Synthesis and Radical Ring-Opening Polymerization of Vinylcyclopropanes Derived from Amino Acids with Hydrophobic Moieties

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ABSTRACT: Six 1,1-disubstituted vinylcyclopropanes (VCP) were synthesized from glycine and amino acids bearing hydrophobic moieties, L-alanine, L-valine, L-leucine, L-isoleucine, and L-phenylalanine. These VCP derivatives efficiently underwent radical ring-opening polymerization to afford the corresponding polymers bearing *trans*-vinylene moiety in the main chains and the amino acid-derived chiral moieties in the side chains. The polymers were film-formable, and in the films of polymers bearing the glycine- and alanine-derived side chains, presence of hydrogen bonding was confirmed by IR analysis. Thermogravimetric analysis of the polymers revealed that the temperatures of 5% weight loss were higher than 300 °C. Differential scanning calorimetry clarified that the polymers were amorphous ones showing glass transition temperatures in a range of 48–80 °C. © 2017 Wiley Periodicals, Inc. J. Polym. Sci., Part A: Polym. Chem. **2017**, *00*, 000–000

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INTRODUCTION Radical ring-opening polymerization (RROP) is one of the attractive methods to synthesize functional polymers bearing heteroatoms and functional groups in their main chains. In general, monomers that undergo RROP efficiently fulfill the following four requirements:¹ (1) Monomers should have a C—C double bond which can accept radical species. (2) They should have distorted ring structures. (3) Preferably, their ring-opening reactions accompany isomerization processes to form thermodynamically more stable structures. (4) The ring-opening reaction results in formation of radicals that can be stabilized by appropriately chosen and located functional groups.

Vinylcyclopropane (VCP) derivatives have attracted considerable attention as cyclic monomers undergoing RROP.¹⁻⁶ Among various VCPs, 1,1-disubstituted ones have been investigated most extensively. Their RROPs give the corresponding polymers bearing C—C double bonds in the main chains, often accompanied by a side reaction giving cyclobutane rings (Scheme 1). The successful RROP of these VCPs arise from the high distortion energy of cyclopropane ring and stabilization of the resulting radical by the two substituents at 1-position such as halogen and ester moieties. Controlled RROPs of VCP derivatives have been also achieved by nitroxide-mediated polymerizations in the presence of 2,2,6,6-tetramethylpiperidin-1-oxyl,⁷ by using the reversible addition-fragmentation chain transfer protocol,⁸ and using the atom transfer radical polymerization protocol.⁹ In addition, copolymerizations with allylic carbonate,¹⁰ methyl acrylate,¹¹ and other VCPs¹² have been reported. The copolymerization with methyl methacrylate afforded the corresponding polymer bearing cycloalkane moieties in the main chain. There have been reports on relatively low volume shrinkage accompanying the RROP of VCPs, which makes VCPs attractive as potential monomers applicable to coating, holographic data storages, and dental materials.¹³⁻¹⁶

Recently, RROPs of VCPs bearing amide moieties at 1position have been reported.^{15,16} Interestingly, the polymerizations were accelerated by hydrogen bonds between amide moieties. 1,1-Disubstituted VCP derivatives, on which α amino acid-derived components are attached through amide

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SCHEME 1 RROP system of VCP derivatives.

bonds, are monomers of high interest, because they can be used for synthesis of functionalized polymers with optically active nature¹⁷⁻²⁰ and C—C double bonds in the main chain that can be chemically modified by various methods.^{21,22}

Herein, we report 1,1-disubstituted VCP monomers **1a-1f** bearing (*L*)-amino acid-derived moieties (Fig. 1). Among various amino acids, glycine and α -alkyl-substituted ones such as alanine, valine, leucine, isoleucine, and phenylalanine were selected and used. The polymerization behaviors of **1a-1f** were studied to find dependence on the structures of each amino acid moiety. These corresponding polymers have chirality arising from amino acid and ester parts available for postmodification.

