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# Novel (Bisarylmethoxy)butylpiperidine Analogues as Neurotransmitter Transporter Inhibitors with Activity at Dopamine Receptor Sites

Sung-Woon Choi,<sup>a</sup> David R. Elmaleh,<sup>a</sup> Robert N. Hanson,<sup>b</sup> Timothy M. Shoup<sup>a</sup> and Alan J. Fischman<sup>a,\*</sup>

<sup>a</sup>Division of Nuclear Medicine, Department of Radiology, Massachusetts General Hospital, Boston, MA 02114, USA <sup>b</sup>Department of Chemistry, Northeastern University, Boston, MA 02115, USA

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Abstract—A series of (bisarylmethoxy)butylpiperidine derivatives was prepared and evaluated in vitro and in vivo to determine the structural requirements necessary for dual activity at the DAT and DA/5-HT receptor sites. These hybrid ligands, constructed by combining pharmacophores specific for the DAT and DA/5-HT receptors, could be useful drugs for treating cocaine addiction by assisting cocaine addicts in maintaining abstinence. The series was evaluated in vitro for DAT and DA/5-HT receptor activity and then selected compounds were tested in vivo for their effects on cocaine-induced hyperlocomotor activity (LMA). The majority of the new compounds demonstrated high to moderate affinity (4–191 nM) for the DAT with 4-hydroxy-4-phenylpiperidine analogues 14 and 15 possessing the greatest affinity. Compounds 15 and 22 exhibited the highest ratio of reuptake inhibition-to-binding (discrimination ratio, DR), 111 and 323, respectively. These derivatives had modest affinity and antagonistic activity for dopamine  $D_2/D_3$  receptors. Compounds 9 and 15 (DR = 0.9 and 111, respectively) stimulated locomotor activity, whereas the other compounds suppressed this response. All compounds tested except for 17 and 21 attenuated cocaine-induced hyperlocomotion. (C) 2002 Elsevier Science Ltd. All rights reserved.

# Introduction

Cocaine addiction represents one of the major public health problems in American society today.<sup>1</sup> Efforts have been directed toward identifying and developing new therapeutic agents that can mediate cocaine's acute behavior and reinforcing effects;<sup>2</sup> however, at present there are no effective long-term pharmacotherapy that can lessen the abuse of cocaine.

Cocaine is pharmacologically a nonselective drug interacting in the central nervous system at the dopamine transporter (DAT), serotonin transporter (SERT), and norepinephrine transporter (NET) sites by inhibiting reuptake of the dopamine (DA), serotonin (5-HT), and norephrine (NE) neurotransmitters, respectively. Substantial evidence indicates that the DAT is a critical recognition site that contributes significantly to the reinforcing properties of cocaine.<sup>3,4</sup> Hence, therapeutic drug development has focused on designing agents that target the DAT.

Dopamine agonists and antagonists as medications for cocaine abuse have been evaluated and extensively reviewed.<sup>5</sup> Although the behavioral effects of cocaine are not consistently altered by selective dopamine antagonists,<sup>6</sup> many studies have reported their mediation of the reinforcing effects of cocaine.<sup>7</sup> The dopamine  $D_3$  receptor has been of particular interest because of its relatively restricted localization within the limbic system compared with the dopamine  $D_2$  receptor and due to its role as a possible target for treatment of schizophrenia and drug abuse.<sup>8</sup> A recent report of selective inhibition of cocaine-seeking behavior by a partial dopamine  $D_3$  receptor agonist suggested that this receptor is an important target for the development of medications for cocaine abuse.<sup>9</sup>

Comparison of  $IC_{50}$  values for inhibition of [<sup>3</sup>H]DA reuptake and  $K_i$  values for DAT ligand-binding affinity (e.g., [<sup>125</sup>I]RTI-55) has been suggested as a method for evaluating cocaine antagonists or partial agonists in

<sup>\*</sup>Corresponding author. Tel.: +1-617-726-8353; fax: +1-617-726-6165; e-mail: fischman@pet.mgh.harvard.edu

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vitro.<sup>10,11</sup> In this context, important cocaine antagonists would display high discrimination ratios (ratio of reuptake inhibition-to-binding, DR) under comparable experimental conditions.<sup>10</sup> Although DR values from in vitro assays have not always been predictive of in vivo activities such as locomotor activation or depression, drug discrimination or substitution for cocaine and anti-cocaine activity, they can serve as an initial in vitro screening process.<sup>11</sup> While a pure cocaine antagonist at the DAT may be of interest in preventing the reinforcing effects of cocaine, a compound with some residual DA reuptake inhibition may have greater potential for the treatment of cocaine abuse. Such agents may serve to provide some relief from cocaine craving and alleviate the withdrawal symptoms encountered during initial stages of abstinence.

Various DAT ligands have been proposed as potential therapeutic agents for cocaine abuse including cocaine analogues<sup>12</sup> and disubstituted piperazine (GBR) analogues.<sup>13,14</sup> GBR analogues are particularly interesting because their pharmacologic properties include slow dissociation from the DAT, high transporter selectivity, and low intrinsic activities.<sup>15</sup> Oxygenated GBR analogues have been evaluated as DAT-specific, potentially long acting, anti-cocaine therapeutics for cocaine addiction.<sup>16</sup> Although the behavioral profile of GBR 12909 (1, Figure 1) was shown to be different from that of cocaine, this compound was reliably self-administered in nonhuman primates and fully substituted for cocaine in drug discrimination studies.<sup>17–20</sup> Such studies indicate that GBR 12909 might have the potential for abuse in humans,

albeit to a lesser extent compared to cocaine. Importantly, when the dopamine D2-selective antagonist, spiperon, was used to pretreat monkeys, the self-administration of GBR 12909 was attenuated.<sup>18</sup> In related studies, the 5-HT agonist, quipazine, reliably attenuated the behavioral-stimulus effects of GBR 12909.20 Therefore, assuming actions at the DAT are responsible for the behavioral effects of GBR 12909, it is possible that combining strong DAT binding properties and dopamine antagonistic and/or 5-HT agonistic properties in one molecule may offer a more effective treatment with less abuse liability potential than GBR 12909 and fewer side-effects observed from dopamine D2, D3 antagonist monotherapy. Although our explanation is speculative without previous neurobehavioral data from dual activity ligands, such compounds should block the binding of cocaine to the DAT with weak dopamine reuptake inhibition. Such residual dopamine uptake inhibiting properties will then be attenuated by activities at the dopamine receptor site(s) as antagonist and/or 5-HT agonist. Therefore, abuse liability that may arise from the activity of dopamine reuptake inhibition can be minimized.

As part of our program to develop potent DAT inhibitors, we have investigated numerous GBR analogues to identify structural features that confer high binding affinity for the DAT but weaker potency for inhibition of DA reuptake.<sup>21,22</sup> By extending our studies to measurements of DA/5-HT receptor activity, we have anticipated that some agents might influence the dopa-



minergic system in novel ways. In the current study, we used a hybrid drug approach which combines pharmacophores specific for the DAT and DA/5-HT receptors.

Structure-activity relationship (SAR) studies have demonstrated that the (bisarylmethoxy)ethyl and diamine moieties of GBR (1) are essential pharmacophores for retaining favorable activities at the DAT.<sup>13,14,22</sup> Also, at least one nitrogen atom (2) attached to the benzyl moiety is required.<sup>14,23</sup> Previously, the rigid piperazine ring of GBR (1) had been substituted with conformationally flexible polymethylene diamines and amine-amide moieties (3) without compromising biologic activities.<sup>22</sup> Tolerance of the amide function at the N2 nitrogen atom was interpreted as due to the possible presence of strong hydrogen-bonding capabilities at binding site(s) on the DAT protein. These acyclic analogues have provided a basis for the design of novel probes that demonstrate selective and high affinity binding to the DAT and weak potency for DA reuptake inhibition. In order to develop compounds with a mixed pharmacologic profile containing both DAT activity (binding/ reuptake inhibition) and dopamine antagonistic/5-HT agonistic properties, we incorporated structural moieties believed to induce dopamine  $D_2/D_3$  antagonism such as 4-aryl-4-piperidinol found in bromperidol and haloperidol and 5-HT activity found in arylpiperazine.24,25 Compound selection was also based on H-bonding and aryl functionality as elucidated in our previous findings with amine-amide analogues.<sup>26</sup> Ahmad et al.<sup>27</sup> reported benzhydroxyalkylpiperidine derivatives that were nonselective reuptake inhibitors of DA, 5-HT, and norepenephrine. As shown in the general structure 4, several important aspects of SAR were identified, including distance between the diaryl and piperidine moieties (*n*, distance as shown by the bold line in 1-3) and the nature of the substituents  $(R_1-R_4)$  on the piperidine ring. A specific three-dimensional arrangement of these groups (distance, basic nitrogen, hydrogen bonding, and Ar function) was deduced to test the hypothesis concerning which chemical groups (H-bonding, Ar function, and distance) are important for a particular biologic activity and what is the transporter-bound conformation of these groups. The results of our work, together with that of other investigators, led to the preparation of several arylpiperidine derivatives. In this study, we report the preparation and pharmacologic evaluation of a series of (bisarylmethoxy)butylpiperidine derivatives having mixed activities at the DAT and DA/5-HT receptor sites.

