

Structure—Activity Relationships and Pharmacophore Model of a Noncompetitive Pyrazoline Containing Class of GluN2C/GluN2D Selective Antagonists

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Supporting Information

ABSTRACT: Here we describe the synthesis and structure—activity relationship for a class of pyrazoline-containing dihydroquinolone negative allosteric modulators of the NMDA receptor that show strong subunit selectivity for GluN2C- and GluN2D-containing receptors over GluN2A- and GluN2B-containing receptors. Several members of this class inhibit NMDA receptor responses in the nanomolar range and are more than 50-fold selective over GluN1/ GluN2A and GluN1/GluN2B NMDA receptors, as well as AMPA, kainate, GABA, glycine, nicotinic, serotonin, and purinergic receptors. Analysis of the purified enantiomers of one of the more potent and selective compounds shows that the S-enantiomer is both more potent

and more selective than the R-enantiomer. The S-enantiomer had an IC $_{50}$ of 0.17-0.22 μ M at GluN2D- and GluN2C-containing receptors, respectively, and showed over 70-fold selectivity over other NMDA receptor subunits. The subunit selectivity of this class of compounds should be useful in defining the role of GluN2C- and GluN2D-containing receptors in specific brain circuits in both physiological and pathophysiological conditions.

INTRODUCTION

Glutamatergic neurotransmission through ionotropic glutamate receptors is the primary means of excitatory synaptic transmission in the mammalian central nervous system (CNS). The receptor family comprises the α -amino-3-hydroxy-5-methyl-4-isoxazole propionate (AMPA), N-methyl-0-aspartate (NMDA), and kainate receptors. NMDA receptors are widely expressed in the CNS and are thought to be involved in a range of important physiological processes including axonal guidance, synaptic plasticity, and memory formation. MDA receptors are also thought to play an important role in pathophysiological conditions including Parkinson's disease, schizophrenia, depression, and ischemia. $^{2,6-8}$

NMDA receptors mediate the slow component of excitatory synaptic transmission and require the binding of both glutamate and glycine for channel activation. Glycine binds to the GluN1 subunits, which have eight splice variants encoded by a single gene. The GluN2 subunits (GluN2A–D) bind glutamate and are encoded by four distinct genes. The GluN2 subunits control many of the functional and pharmacological properties of the receptor, including agonist EC₅₀, single channel open time and open probability, and deactivation time course following removal of glutamate. NMDA receptor deactivation time course determines the time course for the slow, Ca²⁺-permeable component of synaptic transmission. Typically, NMDA receptors are blocked by extracellular Mg^{2+} at resting membrane potentials, and the requirements of the

glutamate release and depolarization-induced relief of Mg^{2+} block have led to the idea that the NMDA receptors act as coincidence detectors in the brain. ^{19,20} The Mg^{2+} IC₅₀ and the kinetics of block and unblock also vary according to the GluN2 subunit. ²¹

The GluN1 subunits are expressed throughout the CNS, but GluN2 subunit composition and expression vary during development and anatomically. The spatially restricted expression patterns, together with distinct functional and pharmacological differences imparted by the GluN2 subunits, make NMDA receptor subunit selective modulators of therapeutic interest for several neurological disorders, including stroke, schizophrenia, treatment-resistant depression, and Parkinson's disease. Subunit selectivity will restrict modulator actions to brain regions that express the subunit of interest, potentially limiting side effects that occur as a result of global NMDA receptor blockade.

We previously have described the discovery, preliminary SAR of 25 compounds, and pharmacological mechanism of a representative member of the dihydroquinolone pyrazoline (DQP) class of GluN2C/D subunit-selective antagonists. In our previous study, the most potent analogue, DQP-1105, had an IC₅₀ of 2.7 μ M at GluN2D-containing NMDA receptors and was 41-fold selective over GluN2B-containing receptors (Figure 1A). In this report we provide an extensive exploration of the

Received: May 2, 2013



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Figure 1. Previously reported best-in-class compound and representative structure for SAR: (A) structure of previously reported representative compound DQP-1105; ³⁰ (B) structure of a general analogue with numbered substituents.

SAR, confirmation of mechanism of action, off-target analysis, an analysis of the stereoselectivity for a representative member of the class, and data regarding metabolic stability and potential for blood—brain barrier (BBB) penetration. These efforts have yielded potent and selective analogues as well as insights into the pharmacophore for these pyrazoline-containing compounds.

■ RESULTS

Chemistry. The structure-activity relationship around the quinolone pyrazoline core was probed by testing the potency and selectivity of analogues that contained a variety of aromatic substitutions in combination with perturbations of the acyl chain moiety (Figure 1B). A representative synthesis of these analogues is shown in Scheme 1. Briefly, anthranilic acids were reacted with triphosgene under standard conditions to yield the isatoic anhydride derivatives 80. These compounds were then converted, via the Weinreb amide 81 and a subsequent lithium halogen exchange reaction with aryl bromides 82, to the appropriate benzophenones, 83.³¹ The substituted quinolone core was accessed by condensation of 83 with ethyl acetoacetate using microwave irradiation, yielding compound 84. The resultant methyl ketone underwent base-catalyzed condensation with an appropriate arylaldehyde, 85, yielding the α,β -unsaturated ketone compounds 86. These intermediates could be treated with hydrazine monohydrate in ethanol, utilizing microwave irradiation, to yield the pyrazolinecontaining compounds typified by 87. The pyrazoline amine was then functionalized with succinic anhydride (88), glutaric

Scheme 1. Synthesis of Dihydroquinolone Pyrazoline Derivatives^a

"(a) Anhydrous THF, triphosgene (warning, triphosgene is toxic; see Experimental Section), reflux; (b) EtOH, Weinreb's HCl salt, reflux; (c) anhydrous THF, n-butyllithium, -78 °C; (d) ethyl acetoacetate, DMF, 4 Å molecular sieves, 180 °C, microwave; (e) 4:3 EtOH/H₂O (0.05 M), 0 °C to rt; (f) hydrazine monohydrate, EtOH, 110 °C, microwave; (g) anhydrous THF, 4 Å molecular sieves, 165 °C, microwave.

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Scheme 2. Modifications to the Acyl Chain^a

"(a) HCl, MeOH; (b) BH₃—Me₂S, anhydrous THF, 0 °C; (c) EDCI, DMAP, NH₃ in dioxane (0.5M), THF; (d) EDCI, DMAP, (*E*)-4-methoxy-4-oxobut-2-enoic acid; (e) anhydrous THF, (*E*)-methyl 4-chloro-4-oxobut-2-enoate, 4 Å molecular sieves, 165 °C, microwave; (f) methyl 4-oxobutanoate, BH₃—Me₂S, THF; (g) EDCI, DMAP, 4-fluorobutanoic acid, DCM; (h) NaOH, EtOH/H₂O.

anhydride (89), or maleic anhydride (90) to yield the fully saturated or cis double bond acyl chain derivatives (Scheme 1).

Standard esterification conditions from compound 4 yielded the saturated monomethyl ester analogue 56 (Scheme 2). Reduction of the acid in compound 26 with BH₃-DMS led to the primary alcohol containing compound, 64; a coupling reaction with compound 26 and NH₃ gave the primary amide compound 65 (Scheme 2). The unsaturated fumaric esters could be accessed under standard amide coupling conditions using (E)-4-methoxy-4-oxobut-2-enoic acid and compound 55f, yielding compound 54g, or with acylation of 22f using (E)-methyl 4-chloro-4-oxobut-2-enoate, yielding compound 60 (Scheme 2). Additionally, the acyl chain was replaced with the alkyl chain by reacting the pyrazoline derivative 26f and methyl 4-oxobutanoate under reductive amination conditions to give compound 66 (Scheme 2). Monofluorobutyrate 68c (Scheme S1) was synthesized in three steps and also coupled to

compound **26f**, yielding **68** as an isostere of the hydroxylcontaining compound **64** (Scheme 2). 32,33 The ester containing compounds **54g**, **60**, and **66** could be saponified under basic conditions yielding the target scaffolds **54**, **58**, and **67** (Scheme 2). All compounds were assayed for activity using two-electrode voltage clamp recordings from *Xenopus laevis* oocytes recombinantly expressing the desired NMDA receptor subtypes (see Experimental Section).

A-Ring Substituent Optimization. We first evaluated the effect of substituents on the A-phenyl ring (Figure 1B) by holding the chlorine substitution on the quinolone core constant and evaluating the substitutions shown in Table 1 and Table S1. At the A-ring, 4-substituted phenyl derivatives resulted in the best potency and selectivity. This observation led to the identification of 5 (Table 1), with a nitro group at R₇, which was the most potent para-substituted analogue at GluN2D-containing receptors compared to the unsubstituted

Table 1. Evaluation of A-Ring Para and Ether Substitutions^a

						IC_{50}	(μM)	
DQP	R_6	R_7	$(2A IC_{50})/(2D IC_{50})$	$(2B IC_{50})/(2D IC_{50})$	GluN2A	GluN2B	GluN2C	GluN2D
1	Н	Н			NE	NE	86	88
2	Н	F	9	6	128	87	23	14
3	Н	Cl			NE	NE	5.5	4.5
4	Н	Br		7	NE	22	3.6	3.1
5	Н	NO_2	85	42	91	45	0.9	1.1
6	Н	СООН			NE	NE	NE	NE
7	H	COOMe			NE	NE	93	32
8	H	CF ₃	20	13	80	54	5	4.1
9	Н	OMe	9	9	197	187	28	21
10	Н	NMe_2			NE	NE	39	19
11	-(OCH ₂ O-		4	NE	90	23	23

 $^{\alpha}$ IC₅₀ values were obtained by fitting the Hill equation to the average composite concentration—effect curves (see Experimental Section). Data are from 7–18 oocytes between 2–4 frogs. NE indicates less than 30% inhibition at 100 μ M.

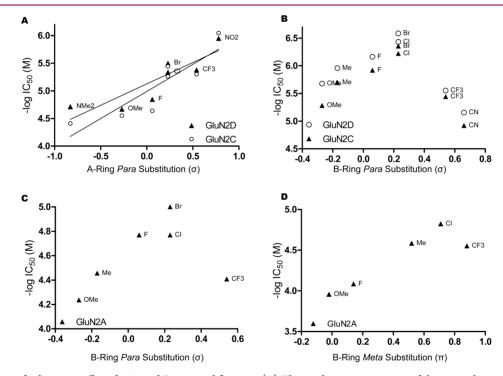


Figure 2. Evaluation of substituent effects for A- and B-ring modifications. (A) The σ substituent constants of the para-substituted A-ring analogues vs activity show a correlation for GluN2C- and GluN2D-containing receptors, when the R₁ position of the C-ring is substituted with chloro: GluN2D r^2 = 0.82, p < 0.05 Pearson two-tailed correlation analysis; GluN2C r^2 = 0.84, p < 0.05 Pearson two-tailed correlation analysis; compounds 2–5 and 8–10). (B) The analysis of the para-substituents on the B-ring as a function of activity at GluN2C- and GluN2D-containing receptors appears parabolic with respect to the σ substituent constants, with an optimal value close to that of the chloro and bromo substitutions (compounds 29–33). (C) The analysis of the para-substituents at GluN2A-containing receptors shows a similar parabolic relationship as observed at the GluN2C- and GluN2D-containing receptors when the activity is plotted as a function of the σ substituent constants (compounds 29–33). (D) The analysis of the substituent effects appears parabolic with respect to the π substituent constants for B-ring meta-substituted compounds at GluN2A-containing receptors, suggesting an optimal hydrophobicity close to that of the chloro substitution. Substituent constants were obtained from the same source (compounds 34–39).

A-ring, compound 1 (1.1 μ M vs 88 μ M, respectively, Table 1). Realizing the substantial liabilities associated with the nitro group, we explored bioisosteres of the substitution, replacing this group with a carboxylic acid 6 (Table 1) which showed no

activity. By contrast, the methyl ester 7 (Table 1) remained active but had decreased potency at GluN2D-containing receptors (32 μ M vs 1.1 μ M). Interestingly, sp³ hybridization is tolerated but not preferred, as can be seen with the

Table 2. A- and B-Ring Modifications^a

						IC ₅₀	(μM)	
DQP	R_3	R_7	$(2A IC_{50})/(2D IC_{50})$	$(2B IC_{50})/(2D IC_{50})$	GluN2A	GluN2B	GluN2C	GluN2D
21	Br	Br	33	59	13	23	0.71	0.39
22	Br	Cl	34	79	10	23	0.56	0.29
23	Br	F	12	26	34	75	3.8	2.9
24	Br	H	7	12	64	113	10	9.1
25	Cl	Br	37	67	19	34	0.95	0.51
26	Cl	Cl	48	50	21	22	0.77	0.44
27	Cl	F	14	26	47	90	4.1	3.4
28	Cl	Н	4	13	49	143	13	11

 $^{^{}a}IC_{50}$ values were obtained by fitting the Hill equation to the average composite concentration—effect curves (see Experimental Section). Data are from 8–18 oocytes between 2–3 frogs. Compound 26 is shown in Table 4 for comparison.

Table 3. B-Ring Modifications^a

						IC ₅₀	(μM)	
DQP	R_2	R_3	$(2A IC_{50})/(2D IC_{50})$	$(2B IC_{50})/(2D IC_{50})$	GluN2A	GluN2B	GluN2C	GluN2D
29	Н	F	36	98	21	57	1.0	0.58
26	H	Cl	48	50	21	22	0.77	0.44
30	Н	Me	24	30	33	42	2.5	1.4
31	H	OMe	27	33	62	75	5	2.3
32	H	CN	22		156	NE	12	7
33	H	CF_3	10	14	29	39	3.6	2.8
34	F	Н	67	101	46	70	1.1	0.69
35	Cl	Н	20	43	20	43	2.1	1.0
36	Me	Н	13	37	24	71	4.0	1.9
37	OMe	Н	24	34	110	152	7.8	4.5
38	CN	Н			NE	NE	19	13
39	CF_3	H	11	18	28	47	3.4	2.6

[&]quot; ${\rm ^{41}C_{50}}$ values were obtained by fitting the Hill equation to the average composite concentration—effect curves (see Experimental Section). Data are from 6–14 oocytes between 2–3 frogs. NE indicates less than 30% inhibition at 100 μ M. Data for **26** are presented in Table 2 and repeated here to facilitate comparisons with data.

trifluoromethyl-containing compound 8 (IC₅₀ = 4.1 μ M, GluN2D, Table 1). Although substitution at any of the three available positions on the A-ring is tolerated, substitution at either the R₅ or R₆ position showed no improvement in potency or selectivity with any of the analogues tested (12–20, 71; Table S1). Similarly, analogues with the A-ring replaced with furan, thiophene, and pyridine substituents were evaluated and were inactive (Table S2).

A manual Hansch analysis, similar to that of the Topliss approach, was employed to better understand the physicochemical properties governing potency. Analysis of the steric, σ , and π substituent effects for the A-ring suggests that only the para- σ contribution is directly associated with the IC₅₀ values at the GluN2C- and GluN2D-containing NMDA receptors (compounds 2–5 and 8–10, Table 1 and Figure 2A). Alpha 34,35

B-Ring Substituent Optimization. We next modified the B-ring substituents with the aim of understanding the substituent effects at the meta- and para-positions (Figure 1B). The p-bromo substitutions at the A- and B-rings in compound 21 showed enhanced potency at GluN2C- and GluN2D-containing receptors, with IC₅₀ values of 0.71 and 0.39 μ M, respectively (Table 2). Interestingly, this compound also showed less selectivity for GluN2D- over GluN2A-containing receptors (33-fold) compared to GluN2B-containing receptors (59-fold), suggesting that a more favorable interaction with GluN2A-containing receptors had been formed (Table 2).

Co-varying A-ring para-substituents with the p-bromo B-ring substitution allowed us to determine that the p-chloro A-ring substitution was optimal for potency (22–24, Table 2). A similar trend was observed using the p-chloro substitution on the B-ring while co-varying the A-ring substituents (25–28,

Table 4. B-Ring Disubstitution^a

							IC ₅₀	(μM)	
DQP	R_2	R_3	R_4	$(2A IC_{50})/(2D IC_{50})$	$(2B IC_{50})/(2D IC_{50})$	GluN2A	GluN2B	GluN2C	GluN2D
40	F	Cl	Н	23	53	12	28	0.91	0.53
41	Cl	F	Н	19	34	19	34	1.4	1.0
42	Cl	Cl	Н	8	22	7.7	20	0.79	0.91
43	F	F	Н	32	108	21	71	0.78	0.66
44	F	Н	F	26	123	21	100	1.1	0.81
45	Cl	Н	Cl	8	19	5.5	13	0.78	0.70

 ${}^{a}\text{IC}_{50}$ values were obtained by fitting the Hill equation to the average composite concentration–effect curves from oocyte recordings (see Experimental Section). Data are from 8–15 oocytes between 2 frogs.

Table 2). Therefore, this *p*-chloro substitution on the A-ring was used for further SAR elaboration. We varied substituents at the meta- and para-position of the B-ring which identified numerous analogues that were highly potent and selective (Table 3). Notably, the *p*-fluoro containing compound **29** and the *m*-fluoro containing compound **34** were both potent congeners that showed over 90-fold selectivity for GluN2D-over GluN2B-containing receptors; compound **34** also showed 67-fold selectivity over GluN2A-containing receptors (Table 3).

The Hansch evaluation of the para- σ substituent effects at the B-ring shows a seemingly parabolic relationship when compared to potency for only GluN2A-, GluN2C-, and GluN2D-containing receptors (compounds 26 and 29-33, Table 3, Figure 2B,C), with an optimal σ value corresponding to the bromo and chloro substitutions at all three receptors. At the meta-position of the B-ring, the correlation between the potency and the hydrophobic π value for substitutions also appears parabolic at GluN2A-containing receptors (compounds 34-39, Table 3, Figure 2D). The decrease in potency at GluN2A-containing receptors observed with compound 39, which was meta-substituted with the CF₃ group (Table 3 and Figure 2D), could be a result of steric clashes with the receptor or, as was observed with the para- σ substituents, could suggest that the optimal hydrophobicity at the GluN2A-containing receptors is attained with the *m*-chloro substitution.

From this analysis of meta- and para-substitutions, we hypothesized that combining an optimal para-substitution for potency at GluN2D-containing receptors with a meta-substitution that was less active at GluN2A-containing receptors might improve selectivity. Both Cl and F substitutions gave optimal potency and selectivity when monosubstituted on the B-ring, leading us to co-vary these groups (Table 4). We synthesized compound 40, which has a *m*-fluoro and *p*-chloro substitution pattern on the B-ring. This compound maintained potency but did not increase selectivity (Table 4). Several other compounds that were disubstituted on the B-ring exhibited submicromolar potency at GluN2D-containing receptors, but all showed modest selectivity over GluN2A-containing receptors (41–45, Table 4).

C-Ring Substitutions with Optimized A- and B-Ring Substituents. We next made a series of substitutions to the Cring on the quinolone core (Table S3). Beginning with a methyl group at R_1 (Figure 1B) in combination with either the p-chloro or the m-fluoro substitution on the B-ring, we

synthesized compounds 46 and 47, which decreased the potency compared to the more favorable compounds with only B-ring and A-ring substitutions (Table S3). Interestingly, the modifications showed variability with regard to the relative selectivity for GluN2A- over GluN2B-containing receptors, suggesting that there remains room in this portion of the binding pocket for potential optimization of selectivity (compounds 48–53, Table S3).

Acyl-Chain Perturbations. We subsequently evaluated a series of perturbations to the acyl chain of the pyrazoline nitrogen (Table 5). Restricting the conformation to a cisconfiguration with the maleate derivative maintained similar potency to the parent compound in each instance tested (55 and 59 vs 4 and 22, respectively, Table 5). The transconfiguration in the fumaric containing derivative 58 was the most potent compound identified but was no more selective over GluN2A- or GluN2B-containing receptors than the saturated derivative 22 (Tables 5 and 2). The succinic ester containing compound 56 was inactive, as was the fumaric ester **60** (Table 5). We also evaluated glutaric-containing derivatives such as compound 61 (Table 5), which showed potencies similar to that of the succinic derivative 22 (Tables 5 and 2) at all receptors tested, suggesting that the length of the acyl chain is not crucial for activity.

The primary alcohol containing compound **64** retained similar activity as that of the parent compound **26** at GluN2C-and GluN2D-containing receptors (IC $_{50}$ of 1.7 and 0.69 μ M, respectively) while improving selectivity over GluN2A-containing receptors to 90-fold (Table 5). The primary amide derivative of the succinate acyl chain in compound **65** retained activity but showed decreased potency and selectivity compared to the alcohol and acid moieties (Table 5). Replacing the amide linkage to the pyrazoline with the alkyl derivative in compound **67** slightly diminished potency at GluN2D-containing receptors over the parent compound **26** but maintained selectivity over the other receptor subtypes (Tables 5 and 2). The monofluoro isostere of the hydroxyl group in compound **64** was tested with compound **68** (Table 5). While this compound retained the ability to accept a hydrogen bond, it was inactive.

Stereochemical Preference of a Representative Analogue. Finally, we evaluated the selectivity and potency of purified enantiomers for a representative member of this class of compounds. The enantiomers of the racemic final product 26 were separable via reverse phase chiral chromatography

Table 5. Acyl Chain Perturbations^a

DQP-	R₁	R ₃	R ₇	Acyl Chain	2A IC ₅₀ 2D IC ₅₀	2B IC ₅₀ 2D IC ₅₀	GluN2A IC ₅₀ (μM)	GluN2B IC ₅₀ (μM)	GluN2C IC ₅₀ (μM)	GluN2D IC ₅₀ (μM)
4	CI	Н	Br	У О О О О О О Н	-	7	NE	22	3.6	3.1
54	CI	Н	Br	O OH	-	-	NE	NE	2.1	1.4
55	CI	Н	Br	²⁵ → OH	15	7	74	37	8.9	5
56	CI	Н	Br	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	-	-	NE	NE	NE	NE
57	CI	Н	Br	O O OH	20	23	78	90	6.4	4.0
22	Н	Br	CI	¥, OH	34	79	10	23	0.56	0.29
58	Н	Br	CI	OH	23	63	4.3	12	0.20	0.19
59	Н	Br	CI	Ö V _Z ——OH	21	30	12	17	1.0	0.57
60	Н	Br	CI	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	-	-	NE	NE	59	95
61	Н	Br	CI	O O O O O O O	33	91	10	29	0.6	0.32
5	CI	Н	NO ₂	ў ОН	85	42	91	45	0.9	1.1
63	CI	Н	NO ₂	O O OH	10	8	109	92	10	11
26	Н	CI	CI	¥, OH	48	50	21	22	0.77	0.44
64	Н	CI	CI	O OH	90	48	62	33	1.7	0.69
65	Н	CI	CI	NH ₂	31	18	34	20	2.0	1.1
66	Н	CI	CI	**************************************	-	-	NE	NE	NE	NE
67	Н	CI	CI	ZZ OH	34	31	58	53	3	1.7
68	Н	CI	CI	22, F	-	-	NE	NE	NE	NE

 $[^]a$ IC $_{50}$ values were obtained by fitting the Hill equation to the average composite concentration—effect curves (see Experimental Section). Data are from 5–24 oocytes between 2–4 frogs. NE indicates less than 30% inhibition at 100 μ M. Data for compounds 4, 22, 5, and 26 were presented in preceding tables and are shown here for comparison.

