

Communication

Discovery of Fluorogenic Diarylsydnone-Alkene Photo-ligation: Conversion of *ortho*-Dual-Twisted Diarylsydnones into Planar Pyrazolines

linmeng Zhang, Xiaocui Zhang, Zhuojun Yao, Shichao Jiang, Jiajie Deng, Bo Li, and Zhipeng Yu J. Am. Chem. Soc., Just Accepted Manuscript • DOI: 10.1021/jacs.8b02493 • Publication Date (Web): 05 Jun 2018 Downloaded from http://pubs.acs.org on June 5, 2018

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Discovery of Fluorogenic Diarylsydnone-Alkene Photo-ligation: Conversion of *ortho*-Dual-Twisted Diarylsydnones into Planar Pyrazolines

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Supporting Information Placeholder

ABSTRACT: A small library of diarylsydnones (DASyds) was constructed based on aryl-pairing combinations and subjected to click reaction toward alkenes under photo-irradiation with high efficiency. We were able to demonstrate the utility of DASyds for highly fluorescent turn-on ligation targeting the TCO moieties on protein.

Photo-induced chemoselective covalent-bond forming reactions (photo-click)¹ which introduce probing functionalities are desired tools for materials science² to functionalize nanoparticles and interfaces with real-time response, as well as for the labeling of oligonucleotides, proteins, and oligosaccharides.³ Photon utilization^{3a, 4} and the precision of the chemistry⁴ are both vital for offering better bioorthogonality⁵ accompanied with higher spatiotemporal resolution. In spite of this need, only a limited number of photo-click methods have been investigated currently including tetrazole-alkene photo-click chemistry (TAP),⁶ photo-initiated thiol-ene,^{4c} -yne,^{4d} -quinone^{4e} reactions, and photo-induced strainpromoted azide-alkyne cycloaddition (photo-SPAAC).^{2b} Since the TAP reaction⁶ was reported as a bioorthogonal reaction, the photo-click strategy, including [3+2]-type: TAP; photo-SPAAC and the [4+2]-type: o-naphthoquinone methides (oNQMs)-ene coupling,4f has been used to visualize proteins in living cells, to study D(R)NA dynamics,^{3c-d} and to track drug molecules.^{6c} In spite of the exciting developments over the past two decades, demand remains high for the discovery of new and high-performing photoclick reactions, especially those with the ability to site-specifically hit the desired targets for use in theranostics.

N-Phenyl-sydnones were discovered to be clickable with alkynes via copper complex mediation by Taran⁷ or strainpromotion by Chin^{8a} and Murphy;^{8b} and tremendous enhancement of cycloaddition rate achieved via introduction of fluorine substitution by Taran,^{8c} elevated this functionality as potential chemical probes to study biomolecular dynamics *in vivo*. Inspired by early sydnone photo-reactions,⁹⁻¹⁰ we investigated the photo-induced diarylsydnone-alkene cycloaddition reaction, which affords pyrazoline fluorophores (Scheme 1), as a potentially bioorthogonal conjugation strategy.⁵ Since nitrile imine (NI) is the reactive dipolar intermediate,¹¹ diarylsydnones (DASyd) demonstrate commendable selectivity toward electron-deficient or ring-strained alkenes through a photo-induced transformation sequence.¹² Unlike those in 2,5-diaryltetrazoles, the two *ortho*-substituted aromatic rings in the DASyd scaffold are twisted around the sydnone core, allowing for unique tuning of reactivity and selectivity of these chemical ligation reagents in 3-D way, as reported herein.