#### **Results and Discussion**

### Chemistry

The preparation of the GBR analogues is depicted in Scheme 1. The requisite starting materials **6** and  $7^{22}$ were prepared according to literature procedures with or without modifications, and **8** was purchased. The *N*-alkylated target compounds were prepared in good yields (73–87%) by alkylation of the appropriate amine with alkylating agents (**6–8**) in the presence of K<sub>2</sub>CO<sub>3</sub> as base and KI as catalyst. The final products were converted into oxalate salts for elemental analysis and biologic evaluation.

## In vitro characterization

This research was designed to develop new agents with higher DAT affinity and low potency for DA reuptake inhibition by selective modification of the substituents on a piperidine ring template structure (4). In order to find new lead compound(s) for potential cocaine antagonists, we focused on the introduction of new moieties for hydrogen bonding and aromatic function. Such modifications are presumed to be critical structural determinants for dual activity at DAT and DA/ 5-HT receptor sites. A new series of (bisarylmethoxy) alkylpiperidine analogues was prepared and characterized. The analogues were evaluated for their ability to displace  $[^{125}I]$ RTI-55, a cocaine analogue (K<sub>i</sub> values in Table 1) as well as for potency of reuptake inhibition of DA, 5-HT and NE (IC<sub>50</sub> values in Table 2) with HEK 293 cells expressing cDNA for human dopamine transporter (hDAT), human serotonin transporter (hSERT) or human norepinephrine transporter (hNET). In addition, some compounds were evaluated for their ability to bind and function at DA and 5-HT receptor sites (Table 3). Our initial targets were dopamine D2 and D3 receptors since the target compounds contain pharmacophores for dopamine receptors as well as 5-HT receptors as non-selective antagonists (mostly dopamine D2 and D3). However, it seems reasonable to test all dopamine receptor subtypes to get a wider view of ligand effects on dopaminergic and serotonergic systems, especially regarding the interpretation of locomotor activities. The final products were submitted to the National Institute on Drug Abuse, Cocaine Treatment Development Program (CTDP) for pharmacologic screening (Table 4).

## Neurotransporter activities<sup>28</sup>

Substitution on the piperidine ring by –OH and –CONH<sub>2</sub> groups (9-11) without an aromatic group produced moderate binding activity ( $K_i = 91 - 114$  nM) with low selectivity at the DAT (DR = 0.6-1.3). The introduction of an acetyl group and aromatic function at the 4-position of the piperidine ring (13) resulted in a compound that retained significant binding activity at the DAT with a DR of 48. Substitution of -COCH<sub>3</sub> with -CN (12, non-H-bonding group) decreased activities dramatically. Further modification of the acetyl group to a hydroxyl group (14) caused a marked increase in DAT affinity with reduced selectivity at the SERT (6-3) and decreased DR (48–15). When a large halogen atom (Br, 15) was introduced at the 4-position of the aromatic ring, selectivity and potency of binding at the DAT were improved (15 vs 14) and a significant increase in DR (15–111) was achieved. A change in alkyl chain length (n=4-2, 15 vs 16) decreased activities at the DAT  $(K_i = 4-191 \text{ nM}, \text{ DR} = 111-37)$ . Substitution of the bisarylmethoxy group with the phthalimide group (24 vs 14) caused loss of activities in some cases. Introduction of the  $-CF_3$  group at the 3-position of the aromatic group (14-17) resulted in weakly selective activities at



Scheme 1. (a) concd  $H_2SO_4$ , PhCH<sub>3</sub>; (b)  $K_2CO_3$ , KI, DMF, 60–70 °C.

the NET site ( $K_i = 10$  nM, IC<sub>50</sub> = 780 nM). When an aryl group was attached to the piperidine by a CH<sub>2</sub> linkage (benzyl group, 18), selective binding at the DAT was maintained but the selectivity of reuptake inhibition was shifted to the NET site from the DAT site (14, 15 vs 18). When the aromatic group was attached directly to the piperidine ring (19, 20) without an H-bonding group, the binding was strongest at the SERT site  $(K_{i} = 18 \text{ and } 25 \text{ nM}, \text{ respectively})$  while selective reuptake inhibition of NE was maintained. Additional aromatic groups on the piperidine ring (20-21) were detrimental and resulted in loss of activity at the DAT. Compounds 22 and 23 contained both H-bonding (as amide) and Ar groups but the activities at the transporters were different for each compound. Compound 22 represents the strongest binding ligand at the DAT  $(\hat{K}_i = 1 \text{ nM})$  and has the highest DR value (323). From this limited data set, it is apparent that the potency in the DAT binding assay and reuptake inhibition are relatively dependent on substitution and position on the piperidine ring.

# DA/5-HT receptor activities<sup>29</sup>

Compounds 14-17, 22 and 23 were evaluated for their ability to bind and function at DA and 5-HT receptor sites which is based on the fact that 4-hydroxy-4-arylpiperidines are the basic pharmacophore for activity at DA receptor sites. The potency and selectivity of the selected target compounds for DA and 5-HT receptors were evaluated using ligand displacement assays (see Table 3 for radioligands and cell types used). D<sub>2</sub> and D<sub>3</sub> antagonistic activities were assessed in vitro by the ability of the compounds to block [<sup>3</sup>H]thymidine incorporation (inhibition of mitogenesis) induced by 10 nM quinpirole in  $CHO_p$ -D<sub>2</sub> and D<sub>3</sub> cells (Table 3). Most compounds evaluated had weak to moderate affinity and antagonistic activity at D<sub>2</sub> and/or D<sub>3</sub> sites. (Bisarylmethoxy)butylpiperidines (4) are nonselective neurotransmitter inhibitors,<sup>27</sup> and bromperidol (5) is a potent nonselective dopamine  $D_2/D_3$  receptor antagonist.<sup>30</sup> The combination of both structural elements in 15 provided a compound that was both a very potent DAT binder (DR value of 111) and a moderate dopamine  $D_2/D_3$  receptor antagonist.

Table 1. Binding affinities and selectivities of bisarylmethoxybutylpiperidine derivatives at the DA, 5-HT, and NE transporters labeled with  $[^{125}I]RTI-55$ 

Compound		Binding $(K_i \pm SD [nM])$	Ratios		
	DAT	SERT	NET	SERT/DAT	NET/DAT
9	$114 \pm 27^{a}$	$423\!\pm\!98^a$	$610 \pm 130^{a}$	3.7	5.4
10	$91 \pm 26$	$437 \pm 66$	$611 \pm 76$	4.8	6.7
11	$94 \pm 42$	$132 \pm 26$	$578 \pm 52$	1.4	6.1
12	$135 \pm 34$	$311 \pm 58$	$368 \pm 14$	2.3	2.7
13	$21.7 \pm 8.0$	$126 \pm 45$	$114 \pm 4.1$	5.8	5.3
14	$6.4 \pm 1.2$	$17.9 \pm 8.8$	$54 \pm 16$	2.8	8.4
15	$3.9 \pm 1.8$	$38 \pm 10$	$250 \pm 110$	9.7	64.1
16	$191 \pm 78$	$590 \pm 180$	$1240 \pm 310$	3.1	6.5
17	$15.8 \pm 6.1$	$57 \pm 16$	$10 \pm 1.9$	3.6	0.6
18	$41 \pm 12$	$62 \pm 27$	$175 \pm 47$	1.5	4.3
19	$66 \pm 20$	$18.4 \pm 4.1$	$260 \pm 41$	0.3	3.9
20	$73 \pm 33$	$25.3 \pm 7.2$	$371 \pm 35$	0.4	5.1
21	$375 \pm 90$	$3000 \pm 1100$	$4600 \pm 1800$	8	12.3
22	$1.32 \pm 0.49$	$3.8 \pm 1.9$	$22.2 \pm 3.6$	2.9	16.8
23	$125 \pm 22$	$192 \pm 95$	$356 \pm 36$	1.5	2.9
24	$>10 \ \mu M$	$1590 \pm 43$	$> 10 \ \mu M$	< 0.2	1
GBR 12909	$27 \pm 8$	$186 \pm 30$	$163 \pm 39$	6.9	6.0
Cocaine <sup>b</sup>	$915 \pm 214$	$419 \pm 12$	$600 \pm 216$		
Cocaine <sup>c</sup>	$258 \pm 23$	$343 \pm 31$	$1740 \pm 180$		
Cocained	$573 \pm 54$	$402 \pm 62$	$2040 \pm 240$		
Cocaine <sup>e</sup>	$350 \pm 45$	$260 \pm 9.4$	$1610 \pm 300$		
Cocainef	$271 \pm 65$	$217 \pm 23$	$1730 \pm 280$		
Cocaine <sup>g</sup>	$621 \pm 45$	$496 \pm 37$	$1400 \pm 320$		

<sup>a</sup>Results are average  $\pm$  SEM of three independent experiments assayed in triplicate.

<sup>b</sup>Cocaine as reference for GBR 12909.

<sup>c</sup>Cocaine as reference for 9, 10, 14, 16, 22.

<sup>d</sup>Cocaine as reference for **11**, **24**.