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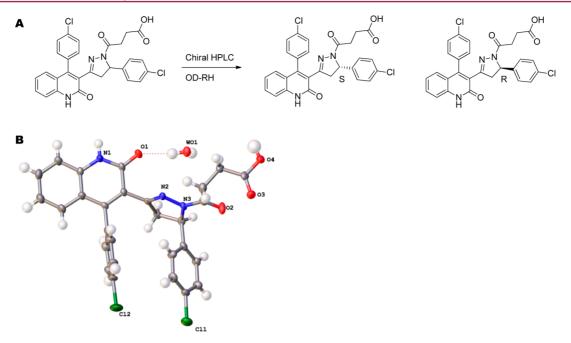


Figure 3. Separation of enantiomers. (A) The enantiomers of the final compound, **26**, could be separated using reverse phase chiral chromatography (see Experimental Section). (B) The crystal structure of the inactive enantiomer, **70** (Table 6), was solved using X-ray diffraction and has the R configuration.

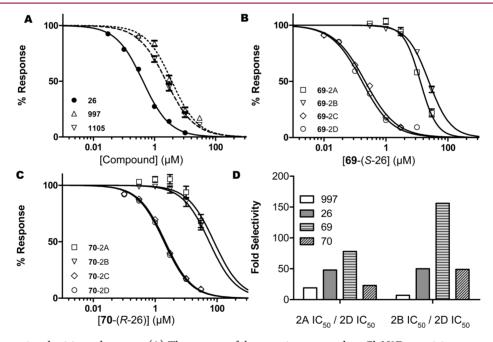


Figure 4. Improvements in selectivity and potency. (A) The potency of the racemic compounds at GluN2D-containing receptors was improved 10-fold over the previous members in the class. (B) The potency of the S-enantiomer of compound 26, compound 69, is 2-fold more potent than the racemic mixture at GluN2D-containing receptors, while the potency at GluN2A- and GluN2B-containing receptors is unaffected, making it more selective for GluN2C- and GluN2D-containing receptors. (C) The potency of the R-enantiomer of compound 26, compound 70, at GluN2C- and GluN2D-containing receptors is diminished compared to the racemate 26, making it less selective over GluN2A- and GluN2B-containing receptors. (D) Bar graph showing the fold-selectivity improvements attained through SAR. Data for compounds 997 and DQP-1105 (A, D) are previously published and shown here for comparison.³⁰

using an OD-RH column (Figure 3, see Experimental Section). Absolute stereochemistry of the second peak to elute during the enantiomeric resolution was assigned using X-ray crystallography as the R-enantiomer (70) (Figure 3, see Experimental Section). Evaluation of the purified enantiomers showed that the S-enantiomer (69) is 11-fold more potent at GluN2D-containing receptors (IC₅₀ = 0.17 μ M) than the R-enantiomer

70 (IC $_{50}$ = 1.9 μ M; Figure 4 and Table 6). In addition, 69 shows enhanced selectivity for GluN2C- and GluN2D- over GluN2A- and GluN2B-containing receptors compared to the racemic 26 and the R-enantiomer 70 due to the enhanced potency at the GluN2D-containing receptors (Figure 4 and Table 6).

Table 6. Stereoselectivity for the Purified Enantiomers of Compound 26^a

						IC_{50}	(μM)	
DQP	R_2	R_4	$(2A IC_{50})/(2D IC_{50})$	$(2B IC_{50})/(2D IC_{50})$	GluN2A	GluN2B	GluN2C	GluN2D
26	Cl	Cl	48	50	21	22	0.77	0.44
69 (S- 26)	Cl	Cl	78	156	13	26	0.22	0.17
70 (R-26)	Cl	Cl	23	49	45	52	2.1	1.9

 $^{{}^{}a}\text{IC}_{50}$ values were obtained by fitting the Hill equation to the average composite concentration—effect curves (see Experimental Section). Data are from 8–17 oocytes between 2–4 frogs. Data for compound 26, which were presented in preceding tables, are shown here for comparison.

Table 7. Off-Target Responses for Compounds 26 and 58^a

		26		58	
receptor	agonist (μM)	$I_{\text{test}}/I_{\text{control}}$ (mean \pm SEM, %)	N	$I_{\text{test}}/I_{\text{control}}$ (mean \pm SEM, %)	N
GluN1/GluN2A	100 glutamate, 30 glycine	89 ± 4.2*	14	89 ± 2.2*	12
GluN1/GluN2B	100 glutamate, 30 glycine	79 ± 2.9*	12	98 ± 1.9	12
GluN1/GluN2C	100 glutamate, 30 glycine	$21 \pm 1.2*$	14	$26 \pm 1.5^*$	11
GluN1/GluN2D	100 glutamate, 30 glycine	14 ± 1.6*	13	14 ± 1.6*	13
GluA1	100 glutamate	97 ± 2.2	6	99 ± 1.1	6
GluA2	100 glutamate	98 ± 0.8	3	96 ± 0.9*	4
GluA3	100 glutamate	99 ± 0.3	4	100 ± 1.1	4
GluA4	100 glutamate	96 ± 1.3	3	97 ± 1.5	4
GluK1	100 glutamate	97 ± 1.0	3	100 ± 4.4	3
GluK2	100 glutamate	97 ± 1.1	4	$97 \pm 0.6*$	4
GluK2/GluK5	100 glutamate	97 ± 1.3	3	95 ± 1.8	3
serotonin 5-HT _{3A}	3 serotonin	95 ± 1.5*	4	95 ± 1.2*	4
GABA _A $\alpha\beta_1\beta_2\gamma_2$ s	20 GABA	97 ± 2.7	4	95 ± 3.1	4
$GABA_{C} (\rho 1)^{(human)}$	2 GABA	99 + 2.1	4	97 + 0.6*	4
glycine α_1	50 glycine	101 ± 1.6	4	99 ± 1.0	4
nicotinic $\alpha_1 \beta_1 \gamma \delta^{(\text{mouse})}$	1 acetylcholine	94 ± 1.2*	6	98 ± 0.7	7
nicotinic $\alpha_4 \beta_2^{\text{(human)}}$	10 acetylcholine	79 ± 4.2*	6	$82 \pm 1.7*$	5
nicotinic $\alpha_3 \beta_4^{\text{(human)}}$	10 acetylcholine	77 ± 2.7*	7	87 ± 2.2*	5
nicotinic $\alpha_7^{(human)}$	300 acetylcholine	82 ± 9.4	3	$64 \pm 7.3*$	3
nicotinic $\alpha_9\alpha_{10}$	100 acetylcholine	67 ± 4.6*	3	72 ± 4.7*	4
purinergic P2 _{X2} ^(human)	9 ATP	$113 \pm 1.8*$	5	100 ± 0.9	4
purinergic P2 _{X2} (rat)	9 ATP	97 + 1.2*	5	96 + 1.0*	4

[&]quot;Agonist-evoked currents were recorded from the receptors listed using the *Xenopus laevis* oocyte expression system under two-electrode voltage clamp ($V_{\text{hold}} = -30 \text{ to } -60 \text{ mV}$) in the absence and presence of 3 μ M 26 or 3 μ M 58. The cDNA origin used was rat unless otherwise indicated (paired *t*-test, (*) p < 0.05).

Mechanism and Site of Action. A previous study of a representative member suggested that this class of compounds inhibits NMDA receptor function in a voltage-independent and noncompetitive manner.³⁰ We confirmed this mechanism for compound 58. Inhibition of GluN1/GluN2D responses by 1 μM 58 was not surmounted by increasing both glutamate and glycine from 30 μ M to 3 mM (4.1 \pm 0.58% of control in 30 μ M, 3.1 \pm 0.50% of control in 3 mM; n = 6, unpaired t test), implying that the compound is noncompetitive at the glutamate and glycine binding sites. Moreover, inhibition produced by 1 μ M 58 was not significantly different at -40 or +30 mV (4.9 \pm 1.3% and 4.8 \pm 1.2% of control, respectively, n = 8, Student's ttest, p = 0.93), suggesting that the receptor blockade by this more potent compound is also voltage-independent. We also examined whether these more potent analogues interacted with the binding site first identified for both the previously described DQP-1105 (Figure 1A) and the quinazolin-4-one (QNZ), QNZ-46.^{30,36} Mutations to the wild type GluN2D receptor (Q801Y, L705F, and A752V), previously shown to decrease sensitivity of the GluN2D receptor to blockade by either DQP-1105 and/or QNZ-46 in the membrane proximal region of the bi-lobed ligand binding domain encoded by the S2 region of the polypeptide chain, were evaluated.^{30,36} A test of the effectiveness of racemic **26** revealed that these mutants each significantly reduced the degree of inhibition, consistent with this compoundnicotinic acetylcholine receptors, which exhibited acting at a similar site as DQP-1105 and QNZ-46 (Figure S1).

Evaluation of Off-Target Effects. We next evaluated the off-target actions for the racemic compounds **26** and **58** in a series of two-electrode voltage-clamp recordings using recombinant ligand-gated ion channels expressed in *Xenopus* oocytes (Table 7, see Experimental Section). Compounds **26** and **58** were tested at 3 μ M on the AMPA receptors (GluA1–

4), kainate receptors (GluK1–2 and GluK2/GluK5), the serotonin receptor (5HT_{3A}), the γ -aminobutyric acid receptors (GABA_A and GABA_C), the glycine receptor (glycine α 1), nicotinic acetylcholine receptors comprising $\alpha_1\beta_1\delta\gamma$, $\alpha_3\beta_4$, $\alpha_4\beta_2$, α_7 , or α_9/α_{10} , and purinergic P2_{X2} receptors. Of the ion channel classes evaluated, compounds **26** and **58** altered agonist-induced currents by less than 10%, with the exception of the nicotinic acetylcholine receptors, which exhibited 13–33% inhibition (Table 7).

We also tested the actions of racemic compounds **26** and **58** at 5 μ M on 42 different ion channels, G-protein-coupled receptors, and transporters via the National Institute of Mental Health (NIMH) psychoactive drug screening program (PDSP; Supporting Information Table S4). The primary binding assay demonstrated that compounds **26** and **58** had a minimal effect on the receptors and transporters, with initial screens showing inhibition of three receptors by **26** (5HT₆, H₂, κ -opioid) and four receptors by **58** (5-HT_{1E}, 5-HT₆, κ -opioid, μ -opioid). For both compounds, the K_i values at these receptors were greater than 10 μ M on all receptors. The data collected from both the two-electrode voltage-clamp experiments and the PDSP demonstrate the utility of this class of compounds as selective inhibitors of the GluN2C- and GluN2D-containing NMDA receptors.

Plasma Stability, Aqueous Solubility, BBB Penetration, and Human Liver Microsomal Stability. Three of the more potent analogues 58, 26, and 64 were evaluated for plasma stability. The compounds showed minimal degradation in human, rat, and mouse plasma over a two hour time-course (Figure S2). The aqueous solubility of compound 26 was evaluated in oocyte recording buffer using nephelometry and assessed to be soluble at >80 μ M (see Experimental Section and Supporting Information p S8).

The topological polar surface area (TPSA) of the carboxylic acid compounds was calculated to be outside the optimal range (<90 Ų) for blood—brain barrier (BBB) penetration.³⁷ However, reduction of the acid to the alcohol moves the properties of this class closer to a typical range for CNS penetration (64, 102.0 Ų, QikProp).³⁷ In order to assess the potential for BBB penetration, compounds 26 and 64 were selected for evaluation in the MDR1-MDCK permeability assay which has been demonstrated to accurately predict BBB penetration because of the overexpression of P-glycoprotein (P-gp) and high transepithelial electrical resistance of the cell line (Table 8).³⁸ As was anticipated with the carboxylic acid containing 26, the potential for BBB penetration was low

Table 8. MDR1-MDCK Permeability^a

			$P_{\rm app} \ (10^{-6} \ {\rm cm/s})$			
test compd	direction	recovery (%)	1	2	av	efflux ratio
64	A to B	43	2.45	2.48	2.46	26
	B to A	73	66.2	62.7	64.5	
26	A to B	73	0.47	0.47	0.47	55
	B to A	76	33.4	17.9	25.6	
68	A to B	43	4.51	3.24	3.88	2.5
	B to A	67	9.32	9.72	9.52	

"The $P_{\rm app}$ and efflux ratio were calculated as described in the Experimental Section. Compounds displaying $P_{\rm app} < 3.0 \times 10^{-6}$ cm/s and an efflux ratio of >10 are interpreted to have a low potential for crossing the BBB. Compounds with $P_{\rm app} > 3.0 \times 10^{-6}$ cm/s and an efflux ratio of <10 are expected to have high brain penetration. ³⁸

(Table 8). The results for the hydroxyl-containing compound 64 also suggested low BBB potential; however, the permeability coefficient ($P_{\rm app}({\rm A-B})$) was much closer to the recommended 3.0×10^{-6} cm/s (64, $P_{\rm app}({\rm A-B})=2.46\times 10^{-6}$ cm/s) than that of the carboxylic acid containing compound (26, $P_{\rm app}({\rm A-B})=0.47\times 10^{-6}$ cm/s), suggesting that efflux may be problematic with this congener. In order to evaluate an analogue with lower TPSA, the monofluoro-containing compound 68 (Table 5, TPSA = 79.08 Ų, QikProp) was assessed in the MDR1-MDCK assay and was classified as being highly brain penetrable ($P_{\rm app}({\rm A-B})=3.88\times 10^{-6}$ cm/s; $P_{\rm app}({\rm B-A})=9.52\times 10^{-6}$ cm/s; Table 8).

The same compounds were also evaluated for metabolic stability using human liver microsomes. While the carboxylic acid containing 26 showed minimal degradation over the 60 min assay, the hydroxyl-containing derivative 64 had a half-life of 13 min (Table 9). The half-life of the monofluoro compound, 68, was determined to be 35 min in the human liver microsomal assay (Table 9). These data suggest that the acyl chain is a candidate for further optimization of desirable pharmacokinetic properties.

DISCUSSION AND CONCLUSION

This study describes the development of potent, selective, and soluble negative allosteric modulators for GluN2C- and GluN2D-containing NMDA receptors that act on the membrane proximal lobe of the GluN2 glutamate binding domain. We describe here several compounds with IC $_{50}$ values in the 100–500 nM range that show 50- to 200-fold selectivity over GluN2A- and GluN2B-containing receptors. We have taken a classical approach to the SAR, allowing proposal of features for a hypothetical pharmacophore.

The A-ring substituents that were explored directly correlate potency with the para- σ substituent coefficients at only GluN2C- and GluN2D-containing receptors, when R₁ was substituted with chloro (Figures 2A and 5A). While we maintained selectivity with many of the analogues, our results suggest that there is a conserved portion of the binding pocket among GluN2A-, GluN2C-, and GluN2D-containing receptors with respect to the para-position of the B-ring; the electronic effects relative to the σ substituent coefficient show similar trends when plotted as a function of potency (Figure 2B,C and Figure 5A). We visualized the electron density of these analogues by carrying out Hartree-Fock calculations using the 6-31G* basis set. While several of the analogues substituted at the para-position of the B-ring could theoretically accept a hydrogen bond, only the bromo- and chloro-containing compounds (26 and 21) exhibit an accessible deficiency of electron density at the terminal position of the substitutions. We hypothesize that the apparent σ hole found at the bromo and chloro atoms could enhance potency by forming a halogen bond with an oxygen electron donor at the receptors (Figure 5B). 39,40 Furthermore, the SAR has revealed that rigidifying the acyl chain in the trans-conformation can enhance potency at the GluN2D-containing receptors, while the length of the chain was found not to be crucial for activity (Figure 5A). The finding that the hydroxyl-containing compound **64** (Table 5) retains both potency and selectivity suggests that the charge on the carboxylic acid is not crucial for either property. While it would be ideal to interpret these data in the context of the receptor, no high quality crystallographic data exist for the region of the LBD where these compounds are thought to act, making docking studies challenging.

Table 9. Human Microsomal Stability

		%					
test compd	0 min	10 min	20 min	30 min	60 min	$half$ - $life^a$ (min)	$CL_{int}^{\ \ b}$ ((mL/min)/mg protein)
64	100	52	31	24	6.5	13	0.110
26	100	101	100	115	86	>60	<0.02
68	100	67	49	49	39	35	0.040

"Half-life was calculated based on $t_{1/2} = 0.693/k$, where k is the elimination rate constant based on the slope of the natural logarithm percent remaining versus incubation time. Intrinsic clearance (CL_{int}) was calculated on $CL_{int} = k/P$, where k is the elimination rate constant and P is the protein concentration in the incubation.

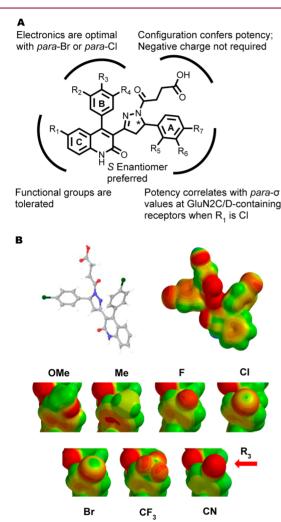


Figure 5. Pharmacophore model and electrostatic potential maps of para-B-ring modifications. (A) The para-substitution of the A-ring shows correlation between the σ substituent constants and activity at GluN2C/D-containing receptors, when R₁ is a chlorine. The length and configuration of the acyl chain is flexible, with the transconfiguration improving potency. B-ring modification shows an optimal para- σ coefficient close to that of chloro and bromo substitutions for GluN2A-, GluN2C-, and GluN2D-containing receptors, suggesting a conserved nature of the binding interaction at each of the three receptors. The C-ring substitutions explored are consistent with this portion of the molecule interacting with a hydrophobic pocket and could allow for improvements in selectivity. (B) The electrostatic potential maps of the para-B-ring modifications evaluated are shown. Only the Cl and Br substituents show significant electron deficiency at the termini of the substituents, suggesting that a potential halogen bond could be responsible for the improved potency of these compounds.

We expect that a reduction in molecular weight in conjunction with further optimization of the topological polar surface area will be required to obtain optimal BBB penetration and pharmacokinetic properties. However, the improvements in potency and selectivity suggest that this class of compounds should be useful as pharmacologic probes to evaluate the contributions of the GluN2C- and GluN2D-containing NMDA receptors in normal and pathophysiologic processes in isolated systems.

The GluN2C- and GluN2D-NMDA receptor subunits remain understudied largely because of a lack of potent and selective pharmacological tools. However, these NMDA receptor subunits reside in a number of brain regions that are highly relevant for neurological disease. For example, expression of functional GluN2D in the subthalamic nuclei raises the possibility that GluN2D-selective inhibitors could attenuate neuronal firing rate and alter firing patterns in subthalamic neurons, which could be of utility in Parkinson's disease. ^{22,24,26,28,41–44} In addition, expression of GluN2Dcontaining receptors in substantia nigra pars compacta neurons raises the possibility that GluN2D-selective antagonists might possess neuroprotective properties in Parkinson's disease by diminishing Ca2+ influx into the dopaminergic substantia nigra pars comapcta neurons, which may lead to neuronal death. 8,45 GluN2C is expressed widely in the cerebellum and has also been suggested to have a role in both emotional learning and schizophrenia. 46-48 The compounds described here could therefore be tools with which to evaluate GluN2C- and GluN2D-containing receptor function in specific circuits implicated in these conditions.

■ EXPERIMENTAL SECTION

Biology. Two-Electrode Voltage-Clamp Electrophysiology. The Emory University Institutional Animal Care and Use Committee approved all protocols involving the use of animals. Xenopus laevis oocytes were isolated and maintained as previously described.³⁶ The cDNAs for the desired NMDA receptor subunits (GenBank accession numbers U11418 and U08261; GluN1, D13211; GluN2A, U11419; GluN2B, M91563; GluN2C, L31611; GluN2D) were obtained from Drs. S. Heinemann (Salk Institute), S. Nakanishi (Kyoto University), and P. Seeburg (University of Heidelberg). Preparation of cRNA, injection of oocytes with RNA, and two-electrode voltage clamp recordings have been described elsewhere. 30,36,49 Briefly, oocytes were placed in a perfusion chamber with recording solution comprising (in mM) 90 NaCl, 1 KCl, 0.5 BaCl₂, 0.005 EDTA, and 10 HEPES, with pH adjusted to 7.4 with NaOH at 23 $^{\circ}$ C. The glass electrodes used had tip resistances of 0.5–2.0 M Ω and were filled with 0.3–3.0 M KCl. Compounds were made as 20 mM stock solutions in DMSO and diluted to final concentrations in recording solution (final DMSO was 0.1-0.05% v/v). The current recordings were performed using a Warner OC-725B or -C amplifier at a holding potential of -40 mV.

Subunit selectivity was determined by recording from various ligand-gated ion channels expressed in *Xenopus laevis* oocytes as previously described. 50 The cDNA encoding GABA_A, GABA_C, and

glycine receptor cDNAs were provided by Dr. Weiss (University of Texas Health Science Center at San Antonio). Nicotinic acetylcholine and 5-HT_{3A} serotonin receptor cDNAs were provided by Drs. Papke and Heinemann (University of Florida and Salk Institute), and purinergic receptor cDNA was provided by Dr. Hume (University of Michigan). The glutamate receptors GluA1-4, GluK1, and GluK2 were activated by 100 µM glutamate. GluK1 and GluK2 expressing oocytes were incubated for 5 min in 1 mg/mL concanavalin A prior to recording. The GluK2/5 receptor was activated with 100 µM AMPA (2-amino-3-(3-hydroxy-5-methylisoxazol-4-yl)propanoic acid). The GABA_A and GABA_C receptors were activated by 20 and 2 μ M γ aminobutyric acid, respectively. Acetylcholine was used activate the nicotinic acetylcholine $\alpha_1\beta_1\delta\gamma$ (1 μ M), $\alpha_3\beta_4$ (10 μ M), $\alpha_2\beta_4$ (10 μ M), α_7 (300 μ M), $\alpha_9\alpha_{10}$ (10 μ M) receptors. The glycine α_1 and 5-HT_{3A} currents were evoked by 50 μ M glycine receptor and 3 μ M serotonin, respectively. The human and rat P2_{X2} purinergic receptors were activated with 9 µM adenosine triphosphate.