RESULTS AND DISCUSSION

Synthesis of VCP Derivatives

Scheme 2 shows the route for the synthesis of the VCP derivatives **1a–1f**. Previously, we reported the synthesis of L-alanine-derived VCP **1b** in this route.²³ 1,1-Diethoxycarbonyl-2-vinylcyclopropane (DECVCP), which was synthesized by the reaction of diethyl malonate with 1,4-dibromobutene, was used as the starting material. Its ethyl esters were hydrolyzed under basic conditions and subsequent acidification with hydrochloric acid to yield 1,1-dicarboxy-2-vinylcy-clopropane (DCAVCP). In the final step, DCAVCP was



FIGURE 1 Molecular structures of VCPs bearing α -amino acidderived moieties.



SCHEME 2 Synthesis of VCP derivatives 1.



FIGURE 2 ¹H NMR spectrum of 1a in CDCl₃.





FIGURE 3 ¹H NMR spectrum of an epimeric mixture of **1c** (in CDCl₃).



condensed with methyl esters of amino acids using *N*,*N*'-dicyclohexylcarbodiimide (DCC), 1-hydroxybenzotriazole (HOBt), and *N*,*N*-diisopropylethylamine (DIPEA) to afford the VCP derivatives **1**. Except the glycine-derived **1a**, the VCPs **1b-1e** bearing L-amino acid-derived moieties, were obtained as 1:1 epimeric mixtures.

Figure 2 shows the ¹H NMR spectrum of glycine-derived VCP **1a.** In the spectrum, the signals of the glycine-derived moiety in the *cis*-configuration to the vinyl moiety were distinguished from those of the other glycine-derived moiety in the *trans*-configuration to the vinyl moiety. For example, there are two signals at 7.9 and 7.2 ppm, which are attributable to the two amide protons *h* and *e*, respectively. For these assignments, we referred our previous investigation on the molecular structure of alanine-derived VCP **1b** by combining X-ray crystallography and NOE analysis, which clarified that the signal for the NH located in the *trans*-configuration to the vinyl moiety appeared in a lower magnetic field than that for the other NH located in the *cis*-configuration to the vinyl moiety.

Figure 3 shows the spectrum of L-valine-derived VCP **1c**, where the presence of two epimers *RSS*-**1c** and *SSS*-**1c** was confirmed. The signals were assigned as indicated in the

TABLE 1	RROP	of '	1
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Figure 3, according to the assignment made for the L-alanine-derived VCP **1b**. ¹H-¹H COSY and ¹H-¹H NOESY analyses of **1c** supported the assignment (Figs. S1 and S2, Supporting Information).

RROP of Vinylcyclopropanes 1

The polymerizations of vinylcyclopropanes 1a-1f were performed with azobisisobutyronitrile (AIBN) as an initiator (Scheme 3, Table 1). The vinylcyclopropanes 1b-1f derived from L-amino acids were used as epimeric mixtures as obtained, because the chirality of the cyclopropane ring would be lost upon its ring-opening reaction during the polymerization to afford the polymers with no chiral centers in the backbone. The solvent was chosen from N,N-dimethylformamide (DMF) and toluene depending on the solubility of the resulting polymers 2: The polymerizations of 1a, 1b, and 1f proceeded in DMF efficiently, leading to the almost quantitative conversions of the monomers. The resulting polymers 2a, 2b, and 2f were soluble in DMF. On the other hand, those of 1c, 1d, and 1e, bearing bulky and highly hydrophobic branching alkyl groups, were performed in toluene. The corresponding polymers 2c, 2d, and 2e were soluble in toluene but insoluble in DMF.

The structures of the polymers were analyzed with ¹H NMR, ¹³C NMR, and IR. The spectra supported the polymer structures depicted in Scheme 3, that is, they were not contaminated by cyclobutane rings and had a *trans*-alkylene backbone (Fig. S3, Supporting Information). As typical examples, ¹H and ¹³C NMR spectra of **2c** obtained by RROP of **1c** (Entry 3 in Table 1) are shown in Figure 4.

In the ¹H NMR spectrum of **2c**, signals attributable to the protons *a* and *b* on the vinylene moiety in the main chain appeared at 2.6 and 5.5 ppm, respectively. As well, in the ¹³C NMR spectrum of **2c**, signals attributable to the carbons *a* and *b* on the vinylene moiety appeared at 52 and 130 ppm, respectively. In addition, in the IR spectrum of **2c**, an absorption corresponding to the *trans*-alkylene moiety in the backbone was observed at 996 cm⁻¹. The structures of the other polymers were also confirmed similarly.