In order to identify a DASyd with excellent photo-response, we consider the electronic effects on the diaryl moieties might be vital to the photo-decarboxylation,12 including the auxochromes, the polycyclic aromatics (PAr) and the electron donors (eD) or acceptors (eA).¹³ Accordingly, a small library of DASyd bearing various aryl substituents on C^4 and N^3 positions were designed (1a-n, Figure 1a; S8-19, Table S8) and constructed.¹⁴ The UV-Vis spectroscopic properties of representative DASyds were evaluated with maximum absorption wavelengths and the top-three ranked maximum extinction coefficients (ε_{max}) were observed for 1k>1j>1h (Figure 1b, Table S1) implying a C⁴-PAr-eA is beneficial. When the N^3 -Ph substituent was an eD, it could offer higher ε value at λ_{max} than seen with an eA moiety (1c vs. 1d). This effect on the ε value was particularly evident when the N³-Ph-eD was coupled with a C^4 -Ph-eA, in contrast to a C^4 -Ph-eD (1e vs. 1j) which processes a bathochromic shifting effect. Intriguingly, the absorption peak tail of the simple DASyd could be observed in the visible region (>400 nm, e.g. 1e and 1f), which opens the window for visible light-mediated triggering.



Scheme 1. The flipping of the diaryl substitutions in the SAP versus TAP.

Subsequent analysis of the photochemistry of the DASyds toward methyl methacrylate in quartz test tube was inquired under exposure to a handheld UV lamp (311 nm, 10.8 mW/cm²) or a LED array (371 nm, 15.4 mW/cm² or 405 nm, 33.6 mW/cm²). Based on HPLC-MS analysis, the experimental results can be summarized as follows: (i) substrates with N^3 -Ph-eA groups (**1b** & **1d**) showed poor reactivity toward alkene (Table 1, entries 2 & 4); (ii) 311 nm irradiation afforded higher conversions for the entire DASyd library (Table 1); (iii) surprisingly, a *p*-aniline moiety on either the N^3 or C^4 positions (e.g., **1e** & **1f**) exhibited very poor photo-response despite their strong absorption at 371 nm and 405 nm (Table 1, entries 5 & 6; Figure S34). Therefore, N^3 -*p*-anisole- C^4 -Ar-eA-Syds are a better choice for continued screening. Partial hydrolysis of the active intermediate was observed for **1j** and **1k**, accounting for the discrepancy between isolated yield and conversion (Table 1, entries 10 & 11, Figure S11-12) despite their higher conversion at 405 nm. Overall, the three best DASyds at 311 nm irradiation were **1g**, **1h** and **1i** in descending order of reactivity (Table 1, entries 7-9), and no competitive cycloadditions were observed in the dark environment.¹⁵



Figure 1. (a) Structures of selected DASyds with the C-H activation crosscoupling yield. (b) Absorption spectra and molar absorption coefficients at designated wavelength for selected DASyds from library in acetonitrile/H₂O (1:1) mixed solvent at 30 μ M (250-450 nm).

 Table 1. Photo-activated 1,3-Dipolar Cycloaddition of 1a-1n

 with Methyl Methacrylate^a



 a 5 mg of 1 and 20 eq. of MMA in 10 mL of ethyl acetate (EtOAc) with 311 nm UV for 2 h unless noted otherwise. ^bIsolated yield. ^c100 μ M 1 and 2 mM 2a in ACN/H₂O (1:1) was irradiated with designated light source for 60s. Determined by LC-MS. See Figure S2-14 for details. ^dN.D. = not detected. ^eAll converted into undesired products. ^fBy-products formed. ^g30 mg of 1 in 50 mL of EtOAc scale. ^hHydrolysis as a predominate side-reaction.

Only a single regioisomer of **3ga** was observed based on the ¹H NMR signals of the Pyr protons and XRD structure, thus verifying the synergistic occurrence of geometric flipping and planarization with the photo-reaction (Table 1; Figure S34). Compared to the DASyd (**1g**, **1i**, **1j**), the corresponding 2,5-diaryltetrazole has an hypsochromic shift of absorbance peak around 37-48 nm, resulting in its complete inertness under excitation by 371 nm LED (Figure S23).

