

Physicochemical characterization of the dimeric lanthanide complexes $[en{Ln(DO3A)(H_2O)}_2]$ and $[pi{Ln(DTTA)(H_2O)}_2]^{2-}$: a variable-temperature ¹⁷O NMR study

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The Gd(III) complexes of the two dimeric ligands [en(DO3A)₂] {*N*,*N*'-bis[1,4,7-tris(carboxymethyl)-1,4,7,10tetraazacyclododecan-10-yl-methylcarbonyl]-N,N'-ethylenediamine} and [pi(DTTA)2]⁸⁻ [bisdiethylenetriaminepentaacetic acid (trans-1,2-cyclohexanediamine)] were synthesized and characterized. The ¹⁷O NMR chemical shift of H₂O induced by $[en{Dy(DO3A)}_2]$ and $[pi{Dy(DTTA)}_2]^{2-}$ at pH 6.80 proved the presence of 2.1 and 2.2 inner-sphere water molecules, respectively. Water proton spin-lattice relaxation rates for $[en{Gd(DO3A)(H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ at $37.0 \pm 0.1 \,^{\circ}C$ and 20 MHz are 3.60 ± 0.05 and 5.25 ± 0.05 mm $^{-1}$ s $^{-1}$ per Gd, respectively. The EPR transverse electronic relaxation rate and ¹⁷O NMR transverse relaxation time for the exchange lifetime of the coordinated H₂O molecule and the ²H NMR longitudinal relaxation rate of the deuterated diamagnetic lanthanum complex for the rotational correlation time were thoroughly investigated, and the results were compared with those reported previously for other lanthanide(III) complexes. The exchange lifetimes for $[en{Gd(DO3A)(H_2O)}_2]$ (769 ± 10 ns) and $[pi{Gd(DTTA)(H_2O)}_2]^2$ (910 ± 10 ns) are significantly higher than those of $[Gd(DOTA)(H_2O)]^-$ (243 ns) and $[Gd(DTPA)(H_2O)]^{2-}$ (303 ns) complexes. The rotational correlation times for $[en{Gd(DO3A)(H_2O)}_2]$ (150 ± 11 ps) and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ (130 ± 12 ps) are slightly greater than those of $[Gd(DOTA)(H_2O)]^-$ (77 ps) and $[Gd(DTPA)(H_2O)]^{2-}$ (58 ps) complexes. The marked increase in relaxivity (r_1) of $[en{Gd(DO3A)(H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ result mainly from their longer rotational correlation time and higher molecular weight. Copyright © 2004 John Wiley & Sons, Ltd.

KEYWORDS: NMR; EPR; ¹H NMR; ²H NMR; ¹⁷O NMR; Gd(III) complexes; paramagnetic complexes; proton relaxation

INTRODUCTION

Several lanthanide [e.g. Gd(III)] and transition metal [e.g. Mn(II) and Fe(III)] complexes of polyaminocarboxylates are either commercially available or in clinical trials for use as magnetic resonance imaging (MRI) contrast agents. The MR signal of body fluids can be altered by the presence of paramagnetic water relaxation agents to result in enhanced image contrast. The general design criteria for safe and efficacious MRI contrast agents have been reviewed by a number of investigators.^{1,2} However, they remain areas for further research and development of MRI contrast agents,

particularly those with enhanced relaxivity and improved tissue targeting functions. The main barrier to this endeavor is the low sensitivity of MRI contrast agents when they are coupled with the low flux of most biochemical processes. This problem can be addressed by developing contrast agents that possess higher spin–lattice relaxivity (r_1). The dimeric MRI contrast agents possess two gadolinium(III) ions and will increase relaxivity.

This has been confirmed for $H_6BO(DO3A)_2$,³ in which the Gd(III) complex incorporates several desirable features. As the ligand is a DOTA [DOTA = 1,4,7,10-tetraaza-1,4,7,10tetrakis(carboxymethyl)cyclododecane] derivative, we can expect high kinetic inertness and thermodynamic stability, and as a neutral complex it is preferable from the point of view of application (a less painful injection because of lower osmolality). The fact that two Gd³⁺ ions are bound to one molecule allows for smaller injection volumes for the same total amount in mmol Gd kg⁻¹ body weight. Finally,

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Scheme 1. Synthesis of en(DO3A)₂.

the increased molecular weight and volume may result in a longer rotational correlation time, and thus in a higher proton relaxivity.