<sup>e</sup>Cocaine as reference for 12, 17, 19.

<sup>f</sup>Cocaine as reference for **13–15**.

<sup>g</sup>Cocaine as reference for **18**, **20**, **21**, **23**.

Table 2.	DA,	5-HT a	und NE	reuptake	inhibitio	n and	ratios	of reupta	ıke in	hibitior	ı to	binding	of (	bisaryl	methox	y)buty	lpiperi	dine/p	oiperazi	ne dei	ci-
vatives at	the D	A tran	sporters																		

Compound	Rei	Discrimination ratios		
	[ <sup>3</sup> H]DA	[ <sup>3</sup> H]5-HT	[ <sup>3</sup> H]NE	[ <sup>3</sup> H]DA reuptake inhibition/DAT binding
9	$101 \pm 11^{a}$	$314 \pm 71^{a}$	$143\pm50^{\mathrm{a}}$	0.9
10	$114 \pm 28$	$860 \pm 180$	$310 \pm 110$	1.3
11	$56 \pm 16$	$203 \pm 78$	$158 \pm 60$	0.6
12	$415 \pm 87$	$2030 \pm 190$	$393 \pm 30$	3.1
13	$1030 \pm 330$	$>10 \ \mu M$	$2250 \pm 950$	47.5
14	$96 \pm 11$	$136 \pm 41$	$179 \pm 47$	15.0
15	$431 \pm 88$	$3070 \pm 830$	$1870 \pm 670$	110.5
16	$7080 \pm 510$	$>10 \ \mu M$	$> 10 \ \mu M$	37.1
17	$1000 \pm 160$	$3800 \pm 1300$	$780 \pm 310$	63.3
18	$184 \pm 47$	$570 \pm 110$	$30.7 \pm 8.3$	4.5
19	$1750 \pm 730$	$2100 \pm 400$	$1410 \pm 300$	26.5
20	$440 \pm 120$	$1700 \pm 390$	$159 \pm 21$	6.0
21	$3100 \pm 1400$	$3070 \pm 740$	$> 10 \ \mu M$	8.3
22	$426 \pm 54$	$570 \pm 210$	$132 \pm 11$	323
23	$3490 \pm 220$	$111 \pm 31$	$64 \pm 19$	27.9
24	$> 10 \ \mu M$	$361 \pm 68$	$> 10 \ \mu M$	1
GBR 12909	$246 \pm 142$	$584 \pm 237$	$532 \pm 183$	9.1
Cocaine <sup>b</sup>	$152 \pm 24$	$425 \pm 114$	$445 \pm 205$	
Cocaine <sup>c</sup>	$276 \pm 22$	$301 \pm 53$	$264 \pm 57$	
Cocained	$237 \pm 41$	$348 \pm 66$	$190 \pm 38$	
Cocainee	$417 \pm 35$	$405 \pm 39$	$239 \pm 26$	
Cocaine <sup>f</sup>	$278 \pm 53$	$189 \pm 31$	$209\pm36$	

<sup>a</sup>Results are average  $\pm$  SEM of three independent experiments assayed in triplicate.

<sup>b</sup>Cocaine as reference for GBR 12909.

<sup>c</sup>Cocaine as reference for 9, 10, 14, 16, 22.

<sup>d</sup>Cocaine as reference for **11**, **24**.

<sup>e</sup>Cocaine as reference for 12, 17, 19.

<sup>f</sup>Cocaine as reference for 13–15.

Table 3.	Receptor	binding profile	and effects of selec	ted target comp	ounds 14–17, 22, and 23
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Receptor		Affinity $(K_i \pm \text{SD [nM]})^a$ Antagonist activity (IC <sub>50</sub> [nM]) <sup>b</sup>								
	14	15	16	17	22	23				
$D_1$	$2460 \pm 38.1^{\circ}$	$3050 \pm 1440$	$172 \pm 10.3$	$1570 \pm 523$	$1640 \pm 527$	$1890 \pm 763$				
$\dot{D_2}$	$22.6 \pm 0.5^{a}$	$27.6 \pm 0.8$	$51.5 \pm 14.6$	$37.4 \pm 5.0$	$17.0 \pm 6.17$	$18.8 \pm 4.32$				
-	$136 \pm 1.3^{b}$	$90.6 \pm 40.3$	$42.3 \pm 13.6$	$70.1 \pm 13.0$	$48.7 \pm 2.67$	$67.8 \pm 1.22$				
$D_3$	$239 \pm 121^{a}$	$56.8 \pm 34.1$	$143 \pm 5.1$	$65.2 \pm 20.9$	$15.8 \pm 1.08$	$33.0 \pm 13.8$				
2	$52.0 \pm 4.8^{b}$	$91.3 \pm 28.8$	$174 \pm 50.2$	$85.9 \pm 30.6$	$87.4 \pm 17.0$	$77.3 \pm 6.26$				
5-HT14	$1400 \pm 247$	$2870 \pm 693$	$>10 \ \mu M$	$1130 \pm 37.5$	$857 \pm 123$	$682 \pm 77.7$				
5-HT <sub>2A</sub>	$566 \pm 45.5$	$580 \pm 44.4$	$639 \pm 107$	$621 \pm 83.3$	$580 \pm 58.1$	$241 \pm 64.7$				
5-HT <sub>2C</sub>	$5870 \pm 4140$	$5790 \pm 419$	$4300 \pm 411$	$6390 \pm 1760$	$3100 \pm 235$	$>10 \ \mu M$				
5-HT <sub>3</sub>	$2470 \pm 46.6$	$5270 \pm 594$	nt <sup>d</sup>	nt	nt	nt				

<sup>a</sup>Binding assay receptors/radioligands: D<sub>1</sub> (human cloned receptors in LHD<sub>1</sub> cells/[<sup>3</sup>H]SCH 23,390); D<sub>2</sub> and D<sub>3</sub> (human cloned receptors in CHO<sub>p</sub> cells/[<sup>3</sup>H]YM-09151–2); 5-HT<sub>1A</sub> (human cloned receptors in HA7 cells/[<sup>3</sup>H]8-OH-DPAT); 5-HT<sub>2A</sub> (rat receptors in NIH-3T3-GF6 cells/[<sup>3</sup>H]ketan-serin); 5-HT<sub>2C</sub> (rat receptors in NIH-3T3-P $\phi$  cells/[<sup>3</sup>H]mesulergine); 5-HT<sub>3</sub> (rat/mouse hybrid receptors in NG108–15 cells/[<sup>3</sup>H]GR65630).

 $^{b}D_{2}$ ,  $D_{3}$  antagonist assays: CHO<sub>p</sub> cells (human receptor), [<sup>3</sup>H]thymidine incorporation (inhibition of mitogenesis), quinpirole as internal standard (ED<sub>50</sub> range 6.5–57 nM for D<sub>2</sub> receptor, ED<sub>50</sub> range 2.8–25 nM for D<sub>3</sub> receptor).

<sup>c</sup>All values represent the mean of at least two determinations.

<sup>d</sup>nt = Not tested.

Thus, the 4-hydroxy-4-phenylpiperidine moiety provides additional binding to the DAT. Analogues 14-16 resemble the antipsychotic bromperidol and were found to be moderate affinity DA antagonists. Addition of aryl and hydroxyl groups at the 4-position of the piperidine ring provided a desirable combination of DAT and DA receptor activity. The 4-aryl-4-piperidinol analogues 14 and 15 demonstrated very weak binding at the  $D_1$  and 5-HT receptor sites. Analogues 17, 22, and 23, each containing a partial structure of a DA antagonist (17, trifluperidol; 22, pimozide; 23, spiperone), also showed moderate activity at the  $D_2/D_3$  receptor sites. Most compounds had selective and weak activity at the 5-HT<sub>2A</sub> receptor ( $K_i = 241-639$  nM). As expected, the bulky bisaryl group was not well tolerated in terms of DA activities.

### In vivo locomotor activity

Based on their in vitro activities at the DAT, dopamine  $D_2/D_3$  receptors and DR values, compounds 15–17 (high DR) and 9, 18, 21 (low DR) were selected for preliminary behavioral screening which involved testing alone and in combination with cocaine for their effect on locomotor activity (LMA) in mice. Likewise, the effects of the dopaminergic antagonists (14–17, and 23) on cocaine-induced hyperactivity were studied. Compound 22 was not studied even though it showed a high DR value at the DAT and SERT because NE reuptake inhibition was strong.<sup>31</sup> As summarized in Table 4, compounds 9 and 15 produced a significant increase in spontaneous LMA with low ED<sub>50</sub> compared with GBR 12909. Significant suppression of spontaneous LMA was demonstrated by all the compounds except 9 and 15. Compound 15 produced a biphasic effect on LMA; activity was increased with low doses and decreased with high doses. Biphasic effects on spontaneous LMA by dopamine  $D_2/D_3$  dopaminergic antagonistic ligands have been previously demonstrated with other compounds.<sup>32</sup> Since this ligand possesses activities at the DAT and is a moderate dopamine  $D_2/D_3$  antagonist, its effects on LMA are probably the result of activities at

both DAT and DA receptor sites. Maximal stimulatory effect on spontaneous LMA by 9 and 15 is only 40-80% of that achieved with cocaine as shown by the ratio of ME/CME in Table 4. Attenuation of cocaine-induced LMA (20 mg/kg cocaine) was demonstrated by most of the compounds. Only compounds 17 and 21 failed to attenuate the hyperlocomotion induced by cocaine. Compound 24 showed very weak receptor activities (2.4  $\mu M$  for  $D_2,$  1.8  $\mu M$  for  $D_3,$  data not shown) and weak 5-HT reuptake inhibition (361 nM, Table 2). No activity at the DAT and NET sites was identified; however, hyper LMA caused by cocaine was attenuated. Correlations between DR values and ED<sub>50</sub> or AD<sub>50</sub> were not found, although other factors such as potency as  $D_2/D_3$  antagonists and pharmacokinetics (uptake rate into brain) might be critical for the potency in LMA studies.

