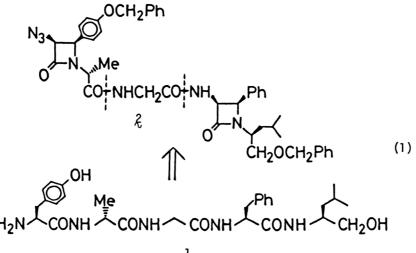
SYNTHESIS OF OPTICALLY PURE ENKEPHALIN ANALOG, [D-A1a², Leu⁵-o1]ENKEPHALIN, USING CHIRAL β -LACTAMS AS SYNTHETIC INTERMEDIATES

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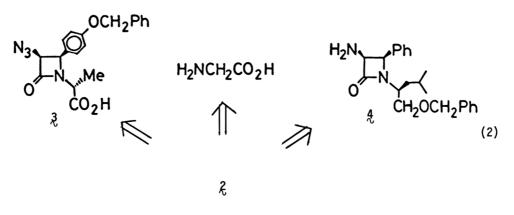
Optically pure pentapeptide, L-Tyr-D-Ala-Gly-L-Phe-L-Leu-ol (1) which is an analog of opioid hormone, enkephalin, was successfully synthesized by using β -lactam building blocks as chiral synthons of peptide units.

Recently, we have developed an entirely new method for peptide synthesis via β -lactam intermediates, ¹⁻⁵which is of considerable advantage to the manipulation of peptides because of the high solubility of the β -lactam intermediates in usual organic solvents and a good performance in chromatography on silica gel. In the preceding paper,⁵ we reported a successful application of the β -lactam method to the synthesis of [Leu⁵]enkephalin t-butyl ester which was, however, racemic except leucine residue. Now, we describe here the synthesis of optically pure [D-Ala², Leu⁵-ol]enkephalin (1) which is a potent long-lasting analog of enkephalin,⁶ an opioid hormone, by using chiral β -lactam building blocks.

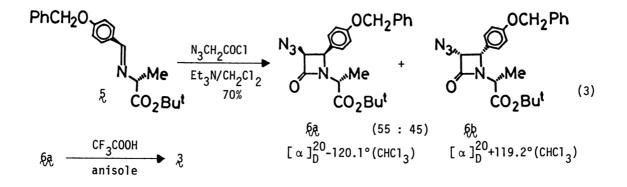
As the final precursor of $[D-Ala^2$, $Leu^5-ol]$ enkephalin (1) we planned, based on the retrosynthetic scheme (eq. 1), to synthesize a chiral bis- β -lactam (2) which would readily be converted to 1 by hydrogenolysis on palladium.¹⁻⁵



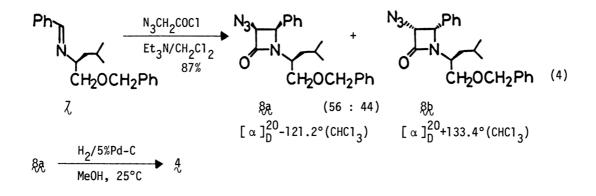
The bis- β -lactam (2) consists of three units, i.e., the β -lactam (3) which is a synthon of Tyr-(D)-Ala, glycine, and the β -lactam (4) which is a synthon of Phe-Leu-ol (eq. 2).



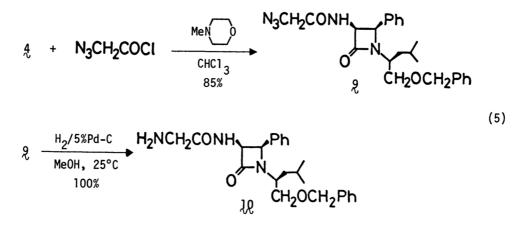
The β -lactam (3) was prepared via the in situ azidoketene addition to t-butyl 4-benzyloxybenzylidene-(D)-alaninate (5) followed by HPLC separation of two diastereomers (β_a and β_b)⁷ on silica gel (n-hexane/AcOEt=2/1) and deprotection of β_a (eq. 3).



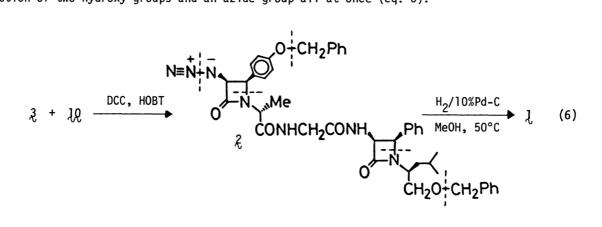
In a similar manner, the β -lactam (4) was prepared from benzylideneleucinol benzyl ether (7) and azidoacetyl chloride followed by HPLC separation of two diastereomers (8a and 8b)⁸ on silica gel (n-hexane/AcOEt=4/1), and reduction of the azide group of 8a (eq. 4).



We employed azidoacetyl chloride as glycine synthon. Thus, the β -lactam (4) was azidoacetylated to give the azidoacetyl- β -lactam (9), which was transformed to the tripeptide synthon (10) by the reduction of azide moiety of 9 (eq. 5).