To expand the scope of reporters¹⁶ paired with 1g, we examined the photo-induced ligation by incubating 1g with a range of representative alkenes upon 311 nm irradiation (10.8 mW/cm²) (Table 2). As expected, electron-deficient alkenes, such as 2a and 2d (diethyl fumarate, DEF), reacted efficiently with 1g within 2h, furnishing high yields, despite concomitant formation of the corresponding pyrazole as an oxidative aromatization product, especially for DEF (Table 2, entry 4, page 153 in SI). Alternatively, alkene 2c (norbornene, NOR) exhibited a relative low yield (66%, Table 2, entry 3, Figure S16) mainly because of side-reactions. The genetic encodable ring-strained trans-cyclooct-4-en-1-ol (TCO)^{4a} appears to be promising, undergoing the photo-click reaction to form 3gb (Table 2, entry 2), superior not only in ligation precision but also satisfying Φ_F (up to 42%). Unexpectedly, **2e** (5-vinyl-2'-deoxyuridine, VdU)¹⁷ behaves as a good ligation substrate with respect to 1g, affording a 68% yield of 3ge with little oxidation. The slightly lower yield is caused by incomplete conversion of 1g because of the extra optical filtering effect from VdU (Table 2, entry 5).18 Significant bathochromic shift of absorption (λ_{max} shifted from 33 to 66 nm), arising in emission maxima (λ_{em} , from 525 to 543 nm), accompanied with large Stokes' shifts (Δ_{λ} , from 158 to 163 nm) were spectroscopic characteristics of fluorophores, 3ga-3ge, upon isolation (Table 2).

 Table 2. Photo-activated 1,3-Dipolar Cycloaddition of 1g with

 Various Alkene Dipolarophiles^a

MeQ CF3						
		$H = \frac{R_2}{R_1} \frac{31}{H}$	1 nm, 25°C EtOAc, 2h CO ₂ Me	F 0-	$R_1 R_2$ (±)	CF3
	1g	2			3g	
Entry	Alkene	$\lambda_{ m max}/\lambda_{ m em}$ (nm)	$\Phi_{\rm F}\!/{\epsilon_{max}}^b$	Stoke shift	Yield ^c (%)	Conv. ^f (%)
1	Me J OMe	367/525	0.56/15.9	158 nm	83	94
2	но [—] 2b	376/534	0.42/11.2	158 nm	85	97
3	() 2c	396/533	0.35/18.9	137 nm	66	91 ^g
4		380/543	0.22/17.1	163 nm	$78^{d} (9^{e})$	95
5	°↓°~↓ 2e	381/534	0.16/15.2	153 nm	68	46 ^{<i>g</i>}

^{*a*}Irradiation 20 mg of **1g** and 5 eq. of alkenes in 50 mL of EtOAc for 2h. ^{*b*} Φ_F denote fluorescence quantum yield, ε_{max} in $10^3 \, \text{M}^{-1} \text{cm}^{-1}$. ^{*c*}Isolated yields. ^{*d*}20 eq. DEF. ^{*c*}Side-product from oxidative aromatization. ^{*f*}100 μ M **1g** and 0.5 mM **2** in ACN/H₂O (1:1), 311 nm UV for 60s. Determined by LC-MS. See Figure S15-18 for details. ^{*g*}Not peak-to-peak conversion.

Encouraged by the high efficiency of the photo-reaction, we continued to determine the magnitude of fluorescence turn-on effect derived from preferred DASyds, **1g-1i**, toward TCO under irradiation at 371 nm (15.4 mW/cm²), appending corresponding photophysical properties of the resultant Pyrs (Table 3). Likewise, the chromophoric and fluorophoric nature of products **3gb-3ib** in the visible range allowed for facile visualization of their formation. Three DASyds show excellent degrees of fluorescence turn-on effect, ranging from 122-fold for **3gb** to 321-fold for **3ib** (Table 3).¹⁵ As displayed in Figure 2, complete photo-conversion to the desired Pyr **3hb** was achieved within 520s as evidenced by the

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59 60 iterative increase in a new absorption peak at 397 nm and a broad emission peak centered at 553 nm simultaneously. Complete conversion to **3gb** requires longer time than **3hb**, but still proceeds within a similar scale (for turn-on ratio in ACN:PBS = 1:2, pH~7.4, also see Figure S19-S21). Overall, DASyds **1g-1i** hold great potential for fluorogenic imaging.