Table 4. Effect of compounds 9, 13, 15–18, 21, 23 and 24 on loco-motor activity in mouse

Compound	$\begin{array}{c} ED_{50} \; (mg/kg)^{a} \\ ID_{50} \; (mg/kg)^{b} \end{array}$	ME/CME <sup>c</sup>	$\begin{array}{c} AD_{50} \\ (mg/kg)^d \end{array}$
9	6.6/S <sup>a</sup>	0.78	77.6
13	170/I <sup>b</sup>		109
15	$2.42/S^{a,e}$	0.38	15.8
16	14.1/I <sup>b</sup>		13.8
17	95.7/I <sup>b</sup>		No attenuation
18	50.7/I <sup>b</sup>		16.4
21	132/I <sup>b</sup>		No attenuation
23	49.2/I <sup>b</sup>		45.7
24	82.8/I <sup>b</sup>		40.1
GBR 12909	$6.24/S^{a}$	1.22	<b>Biphasic</b> <sup>f</sup>

 $^{a}\text{ED}_{50}\text{=}\text{dose}$  producing 1/2 maximal stimulant activity, S=stimulation.

 $^{b}ID_{50}$  = dose-producing 1/2 maximal depressant activity, where maximal depression = 0 counts/30 min, I = inhibition.

°Maximal effect (ME)/cocaine maximal effect (CME)

 ${}^d\mathrm{AD}_{50}\!=\!\mathrm{dose}$  attenuating cocaine-induced stimulation by 50%, ligand–cocaine interaction study.

<sup>e</sup>Ligand study, see text for details; in general low doses produce stimulation and high doses inhibition.

<sup>f</sup>10 mg/kg enhanced and 50 mg/kg depressed 30-60 min postinjection.

It must be noted that  $ID_{50}$  is similar to  $ED_{50}$  (LMA increase or decrease) and a high  $ID_{50}$  value means low intrinsic activities of LMA inhibition for the compound alone, whether as a weak DA receptor antagonist or with other unknown activities. But if a compound also has a low  $AD_{50}$ , this means a strong counter action against cocaine at low dosage. Such activities can be explained with other in vitro activities and with other pharmacophysical properties of the ligand. High  $ID_{50}$  and low  $AD_{50}$  values (discrimination and self-administration) are currently important criteria for the selection of ligands for in vivo testing by NIDA.

Biphasic effects on spontaneous LMA by  $D_2/D_3$  dopaminergic antagonistic ligands have been previously demonstrated with other compounds. Activities at the  $D_2/D_3$  autoreceptors might explain some of possible underlying mechanisms of biphasic activities. Since these ligands possess activities at the DAT and are moderate inhibitors of DA reuptake, their effects on LMA are probably the result of activities at both DAT and DA receptor sites. The biphasic activity as stimulant or inhibitant cannot be explained with only in vitro receptor activities and DAT activities because the other sites also can modulate dopaminergic activities. However, the time course LMA studies will clearly demonstrate whether such compounds have beneficial long-term effects or not which also can explain pharmacokinetic factors.

### Conclusions

The introduction of hydroxyl and/or aromatic substituents to the piperidine ring of template structure 4 produced retention of transporter affinity and DA reuptake inhibition. Such modification resulted in DA receptor affinity and selectivity when compared with 5-HT receptors. These data give insight into possible structural similarities between ligand-binding sites on the DAT and DA receptors. The current studies have provided additional information on pharmacophores that are responsible for selective binding and potency of reuptake inhibition at the DAT and on activities at the DA/5-HT receptor sites. Although correlation between in vitro DR values and in vivo potency in LMA activities was not observed, these new analogues inconjuction with the results of the LMA studies may provide a basis for the design of other ligands with strong binding to the DAT, low potency inhibition of DA reuptake, and binding to DA/5-HT receptor sites.

#### Experimental

Melting points were determined with a Thomas-Hoover melting point apparatus and are uncorrected. Elemental analyses, performed by Atlantic Microlabs, Atlanta, GA, were within 0.4% of calculated values. <sup>1</sup>H NMR spectra were recorded on a Varian XL-500 spectrometer. In general, CDCl<sub>3</sub> was used for the free bases and DMSO- $d_6$  for salts. Thin layer chromatography (TLC) was performed on 250-µm-thick silica gel plates or alumina precoated plates (Whatman, AL SIL G/UV or J.T. Baker, Baker-flex, SILICA GEL IB-F) containing fluorescent indicator ( $2 \times 8$  cm). Column chromatography was performed on silica gel (Baker, 40 µm Flash chromatography). Fractions were analyzed using TLC and compounds were visualized with ninhydrin (0.5 g/ 100 mL methanol) or iodine vapor for primary and secondary amine(s). Free bases were dissolved in ethyl acetate and/or diethyl ether, filtered, and precipitated by addition of a solution of oxalic acid. The resulting solids were collected by filtration and recrystallized.

# Synthetic procedures

**1-[bis(4-Fluorophenyl)methoxy]-4-chlorobutane (6).** A mixture of 4-chloro-1-butanol (6.51 g, 60 mmol), 1 mL concentrated sulfuric acid, and 4,4'-difluorobenzhydrol (2.2 g, 10 mmol) in 50 mL toluene was heated at reflux for 12 h. The reaction mixture was cooled, washed successively with saturated sodium bicarbonate solution (50 mL) and water (50 mL). The organic layer was dried over magnesium sulfate, filtered, and the solvent was removed by rotary evaporation. The resultant brown oil was purified on a silica gel column. Elution with 2% ethyl acetate–hexane afforded 2.33 g (7.5 mmol, 75%) of the product as a colorless oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.71–1.80 (2H, m), 1.84–1.93 (2H, m), 3.43 (2H, t, J=6.0), 3.52 (2H, t, J=6.4), 5.30 (1H, s) 6.95–7.02 (4H, m), 7.24–7.29 (4H, m).

**1-[bis(4-Fluorophenyl)methoxy]-2-bromoethane (7).** As described for **6**, **7** was prepared from 2-bromo-1-ethanol and 4,4'-difluorobenzhydrol.

*N*-(4-Bromobutyl)-phthalimide (8). This compound was purchased from Sigma Chemical Company.

