT-IR, Fourier transform infrared; UV-vis, Ultraviolet-visible; TEM, Transmission electron microscope; EDTA-DA, 4-[2-(2,6-dioxomorpholin-4-yl) ethyl] morpholine-2,6-dione; Py, pyridine; DFT, Density functional theory; B3LYP, Becke-3-Lee-Yang-Parr; MOs, Molecular orbitals; HOMO, Highest Occupied Molecular Orbital; LOMO, Lowest Unoccupied Molecular Orbital; HFIP, 1,1,1,3,3,3-hexafluoro-2-propanol; ThT, Thioflavin T; CHEQ, Chelation-enhanced fluorescence quenching; ISC, Intersystem crossing; K_a , Binding association constant; B-H, equation Benesi-Hildebrand equation

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models and AD patients. CQ was proved to be capable of dissolving A β deposits in transgenic mice and it can slow the cognitive decline associated with AD in some cases, and PBT2 could decrease the levels of A β in AD patients. Yet, all these metal chelators have not yielded promising results. The unpredictable side effects of these metal chelators, including drug-resistance and subacute myelo-optic neuropathy, limit their universal clinical applications. Besides, most of these chelators exhibited limited application due to problems such as complicated synthetic procedures, poor water solubility, and high interference by co-existing metal ions [21–24]. Therefore, bearing the above considerations in mind, the functional chelator designs are still novel, and useful methods. It is of great significance and necessary to develop a novel metal chelator which exhibits high selectivity towards Cu²⁺ in aqueous solution via a simply synthetic method.

Interestingly, the well-known chelator, EDTA, has attracted considerable attention in recent years [25–27]. Despite its limitations in clinical studies, its EDTA backbone inspires us to functionalize it further to improve its possible properties. We have started to construct a modified EDTA derivative as a functional metal chelator. 4-aminosalicylic acid (4-ASA) has been widely utilized for the treatment of inflammatory diseases since the 1940s [28–31]. ASA conjugates of EDTA were reported as promising anti-inflammatory prodrugs [32]. The compound 4-ASA possesses a fluorescent moiety which is useful for probing metal ions. Furthermore, 4-ASA conjugates of EDTA derivative as a chelator for selective recognition of Cu²⁺ in AD therapy has not been reported [33,34]. From the above, we have herein proposed to conjugate 4-ASA to EDTA with the amide linkage (-CONH) as a bridging unit to acquire the desired chelator, which will show water solubility improvement, strong fluorescence, and be capable for specific recognition of Cu²⁺ in aqueous solution.

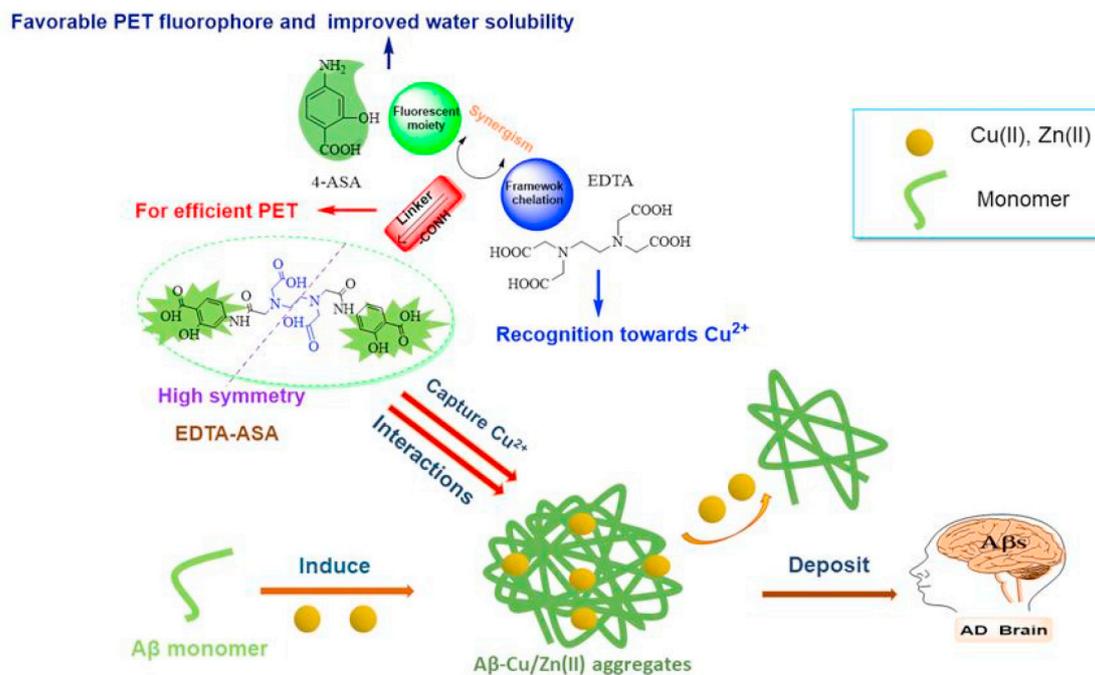
We strategically designed and synthesized a modified EDTA derivative 4,4'-((2,2'-(ethane-1,2-diylbis((carboxymethyl)azanediyl)) bis(acetyl)) bis(azanediyl))bis(2-hydroxybenzoic acid) (**EDTA-ASA**) (scheme. 1). It is based on EDTA as receptor and modified with 4-ASA as fluorophore. **EDTA-ASA** comprising both 4-ASA and EDTA-backbone group could provide excellent coordinating sites, such as electron-rich N (amino-nitrogen, -NH-) and O (hydroxyl and carboxyl groups, -OH, -COOH) atoms. Both the framework of EDTA and side cores of 4-ASA have the potential to chelate Cu²⁺. **EDTA-ASA** presents a highly

symmetrical structure, which is hoped to provide a special configuration and binding cavity for Cu²⁺. In addition, the groups of -COOH and -OH on 4-ASA could increase the solubility in aqueous solution. The -CONH extends the distance between the two functional groups thereby adjusting the effect of photoinduced electron transfer (PET) of secondary amines of 4-ASA to EDTA group. Furthermore, the -COOH and the -OH group in **EDTA-ASA** may have electrostatic interactions and hydrogen bonds with the carbonyl groups of N-terminal residues at A β . Besides the chelating interaction with Cu²⁺, **EDTA-ASA** is expected to have other synergistic effects to disaggregate the Cu (II)/Zn (II)-mediated A β aggregates. Collectively, **EDTA-ASA** is anticipated to have a wide potential application as potential chelator agents in the field of AD therapy.