To extend these observations, $[en(DO3A)_2]^{6-}$ and $[pi(DTTA)_2]^{8-}$, the derivatives of DO3A (1,4,7,10-tetraazacyclododecane-1,4,7-triaacetic acid) and DTPA [1,1,4,7,7-pentakis(carboxymethyl)-1,4,7-triazaheptane] were synthesized. The number of inner-sphere water molecules was determined from the ¹⁷O NMR chemical shift of the water as a function of Dy(III) concentration. The water proton spin–lattice relaxivity r_1 of the $[en{Gd(DO3A)(H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ complexes at various temperatures and pH values are described. The EPR and ¹⁷O NMR transverse relaxation rate data were analyzed together in a simultaneous multiple-parameter least-squares fitting procedure to determine the water residence lifetime. ²H NMR spectroscopy was used to determine the rotational correlation time.

EXPERIMENTAL

Materials

The acid forms of the free ligand, DO3A, were synthesized and characterized in accordance with the literature.⁴

Ethylenediaminediacetic acid dimethyl ester (1).

To a solution of EDDA (ethylenediaminediacetic acid, 28.41 mmol) in MeOH (150 ml) warmed to 40-50 °C was added SOCl₂ (8.24 ml, 113.64 mmol) dropwise slowly. After

20 h, solvent was removed from the fraction containing the product by rotary evaporation. The residue was dried under vacuum and a white powder was obtained (4.76 g, 80.21%). ¹H NMR (D₂O), δ (ppm): 3.48 (NCH₂CH₂N), 3.75 (NCH₂COOCH₃), 4.03 (NCH₂COOCH₃).

$en(DO3A)_2$ (2) (Scheme 1).

DO3A (1.27 g, 3.66 mmol) and anhydrous methanol (120 ml) were mixed in a single-necked flask and the pH of the solution was adjusted to 11.59 with NH₄OH. A water-bath was used to keep the temperature at 40-50 °C. Compound 1 (0.49 g, 2.4 mmol) was dissolved in anhydrous methanol (240 ml) and slowly added to a single-necked flask which contained DO3A and K₂CO₃ (10 g). After 48 h, the reaction solution was concentrated under reduced pressure to a pale yellow oil. The residue was dissolved in 50 ml of distilled water and made alkaline to pH 11.0 with ammonia solution. The solution was then applied to an AG1-X8 anion-exchange resin column (200-400 mesh, HCO2⁻ form, 60 ml of resin and 3.0 cm column diameter). After passing through an anion-exchange resin column, the product eluted in the 0.035 M formic acid fraction. Solvent was removed from the fraction containing the product by rotary evaporation and co-evaporated five times with 200 ml of water to remove the formic acid. The residue was dried in vacuum and a pale yellow hygroscopic powder was obtained (1.374 g, 88%) ¹³C NMR (50 MHz, D₂O) (ppm): 179.2, 178.0, 173.0, 166.7, 58.6, 57.4, 54.2, 53.1, 51.8, 50.8, 47.1, 46.8, 45.0, 42.6. Anal.





Scheme 2. Synthesis of pi(DTTA)₂.

Calculated for $C_{35}H_{64}N_{10}O_{15}$: C, 48.60; H, 7.46; N, 16.19. Found: C, 48.55; H, 7.40; N, 16.10%.

Diethylenetriamine-N'-acetic acid-N,N"-dianhydride (**3**). Pyridine (50 ml) and acetonitrile (50 ml) warmed to 50 °C were mixed with a solution of DTPA (50 mmol, 19.7 g) in acetic anhydride (318 mmol, 32.4 g). After 24 h, solvent was removed from the fraction and the residue was washed with acetic anhydride and diethyl ether. The solid was then dried under vacuum and a white powder was obtained (16.6 g, 92%). ¹H NMR (200 MHz, DMSO-*d*₆), δ (ppm): 3.71 (s, 8H, terminal NC*H*₂CO₂), 3.30 (s, 2H, central NC*H*₂CO₂), 2.72 (t, 4H, NC*H*₂CH₂N), 2.59 (t, 4H, NC*H*₂C*H*₂N).

$[pi(DTTA)_2]$ (4) (Scheme 2).