1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-hydroxypiperidine (9). 4-Hydroxypiperidine (1.2 g, 5 mmol) was dissolved in 20 mL anhydrous DMF and stirred with K<sub>2</sub>CO<sub>3</sub> powder (2.76 g, 20 mmol) and KI (100 mg, 0.6 mmol) for 0.5 h. To this turbid solution, 6 (1.55 g, 5 mmol) in 10 mL DMF was added dropwise. The reaction mixture was stirred for 72 h at 60-70 °C. The turbid reaction mixture was poured into 200 mL ethyl acetate, washed with saturated sodium chloride solution (60  $mL \times 3$ ), dried over magnesium sulfate, and evaporated to dryness. The crude oil was applied to a silica gel column for purification. Elution with CHCl<sub>3</sub>/MeOH (98:2) afforded the desired alkylated product 9 as a colorless oil (87% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 1.54–1.66 (6H, m), 1.86-1.89 (2H, m), 2.09 (2H, t, J=10.0), 2.31 (3H, t, J = 7.5 overlap with br s), 2.74–2.76 (2H, m), 3.42 (2H, t, J = 6.3), 3.66 (1H, heptet, J = 4.5), 5.28 (1H, s), 6.98– 7.02 (4H, m), 7.25–7.29 (4H, m); mp (oxalate salt): 117– 120°C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-3-hydroxypiperidine (10).** As described for **9**, **10** was prepared from **6** and 3-hydroxypiperidine as a colorless oil (87% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.48–1.67 (7H, m), 1.76–1.79 (1H, m), 2.26 (1H, br s), 2.33 (2H, t, *J*=7.3), 2.41–2.45 (4H, m), 3.42 (2H, t, *J*=6.3), 3.80 (1H, br s), 5.28 (1H, s), 6.99–7.03 (4H, m), 7.26–7.29 (4H, m); mp (oxalate salt): 106–109 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-3-piperidine carboxamide (11).** As described for compound **9**, **11** was prepared from **6** and nipecotamide (Sigma Chemical Company) as a colorless oil (78% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.48–1.67 (7H, m), 1.76–1.79 (1H, m), 2.26 (1H, br s), 2.33 (2H, t, *J* = 7.3), 2.41–2.45 (4H, m), 3.42 (2H, t, *J* = 6.3), 3.80 (1H, br s), 5.28 (1H, s), 6.99–7.03 (4H, m), 7.26–7.29 (4H, m); mp (oxalate salt): 143–146 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-phenylpiperidine-4-carbonitrile (12).** As described for compound **9**, **12** was prepared from **6** and 4-cyano-4-phenylpiperidine hydrochloride as a colorless oil (83% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.60–1.70 (4H, m), 2.05–2.12 (4H, m), 2.42–2.48 (4H, m), 2.99 (2H, br d, J=12.5), 3.44 (2H, t, J=6.5), 5.29 (1H, s), 6.96–7.00 (4H, m), 7.27–7.30 (5H, m), 7.35–7.38 (2H, m), 7.48–7.50 (2H, m); mp (oxalate salt): 176–178 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-acetyl-4-phenylpiperidine (13).** As described for **9**, **13** was prepared from **6** and 4-acetyl-4-phenylpiperidine hydrochloride as a colorless oil (73% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.56–1.68 (4H, m), 1.90 (3H, s), 2.06 (2H, t d, *J*=11.8, 2.7), 2.21 (2H, br t, *J*=10.8), 2.30 (2H, t, *J*=7.1), 2.46 (2H, br d, *J*=11.7), 2.71 (2H, br d, *J*=10.5), 3.42 (2H, t, *J*=5.9), 5.28 (1H, s), 6.96–7.02 (4H, m), 7.25–7.37 (9H, m); mp (oxalate salt): 138–140 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-phenylpiperidin-4-ol (14).** As described for **9**, **14** was prepared from **6** and 4-phenyl-4-piperidinol as a colorless oil (79% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.58–1.73 (6H, m), 2.13 (2H, t d, *J*=13.0, 4.5), 2.37 (2H, t, *J*=7.2), 2.43 (2H, br d, *J*=11.8), 2.76 (2H, br d, *J*=11.1), 3.42 (2H, t, *J*=5.6), 5.28 (1H, s), 6.96–7.03 (4H, m), 7.22–7.34 (7H, m), 7.49 (2H, d, *J*=7.9); mp (oxalate salt): 168–170 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-(4-bromophenyl)-4-piperidinol (15).** As described for **9**, **15** was prepared from **6** and 4-(4-bromophenyl)-4-piperidinol as a colorless oil (80% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.60–1.73 (6H, m), 2.12 (2H, t d, J=12.8, 2.9), 2.39–2.45 (4H, m), 2.80 (2H, br d, J=10.9), 3.43 (2H, t, J=5.4), 5.29 (1H, s), 6.97–7.03 (4H, m), 7.26–7.34 (4H, m), 7.37 (2H, d, J=8.7), 7.46 (2H, d, J=8.4); mp (oxalate salt): 156–158 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]ethyl]-4-(4-bromophenyl)-4-piperidinol (16).** As described for **9**, **16** was prepared from **7** and 4-(4-bromophenyl)-4-piperidinol as a color-less oil (76% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.67 (2H, br d, J = 12.0), 2.07 (2H, t d, J = 12.1, 4.0), 2.14 (1H, br s), 2.52 (2H, t d, J = 11.8, 2.5), 2.71 (2H, t, J = 5.8), 2.78 (2H, br d, J = 12.0), 3.57 (2H, t, J = 6.0), 5.32 (1H, s), 6.97–7.02 (4H, m), 7.26–7.29 (4H, m), 7.35 (2H, d, J = 8.5), 7.44 (2H, d, J = 8.5); mp (oxalate salt): 148–151 °C.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-(3-trifluoromethylphenyl)-4-piperidinol (17).** As described for **9**, **17** was prepared from **6** and 4-[3-(trifluoromethyl)phenyl]- 4-piperidinol as a colorless oil (81% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.56–1.67 (4H, m), 1.71 (2H, d, *J*=12.0), 2.13 (2H, t t, *J*=13.1, 4.0), 2.36–2.45 (4H, m), 2.79 (2H, br d, *J*=11.0), 3.42 (2H, t, *J*=6.3), 5.29 (1H, s), 6.97–7.01 (4H, m), 7.26–7.30 (4H, m), 7.42 (1H, t, *J*=7.8), 7.48 (1H, d, *J*=7.0), 7.67 (1H, d, *J*=8.0), 7.83 (1H, s); mp (oxalate salt): 148–150 °C.

**4-Benzyl-1-[4-[bis(4-fluorophenyl)methoxy]butyl]-4-piperidinol (18).** As described for **9**, **18** was prepared from **6** and 4-benzyl-4-hydroxypiperidine as a colorless oil (73% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.21 (1H, br s), 1.51 (2H, br d, J=12.5), 1.56–1.66 (4H, m), 1.74 (2H, t d, J=12.9, 3.8), 2.26 (2H, br t, J=11.0), 2.35 (2H, t, J=7.3), 2.65 (2H, br d, J=11.5), 2.75 (2H, s), 3.42 (2H, t, J=5.8), 5.28 (1H, s), 6.97–7.02 (4H, m), 7.19–7.21 (2H, m), 7.23–7.32 (7H, m); mp (oxalate salt): 136–138 °C.

**2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-1,2,3,4-tetrahydroisoquinoline (19).** As described for **9**, **19** was prepared from **8** and 1,2,3,4-tetrahydroisoquinoline as a slightly yellow oil (79% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.68–1.71 (4H, m), 2.50 (2H, br t, *J*=7.0), 2.69 (2H, t, *J*=5.8), 2.88 (2H, t, *J*=5.8), 3.45 (2H, br t, *J*=5.5), 3.60 (2H, s), 5.28 (1H, s), 6.97–7.00 (5H, m), 7.06–7.11 (3H, m), 7.26–7.28 (4H, m); mp (oxalate salt): 135–137 °C.

**2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline (20).** As described for **9**, **20** was prepared from **8** and 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline hydrochloride as a colorless oil (81% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.69–1.71 (4H, m), 2.50 (2H, br t, *J* = 6.8), 2.69 (2H, t, *J* = 5.8), 2.81 (2H, t, *J* = 5.8), 3.45 (2H, t, *J* = 5.8), 3.53 (2H, s), 3.83 (3H, s), 3.84 (3H, s), 5.30 (1H, s), 6.50 (1H, s), 6.59 (1H, s), 6.98–7.02 (4H, m), 7.26–7.30 (4H, m); mp (oxalate salt): 88–102 °C.

**2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-6,7-dimethoxy-1-phenyl-1,2,3,4-tetrahydroisoquinoline (21).** As described for **9, 21** was prepared from **6** and 6,7-dimethoxy-1phenyl-1,2,3,4-tetrahydroisoquinoline hydrochloride as a colorless oil (75% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.48–1.63 (4H, m), 2.28–2.33 (1H, m), 2.49–2.56 (2H, m), 2.75 (1H, d t, *J* = 16.5, 4.7), 2.93–2.99 (1H, m), 3.14 (1H, d t, *J* = 12.0, 5.3), 3.27–3.34 (2H, m), 3.58 (3H, s), 3.84 (3H, s), 4.46 (1H, s), 5.20 (1H, s), 6.16 (1H, s), 6.60 (1H, s), 6.96–7.00 (4H, m), 7.20–7.27 (9H, m); mp (oxalate salt): 115–118 °C.

**1-[1-[4-[bis(4-Fluorophenyl)methoxy]butyl]piperidin-4-yl]-1,3-dihydrobenzoimidazol-2-one (22).** As described for **9, 22** was prepared from **6** and 4-(2-keto-1-benzimidazolinyl)piperidine as a colorless oil (80% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.64–1.70 (4H, m), 1.82 (2H, br d, *J*=10.0), 2.13 (2H, t, *J*=11.5), 2.42 (2H, t, *J*=7.0), 2.50 (2H, q d, *J*=12.7, 3.4), 3.10 (2H, br d, *J*=11.0), 3.45 (2H, t, *J*=5.8), 4.40 (1H, t t, *J*=12.3, 4.0), 5.30 (1H, s), 6.98– 7.04 (6H, m), 7.12–7.13 (1H, m), 7.53–7.30 (5H, m), 11.09 (1H, br s); mp (oxalate salt): 225–227 °C.

**8-[4-[bis(4-Fluorophenyl)methoxy]butyl]-1-phenyl-1,3,8triazaspiro-[4,5]decan-4-one (23).** As described for **9**, **23** was prepared from **6** and 1-phenyl-1,3,8-triazaspiro-[4,5]decan-4-one as a colorless oil (79% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.59–1.70 (4H, m), 1.72 (2H, br d, J=13.5), 2.43 (2H, t, J=7.5), 2.66 (2H, t d, J=12.7, 5.5), 2.75– 2.84 (4H, m), 3.44 (2H, t, J=6.3), 4.72 (2H, s), 5.29 (1H, s), 6.85 (1H, t, J=7.3), 6.91 (2H, d, J=8.0), 6.97–7.02 (4H, m), 7.25–7.30 (6H, m), 8.08 (1H, s); mp (oxalate salt): 106–110 °C.