2. Experimental section

2.1. Materials and methods

All chemical reagents (analytic grade or molecular biology grade) purchased commercially were available and can be used as is unless otherwise specified. The solutions of metal cations were performed from their corresponding salts, such as LiCl, NaCl, KCl, MgCl₂·6H₂O, CaCl₂·4H₂O, AlCl₃·6H₂O, BaCl₂, ZnCl₂, AgNO₃, MnCl₂·4H₂O, Pb(NO₃)₂, CrCl₃·6H₂O, CoCl₂·6H₂O, NiCl₂·6H₂O, HgCl₂, and CuCl₂·3H₂O. For the investigation of anions on the effect of recognition, the different water-soluble chloride, sulphate, nitrate, bromide, perchlorate, and acetate salts of copper (CuCl₂, CuSO₄, Cu(NO₃)₂, CuBr₂, Cu(ClO₄)₂, and Cu(OAc)₂) were used. The water used in this work is ultrapure water (18.2 M Ω cm) deionized by the Milli-Q system. A β ₄₀ is the most predominant forms of A β in vivo. A β ₄₀ peptide (free acid terminal) was commercially purchased from Sangon Biotech (Shanghai, China) and kept at -20 °C. The sequence of amino acid for the A β ₄₀ is DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVV. The nuclear magnetic resonance (NMR) spectra were employed on Bruker AVANCE III HD 400 in D₂O. Electrospray ionization mass spectra (ESI-MS) were determined on a Shimadzu, Japan LCMS-2020 Liquid Chromatograph-Mass Spectrometer using methanol as a solvent. The Fourier transform infrared (FT-IR) spectra were determined ranging from 4000 to 400 cm⁻¹ on a THEMOR-FILSHER iS50R instrument using KBr pellets.



Scheme 1. Schematic illustration of the design strategy of EDTA-ASA and its disaggregation in A β -Cu (II)/ Zn (II) aggregates.

Ultraviolet-visible (UV-vis) spectra were measured on PerkinElmer Lambda 950 spectrophotometer. Fluorescence spectra were recorded on a HORIBA Jobin Yvon FluoroMax-4 Fluorescence Spectrometer. Both absorption and emission spectra were acquired at ambient temperature. All tests were performed in parallel three times, and data processing were expressed as the mean and standard deviation. Transmission electron microscope (TEM) analysis was performed on the Shimadzu, Japan HT7700.

2.2. Synthesis of 4,4'-((2,2'-(ethane-1,2-diylbis((carboxymethyl)azanediyl)) bis(acetyl))bis(azanediyl))bis(2-hydroxybenzoic acid) (EDTA-ASA)

The 4-[2-(2,6-dioxomorpholin-4-yl) ethyl] morpholine-2,6-dione (EDTA-DA) (2.56 g (0.01 mol)), and 4-ASA (3.06 g (0.02 mol)) were combined in 20 mL DMF under nitrogen atmosphere. 8 mL of pyridine (Py) was sequentially added dropwise to the mixed solution. The solid was completely dissolved for 15 min, and the reaction was stirred at 30 °C for 24 h. After the reaction was completed, the reaction mixture was poured into CHCl_3 to give a white precipitate. The obtained product was collected by filtration under vacuum and washed with anhydrous ethanol. After vacuum drying at 60 °C for 5 h, a white powder was obtained. This white powder was dissolved in an appropriate amount of NaOH and then was followed by acidification by adding 0.1 M HCl dropwise. The pure white precipitate of EDTA-ASA was finally obtained by vacuum filtration in a yield of 92%.

2.3. UV-vis spectra and fluorescence measurements of EDTA-ASA with metal ions

The metal ions stock solution was prepared in ultrapure water using the corresponding metal salts. This stock solution of EDTA-ASA was prepared and diluted to desired concentration with Tris-HCl buffer solution (10 mM, pH 7.4). Both UV-vis spectrum and fluorescence emission spectrum were performed according to the following procedures. In the UV-visible experiment, the EDTA-ASA solution was placed in a quartz cell with an optical path of 1.0 cm to achieve an absorption spectrum. For emission experiment, the solution of EDTA-ASA 2000 μL was filled in a quartz cell with 1.0 cm optical path, the solution of different metal ions was added and gradually mixed with a micropipette for 3 min. The fluorescence titration experiment was measured by mixing EDTA-ASA with a different molar ratio of Cu^{2+} solution.

2.4. Computational details

Density functional theory (DFT) theoretical calculation was employed to investigate the binding mode between EDTA-ASA and Cu^{2+} , based on the 6-31G* in conjunction with Becke-3-Lee-Yang-Parr (B3LYP) basis set [35–37]. The 6-31G* basis set was selected for non-metallic elements such as the C, H, O, and N atoms; the SDD basis set was for the Cu atom. The structures of EDTA-ASA and EDTA-ASA-Cu complex were optimized with the DFT calculations. Vibrational frequencies of the structure have been analyzed to confirm the minima optimized structures. All theoretical calculations were accomplished on the Gaussian 09 programs. The molecular orbitals (MOs) of EDTA-ASA and EDTA-ASA-Cu were visualized and plotted with the Gauss-View program.

2.5. HFIP treatment of $\text{A}\beta_{40}$

The initial powder of $\text{A}\beta_{40}$ was dissolved in cooled 1,1,1,3,3,3-hexafluoro-2-propanol (HFIP) with constant agitation for 6 h to obtain a homogeneous $\text{A}\beta_{40}$ monomeric solution. The solution was blowing with argon gas stream to evaporate the HFIP. The obtained thin white films of the $\text{A}\beta_{40}$ monomer were stored at -20 °C. Monomeric films were

dissolved in DMSO with agitation to acquire a final concentration of $1.0 \text{ mg}\cdot\text{mL}^{-1}$. The stock solution of $\text{A}\beta_{40}$ was diluted with Tris-HCl buffer solution (10 mM, pH 7.4) to desired concentrations for the following experimental.

2.6. Thioflavin T (ThT) fluorescence study

ThT fluorescence investigations were employed with a fluorescence spectrometer. In the inhibition studies, freshly prepared $\text{A}\beta_{40}$ (25 μM) in the presence or absence of Cu (II) or Zn (II) (25 μM) was mixed with or without EDTA and EDTA-ASA (25 μM) and incubated at 37 °C with constant agitation for 24 h. In the disaggregation studies, metal-free and metal treated $\text{A}\beta_{40}$ (25 μM) aggregates were generated by incubating mixtures of freshly prepared $\text{A}\beta_{40}$ (25 μM) in the presence or absence of Cu (II) or Zn(II) (25 μM) at 37 °C with agitation. After 24 h, the samples were treated with EDTA and EDTA-ASA (25 μM) and incubated for another 24 h. After incubation, all of the samples were treated with ThT for the following ThT studies ($\lambda_{\text{ex}} = 404 \text{ nm}$, $\lambda_{\text{em}} = 487 \text{ nm}$).

2.7. Transmission electron microscopy (TEM)

Samples for TEM were prepared by the following steps. The glow-discharged grid (Formar/Carbon 300-mesh, Electron Microscopy Sciences) was treated with the corresponding $\text{A}\beta_{40}$, and $\text{A}\beta_{40}$ -Cu (II)/Zn (II) aggregates samples (25 μM , 5 μL) for 2 min at ambient temperature. Excess samples were removed with filter paper and then washed twice with ultrapure water. Each grid dried for 30 min at room temperature. Images from each sample were captured by a Shimadzu, Japan HT7700 (40–120 kV, Image rotation: maximum range X1000-X40000 magnification).