Compound **3** (2.3 g, 6.88 mmol), K_2CO_3 (10 g) and DMSO (100 ml) were mixed in a single-necked flask and *trans*-1,2-cyclohexanediamine (0.33 ml, 2.75 mmol) was slowly added. After 12 h, the solution was fractioned and filtered by ultrafiltration using ultrafiltration membranes YM3 (diameter 25 mm, MW cut-off = 3000) and YM1 (diameter 25 mm, MW cut-off = 1000). Solvent was removed by rotary evaporation. The residue was dried under vacuum and a pale-yellow oil was obtained (1.37 g, 43%).¹³C NMR (100 MHz, D₂O), δ (ppm): 172.91, 170.80, 166.81, 166.72, 56.64, 56.22, 54.40, 52.93, 52.44, 52.34, 53.03, 50.67, 50.38, 49.89, 31.28,

24.04. Anal. Calculated for C₃₄H₅₆N₈O₁₈: C, 47.22; H, 6.53; N, 12.96. Found: C, 47.16; H, 6.50; N, 12.64%.

General

GdCl₃·6H₂O (99.9%), DyCl₃·6H₂O (99.9%) and LaCl₃·7H₂O (99.9%) were obtained from Aldrich and used without further purification. The concentrations of Gd³⁺, Dy³⁺ and La³⁺ were determined by chelatometric titration with EDTA using xylenol orange as indicator. All other reagents used for the synthesis of the ligand were purchased from commercial sources unless noted otherwise. ¹H and ¹³C NMR spectra and elemental analyses were used to confirm the composition of the products. ¹⁷O-enriched water (20.1%) was purchased from Isotec.

Deuteration

The lanthanum complexes were synthesized by reaction of La_2O_3 with ligands in water and precipitated by addition of acetone. Deuteration of lanthanum complexes at the α -position with respect to carboxylate groups was performed using the procedure described by Wheeler and Legg.⁵ Deuteration was confirmed by ¹H NMR spectroscopy.

Complexation

The Dy(III) and Gd(III) complexes were prepared by mixing solutions of hydrated $LnCl_3$ (10 mM) and ligand (10 mM) in a

2:1 ratio. The pH was maintained at 7.50 with 1.0 M NaOH. Complex formation was instantaneous at room temperature. The solution was then evaporated under reduced pressure and the residue dried overnight at 60 °C.

Proton *T*¹ measurements

The samples were prepared by dissolving a measured amount of the Gd(III) chelates in water at pH 6.80 using the buffer solution (0.10 M) PIPES (PIPES = piperazine-N,N'-bis-2-ethanesulfonic acid)–NaOH. The buffer solution was used to maintain a constant ionic strength (i.e. 0.10M). The 0.10 M buffer was sufficient to maintain the solution pH at 6.80. The buffered Gd(III) chelate solutions were all allowed to equilibrate for at least 2 h. The pH of these solutions was determined immediately before relaxation time (T_1) measurements.

Relaxation times of aqueous solutions of gadolinium(III) complexes with $[en(DO3A)_2]^{6-}$ and $[pi(DTTA)_2]^{8-}$ were measured to determine the relaxivity r_1 . All measurements were made at 20 MHz as a function of temperature on a Bruker Minispec NMS-120 NMR spectrometer. The samples were contained in 5 mm glass tubes. The spectrometer was tuned and calibrated before each measurement. The values of T_1 were measured from eight data points generated by an inversion–recovery pulse sequence. The slope of plots of $1/T_1$ versus the concentration of Gd(III) complex gives r_1 in mM⁻¹ s⁻¹.

EPR measurements

EPR spectra were recorded at the X-band (0.34 T) using a Bruker ER 200D-SRC spectrometer operated in the continuous-wave mode. The samples were contained in the 1 mm glass tubes. The cavity temperature was stabilized using electronic temperature control of the gas flowing through the cavity. Temperature was verified by substituting a thermometer for the sample tube. Measurements were made from 273 to 363 K. The peak-to-peak linewidth was measured from the recorded spectra using the instrument's software.

¹⁷O NMR

The hydration numbers of $[en{Dy(DO3A)}_2]$ and $[pi{Dy(DTTA)}_2]^{2-}$ were determined using the method described by Alpoim *et al.*⁶ The ¹⁷O NMR spectra were recorded on a Varian **Gemini-400** spectrometer at 25 °C. Induced ¹⁷O shift (d.i.s: dysprosium(III) induced ¹⁷O NMR water chemical shift) measurements were made using D₂O as an external standard. Dy(III) chelate solutions were prepared by combining solutions of Dy(III) and ligand in a 2 : 1 ratio, and a stoichiometric amount of standardized NaOH was added so that the complex was fully formed. Six solutions of various dysprosium(III) concentrations were prepared by serial dilution of the stock solution.