**2-[4-[4-Bromophenyl]-4-hydroxypiperidin-1-yl]-butyl]**isoindole-1,3-dione (24). As described for 9, 24 was prepared from 8 and 4-(4-bromophenyl)-4-piperidinol as a white solid (80% yield). <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  1.47 (2H, p, J=7.3), 1.55 (2H, br d, J=12.5), 1.64 (2H, p, J=7.3), 1.85 (2H, t d, J=12.8, 3.8), 2.31–2.36 (4H, m), 2.61 (2H, br d, J=10.5), 3.42 (1H, br s), 3.61 (2H, t, J=7.0), 7.42 (2H, d, J=9.0), 7.48 (2H, d, J=9.0), 7.83– 7.88 (4H, m); mp (oxalate salt): 223–225 °C.

### **Biologic methods**

**In vitro binding and reuptake inhibition assays.** In vitro assays for [<sup>125</sup>I]RTI-55-binding inhibition and [<sup>3</sup>H]neuro-transmitter-uptake inhibition using HEK 293 cells expressing recombinant biogenic amine transporters were carried out according to the procedures described previously.<sup>22,28</sup>

In vitro receptor binding and functional biochemical assays.<sup>29</sup> Binding assay for the  $D_1$  receptor LHD<sub>1</sub> cells (human receptor) were grown to confluence on 100×20 mm plates in DMEM containing 10% fetal calf serum, 0.05% penicillin-streptomycin, and 400 µg/mL G418 sulfate. The cells were scraped from the plates and centrifuged at 500g for 5 min. The pellet was homogenized in 50 mM Tris-HCl (pH 7.7) by polytron, centrifuged at 27,000g, and resuspended at 10 mg protein/mL in the same buffer. The homogenate was then stored at -70 °C in 1-mL aliquots. When needed, the thawed cells were washed once and resuspended at 5 mg protein/80 mL 50 mM Tris-HCl, containing 120 mM NaCl, 5 mM KCl, 2 mM CaCl<sub>2</sub>, and 1 mM MgCl<sub>2</sub> (pH 7.4). The assay was performed in triplicate in 96-well plates. To wells containing 100 µL test drug or buffer and 100 µL <sup>3</sup>H]SCH 23,390 (0.18 nM final concentration), 0.8 mL cell homogenate (0.05 mg protein/well) was added, and the plates were incubated at 25 °C for 60 min. SCH 23,390 at 1 µM was used to determine nonspecific binding.

Binding assay for the  $D_2$  and  $D_3$  receptors. CHOp cells (human receptors) were grown to confluence on  $100 \times 20$ mm plates in  $\alpha$  minimum essential medium ( $\alpha$ MEM), containing 10% fetal calf serum, 0.05% penicillin– streptomycin, and 600 µg/mL G418 sulfate. The cells were scraped from the plates and centrifuged at 500g for 5 min. The pellet was homogenized by polytron in 50 mM Tris–HCl (pH 7.7) and centrifuged at 27,000g for 12 min. The pellet was resuspended in 50 mM Tris to contain  $D_2$  at 5 mg protein/mL or  $D_3$  at 1 mg protein/ mL and stored at -70 °C in 1-mL aliquots. On the day of the experiment, CHOp- $D_2$  or CHOp- $D_3$  cells were thawed, resuspended in 50 mM Tris, and centrifuged at 27,000g for 12 min. The pellet was resuspended at 5 mg protein/80 mL for  $D_2$  and 1 mg protein/80 mL for  $D_3$  in 50 mM Tris, containing 120 mM NaCl, 5 mM KCl, 1.5 mM CaCl<sub>2</sub>, 4 mM MgCl<sub>2</sub>, and 1 mM EDTA (pH 7.4). Then 0.8 mL cell homogenate (0.05 and 0.01 mg protein/ well for D<sub>2</sub> and D<sub>3</sub>, respectively) was added to wells containing 100  $\mu$ L test drug or buffer and 100  $\mu$ L [<sup>3</sup>H]YM-09151–2 (0.21 nM final concentration). Chlorpromazine at 1  $\mu$ M was used to determine nonspecific binding. The plates were incubated at 25 °C for 60 min. The incubation was terminated by rapid filtration through Whatman GF/B filter paper soaked in 0.1% polyethylenimine (PEI) on a Brandel cell harvester. The filters were washed three times with ice-cold 50 mM Tris–HCl (pH 7.7) and soaked overnight in scintillation cocktail before counting for 2 min on a Beckman LS 6000 scintillation counter.

Binding assay for the 5-HT<sub>1A</sub> receptor. HA7 cells (human receptor) were grown and harvested as above. The pellet was homogenized in 50 mM Tris-HCl (pH 7.7) by polytron, centrifuged at 27,000g, and resuspended at 10 mg protein/mL in the same buffer. The homogenate was then stored at -70 °C in 1-mL aliquots. When needed, the thawed cells were washed once and resuspended at 10 mg protein/80 mL in 25 mM Tris-HCl, containing 100 µM ascorbic acid and 10 µM nialamide (pH 7.4). The assay was performed in triplicate in 96-well plates. To each well, 100 µL [<sup>3</sup>H]8-OH-DPAT (0.5 nM final concentration), 100 µL test compound or buffer, and 0.8 mL cell homogenate (0.1 mg protein/well) were added by a Tomtec Quadra 96. Dihydroergotamine at 1 µM was used to determine nonspecific binding. The plates were incubated at 25 °C for 60 min. The incubation was terminated by rapid filtration through glass-fiber filter paper on a Tomtec cell harvester. The filters were washed four times with ice-cold 50 mM Tris-HCl (pH 7.7), dried overnight, and mixed with 10 mL scintillation cocktail before counting for 2 min on a Wallac Betaplate 1205 liquid scintillation counter.

Binding assay for the 5-HT<sub>2A</sub> receptor. NIH-3T3-GF6 cells (rat receptor) were grown as above. On the day of the experiment, the cells were thawed, resuspended in 50 mM Tris–HCl, and centrifuged at 27,000g for 12 min. The pellet was resuspended at 1 mg protein/80 mL 25 mM Tris–HCl (pH 7.7) and 0.8 mL cell homogenate (0.01 mg protein/well) was added to wells containing 100  $\mu$ L test drug or buffer and 100  $\mu$ L [<sup>3</sup>H]ketanserin (0.40 nM final concentration). Ketanserin at 1  $\mu$ M was used to determine nonspecific binding. The plates were incubated at 25°C for 60 min and processed as above for the 5-HT<sub>1A</sub> receptor.

Binding assay for the 5-HT<sub>2C</sub> receptor. NIH-3T3-P $\phi$  cells (rat receptor) were grown as above. The final pellet was resuspended at 3 mg protein/80 mL 50 mM Tris–HCl, containing 4 mM CaCl<sub>2</sub>, 10  $\mu$ M pargyline, and 0.1% ascorbic acid (pH 7.7). Wells with 100  $\mu$ L test drug or buffer, 100  $\mu$ L [<sup>3</sup>H]mesulergine (0.4 nM final concentration), and 0.8 mL cell homogenate (0.03 mg protein/well) were incubated at 25 °C for 60 min. Mesulergine at 10  $\mu$ M was used to determine non-specific binding. After incubation, the plates were processed as above for the 5-HT<sub>1A</sub> receptor.

Binding assay for the 5-HT<sub>3</sub> receptor. NG108–15 cells (rat/mouse hybrid) were grown to confluence on  $100 \times 20$  mm plates in DMEM with HAT supplement and 10% fetal calf serum. The cells were washed from the plates, centrifuged, homogenized as described above, and stored at -70 °C in aliquots of 15 plates/4 mL. When needed, the thawed cells were washed once and resuspended at 15 plates/20 mL 25 mM Tris-HCl (pH 7.7). The assay was performed by adding 50  $\mu$ L test drug or buffer, 50 µL [3H]GR65630 (1.6 nM final concentration), and 0.4 mL cell homogenate (0.13 mg protein/ tube) to each tube. The tubes were then incubated at 25 °C for 45 min. Zacopride at 1 µM was used to determine nonspecific binding. The incubation was terminated by rapid filtration through Whatman GF/B filter paper soaked in 0.1% polyethylenimine (PEI) on a Brandel cell harvester. The filters were washed three times with ice-cold 50 mM Tris-HCl (pH 7.7) and soaked overnight in scintillation cocktail before counting for 2 min on a Beckman LS 6000 scintillation counter.

 $D_2$  and  $D_3$  functional mitogenesis assay. To measure  $D_2$ and D<sub>3</sub> inhibition of quinpirole stimulation of mitogenesis (antagonist assay), CHOp cells (human receptor) were seeded in 96-well plates at 5000 cells/well. The cells were incubated at 37 °C in αMEM with 10% FBS, 0.05% penicillin-streptomycin, and 200 µg/mL G418 sulfate. After 48 h, the wells were rinsed twice with 100  $\mu$ L serum-free  $\alpha$ -MEM and incubated for 24 h at 37 °C in serum-free  $\alpha$ -MEM. The medium was then removed and replaced with 90  $\mu$ L serum-free  $\alpha$ -MEM and 10  $\mu$ L drug in sterile water plus quinpirole. After another 24 h incubation at 37 °C, 0.25  $\mu$ Ci [<sup>3</sup>H]thymidine was added to each well. The cells were incubated for 2 h at 37 °C. Then, 10  $\mu$ L/well 10X trypsin (trypsin–EDTA solution: 5 g trypsin in 20 mL) was added to release the cells, and the plates were filtered using a Tomtec cell harvester. The filters were washed four times with deionized water, dried overnight, and mixed with 10 mL scintillation cocktail before counting for 2 min on a Wallac Betaplate 1205 liquid scintillation counter. Quinpirole was run on every plate as an internal standard.