3. Results and discussion

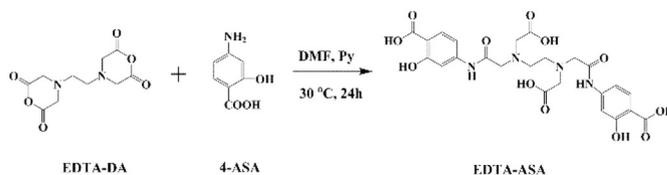
3.1. Synthesis and general characterization of EDTA-ASA

EDTA-ASA was synthesized in 92% yield via one-step reaction from the starting materials EDTA-DA and 4-ASA respectively (as illustrated in Scheme 2). The structure of EDTA-ASA was well characterized by ESI-MS, FT-IR, ^1H NMR, and ^{13}C NMR. All data match well with the corresponding structure.

3.2. The spectroscopic studies of EDTA-ASA towards Cu^{2+} recognition

In the UV-vis spectra, EDTA-ASA showed two main absorption bands located at 260 nm and 302 nm in Tris-HCl buffer solution (Fig. S5, ESI †). The UV-vis spectrum of 4-ASA exhibited a similar major absorption band at 263 nm and 298 nm in Tris-HCl buffer solution (Fig. S6, ESI †). The maximum absorbance band located at 260 nm corresponds to benzene ring π - π^* transition and the band centered at 302 nm is attributed to the charge transition from benzene ring to an amino group. Therefore, we choose the band centered at 260 nm as the excitation wavelength in the subsequent fluorescent studies.

To investigate the selectivity of EDTA-ASA, the tested metal ions including Cu^{2+} , Zn^{2+} , Li^+ , Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Al^{3+} , Ba^{2+} , Ag^+ , Mn^{2+} , Pb^{2+} , Cr^{3+} , Co^{2+} , Ni^{2+} and Hg^{2+} were employed in our study. Fig. 1 shows the fluorescence emission spectra of EDTA-ASA in the



Scheme 2. The synthesis procedure of EDTA-ASA

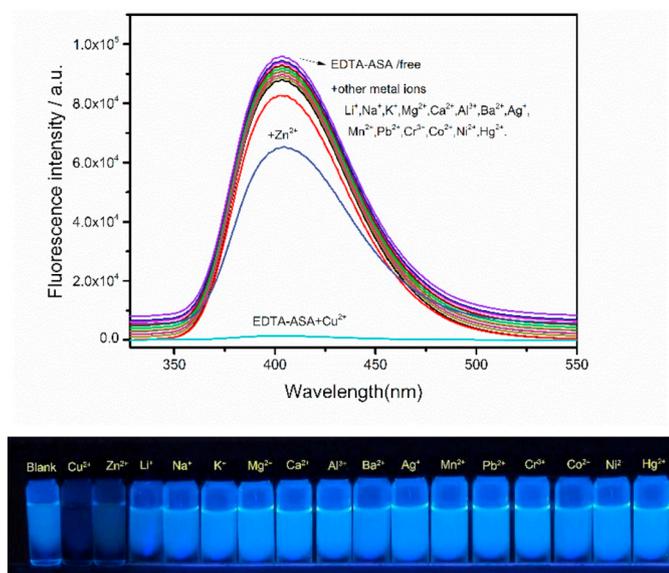


Fig. 1. Fluorescence spectrum of EDTA-ASA (10 μ M) on addition of different metal ions (10 μ M) of Cu^{2+} , Zn^{2+} , Li^+ , Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Al^{3+} , Ba^{2+} , Ag^+ , Mn^{2+} , Pb^{2+} , Cr^{3+} , Co^{2+} , Ni^{2+} and Hg^{2+} in Tris-HCl buffer solution (10 mM, pH 7.4). Image: Selective fluorescent response of EDTA-ASA in the presence of metal ions (under UV light).

presence of 1.0 equal different metal ions. Collectively, EDTA-ASA alone had relatively strong fluorescence centered at near 403 nm in Tris-HCl buffer solution. Upon addition with 1.0 equal Cu^{2+} , while, the fluorescence at 403 nm was almost wholly quenched. The fluorescence changed from “on” to “off.” In contrast, the other metal ions including Zn^{2+} failed to show any dramatic fluorescence change in fluorescence behavior of EDTA-ASA. The UV-vis spectra of EDTA-ASA on addition of different metal ions in Tris-HCl buffer solution were also employed (Fig. S7, ESI[†]). For comparison purpose, we also investigated the fluorescence properties of control compound 4-ASA in Tris-HCl buffer solution. A negligible response of 4-ASA was observed towards all the tested metal ions (Fig. S8, ESI[†]). Therefore, the results confirmed that EDTA-ASA exhibited good selectivity and displayed a dynamic “on-off” behavior towards Cu^{2+} in Tris-HCl buffer solution.

Notably, the efficient quenching of fluorescence indicated that EDTA-ASA showed a specific response to Cu^{2+} . Before coordination with Cu^{2+} , EDTA-ASA have strong fluorescence. However, when EDTA-ASA was coordinated with Cu^{2+} , the fluorescence is quenched, which is probably due to the formation of a non-fluorescent complex EDTA-ASA-Cu. This “on-off” switching could be explained with chelation-enhanced fluorescence quenching (CHEQ) effect and PET-enhanced process [38–40]. These metal ions differ too much in many ways such as the orbital shape, electron density, size of the metal ion, and they can establish different coordination interactions. The metal ions that failed to cause a significant change in fluorescence intensity may be due to the inappropriate coordination conformation and unsuitable ion radius. This phenomenon is following the case of d^9 Cu (II), it is the highest in the Irving-Williams series [41]. Moreover, due to the Jahn-Teller effect, the complex EDTA-ASA-Cu has higher stability [42]. Furthermore, Cu^{2+} is paramagnetic with an empty shell, the intrinsic paramagnetic nature of Cu^{2+} from spin-orbit coupling could actively quench the emission of a nearby fluorophore via non-radiative intersystem crossing (ISC) transition process [43,44].

The fluorescence titration investigations were used to explore the response properties of EDTA-ASA to Cu^{2+} . As is presented in Fig. 2(a), EDTA-ASA displayed strong fluorescence at 403 nm. Upon adding various molar of Cu^{2+} (0–1 equal) into EDTA-ASA, the fluorescence intensity gradually decreased, and the fluorescence intensity nearly remains unchanged even 1.0 equal Cu^{2+} was added. The results

indicated the formation of 1:1 chelation ratio of EDTA-ASA and Cu^{2+} . The 1:1 binding mode between EDTA-ASA and Cu^{2+} was also established using Job's plot methods. The fluorescence emission intensity got a minimum when the molar ratio fraction was 0.5, which further verified a 1:1 stoichiometry of EDTA-ASA and Cu^{2+} . Cu^{2+} may be chelated at the N (-NH-, amino-nitrogen) and O (-COOH, carboxyl group) atoms of EDTA-ASA to satisfy the need for saturated 1:1 coordination by forming stable EDTA-ASA-Cu complex. Theoretical calculations based on DFT principles were employed to better understand the phenomenon of fluorescence quenching and the chelation interaction between EDTA-ASA and Cu^{2+} in the following discussions.