The receptor binding profiles (Supporting Information Table S2) and K_i determinations for compounds **26** and **58** were generously provided by the National Institute of Mental Health psychoactive drug screening program, Contract HHSN-271-2008-025C (NIMH PDSP). The NIMH PDSP is directed by Bryan L. Roth M.D., Ph.D. at the University of North Carolina at Chapel Hill and Project Officer Jamie Driscoll at NIMH, Bethesda, MD, U.S. Data were collected using 5 μ M **26** and **58**.

MDR1-MDCK Permeability. Cell monolayers were grown to confluence on collagen-coated, microporous, polycarbonate membranes in 12-well Costar Transwell plates. The permeability assay buffer was Hanks' balanced salt solution containing 10 mM HEPES and 15 mM glucose at pH 7.4. The buffer in the receiver chamber also contained 1% bovine serum albumin. The dosing solution concentration in the assay buffer was 5 μ M for each compound tested. The cell monolayers were dosed on the apical side (A to B) or basolateral side (B to A) and incubated at 37 °C with 5% CO₂ in a humidified incubator. Samples were taken from the donor and receiver chambers at 120 min. Each determination was performed in duplicate. All samples were assayed by LC-MS/MS using electrospray ionization. The apparent permeability ($P_{\rm app}$) and percent recovery were calculated as follows.

$$P_{\rm app} = (dC_{\rm r}/dt)V_{\rm r}/(AC_{\rm A}) \tag{1}$$

percent recovery =
$$100 \times [(V_r C_r^{\text{final}}) + (V_d C_d^{\text{final}}) + (V_d C_d^{\text{final}})]$$

/ $(V_d C_N)$ (2)

where $\mathrm{d}C_r/\mathrm{d}t$ is the slope of the cumulative concentration in the receiver compartment versus time in $\mu\mathrm{M}$ s⁻¹, V_r is the volume in the receiver compartment in cm³, V_d is the volume in the donor compartment in cm³, V_d is the area of the insert (1.13 cm² for 12-well Transwell), V_d is the average of the nominal dosing concentration and the measured 120 min donor concentration in $\mu\mathrm{M}$, V_d is the nominal concentration of the dosing solution in v_d , v_d is the cumulative receiver concentration in v_d at the end of the incubation period, and v_d is the concentration of the donor in v_d at the end of the incubation period. The MDR1-MDCK permeability assays were performed by Absorption Systems.

Human Liver Microsomal Stability. Human liver microsomes were obtained from XenoTech. The reaction mixture was prepared with 0.5 mg/mL human liver microsomes, 100 mM potassium phosphate (pH 7.4), 5 mM magnesium chloride, and 1 μ M test compound. The reaction mixture was incubated in a shaking water bath at 37 °C for 3 min prior to the addition of NADPH (1 mM). Testosterone was run simultaneously in a separate vessel as a control. Then 100 μ L aliquots were taken at 0, 10, 20, 30, and 60 min for both test compound and testosterone. The aliquots were combined immediately with 400 μ L of ice cold 50/50 acetonitrile/deionizd H_2O containing 0.1% formic acid and internal standard to terminate the reaction. The samples were then mixed and centrifuged to precipitate microsomal proteins. All samples were assayed by LC–MS/MS using electrospray ionization and multiple reaction monitor-

ing, and the peak area responses to internal standard of the compounds at each time point were compared to the peak area response at time 0 to determine the percent compound remaining. The human liver microsomal stability assays were performed by Absorption Systems.

Data Analysis. Potency of compounds was assessed by fitting the composite concentration—response curve obtained from the average of multiple recordings with the equation

response =
$$100/\{1 + [(inhibitor concentration)/IC_{50}]^N\}$$
 (3)

where ${\rm IC}_{50}$ is the concentration of compound that is required to inhibit the response half-maximally and N is the Hill slope. Saturating conditions were assumed to give complete inhibition. Data for compounds that did not inhibit the steady-state current by at least 30% were not fit by the above equation and are designated as NE in the data tables.

Chemistry. Commercial vendors provided compounds 4, 5, 9-11, 14, 16, 17, 20, 57, and 62, which are not described below and which were ≥90% purity, as provided by the vendor or determined independently as below. All reagents were obtained from commercial suppliers and used without further purification unless otherwise noted. Reaction progress was monitored by thin layer chromatography (TLC) on precoated glass plates (silica gel 60 F254, 0.25 mm). Proton, carbon, and fluorine NMR spectra were recorded on INOVA-400 (400 MHz), VNMRS 400 (400 MHz), UNITY-600 (600 MHz), or INOVA-600 (600 MHz) instrument. Proton and carbon spectra were referenced to the residual solvent peak, while fluorine spectra were referenced to trifluoroacetic acid residual peak. The Emory University Mass Spectrometry Center collected mass spectral data on either a VG 70-S Nier Johnson or JEOL instrument. Compound purity was assessed by reverse phase liquid chromatography using an Agilent Zorbax, 3.5 μ m, XD B-C18 column, 4.6 mm \times 50 mm (254 nm), or by elemental analyses, performed by Atlantic Microlab Inc. Purity for all compounds synthesized and tested was at or above 95% unless otherwise noted. Flash chromatography was performed on a Teledyne ISCO Combiflash Companion with prepackaged disposable normal phase silica columns.

Computational Analysis. Energy minimized conformations of the compounds being analyzed were generated using the OPLS_2005 force field in MacroModel (*MacroModel*, version 9.9; Schrödinger, LLC: New York, NY, 2011). The TPSA approximations were obtained from QikProp (*QikProp*, version 3.4; Schrödinger, LLC: New York, NY, 2011) using energy minimized conformations, as above. The Hartree–Fock calculations were carried out using the neutral compounds and the 6-31G* basis set in Spartan '10 (*Spartan '10*; Wavefunction, Inc.: Irvine, CA). The equilibrium geometry at the ground state in vacuum was calculated. For visualization purposes, the energy range in the electrostatic potential maps was limited from –100.00 to 280.00 kJ/mol.

Separation and X-ray Crystallography of Enantiomers. Separation of the final compounds used for biological testing from the racemic 26 was obtained using a ChiralPak OD-RH 30 mm × 250 mm, 5 μ m column using the following conditions: flow rate 10 mL/ min, injection volume 4-6 mL (2 mg/mL), 60% acetonitrile (0.1% formic acid)/40% H₂O (0.1% formic acid); **69** t_R = 21.8; **70** t_R = 25.1 min. The enantiomeric excess (ee) of the enantiomers 69 and 70 was determined using an Agilent 1200 HPLC pump on a ChiralPak OD-RH column (4.6 mm \times 150 mm, 5 μ m) using the following conditions: flow rate 0.5 mL/min, injection volume 10 μ L, 60% acetonitrile (0.1% formic acid)/40% H₂O (0.1% formic acid); 69 (S-**26**, $[\alpha]_D^{20}$ –34.0 (c = 0.32, chloroform), t_R = 7.47 min, 100% ee. **70** (R-**26**) $[\alpha]_D^{20} + 36.0$ (c = 0.25, MeOH), $t_R = 8.79$ min, 98% ee. Optical rotation data were collected using a Perkin-Elmer 314 instrument. The proton NMR spectrum was identical to that of racemic 26 for each enantiomer. Single crystals of the second peak to elute from the separation of racemic 26 (70 $t_R = 25.1$ min) were grown by slow evaporation of a solution of the compound in a mixture of methanol and water. Crystal data for $C_{28}H_{23}Cl_2N_3O_5$ (M = 552.39): 1.124 × 0.087×0.056 , orthorhombic, space group $P2_12_12_1$ (No. 19), a = 8.0529(5) Å, b=10.2097(5) Å, c=31.2978(13) Å, V=2573.2(2) ų, Z=4, $\mu(\text{Mo K}\alpha)=0.315~\text{mm}^{-1}$, $D_{\text{calc}}=1.490~\text{g/mm}^3$, temperature 173 K. Intensity data were collected on a Bruker APEX II CCD diffractometer with monochromated Mo K α radiation ($\lambda=0.710.73$ Å) at 173 K in the 2θ range $2.6-53.4^\circ$. The user interface Olex2 was used for the crystallographic calculations and crystal structure visualization. The structure was solved with Superflip by charge flipping and refined by least-squares minimization using SHELXL. A total of 15.745 reflections were measured ($2.602 \le 2\theta \le 53.41$), while 5408 unique data ($R_{\text{int}}=0.124$) were used in the refinements. The final R_1 was $0.0590~(I>2\sigma(I))$ and the weighted R value wR2 was 0.0874~(all data).

General Procedure for the Synthesis of Acylated Quinolone Pyrazoline Products. Procedure G. In an appropriately sized microwaveable vessel, the pyrazol-3-ylquinolin-2(1H)-one intermediate (1.00 equiv) was dissolved in anhydrous tetrahydrofuran (THF) (0.15 M) with 4 Å molecular sieves present. The appropriate anhydride (1.00 equiv) was added. The solution was microwaved (Biotage Initiator) with stirring for 20 min at 165 °C. The THF was removed under vacuum, and the organics were dissolved in dichloromethane (DCM), washed three times with acidified (pH 2, HCl) brine, dried over magnesium sulfate, filtered, concentrated under reduced pressure, and subjected to flash column chromatography using a 0–10% MeOH/DCM gradient unless otherwise noted.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(4-fluorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (2). Compound 2 was prepared according to general procedure G using succinic anhydride (88) (0.062 g, 0.622 mmol) and 2f (0.260 g, 0.622 mmol). After removal of the THF, the residue was dissolved in hot EtOAc and small portions of hexanes were added until a solid began to form. The solid was filtered and column-chromatographed using 0-10% MeOH/DCM, and the title compound was obtained as a yellow solid. Yield 0.236 g, 73.2%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.42 (s, 1H), 12.15 (s, 1H), 7.68–7.38 (m, 6H), 7.27 (d, J = 7.4 Hz, 1H), 7.04 (td, J = 8.8, 2.4 Hz, 2H), 6.93 (d, J = 2.5 Hz, 1H), 6.85-6.76(m, 2H), 5.35-5.30 (m, 1H), 3.80-3.67 (m, 1H), 2.83-2.74 (m, 1H), 2.48-2.39 (m, 2H), 2.34-2.25 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.59, 168.66, 160.13, 152.45, 149.96, 138.39, 137.34, 134.56, 131.25, 129.47, 128.51, 127.55, 126.09, 124.64, 120.68, 117.65, 115.26, 115.05, 58.24, 45.23, 28.92, 28.59, 28.22. ¹⁹F NMR (376 MHz, DMSO- d_6) δ -116.13 to -116.20 (m). HRMS (m/z): [M - H] calcd for C28H20ClN3O4F, 516.11319; found, 516.11246. Anal. Calcd for C₂₈H₂₁ClN₃O₄F: C, 64.93; H, 4.09; N, 8.11. Found: C, 61.01; H, 4.22; N, 7.15. HPLC 85% MeOH/ H_2O (0.1% formic acid) $t_R = 0.95$ min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 0.72$ min; >95% purity.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (3). Compound 3 was prepared according to general procedure G using 87 (0.046 g, 0.460 mmol) and 3f (0.200 g, 0.460 mmol). The title compound was obtained by removing the THF, dissolving the crude mixture into hot EtOAc, and adding small portions of hot hexanes until a yellow precipitate formed. The mixture was allowed to cool and filtered to give the title compound as a yellow solid. Yield 0.187 g, 76.0%. ¹H NMR (400 MHz DMSO- d_6) δ 12.42 (s, 1H), 12.15 (s, 1H), 7.64 (dd, J = 8.7, 2.5 Hz, 1H), 7.61–7.38 (m, 5H), 7.27 (d, J =7.9 Hz, 3H), 6.94 (d, J = 2.5 Hz, 1H), 6.79 (d, J = 8.2 Hz, 2H), 5.33 (dd, J = 12.1, 4.7 Hz, 1H), 3.75 (dd, J = 18.5, 12.0 Hz, 1H), 2.78 (dd, J = 18.4, 4.7 Hz, 1H), 2.49-2.23 (m, 4H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.55, 168.67, 160.09, 152.42, 149.96, 141.13, 137.32, 134.54, 131.59, 131.24, 129.47, 128.57, 128.46, 128.37, 127.38, 126.13, 126.06, 124.56, 120.66, 117.64, 58.29, 45.12, 28.57, 28.23. HRMS (*m*/ z): $[M + H]^+$ calcd for $C_{28}H_{22}Cl_2N_3O_4$, 534.09819; found, 534.09774. Anal. Calcd for C₂₈H₂₁Cl₂N₃O₄·0.40H₂O: C, 62.09; H, 4.06; N: 7.76. Found: C, 61.90; H, 4.13; N: 7.64.

4-[1-(3-Carboxypropanoyl)-3-(6-chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1*H*-pyrazol-5-yl]benzoic Acid (6). Compound 6 was prepared according to general procedure G using 87 (0.045 g, 0.451 mmol) and 6f (0.200 g, 0.451 mmol). The solvent was removed, and the product was obtained by dissolving in

hot EtOAc and adding hexanes until a solid began to precipitate. The solution was cooled and the product filtered to yield a white solid. Yield 0.040 g, 16.3%. $^1{\rm H}$ NMR (400 MHz, DMSO- d_6) δ 12.42 (bs, 3H), 7.79 (d, J=8.0 Hz, 2H), 7.64 (dd, J=8.7, 2.4 Hz, 1H), 7.60—7.41 (m, 5H), 7.27 (d, J=7.4 Hz, 1H), 6.96—6.82 (m, 3H), 5.39 (dd, J=12.0, 4.8 Hz, 1H), 3.79 (dd, J=18.4, 12.2 Hz, 1H), 2.79 (dd, J=18.4, 4.7 Hz, 1H), 2.61—2.37 (m, 2H), 2.30 (t, J=6.9 Hz, 2H). $^{13}{\rm C}$ NMR (150 MHz, DMSO- d_6) δ 173.51, 172.19, 168.72, 167.04, 160.09, 152.41, 149.99, 146.92, 137.33, 134.56, 131.25, 129.68, 129.55, 129.51, 128.52, 128.48, 128.38, 126.12, 126.07, 125.54, 124.53, 120.68, 117.65, 58.71, 45.12, 28.53, 28.22. HRMS (m/z): [M + Na]+ calcd for $\rm C_{29}H_{22}{\rm ClN}_3O_6Na$, 566.10893; found, 566.10923. HPLC 85% MeOH/ $\rm H_2O$ (0.1% formic acid) $t_{\rm R}=0.71$ min; >95% purity; 75% ACN/H₂O (0.1% formic acid) $t_{\rm R}=0.53$ min; >95% purity.

4-{3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-[4-(methoxycarbonyl)phenyl]-4,5-dihydro-1H-pyrazol-1-yl}-4oxobutanoic Acid (7). Compound 7 was prepared according to general procedure G using 87 (0.055 g, 0.546 mmol) and 7f (0.250 g, 0.546 mmol). The title compound was purified using flash chromatography (2-10% MeOH/DCM), followed by precipitation from hot EtOAc using hot hexanes. Yield 0.040 g, 13.1%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.38 (s, 1H), 12.06 (s, 1H), 7.84–7.72 (m, 2H), 7.64-7.35 (m, 6H), 7.22 (d, J = 7.4 Hz, 1H), 6.87 (d, J = 8.4 Hz, 3H), 5.36 (dd, J = 12.3, 4.6 Hz, 1H), 3.90–3.69 (m, 4H), 2.74 (dd, J =18.5, 4.6 Hz, 1H), 2.51-2.33 (m, 2H), 2.30-2.21 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.53, 168.77, 165.96, 160.10, 152.43, 150.02, 147.43, 137.34, 134.59, 131.29, 129.44, 128.41, 126.14, 125.76, 124.50, 120.67, 117.67, 58.68, 52.16, 45.09, 28.52, 28.22. HRMS (m/ z): [M + Na]⁺ calcd for C₃₀H₂₄ClN₃O₆Na, 580.12458; found, 580.12484. Anal. Calcd for C₃₀H₂₄ClN₃O₆·0.80H₂O: C, 62.95; H, 4.51; N, 7.34. Found: C, 63.04; H, 4.55; N, 7.36.

4-{3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-[4-(trifluoromethyl)phenyl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (8). Compound 8 was prepared according to general procedure G using 87 (0.043 g, 0.427 mmol) and 8f (0.200 g, 0.427 mmol). The title compound was obtained after flash chromatography (2-10% MeOH/DCM) followed by trituration from EtOAc as a yellow solid. Yield 0.030 g, 12.4%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.42 (s, 1H), 12.04 (s, 1H), 7.65 (dd, J = 8.7, 2.4 Hz, 1H), 7.61-7.40 (m, 7H), 7.26 (d, J = 7.4 Hz, 1H), 6.99 (d, J = 8.0 Hz, 2H), 6.93 (d, J = 8.0 H = 2.3 Hz, 1H), 5.43 (dd, J = 12.2, 4.7 Hz, 1H), 3.79 (dd, J = 18.5, 12.2)Hz, 1H), 2.80 (dd, J = 18.6, 4.7 Hz, 1H), 2.60–2.40 (m, 2H), 2.29 (t, J= 6.8 Hz, 2H). 13 C NMR (150 MHz, DMSO- d_6) δ 174.14, 169.46, 160.72, 153.09, 150.68, 147.32, 138.01, 135.23, 131.92, 130.09, 129.22, 129.12, 129.05, 126.88, 126.79, 126.72, 126.05, 125.13, 121.31, 118.31, 59.19, 45.74, 29.19, 28.89. HRMS (m/z): $[M + H]^+$ calcd for $C_{29}H_{22}ClN_3O_4F_3,\ 568.12455;\ found,\ 568.12554.$ C₂₉H₂₁ClN₃O₄F₃·0.07EtOAc; HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 1.1 \text{ min}$; >95% purity; 75% ACN/H₂O (0.1% formic acid) t_R = 0.86 min; >95% purity.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(4-methoxyphenyl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (9). Compound 9 was prepared according to general procedure G using 87 (0.058 g, 0.582 mmol) and 9f (0.250 g, 0.582 mmol). After removal of the THF, the title compound was obtained by dissolving the crude mixture into hot EtOAc and adding small portions of hot hexanes until a yellow precipitate formed. The mixture was allowed to cool and filtered to give the title compound as a yellow solid. Yield 0.132 g, 42.8%. 1 H NMR (400 MHz, DMSO- d_6) δ 12.40 (s, 1H), 12.16 (s, 1H), 7.64 (dd, J = 8.9, 2.4 Hz, 1H), 7.60–7.49 (m, 3H), 7.45 (d, J = 8.7 Hz, 1H), 7.43-7.38 (m, 1H), 7.29 (d, J = 7.0 Hz, 1H), 6.93 (d, I = 2.5 Hz, 1H), 6.82-6.66 (m, 4H), 5.24 (dd, I = 12.1, 4.6 Hz, 1H), 3.89–3.62 (m, 4H), 2.78 (dd, J = 18.3, 4.6 Hz, 1H), 2.45 (t, J = 7.0 Hz, 2H), 2.27 (t, J = 6.9 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.58, 168.49, 160.14, 158.22, 152.39, 149.86, 137.30, 134.58, 134.31, 131.22, 129.42, 128.57, 128.43, 128.35, 126.77, 126.09, 126.02, 124.74, 120.71, 117.63, 113.71, 58.47, 55.10, 45.27, 28.63, 28.24. HRMS (m/z): $[M + Na]^+$ calcd for $C_{29}H_{24}ClN_3O_5Na$, 552.12967; found, 552.13018. Anal. Calcd for

