### Accepted Manuscript

Surveying heterocycles as amide bioisosteres within a series of mGlu<sub>7</sub> NAMs: discovery of VU6019278

Carson W. Reed, Jordan P. Washecheck, Marc C. Quitlag, Matthew T. Jenkins, Alice L. Rodriguez, Darren W. Engers, Anna L. Blobaum, P. Jeffrey Conn, Colleen M. Niswender, Craig W. Lindsley

PII: DOI: Reference:	S0960-894X(19)30147-7 https://doi.org/10.1016/j.bmcl.2019.03.016 BMCL 26330
To appear in:	Bioorganic & Medicinal Chemistry Letters
Received Date:	31 January 2019

Revised Date:18 February 2019Accepted Date:13 March 2019



Please cite this article as: Reed, C.W., Washecheck, J.P., Quitlag, M.C., Jenkins, M.T., Rodriguez, A.L., Engers, D.W., Blobaum, A.L., Jeffrey Conn, P., Niswender, C.M., Lindsley, C.W., Surveying heterocycles as amide bioisosteres within a series of mGlu<sub>7</sub> NAMs: discovery of VU6019278, *Bioorganic & Medicinal Chemistry Letters* (2019), doi: https://doi.org/10.1016/j.bmcl.2019.03.016

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

#### **Graphical Abstract**

To create your abstract, type over the instructions in the template box below. Fonts or abstract dimensions should not be changed or altered. $9.0_{0}$ 





Bioorganic & Medicinal Chemistry Letters journal homepage: www.elsevier.com

# Surveying heterocycles as amide bioisosteres within a series of mGlu<sub>7</sub> NAMs: discovery of VU6019278

Carson W. Reed,<sup>a,c</sup> Jordan P. Washecheck,<sup>a,c</sup> Marc C. Quitlag,<sup>a,b</sup> Matthew T. Jenkins,<sup>a,b</sup> Alice L. Rodriguez,<sup>a,b</sup> Darren W. Engers,<sup>a,b</sup> Anna L. Blobaum,<sup>a,b</sup> P. Jeffrey Conn,<sup>a,b,e</sup> Colleen M. Niswender,<sup>a,b,e\*</sup> and Craig W. Lindsley<sup>a,b,c,d\*</sup>

<sup>a</sup>Vanderbilt Center for Neuroscience Drug Discovery, Vanderbilt University Medical Center, Nashville, TN 37232, USA <sup>b</sup>Department of Pharmacology, Vanderbilt University School of Medicine, Nashville, TN 37232, USA <sup>c</sup>Department of Chemistry, Vanderbilt University, Nashville, TN 37232, USA

<sup>d</sup>Department of Biochemistry, Vanderbilt University, Nashville, TN 37232, USA

eVanderbilt Kennedy Center, Vanderbilt University School of Medicine, Nashville, TN 37232, USA

\*To whom correspondence should be addressed: <u>colleen.m.niswender@vanderbilt.edu</u> or <u>craig.lindsley@vanderbilt.edu</u>

#### ARTICLE INFO

ABSTRACT

Article history:	
Received	
Revised	
Accepted	
Available online	
Keywords:	
mGlu <sub>7</sub>	
metabotropic glutamate receptor	
negative allosteric modulator (NAM)	
Structure-Activity Relationship (SAR)	
VU6019278	

This letter describes a diversity-oriented library approach to rapidly assess diverse heterocycles as bioisosteric replacements for a metabolically labile amide moiety within a series of mGlu<sub>7</sub> negative allosteric modulators (NAMs). SAR rapidly honed in on either a 1,2,4- or 1,3,4- oxadizaole ring system as an effective bioisostere for the amide. Further optimization of the southern region of the mGlu<sub>7</sub> NAM chemotype led to the discovery of VU6019278, a potent mGlu<sub>7</sub> NAM (IC<sub>50</sub> = 501 nM, 6.3% L-AP<sub>4</sub> Min) with favorable plasma protein binding (rat  $f_u$ = 0.10), low predicted hepatic clearance (rat CL<sub>hep</sub> = 27.7 mL/min/kg) and high CNS penetration (rat K<sub>p</sub> = 4.9, K<sub>p,uu</sub> = 0.65).

2019 Elsevier Ltd. All rights reserved.

Metabotropic glutamate receptor subtype 7 (mGlu<sub>7</sub>) is an attractive therapeutic target for a number of CNS disorders, including ADHD, schizophrenia, depression, Rett syndrome, epilepsy, anxiety and autism.<sup>1-13</sup> However, of the eight mGlu receptors (mGlu<sub>1-8</sub>), a lack of highly selective in vivo tool compounds has severely hindered the field from assessing the true therapeutic potential of activating or inhibiting mGlu<sub>7</sub> in the CNS.<sup>14,15</sup> In order to provide subtype selectivity amongst the mGlu family, efforts have largely focused on target modulation through allosteric mechanisms. To date, there are no selective mGlu<sub>7</sub> positive allosteric modulators (PAMs), but both pan-Group III PAMs and mGlu7-preferring PAMs have been reported.<sup>16</sup> The literature with mGlu<sub>7</sub> negative allosteric modulators (NAMs) is more mature (e.g. 1-5),<sup>17-22</sup> but only recently has a robust in vivo tool compound, 6 (VU6012962) been reported (Fig. 1).<sup>23</sup> Prior to the discovery of 6, poor physiochemical and DMPK properties precluded key in vivo proof of concept studies. Along the path to 6, NAM 5 was an important step,<sup>22</sup> demonstrating robust efficacy in native tissues (i.e., electrophysiology), but metabolic instability of the amide linker limited in vivo studies. Further optimization of 5 followed two parallel tracks: 1) functionalization of the eastern aryl ring to improve DMPK properties and 2) replacement of the metabolically labile amide linker with bioisosteres. Recently, we reported on the first approach, which led to the discovery of  $6^{23}$  Here, we will describe efforts examining a diverse array of small heterocycles to serve as replacement bioisosteres for the amide linker and the identification of an additional new tool compound and potential lead.