The binding association constant (K_a) is used to evaluate the stability of interaction between the chelator and metal ions together in a solution. We adopted the Benesi-Hildebrand eq. (B–H equation) to determine the K_a of EDTA-ASA and Cu^{2+} in Tris-HCl buffer solution [45]. There is a great linear relationship with the R^2 value of 0.9984 in Fig. 2(b). The K_a value was counted as $6.852 \times 10^7 \text{ M}^{-1}$ for EDTA-ASA-Cu complex, indicating a robust binding association between EDTA-ASA and Cu^{2+} . Moreover, the EDTA-ASA-Zn complex showed the same 1:1 stoichiometry, and the K_a was calculated to be $9.765 \times 10^5 \text{ M}^{-1}$, which is far below that of EDTA-ASA-Cu complex (Fig. S9–S11, ESI[†]).

Competition experiments study further assessed the specificity of EDTA-ASA for Cu^{2+} . As shown in Fig. 3(a), there was almost no significant decrease in fluorescence intensity when EDTA-ASA was treated with 1 equal of other metal ions (Zn^{2+} , Li^+ , Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Al^{3+} , Ba^{2+} , Ag^+ , Mn^{2+} , Pb^{2+} , Cr^{3+} , Co^{2+} , Ni^{2+} , and Hg^{2+}). However, upon addition of another 1 equal Cu^{2+} to the mixed solution of EDTA-ASA with other metal ions, an apparent fluorescence quenching was noticed. It was apparent that the quenching of fluorescence aroused by the mixture of Cu^{2+} with other metal ions was almost the same as that induced by Cu^{2+} only. As shown in Fig. 3(b), after EDTA-ASA was treated with of 1 equiv. Cu^{2+} , other excess 10 equiv. of competitive metal ions were added, there is almost no interference in fluorescence signal. Therefore, these consequences demonstrated that other common competitive metal ions do not significantly influence the recognition of EDTA-ASA for Cu^{2+} , which confirms that EDTA-ASA behaves high selectivity for Cu^{2+} in Tris-HCl buffer solution.

The fluorescence emission spectra of EDTA-ASA with different counter anions (SO_4^{2-} , Cl^- , NO_3^- , Br^- , ClO_4^- , and OAc^-) was investigated to determine the influence of interference anions on the recognition efficiency. This result illustrated that these anions had almost no interference with the recognition of Cu^{2+} (Fig. S12, ESI[†]). It is the chelation of Cu^{2+} and EDTA-ASA that leads to the efficient fluorescence quenching of EDTA-ASA, these anions exhibit no distinct effect on the recognition selectivity.

The reversibility of the fluorescence response process of EDTA-ASA towards Cu^{2+} was also explored with EDTA. After addition of excess EDTA to the EDTA-ASA and Cu^{2+} mixed solution, the fluorescence returned to the original fluorescence of EDTA-ASA, which strongly reveals that the Cu^{2+} recognition is a reversible chelation process (Fig. S13, S14, ESI[†]). These results may be connected with the different K_a between the chelators and Cu^{2+} . Though its binding affinity of Cu^{2+} with EDTA-ASA (EDTA-ASA-Cu, $K_a = 6.852 \times 10^7 \text{ M}^{-1}$, namely $\lg K_a = 7.84$) is not as good as EDTA (EDTA-Cu, namely $\lg K_a = 18.7$), it exhibits a high selectivity towards Cu^{2+} with much higher efficiency than EDTA.

It is well known that the stability of the metal chelator is of significance for further practical application. Thus, the interrelationship between the fluorescence of EDTA-ASA and pH variation was investigated to confirm the optimum pH range in application. When pH is in the range of 2–4, the fluorescence intensity of the fluorescence spectrum is nearly zero (Fig. S15, ESI[†]). This is probably due to the weak solubility of EDTA-ASA in Tris-HCl buffer solution. The fluorescence intensity of EDTA-ASA increased moderately with the change of pH from 4 to 7 and maintained relative stability in the pH range of

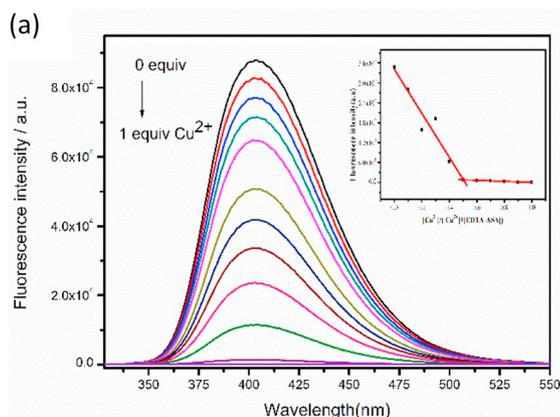
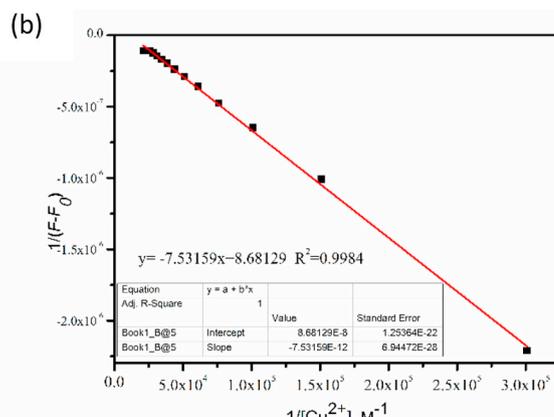


Fig. 2. (a) Fluorescence spectra of EDTA-ASA with addition of various concentrations of Cu^{2+} (0–10 equiv, the total concentration of EDTA-ASA and Cu^{2+} was $10 \mu\text{M}$) in Tris-HCl buffer solution (10 mM , $\text{pH } 7.4$). Inset: the drawing is Job's plot of the complexation between the EDTA-ASA and Cu^{2+} . (b) Benesi-Hildebrand plot of EDTA-ASA-Cu complex in Tris-HCl buffer solution (10 mM , $\text{pH } 7.4$).



7–12, which is ascribed to the protonation and deprotonation of EDTA-ASA with multiple hydroxyl and amino groups.

Furthermore, the influence of the time on the fluorescence intensity of EDTA-ASA over the range from 0 to 72 h in Tris-HCl buffer solution was also evaluated. The results reveal that the fluorescence intensity barely changes in a 10% range within a period of time (Fig. S16, ESI†). Namely, the aqueous solution of EDTA-ASA and its fluorescence properties are relatively stable, which means it may overcome the physiological fluctuations and maintain expected performances.