Entry	1	Solvent	Conversion of 1 (%) ^a	2	Yield (%)	<i>M</i> _n (×10 ³)	$M_{\rm w}/M_{\rm n}$
1	1a	DMF	97	2a ^c	49	20 ^d	3.9 ^d
2	1b	DMF	>99	2b ^c	66	22 ^d	2.4 ^d
3	1c	Toluene	92	2c ^b	53	21 ^e	1.9 ^e
4	1d	Toluene	86	2d ^b	45	15 ^e	1.6 ^e
5	1e	Toluene	87	2e ^b	71	17 ^e	1.9 ^e
6	1f	DMF	>99	2f ^c	52	31 ^d	2.1 ^d

DMF containing 10 mM LiBr).

chloroform).

 $^{\rm a}$ Determined by calculating the ratio between vinyl moiety and methoxy moiety of $^{\rm 1}{\rm H}$ NMR.

^b Isolated as insoluble parts in a mixture of diethyl ether and hexane (1:3).

^c Isolated as insoluble parts in diethyl ether.



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^d Estimated by SEC (calibrated by polystyrene standards, eluted with

^e Estimated by SEC (calibrated by polystyrene standards, eluted with



FIGURE 4 ¹H and ¹³C NMR spectra of 2c in CDCl₃.

Properties of Polymers 2

The resulting polymers were subjected to thermogravimetric analysis (TGA) to evaluate their thermal stability. The thermograms are shown in Figure S4, Supporting Information. For all the polymers, the temperatures of 5% weight loss by thermal degradation (T_{d5}) were higher than 300 °C (Table 2).

The glass transition temperatures (T_g) of the polymers were determined by differential scanning calorimetry (DSC) in a dynamic mode with heating rate of 10 °C/min. The polymers were heated up to 200 °C, cooled to -110 °C, and then analyzed with DSC. The resulting thermograms are shown in Figure 5. The T_g values decreased as the bulkiness of the amino acid-derived substituent increased, implying that the bulky substituents in the side chains hindered intermolecular hydrogen bonding.^{24,25} The relatively high T_g of **2f** would be attributable to the rigidity of the phenyl group.

For investigating the presence and degree of hydrogen bonding in the resulting polymers **2**, their films were prepared and analyzed by IR with focusing on the absorptions attributable to the amide NH in the polymers (Fig. 6). Each of the

TABLE 2 Thermal properties of 2

Entry	2	T _{d5} (°C) ^a	τ _g (°C) ^k
1	2a	300	80
2	2b	305	74
3	2c	323	60
4	2d	310	57
5	2e	323	48
6	2f	310	64

^a Determined by TGA.

^b Determined by DSC.

(2a) 80 °C (2b) '4 °C (2c) Exothermic 60 °C (2d) 57 °C (2e) 48 °C (2f) 64 °C 0 40 80 120

FIGURE 5 DSC thermograms of polymers **2** along with their glass transition temperatures.

Temperature /

°C

polymers were fabricated into two films by two different processes: (1) One film (film A) was prepared by casting a polymer from its chloroform solution on the ATR prism of the IR instrument with rapid evaporation of chloroform within 1 min, and was analyzed with IR immediately. (2) The other one (film B) was prepared by casting the same solution on a Teflon petri dish with slow evaporation of chloroform taking 24 h in an incubating chamber, and then was analyzed with IR.

In the IR spectrum of film A of 2a [Fig. 6(a), in a solid line], the amide NH absorption appeared at 3351 cm^{-1} . On the other hand, in the IR spectrum of film B of 2a [Fig. 6(a)], in a dotted line), the amide NH absorption appeared at 3337 cm^{-1} . The difference in wavenumber indicated that the degree of hydrogen bonding in film A was less than that in film B, that is, the rapid evaporation of chloroform led to the formation of a polymer film with insufficiently grown hydrogen bonding, while the slow evaporation of chloroform permitted more efficient growth of hydrogen bonding. A similar red shift of the amide NH absorption raised by the slow evaporation was also observed for 2b. In addition, in the IR spectrum of **2b**, the absorption at 1665 cm^{-1} attributable to the amide carbonyl stretching slightly red-shifted with accompanying a newly appeared shoulder in the red side (Fig. S5, Supporting Information), supporting the formation of hydrogen bond between amide NH and amide carbonyl moieties.