C₂₉H₂₄ClN₃O₅·1.00H₂O: C, 63.56; H, 4.78; N, 7.67. Found: C, 63.68; H, 4.57; N, 7.59.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroguinolin-3-yl)-5-(3-fluorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (12). Compound 12 was prepared according to general procedure G using 87 (0.120 g, 1.20 mmol) and 12f (0.500 g, 1.20 mmol). After removal of the THF, the title compound was obtained by precipitation from EtOAc using hexanes, as a yellow solid. Yield 0.380 g, 62%. 1 H NMR (400 MHz, DMSO- d_{6}) δ 12.42 (s, 1H), 12.15 (s, 1H), 7.65 (dd, J = 8.7, 2.6 Hz, 1H), 7.53 (m, 2H), 7.44 (m, 3H), 7.26 (d, J = 7.0 Hz, 2H), 7.03 (t, J = 8.7 Hz, 1H), 6.93 (d, J = 2.3 Hz, 1H),6.64 (dd, J = 12.3, 8.8 Hz, 2H), 5.35 (dd, J = 12.3, 4.4 Hz, 1H), 3.75(dd, J = 18.6, 12.1 Hz, 1H), 2.81 (dd, J = 17.7, 4.7 Hz, 1H), 2.60-2.23(m, 4H). 13 C NMR (100 MHz, DMSO- d_6) δ 173.57, 168.75, 163.36, 160.94, 160.12, 152.47, 149.98, 144.95, 144.88, 137.33, 134.56, 131.25, 130.52, 130.45, 129.40, 128.56, 128.42, 128.38, 128.29, 126.14, 126.07, 124.53, 121.50, 120.65, 117.65, 114.00, 113.79, 112.34, 112.13, 58.41, 45.14, 28.53, 28.20. HRMS (m/z): $[M - H]^-$ calcd for C₂₈H₂₀ClN₃O₄F; 516.11319; found, 516.11239. Anal. Calcd for C₂₈H₂₁ClN₃O₄F·0.40H₂O: C, 64.93; H, 4.09; N, 8.11. Found: C, 64.11; H, 4.03; N, 7.94. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_{\rm R} = 0.97 \text{ min}; >95\% \text{ purity}; 75\% \text{ ACN/H}_2\text{O} (0.1\% \text{ formic acid}) t_{\rm R} =$ 0.73 min; >95% purity.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroguinolin-3-yl)-5-(3-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (13). Compound 13 was prepared according to general procedure G using 87 (0.046 g, 0.460 mmol) and 13f (0.200 g, 0.460 mmol). After removal of the THF, the title compound was obtained by precipitating from EtOAc using hexanes, as a yellow solid. Yield 0.156 g, 63.4%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.39 (s, 1H), 12.04 (s, 1H), 7.64–7.57 (m, 1H), 7.56–7.34 (m, 5H), 7.27–7.17 (m, 3H), 6.97-6.87 (m, 2H), 6.68 (d, J = 6.8 Hz, 1H), 5.29 (dd, J = 12.2, 4.8 Hz, 1H), 3.71 (dd, J = 18.5, 12.0 Hz, 1H), 2.84-2.73 (m, 1H), 2.50-2.32 (m, 2H), 2.25 (t, J = 6.7 Hz, 2H). 13 C NMR (100 MHz, DMSO- d_6) δ 173.55, 168.78, 152.53, 150.00, 144.54, 137.35, 134.62, 133.05, 130.47, 129.37, 128.60, 128.45, 127.14, 126.15, 125.51, 124.51, 124.11, 120.64, 58.40, 45.10, 28.53, 28.18. HRMS (m/z): [M + Na] calcd for C₂₈H₂₁Cl₂N₃O₄Na; 556.08013; found, 556.07988. Anal. $\label{eq:calcd} \text{Calcd for } C_{28}H_{21}\text{Cl}_2N_3O_4\cdot 0.20H_2O\text{: } C\text{, } 61.68\text{; } H\text{, } 4.10\text{; } N\text{, } 7.71\text{.}$ Found: C, 61.74; H, 4.27; N, 7.32.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-1-yl]-4-oxobutanoic Acid (15). Compound 15 was prepared according to general procedure G using 87 (0.029 g, 0.29 mmol) and 15f (0.13 g, 0.29 mmol). The title compound was purified using flash chromatography (2-10% MeOH/DCM) and isolated as a yellow solid. Yield 0.090 g, 58.7%. 1 H NMR (400 MHz, DMSO- d_{6}) δ 12.41 (s, 1H), 12.08 (s, 1H), 7.64 (dt, J = 8.8, 2.1 Hz, 1H), 7.58–7.42 (m, 4H), 7.39 (d, J = 7.3Hz, 1H), 7.27 (d, J = 7.3 Hz, 1H), 7.13 (t, J = 7.9 Hz, 1H), 6.93 (d, J =2.2 Hz, 1H), 6.77 (dd, J = 8.2, 2.4 Hz, 1H), 6.59 (s, 1H), 6.32 (d, J = 7.7 Hz, 1H), 5.27 (dd, J = 12.0, 4.7 Hz, 1H), 3.77–3.63 (m, 4H), 2.85 (dd, J = 18.4, 4.7 Hz, 1H), 2.48-2.36 (m, 2H), 2.28 (t, J = 6.8 Hz, 1.48 Hz)2H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.54, 168.64, 160.16, 159.25, 152.47, 149.91, 143.75, 137.32, 134.64, 131.21, 129.59, 129.15, 128.59, 128.48, 128.37, 128.30, 126.09, 124.56, 120.68, 117.63, 117.39, 112.37, 111.35, 58.88, 54.98, 45.23, 28.56, 28.22. HRMS (m/z): [M -H]⁻ calcd for C₂₉H₂₃ClN₃O₅, 528.13317; found, 528.13351. Anal. Calcd for C₂₉H₂₃Cl₁N₃O₅·0.70H₂O: C, 64.32; H, 4.54; N, 7.76. Found: C, 64.38; H, 4.67; N: 7.67.

4-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(2-chlorophenyl)-4,5-dihydro-1H-pyrazol-1-yl]-4-oxobutanoic Acid (18). Compound 18 was prepared according to general procedure G using 87 (0.046 g, 0.460 mmol) and 18f (0.200 g, 0.460 mmol). After removal of the THF, the title compound was obtained by precipitation from EtOAc using hexanes, as a yellow solid. Yield 0.136 g, 55.3%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.40 (s, 1H), 12.13 (s, 1H), 7.63 (dd, J= 8.6, 1.6 Hz, 1H), 7.56–7.49 (m, 2H), 7.45–7.41 (m, 4H), 7.26 (t, J= 7.6 Hz, 1H), 7.19–7.15 (m, 2H), 6.93 (s, 1H), 6.41 (d, J= 7.4 Hz, 1H), 3.87 (dd, J=18.4, 12.4 Hz, 1H), 2.75 (dd, J= 18.2, 4.7 Hz, 1H), 2.60–2.54 (m, 1H), 2.46–2.32 (m, 2H), 2.39–2.30 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.55,

168.78, 160.02, 152.56, 150.10, 138.66, 134.68, 131.27, 130.52, 129.36, 128.78, 128.37, 128.58, 126.13, 125.90, 124.33, 120.66, 117.64, 56.40, 44.20, 28.43, 28.22. HRMS (m/z): $[M + Na]^+$ calcd for $C_{28}H_{21}Cl_2N_3O_4Na$, 556.08013; found 556.08038. Anal. Calcd for $C_{28}H_{21}Cl_2N_3O_4\cdot 0.30H_2O$: C, 62.30; H, 4.03; N, 7.78. Found: C, 62.27; H, 4.32; N, 7.49.

4-[5-(2-Bromophenyl)-3-(6-chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (19). Compound 19 was prepared according to general procedure G using 87 (0.063 g, 0.627 mmol) and 19f (0.300 g, 0.627 mmol). After removal of the THF, the title compound was obtained by precipitation from EtOAc and hexanes, as a yellow solid. Yield 0.140 g, 38.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.41 (s, 1H), 12.05 (s, 1H), 7.64 (dd, J = 8.9, 2.5 Hz, 1H), 7.59 (d, J = 7.5 Hz, 1H), 7.53 (p, J = 7.6 Hz, 2H), 7.42 (dd, J = 11.1, 7.9 Hz, 3H), 7.26–7.15 (m, 3H), 6.93 (d, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 5.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 5.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.48 (dd, J = 2.3 Hz, 1H), 6.39-6.32 (m, 1H), 6.39-6.12.1, 4.6 Hz, 1H), 3.87 (dd, *J* = 18.3, 12.0 Hz, 1H), 2.73 (dd, *J* = 18.4, 4.6 Hz, 1H), 2.62-2.51 (m, 1H), 2.49-2.28 (m, 3H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.53, 168.76, 160.06, 152.42, 150.11, 140.19, 137.33, 132.58, 131.25, 129.08, 128.41, 126.12, 124.30, 120.64, 117.63, 58.69, 44.33, 28.41, 28.21. HRMS (m/z): $[M + Na]^+$ calcd for C28H21ClBrN3O4Na; 600.02962, found 600.02945. Anal. Calcd for C₂₈H₂₁ClBrN₃O₄: C, 58.10; H, 3.66; N, 7.26. Found: C, 54.48; H, 3.54; N, 6.55. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 1.17$ min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 1.17$ min;

4-{5-(4-Bromophenyl)-3-[4-(4-bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (21). Compound 21 was prepared according to general procedure G using 87 (0.057 g, 0.573 mmol) and 21f (0.300 g, 0.573 mmol). There was a yellow solid present in the reaction vessel which was filtered, dried, and determined to be the title compound. Yield 0.320 g, 90%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.30 (s, 1H), 12.18 (s, 1H), 7.75 (d, J = 8.2 Hz, 1H), 7.66 (d, J = 8.2 Hz, 1H), 7.58 (t, J =7.8 Hz, 1H), 7.48–7.39 (m, 3H), 7.36 (d, J = 8.1 Hz, 1H), 7.22 (d, J =8.1 Hz, 1H), 7.15 (t, J = 7.7 Hz, 1H), 7.03 (d, J = 8.2 Hz, 1H), 6.74 (d, J = 7.9 Hz, 2H), 5.33 (dd, J = 11.8, 4.4 Hz, 1H), 3.74 (dd, J = 18.7, 12.1 Hz, 1H), 2.76 (dd, J = 18.5, 4.4 Hz, 1H), 2.65–2.39 (m, 2H), 2.39–2.30 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.52, 168.65, 160.14, 152.69, 149.89, 141.52, 138.55, 134.40, 131.73, 131.45, 131.24, 130.82, 127.63, 127.28, 123.41, 122.41, 121.82, 120.09, 119.07, 115.59, 58.28, 45.14, 28.49, 28.23. HRMS (m/z): $[M - H]^-$ calcd for C₂₈H₂₀Br₂N₃O₄, 619.98260; found, 619.98231. Anal. Calcd for C₂₈H₂₁Br₂N₃O₄: C, 53.96; H, 3.40; N, 6.74. Found: C, 52.23; H, 3.67; N, 5.55. HPLC 85% MeOH/H₂O 0.1% formic acid) $t_R = 1.2$ min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 0.84$ min; >94% purity.

4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroguinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (22). Compound 22 was prepared according to general procedure G using 87 (0.105 g, 1.04 mmol) and 22f (0.500 g, 1.044 mmol). After removal of the THF, the residue was partitioned between EtOAc and acidified brine. The organics were washed three times, dried over magnesium sulfate, and concentrated under vacuum. The title compound was obtained after column chromatography using 10% MeOH/DCM. Yield 0.240 g, 39.7%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.05 (s, 1H), 7.74 (dd, J = 8.1, 2.2 Hz, 1H), 7.65 (dd, J = 8.1, 2.2 Hz, 1H), 7.58 (t, J = 7.6 Hz, 1H), 7.42 (d, J= 8.3 Hz, 1H), 7.36 (dd, J = 8.1, 2.3 Hz, 1H), 7.34-7.25 (m, 2H), 7.22(dd, J = 8.2, 2.3 Hz, 1H), 7.15 (t, J = 7.6 Hz, 1H), 7.03 (d, J = 8.1 Hz,1H), 6.85-6.75 (m, 2H), 5.35 (dd, J = 11.9, 4.4 Hz, 1H), 3.73 (dd, J =18.4, 12.1 Hz, 1H), 2.76 (dd, J = 18.6, 4.4 Hz, 1H), 2.65–2.42 (m, 2H), 2.34 (t, J = 6.7 Hz, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.54, 168.69, 160.17, 152.70, 149.90, 141.10, 138.58, 134.42, 131.61, 131.45, 131.27, 130.84, 128.34, 127.30, 123.44, 122.42, 119.08, 115.60, 58.23, 45.16, 28.48, 28.23. HRMS (m/z): $[M - H]^-$ calcd for C₂₈H₂₀BrClN₃O₄, 576.03312; found, 576.03267. Anal. Calcd for C₂₈H₂₁BrClN₃O₄·0.50H₂O: C, 57.21; H, 3.77; N, 7.15. Found: C, 57.02; H, 3.72; N, 7.05.

4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-fluorophenyl)-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (23). Compound 23 was prepared according to general procedure G using 87 (0.054 g, 0.541 mmol) and 23f (0.250 g, 0.541 mmol). The title compound, a yellow solid, was obtained after purifying using flash chromatography (0-10% MeOH/DCM). Yield 0.100 g, 32.9%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.11 (s, 1H), 7.74 (d, J = 8.0 Hz, 1H), 7.66 (d, J = 8.2 Hz, 1H), 7.58 (t, J = 7.7 Hz, 1H), 7.43 (d, J = 8.3 Hz, 1H), 7.36 (d, J = 8.0 Hz, 1H),7.23 (d, J = 8.2 Hz, 1H), 7.15 (t, J = 7.6 Hz, 1H), 7.05 (t, J = 8.9 Hz, 3H), 6.83 (dd, J = 8.4, 5.2 Hz, 2H), 5.35 (dd, J = 12.0, 4.4 Hz, 1H), 3.72 (dd, I = 18.5, 12.0 Hz, 1H), 2.78 (dd, I = 18.4, 4.4 Hz, 1H), 2.65-2.43 (m, 2H), 2.34 (t, J = 6.7 Hz, 2H). ¹³C NMR (100 MHz, DMSO d_6) δ 173.56, 168.64, 160.16, 152.71, 149.87, 138.56, 138.36, 134.42, 131.69, 131.46, 131.24, 130.86, 127.46, 127.38, 127.29, 123.47, 122.43, 121.80, 119.08, 115.59, 115.18, 114.96, 58.15, 45.26, 28.50, 28.23. ¹⁹F NMR (376 MHz, DMSO- d_6) δ –116.06 to –116.19 (m). HRMS (m/ z): [M + H]⁺ calcd for C₂₈H₂₂BrFN₃O₄, 562.07722; found, 562.07669. Anal. Calcd for C₂₈H₂₁BrFN₃O₄: C, 59.80; H, 3.76; N, 7.47. Found: C, 50.97; H, 3.55; N, 6.00. HPLC 75-95% MeOH/H₂O (0.1% formic acid) $t_R = 0.89 \text{ min}$; >95% purity; 75% ACN/H₂O (0.1% formic acid) $t_{\rm R} = 0.67 \text{ min; } > 95\% \text{ purity.}$

4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5phenyl-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (24). Compound 24 was prepared according to general procedure G using 87 (0.023 g, 0.23 mmol) and 24f (0.100 g, 0.23 mmol). The title compound was obtained after purifying using flash chromatography (0-10% MeOH/DCM) as an off-white solid. Yield 0.110 g, 86%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.20 (s, 1H), 7.76 (d, J = 8.0 Hz, 1H), 7.67-7.54 (m, 2H), 7.47-7.34 (m, 2H), 7.23 (d, J = 1.00 m)5.4 Hz, 4H), 7.15 (t, J = 7.6 Hz, 1H), 7.03 (d, J = 8.3 Hz, 1H), 6.77 (d, J = 6.7 Hz, 2H), 5.32 (dd, J = 11.8, 4.7 Hz, 1H), 3.75 (dd, J = 18.2, 1.2 Hz) 11.9 Hz, 1H), 2.77 (dd, J = 18.3, 4.5 Hz, 1H), 2.62–2.41 (m, 2H), 2.33 (t, I = 6.7 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.54, 168.57, 160.17, 152.68, 149.84, 142.25, 138.54, 134.42, 131.74, 131.40, 131.25, 131.16, 130.84, 128.36, 127.28, 127.03, 125.33, 123.55, 122.39, 121.77, 119.11, 115.57, 58.87, 45.39, 28.54, 28.24. HRMS (m/z): [M - H] calcd for C₂₈H₂₁BrN₃O₄, 542.07209; found, 542.07235. Anal. Calcd for C₂₈H₂₂BrN₃O₄: C, 61.89; H, 3.89; N, 7.73. Found: C, 56.02; H, 4.02; N, 6.78. HPLC 95% MeOH/H₂O (0.1% formic acid) t_R = 0.91 min; >95% purity; 65% ACN/ H_2O (0.1% formic acid) $t_R = 0.70$ min; >95% purity.

4-{5-(4-Bromophenyl)-3-[4-(4-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (25). Compound 25 was prepared according to general procedure G using 87 (0.031 g, 0.31 mmol) and 25f (0.150 g, 0.31 mmol). The title compound was obtained after purifying using flash chromatography (0-10% MeOH/DCM) as an off-white solid. Yield 0.048 g, 27%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.30 (s, 1H), 7.64– 7.48 (m, 2H), 7.42 (t, J = 7.9 Hz, 5H), 7.28 (d, J = 8.2 Hz, 1H), 7.15 (t, J = 7.6 Hz, 1H), 7.02 (d, J = 8.1 Hz, 1H), 6.74 (d, J = 8.0 Hz, 2H),5.33 (dd, J = 11.9, 4.4 Hz, 1H), 3.72 (dd, J = 18.5, 12.0 Hz, 1H), 2.79– 2.69 (m, 1H), 2.53–2.36 (m, 2H), 2.32 (t, J = 6.9 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 174.14, 169.65, 160.19, 152.36, 149.87, 141.71, 138.60, 133.96, 133.25, 131.59, 131.44, 131.23, 130.56, 128.37, 127.64, 127.26, 123.65, 122.43, 120.06, 119.18, 115.66, 58.23, 45.12, 30.27, 29.69. HRMS (m/z): $[M + H]^+$ calcd for $C_{28}H_{22}BrClN_3O_4$, 578.04767; found, 578.04719. Anal. Calcd for C₂₈H₂₁BrClN₃O₄·1.20H₂O: C, 56.01; H, 3.93; N, 7.00. Found: C, 56.02; H, 4.02; N, 6.78.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (26). Compound 26 was prepared according to general procedure G using 87 (0.046 g, 0.460 mmol) and 26f (0.200 g, 0.460 mmol). The title compound was obtained after removing the THF under vacuum, precipitating from EtOAc with hexanes and further purification using flash chromatography (2–10% MeOH/DCM) as a yellow solid. Some compound was lost because of spillage. Yield 0.054 g, 22%. 1 H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.06 (s, 1H), 7.65–7.55 (m, 2H), 7.52 (dd, J = 8.2, 2.3 Hz, 1H),

7.48–7.38 (m, 2H), 7.34–7.24 (m, 3H), 7.15 (t, J = 7.6 Hz, 1H), 7.03 (d, J = 8.1 Hz, 1H), 6.82 (d, J = 8.5 Hz, 2H), 5.35 (dd, J = 11.9, 4.5 Hz, 1H), 3.73 (dd, J = 18.5, 12.0 Hz, 1H), 2.77 (dd, J = 18.4, 4.4 Hz, 1H), 2.65–2.43 (m, 2H), 2.34 (t, J = 6.7 Hz, 2H). 13 C NMR (100 MHz, DMSO- d_6) δ 173.54, 168.67, 160.17, 152.72, 149.89, 141.09, 138.57, 134.04, 133.22, 131.59, 131.46, 130.55, 128.32, 127.31, 122.42, 119.15, 115.59, 58.21, 45.16, 28.47, 28.20. HRMS (m/z): [M + H]⁺ calcd for $C_{28}H_{22}Cl_2N_3O_4$, 534.09819; found, 534.09787. Anal. Calcd for $C_{28}H_{21}Cl_2N_3O_4$: C, 62.93; H; 3.96; N, 7.86. Found: C, 62.38; H, 4.03; N, 7.73. HPLC 85% MeOH/H₂O (0.1% formic acid) t_R = 0.61 min; >95% purity; 75% ACN/H₂O (0.1% formic acid) t_R = 0.61 min; >95% purity.

4-{3-[4-(4-Chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-fluorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (27). Compound 27 was prepared according to general procedure G using 87 (0.029 g, 0.287 mmol) and 27f (0.120 g, 0.287 mmol). The title compound was obtained after removal of the residual solvent, dissolving the crude material in DCM and washing 3× with acidified brine. The organics were collected, dried over magnesium sulfate, and concentrated to yield the title compound as a brown solid. Yield 0.085 g, 57%. $^1\mathrm{H}$ NMR (400 MHz, DMSO- $d_6)$ δ 12.30 (s, 1H), 12.08 (s, 1H), 7.65-7.49 (m, 3H), 7.47-7.34 (m, 2H), 7.34-7.26 (m, 1H), 7.15 (t, J = 7.7 Hz, 1H), 7.10-7.00 (m, 3H), 6.88-6.80 (m, 2H), 5.35 (dd, J = 11.7, 4.3 Hz, 1H), 3.72 (dd, J = 18.1, 12.0 Hz, 1H), 2.78 (dd, J = 18.5, 4.3 Hz, 1H), 2.64-2.42 (m, 2H), 2.33 (t, I = 6.6 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 174.19, 169.29, 160.82, 153.38, 150.51, 139.22, 138.99, 134.70, 133.85, 132.08, 131.24, 128.97, 128.11, 128.05, 127.94, 124.20, 123.06, 119.81, 116.25, 115.76, 115.62, 58.80, 45.91, 29.16, 28.87. HRMS (m/z): $[M - H]^{-1}$ calcd for C₂₈H₂₀ClFN₃O₄, 516.11319; found, 516.11362. Anal. Calcd for C₂₈H₂₁ClFN₃O₄·0.70DCM: C, 59.70; H, 3.91; N, 7.28. Found: C, 59.54; H, 4.15; N, 7.20.

4-{3-[4-(4-Chlorophenyl)-2-oxo-1,2-dihydroguinolin-3-yl]-5phenyl-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (28). Compound 28 was prepared according to general procedure G using 87 (0.055 g, 0.550 mmol) and 28f (0.220 g, 0.550 mmol). The title compound was obtained by filtering from DCM after removal of the THF in vacuo. Yield 0.204 g, 74%. ${}^{\rm i}$ H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.13 (s, 1H), 7.66-7.54 (m, 2H), 7.54-7.39 (m, 3H), 7.32-7.11 (m, 5H), 7.03 (d, I = 8.1 Hz, 1H), 6.77 (dd, I = 6.5, 3.0 Hz, 2H), 5.32 (dd, *J* = 11.9, 4.3 Hz, 1H), 3.75 (dd, *J* = 18.4, 12.1 Hz, 1H), 2.77 (dd, J = 18.3, 4.3 Hz, 1H), 2.64-2.39 (m, 2H), 2.33 (t, J = 6.6 Hz, 1.00 Hz)2H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.55, 168.57, 160.17, 152.70, 149.84, 142.23, 138.54, 134.03, 133.17, 131.48, 131.40, 130.57, 128.33, 128.25, 127.28, 127.03, 125.33, 123.62, 122.39, 119.18, 115.57, 58.86, 45.39, 28.53, 28.23. HRMS (m/z): $[M - H]^-$ calcd for C₂₈H₂₁ClN₃O₄, 498.12261; found, 498.12276. HPLC 85% MeOH/ H_2O (0.1% formic acid) t_R = 0.87 min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 0.69 \text{ min; } >95\% \text{ purity.}$

4-{5-(4-Chlorophenyl)-3-[4-(4-fluorophenyl)-2-oxo-1,2-dihydroguinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (29). Compound 29 was prepared according to general procedure G using 87 (0.072 g, 0.718 mmol) and 29f (0.300 g, 0.718 mmol). The title compound was obtained by filtering from DCM after removal of the THF in vacuo. Yield 0.210 g, 57%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.27 (s, 1H), 12.09 (s, 1H), 7.58 (t, J = 7.7Hz, 1H), 7.43 (d, J = 7.6 Hz, 2H), 7.40–7.25 (m, 5H), 7.15 (t, J = 7.7Hz, 1H), 7.04 (d, J = 8.2 Hz, 1H), 6.86 (d, J = 8.2 Hz, 2H), 5.34 (dd, J = 8.2 Hz, 2H), 5.34 (dd = 11.9, 4.6 Hz, 1H), 3.74 (dd, J = 18.5, 11.9 Hz, 1H), 2.79 (dd, J = 18.5, 11.9 Hz, 1H)18.4, 4.6 Hz, 1H), 2.64–2.42 (m, 2H), 2.33 (t, J = 6.8 Hz, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.54, 168.67, 160.21, 152.82, 150.17, 141.11, 138.53, 131.57, 131.44, 130.80, 128.30, 127.36, 123.62, 122.38, 119.40, 115.57, 115.31, 115.10, 58.21, 45.16, 28.50, 28.20. HRMS (*m*/ z): [M + Na]⁺ calcd for C₂₈H₂₁ClFN₃O₄Na, 540.10968; found, 540.10938. Anal. Calcd for $C_{28}H_{21}CIFN_3O_4\cdot 0.90H_2O$: C, 62.96; H, 4.30; N, 7.86. Found: C, 62.80; H, 4.06; N: 7.80.