Measurement of the ¹⁷O transverse relaxation rate was carried out with a Varian **Gemini-300** (7.05 T, 40.65 MHz) spectrometer, equipped with a 5 mm probe, by using an external D₂O lock. Experimental settings were spectral width 10 000 Hz, pulse width 7 μ s, acquisition time 10 ms and no sample spinning. A Varian VT-J103 temperature control





Figure 1. Dy(III)-induced water ¹⁷O NMR shift versus Dy(III) chelate concentration in D_2O at 25.0 ± 0.1 °C.

unit was used to stabilize the temperature. The value of the transverse relaxation rate was obtained by evaluating the linewidth at half-height ($\Delta v_{1/2}$) of the water ¹⁷O signal ($R_2 = \pi \Delta v_{1/2}$). Solutions containing 2.6% of the ¹⁷O isotope were used.

²H NMR

The rotational correction time values of $[en{La(DO3A)}_2]$ and $[pi{La(DTTA)}_2]^{2-}$ were determined by ²H NMR spectroscopy. The samples were prepared by dissolving the La³⁺ complexes in D₂O at pH 6.80. The measurement was carried out in a 10 mm o.d. tube on a Varian Gemini-400 (9.4 T) spectrometer equipped with a broadband probe and measured by a substitution technique as described elsewhere.⁵

RESULTS AND DISCUSSION

Dy(III)-induced water ¹⁷O NMR shifts

Figure 1 shows the Dy(III)-induced water ¹⁷O NMR shifts versus Dy(III) chelate concentration for solutions of DyCl₃, [en{Dy(DO3A)}₂] and [pi{Dy(DTTA)}₂]²⁻ in D₂O at 25 °C. The slopes obtained for [en{Dy(DO3A)}₂] and [pi{Dy(DTTA)}₂]²⁻ at pH 6.80 are -102.6 ppm mM⁻¹ ($r^2 = 0.9764$) and -107.8 ppm mM⁻¹ ($r^2 = 0.9762$). On the other hand, the slope for DyCl₃ is -382.8 ppm mM⁻¹ ($r^2 = 0.999$), and eight hydration numbers have been proposed for the dysprosium(III) ion.⁷⁻⁹ Therefore, [en{Dy(DO3A)}₂] and [pi{Dy(DTTA)}₂]²⁻ complexes contain 2.1 and 2.2 innersphere water molecules, respectively, at pH 6.80. The actual number of inner sphere water molecules coordinated to the metal center for [en{Dy(DO3A)}₂] and [pi{Dy(DTTA)}₂]²⁻ is one per Dy(III) ion. This result is similar to those for [pip{Gd(DO3A)(H₂O)]₂] and [bisoxa{Gd(DO3A)(H₂O)]₂].¹⁰



Relaxometric studies of the gadolinium(III) complexes

The longitudinal relaxivity r_1 values of $[en{Gd(DO3A) (H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ are 3.60 mm⁻¹s⁻¹ per Gd and 5.25 mm⁻¹s⁻¹ per Gd at pH 6.80, 37.0 ± 0.1 °C and 20 MHz, respectively. The r_1 value of $[en{Gd(DO3A)(H_2O)}_2]$ is significantly higher than those of $[Gd(DOTA)]^-$ (3.38 mm⁻¹s⁻¹ per Gd, 37.0 °C)¹¹ and $[BO{Gd(DO3A)(H_2O)}_2]$ (4.61 mm⁻¹s⁻¹ per Gd, 37.0 ± 0.1 °C)³ but lower than those of $[pip{Gd(DO3A)(H_2O)}_2]$ (5.8 mm⁻¹s⁻¹ per Gd, 40.0 °C)¹² and $[bisoxa{Gd(DO3A)(H_2O)}_2]$ (4.9 mm⁻¹s⁻¹ per Gd, 40.0 °C)¹² and $[bisoxa{Gd(DO3A)(H_2O)}_2]$ (4.9 mm⁻¹s⁻¹ per Gd, 40.0 °C)¹² and $[bisoxa{Gd(DO3A)(H_2O)}_2]^{2-}$ is significantly higher than that of the monomer $[Gd(DTPA)]^{2-}$.