On the other hand, such clear red shifts were not observed in the IR spectra of films of **2c**, **2d**, and **2e**, regardless of film fabrication process, implying that hydrogen bonding in these polymers was sterically hindered by the bulkier substituents such as branched alkyls and benzyl in the amino acid-derived moieties [Fig. 6(c, d, and e)].



FIGURE 6 Expanded IR spectra of **2a** (a), **2b** (b), **2c** (c), **2d** (d), **2e** (e), and **2f** (f) in a range from 3200 to 3500 cm⁻¹. Solid lines indicate the IR spectra of thin films prepared on cell plate by casting from chloroform and evaporated in one minute. Dotted lines indicate the IR spectra of films prepared by casting from chloroform in an incubating chamber for 24 hours.

CONCLUSION

A series of vinylcyclopropanes bearing amino acid-derived moieties were synthesized and their radical ring-opening polymerizations (RROPs) were performed. The polymerizations proceeded efficiently to afford the corresponding polymers, of which side chains inherited the amino acid-derived moieties from the monomers. The NMR analyses of the polymers confirmed the well-defined structures of the main chains and side chains. This study using L-amino acidderived vinylcyclopropanes has opened up possibilities of RROP as a new efficient and reliable approach to optical polymers, which are expected as new functional materials for chiral recognition, chiral discrimination, and chiral catalysis.

EXPERIMENTAL

Instruments

¹H and ¹³C NMR spectra were recorded with a JEOL ECS-400 (400 MHz) spectrometer, and chemical shifts were recorded in ppm units using tetramethylsilane or a solvent signal as an internal standard. IR spectra were recorded on Thermo Scientific Nicolet iS10 spectrometer equipped with a Smart iTR Sampling Accessory using attenuated total reflection (ATR) method. For the estimation of number average molecular weight (M_n) and weight average molecular weight (M_w) of the synthesized polymers, size-exclusion chromatography (SEC) was performed on a TOSOH HLC-8220 system equipped with three consecutive polystyrene gel columns [TSK-gels (bead size, exclusion limited molecular weight); super-AW4000 (6 μ m, > 4 \times 10⁵), super-AW3000 (4 μ m, > 6 \times 10⁴), and super-AW2500 (4 μ m, >2 \times 10³)] and refractive index and ultraviolet detectors. The system was operated at 40 °C using 10 mM LiBr in DMF as the eluent, at a flow rate of 0.5 mL/min. Polystyrene standards were used



for calibration. The SEC using chloroform as eluent was also performed on Tosoh TSKgel SuperHM-H column (6.0 mm $\varphi \times 15$ cm, 3 and 5 μ m bead sizes) at a flow rate of 0.5 mL/min at 40 °C. High-resolution mass spectra (HRMS) were obtained on a JEOL JMN-700 spectrometer using electron impact (EI) ionization mode. DSC was carried out using Seiko Instrument DSC-6200R in an aluminum pan at a heating rate 10 °C/min under a nitrogen flow of 50 mL/min. TGA was performed on a Seiko Instrument TG-DTA 6200 using an aluminum pan under nitrogen flow of 100 mL/min at a heating rate of 10 °C/min.

Chemicals

L-Amino acids, sodium hydroxide, NaH (60% dispersion in mineral oil), diethylmalonate, thionyl chloride, HOBt monohydrate, DCC, and all solvents were purchased from Wako Pure Chemical Industry (Osaka, Japan). *N*,*N*-Diisopropylethylamine, 1,4-dibromo-2-butene, and sodium hydrate were purchased from Tokyo Chemical Industry (Tokyo Japan). Tetrahydrofurane (THF), toluene, and DMF were distilled prior to use. Other solvents and reagents were used as received.