4-[5-(4-Chlorophenyl)-3-(2-oxo-4-p-tolyl-1,2-dihydroquino-lin-3-yl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (30). Compound 30 was prepared according to general procedure G using 87 (0.060 g, 0.604 mmol) and 30f (0.250 g, 0.604 mmol). The title

compound was obtained as a yellow solid after purifying using flash chromatography (2–10% MeOH/DCM) followed by precipitation from EtOAc using hexanes. Yield 0.260 g, 84%. 1 H NMR (400 MHz, DMSO- d_6) δ 12.24 (s, 1H), 12.08 (s, 1H), 7.57 (t, J=7.5 Hz, 1H), 7.42 (d, J=8.2 Hz, 1H), 7.36–7.21 (m, 5H), 7.18–7.04 (m, 3H), 6.84 (dd, J=8.5, 2.3 Hz, 2H), 5.32 (dd, J=11.9, 4.4 Hz, 1H), 3.68 (dd, J=18.4, 12.0 Hz, 1H), 2.74 (dd, J=18.4, 4.5 Hz, 1H), 2.63–2.39 (m, 5H), 2.31 (t, J=6.7 Hz, 2H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.55, 168.65, 160.25, 152.86, 151.26, 141.13, 138.55, 137.66, 132.18, 131.51, 131.28, 129.24, 128.79, 128.69, 128.54, 128.26, 127.45, 123.25, 122.23, 119.43, 115.52, 58.17, 45.20, 28.79, 28.60, 28.24, 20.90. HRMS (m/z): [M + H] $^+$ calcd for $\rm C_{29}H_{25}ClN_3O_4$, 514.15281; found, 514.15260. Anal. Calcd for $\rm C_{29}H_{24}ClN_3O_4$ -0.80H₂O: C, 65.92; H, 4.88; N, 7.95. Found: C, 65.86; H, 4.84; N, 7.75.

4-{5-(4-Chlorophenyl)-3-[4-(4-methoxyphenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (31). Compound 31 was prepared according to general procedure G using 87 (0.047 g, 0.465 mmol) and 31f (0.200 g, 0.465 mmol). The title compound was obtained as a white solid by filtering from DCM after removal of the THF in vacuo. Yield 0.165 g, 67%. ¹H NMR (400 MHz, DMSO- d_6) δ 14.05 (s, 1H), 12.24 (s, 1H), 7.57 (t, J $= 7.7 \text{ Hz}, 1\text{H}, 7.42 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{H}), 7.38 - 7.06 \text{ (m, 6H)}, 7.01 \text{ (d, } J = 8.3 \text{ Hz}, 2\text{ (d, } J = 8.3 \text{$ = 8.5 Hz, 1H), 6.83 (d, J = 7.8 Hz, 3H), 5.33 (dd, J = 11.8, 4.4 Hz,1H), 3.85 (s, 3H), 3.67 (dd, J = 18.4, 12.1 Hz, 1H), 2.78-2.53 (m, 2H), 2.41–2.22 (m, 2H). 13 C NMR (100 MHz, DMSO- d_6) δ 175.22, 173.68, 168.75, 160.42, 160.28, 159.20, 152.91, 151.06, 141.19, 138.54, 131.50, 131.26, 131.01, 130.92, 130.00, 128.39, 128.24, 127.68, 127.44, 127.00, 123.52, 122.23, 119.66, 115.54, 113.68, 113.56, 58.16, 55.17, 45.24, 32.60, 28.76, 28.44. HRMS (m/z): $[M + K]^+$ calcd for C₂₉H₂₄ClN₃O₅, 568.10361; found, 568.10341. Anal. Calcd for C₂₉H₂₄ClN₃O₅: C, 65.72; H, 4.56; N, 7.92. Found: C, 57.27; H, 4.20; N, 6.50. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 0.89$ min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 0.67$ min; >95% purity.

4-{5-(4-Chlorophenyl)-3-[4-(4-cyanophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (32). Compound 32 was prepared according to general procedure G using 87 (0.059 g, 0.588 mmol) and 32f (0.150 g, 0.588 mmol). The title compound was obtained as a yellow solid after precipitation from EtOAc using hexanes followed by purification using flash chromatography with 10% MeOH/DCM as a yellow solid. Yield 0.150 g, 48.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.33 (s, 1H), 12.09 (s, 1H), 7.97 (ddd, J = 11.8, 7.7, 1.7 Hz, 2H), 7.64–7.56 (m, 2H), 7.50 (dd, J = 7.8, 1.7 Hz, 1H), 7.43 (d, J = 8.1 Hz, 1H), 7.35– 7.27 (m, 2H), 7.14 (t, J = 7.7 Hz, 1H), $6.95 \text{ (dd, } J = 8.2, } 1.2 \text{ Hz, } 1\text{H)}$, 6.89-6.84 (m, 2H), 5.35 (dd, J = 12.0, 4.4 Hz, 1H), 3.79 (dd, J = 18.5, 12.0 Hz, 1H), 2.89 (dd, I = 18.5, 4.5 Hz, 1H), 2.57–2.23 (m, 4H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.45, 168.70, 160.12, 152.50, 149.37, 140.99, 140.55, 138.61, 132.20, 131.62, 130.59, 130.50, 129.79, 128.37, 128.25, 127.32, 127.23, 123.18, 122.52, 118.75, 118.63, 115.65, 111.18, 58.30, 45.00, 28.77, 28.22. HRMS (m/z): $[M - H]^-$ calcd for C₂₉H₂₀ClN₄O₄, 523.11786; found, 523.11828. HPLC 85% MeOH/ H_2O (0.1% formic acid) $t_R = 0.67$ min; 87% purity; 75% ACN/ H_2O (0.1% formic acid) $t_{\rm R} = 0.89$ min; 85% purity.

4-[5-(4-Chlorophenyl)-3-{2-oxo-4-(4-(trifluoromethyl)phenyl)-1,2-dihydroquinolin-3-yl}-4,5-dihydro-1*H*-pyrazol-1yl]-4-oxobutanoic Acid (33). Compound 33 was prepared according to general procedure G using 87 (0.039 g, 0.385 mmol) and 33f (0.180 g, 0.220 mmol). The title compound was obtained after flash column chromatography using 0-10% MeOH/DCM and precipitation from EtOAc using hexanes as a yellow solid. Yield 0.125 g, 57.2%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.34 (s, 1H), 12.07 (s, 1H), 7.90 (d, J = 8.1 Hz, 1H), 7.85 (d, J = 8.1 Hz, 1H), 7.65(d, J = 8.1 Hz, 1H), 7.65 (d, J = 8.1 Hz, 1H), 7.59 (t, J = 7.6 Hz, 1H),7.53 (d, J = 8.1 Hz, 1H), 7.45 (d, J = 8.2 Hz, 1H), 7.26 (d, J = 8.2 Hz, 2H), 7.15 (t, J = 7.7 Hz, 1H), 6.98 (d, J = 8.1 Hz, 1H), 6.86 (d, J = 8.2Hz, 2H), 5.36 (dd, J = 12.0, 4.6 Hz, 1H), 3.80 (dd, J = 18.4, 12.0 Hz, 1H), 2.91 (dd, J = 18.5, 4.8 Hz, 1H), 2.52–2.21 (m, 4H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.42, 168.68, 160.19, 152.58, 149.61, 141.06, 139.85, 138.62, 131.62, 130.35, 129.69, 128.31, 127.27, 125.20,

123.28, 122.54, 118.95, 115.63, 58.23, 45.04, 28.32, 28.05. $^{19}\mathrm{F}$ NMR (376 MHz, DMSO- d_6) δ –61.595 (s). HRMS (m/z): [M – H] $^-$ calcd for C₂₉H₂₀ClF₃N₃O₄, 566.10999; found, 566.11036. Anal. Calcd for C₂₉H₂₁ClF₃N₃O₄-0.40H₂O: C, 60.56; H, 3.82; N, 7.31. Found: C, 60.57; H, 4.00; N: 7.23.

4-{5-(4-Chlorophenyl)-3-[4-(3-fluorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (34). Compound 34 was prepared according to general procedure G using 87 (0.060 g, 0.598 mmol) and 34f (0.250 g, 0.367 mmol). The title compound was obtained after precipitating from EtOAc using hexanes followed by flash column chromatography using 10% MeOH/DCM, as a yellow solid. Yield 0.190 g, 61.3%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.24 (s, 1H), 12.08 (s, 1H), 7.57 (t, J = 7.5 Hz, 1H), 7.42 (d, J = 8.2 Hz, 1H), 7.36-7.21 (m, 5H), 7.18-7.04 (m, 3H), 6.84 (d, J = 8.2 Hz, 2H), 5.32 (dd, J = 12.0, 4.5 Hz, 1H), 3.68 (dd, J = 18.5, 12.0 Hz, 1H), 2.74 (dd, J = 18.5, 4.5 Hz, 1H), 2.52-2.39 (m, 4H). 13 C NMR (100 MHz DMSO- d_6) δ 173.48, 168.60, 160.19, 149.62, 141.15, 138.53, 131.53, 131.45, 130.35, 128.37, 127.33, 127.16, 122.44, 119.07, 115.57, 58.27, 45.11, 28.79, 28.47, 28.18. ¹⁹F NMR (376 MHz, DMSO- d_6) δ –113.29 to –113.63 (m). HRMS (m/z): [M + Na]⁺ calcd for C₂₈H₂₁ClFN₃O₄Na, 540.10968; found, 540.11014. Anal. Calcd for C₂₈H₂₁ClFN₃O₄·0.90H₂O: C, 62.96; H, 4.30; N, 7.87. Found: C, 63.04; H, 4.38; N, 7.55.

4-{5-(4-Chlorophenyl)-3-[4-(3-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (35). Compound 35 was prepared according to general procedure G using 87 (0.069 g, 0.691 mmol) and 35f (0.300 g, 0.691 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 2-10% MeOH/DCM. Yield 0.170 g, 46.1%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.25 (s, 1H), 12.04 (s, 1H), 7.58-7.51 (m, 2H), 7.51-7.46 (m, 1H), 7.39 (d, J = 8.2Hz, 1H), 7.36–7.32 (m, 1H), 7.25 (ddt, J = 7.1, 4.9, 2.5 Hz, 2H), 7.19 (d, J = 7.5 Hz, 1H), 7.12 (t, J = 7.7 Hz, 1H), 7.01-6.94 (m, 1H),6.81-6.73 (m, 2H), 5.34-5.28 (m, 1H), 3.83-3.74 (m, 1H), 2.84-2.70 (m, 1H), 2.52-2.34 (m, 2H), 2.31-2.24 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.50, 168.64, 160.20, 152.66, 149.46, 141.16, 138.53, 137.44, 137.28, 133.18, 131.56, 131.47, 130.15, 130.00, 129.59, 128.49, 128.40, 127.29, 127.16, 123.43, 122.48, 119.17, 115.58, 58.30, 45.10, 28.78, 28.49, 28.19. HRMS (m/z): $[M + Na]^+$ calcd for C₂₈H₂₁Cl₂N₃O₄Na, 556.08013; found, 556.07990. Anal. Calcd for C₂₈H₂₁Cl₂N₃O₄·0.70H₂O: C, 61.48; H, 4.13; N, 7.68. Found: C, 61.51; H, 4.11; N: 7.48.

4-{5-(4-Chlorophenyl)-3-[2-oxo-4-(m-tolyl)-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (36). Compound 36 was prepared according to general procedure G using 87 (0.060 g, 0.604 mmol) and 36f (0.250 g, 0.604 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0-10% MeOH/DCM followed by precipitation from hot EtOAc and hexanes. Yield 0.240 g, 77.0%. 1H NMR (400 MHz, DMSO- d_6) δ 12.21 (s, 1H), 12.05 (s, 1H), 7.53 (t, J = 7.6Hz, 1H), 7.42–7.27 (m, 3H), 7.27–7.17 (m, 2H), 7.16–7.06 (m, 2H), 7.00 (q, J = 10.2, 8.9 Hz, 2H), 6.80-6.67 (m, 2H), 5.33-5.23 (m, 1H), 3.80-3.62 (m, 1H), 2.76 (dd, J = 18.4, 4.6 Hz, 0.5H), 2.64 (dd, J= 18.5, 4.6 Hz, 0.5H), 2.57–2.34 (m, 5H), 2.31–2.18 (m, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.61, 173.54, 168.63, 168.57, 160.29, 152.88, 152.70, 151.33, 151.24, 141.25, 141.20, 138.52, 137.49, 137.43, 135.17, 135.03, 131.53, 131.30, 130.25, 128.84, 128.36, 128.32, 128.13, 127.50, 127.41, 127.34, 127.26, 126.39, 125.46, 123.16, 122.28, 119.42, 115.55, 115.51, 58.28, 58.23, 45.25, 28.90, 28.79, 28.67, 28.57, 28.35, 28.24, 28.14, 21.09, 21.02, 20.97, 20.91. HRMS (m/z): $[M + H]^+$ calcd for C₂₉H₂₅ClN₃O₄, 514.15281; found, 514.15263. Anal. Calcd for C₂₉H₂₄ClN₃O₄·1.50H₂O·0.06EtOAc: C, 64.29; H, 5.07; N, 7.69. Found: C, 64.65; H, 4.90; N, 7.29.

4-{5-(4-Chlorophenyl)-3-[4-(3-methoxyphenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (37). Compound 37 was prepared according to general procedure G using 87 (0.070 g, 0.698 mmol) and 37f (0.300g, 0.698 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0–10% MeOH/DCM. Yield 0.090 g, 24.3%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.23 (s, 1H), 12.03 (s, 1H), 7.57 (t, J = 7.8 Hz, 1H), 7.48–7.35 (m, 2H), 7.32–7.21 (m, 2H),

7.18–7.12 (m, 1H), 7.09 (t, J = 8.3 Hz, 2H), 6.95 (d, J = 9.9 Hz, 1H), 6.85–6.72 (m, 3H), 5.37–5.30 (m, 1H), 3.84–3.64 (m, 4H), 2.83–2.72 (m, 1H), 2.60–2.51 (m, 2H), 2.31 (t, J = 7.1 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.52, 168.60, 160.26, 159.03, 152.90, 150.95, 141.24, 141.19, 138.52, 138.50, 136.50, 136.46, 131.56, 131.48, 131.33, 131.31, 129.43, 129.38, 128.35, 128.21, 128.14, 127.46, 127.43, 127.19, 123.24, 123.14, 122.32, 121.65, 120.70, 119.31, 119.27, 115.51, 115.13, 114.02, 113.88, 113.58, 58.25, 55.26, 54.99, 45.30, 45.19, 28.59, 28.22. HRMS (m/z): [M + H]⁺ calcd for C₂₉H₂₅ClN₃O₅, 530.14773; found, 530.14715. Anal. Calcd for C₂₉H₂₄ClN₃O₅: C, 65.72; H, 4.56; N, 7.93. Found: C, 65.52; H, 4.72; N, 7.97.

4-{5-(4-Chlorophenyl)-3-[4-(3-cyanophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (38). Compound 38 was prepared according to general procedure G using 87 (0.021g, 0.212 mmol) and 38f (0.090 g, 0.212 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0-10% MeOH/DCM. Yield 0.034 g, 31.0%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.35 (s, 1H), 12.10 (s, 1H), 8.04–7.91 (m, 1H), 7.84–7.67 (m, 2H), 7.65–7.55 (m, 2H), 7.43 (d, J = 8.2 Hz, 1H), 7.31 (d, J = 8.3 Hz, 2H), 7.16 (t, J = 7.7Hz, 1H), 6.97 (d, J = 8.2 Hz, 1H), 6.82 (m, 2H), 5.40-5.32 (m, 1H), 3.96-3.81 (m, 1H), 2.87 (dt, J = 18.3, 4.1 Hz, 1H), 2.42 (d, J = 7.6 Hz, 2H), 2.35–2.26 (m, 2H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.58, 173.42, 168.61, 160.13, 152.58, 148.90, 141.02, 138.54, 136.80, 136.70, 134.66, 133.48, 133.05, 132.18, 132.09, 131.73, 131.57, 129.51, 129.32, 128.47, 128.41, 127.34, 127.25, 127.04, 123.64, 123.53, 122.54, 119.03, 118.52, 115.59, 111.60, 111.40, 58.27, 45.03, 28.75, 28.32, 28.10. HRMS (m/z): $[M - H]^-$ calcd for $C_{29}H_{20}ClN_4O_4$, 523.11786; found, 523.11778. Anal. Calcd for C₂₉H₂₁ClN₄O₄: C, 66.35; H, 4.03; N, 10.67. Found: C, 63.97; H, 4.59; N, 9.39. HPLC 85% MeOH/H2O (0.1% formic acid) t_R = 0.68 min; >95% purity; 75% ACN/H₂O (0.1%) formic acid) $t_R = 0.59$ min; >95% purity.

4-[5-(4-Chlorophenyl)-3-{2-oxo-4-[3-(trifluoromethyl)phenyl]-1,2-dihydroquinolin-3-yl}-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoic Acid (39). Compound 39 was prepared according to general procedure G using 87 (0.054 g, 0.53 mmol) and 39f (0.250 g, 0.53 mmol). The title compound was obtained as a yellow solid using a 0-8% MeOH/DCM. Yield 0.073 g, 24%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.34 (s, 1H), 12.06 (s, 1H), 7.96– 7.84 (m, 1H), 7.82–7.66 (m, 2H), 7.59 (t, J = 8.4 Hz, 2H), 7.44 (d, J =8.3 Hz, 1H), 7.28 (d, J = 8.2 Hz, 1H), 7.23 (d, J = 8.1 Hz, 1H), 7.20-7.13 (m, 1H), 6.98 (d, J = 8.2 Hz, 0.5H), 6.92 (d, J = 8.2 Hz, 0.5H), 6.84 (d, J = 8.1 Hz, 1H), 6.76 (d, J = 8.0 Hz, 1H), 5.34 (dt, J = 12.2, 4.3 Hz, 1H), 3.95-3.70 (m, 1H), 3.00-2.74 (m, 1H), 2.54-2.23 (m, 4H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.40, 173.34, 168.62, 168.52, 160.20, 160.17, 152.67, 152.61, 149.38, 141.10, 141.03, 138.57, 136.57, 136.47, 133.59, 132.57, 131.60, 131.51, 129.50, 129.33, 129.15, 128.92, 128.49, 128.38, 128.30, 127.67, 127.25, 127.17, 127.13, 126.47, 125.07, 123.62, 123.45, 122.54, 122.36, 119.26, 119.11, 115.61, 58.26, 58.21, 45.12, 45.01, 28.37, 28.29, 28.19, 28.08. HRMS (m/z): [M + H]⁺ calcd for C₂₉H₂₂ClF₃N₃O₄, 568.12455; found, 568.12417. Anal. Calcd for C₂₉H₂₁ClF₃N₃O₄·0.20H₂O; C, 60.94; H, 3.77; N, 7.35. Found: C, 60.79; H, 3.96; N, 7.33.

4-{3-[4-(4-Chloro-3-fluorophenyl)-2-oxo-1,2-dihydroguinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (40). Compound 40 was prepared according to general procedure G using 87 (0.040 g, 0.398 mmol) and 40f (0.180 g, 0.398 mmol). The title compound was obtained after flash column chromatography using 0-10%MeOH/DCM as a yellow solid followed by precipitation from EtOAc and hexanes. Yield 0.096 g, 43.7%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.33 (s, 1H), 12.12 (s, 1H), 7.79– 7.64 (m, 1H), 7.62–7.51 (m, 2H), 7.46–7.38 (m, 2H), 7.33–7.23 (m, 2H), 7.21-7.00 (m, 2H), 6.93-6.78 (m, 2H), 5.39 (dt, J = 12.1, 4.4Hz, 1H), 3.85-3.72 (m, 1H), 2.91-2.79 (m, 1H), 2.66-2.39 (m, 2H), 2.34 (td, J = 7.0, 2.2 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.61, 173.47, 173.44, 168.68, 168.66, 168.09, 168.05, 160.11, 157.84, 156.19, 152.77, 152.70, 152.63, 148.67, 141.03, 140.99, 138.55, 136.45, 136.36, 136.30, 131.63, 131.57, 130.63, 130.53, 128.34, 128.28, 127.32, 127.21, 127.12, 127.00, 126.21, 123.57, 123.45, 122.54, 119.69, 119.58, 118.92, 117.28, 115.58, 58.28, 58.17, 45.15, 28.77, 28.39, 28.17. ¹⁹F

NMR (376 MHz, DMSO- d_6) δ –116.32 to –116.51 (m). HRMS (m/z): [M + K]⁺ calcd for C₂₈H₂₀Cl₂FN₃O₄K, 590.04465; found, 590.04631. Anal. Calcd for C₂₈H₂₀Cl₂FN₃O₄·0.80H₂O·0.10EtOAc: C, 59.26; H, 3.92; N, 7.30. Found: C, 59.16; H, 4.21; N, 7.14.