The plausible mechanism of the DASyd-alkene photo-reaction was proposed,¹² but the reaction rate as a ligation method has not been investigated. We probed the rate profile by capturing timedependent fluorescence evolution under LED irradiation (371nm, 28.0 mW/cm², Figure S1) via monitoring the λ_{em} signal of corresponding Pyr product. Unexpectedly, the apparent first-order rate constant, k_{obs} , detected for **1g-1i** with either TCO or NOR were almost equal $(k_{obs1g-TCO} = 0.045 \text{ s}^{-1} \text{ vs. } k_{obs1g-NOR} = 0.044 \text{ s}^{-1}$, Figure S1), independent on the alkene type or concentration but dependent on the DASyd itself ($k_{obs1g-TCO} = 0.045 \text{ s}^{-1} \text{ vs. } k_{obs1i-TCO} =$ 0.034 s^{-1} , Table 3), implying that the decarbonylation of 4,5diaryl-2-oxa-1,5-diazabicyclo[2.1.0]pentan-3-one^{12b} (Figure S1) might become the rate determining step instead of the ultra-fast [3+2] cycloaddition¹¹ (Figure S1). The quantum yields of the photo-chemical reaction for 1g, 1h, and 1i were determined to be 0.20, 0.10, and 0.12, respectively (for yield in ACN:PBS = 1:2,)pH~7.4, also see Figure S25).

Table 3. Photophysical Properties of the TCO-Pyrs and Fluorescence Turn-On Efficiency^{*a*}

HO" _______ 371 nm -CO; ACN/H2O (1:1) -0 MeO (±) 1g/3gb: R¹=H; R²=CF₃; R³=H Fluorescence turn-on DASyds 1h/3hb; R1=F; R2=CF3; R3=F R¹=CF₃; R²=H; R³=CF₃ Pyr 1i/3ib: $\lambda_{max}/\lambda_{cm}$ Fluorescence kobs Comp $\Phi_{\rm F}$ Φ_{p} Emax \mathcal{E}_{400nm} (s⁻¹) (nm) turn-on^f 13,573 347 N.D. 331/-1g 0.20 0.045 122-fold 376/532 0.42 3gb 11.156 8.100 N.D. 334/-15,333 213 1h 0.10 0.043 205-fold 3hb 396/552 18,533 18,400 0.33 331/-11,853 187 N.D. 1i 0.12 0.034 321-fold 383/543 13 178 11 611 0.28 3ib

^{*a*}25 µM **1g-1i** in ACN/H₂O (1:1). ^{*b*} λ_{max} in the 300-500 nm region. ^cM⁻¹cm⁻¹. ^{*d*}Fluorescence quantum yield in EtOAc. ^cReaction quantum yield. ^fObtained by comparing the emission intensity of Pyr to that of the initial reactants mixture at λ_{em} ; $\lambda_{ex} = 400$ nm.



Figure 2. Time course of photo-ligation of TCO-DASyd as monitored by UVvis (dashed lines) and fluorescence (solid lines). **1h** in ACN/H₂O (1:1) to derive concentrations of 25 μ M, and the activation wavelength was set at 365 nm (0.95 mW/cm²). For fluorescence, $\lambda_{ex} = 400$ nm.

To demonstrate the utility of the DASyds for bioconjugation, we first appended the TCO moiety to lysozyme (lyso-TCO, M.W. ~14.3 kDa) via chemical tagging and to sfGFP via genetic incorporation (sfGFP-Q204TCOK, M.W. ~27.9 kDa).¹⁹ Then, the residue-specific protein modification was investigated by reacting the resultant proteins with **1g** and **1k** under irradiation of 311 nm and 405 nm light for 2 min, respectively. The resultant mixture was profiled by in-gel fluorescence imaging (Figure 3a-b) and LC–MS (Figure 3c). The conversions into lyso-Pyr-**1g** or lyso-Pyr-**1k** or

sfGFP-Pyr-1g were estimated to be 40%, 32% and 14% (Figure S26-32, S35-36) with 1.8%, 4.4% and 8.1% of non-specific reactions respectively. The LC-MS/MS analysis revealed that the photo-ligation reaction of 1g has a highly spatial preference toward K97-TCO residue of lyso-TCO (Figure S33). In-gel fluorescence imaging indicated obvious fluorescent turn-on bands only for lanes meeting all requirements for the photo-ligation (Figure 3a-b).