Synthesis

Synthesis of Vinylcyclopropanes 1 Typical procedure

To a solution of DCAVCP (989 mg, 6.34 mmol), which was synthesize by previously reported method,²³ in DMF (15 mL), a solution of glycine methyl ester hydrochloride (1.64 g, 11.8 mmol) and DIPEA (2.1 mL, 12.1) in DMF (25 mL) was added. The resulting mixture was cooled to 0 °C, then DCC (2.57 g, 13.4 mmol) and HOBt monohydrate (1.74 g, 12.9 mmol) were added to the mixture. After stirring the mixture at room temperature for 24 h, 2% hydrochloric acid was added and the resulting precipitate was removed by filtration. The filtrate was diluted with chloroform,

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washed by sat. NaHCO₃ aq., and brine. The organic layer was dried over Na₂SO₄, flittered, and evaporated. The residue was fractionated by flash silica-gel column chromatography (eluent: ethyl acetate and hexane), and the fraction containing 1a were collected, evaporated, and dried under vacuum to obtain **1a** (1.19 g, 3.99 mmol, 63%): ¹H NMR (CDCl₃): δ 7.87 (1H, br t, J = 6.0 Hz, NH anti to vinyl), 7.16 (1H, br t, J = 6.0 Hz, NH syn to vinyl), 5.53–5.44 (1H, m, CH=CH₂), 5.31 (1H, d, J = 15.6 Hz, C=CH (trans)), 5.14 (1H, d, J = 7.8 Hz, C=CH (cis)), 4.25 and 4.15 (1H + 1H, dd, J = 6.4, 18.4 Hz, N—CH \times 2), 3.96 and 3.89 (1H + 1H, dd, J = 5.2, 18.4 Hz, N—CH \times 2), 3.77 and 3.75 (3H + 3H, s, OCH₃ \times 2), 2.48– 242 (1H, m, C=C-CH \times 2), 1.76 (1H, dd, J = 6.4, 18.0 Hz, CH trans to vinyl), 1.61 (1H, dd, J = 5.2, 18.0 Hz, CH cis to vinyl); HRMS (EI): found m/z 298.1165, calcd. for C₁₅H₂₂N₂O₆; M⁺: 298.1165.

1b was Synthesized by Previously Reported Method.²³

According to the typical procedure, DCAVCP and L-valine methyl ester hydrochloride were condensed to obtain **1c** (2.47 g, 6.47 mmol, 68%): ¹H NMR (CDCl₃): δ 7.66 and 7.53 (1H + 1H, d, *J* = 8.0 Hz, NH *anti* to vinyl × 2), 7.00 and 6.80 (1H + 1H, d, *J* = 7.6, NH *syn* to vinyl × 2), 5.63-5.54, 5.47-5.39 (1H, m, CH=CH₂ × 2), 5.35-5.28 (2H, m, C=CH (*trans*) × 2), 5.20-5.11 (2H, m, C=CH (*cis*) × 2), 4.53-4.45 (4H, m, N-CH × 4), 3.76, 3.74₉, 3.74₅, 3.73 (each 3H, s, OCH₃ × 4), 2.51-2.45, 2.36-2.30 (1H + 1H, m, C=C-CH × 2), 2.28-2.15 (4H, m, CH(CH₃)₂ × 4), 1.83, 1.70 (1H + 1H, dd, *J* = 5.2, 9.2 Hz, CH (*cis* to vinyl) × 2), 1.65, 1.55 (1H + 1H, dd, *J* = 5.2, 6.8 Hz, CH (*trans* to vinyl)), 0.983-0.918 (12H, m, CH(CH₃)₂ × 2): HRMS (EI): found *m*/*z* 382.2105, calcd. for C₁₅H₂₂N₂O₆; M⁺: 382.2104.

According to the typical procedure, DCAVCP and L-leucine methyl ester hydrochloride were condensed to obtain **1d** (2.34 g, 5.71 mmol, 85%): ¹H NMR (CDCl₃): δ 7.87, 7.40 (1H + 1H, d, *J* = 7.2 Hz, NH *anti* to vinyl × 2), 7.04, 7.01 (1H + 1H, d, *J* = 7.2, NH *syn* to vinyl × 2), 5.62-5.53 (1H, m, C<u>H</u>=CH₂), 5.39-5.26 (3H, m, C<u>H</u>=CH₂ + CH=C<u>H</u> (*trans*) × 2), 5.16-5.07 (2H, m, CH=C<u>H</u> (*cis*) × 2), 4.62-4.52 (4H, m, N—CH × 4), 3.76, 3.75, 3.74, 3.73 (each 3H, s, OCH₃ × 4), 2.61-2.55, 2.35-2.29 (1H + 1H, m, C=C—CH × 2), 1.80-1.60 (15H, m, C=C—CH—C<u>H</u> × 3 + C<u>H₂CH</u>(CH₃)₂ × 4), 1.48 (1H, dd, *J* = 5.2, 7.2 Hz, CH (*trans* to vinyl)), 0.96-0.90 (12H, m, CH(C<u>H₃)₂ × 2</u>); HRMS (EI): found *m*/*z* 410.2416, calcd. for C₁₅H₂₂N₂O₆; M⁺: 410.2417.