4-{3-[4-(3-Chloro-4-fluorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (41). Compound 41 was prepared according to general procedure G using 87 (0.040 g, 0.398 mmol) and 41f (0.180 g, 0.398 mmol). The title compound was obtained after purifying using flash column chromatography using 0-10% MeOH/DCM as a yellow solid. Yield 0.120 g, 54.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.32 (s, 1H), 12.12 (s, 1H), 7.73-7.50 (m, 3H), 7.44 (dd, J = 8.2, 1.7 Hz, 2H), 7.29 (td, J = 8.8, 2.0 Hz, 2H), 7.17 (t, J = 7.6 Hz, 1H), 7.09-7.01 (m, 1H), 6.88 (dt, J = 8.6, 2.4 Hz, 2H), 5.38 (dd, J = 10.4, 4.6 Hz, 1H), 3.90-3.76 (m, 1H), 2.93-2.77 (m, 1H), 2.66-2.40 (m, 2H), 2.40-2.28 (m, 2H). 13 C NMR (100 MHz, DMSO- d_6) δ 173.63, 173.48, 173.44, 168.68, 160.15, 155.93, 152.68, 152.60, 148.76, 148.72, 141.01, 138.50, 133.00, 132.90, 131.93, 131.62, 131.57, 131.50, 130.63, 130.51, 129.53, 128.39, 128.34, 127.38, 127.23, 127.11, 123.74, 123.66, 122.50, 119.77, 119.46, 119.24, 116.87, 116.66, 115.56, 58.25, 45.10, 28.78, 28.43, 28.38, 28.15. HRMS (m/z): $[M + H]^+$ calcd for C₂₈H₂₁Cl₂FN₃O₄, 552.08877; found, 552.09030. Anal. Calcd for C₂₈H₂₀Cl₂FN₃O₄: C, 60.88; H, 3.64; N, 7.61. Found: C, 58.29; H, 4.04; N, 6.66. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 0.93$ min; >95% purity; 75% ACN/H₂O (0.1% formic acid) t_R = 0.55 min; >95% purity.

4-{5-(4-Chlorophenyl)-3-[4-(3,4-dichlorophenyl)-2-oxo-1,2dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (42). Compound 42 was prepared according to general procedure G using 87 (0.043 g, 0.427 mmol) and 42f (0.200 g, 0.427 mmol). The title compound was obtained after purifying flash column chromatography using 0-10% MeOH/DCM as a yellow solid. Yield 0.102 g, 42.0%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.31 (s, 1H), 12.10 (s, 1H), 7.83-7.70 (m, 1H), 7.63-7.54 (m, 2H), 7.46-7.37 (m, 2H), 7.32-7.22 (m, 2H), 7.16 (t, J = 7.7 Hz, 1H), 7.04 (t, J = 7.0 Hz, 1H), 6.87-6.80 (m, 2H), 5.37 (dd, J = 12.0, 4.3 Hz, 1H), 3.88-3.74(m, 1H), 2.90–2.74 (m, 1H), 2.53–2.39 (m, 2H), 2.38–2.29 (m, 2H). 13 C NMR (150 MHz, DMSO- d_6) δ 174.04, 169.33, 160.76, 153.24, 149.13, 141.66, 139.20, 136.68, 132.40, 132.27, 132.19, 131.96, 131.08, 130.76, 129.73, 129.05, 128.98, 127.99, 127.83, 127.74, 123.18, 119.69, 116.23, 58.93, 45.75, 29.62, 29.11, 29.07, 28.86. HRMS (m/z): [M +H]⁺ calcd for C₂₈H₂₁Cl₃N₃O₄, 568.05922; found, 568.06168. Anal. Calcd for $C_{28}H_{20}Cl_3N_3O_4\cdot 0.80H_2O$: C, 57.66; H, 3.73; N, 7.20. Found: C, 57.70; H, 3.65; N: 6.94.

4-{5-(4-Chlorophenyl)-3-[4-(3,4-difluorophenyl)-2-oxo-1,2dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (43). Compound 43 was prepared according to general procedure G using 87 (0.0427 g, 0.427 mmol) and 43f (0.200 g, 0.427 mmol). The title compound was obtained after flash column chromatography using 0-10% MeOH/DCM as a yellow solid. Yield 0.110 g, 68.8%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.31 (s, 1H), 12.13 (s, 1H), 7.63-7.52 (m, 3H), 7.46-7.39 (m, 1H), 7.35-7.28 (m, 1H), 7.29-7.23 (m, 2H), 7.20-7.10 (m, 1H), 7.06 (d, J = 8.2 Hz, 1H), 6.95-6.84 (m, 2H), 5.43-5.33 (m, 1H), 3.88-3.73 (m, 1H), 2.86 (dd, J = 18.5, 4.4 Hz, 1H), 2.66–2.29 (m, 4H). ¹³C NMR (150 MHz, DMSO- d_6) δ 174.08, 169.36, 160.79, 153.45, 149.53, 141.69, 139.20, 139.20, 132.28, 132.14, 128.99, 128.91, 127.98, 127.80, 124.26, 123.12, 119.83, 116.22, 58.92, 45.79, 29.07, 28.84. ¹⁹F NMR (376 MHz, DMSO- d_6) δ -5.88 to -6.23 (m), -6.44 to -6.75 (m). HRMS (m/z): $[M - H]^-$ calcd for $C_{28}H_{19}ClF_2N_3O_4$, 534.10376; found, 534.10358. Anal. Calcd for C₂₈H₂₀ClF₂N₃O₄·0.40H₂O: C, 61.92; H, 3.86; N, 7.74. Found: C, 61.76; H, 4.15; N, 7.52.

4-{5-(4-Chlorophenyl)-3-[4-(3,5-difluorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (44). Compound 44 was prepared according to general procedure G using 87 (0.041 g, 0.413 mmol) and 44f (0.180 g, 0.413 mmol). The title compound as a yellow solid was obtained after purifying using flash column chromatography with 0–10% MeOH/DCM. Yield 0.133 g, 56.5%. 1 H NMR (400 MHz, DMSO- 4 6) δ 12.33 (s, 1H), 12.07 (s, 1H), 7.59 (t, 1 = 7.7 Hz, 1H), 7.47–7.33 (m, 2H), 7.29 (d, 1 = 8.2 Hz, 2H), 7.23 (d, 1 = 8.5 Hz, 1H), 7.17 (t, 1 = 7.6 Hz,

1H), 7.08 (d, J = 8.0 Hz, 2H), 6.89 (d, J = 8.1 Hz, 2H), 5.40 (dd, J = 12.0, 4.6 Hz, 1H), 3.86 (dd, J = 18.4, 12.0 Hz, 1H), 2.90 (dd, J = 18.5, 4.6 Hz, 1H), 2.59–2.41 (m, 2H), 2.33 (t, J = 6.8 Hz, 2H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.65, 173.46, 168.64, 163.55, 163.43, 161.11, 160.95, 160.11, 152.68, 148.60, 141.09, 139.05, 138.95, 138.52, 131.58, 128.55, 128.40, 127.26, 127.13, 123.41, 122.57, 118.79, 115.58, 113.31, 113.12, 112.32, 112.12, 103.85, 58.30, 45.02, 28.80, 28.37, 28.15. ¹⁹F (376 MHz, DMSO- d_6) δ –109.98 to –110.05 (m). HRMS (m/z): [M – H]⁻ calcd for C₂₈H₁₉ClF₂N₃O₄, 534.10376; found, 534.10402. Anal. Calcd for C₂₈H₂₀ClF₂N₃O₄, 0.50H₂O: C, 61.71; H, 3.88; N, 7.71. Found: C, 61.52; H, 4.05; N, 7.42.

4-{5-(4-Chlorophenyl)-3-[4-(3,5-dichlorophenyl)-2-oxo-1,2dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (45). Compound 45 was prepared according to general procedure G using 87 (0.038 g, 0.384 mmol) and 45f (0.180 g, 0.384 mmol). The title compound was obtained as a yellow solid after purifying using flash column chromatography with 0-10% MeOH/ DCM. Yield 0.080 g, 36.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.33 (s, 1H), 12.10 (s, 1H), 7.84–7.71 (m, 1H), 7.64–7.56 (m, 2H), 7.51– 7.39 (m, 2H), 7.29 (t, J = 7.3 Hz, 2H), 7.17 (t, J = 7.6 Hz, 1H), 7.06 (t, J = 7.6 Hz, 1H), 7.06 (t, J = 7.8 Hz, JI = 6.9 Hz, 1H), 6.84 (d, I = 8.1 Hz, 2H), 5.39 (dd, I = 12.0, 4.3 Hz, 1H), 3.89–3.75 (m, 1H), 2.85–2.67 (m, 1H), 2.64–2.30 (m, 4H). ¹³C NMR (100 MHz, DMSO- d_6) δ 173.46, 168.66, 160.11, 152.60, 152.53, 148.48, 141.01, 138.52, 136.01, 131.77, 131.59, 131.31, 131.11, 130.39, 130.13, 129.07, 128.40, 128.33, 127.35, 127.17, 127.06, 122.55, 119.02, 115.57, 58.24, 45.09, 28.43, 28.38, 28.17. HRMS calcd for $C_{28}H_{19}Cl_3N_3O_4 \ [M-H]^-$, 566.04466; found, 566.04483. Anal. Calcd for C₂₈H₂₀Cl₃N₃O₄·0.30H₂O: C, 58.57; H, 3.62; N, 7.32. Found: C, 58.58; H, 3.75; N, 7.23.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-6-methyl-2oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4oxobutanoic Acid (46). Compound 46 was prepared according to general procedure G using 87 (0.039 g, 0.39 mmol) and 46f (0.175 g, 0.39 mmol). The title compound was obtained after flash column chromatography using 0-8% MeOH/DCM as a yellow solid. Yield 0.114 g, 53.3%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.18 (s, 1H), 11.79 (s, 1H), 7.56 (dt, I = 8.2, 2.3 Hz, 1H), 7.48 (dt, I = 8.2, 2.3 Hz, 1H), 7.41-7.34 (m, 2H), 7.30 (dd, J = 8.3, 2.2 Hz, 1H), 7.27-7.20 (m, 3H), 6.80-6.72 (m, 3H), 5.35-5.25 (m, 1H), 3.75-3.59 (m, 1H), 2.76–2.65 (m, 1H), 2.61–2.33 (m, 2H), 2.26–2.32 (m, 2H), 2.19 (s, 3H). 13 C NMR (100 MHz, DMSO- d_6) δ 173.54, 168.64, 160.00, 152.85, 149.66, 141.10, 136.63, 134.07, 133.16, 132.79, 131.56, 131.48, 131.40, 130.53, 128.37, 128.30, 127.30, 126.53, 123.47, 119.05, 115.59, 58.18, 45.18, 28.47, 28.19, 20.58. HRMS (m/z): $[M - H]^-$ calcd for C₂₉H₂₂Cl₂N₃O₄, 546.09929; found, 546.09872. Anal. Calcd for $C_{29}H_{23}Cl_2N_3O_4\cdot 0.50H_2O\colon \ C,\ 62.49;\ \ H,\ \ 4.34;\ \ N,\ \ 7.54.\ \ Found:\ \ C,$ 62.44; H, 4.55; N 7.39.

4-{5-(4-Chlorophenyl)-3-[4-(3-fluorophenyl)-6-methyl-2oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4**oxobutanoic Acid (47).** Compound 47 was prepared according to general procedure G using 87 (0.037 g, 0.37 mmol) and 47f (0.160 g, 0.37 mmol). The title compound was obtained after flash column chromatography using 0-10% MeOH/DCM as a brown solid. Yield 0.094 g, 47.7%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.24 (s, 1H), 12.12 (s, 1H), 7.64–7.47 (m, 1H), 7.42 (d, J = 8.5 Hz, 1H), 7.40–7.32 (m, 2H), 7.32-7.22 (m, 3H), 7.11 (dd, J = 23.8, 8.5 Hz, 1H), 6.86-6.77 (m, 3H), 5.35 (dt, J = 12.5, 3.4 Hz, 1H), 3.80 (dd, J = 18.6, 12.1)Hz, 1H), 2.81 (dt, I = 18.5, 5.2 Hz, 1H), 2.53–2.39 (m, 2H), 2.31 (t, I= 6.8 Hz, 2H), 2.23 (s, 3H). 13 C NMR (150 MHz, DMSO- d_6) δ 173.46, 168.57, 162.67, 161.11, 160.03, 152.88, 152.81, 149.39, 141.15, 137.59, 136.61, 132.77, 131.58, 131.52, 131.38, 130.25, 128.36, 128.20, 127.33, 127.15, 126.57, 126.52, 125.83, 124.79, 123.38, 123.30, 118.98, 116.66, 116.52, 115.57, 115.19, 115.05, 58.25, 45.16, 28.98, 28.47, 28.19, 20.61. $^{19}\mathrm{F}$ NMR (376 MHz, DMSO- $d_{6})~\delta$ -113.28 to -113.50(m). HRMS (m/z): $[M - H]^-$ calcd for $C_{29}H_{22}CIFN_3O_4$, 530.12884; found, 530.12883. Anal. Calcd for C₂₉H₂₃ClFN₃O₄·1.20H₂O: C, 62.92; H, 4.62; N, 7.59. Found: C, 62.82; H, 4.37; N, 7.34.

4-{3-[6-Chloro-4-(4-chlorophenyl)-2-oxo-1,2-dihydroquino-lin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-ox-obutanoic Acid (48). Compound 48 was prepared according to general procedure G using 87 (0.038 g, 0.38 mmol) and 48f (0.180 g,

0.38 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0–8% MeOH/DCM. Yield 0.033 g, 15.1%. $^1\mathrm{H}$ NMR (400 MHz, DMSO- d_6) δ 12.44 (s, 1H), 12.08 (s, 1H), 7.69–7.58 (m, 2H), 7.54 (dd, J = 8.2, 2.3 Hz, 1H), 7.44 (dd, J = 8.5, 2.7 Hz, 2H), 7.35–7.23 (m, 3H), 6.93 (d, J = 2.3 Hz, 1H), 6.85–6.76 (m, 2H), 5.35 (dd, J = 12.0, 4.4 Hz, 1H), 3.72 (dd, J = 18.5, 12.0 Hz, 1H), 2.76 (dd, J = 18.5, 4.4 Hz, 1H), 2.64–2.38 (m, 2H), 2.33 (t, J = 6.7 Hz, 2H). $^{13}\mathrm{C}$ NMR (150 MHz, DMSO- d_6) δ 173.50, 168.71, 159.96, 152.33, 148.76, 141.00, 137.33, 133.52, 133.36, 131.61, 131.48, 131.38, 130.52, 128.52, 128.32, 127.27, 126.25, 125.91, 124.80, 120.43, 117.67, 58.25, 45.08, 28.47, 28.19. HRMS (m/z): [M + H] $^+$ calcd for $\mathrm{C_{28}H_{21}Cl_3N_3O_4}$, 568.05922; found, 568.05881. Anal. Calcd for $\mathrm{C_{28}H_{20}Cl_3N_3O_4}$ ·0.40H $_2\mathrm{O}$: C, 58.38; H, 3.64; N, 7.29. Found: C, 58.29; H, 3.73; N, 7.24.

4-{3-[6-Chloro-4-(3-fluorophenyl)-2-oxo-1,2-dihydroguinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (49). Compound 49 was prepared according to general procedure G using 87 (0.035 g, 0.35 mmol) and 49f (0.160 g, 0.35 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0-8% MeOH/DCM. Yield 0.039 g, 20.0%. 1 H NMR (600 MHz, DMSO- d_{6}) δ 12.45 (s, 1H), 12.05 (s, 1H), 7.65 (dd, J = 8.8, 2.5 Hz, 1H), 7.63–7.53 (m, 1H), 7.45 (d, J =8.8 Hz, 1H), 7.42-7.32 (m, 2H), 7.30 (d, J = 8.2 Hz, 1H), 7.29-7.25(m, 1H), 7.19 (dd, J = 9.2, 2.1 Hz, 1H), 7.12 (d, J = 7.5 Hz, 1H), 6.94(d, J = 2.4 Hz, 1H), 6.83 (dd, J = 11.9, 8.3 Hz, 2H), 5.36 (dt, J = 12.2,4.6 Hz, 1H), 2.88-2.80 (m, 1H), 2.51-2.44 (m, 2H), 2.31 (t, J = 6.9Hz, 2H). 13 C NMR (150 MHz, DMSO- d_6) δ 174.13, 169.33, 163.45, 161.82, 160.66, 153.02, 152.95, 149.14, 141.73, 137.95, 137.52, 132.03, 131.21, 129.05, 127.96, 127.78, 126.91, 126.57, 125.39, 125.30, 121.01, 118.31, 116.19, 58.96, 45.71, 29.12, 28.85. HRMS (m/z): $[M - H]^{-1}$ calcd for C₂₈H₁₉Cl₂FN₃O₄, 550.07421; found, 550.07485. Anal. Calcd for C₂₈H₂₀Cl₂FN₃O₄·1.00H₂O: C, 58.96; H, 3.89; N, 7.37. Found: C, 58.98; H, 3.78; N, 7.06.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-6-fluoro-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanoic Acid (50). Compound 50 was prepared according to general procedure G using 87 (0.035 g, 0.35 mmol) and 50f (0.19 g, 0.42 mmol). The title compound was obtained after flash column chromatography using 0-10% MeOH/DCM as a yellow solid. Yield 0.151 g, 65.1%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.37 (s, 1H), 12.07 (s, 1H), 7.61 (dd, J = 8.2, 2.4 Hz, 1H), 7.56–7.41 (m, 4H), 7.34-7.25 (m, 3H), 6.85-6.78 (m, 2H), 6.71 (dd, J = 9.7, 2.9 Hz, 1H), 5.36 (dd, J = 12.0, 4.5 Hz, 1H), 3.73 (dd, J = 18.5, 12.1 Hz, 1H), $2.78 \text{ (dd, } J = 18.5, 4.4 \text{ Hz, } 1\text{H}), 2.62-2.54 \text{ (m, } 1\text{H}), 2.53-2.40 \text{ (m, } 1\text{H})}$ 1H), 2.33 (t, I = 6.7 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.48, 168.69, 159.89, 157.84, 156.25, 152.46, 149.03, 141.01, 135.30, 133.52, 133.45, 131.59, 131.40, 130.47, 128.48, 128.30, 127.28, 124.74, 119.94, 119.78, 119.61, 117.67, 117.62, 111.93, 111.77, 58.24, 45.07, 28.46, 28.18. ¹⁹F NMR (376 MHz, DMSO- d_6) δ –120.13 to –120.29 (m). HRMS (m/z): $[M - H]^-$ calcd for $C_{28}H_{19}Cl_2FN_3O_4$, 550.07421; found, 550.07419. Anal. Calcd for C₂₈H₂₀Cl₂FN₃O₄: C, 60.88; H, 3.65; N, 7.61. Found: C, 60.16; H, 3.98; N, 7.28. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 1.10 \text{ min}$; >95% purity; 75% ACN/H₂O (0.1% formic acid) $t_R = 0.81$ min; >95% purity.

4-{5-(4-Chlorophenyl)-3-[6-fluoro-4-(3-fluorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobutanoic Acid (51). Compound 51 was prepared according to general procedure G using 87 (0.041 g, 0.41 mmol) and 51f (0.180 g, 0.41 mmol). The title compound was obtained after flash column chromatography using 0-10% MeOH/DCM as a yellow solid. Yield 0.076 g, 34.3%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.34 (s, 1H), 12.01 (s, 1H), 7.59-7.39 (m, 3H), 7.37-7.19 (m, 3H), 7.14 (t, J = 7.8Hz, 1H), 7.07 (t, J = 7.2 Hz, 1H), 6.83-6.75 (m, 1H), 6.74-6.63 (m, 1H), 5.40-5.33 (m, 1H), 3.81 (dd, J = 18.5, 12.1 Hz, 2H), 2.85-2.75(m, 1H), 2.46-2.36 (m, 2H), 2.27 (t, J = 7.0 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.47, 168.66, 162.80, 161.17, 161.11, 159.93, 157.85, 156.26, 152.52, 152.44, 148.77, 141.08, 137.08, 137.02, 136.97, 135.29, 131.62, 131.56, 130.55, 128.39, 127.39, 127.25, 127.21, 127.08, 125.75, 124.69, 124.59, 119.90, 119.83, 117.66, 116.65, 115.50, 115.36, 111.87, 58.36, 58.24, 45.07, 28.46, 28.18. ¹⁹F NMR (376 MHz,

DMSO- d_6) δ –112.89 to –113.28 (m), –120.04 to –120.29 (m). HRMS (m/z): [M – H]⁻ calcd for C₂₈H₁₉ClF₂N₃O₄, 534.10376; found, 534.10345. Anal. Calcd for C₂₈H₂₀ClF₂N₃O₄: C, 62.75; H, 3.76; N, 7.84. Found: C, 61.88; H, 3.98; N: 7.67.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-6-methoxy-2oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4oxobutanoic Acid (52). Compound 52 was prepared according to general procedure G using 87 (0.039 g, 0.39 mmol) and 52f (0.180 g, 0.39 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0-10% MeOH/DCM. Yield 0.095 g, 43.4%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.19 (s, 1H), 12.09 (s, 1H), 7.61 (dd, J = 8.2, 2.4 Hz, 1H), 7.52 (dd, J = 8.2, 2.4 Hz, 1H), 7.42 (dd, J = 8.2, 2.3 Hz, 1H), 7.39 (d, J = 8.8 Hz, 1H), 7.32-7.25 (m, 4H), 6.82 (d, J = 8.5 Hz, 2H), 6.42 (d, J = 2.8 Hz, 1H), 5.34 (dd, J = 11.9, 4.5 Hz, 1H), 3.73 (dd, J = 18.4, 12.0 Hz, 1H), 3.60 (s, 1)3H), 2.77 (dd, J = 18.4, 4.4 Hz, 1H), 2.63–2.44 (m, 2H), 2.33 (t, J = 6.6 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.52, 168.66, 159.69, 154.24, 152.88, 149.28, 141.09, 134.02, 133.23, 133.15, 131.57, 131.42, 130.51, 128.39, 128.29, 128.12, 127.31, 123.95, 120.17, 119.68, 117.01, 109.05, 58.20, 55.32, 45.15, 28.48, 28.21. HRMS (m/z): [M – H]⁻ calcd for $C_{29}H_{22}Cl_2N_3O_5$, 562.09420; found, 562.09430. Anal. Calcd for C₂₉H₂₃Cl₂N₃O₅: C, 61.71; H, 4.11; N, 7.44. Found: C, 61.47; H, 4.06; N, 7.36.