The origin of paramagnetic relaxation enhancement is generally divided into two components, inner-sphere and outer-sphere:¹³

$$(1/T_i)_p = (1/T_i)_{inner-sphere} + (1/T_i)_{outer-sphere}$$
 $i = 1, 2$ (1)

Inner-sphere relaxation refers to relaxation enhancement of a solvent molecule directly coordinated to the paramagnetic ion, and outer-sphere relaxation refers to relaxation enhancement of solvent molecules in the second coordination sphere and beyond (i.e. bulk solvent). The inner-sphere relaxation contribution is obtained with the equation¹⁴

$$r_{1p}^{\rm is} = Cq / [55.6(T_{\rm 1M} + \tau_{\rm M})]$$
(2)

where *C* is the molar concentration of the gadolinium(III) complex, *q* is the number of water molecules bound to metal ion, T_{1M} is the longitudinal relaxation time of the bound water protons and τ_M^{298} is the residence lifetime of the bound water. Because of the inverse temperature dependence of T_{1M} and τ_M^{298} , two cases can be considered: (1) fast water-exchange $(T_{1M} \gg \tau_M)$, r_{1p}^{is} increases as temperature decreases; (2) slow water-exchange $(T_{1M} \ll \tau_M)$, r_{1p}^{is} decreases as temperature decrease of observed relaxivity with increasing temperature in the range 278–343 K. This is characteristic of fast chemical exchange behavior, occurring when the τ_M^{298} of the coordinated water molecule is much shorter than T_{1M} of the bound water proton. In fact, Eqn (3)¹⁵ can express T_{1M} :

$$\frac{1}{T_{1M}} = \frac{2}{15} \frac{\gamma_{\rm H}^2 g^2 S(S+1) \beta^2}{r_{\rm H}^6} \left(\frac{3\tau_{\rm C1}}{1+\omega_{\rm H}^2 \tau_{\rm C1}^2} + \frac{7\tau_{\rm C2}}{1+\omega_{\rm S}^2 \tau_{\rm C2}^2} \right) (3)$$
$$\frac{1}{\tau_{\rm Ci}} = \frac{1}{\tau_{\rm R}} + \frac{1}{\tau_{\rm M}} + \frac{1}{\tau_{\rm Si}} \tag{4}$$

where *S* is the electron spin quantum number (7/2 for Gd³⁺), $\gamma_{\rm H}$ is the proton nuclear magnetogyric ratio, β is the Bohr magneton, *g* is the Landé factor for the free electron, $r_{\rm H}$ is the distance between the metal ion and the bound water protons, $\omega_{\rm H}$ and $\omega_{\rm S}$ are the respective proton and electron Larmor frequencies and $\tau_{\rm Ci}$ (*i* = 1, 2) is the correlation time of the modulation of the dipolar electron-proton coupling. The overall correlation time $\tau_{\rm Ci}$ receives contributions from $\tau_{\rm M}^{298}$, $\tau_{\rm R}^{298}$ and $\tau_{\rm S}$ (the electronic relaxation time of the metal ion) [Eqn (4)]. To understand how $\tau_{\rm M}^{298}$ and $\tau_{\rm R}^{298}$ influence



Figure 2. Temperature dependence of the relaxivity for $[en{Gd(DO3A)(H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ at pH 6.80 and 20 MHz.

 r_1 of [en{Gd(DO3A)(H₂O)}₂] and [pi{Gd(DTTA)(H₂O)}₂]^{2–}, ¹⁷O and ²H NMR spectra were used to determine the values of τ_M and τ_R .

Water-exchange lifetime studies of Gd(III) complexes

The measured peak-to-peak line widths, ΔH_{pp} , of the derivative spectrum can be related to the overall transverse electronic relaxation rate, $1/T_{2e}$, vià Eqn (5), where g_L is the isotropic Landé *g* factor ($g_L = 2.0$ for Gd³⁺):¹⁶

$$\frac{1}{T_{2e}} = \frac{g_{\rm L}\mu_{\rm B}\pi\sqrt{3}}{h}\Delta H_{\rm pp} \tag{5}$$

The temperature dependence of transverse electronic relaxation rates at the X-band (0.34 T) at pH 6.80 for 50 mM solution of $[en{Gd(DO3A)(H_2O)}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ are shown in Figs 3 and 4. The data were fitted simultaneously with the following ¹⁷O NMR results. Analysis of the temperature dependence of the transverse relaxation rate for the ¹⁷O water nuclei is the most accurate method for evaluating the exchange lifetime of the water molecules directly coordinated to the metal in a paramagnetic Gd³⁺ chelate.¹⁷ According to the Swift and Connick theory,¹⁴ the paramagnetic contribution (R_{2p}^{O}) to the observed transverse relaxation rate is given by

$$R_{2p}^{O} = \frac{Cq}{55.6} (\tau_{M}^{O})^{-1} \frac{R_{2M}^{O^{2}} + (\tau_{M}^{O})^{-1} R_{2M}^{O} + \Delta \omega_{M}^{O^{2}}}{\left[R_{2M}^{O} + (\tau_{M}^{O})^{-1}\right]^{2} + \Delta \omega_{M}^{O^{2}}}$$
(6)



Figure 3. Temperature dependence of transverse electronic relaxation rates at the X-band (0.34 T) and pH 6.80 for a 50 mm solution of $[en{Gd(DO3A)(H_2O)}_2]$.