According to the typical procedure, DCAVCP and L-isoleucine methyl ester hydrochloride were condensed to obtain **1e** (2.80 g, 6.83 mmol, 87%): ¹H NMR (CDCl₃): δ 7.70, 7.53 (1H + 1H, d, *J* = 7.6 Hz, NH *anti* to vinyl × 2), 7.00, 6.84 (1H + 1H, d, *J* = 8.0 Hz, NH *syn* to vinyl × 2), 5.56-5.51 (1H, m, CH=CH₂), 5.46-5.37 (1H, m, CH=CH₂), 5.34-5.27 (2H, m, C=CH (*trans*) × 2), 5.19-5.10 (2H, m, CH=CH (*cis*) × 2), 4.57-4.46 (4H, m, N-CH × 4), 3.76, 3.74, 3.74, 3.73 (each 3H, s, OCH₃ × 4), 2.49-242, 2.35-2.29 (1H + 1H, m, C=C-CH × 2), 2.01-1.86 (4H, m, CH(CH₃)CH₂ × 4) 1.81, 1.69 (1H + 1H, dd, *J* = 5.2, 8.8 Hz, CH (*cis* to vinyl) × 2) 1.62

and 1.53 (1H, dd, J = 4.8, 7.6 and 5.6, 6.8 Hz, CH (*trans* to vinyl)), 0.95-0.81 (12H, m, CH(CH₃)₂ × 2): HRMS (EI): found m/z 410.2417, calcd. for C₁₅H₂₂N₂O₆; M⁺: 410.2417.

According to the typical procedure, DCAVCP and L-phenylalanine methyl ester hydrochloride were condensed to obtain **1f** (1.07 g, 2.2 mmol, 60%): ¹H NMR (CDCl₃): δ 7.69, 7.40 (1H + 1H, d, *J* = 8.0 Hz, NH *anti* to vinyl × 2), 7.29-7.11 (10H, m, aromatic) 6.87, 6.81 (1H + 1H, d, *J* = 7.2, NH *syn* to vinyl × 2), 5.20-5.09, 5.02–4.99, 4.91-4.88 (4H + 1H + 1H, m, C<u>H=CH₂ × 2)</u>, 4.89-4.73 (4H, m, N-CH × 4), 3.76, 3.73, 3.70, 3.69 (each 3H, s, OCH₃ × 4), 3.23-3.11, 3.07-2.96 (4H + 4H, m, C<u>H₂-phenyl</u>), 2.35-2.30, 2.11-2.05 (1H + 1H, m, C=C-CH × 2), 1.69, 1.49, 1.31 (1H + 2H + 1H, m, CH (*cis* to vinyl) × 2 + CH (*trans* to vinyl) × 2). HRMS (EI): found *m/z* 478.2105, calcd. for C₂₇H₃₀N₂O₆; M⁺: 478.2104.

Radical ROP of Vinylcyclopropanes 1 Typical procedure

Vinylcyclopropane **1a** (150.0 mg, 503 µmol) and AIBN (2.4 mg, 15.1 µmol, 3 mol% to **1a**) were dissolved in 0.5 µL of distilled DMF. The solution was degassed by three freeze-thaw cycles, sealed, and heated at 60 °C. After 20 h, the solution was diluted with chloroform (5 mL) and added to diethyl ether (300 mL). The resulting white precipitates were collected by filtration with suction and dried under vacuum to obtain polymer **2a** (101.1 mg, 67%): ¹H NMR (CDCl₃): δ 7.65 (2H, brs, NH × 2), 5.54 (2H, brs, CH=CH), 3.99 (4H, d, *J* = 4.4 Hz, N–CH₂ × 2), 3.72 (6H, brs, OCH₃ × 2), 2.56 (4H, brs, CH₂–CH=CH–CH₂); ¹³C NMR (CDCl₃): δ 172.66, 170.70, 129.44, 56.81, 52.40, 41.59, 39.05; IR (ATR): ϵ 3351 (NH), 1742 (C=O ester), 1662 (C=O amide), 978 (C=CH *trans*).