The coupling of Tyr-(D)-Ala synthon (\mathfrak{Z}) and Gly-Phe-Leu-ol synthon (\mathfrak{IQ}) by using dicyclohexylcarbodiimide(DCC) and l-hydroxybenzotriazole(HOBT) in dimethylformamide gave the bis- β -lactam (\mathfrak{Z}),⁹ which is the planned final precursor of \mathfrak{I} , in 84% yield after purification on silica gel column (AcOEt). Then, the pentapeptide synthon (\mathfrak{Z}) was submitted to hydrogenolysis on 10% Pd-C in methanol at 50°C to give \mathfrak{I}^{10} in 85% yield through the reductive cleavage of two β -lactam rings and the deprotection of two hydroxy groups and an azide group all at once (eq. 6).



It is noteworthy that the β -lactam ring of 3 acts not only as tyrosine synthon but also as excellent protecting group of D-alanine. According to the established rationale of the mechanism of racemization during peptide synthesis, the formation of oxazolone using an acylamino proton or an alkoxycarbonylamino proton is crucial,¹¹ which is more or less inevitable as far as ordinary protecting groups are employed. However, in the Tyr-(D)-Ala synthon (3), two amino protons of (D)alanine are protected by the β -lactam ring; the racemization at chiral center cannot take place via oxazolone formation. Actually, no racemization was detected during the DCC-HOBT coupling of 3 and 10. This must be another advantageous feature of the β -lactam method. As we have already disclosed, β -lactam building blocks exhibited good solubility toward usual organic solvents, readily purified on usual silica gel column, and fully characterized by spectroscopic analyses. In addition to these, i) the β -lactam building blocks, $\beta \beta$ and $\beta \beta$, which were not used in the present synthesis, act as the chiral synthons of (D)-Tyr-(D)-Ala and (D)-Phe-Leu-ol, respectively: These are useful for the synthesis of other analogs, and ii) a variety of aromatic substituents, e.g., p-fluorophenyl, 3,4-dihydroxyphenyl, indole, furan, and pyrrole, etc., can be introduced to the β -lactam building blocks simply by using the corresponding aromatic aldehydes. These characteristics together with the stereochemical stability of β -lactam building blocks may provide a powerful device for the synthesis of various physiologically active oligopeptides.

References

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- 7. The absolute configurations of 6a and 6b were unambiguously determined on the basis of NMR and HPLC analyses by the comparison of the two diastereomers of Tyr-(D)-Ala-O^tBu which were obtained by the hydrogenolysis of 6a and 6b following our method, $^{1-5}$ with the authentically prepared (L)-Tyr-(D)-Ala-O^tBu.
- 8. The absolute configurations of &a and &b were unambiguously determined in a manner similar to the case of &a and &b on the basis of NMR and HPLC analyses by using authentically prepared (L)-Phe-(L)-Leu-ol; It was found that &a gave (L)-Phe-(L)Leu-ol and &b gave (D)-Phe-(L)-Leu-ol by hydrogenolysis.
- 9. Hygroscopic colorless solid; NMR(CDCl₃) δ 0.92 (d, J=5.5Hz, 3H), 0.94 (d, J=5.5Hz, 3H), 1.07 (d, J=7Hz, 3H), 1.14-2.05 (m, 3H), 3.14-3.42 (m, 2H), 3.56 (d, J=6Hz, 2H), 3.80 (m, 1H), 4.31 (q, J=7Hz, 1H), 4.24 (s, 2H), 4.95 (d, J=5Hz, 1H), 5.01 (ABq, J=5Hz, 2H), 5.03 (s, 2H), 5.41 (d of d, J=5Hz, 9Hz, 1H), 6.85-7.50 (m, 21H); IR(KBr) 3320 ($^{\nu}$ NH), 2110 ($^{\nu}$ N₃), 1750 ($^{\nu}$ C=0), 1660 (Amide I), 1540 (Amide II) cm⁻¹; Anal. Calcd. for C₄₃H₄₇N₇O₆.3/2H₂O; C, 65.80; H, 6.42; N, 12.49. Found: C, 65.58; H, 6.23; N, 12.38.; [α]_D²⁰-55.3° (c=1.07, CHCl₃).
- 10. Extremely hygroscopic colorless solid; IR(KBr) 3300 ($^{\nu}$ NH, $^{\nu}$ OH), 1655 (Amide I), 1540 (Amide II) cm⁻¹; Anal. Calcd. for C₂₉H₄₁N₅O₆.5H₂O: C, 53.94; H, 7.90; N, 10.84. Found: C, 53.75; H, 7.47; N, 10.44.; [α]_D²⁰ +15.8° (c=0.91, MeOH). The product was also identified by comparing the HPLC chromatogram with that of authentically prepared HCl.H-(L)-Tyr-(D)-Ala-Gly-(L)-Phe-(L)-Leu-ol. HPLC analysis was carried out using a column packed with TOYO SODA LS 410K (ODS SIL) and MeOH-H₂O.
- e.g., M. Bodanszky in "The Peptides", Vol. 1, ed. by E. Gross and J. Meienhofer, Academic Press, New York, 1979, pp 152-156.

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