3.3. Theoretical calculations

The outstanding performances of EDTA-ASA in the recognition of Cu^{2+} drive us to explore its binding mode. Based on the results of fluorescence titration experiments and Job's plot, we hereby got the optimized energy-minimized structures of EDTA-ASA and EDTA-ASA-Cu by DFT optimization simulation calculations (as shown in Fig. 4). Fig. S17, and Fig. S18 ESI† illustrated the energy optimization curves under the best binding mode between EDTA-ASA and Cu^{2+} . The optimized structure of EDTA-ASA showed that it had a symmetrical plane containing the fluorophore and the receptor. The Cu^{2+} coordination changes the conformational states of EDTA-ASA. The structure of EDTA-ASA-Cu displays that the Cu^{2+} binds to EDTA-ASA at the amino nitrogen atoms and the carboxyl groups of core framework EDTA through four coordination sites. And, the Cu – O bond lengths are 2.03991 \AA and 2.13863 \AA , the Cu – N bond lengths are 2.03111 \AA and 2.03989 \AA , respectively, which means a strong coordination bond interaction between EDTA-ASA and Cu^{2+} . The coordination distances are suitable for Cu^{2+} binding and utilized exclusively for Cu^{2+} . These data indicated that the structure of EDTA-ASA could effectively provide

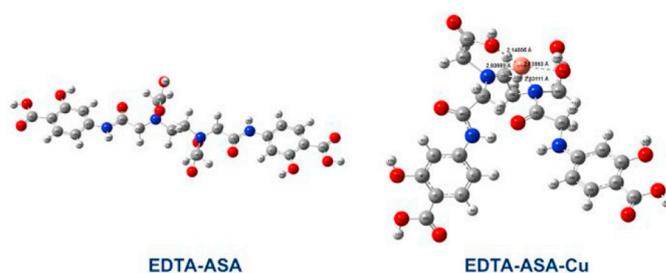


Fig. 4. Optimized energy-minimized structure of EDTA-ASA and EDTA-ASA-Cu (C, N, O, H, and Cu atom are represented as grey, blue, red, white-grey, and orange respectively).

appropriate space to accommodate the Cu^{2+} and a cavity-like structure was generated to tether Cu^{2+} successfully.

In addition to the localization of the MOs (molecular orbital), E_{HOMO} , E_{LUMO} , and the HOMO (the highest occupied molecular orbital)-LUMO (the lowest unoccupied molecular orbital) band gap ($\Delta E = E_{\text{LUMO}} - E_{\text{HOMO}}$) are commonly used parameters to interpret the electronic properties and chelation behavior. Therefore, the orbital energies and the spatial electronic cloud distributions of the HOMO and the LUMO of EDTA-ASA and EDTA-ASA-Cu complexes were shown in Fig. 5. The HOMO of EDTA-ASA is primarily delocalized on the fluorophore 4-ASA part, the LUMO is the same as the distribution of HOMO. There is no electron transfer, and the PET process is prohibited, EDTA-ASA showed fluorescence. However, when EDTA-ASA is converted to EDTA-ASA-Cu after coordinating with Cu^{2+} , the HOMO of EDTA-ASA-Cu is mostly delocalized on the Cu^{2+} and nearby EDTA part, while the LUMO is distributed on the fluorophore 4-ASA part. Under this

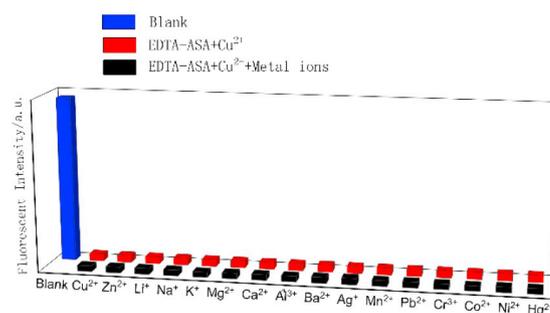
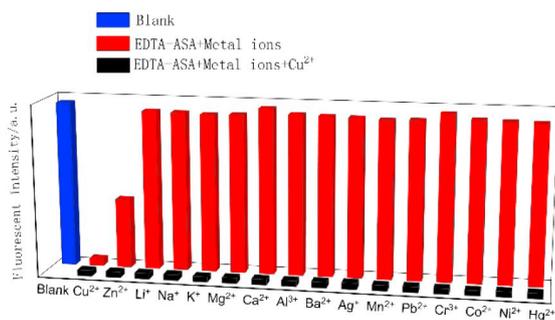


Fig. 3. (a) The fluorescence intensity at 403 nm of EDTA-ASA ($10 \mu\text{M}$) with metal ions in Tris-HCl buffer solution (10 mM , $\text{pH } 7.4$) and then with addition of equal molar of Cu^{2+} . (b) the fluorescence intensity at 403 nm of EDTA-ASA ($10 \mu\text{M}$) with the addition of equal molar of Cu^{2+} and then addition of excess 10 equiv. of other metal ions in Tris-HCl buffer solution (10 mM , $\text{pH } 7.4$).

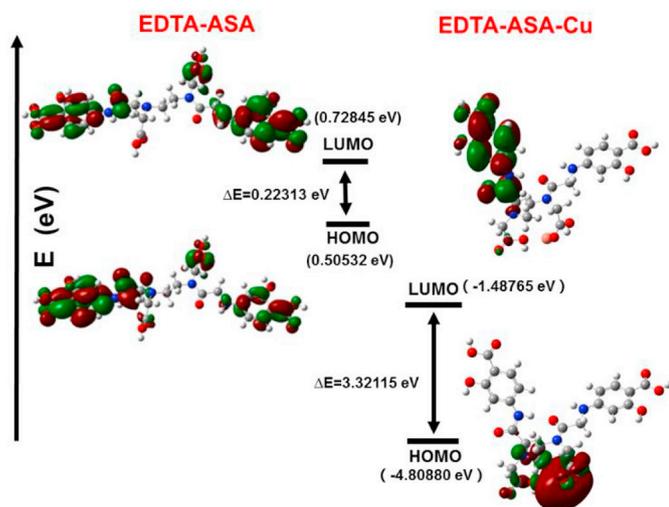


Fig. 5. Optimized frontier molecular orbital profiles for EDTA-ASA and its complex EDTA-ASA-Cu based on DFT (B3LYP/6-31G* set) calculation.

circumstance, this corresponds to the electron cloud distribution of the PET-enhanced process between the receptor and the fluorophore 4-ASA.

The ΔE between HOMO and LUMO of EDTA-ASA and EDTA-ASA-Cu was calculated as 0.22313 eV and 3.32115 eV, respectively. The ΔE between HOMO and LUMO of the EDTA-ASA-Cu complex had increased 3.09802 eV as compared to that of EDTA-ASA. Moreover, the geometry energy of EDTA-ASA-Cu was lower than EDTA-ASA (Fig. S17, Fig. S18, Table 1, ESI[†]). Therefore, the conversion of EDTA-ASA to EDTA-ASA-Cu becomes more accessible to form a stable EDTA-ASA-Cu complex by broadening the energy gap between HOMO and LUMO, which is also in accordance with Fang, Kim, Anand, and Hou's researches [46–49].