Figure 4. Temperature dependence of transverse electronic relaxation rates at the X-band (0.34 T) and pH 6.80 for a 50 mm solution of $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$.

where R_{2M}^{O} represents the ¹⁷O transverse relaxation rate of the coordinated water molecule and $\Delta \omega_{M}^{O}$ the chemical shift difference between the coordinated and bulk water ¹⁷O NMR resonances.



 R_{2M}^{O} is expressed by

$$R_{\rm 2M}^{\rm O} = \frac{1}{3} \left(\frac{A}{\hbar}\right)^2 S(S+1) \left(\tau_{\rm e1} + \frac{\tau_{\rm e2}}{1+\omega_{\rm s}^2 \tau_{\rm e2}^2}\right)$$
(7)

and

$$\tau_{ei}^{-1} = \tau_{\rm M}^{\rm O^{-1}} + T_{ie}^{-1} \tag{8}$$

where *S* is the electronic spin quantum number [7/2 for Gd(III)], A/\hbar is the Gd–¹⁷O scalar coupling constant and τ_{ei} (i = 1, 2) represents the correlation time of the processes modulating the scalar interaction. This modulation may occur through both the longitudinal and the transverse average electronic relaxation times (T_{1e} and T_{2e}) and the mean residence lifetime (τ_{M}^{O}) of the water molecule at the paramagnetic site.

The temperature dependence of R_{2M}^{O} is determined by the temperature effect on τ_{M}^{O} , τ_{v} (the correlation time for modulation of the zero field splitting interaction) and $\Delta \omega_{M}^{O}$ according to

$$(\tau_j)_T^{-1} = \frac{(\tau_j^{-1})^{298.15}T}{298.15} \exp\left[\frac{\Delta H_j}{R}\left(\frac{1}{298.15} - \frac{1}{T}\right)\right]$$
 (9)

$$\Delta\omega_{\rm M}^{\rm O} = \frac{g_{\rm L}\mu_{\rm B}S(S+1)B}{3k_{\rm B}T}\frac{A}{\hbar} \tag{10}$$

where the subscript *j* refers to the different correlation times, ΔH_j is the activation enthalpy for the corresponding dynamic process, *B* is the applied magnetic field strength and k_B is the Boltzmann constant.

The water-exchange rates for [en{Gd(DO3A)(H₂O)}₂] and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ were obtained by measuring the ¹⁷O NMR transverse relaxation rate (R_{2p}^{O}) as a function of temperature. The data and its best simulation according to Eqns $(5)-(10)^{16,18}$ are shown in Figs 5 and 6. As there are a large number of parameters to be determined in the quantitative analysis of the ¹⁷O NMR transverse relaxation rate (R_{2p}^{O}) versus T profiles, it is convenient to fix some of them. On this basis, in addition to the values of q and $A/\hbar(-3.8 \times 10^6 \text{ rad s}^{-1})$, the value of $\Delta H_{\rm M}$ is fixed at 30 kJ mol⁻¹.¹⁰ The parameters which provide the best fit of the data for $[en\{Gd(DO3A)(H_2O)\}_2]$ and $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ are listed in Table 1. By varying the temperatures over a wide range, R_{2p}^{O} is dominated by $1/\tau_{\rm M}$ in the slow kinetic region at low temperatures and is dominated by $1/\tau_{ei}$ in the fast kinetic region at high temperature.

As shown in Table 1, the water-exchange lifetime τ_{M}^{298} of $[en{Gd(DO3A)(H_2O)}_2]$ (769 ± 10 ns) is similar to those of $[pip{Gd(DO3A)(H_2O)}_2]$ (666 ns)¹⁰ and $[bisoxa{Gd(DO3A)(H_2O)}_2]$ (714 ns)¹² but higher than that of $[Gd(DOTA)(H_2O)]^{2-}$ (243 ns).¹⁸ The higher water-exchange lifetimes for $[en{Gd(DO3A)(H_2O)}_2]$, $[pip{Gd(DO3A)(H_2O)}_2]$ and $[bisoxa{Gd(DO3A)(H_2O)}_2]$ is perhaps due to the decreased number of carboxylate moieties which bind to the Gd(III) ion, so that the ligand is pulled less tightly around the metal center and is therefore less crowded around the water binding site.¹⁰ The water-exchange lifetime τ_{M}^{298} of $[pi{Gd(DTA)(H_2O)}_2]^{2-}$ (910 ± 10 ns) is significantly higher