# Locomotor activity<sup>22</sup>

This study was conducted using 16 Digiscan locomotor activity testing chambers ( $40.5 \times 40.5 \times 30.5$  cm). Panels of infrared beams (16 beams) and corresponding photodetectors were located in the horizontal direction along the sides of each activity chamber. Separate groups of eight nonhabituated male Swiss-Webster mice were injected via the intraperitoneal route with either vehicle (methylcellulose, saline, or distilled water) or compound (3, 10, 30, or 100 mg/kg), 20 min prior to locomotor activity testing. Just prior to placement in the apparatus, all mice received a saline injection intraperitoneally. In all studies, horizontal activity (interruption of 1 photocell beam) was measured for 1 h within 10-min periods. Testing was conducted with one mouse per activity chamber. For the cocaine/compound interaction study, 20 min following administration of vehicle or compound intraperitoneally (3, 10, 30, or 100 mg/ kg), groups of eight nonhabituated male Swiss-Webster mice were injected with either 0.9% saline or 20 mg/kg cocaine intraperitoneally and placed in the Digiscan apparatus for a 1-h session.

Maximal effects of cocaine and stimulant test compound (independent studies of cocaine and test compound). One 30-min time period in which maximal effects of cocaine/ test compound were evident at most doses was selected. The mean  $(\pm SEM)$  maximal stimulant activity (total counts in the 30-min period divided by 3) was plotted versus dose. A log<sub>10</sub> transformation of the 30-min period average counts for individual subjects was performed to homogenize variances for subsequent analyses. ANOVA was used and each dose of cocaine/test compound (specified a priori) was contrasted to saline (the vehicle) to determine significant (P < 0.05) dose effects. The 30-min period average counts across subjects was fit to a function of  $\log_{10}$  dose using least-squares curve-fitting analyses (i.e., TableCurve software from Jandel). The maximum effect from the resultant dose-response curve was estimated. A test compound maximal effect/cocaine maximum effect (ME/CME) ratio was calculated and the  $ED_{50}$  (dose that produces 1/2 maximal stimulant activity) was determined from a linear regression analysis of the ascending portion (up to the dose that produced a maximal effect) of the curve. The mean maximal effect for cocaine was calculated for each mouse.

Maximal effects of depressant test compound (studies of test compound alone). A 30-min time period was selected in which cocaine (20 mg/kg) produced its maximal effects as determined from the studies with cocaine alone. The mean  $(\pm SEM)$  activity (total counts in the 30-min period divided by 3) was plotted versus dose. A log<sub>10</sub> transformation of the 30-min period average counts for individual subjects was performed to homogenize variances for subsequent analyses. ANOVA was used and each dose of the test compound (specified a priori) was contrasted to the vehicle to determine significant (P < 0.05) dose effects. A linear least-squares regression analysis was conducted and the 30-min period average counts across subjects were regressed over the descending potion of the curve against the  $\log_{10}$  dose of the test compound. The  $ID_{50}$  (dose that produces 1/2) maximal depressant activity where maximum depression = 0) was calculated from the linear regression analysis.

Maximal effects of test compound-cocaine interaction studies. A 30-min time period was selected in which cocaine (20 mg/kg) produced its maximal activity as determined from the studies with cocaine alone. The mean  $(\pm SEM)$  maximal activity (total counts in the 30-min period divided by 3) for vehicle, vehicle pretreatment plus cocaine (20 mg/kg), and test compound pretreatment plus cocaine (20 mg/kg) were plotted in a histogram. A  $\log_{10}$  transformation of the 30-min period average counts for individual subjects was performed to homogenize variances for subsequent analyses. ANOVA was used and vehicle and each dose of the test compound plus cocaine to cocaine alone (specified a priori) were contrasted to determine significant (P < 0.05) dose effects. A linear least-squares regression analysis was performed and the 30-min period average counts across subjects were regressed over the descending portion of the curve against the  $\log_{10}$  dose of the test compound. The AD<sub>50</sub> (dose that attenuates cocaineinduced stimulation by 50%) was calculated from the linear regression analysis.

## **Elemental Analysis**

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-hydroxypiperidine (9).** Anal. calcd for  $C_{24}H_{31}NO_7F_2$ : C, 59.62; H, 6.46; N, 2.90. Found: C, 60.12; H, 6.08; N, 2.84.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-3-hydroxypiperidine (10).** Anal. calcd for  $C_{24}H_{29}NO_6F_2$ : C, 61.93; H, 6.28; N, 3.01. Found: C, 61.84; H, 6.23; N, 2.99.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-3-piperidine carboxamide (11).** Anal. calcd for  $C_{25}H_{32}N_2O_7F_2$ : C, 58.82; H, 6.32; N, 5.49. Found: C, 59.02; H, 5.96; N, 5.29.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-phenylpiperidine-4-carbonitrile (12).** Anal. calcd for  $(C_{31}H_{32}N_2O_5F_2)$ : C, 67.62; H, 5.86; N, 5.09. Found: C, 67.53; H, 5.91; N, 5.05.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-acetyl-4-phenylpiperidine (13).** Anal. calcd for C<sub>32</sub>H<sub>35</sub>NO<sub>6</sub>F<sub>2</sub>: C, 67.71; H, 6.22; N, 2.47. Found: C, 67.86; H, 6.32; N, 2.50.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-phenylpiperidin-4-ol (14).** Anal. calcd for  $C_{30}H_{33}NO_6F_2$ : C, 66.53; H, 6.14; N, 2.59. Found: C, 66.66; H, 6.25; N, 2.59.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-(4-bromophenyl)-4-piperidinol (15).** Anal. calcd for  $C_{30}H_{32}BrNO_6F_2$ : C, 58.07; H, 5.20; N, 2.26. Found: C, 58.09; H, 5.28; N, 2.24.

**1-[4-[bis(4-Fluorophenyl)methoxy]ethyl]-4-(4-bromophenyl)-4-piperidinol (16).** Anal. calcd for  $C_{28}H_{28}BrNO_6F_2$ : C, 56.77; H, 4.76; N, 2.36. Found: C, 56.00; H, 4.96; N, 2.43.

**1-[4-[bis(4-Fluorophenyl)methoxy]butyl]-4-(3-trifluoromethylphenyl)-4-piperidinol (17).** Anal. calcd for  $C_{31}H_{32}NO_6F_5$ : C, 61.08; H, 5.29; N, 2.30. Found: C, 60.96; H, 5.31; N, 2.26.

**4-Benzyl-1-[4-[bis(4-fluorophenyl)methoxy]butyl]-4-piperidinol (18).** Anal. calcd for  $C_{32}H_{42}NO_8F_2$ : C, 61.62; H, 6.95; N, 2.25. Found: C, 61.42; H, 5.82; N, 2.14.

**2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-1,2,3,4-tetrahydroisoquinoline (19).** Anal. calcd for  $C_{28}H_{29}NO_5F_2$ : C, 67.59; H, 5.88; N, 2.82. Found: C, 67.54; H, 5.86; N, 2.79.

**2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-6,7-dimethoxy-1,2,3,4- tetrahydroisoquinoline (20).** Anal. calcd for  $C_{30}H_{33}NO_7F_2$ : C, 64.62; H, 5.97; N, 2.51. Found: C, 64.96; H, 5.88; N, 2.24. **2-[4-[bis(4-Fluorophenyl)methoxy]butyl]-6,7-dimethoxy-1-phenyl-1,2,3,4- tetrahydroisoquinoline (21).** Anal. calcd for  $C_{36}H_{37}NO_7F_2$ : C, 68.23; H, 5.89; N, 2.21. Found: C, 67.69; H, 6.02; N, 2.14.

**1-[1-[4-[bis(4-Fluorophenyl)methoxy]butyl]piperidin-4-yl]-1,3-dihydrobenzoimidazol-2-one (22).** Anal. calcd for  $C_{31}H_{33}N_3O_6F_2$ : C, 64.02; H, 5.72; N, 7.22. Found: C, 63.93; H, 5.67; N, 7.19.

**8-[4-[bis(4-Fluorophenyl)methoxy]butyl]-1-phenyl-1,3,8triazaspiro.**[4,5]decan-4-one (23) Anal. calcd for  $C_{32}H_{35}N_3O_6F_2$ : C, 64.53; H, 5.92; N, 7.05. Found: C, 63.75; H, 6.01; N, 6.86.

**2-[4-[4-[4-Bromophenyl]-4-hydroxypiperidin-1-yl]-butyl]isoindole-1,3-dione (24).** Anal. calcd for C<sub>25</sub>H<sub>27</sub>BrN<sub>2</sub>O<sub>7</sub>: C, 54.85; H, 4.97; N, 5.12. Found: C, 54.33; H, 4.49; N, 5.24.