According to the typical procedure, **1c** was polymerized in toluene to obtain polymer **2c** (285.6 mg, 53%): ¹H NMR (CDCl₃): δ 7.46 (2H, brd, *J* = 7.6 Hz, NH × 2), 5.52 (2H, brs, CH=CH), 4.43 (2H, dd, *J* = 5.2, 7.6 Hz, N—CH × 2), 3.73 (6H, brs, OCH₃ × 2), 2.55 (4H, brs, CH₂—CH=CH—CH₂), 2.20-2.12 (2H, m, CH(CH₃)₂ × 2), 0.90 (12H, t, *J* = 6.0Hz, CH₃ × 4); ¹³C NMR (CDCl₃): δ 172.01, 171.75, 129.69, 57.84, 56.81, 52.17, 39.78, 30.61, 19.05, 18.06; IR (ATR): ε 3358 (NH), 1740 (C=O ester), 1663 (C=O amide), 993 (C=CH *trans*).

According to the typical procedure, **1d** was polymerized in toluene to obtain polymer **2d** (259.3 mg, 45%): ¹H NMR (CDCl₃): δ 7.32 (2H, br d, *J* = 6.8 Hz, NH × 2), 5.51 (2H, brs, CH=CH), 4.50 (2H, br t, *J* = 6.8 Hz, N—CH × 2), 3.72 (6H, br s, OCH₃ × 2), 2.51 (4H, brs, CH₂—CH=CH—CH₂), 1.60 (6H, brs, CH₂CH(CH₃)₂ × 2), 0.92 (12H, t, *J* = 4.0Hz, CH₃ × 4); ¹³C NMR (CDCl₃): δ 173.65, 171.93, 129.62, 56.98, 52.36, 51.347, 40.167, 39.57, 24.91, 22.94, 21.48; IR (ATR): ϵ 3349 (NH), 1740 (C=O ester), 1668 (C=O amide), 981 (C=CH *trans*).

According to the typical procedure, **1e** was polymerized in toluene to obtain polymer **2e** (639.3 mg, 71%). ¹H NMR (CDCl₃): δ 7.46 (2H, br d, *J* = 6.8 Hz, NH × 2), 5.52 (2H, br s, CH=CH), 4.47 (2H, br t, *J* = 6.0 Hz, N–CH × 2), 3.73 (6H,

br s, OCH₃ × 2), 2.54 (4H, br s, <u>CH₂</u>—CH=CH—CH₂), 1.88 (2H, br s, NCHCH × 2), 1.43-1.11 (4H, m, (CH₂CH₃) × 2) 0.92-0.86 (12H, m, J = 4.0Hz, CH₃ × 4); ¹³C NMR (CDCl₃): 171.98, 171.58, 129.76, 57.07, 56.72, 52.11, 39.75, 37.20, 25.33, 15.58, 11.58; IR (ATR): ε 3368 (NH), 1739 (C=0 ester), 1672 (C=0 amide), 985 (C=CH *trans*).

According to the typical procedure, **1f** was polymerized in DMF to obtain polymer **2f** (124.7 mg, 52%): ¹H NMR (CDCl₃): δ 7.35 (2H, br d, J = 6.4 Hz, NH × 2), 7.25-7.06 (20H, m, aromatic), 5.16 (2H, brs, CH=CH), 4.47 (2H, brt, J = 6.4, 6.4 Hz, N—CH × 2), 3.62 (6H, brs, OCH₃ × 2), 3.09-2.96 (4H, m, CH₂-phenyl × 2) 2.24 (4H, brs, <u>CH₂</u>—CH=CH-CH₂); ¹³C NMR (CDCl₃): δ 171.89, 171.45, 136.18, 129.11, 128.56, 128.31, 127.07, 56.33, 54.02, 52.29, 39.18, 37.41; IR (ATR): ϵ 3358 (NH), 1740 (C=O ester), 1663 (C=O amide), 983 (C=CH *trans*).

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