3.4. Potential application of EDTA-ASA in disaggregating A β ₄₀-Cu (II)/Zn (II) aggregates

The ThT fluorescence study was explored to evaluate the ability of EDTA-ASA to inhibit and disaggregate Cu²⁺/Zn²⁺-induced A β ₄₀ aggregates (A β ₄₀-Cu (II)/Zn (II) aggregates). ThT is a benzothiazole fluorescent dye with a high affinity for β -sheet amyloid fibers, and it can be utilized to quantify the β -sheet content of A β aggregates [50]. As is clearly shown in Fig. S19–S21 ESI[†], there's no fluorescence signal of the ThT in the fresh A β ₄₀ solution. The solution of self-aggregated A β ₄₀ sample only developed a limited increase in fluorescence intensity. In contrast, in the presence of Cu²⁺/Zn²⁺, there is a significant increase of approximately more than 4-fold in fluorescence signal, compared with the self-aggregated A β ₄₀ aggregates, which suggest that Cu²⁺/Zn²⁺ could induce the aggregation of A β ₄₀ and the generation of the β -sheet structure. Although precise interaction mechanism of the Cu²⁺/Zn²⁺ with A β ₄₀ is still unclear, it is generally conjectured that these may result from the electrostatic and chelating interactions between Cu²⁺/

Table 1

The disaggregation results of EDTA-ASA and EDTA on corresponding A β ₄₀ and A β ₄₀-Cu (II)/Zn (II) aggregates.

Samples	DR ^a (%)	
	EDTA	EDTA-ASA
A β ₄₀ aggregates	19.60 ± 0.23	44.20 ± 0.31
A β ₄₀ -Cu (II) aggregates	26.22 ± 0.21	62.20 ± 0.36
A β ₄₀ - Zn (II) aggregates	8.20 ± 0.27	32.30 ± 0.33

DR^a stands for disaggregation rate.

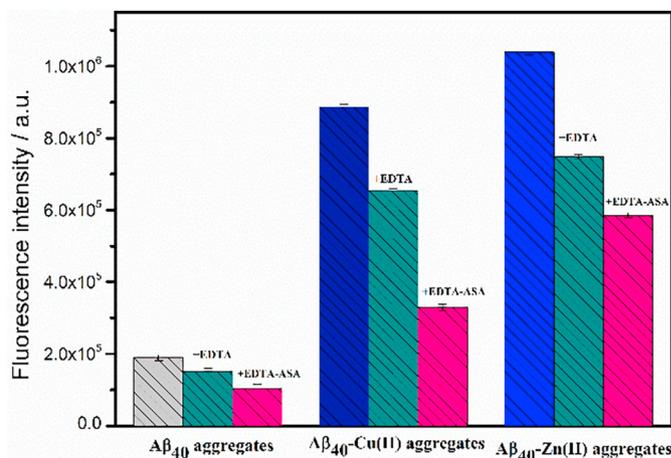


Fig. 6. The disaggregation of EDTA-ASA (Pink) and EDTA (Cyan) on corresponding A β ₄₀ and A β ₄₀-Cu (II)/Zn (II) aggregates in Tris-HCl buffer solution (10 mM, pH 7.4, λ_{ex} 440 nm, λ_{em} 487 nm), were monitored by ThT fluorescence assay.

Zn²⁺ and A β ₄₀. It is generally assumed that the Cu²⁺ can coordinate with A β ₄₀ at N-terminal residues, such as His6, His13, His14, and thereby promote the aggregation of A β ₄₀. Likewise, A β ₄₀ could form a 1: 1 complex with Zn²⁺ [4,5,51]. When EDTA-ASA was added to the sample of A β ₄₀-Cu (II)/Zn (II) aggregates, a decline in fluorescence was observed, which indicated that EDTA-ASA could inhibit the Cu²⁺/Zn²⁺-induced A β ₄₀ aggregation. Moreover, EDTA-ASA also disaggregate A β ₄₀-Cu (II)/Zn (II) aggregates. It is worth noted that the ability of EDTA-ASA to depolymerize A β ₄₀-Cu (II)/Zn (II) aggregates is much stronger than the ability to inhibit aggregation, so EDTA-ASA was evaluated for its disaggregation capacity on self-induced A β ₄₀, and Cu²⁺/Zn²⁺-induced A β ₄₀ aggregates in subsequent investigations.

As shown in Fig. 6, the depolymerization rate of EDTA-ASA for A β ₄₀-Cu (II) aggregates was 62.20%, which is 1.37-fold more effective than EDTA. EDTA, a natural product that is known to disaggregate A β -M (II) aggregates, was used as the reference compound [52]. We found that EDTA disaggregate A β ₄₀-Cu (II) aggregates by a percentage similar to previous reports. (26.22% vs 24.50% disaggregation). EDTA-ASA was more effective at dissolving A β ₄₀-Cu (II) aggregates compared with the control EDTA. Moreover, the consequence of EDTA-ASA to depolymerize in A β ₄₀-Zn (II) aggregates is not as good as that of A β ₄₀-Cu (II) aggregates. The ThT disaggregation results of EDTA-ASA and EDTA on the corresponding A β ₄₀ aggregates are listed clearly in Table 1.

TEM was exerted to directly study the morphology of corresponding A β ₄₀ aggregates and visualize the disassembly effect of EDTA-ASA. As shown in Fig. 7, regular linear fibers morphology and little-fibrillary aggregates were detected in the samples of the self-aggregated A β ₄₀, Cu²⁺-induced A β ₄₀ formed a denser aggregate than that of self-aggregated A β ₄₀, larger aggregates were observed in Zn²⁺-induced A β ₄₀ aggregates. In the presence of EDTA-ASA, the large A β ₄₀-Cu (II) aggregates were broken down into much smaller amorphous aggregates compared with the EDTA. The morphology of A β ₄₀-Cu (II) aggregates became smaller, and the structure was looser. All these findings are consistent with the above ThT fluorescent analysis. Consequently, it is assumed that the collapse of the A β ₄₀-Cu (II) aggregates is mainly due to the capability of EDTA-ASA to chelate Cu²⁺, and thus loosening the interactions of Cu²⁺ with the A β ₄₀. At pH 7.4, the logarithm of the K_a of A β ₄₀ for Cu²⁺ (lgK_a [Cu²⁺-A β] = 5.80) is 5.80 [53]. This value is much lower compared with the affinity of EDTA-ASA for Cu²⁺ that has been obtained in the above discussions, (lgK_a [Cu²⁺-EDTA-ASA] = 7.84). Indeed, the affinity of EDTA-ASA to Cu²⁺ is greater than that of A β ₄₀ (lgK_a [Cu²⁺-A β] = 5.80, lgK_a [Cu²⁺-EDTA-ASA] = 7.84). Consequently, EDTA-ASA can extract Cu²⁺ complexed by A β ₄₀ aggregates, resulting in the disassembly of the A β ₄₀-Cu (II) aggregates. Also, the

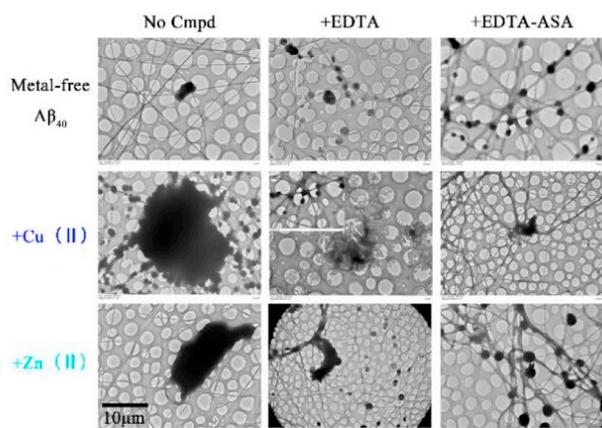


Fig. 7. Morphologies of the $A\beta_{40}$ aggregates after treatment with EDTA and EDTA-ASA as observed using TEM (scale bar 10 μm).