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#### **References and Notes**

1. (a) Das, G. J. Clin. Pharmacol. **1993**, 33, 296. (b) Substance Abuse and Mental Health Services Administration National Household Survey on Drug Abuse: Main Findings 1997; Department of Health and Human Services: Washington, D.C., 1998. (c) McCance, E. F. *NIDA Res. Monogr.* **1997**, *175*, 36.

2. For reviews, see (a) Carroll, F. I.; Lewin, A. H.; Boja, J. W.; Kuhar, M. J. J. Med. Chem. **1992** 35 969, and references therein (b) Carroll, F. I.; Howell, L. L.; Kuhar, M. J. J. Med. Chem. **1999**, 42, 2721. (c) Howell, L. L; Wilcox, K. M. Prespectives in Pharmacology **2001**, 28, 1.

3. (a) Ritz, M. C.; Lamb, R. J.; Goldberg, S. R.; Kuhar, M. J. *Science* **1987**, *237*, 1219. (b) Kuhar, M. J.; Ritz, M. C.; Boja, J. W. *Trends Neurosci.* **1991**, *14*, 299. (c) Giros, B.; Jaber, M.; Jones, S. R.; Wightman, R. M.; Caron, M. G. *Nature* **1996**, *379*, 606.

4. (a) Reith, M. E. A.; Sershen, H.; Lajtha, A. *Life Sci.* **1980**, 27, 1055. (b) Reith, M. E. A.; Meisler, B. E.; Sershen, H.; Lajtha, A. *Brain Res.* **1985**, *342*, 145.

5. (a) Witkin, J. M. *Neurosci. Biobehav. Rev.* **1994**, *18*, 121. (b) Mello, N. K.; Negus, S. S. *Neuropsychopharmacology* **1996**, *14*, 375.

6. (a) Spealman, R. D. *Psychopharmacology* 1990, *101*, 142.
(b) Witkin, J. M.; Schindler, C. W.; Tella, S. R.; Goldberg, S. R. *Psychopharmacology* 1991, *104*, 425.

7. (a) Corrigall, W. A.; Coen, K. M. Pharmacol. Biochem.

*Behav.* **1991**, *39*, 799. (b) Britton, D. R.; Curzon, P.; Mac-Kenzie, R. G.; Kebabian, J. W.; Williams, J. E. G.; Kerkman, D. *Pharmacol. Biochem. Behav.* **1991**, *39*, 911.

 Shafer, R. A.; Levant, B. *Psychopharmacology* **1998**, *135*, 1.
 Pilla, M.; Perachon, S.; Sautel, F.; Garrido, F.; Mann, A.; Wermuth, C. G.; Schwartz, J.-C.; Everitt, B. J.; Sokoloff, P. *Nature* **1999**, *400*, 371.

10. Rothman, R. B.; Becketts, K. M.; Radesca, L. R.; de Costa, B. R.; Rice, K. C.; Carroll, F. I.; Dersch, C. M. *Life Sci.* **1993**, *53*, PL.

11. Deutsch, H. M.; Collard, D. M.; Zhang, L.; Burnham, K. S.; Deshpande, A. K.; Holtzman, S. G.; Schweri, M. M. J. *Med. Chem.* **1999**, *42*, 882.

12. (a) Simoni, D.; Stoelwinder, J.; Kozikowski, A. P.; Johnson, K. M.; Bergmann, J. S.; Ball, R. G. J. Med. Chem. **1993**, *36*, 3975. (b) Nader, M. A.; Grant, K. A.; Davies, H. M. L.; Mach, R. H.; Childers, S. R. J. Pharmacol. Exp. Ther. **1997**, *280*, 541.

(a) Matecka, D.; Rothman, R. B.; Radesca, L.; de Costa,
 B. R.; Dersch, C. M.; Partilla, J. S.; Pert, A.; Glowa, J. R.;
 Wojnicki, F. H. E.; Rice, K. C. *J. Med. Chem.* **1996**, *39*, 4704.
 (b) Matecka, D.; Lewis, D.; Rothman, R. B.; Dersch, C. M.;
 Wojnicki, F. H. E.; Glowa, J. R.; DeVries, A. C.; Pert, A.;
 Rice, K. C. *J. Med. Chem.* **1997**, *40*, 705. (c) Zhang, Y. *Med. Chem. Res.* **1998**, *8*, 66.

14. (a) Madras, B. K.; Reith, M. E. A.; Meltzer, P. C.; Dutta, A. K. *Eur. J. Pharmacol.* **1994**, *267*, 167. (b) Dutta, A. K.; Coffey, L. L.; Reith, M. E. A. *J. Med. Chem.* **1997**, *40*, 35.

 (a) van der Zee, P.; Koger, H. S.; Gootjes, J.; Hespe, W. *Eur. J. Med. Chem.* **1980**, *15*, 363. (b) Rothman, R. B.; Mele, A.; Reid, A. A.; Akunne, H. C.; Greig, N.; Thurkauf, A.; de Costa, B. R.; Rice, K. C.; Pert, A. *Pharmacol. Biochem. Behav.* **1991**, *40*, 387. (c) Baumann, M. H.; Char, G. U.; de Costa, B. R.; Rice, K. C.; Rothman, R. B. *J. Pharmacol. Exp. Ther.* **1994**, *271*, 1216. (d) Tella, S. R.; Ladenheim, B.; Andrews, A. M.; Goldberg, S. R.; Cadet, J. L. *J. Neurosci.* **1996**, *16*, 7416.
 Lewis, D. B.; Matecka, D.; Zhang, Y.; Hsin, L.-W.; Dersch, C. M.; Stafford, D.; Glowa, J. R.; Rothman, R. B.; Rice, K. C. *J. Med. Chem.* **1999**, *42*, 5029.

17. Bergman, J.; Madras, B. K.; Johnson, S. E.; Spealman, R. D. J. Pharmacol. Exp. Ther. **1989**, 251, 150.

18. Howell, L. L.; Byrd, L. D. J. Pharmacol. Exp. Ther. 1991, 258, 178.

19. Katz, J. L.; Izenwasser, S.; Terry, P. *Psychopharmacology* **2000**, *148*, 90.

20. Howell, L. L.; Czoty, P. W.; Byrd, L. D. Psychopharmacology **1997**, 131, 40.

21. Deutsch, H. M.; Schweri, M. M. Life Sci. 1994, 55, PL.

22. (a) Hanson, R. N.; Choi, S.-W.; Elmaleh, D. R.; Fischman, A. J. Bioorg. Med. Chem. Lett. **1997**, 7, 2559. (b) Choi, S.-W.; Elmaleh, D. R.; Hanson, R. N.; Fischman, A. J. J. Med. Chem. **1999**, 42, 3647.

23. Dutta, A. K.; Meltzer, P. C.; Madras, B. K. Med. Chem. Res. 1993, 3, 209.

24. Taverne, T.; Diouf, O.; Depreux, P.; Poupaert, J. H.; Lesieur, D.; Guardiola-Lemaitre, B.; Renard, P.; Rettori, M-C.; Caignard, D-H.; Pfeiffer, B. J. Med. Chem. **1998**, 41, 2010. 25. Caliendo, G.; Fiorino, F.; Grieco, P.; Perissutti, E.; Santagada, V.; Severino, B.; Bruni, G.; Romeo, M. R. *Bioorg.* Med. Chem. **2000**, 8, 533.

26. Results from an arylpiperazine series are currently under manuscript preparation.

27. Ahmad, Y. E.; Maillet, P.; Laurent, E.; Talab, A.; Teste, J. F.; Cedat, M. J.; Fiez-Vandal, P. Y.; Dokhan, R.; Ollivier, R. *Eur. J. Med. Chem.* **1997**, *32*, 205.

28. Eshleman, A. J.; Carmolli, M.; Cumbay, M.; Martens, C. R.; Neve, K. A.; Janowsky, A. J. Pharmacol. Exp. Ther. **1999**, 289, 877.

29. (a) In vitro pharmacological assays for receptors by NIDA/MDD (NIDA contract NO1DA-7–8072, Contract title: 'Receptor Activity Testing for Medication Discovery'. (b) Chio, C. L.; Lajiness, M. E.; Huff, R. M. *Mol. Pharmacol.* **1994**, *45*, 51.

30. Dubinsky, B.; McGuire, J. L.; Niemegeers, C. J. E.; Janssen, P. A. J.; Weintraub, H. S.; McKenzie, B. E. *Psychopharmacology* **1982**, *78*, 1.

31. After testing compounds **18** and **23** which are also potent NE reuptake inhibitor, NIDA indicated that they are looking for ligands that posses a high DR value and a high selectivity but with a low NE reuptake inhibition. Also, compound **22** contained a partial structure of a strong dopamine D2 receptor antagonist, and thus, metabolism might generate a *N*-dealkylation product, which has freezing activities that can be correlated to the EPS of conventional antipsychotics.

32. Manzanedo, C.; Aguilar, M. A.; Miñarro, J. Psychopharmacology 1999, 143, 82.