4-{5-(4-Chlorophenyl)-3-[4-(3-fluorophenyl)-6-methoxy-2oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4oxobutanoic Acid (53). Compound 53 was prepared according to general procedure G using 87 (0.027 g, 0.27 mmol) and 53f (0.120 g, 0.27 mmol). The title compound was obtained as a yellow solid after flash column chromatography using 0-10% MeOH/DCM. Yield 0.040 g, 27.7%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.21 (s, 1H), 7.64–7.48 (m, 1H), 7.43–7.07 (m, 6H), 6.87–6.77 (m, 2H), 6.45– 6.40 (m, 1H), 5.40-5.30 (m, 1H), 3.87-3.74 (m, 2H), 3.60 (s, 3H), 2.83 (dt, J = 18.4, 5.2 Hz, 1H), 2.54-2.39 (m, 2H), 2.31 (t, J = 6.9 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.55, 168.62, 162.77, 162.71, 161.15, 161.08, 159.73, 154.23, 152.93, 152.85, 149.03, 141.16, 137.56, 137.51, 133.12, 131.59, 131.52, 130.38, 128.41, 128.32, 127.44, 127.26, 127.07, 125.78, 124.73, 123.87, 123.78, 120.06, 119.63, 116.98, 115.17, 109.18, 58.35, 58.17, 55.38, 55.20, 45.10, 28.49, 28.23. ¹⁹F NMR (376 MHz, DMSO- d_6) δ –113.15 to –113.37 (m). HRMS (m/z): $[M - H]^-$ calcd for $C_{29}H_{22}ClFN_3O_5$, 546.12375; found, 546.12384. Anal. Calcd for C₂₉H₂₃ClFN₃O₅: C, 63.56; H, 4.23; N, 7.67. Found: C, 52.15; H, 3.91; N, 5.95. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 0.809 \text{ min}$; >95% purity; 75% ACN/H₂O (0.1% formic acid) $t_R = 0.625 \text{ min}; >95\% \text{ purity}.$

(E)-4-[5-(4-Bromophenyl)-3-(6-chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1H-pyrazol-1-yl]-4-oxobut-2enoic Acid (54). Compound 54 was prepared from 54g (0.190 g, 0.322 mmol) and 1 M NaOH (1.22 mL, 1.22 mmol), which were stirred to give a yellow solution. After 4 h, 1 M HCl (1.22 mL) was added and a yellow solid precipitated. The solid was filtered and rinsed with water to give the title compound as a yellow solid. Yield 0.170 g, 92.0%. ¹H NMR (400 MHz, DMSO- d_6) δ 13.03 (s, 1H), 12.46 (s, 1H), 7.66 (dd, J = 8.8, 2.4 Hz, 1H), 7.61–7.38 (m, 7H), 7.31 (dt, J =5.6, 2.5 Hz, 1H), 7.26 (d, *J* = 15.7 Hz, 1H), 6.94 (d, *J* = 2.4 Hz, 1H), 6.78 (d, J = 8.3 Hz, 2H), 6.45 (d, J = 15.7 Hz, 1H), 5.45 (dd, J = 11.8,)4.5 Hz, 1H), 3.80 (dd, J = 18.7, 11.9 Hz, 1H), 2.88 (dd, J = 18.7, 4.5 Hz, 1H). $^{13}{\rm C}$ NMR (150 MHz, DMSO- $d_6)$ δ 167.81, 166.11, 160.30, 159.96, 155.82, 154.41, 150.24, 140.70, 137.39, 134.48, 132.67, 131.83, 131.42, 129.19, 128.63, 128.51, 128.36, 127.93, 127.83, 126.17, 126.05, 124.09, 120.60, 120.44, 117.66, 58.69, 58.63, 45.11. HRMS (*m/z*): [M - H]⁻ calcd for C₂₈H₁₈ClBrN₃O₄, 574.01747; found, 574.01750. Anal. Calcd for C₂₈H₁₉ClBrN₃O₄·1.00H₂O: C, 56.54; H, 3.56; N, 7.06. Found: C, 56.66; H, 3.76; N: 6.93.

(*Z*)-4-[5-(4-Bromophenyl)-3-(6-chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobut-2-enoic Acid (55). Compound 55 was prepared according to general procedure G using furan-2,5-dione (90) (0.061 g, 0.63 mmol) and 55f (0.300 g, 0.63 mmol). The title compound was obtained after filtration from the cooled reaction medium and rinsed with THF. Yield 0.200 g, 55.3%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.69 (s, 1H), 12.41 (s, 1H), 7.64 (d, J = 8.7 Hz, 1H), 7.60 – 7.36 (m, 7H), 7.25 (d, J = 7.2 Hz,

1H), 6.92 (s, 1H), 6.78 (d, J = 8.1 Hz, 2H), 6.40 (d, J = 12.1 Hz, 1H), 6.16 (d, J = 12.1 Hz, 1H), 5.37 (dd, J = 12.0, 4.9 Hz, 1H), 3.78 (dd, J = 18.5, 12.0 Hz, 1H), 2.79 (dd, J = 18.7, 4.8 Hz, 1H). 13 C NMR (100 MHz, DMSO- d_6) δ 166.77, 161.77, 160.01, 153.58, 150.11, 140.93, 137.36, 134.39, 131.31, 129.57, 129.46, 128.64, 128.43, 127.96, 126.16, 126.08, 124.26, 120.62, 120.29, 117.67, 58.43, 45.17. HRMS (m/z): [M - H] $^-$ calcd for $C_{28}H_{18}$ ClBrN₃O₄, 574.01747; found, 574.01654. Anal. Calcd for $C_{28}H_{19}$ BrClN₃O₄: C, 58.30; H, 3.32; N, 7.28. Found: C, 43.78; H, 3.00; N: 5.57. HPLC 85% MeOH/H₂O (0.1% formic acid) t_R = 1.18 min; >95% purity; 75% ACN/H₂O (0.1% formic acid) t_R = 1.06 min; >95% purity.

Methyl 4-[5-(4-Bromophenyl)-3-(6-chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1*H*-pyrazol-1-yl]-4-oxobutanoate (56). Compound 56 was prepared from 56g (0.400 g, 0.69 mmol) in the following manner. 56g (0.400 g, 0.69 mmol) was dissolved in 6.9 mL of THF, and freshly prepared HCl (acetyl chloride added to methanol) in MeOH was added dropwise to the reaction vessel with stirring until TLC indicated completion. Upon completion, the THF was removed under vacuum, the residue dissolved in DCM, washed 3× with acidified brine, and column-chromatographed using a 0-10% MeOH gradient in DCM to give the title compound as a yellow solid. Yield 0.100 g, 24.4%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.42 (s, 1H), 7.64 (dd, J = 8.7, 2.4 Hz, 1H), 7.54–7.49 (m, 3H), 7.48-7.38 (m, 4H), 7.31-7.24 (m, 1H), 6.93 (d, J = 2.3 Hz, 1H), 6.77-6.70 (m, 2H), 5.31 (dd, J = 12.0, 4.6 Hz, 1H), 3.76 (dd, J = 18.5, 12.2 Hz, 1H), 3.59 (s, 1H), 3.54 (s, 2H), 2.79 (dd, I = 18.5, 4.6 Hz, 1H), 2.61–2.42 (m, 2H), 2.37 (t, J = 6.7 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 172.44, 168.37, 168.08, 160.09, 152.81, 152.61, 149.98, 141.49, 137.32, 134.58, 131.30, 129.45, 128.48, 128.36, 127.78, 127.67, 126.13, 126.01, 124.50, 120.65, 120.14, 117.65, 58.43, 58.30, 51.35, 51.24, 45.23, 45.08, 28.46, 27.96. HRMS (m/z): $[M - H]^{-1}$ calcd for C₂₉H₂₂ClBrN₃O₄, 590.04877; found, 590.04851. Anal. Calcd for C₂₉H₂₃ClBrN₃O₄: C, 58.75; H, 3.91; N, 7.09. Found: C, 58.35; H, 4.08; N. 6.66.

(E)-4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobut-2-enoic Acid (58). Compound 58 was prepared from compound 60 using the following method. In a 50 mL round-bottomed flask, ethanol (28.2 mL), 60 (0.500 g, 0.850 mmol), and 1 M NaOH (3.22 mL, 3.22 mmol) were stirred to give a yellow solution. After 4 h, 1 M HCl (3.22 mL) was added and a yellow solid precipitated. The solid was filtered and rinsed with water. The resulting solid was dissolved in DCM, washed with brine, and the organics were dried over magnesium sulfate in vacuo. The title compound was obtained from column chromatography (0–10% MeOH in DCM) as an off-white solid. Yield 0.320 g, 65.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.98 (s, 1H), 12.28 (s, 1H), 7.71-7.51 (m, 3H), 7.48-7.19 (m, 6H), 7.12 (t, J = 7.6Hz, 1H), 7.02 (d, J = 8.2 Hz, 1H), 6.85 (d, J = 8.2 Hz, 2H), 6.44 (d, J $= 15.7 \text{ Hz}, 1\text{H}), 5.45 \text{ (dd, } J = 11.7, 4.4 \text{ Hz}, 1\text{H}), 3.73 \text{ (dd, } J = 18.7, 1.7)}$ 11.8 Hz, 1H), 2.89 (dd, J = 18.6, 4.4 Hz, 1H). ¹³C NMR (150 MHz, DMSO- d_6) δ 168.08, 166.05, 160.24, 160.04, 154.75, 152.81, 152.77, 150.29, 140.25, 138.65, 134.49, 132.80, 131.95, 131.60, 131.36, 131.26, 130.74, 128.51, 127.55, 127.43, 122.92, 122.47, 121.84, 119.01, 115.61, 58.50, 45.19. HRMS (m/z): $[M + H]^+$ calcd for $C_{28}H_{20}ClBrN_3O_4$, 576.03202; found, 576.03294. Anal. Calcd for C₂₈H₁₉ClBrN₃O₄·0.40H₂O: C, 57.58; H, 3.42; N, 7.19. Found: C, 57.46; H, 3.50; N, 7.23.

(*Z*)-4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1*H*-pyrazol-1-yl}-4-oxobut-2-enoic Acid (59). Compound 59 was prepared according to general procedure G using 89 (0.061 g, 0.63 mmol) and 22f (0.300 g, 0.630 mmol). The THF was removed under vacuum, and the resultant residue was dissolved in hot EtOAc. A yellow solid was present upon cooling which was filtered and determined to be the desired product. Yield 0.171 g, 47.3%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.74 (s, 1H), 12.30 (s, 1H), 7.75 (d, J = 8.3 Hz, 1H), 7.67 (d, J = 8.4 Hz, 1H), 7.58 (t, J = 7.9 Hz, 1H), 7.43 (d, J = 8.4 Hz, 1H), 7.32 (t, J = 8.6 Hz, 3H), 7.22 (d, J = 8.2 Hz, 1H), 7.15 (t, J = 7.8 Hz, 1H), 7.03 (d, J = 8.2 Hz, 1H), 6.87 (d, J = 8.1 Hz, 2H), 6.48 (d, J = 12.2 Hz, 1H), 6.20 (d, J = 12.1 Hz, 1H), 5.47–5.38 (m, 1H), 3.76 (dd, J = 18.8, 11.9 Hz, 1H),

2.84–2.74 (m, 1H). 13 C NMR (100 MHz, DMSO- d_6) δ 166.80, 161.77, 160.05, 153.90, 150.03, 140.50, 138.60, 134.27, 131.76, 131.69, 131.55, 131.28, 130.82, 129.68, 129.41, 128.36, 127.51, 127.32, 123.10, 122.47, 121.89, 119.04, 115.61, 93.88, 67.04, 58.28, 45.27, 25.15. HRMS (m/z): [M - H] $^-$ calcd for C₂₈H₁₈ClBrN₃O₄, 576.03202; found, 576.03371. Anal. Calcd for C₂₈H₁₉ClBrN₃O₄·0.60H₂O: C, 57.23; H, 3.46; N, 7.15. Found: C, 57.22; H, 3.54; N, 6.91.

(E)-Methyl 4-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1H-pyrazol-1-yl}-4oxobut-2-enoate (60). Compound 60 was prepared from 22f (0.750g, 1.60 mmol) and (E)-methyl 4-chloro-4-oxobut-2-enoate (0.280 g, 1.90 mmol) using standard procedure G. The THF was removed under vacuum. The residue was dissolved in DCM and washed 3× with brine, and the organics were concentrated. The title compound was obtained as a yellow solid by flash chromatography using a 0-10% MeOH gradient in DCM. Yield 0.546 g, 59.0%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.32 (s, 1H), 7.70 (dt, I = 8.3, 1.9 Hz, 1H), 7.67 (dt, J = 8.1, 2.0 Hz, 1H), 7.60 (t, J = 7.8 Hz, 1H), 7.44 (d, J= 8.3 Hz, 1H), 7.37-7.32 (m, 3H), 7.30 (dt, J = 8.2, 2.0 Hz, 1H),7.27-7.22 (m, 1H), 7.16 (t, J = 7.6 Hz, 1H), 7.08 (d, J = 8.2 Hz, 1H), 6.95 (d, J = 7.6 Hz, 2H), 6.53 (dd, J = 15.5, 1.0 Hz, 1H), 5.49 (dd, J = 15.5) 11.6, 4.3 Hz, 1H), 3.86–3.73 (m, 4H), 3.03 (dd, *J* = 18.6, 4.2 Hz, 1H). 13 C NMR (100 MHz, DMSO- d_6) δ 165.13, 160.11, 159.92, 154.93, 150.40, 140.16, 138.71, 134.71, 133.36, 132.02, 131.69, 131.37, 131.30, 131.11, 130.80, 129.93, 128.65, 128.56, 127.57, 127.46, 122.77, 122.50, 121.77, 119.01, 115.63, 58.56, 52.16, 45.15. HRMS (m/z): $[M + H]^+$ calcd for C₂₉H₂₂ClBrN₃O₄, 590.04767; found, 590.04887. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 1.56$ min; >90% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 1.39$ min; >90% purity.

5-{3-[4-(4-Bromophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-5-(4-chlorophenyl)-4,5-dihydro-1H-pyrazol-1-yl}-5-oxopentanoic Acid (61). Compound 61 was prepared according to general procedure G using 88 (0.071 g, 0.63 mmol) and 22f (0.30 g, 0.63 mmol). Upon completion, the THF was removed under vacuum and the residue was dissolved in hot EtOAc. When the mixture was cooled, a yellow solid formed which was filtered, dried under vacuum, and determined to be the desired product. Yield 0.204 g, 54.9%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 12.09 (s, 1H), 7.77 (dd, J =8.2, 2.1 Hz, 1H), 7.70 (dd, J = 8.2, 2.1 Hz, 1H), 7.58 (t, J = 7.7 Hz, 1H), 7.43 (d, I = 8.3 Hz, 1H), 7.36 (dd, I = 8.2, 2.2 Hz, 1H), 7.34— 7.28 (m, 2H), 7.23 (dd, J = 8.1, 2.3 Hz, 1H), 7.15 (t, J = 7.6 Hz, 1H), 7.04 (d, J = 8.2 Hz, 1H), 6.86-6.76 (m, 2H), 5.36 (dd, J = 12.0, 4.4Hz, 1H), 3.81-3.68 (m, 1H), 2.80 (dd, J = 18.7, 4.3 Hz, 1H), 2.40 (dt, I = 15.3, 7.4 Hz, 1H), 2.29-2.06 (m, 3H), 1.68-1.51 (m, 2H).NMR (100 MHz, DMSO- d_6) δ 174.11, 169.29, 160.18, 152.73, 149.89, 141.29, 138.57, 134.59, 131.73, 131.64, 131.47, 131.30, 131.25, 130.88, 128.40, 127.31, 127.27, 123.41, 122.44, 121.77, 119.09, 115.60, 58.13, 45.10, 32.96, 32.58, 19.82. HRMS (m/z): $[M + H]^+$ calcd for C₂₉H₂₄ClBrN₃O₄, 592.06332; found, 592.06461. Anal. Calcd for C₂₉H₂₃ClBrN₃O₄: C, 58.75; H, 3.91; N, 7.09. Found: C, 58.47; H, 3.96; N. 6.91.

5-[3-(6-Chloro-2-oxo-4-phenyl-1,2-dihydroquinolin-3-yl)-5-(4-nitrophenyl)-4,5-dihydro-1H-pyrazol-1-yl]-5-oxopentanoic Acid (63). Compound 63 was prepared according to general procedure G using glutaric anhydride (89) (0.077 g, 0.67 mmol) and 63f (0.30 g, 0.67 mmol). The title compound was obtained after being dissolved in hot EtOAc followed by slow addition of hot hexanes to yield the title compound as a yellow solid. Yield 0.220 g, 58.4%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.44 (s, 1H), 12.12 (s, 1H), 8.12 (d, J 6.6 Hz, 1H), 7.07 (d, I = 8.5 Hz, 2H), 6.95 (d, I = 2.5 Hz, 1H), 5.50 (dd, *J* = 12.3, 4.8 Hz, 1H), 3.83 (dd, *J* = 18.6, 12.2 Hz, 1H), 2.86 (dd, *J* = 18.6, 4.9 Hz, 1H), 2.45–2.30 (m, 1H), 2.29–2.17 (m, 1H), 2.13 (t, J = 7.4 Hz, 2H), 1.65–1.45 (m, 2H). 13 C NMR (100 MHz, DMSO- d_6) δ 174.07, 169.49, 160.10, 152.49, 149.59, 146.59, 137.35, 131.30, 129.39, 128.47, 126.68, 126.08, 124.34, 123.82, 120.63, 117.67, 58.32, 44.90, 32.94, 32.53, 19.70. HRMS (m/z): $[M + H]^+$ calcd for $C_{29}H_{24}ClN_4O_6$, 559.13789; found, 559.13826. Anal. Calcd for C₂₉H₂₃ClN₄O₆: C, 62.31; H, 4.15; N, 10.02. Found: C, 62.42; H, 4.39; N, 9.72.

4-(4-Chlorophenyl)-3-[5-(4-chlorophenyl)-1-(4-hydroxybutanoyl)-4,5-dihydro-1H-pyrazol-3-yl]quinolin-2(1H)-one (64). In a flame-dried 25 mL round-bottomed flask, 26 (0.300 g, 0.560 mmol) was dissolved in THF (10 mL) and cooled on an ice bath to 0 °C under nitrogen with stirring. BH3-Me2S (2.0 M in hexanes, 0.561 mL, 2 equiv) was added dropwise. The mixture was stirred for 30 min, quenched with MeOH, and the solvent was removed under vacuum. The resultant residue was dissolved in DCM, washed three times with brine, and the organics were combined, dried over magnesium sulfate, concentrated in vacuo, and column-chromatographed using a 0-8% gradient of MeOH in DCM to give the title compound as a yellow solid. Yield 0.063 g, 21.6%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 7.65–7.49 (m, 3H), 7.46–7.24 (m, 5H), 7.20–7.00 (m, 2H), 6.89-6.78 (m, 2H), 5.35 (dd, I = 12.0, 4.4 Hz, 1H), 4.45 (t, I = 5.2 Hz, 1H), 3.74 (dd, J = 18.5, 12.1 Hz, 1H), 3.41-3.27 (m, 2H), 2.80 (dd, J= 18.5, 4.5 Hz, 1H), 2.44-2.19 (m, 2H), 1.59-1.39 (m, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 169.82, 160.17, 152.52, 149.83, 141.31, 138.53, 134.18, 133.14, 131.58, 131.43, 131.39, 130.57, 129.11, 128.33, 127.28, 123.50, 122.41, 119.14, 115.57, 60.19, 58.10, 45.06, 30.14, 27.71. HRMS (m/z): $[M + H]^+$ calcd for $C_{28}H_{24}Cl_2N_3O_3$, 520.11892; found, 520.11993. Anal. Calcd for C₂₈H₂₃Cl₂N₃O₃·0.40H₂O: C, 63.74; H, 4.55; N, 7.96. Found: C, 63.75; H, 4.33; N, 7.89.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}-4-oxobutanamide (65). In a flame-dried 25 mL round-bottomed flask, 26 (0.300 g, 0.560 mmol), 4-dimethylaminopyridine (0.069 g, 0.560 mmol), and N1-((ethylimino)methylene)-N3,N3-dimethylpropane-1,3-diamine (0.096 g, 0.618 mmol) were added to THF (11.23 mL) at 0 °C and stirred for 45 min. Ammonia (0.5 M in dioxane, 1.0 equiv, 1.1 mL) was added to the flask, and the reaction mixture was stirred overnight while being allowed to warm to room temperature. The reaction was quenched with dilute HCl (0.1 M), and the organics were removed under vacuum. The resultant residue was dissolved in DCM, washed 3× with brine, and the organics were dried over magnesium sulfate and concentrated in vacuo prior to column chromatography using a 0-8% MeOH gradient in DCM (0.1% Et₃N). The title compound was obtained as a white solid. Yield 0.019 g, 6.35%. ¹H NMR (400 MHz, DMSO- d_6) δ 12.29 (s, 1H), 7.66–7.56 (m, 2H), 7.55–7.49 (m, 1H), 7.43 (d, J = 8.4 Hz, 2H), 7.28 (dd, J = 8.2, 4.4 Hz, 3H), 7.24 (s, 1H), 7.15 (t, I = 7.6 Hz, 1H), 7.03 (d, I = 8.2 Hz, 1H), 6.79 (d, I = 8.5 Hz, 2H), 6.73 (s, 1H), 5.34 (dd, J = 12.1, 4.5 Hz, 1H), 3.71 (dd, J = 18.4, 12.2 Hz, 1H), 2.72 (dd, J = 18.4, 4.5 Hz, 1H), 2.59–2.50 (m, 2H), 2.20 (t, J = 7.2 Hz, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 173.69, 169.78, 160.79, 153.18, 150.49, 141.85, 139.21, 134.63, 133.87, 132.22, 132.19, 132.09, 131.23, 128.99, 128.95, 127.94, 124.23, 123.07, 119.80, 116.24, 58.83, 46.27, 45.80, 29.94, 29.44. HRMS (m/z): $[M + H]^+$ calcd for $C_{28}H_{23}Cl_2N_4O_3$, 533.11417; found, 533.11517. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_R = 0.89$ min; >90% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 1.00$ min; >95% purity.