Figure 3. Selective photo-induced fluorogenic labeling of proteins by **1g** *in vitro*. Coomassie blue stain (top panel) and in-gel fluorescence (bottom panel) of the same protein gel after resolved by SDS-PAGE (a) lyso (b) sfGFP. (c) Deconvoluted mass spectra of control protein (red) and the TCO-protein mixture (blue) showing the formation of the Pyr adduct (green). NR. = non-specific reaction (purple).

In summary, with a small library of photo-activatable DASyds in hand, we were able to identify **1g-1i** as excellent candidates for photo-clickable fluorogenic ligation. The **1g** showed high reactivity toward TCO and TCO-containing proteins when exposed to near-UV light, giving rise to fluorescent Pyr cycloadducts with outstanding ratio of fluorescence turn-on effect. We will consider the applications of DASyd derivatives as light-activable reagents for ligation reaction in living systems.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI:

Experimental details, characterization of all new compounds, and computational results (PDF).

X-ray crystal structure of **1f** (CCDC-1556441) (CIF) X-ray crystal structure of **1g** (CCDC-1556442) (CIF) X-ray crystal structure of **1m** (CCDC-1823237) (CIF) X-ray crystal structure of **3ga** (CCDC-1556443) (CIF)

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Notes

The authors declare no competing financial interests.

ACKNOWLEDGMENT

We thank Dr. Jason J. Chruma (Sichuan University) for valuable help and suggestions in manuscript preparation. We thank Dr. Wenshe Liu (Texas A&M) and Dr. Qing Lin (SUNY-UB) for generously providing us the plasmids pEvol-PyIT-PyIRS and pET-sfGFP-Q204TAG. We thank Dr. Pengchi Deng (Analytical & Testing Center, SCU) for help on NMR spectra collection. Financial support was provided by the National Natural Science Foundation of China (21502130), the "1000-Youth Talents Pro-

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gram", and the Fundamental Research Funds for the Central Universities.

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(15) The competitive thermal cycloaddition reactions with TCO and MMA in dark are extremely slow at 25 $^{\circ}$ C (Figure S22).

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411x186mm (144 x 144 DPI)



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Figure 1. (a) Structures of selected DASyds with the C-H activation cross-coupling yield.

339x221mm (144 x 144 DPI)



Figure 1. (b) Absorption spectra and molar absorption coefficients at designated wavelength for selected DASyds from library in acetonitrile/ H_2O (1:1) mixed solvent at 30 μ M (250-450 nm).

409x197mm (144 x 144 DPI)





Table 2. Photo-activated 1,3-Dipolar Cycloaddition of **1g** with Various Alkene Dipolarophiles

379x95mm (144 x 144 DPI)







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Figure 2. Time course of photo-ligation of TCO-DASyd as monitored by UV-vis (dashed lines) and fluorescence (solid lines). DASyd in ACN/H₂O (1:1) to derive concentrations of 25 μ M, and the activation wavelength was set at 365 nm (0.95 mW/cm²). For fluorescence, $\lambda_{ex} = 400$ nm.

414x171mm (144 x 144 DPI)





Figure 3. Selective photo-induced fluorogenic labeling of proteins by **1g** *in vitro*. Coomassie blue stain (top panel) and in-gel fluorescence (bottom panel) of the same protein gel after resolved by SDS-PAGE (a) lyso (b) sfGFP. (c) Deconvoluted mass spectra of control protein (red) and the TCO-protein mixture (blue) showing the formation of the Pyr adduct (green). NR. = non-specific reaction (purple).

413x178mm (144 x 144 DPI)



Table of Contents

403x137mm (144 x 144 DPI)