Complex	$\Delta^2~(\mathrm{s}^{-2}\times 10^{20})$	${\tau_{\rm v}}^{298}~{ m (ps)}$	$\tau_{\rm M}^{298}$ (ms)	τ_R^{298} (ps)	$\Delta H (\mathrm{kJ}\mathrm{mol}^{-1})$
Gd ^{3+a}	1.19	7.3	1.2	41	15.3
$[Gd(DOTA)(H_2O)]^{-a}$	0.16	11	243	77	49.8
$[Gd(DTPA)(H_2O)]^{2-b}$	0.46	25	303	58	51.6
$[pip{Gd(DO3A)(H_2O)}_2]^b$	0.17 ± 0.01	19 ± 2	666	171 ± 12	34.2 ± 1.8
$[bisoxa{Gd(DO3A)(H_2O)}_2]^b$	0.21 ± 0.02	15 ± 1	714	106 ± 14	38.5 ± 1.8
$[en{Gd(DO3A)(H_2O)}_2]$	0.49 ± 0.02	17 ± 1	769 ± 10	105 ± 11	35.2 ± 1.0
$[pi\{Gd(DTTA)(H_2O)\}_2]^{2-}$	0.90 ± 0.01	14 ± 2	910 ± 10	130 ± 12	45.0 ± 1.8

Table 1. Kinetic and NMR parameters obtained from the simultaneous fit of ¹⁷O NMR and EPR data for Gd(III) complexes

^a Data from Ref. 11.

^b Data from Ref. 15.



Figure 5. Temperature dependence of the transverse water 17 O relaxation rate at 7.05 T and pH 6.80 for a 50 mM solution of [en{Gd (DO3A)(H₂O)}₂].

than that of $[Gd(DTPA)(H_2O)]^{2-}$ (303 ns).¹⁰ As described in the literature,¹⁹ a τ_M^{298} of 1000 ns creates a situation wherein the exchange rate of water is a significant limiting factor determining relaxivity (r_1) when r_1 increases due to the τ_R^{298} increase in these multidentate chelates, making a major contribution even for dimers. Therefore, the difference in r_1 is not only one between rigid and non-rigid linkages, but is also due to τ_R^{298} in the higher molecular weight molecule.¹⁸

Rotational correlation time studies of La(III) complexes

In diamagnetic molecules, the relaxation rate of the ²H nucleus is predominantly determined by the quadrupolar mechanism,⁵ which is strictly intramolecular and modulated by the sole rotation of the molecule. For fast-tumbling systems, the relaxation rate is thus directly related to the rotational correlation time (τ_R^{298}):

$$R_{1} = \frac{1}{T_{1}} = \frac{3}{8} \left(\frac{e^{2}qQ}{h}\right)^{2} \tau_{\rm R}$$
(11)

where the quadrupolar coupling constant (e^2qQ/h) depends on the hybridization state of the C-atom carrying the ²H



Figure 6. Temperature dependence of the transverse water ¹⁷O relaxation rate at 7.05 T and pH 6.80 for a 50 mM solution of $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$. The line represents the simultaneous least-squares fit to all data points as described in the text.

atom, its value being \sim 170 kHz in the case of an sp³ Catom. The measurement was performed on diamagnetic lanthanum(III) complexes deuterated in the α -position to the carboxylate groups. The values of τ_R^{298} for La(III) complexes with en(DO3A)₂⁶⁻, pip(DO3A)₂^{6-,12} bisoxa(DO3A)₂^{6-,11} [pi(DTTA)₂]⁸⁻, DTPA¹⁰ and DOTA¹¹ at 310 K are given in Table 1. The τ_R^{298} values for Gd(III) dimers are significantly higher than those for Gd(III) monomers, $[La((^{2}H_{10})DOTA)]^{-}$ $(77 \text{ ps})^{11}$ and $[La((^{2}H_{10})DTPA)]^{2-}$ (58 ps).¹¹ Thus, the change in the molecular weight in these complexes alters τ_R significantly. On the other hand, the lower rotational correlation time (τ_R^{298}) values for $[en\{La((^2H_{10})(DO3A))\}_2]$ and [bisoxa{La((²H₁₀)(DO3A))}₂] compared with that of $[pip{La((^{2}H_{10})(DO3A))}_{2}]^{18}$ indicates that the more flexible linker between the two macrocyclic chelating moieties in en(DO3A)₂⁶⁻ and bisoxa(DO3A)₂⁶⁻ decreases the $\tau_{\rm R}$ value and causes the lower relaxivity (r_1) of the Gd(III) complexes. The relationship between r_1 and the molecular weight of





Figure 7. Correlation of molecular weight of gadolinium(III) complexes with HP-DO3A (\blacksquare), PA-DO3A (\blacktriangle), B22F (\bullet), B22 ($_{O}$), TU1 (×), TU2 ($_{\Delta}$), en(DO3A)₂⁶⁻ (\Box) and [pi(DTTA)₂]⁸⁻ (+) with relaxivity.