Methyl 4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1*H*-pyrazol-1-yl}butanoate (66). In a 20 mL round-bottomed flask, 26f (0.500 g, 1.151 mmol) and methyl 4-oxobutanoate (0.121 mL, 1.151 mmol) were dissolved in DCE (11.51 mL). The mixture was allowed to stir at room temperature for 4 h, and sodium triacetoxyborohydride (0.293 g, 1.381 mmol) was added in one portion. The reaction was monitored by TLC and HPLC-MS. Upon completion, the DCE was removed under vacuum. The residue was diluted with DCM and washed 3× with brine. The organics were concentrated, and the title compound was obtained from flash column chromatography using 0-10% MeOH in DCM and trituration of the compound from ether as a yellow solid. Yield 0.150 g, 24.4%. ¹H NMR (400 MHz, CDCl₃) δ 12.22 (s, 1H), 7.55-7.45 (m, 3H), 7.40 (s, 1H), 7.34-7.26 (m, 5H), 7.22 (s, 2H), 7.13 (t, J = 7.6 Hz, 1H), 4.11 (dd, J = 13.6, 10.1 Hz, 1H), 3.62 (s, 3H), 3.41 (dd, J = 16.4, 10.1 Hz, 1H), 2.91–2.78 (m, 1H), 2.70–2.53 (m, 2H), 2.30-2.08 (m, 2H), 1.84-1.66 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 174.18, 162.94, 150.63, 146.34, 139.59, 138.08, 135.07, 134.23, 133.49, 131.12, 130.90, 129.08, 128.89, 128.51, 128.33, 127.82, 124.96, 122.96, 120.65, 116.18, 70.81, 52.20, 51.66, 46.60, 31.30, 23.04. HRMS (m/z): $[M + H]^+$ calcd for $C_{29}H_{26}Cl_2N_3O_3$, 534.13457;

found, 534.13624. HPLC 85% MeOH/H₂O (0.1% formic acid) $t_{\rm R}$ = 2.35 min; >95% purity; 75% ACN/H₂O (0.1% formic acid) $t_{\rm R}$ = 2.09 min; >95% purity.

4-{5-(4-Chlorophenyl)-3-[4-(4-chlorophenyl)-2-oxo-1,2-dihydroquinolin-3-yl]-4,5-dihydro-1H-pyrazol-1-yl}butanoic Acid (67). In a 10 mL round-bottomed flask, compound 66 (0.100 g, 0.187 mmol) was dissolved in NaOH (0.711 mL, 0.711 mmol), H₂O (10.00 mL), and ethanol (6.24 mL). The mixture was stirred at room temperature for 4 h and monitored by TLC/LC-MS. Upon completion, HCl (0.711 mL, 0.711 mmol) was added, giving a bright yellow solid which was filtered, dissolved in DCM, and washed 3× with acidified (pH 2, HCl) brine. The organics were combined, dried over magnesium sulfate and concentrated in vacuo. The title compound was obtained by flash chromatography using 0-8% MeOH in DCM as a yellow solid. Yield 0.050 g, 51.3%. ¹H NMR (600 MHz, DMSO- d_6) δ 12.16 (s, 1H), 11.92 (s, 1H), 7.59–7.49 (m, 3H), 7.42-7.37 (m, 3H), 7.35-7.29 (m, 4H), 7.13 (t, J = 7.6 Hz, 1H), 7.07 (d, I = 8.2 Hz, 1H), 4.03 (dd, I = 13.8, 10.2 Hz, 1H), 3.33 (dd, I = 13.8, 1H), 3.33 (dd, I = 13.8), 1H) 16.4, 10.2 Hz, 1H), 2.73 (dd, J = 16.5, 13.8 Hz, 1H), 2.54-2.46 (m, 1H), 2.40–2.31 (m, 1H), 2.12–1.96 (m, 2H), 1.55–1.39 (m, 2H). ¹³C NMR (150 MHz, DMSO- d_6) δ 174.25, 160.57, 148.75, 146.65, 140.21, 138.31, 135.02, 132.63, 131.86, 131.23, 131.19, 130.84, 130.75, 129.09, 128.44, 128.08, 127.81, 127.04, 125.16, 122.15, 119.31, 115.43, 69.48, 52.16, 46.06, 30.83, 22.35. HRMS (m/z): $[M + H]^+$ calcd for C₂₈H₂₄Cl₂N₃O₃, 520.11892; found, 520.11970. HPLC 85% MeOH/ H_2O (0.1% formic acid) $t_R = 1.33$ min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 0.98 \text{ min; } >95\% \text{ purity.}$

4-(4-Chlorophenyl)-3-[5-(4-chlorophenyl)-1-(4-fluorobutanoyl)-4,5-dihydro-1H-pyrazol-3-yl]quinolin-2(1H)-one (68). In a 10 mL round-bottomed flask, 4-fluorobutanoic acid 68g (0.100 g, 0.943 mmol), DMAP (0.127 g, 1.037 mmol), and EDCI (0.199 g, 1.037 mmol) were dissolved in DCM (9.43 mL) which had been precooled to 0 °C. The mixture was stirred for 45 min prior to the addition of the 26f (0.409 g, 0.943 mmol). The mixture was allowed to warm to room temperature and monitored by TLC. The reaction was quenched with 0.2 N HCl and extracted into DCM. The organics were washed 3× with brine, dried over magnesium sulfate, and concentrated. The title compound was obtained as a yellow solid after column chromatography using a gradient of 0-50% EtOAc in DCM as a yellow solid. Yield 0.050 g, 21.6%. ¹H NMR (400 MHz, CDCl₃) δ 13.16 (s, 1H), 7.62–7.43 (m, 4H), 7.38 (d, J = 8.2 Hz, 1H), 7.34-7.16 (m, 5H), 7.02 (d, J = 8.3 Hz, 2H), 5.42 (dd, J = 11.8, 4.0Hz, 1H), 4.44 (dtd, J = 47.3, 5.8, 1.7 Hz, 2H), 3.70 (dd, J = 18.2, 11.8 Hz, 1H), 3.15 (dd, J = 18.3, 4.1 Hz, 1H), 2.68-2.45 (m, 2H), 2.18-1.82 (m, 2H). 13 C NMR (150 MHz, CDCl₃) δ 170.45, 162.91, 152.52, 151.98, 151.92, 140.41, 138.42, 135.43, 134.99, 133.97, 133.56, 132.00, 130.81, 130.72, 130.68, 130.36, 129.05, 128.96, 128.85, 128.73, 128.70, 128.04, 127.60, 127.52, 126.86, 123.47, 123.09, 120.36, 116.41, 84.06, 82.97, 59.35, 45.44, 29.91, 29.88, 25.69, 25.62, 25.56. ¹⁹F NMR (282 MHz, CDCl₃) δ –220.53 (tt, J = 47.3, 26.3 Hz). HRMS (m/z): [M + H]+ calcd for C₂₈H₂₃Cl₂N₃O₂F, 522.11459; found, 522.11462. HPLC 85% MeOH/ H_2 O (0.1% formic acid) t_R = 1.46 min; >95% purity; 75% ACN/ H_2O (0.1% formic acid) $t_R = 1.32$ min; >95% purity.

ASSOCIATED CONTENT

Supporting Information

Experimental details for the synthesis of all intermediates, solubility determination methods and data analysis, Table S1 with A-ring modifications, Table S2 with heteroaromatic C-ring substitutions, Table S3 with C-ring modifications, Table S4 with off-target data from NIMH-PDSP, Figure S1 showing mutant receptor responses, Figure S2 showing plasma stability, compound solubility, and Scheme S1 showing the synthesis of the monofluorobutyrate. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare the following competing financial interest(s): Several of the authors are co-inventors (T.M.A., D.C.L., S.F.T.) of Emory University owned patent-pending technology and have an equity position (D.C.L., S.F.T., J.P.S.), are board members (D.C.L.), or are paid consultants (S.F.T.) for a company that has licensed this technology.

ACKNOWLEDGMENTS

The authors thank the Emory Institute for Drug Development for performing the plasma stability assays, Dr. Terry W. Moore, Dr. Alexandra Orchard, and Eric Miller for helpful discussions and comments on the manuscript, Drs. Shaoxiong Wu and Fred Strobel for excellent core facility support, and Phuong Le and Jing Zhang for excellent technical assistance. This work was support by the National Institutes of Health (NIH-NINDS Grant NS071802 (T.M.A.) and Grants NS065371, NS036654 (S.F.T.)) and an NIH Graduate Training in the Pharmacological Sciences (Grant GM008602 (T.M.A.)), an NIH Translational Research in Neurology (Grant 5T32-NS007480-07 (K.M.V.)), the University of KwaZulu-Natal and the NRF (C.S.), and Lundbeck A/S and Pfizer Inc. research grants to Emory University.

ABBREVIATIONS USED

Ca²⁺, calcium ion; Mg²⁺, magnesium ion; ATD, amino-terminal domain; LBD, ligand-binding domain; DQP, dihydroquinolone pyrazoline; BH₃–DMS, borane–dimethylsulfide; NH₃, ammonia; μ M, micromolar; mV, millivolt; QNZ, quinazoline-4-one; S2, segment 2; TPSA, topological polar surface area; $P_{\rm app}$, permeability coefficient; MDR1-MDCK, multidrug resistance gene 1 Madin–Darby canine kidney; cRNA, complementary RNA; NaCl, sodium chloride; KCl, potassium chloride; BaCl₂, barium chloride; NaOH, sodium hydroxide; M Ω , megaohm; NIMH PDSP, National Institute of Mental Health psychoactive drug screening program; ACN, acetonitrile; EtOAc, ethyl acetate

REFERENCES

- (1) Mayer, M. L. Glutamate receptor ion channels. Curr. Opin. Neurobiol. 2005, 15, 282–288.
- (2) Traynelis, S. F.; Wollmuth, L. P.; McBain, C. J.; Menniti, F. S.; Vance, K. M.; Ogden, K. K.; Hansen, K. B.; Yuan, H.; Myers, S. J.; Dingledine, R.; Sibley, D. Glutamate receptor ion channels: structure, regulation, and function. *Pharmacol. Rev.* **2010**, *62*, 405–496.
- (3) Wang, P. Y.; Petralia, R. S.; Wang, Y.-X.; Wenthold, R. J.; Brenowitz, S. D. Functional NMDA receptors at axonal growth cones of young hippocampal neurons. *J. Neurosci.* **2011**, *31*, 9289–9297.
- (4) Cull-Candy, S. G.; Leszkiewicz, D. N. Role of distinct NMDA receptor subtypes at central synapses. *Sci. STKE* **2004**, 2004, re16.
- (5) Pérez-Otaño, I.; Ehlers, M. D. Homeostatic plasticity and NMDA receptor trafficking. *Trends Neurosci.* **2005**, 28, 229–238.
- (6) Mony, L.; Kew, J. N. C.; Gunthorpe, M. J.; Paoletti, P. Allosteric modulators of NR2B-containing NMDA receptors: molecular mechanisms and therapeutic potential. *Br. J. Pharmacol.* **2009**, *157*, 1301–1317.
- (7) Chen, H.-S. V.; Lipton, A. S. The chemical biology of clinically tolerated NMDA receptor antagonists. *J. Neurochem.* **2006**, 97, 1611–1626

- (8) Hallett, P. J.; Standaert, D. G. Rationale for and use of NMDA receptor antagonists in Parkinson's disease. *Pharmacol. Ther.* **2004**, 102. 155–174.
- (9) Hollmann, M.; Heinemann, S. Cloned glutamate receptors. *Annu. Rev. Neurosci.* **1994**, *17*, 31–108.
- (10) Durand, G. M.; Gregor, P.; Zheng, X.; Bennett, M. V.; Uhl, G. R.; Zukin, R. S. Cloning of an apparent splice variant of the rat *N*-methyl-D-aspartate receptor NMDAR1 with altered sensitivity to polyamines and activators of protein kinase C. *Proc. Natl. Acad. Sci. U.S.A.* **1992**, *89*, 9359–9363.
- (11) Furukawa, H.; Gouaux, E. Mechanisms of activation, inhibition and specificity: crystal structures of the NMDA receptor NR1 ligand-binding core. *EMBO J.* **2003**, *22*, 2873–2885.
- (12) Furukawa, H.; Singh, S. K.; Mancusso, R.; Gouaux, E. Subunit arrangement and function in NMDA receptors. *Nature* **2005**, 438, 185–192.
- (13) Chen, P. E.; Wyllie, D. J. Pharmacological insights obtained from structure—function studies of ionotropic glutamate receptors. *Br. J. Pharmacol.* **2006**, *147*, 839—853.
- (14) Paoletti, P.; Neyton, J. NMDA receptor subunits: function and pharmacology. *Curr. Opin. Pharmacol.* **2007**, *7*, 39–47.
- (15) Vance, K. M.; N., S.; Traynelis, S. F.; Furukawa, H. Ligand-specific deactivation time course of GluN1/GluN2D NMDA receptors. *Nat. Commun.* **2011**, *2*, 294.
- (16) Gielen, M.; Retchless, B. S.; Mony, L.; Johnson, J. W.; Paoletti, P. Mechanism of differential control of NMDA receptor activity by NR2 subunits. *Nature* **2009**, *459*, 703–707.
- (17) Yuan, H.; Hansen, K. B.; Vance, K. M.; Ogden, K. K.; Traynelis, S. F. Control of NMDA receptor function by the NR2 subunit aminoterminal domain. *J. Neurosci.* **2009**, *29*, 12045–12058.
- (18) Lester, R. A. J.; Clements, J. D.; Westbrook, G. L.; Jahr, C. E. Channel kinetics determine the time course of NMDA receptor-mediated synaptic currents. *Nature* **1990**, *346*, 565–567.
- (19) Nowak, L.; Bregestovski, P.; Ascher, P.; Herbet, A.; Prochiantz, A. Magnesium gates glutamate-activated channels in mouse central neurons. *Nature* **1984**, *307*, 462–465.
- (20) Mayer, M. L.; Westbrook, G. L.; Guthrie, P. B. Voltage-dependent block by Mg²⁺ of NMDA responses in spinal-cord neurons. *Nature* **1984**, *309*, 261–263.
- (21) Clarke, R. J.; Johnson, J. W. NMDA receptor NR2 subunit dependence of the slow component of magnesium unblock. *J. Neurosci.* **2006**, *26*, 5825–5834.
- (22) Monyer, H.; Burnashev, N.; Laurie, D. J.; Sakmann, B.; Seeburg, P. H. Developmental and regional expression in the rat brain and functional properties of four NMDA receptors. *Neuron* **1994**, *12*, 529–540.
- (23) Akazawa, C.; Shigemoto, R.; Bessho, Y.; Nakanishi, S.; Mizuno, N. Differential expression of five *N*-methyl-D-aspartate receptor subunit mRNAs in the cerebellum of developing and adult rats. *J. Comp. Neurol.* **1994**, 347, 150–160.
- (24) Galvan, A.; Wichmann, T. Pathophysiology of parkinsonism. Clin. Neurophysiol. 2008, 119, 1459–1474.
- (25) Bräuner-Osborne, H.; Egebjerg, J.; Nielsen, E. Ø.; Madsen, U.; Krogsgaard-Larsen, P. Ligands for glutamate receptors: design and therapeutic prospects. *J. Med. Chem.* **2000**, *43*, 2609–2645.
- (26) Standaert, D. G.; Testa, C. M.; Penney, J. B., Jr; Young, A. B. Alternatively spliced isoforms of the NMDAR1 glutamate receptor subunit: differential expression in the basal ganglia of the rat. *Neurosci. Lett.* **1993**, *152*, 161–164.
- (27) Laurie, D. J.; Seeburg, P. H. Regional and developmental heterogeneity in splicing of the rat brain NMDAR1 mRNA. *J. Neurosci.* **1994**, *14*, 3180–3194.
- (28) Standaert, D. G.; Testa, C. M.; Young, A.; Penney, J. B., Jr. Organization of *N*-methyl-D-aspartate glutamate receptor gene expression in the basal ganglia of the rat. *J. Comp. Neurol.* **1994**, 343, 1–16.
- (29) Goff, D. C.; Cather, C.; Gottlieb, J. D.; Evins, A. E.; Walsh, J.; Raeke, L.; Otto, M. W.; Schoenfeld, D.; Green, M. F. Once-weekly Dcycloserine effects on negative symptoms and cognition in

- schizophrenia: an exploratory study. Schizophr. Res. 2008, 106, 320–327.
- (30) Acker, T. M.; Yuan, H.; Hansen, K. B.; Vance, K. M.; Ogden, K. K.; Jensen, H. S.; Burger, P. B.; Mullasseril, P.; Snyder, J. P.; Liotta, D. C.; Traynelis, S. F. Mechanism for noncompetitive inhibition by novel GluN2C/D *N*-methyl-D-aspartate receptor subunit-selective modulators. *Mol. Pharmacol.* **2011**, *80*, 782–795.
- (31) Frye, S. V.; Johnson, M. C.; Valvano, N. L. Synthesis of 2-aminobenzophenones via rapid halogen—lithium exchange in the presence of a 2-amino-*N*-methoxy-*N*-methylbenzamide. *J. Org. Chem.* **1991**, *56*, 3750–3752.
- (32) Kim, D. W.; Jeong; Lim, S. T.; Sohn, M.-H.; Katzenellenbogen, J. A.; Chi, D. Y. Facile nucleophilic fluorination reactions using *tert*-alcohols as a reaction medium: significantly enhanced reactivity of alkali metal fluorides and improved selectivity. *J. Org. Chem.* **2008**, 73, 957–962.
- (33) O'Hagan, D. Preparation of monofluorocarboxylic acids using *N*,*N*-diethyl-1,1.2,3,3,3-hexafluoropropylamine. *J. Fluorine Chem.* **1989**, 43, 371–377.
- (34) Topliss, J. G. A manual method for applying the Hansch approach to drug design. J. Med. Chem. 1977, 20, 463-469.
- (35) Hansch, C.; Leo, A. J. Substituent Constants for Correlation Analysis in Chemistry and Biology; John Wiley and Sons: New York, 1979; pp 67–161.
- (36) Hansen, K. B.; Traynelis, S. F. Structural and mechanistic determinants of a novel site for noncompetitive inhibition of GluN2D-containing NMDA receptors. *J. Neurosci.* **2011**, *31*, 3650–3661.
- (37) Hitchcock, S. A.; Pennington, L. D. Structure-brain exposure relationships. *J. Med. Chem.* **2006**, *49*, 7559–7583.
- (38) Wang, Q.; Rager, J. D.; Weinstein, K.; Kardos, P. S.; Dobson, G. L.; Li, J.; Hidalgo, I. J. Evaluation of the MDR-MDCK cell line as a permeability screen for the blood—brain barrier. *Int. J. Pharm.* **2005**, 288, 349—359.
- (39) Hardegger, L. A.; Kuhn, B.; Spinnler, B.; Anselm, L.; Ecabert, R.; Stihle, M.; Gsell, B.; Thoma, R.; Diez, J.; Benz, J.; Plancher, J.-M.; Hartmann, G.; Banner, D. W.; Haap, W.; Diederich, F. Systematic investigation of halogen bonding in protein—ligand interactions. *Angew. Chem., Int. Ed.* **2011**, *50*, 314—318.
- (40) Parisini, E.; Metrangolo, P.; Pilati, T.; Resnati, G.; Terraneo, G. Halogen bonding in halocarbon—protein complexes: a structural survey. *Chem. Soc. Rev.* **2011**, *40*, 2267—2278.
- (41) Wilson, C. J.; Bevan, M. D. Intrinsic dynamics and synaptic inputs control the activity patterns of subthalamic nucleus neurons in health and in Parkinson's disease. *Neuroscience* **2011**, *198*, 54–68.
- (42) Wichmann, T.; DeLong, M. R. Models of basal ganglia function and pathophysiology of movement disorders. *Neurosurg. Clin. North Am.* 1998, 9, 223–226.
- (43) Bolam, J. P.; Hanley, J. J.; Booth, P. A. C.; Bevan, M. D. Synaptic organisation of the basal ganglia. *J. Anat.* **2000**, *196*, 527–542.
- (44) Mullasseril, P.; Hansen, K. B.; Vance, K. M.; Ogden, K. K.; Yuan, H.; Kurtkaya, N. L.; Santangelo, R.; Orr, A. G.; Le, P.; Vellano, K. M.; Liotta, D. C.; Traynelis, S. F. A subunit-selective potentiator of NR2C- and NR2D-containing NMDA receptors. *Nat. Commun.* **2010**, *1*, 90.
- (45) Surmeier, D. J.; Mercer, J. N.; Chan, C. S. Autonomous pacemakers in the basal ganglia: Who needs excitatory synapses anyway? *Curr. Opin. Neurobiol.* **2005**, *15*, 312–318.
- (46) Dravid, S. M.; Burger, P. B.; Prakash, A.; Geballe, M. T.; Yadav, R.; Le, P.; Vellano, K.; Snyder, J. P.; Traynelis, S. F. Structural determinants of D-cycloserine efficacy at the NR1/NR2C NMDA receptors. *J. Neurosci.* **2010**, *30*, 2741–2754.
- (47) Lisman, J. E.; Coyle, J. T.; Green, R. W.; Javitt, D. C.; Benes, F. M.; Heckers, S.; Grace, A. A. Circuit-based framework for understanding neurotransmitter and risk gene interactions in schizophrenia. *Trends Neurosci.* **2008**, *31*, 234–242.
- (48) Hillman, B. G.; Gupta, S. C.; Stairs, D. J.; Buonanno, A.; Dravid, S. M. Behavioral analysis of NR2C knockout mouse reveals deficit in acquisition of conditioned fear and working memory. *Neurobiol. Learn. Mem.* **2011**, *95*, 404–414.

- (49) Mosley, C. A.; Acker, T. M.; Hansen, K. B.; Mullasseril, P.; Andersen, K. T.; Le, P.; Vellano, K. M.; Bräuner-Osborne, H.; Liotta, D. C.; Traynelis, S. F. Quinazolin-4-one derivatives: a novel class of noncompetitive NR2C/D subunit-selective N-methyl-D-aspartate receptor antagonists. *J. Med. Chem.* **2010**, *53*, 5476–5490.
- (50) Santangelo Freel, R. M.; Ogden, K. K.; Strong, K. L.; Khatri, A.; Chepiga, K. M.; Jensen, H. S.; Traynelis, S. F.; Liotta, D. C. Synthesis and structure activity relationship of tetrahydroisoquinoline-based potentiators of GluN2C and GluN2D containing *N*-methyl-D-aspartate receptors. *J. Med. Chem.* **2013**, *56*, 5351–5381.
- (51) Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. OLEX2: a complete structure solution, refinement and analysis program. *J. Appl. Crystallogr.* **2009**, 42, 339–341.
- (52) Sheldrick, G. A short history of SHELX. Acta Crystallogr., Sect. A: Found. Crystallogr. 2008, 64, 112–122.
- (53) Petříček, V.; Dušek, M.; Palatinus, L. *JANA2006. The Crystallographic Computing System*; Institute of Physics: Prague, Czech Republic, 2006.