disaggregation ability of EDTA-ASA to $A\beta_{40}$ -Cu (II) aggregates depends not only on the binding affinity of the EDTA-ASA to Cu^{2+} . The interactions of the electrostatic, hydrophilic, and hydrogen bonds of the EDTA-ASA with $A\beta_{40}$ aggregates are also possible [54,55], since the EDTA-ASA also disassembled $A\beta_{40}$ aggregates containing no Cu^{2+} / Zn^{2+} . In summary, EDTA-ASA has the potential to regulate dissociation of $A\beta_{40}$ -Cu (II) aggregates perhaps by capturing aggregates containing Cu^{2+} and interacting with $A\beta_{40}$ aggregates.

4. Conclusion

In conclusion, the high selectivity for Cu^{2+} and disaggregation of $A\beta_{40}$ -Cu (II)/Zn (II) aggregates have been achieved by developing a novel metal chelator EDTA-ASA based on EDTA as receptor and modified with 4-ASA as fluorophore. The EDTA-ASA exhibited exclusive selectivity towards Cu^{2+} and was not affected by other competitive metal ions in Tris-HCl buffer solution. Besides, EDTA-ASA displayed a dynamic “On-Off” response to Cu^{2+} owing to the CHEQ and PET process. The Job plot indicated that EDTA-ASA bound with Cu^{2+} in a 1:1 stoichiometry and the K_a was $6.852 \times 10^7 \text{ M}^{-1}$. DFT calculations provided an optimized structure of EDTA-ASA-Cu and further confirmed the 1:1 binding mode between EDTA-ASA and Cu^{2+} in a cavity-like structure. Thus, it can capture Cu^{2+} from $A\beta_{40}$ -Cu (II) aggregates effectively. The findings indicate that this simple, practicable, designed chelator EDTA-ASA has good potential for the recognition for Cu^{2+} in disaggregating $A\beta_{40}$ -Cu (II) aggregates. All these investigations propose a novel strategy for chelator design by introducing additional function group and open an avenue for promising exploration of more AD treatment chelators.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jinorgbio.2019.110929>.

References

- [1] Alzheimer's Dement. 13 (2017) 325–373.
- [2] J.A. Hardy, G.A. Higgins, Science 256 (1992) 184–186.
- [3] F. Panza, M. Lozupone, D. Seripa, B.P. Imbimbo, Ann Neurol. 85 (2019) 303–315.
- [4] J. Hardy, D. Allsop, Trends Pharmacol sci. 12 (1991) 383–388.
- [5] G.F. Chen, T.H. Xu, Y. Yan, Y.R. Zhou, Y. Jiang, K. Melcher, H.E. Xu, Acta Pharmacol Sin. 38 (2017) 1205–1235.
- [6] V. Lanza, D. Milardi, G. Di Natale, G. Pappalardo, Curr Med Chem. 25 (2018) 525–539.
- [7] J.A. Duce, A.I. Bush, Prog Neurobiol. 92 (2010) 1–18.
- [8] S. Jones, X. Zhang, D.W. Parsons, J.C.H. Lin, R.J. Leary, P. Angenendt, P. Mankoo, H. Carter, H. Kamiyama, A. Jimeno, S.M. Hong, B. Fu, M.T. Lin, E.S. Calhoun, M. Kamiyama, K. Walter, T. Nikolskaya, Y. Nikolsky, J. Hartigan, D.R. Smith, M. Hidalgo, S.D. Leach, A.P. Klein, E.M. Jaffe, M. Goggins, A. Maitra, C. Iacobuzio-Donahue, J.R. Eshleman, S.E. Kern, R.H. Hruban, R. Karchin, N. Papadopoulos, G. Parmigiani, B. Vogelstein, V.E. Velculescu, K.W. Kinzler, Science. 321 (2008) 1801–1806.
- [9] A. Gnjec, J.A. Fonte, C. Atwood, R.N. Martins, Front Biosci. 7 (2002) 1016–1023.
- [10] H. Zheng, L.M. Weiner, O. Bar-Am, S. Epsztejn, Z.I. Cabantchik, A. Warshawsky, M.B.H. Youdim, M. Fridkin, Bioorg Med Chem. 13 (2005) 773–783.
- [11] H. Zheng, M.B.H. Youdim, M. Fridkin, J. Med. Chem. 52 (2009) 4095–4098.
- [12] P.A. Adlard, A.I. Bush, J. Alzheimers Dis. 62 (2018) 1369–1379.
- [13] W. Huang, W. Wei, Z. Shen, RSC Adv. 4 (2014) 52088–52099.
- [14] K. Kaur, M. Jain, R.P. Reddy, R. Jain, Eur J Med Chem. 45 (2010) 3245–3264.
- [15] (a) M. Di Vaira, C. Bazzicalupi, P. Orioli, L. Messori, B. Bruni, P. Zatta, Inorg Chem. 43 (2004) 3795–3797; (b) M. Wehbe, A.K. Malhotra, M. Anantha, C. Lo, W.H. Dragowska, N. Dos Santos, M.B. Bally, Drug Deliv Transl Re. 8 (2018) 239–251; (c) C.W. Ritchie, A.I. Bush, A. Mackinnon, S. Macfarlane, M. Mastwyk, L. MacGregor, L. Kiers, R. Cherny, Q.X. Li, A. Tammer, D. Carrington, C. Mavros, I. Volitakis, M. Xilinas, D. Ames, S. Davis, K. Beyreuther, R.E. Tanzi, C.L. Masters, Arch. Neurol. 60 (2003) 1685–1691.
- [16] (a) L. Nguyen, J. Vendier, B. Stigliani, Meunier and A. Robert, Eur J, Inorg Chem. 2017 (2017) 600–608; (b) L. Lannfelt, K. Blennow, H. Zetterberg, S. Batsman, D. Ames, J. Harrison, C.L. Masters, S. Targum, A.I. Bush, R. Murdoch, J. Wilson, C.W. Ritchie, Lancet Neurol. 7 (2008) 779–786.
- [17] L. Lannfelt, K. Blennow, H. Zetterberg, S. Batsman, D. Ames, J. Harrison, C.W. Ritchie, Lancet Neurol. 7 (2008) 779–786.
- [18] V. Guérineau, M. Rollet, S. Viel, B. Lepoittevin, L. Costa, P. Saint-Aguet, R. Laurent, P. Roger, D. Gimes, C. Martini, V. Huc, Nat Commun. 10 (2019) 113.
- [19] (a) S. Kimura, A. Kiriya, K. Araki, M. Yoshizumi, D. Inoue, T. Furubayashi, R. Yutani, R. Teraoka, A. Tanaka, K. Kusamori, H. Katsumi, A. Yamamoto, K. Iga, T. Sakane, Eur J Pharm Biopharm. 122 (2018) 1–5; (b) Y. Chong, Org. Biomol. Chem. 13 (2015) 11194–11199.
- [20] A.N. Butkevich, G.Y. Mitronova, S.C. Sidenstein, J.L. Klocke, D. Kamin, D.N.H. Meineke, E. D'Este, P. Kraemer, J.G. Danzl, V.N. Belov, S.W. Hell, Angew Chem Int Edit. 128 (2016) 3350–3355.
- [21] D. Ren, Y. Liu, X. Liu, Z. Li, H. Li, X. Yang, Sens. Actuators B. 255 (2018) 2321–2328.
- [22] X. Ma, J. Wang, Q. Shan, Z. Tan, G. Wei, D. Wei, Y. Du, Org. Lett. 14 (2012) 820.
- [23] A. Gaeta, R.C. Hider, Brit. J. Pharmacol. 146 (2005) 1041–1059.
- [24] A.I. Bush, Neurobiol Aging. 23 (2002) 1031–1038.
- [25] J. Yang, Z.-L. Yuan, G.-Q. Yu, S.-L. He, Q.-H. Hu, Q. Wu, B. Jiang, G. Wei, J. Fluoresc. 26 (2016) 43–51.
- [26] R.S. Juang, L.D. Shiau, Ind Eng Chem Res. 37 (1998) 555–560.
- [27] S. Miah, I.M.M. Rahman, M. Takemura, S. Fukiagi, A.S. Mashio, T. Maki, H. Hasegawa, Talanta. 194 (2019) 980–990.
- [28] A. Fulgenzi, M.E. Ferrero, Int J Mol Sci. 20 (2019) 1019.
- [29] Z.B. Zhao, H.X. Zheng, Y.G. Wei, J. Liu, Chinese, Chem Lett. 18 (2007) 639–642.
- [30] G. Verreck, A. Decorte, K. Heymans, J. Adriaensens, D. Liu, D. Tomasko, A. Arien, J. Peeters, G. Van den Mooter, M.E. Brewster, Int J Pharm. 327 (2006) 45–50.
- [31] V. André, O. Shemchuk, F. Grepioni, D. Braga, M.T. Duarte, Cryst Growth Des. 17 (2017) 6417–6425.
- [32] M.A. Bailey, M.J. Ingram, D.P. Naughton, K.J. Rutt, H.T. Dodd, Transit Metal Chem. 33 (2008) 195–202.
- [33] A. Goyanes, A.B.M. Buanz, G.B. Hatton, S. Gaisford, A.W. Basit, Eur J Pharm Biopharm. 89 (2015) 157–162.
- [34] N. Oturan, C.T. Aravindakumar, H. Olvera-Vargas, M.M. Sunil Paul, M.A. Oturan, Environ Sci Pollut R. 25 (2018) 20363–20373.
- [35] S.M. Bouzzine, S. Bouzakraoui, M. Bouachrine, M. Hamidi, J. Mol. Struct-Theochem. 726 (2005) 271–276.
- [36] H. Derouiche, V. Djara, Sol Energ Mat Sol C. 91 (2007) 1163–1167.
- [37] G.V. Loukova, V.P. Vasiliev, A.A. Milov, V.A. Smirnov, V.I. Minkin, J Photoch Photobio A. 327 (2016) 6–14.
- [38] A. Paul, S. Anbu, G. Sharma, M.L. Kuznetsov, M.F.C. Guedes Da Silva, B. Koch, A.J.L. Pombeiro, Dalton Trans. 44 (2015) 16953–16964.
- [39] Q. Dai, H. Liu, C. Gao, W. Li, C. Zhu, C. Lin, Y. Tan, Z. Yuan, Y. Jiang, New. J. Chem. 46 (2018) 613–618.
- [40] S.K. Sahoo, D. Sharma, R.K. Bera, G. Crisponic, J.F. Callan, Chem. Soc. Rev. 41 (2012) 7195.
- [41] (a) H. Irving, R.J.P. Williams, Nature 162 (1948) 746; (b) H. Irving, R.J.P. Williams, Journal of the Chemical Society (Resumed). 637 (1953) 3192–3210.