Gd(III) monomer and dimer complexes¹⁹ is shown in Fig. 7. The results show that there is a strong correlation between r_1 and molecular weight, which means that the relaxivity value of the monomer generally increases with molecular weight. However, in order to maximize the relaxivity gain, the linking group and molecular weight must be taken into account for Gd(III) dimer complexes.

CONCLUSION

From analysis of the ¹⁷O NMR relaxometric properties, the larger water-exchange lifetime τ_M^{298} for [en{Gd(DO3A) (H_2O)] is perhaps due to a decrease in the number of carboxylate moieties, so that the ligand is pulled less tightly around the metal center and there is less crowding around the water binding site. We have demonstrated relaxivity $(r_1/\text{Gd m}M^{-1} \text{ s}^{-1})$ enhancement through the incorporation of rigidifying elements in the linkers or increasing the molecular weight of chelates. The water proton spin-lattice relaxivity of $[pi{Gd(DTTA)(H_2O)}_2]^{2-}$ is higher than that for the monomer $[Gd(DTPA)]^{2-}$ owing to its longer rotational correlation time and greater molecular weight. Furthermore, approaches aimed at enhancing relaxivity by modulating the water-exchange lifetime, $\tau_{\rm M}^{298}$, will be important for the future development of molecular MRI contrast agents used in imaging biochemical processes.

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REFERENCES

- 1. Reichert DE, Welch MJ. Coord. Chem. Rev. 2001; 212: 111.
- Tóth É, Burai L, Merbach AE. Coord. Chem. Rev. 2001; 216–217: 363.
- Tóth É, Vauthey S, Pubanz D, Merbach AE. *Inorg. Chem.* 1996; 35: 3375.
- Dischino DD, Delaney EJ, Emswiler JE, Gaughan GA, Prasad JS, Srivastava SK, Tweedle MF. *Inorg. Chem.* 1991; 30: 1265.
- 5. Wheeler WD, Legg JJ. Inorg. Chem. 1985; 24: 1292.
- 6. Alpoim MC, Urbano AM, Geraldes CFGC, Peters JA. J. Chem. Soc., Dalton Trans. 1992; 463.
- Kowall T, Foglia F, Helm L, Merbach AE. J. Am. Chem. Soc. 1995; 117: 3790.
- Cossy C, Helm L, Powell DH, Merbach AE. New J. Chem. 1995; 19: 27.
- Cossy C, Barnes AC, Enderby JE, Merbach AE. J. Chem. Phys. 1989; 90: 3254.
- Powell DH, Ni Dhubhghaill OM, Pubanz D, Helm L, Lebedev YS, Schlaepfer W, Merbach AE. J. Am. Chem. Soc. 1996; 118: 9333.
- 11. Micskei K, Helm L, Brücher E, Merbach AE. *Inorg. Chem.* 1993; 32: 3844.
- Carvalho J, Watson AD, Fellmann JD, Koo MD. US Patent 5 650 133, 1988.
- Caravan P, Ellison JJ, McMurry TJ, Lauffer RB. *Chem. Rev.* 1999; 99: 2293.
- 14. Swift TJ, Connick RE. J. Chem. Phys. 1962; 37: 307.
- Aime S, Gianolio E, Terreno E, Giovenzana GB, Pagliarin R, Sisti M, Palmisano G, Botta M, Lowe MP, Parker D. J. Biol. Inorg. Chem. 2000; 5: 488.
- 16. Reuben J. J. Phys. Chem. 1971; 75: 3164.
- Aime S, Botta M, Crich SG, Giovenzana G, Pagliarin R, Sisti M, Terreno E. Magn. Reson. Chem. 1998; 36: S200.
- Aime S, Botta M, Fasano M, Terreno E. Acc. Chem. Res. 1999; 32: 941.
- Ranganathan RS, Fernandez ME, Kang SI, Nunn AD, Ratsep PC, Pillai KMR, Zhang X, Tweedle MF. *Invest. Radio.* 1998; 11: 779.