- [42] R. Englman, J.D. Dow, *Phys Today*. 27 (1974) 57.
- [43] S. Liu, Y.M. Wang, J. Han, *J. Photoch. Photobio. C*. 32 (2017) 78–103.
- [44] M. Formica, V. Fusi, L. Giorgi, M. Micheloni, *Coord Chem Rev.* 256 (2012) 170–192.
- [45] H. Lineweaver, D. Burk, *J. Am. Chem. Soc.* 56 (1934) 658–666.
- [46] Y. Zhao, X.B. Zhang, Z.X. Han, L. Qiao, C.Y. Li, L.X. Jian, G.L. Shen, R.Q. Yu, *Anal. Chem.* 81 (2009) 7022–7030.
- [47] W. Cao, X.J. Zheng, C. Feng, L.P. Jin, *Dalton Trans.* 44 (2015) 5191–5196.
- [48] T. Anand, G. Sivaramana, M. Iniya, A. Siva, D. Chellappa, *Anal. Chim. Acta.* 876 (2015) 1–8.
- [49] F. Hou, L. Huang, P. Xi, J. Cheng, X. Zhao, G. Xie, Y. Shi, F. Cheng, X. Yao, D. Bai, Z. Zeng, *Inorg Chem.* 51 (2012) 2454–2460.
- [50] M. Biancalana, S. K. *Biochimica. BBA-Proteins. Proteom.* 1804 (2010) 1405–1412.
- [51] J.H. Viles, *Coordin Chem Rev.* 256 (2012) 2271–2284.
- [52] E. House, J. Collingwood, A. Khan, O. Korchazkina, G. Berthon, C. Exley, *J. Alzheimers Dis.* 6 (2004) 291–301.
- [53] (a) W. Garzon-Rodriguez, A.K. Yatsimirsky, C.G. Glabe, *Bioorg Med Chem Lett.* 9 (1999) 2243–2248;
(b) L.Q. Hatcher, L. Hong, W.D. Bush, T. Carducci, J.D. Simon, *J. Phys Chem B.* 112 (2008) 8160–8164;
(c) B. Alies, E. Renaglia, M. Rozga, W. Bal, P. Faller, *Anal Chem* 85 (3) (2013) 1501–1508;
(d) K.P. Kepp, *Coordin. Chem. Rev.* 351 (2017) 127–159.
- [54] E. Gazit, *Faseb. J.* 16 (2002) 77–83.
- [55] (a) L. Zhu, Y. Han, C. He, X. Huang, Y. Wang, *J. Phys Chem B* 118 (2014) 9298–9305;
(b) K. Murakami, T. Yoshioka, S. Horii, M. Hanaki, S. Midorikawa, S. Taniwaki, H. Gunji, K. Akagi, T. Kawase, K. Hirose, K. Irie, *Chem Commun.* 54 (2018) 6272–6275.