Figure 1. Structures of reported  $mGlu_7$  NAMs 1-5, and the key *in vivo* tool compound 6.

The optimization plan is depicted in **Figure 2**, where we envisioned surveying a diverse array of heterocycles as amide bioisosteres to afford analogs **7**. If a viable heterocyclic bioisostere could be identified, we would then optimize the substituents on the eastern aryl ring, and ensure that the southern 1,2,4-triazole remained the optimal moiety in the context of analogs **5**.<sup>22</sup>



**Figure 2.** Optimization plan for the labile amide containing **5**. First, heterocyclic bioisoteres for the amide in **5** would be evaluated, followed by optimization of the southern 1,2,4-triazole and the eastern 3,4-dimethoxy phenyl ring.

To survey a wide array of potential heterocyclic bioisosteres for the amide linker in **5**, we held the western (R<sub>1</sub>) and eastern (R<sub>2</sub>) moieties of **5** constant, and surveyed 13 heterocycles with varying hydrogen-bond donating and accepting capabilities in analogs **7** (**Fig. 3**). SAR was steep, with many inactive analogs (mGlu<sub>7</sub> IC<sub>50</sub> > 30  $\mu$ M, e..g, **7a**, **7j-m**), a number of weak NAMs (e.g., **7b-d**) and a few with modest activity (e.g., **7e-g**). Interestingly, a 1,3,4-oxadiazole analog **7h** proved to be an acceptable amide replacement (mGlu<sub>7</sub> IC<sub>50</sub> = 1.58  $\mu$ M, 7.1% L-AP<sub>4</sub> Min), as did a 1,2,4-oxadiazole **7i** (mGlu<sub>7</sub> IC<sub>50</sub> = 1.32  $\mu$ M, 14.5% L-AP<sub>4</sub> Min); moreover, both analogs were within two-fold of the parent **5**.<sup>22</sup> Prior to optimizing the R<sub>1</sub> and R<sub>2</sub> positions of these oxadiazole biosiosteres, we evaluated their *in vitro* DMPK properties to ensure further consideration was warranted.



Figure 3. Diversity library assessing a wide array of heterocycles 7 as

replacements for the amide linker in **5**. A 1,3,4-oxadiazole **7h** and a 1,2,4-oxadiazole **7i** stood out as viable replacements.

Both NAMs displayed favorable profiles, with predicted hepatic clearances ( $CL_{hep}$ ) in rat of 27.2 and 26.8 mL/min/kg for **7h** and **7i**, respectively, moderate plasma protein binding (rat  $f_{us}$  of 0.14 and 0.31 for **7h** and **7i**, respectively) and, in the case of **7h**, favorable rat brain homogenate binding ( $f_u = 0.022$ ).<sup>24</sup> Based on these data, further optimization was pursued around both **7h** and **7i**.

SAR around the 1,2,4-oxadiazole core was steep, with very few analogs displaying even modest mGlu<sub>7</sub> NAM potency, and no analogs were more potent than **7i**. Therefore, all attention shifted to a focus on optimization of the 1,3,4-oxadiazole **7h**. A rapid, four-step, microwave-assisted route to analogs **8** was developed (**Scheme 1**) to quickly assess diverse aryl ( $R_2$ ) moieties. Commercial esters **9** were treated with hydrazine under microwave conditions to provide the analogous acyl hydrazides **10** in 81-88% yields. In parallel, commercial 2-fluoro-5-(trifluoromethoxy)benzonitrile **11** undergoes an  $S_NAr$  reaction with 1*H*-1,2,4-triazole to deliver **12** in 67% yield, followed by hydrolysis of the nitrile to the acid **13**. Finally, condensation of acyl hydrazides **10** with acid **13** with T3P® under microwave conditions provided analogs **8**.

Scheme 1. Synthesis of putative mGlu<sub>7</sub> NAM analogs 8.<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a)  $H_2NNH_2$ · $H_2O$ , EtOH, 160 °C, mw, 1h, 81-88%; (b) 1*H*-1,2,4-triazole,  $K_2CO_3$ , DMF, 150 °C, mw, 30 min, 67%; (c) KOH, EtOH: $H_2O$  (1:1), 160 °C, mw, 30 min, 51%; (d) **10**, propylphosphonic anhydride (T3P®), Et<sub>3</sub>N, 150 °C, mw, 1h, 27-42%.

Table 1. Structure and mGlu<sub>7</sub> NAM activities of analogs 8.



C 1	TT /	01	01
Cmpa	Het	mGlu <sub>7</sub>	mGlu <sub>7</sub>
		$IC_{50} (\mu M)^{a}$	% L-AP4 Min
8a	OCH3	1.31	13.2
8b	OCH3	1.55	10.4
8c	,0,	>30	-

0.1		20	
8d		>30	-
	≥ √ − F		
8e	<u></u>	>30	-
8f	F, F	>30	-
	Е ССН.		
8g	OCH3	4.38	13.5
	Е → ОСН3		
	ʻ ∖= <sub>N</sub>		
8h	OCH3	>30	-
	₽ ₽ ₽		
8i	F	>30	-
	€-<->−OCH3		
8j	N N	>30	-
	}OCH3		
8k	,OCH₃	>10	68.9
	$\sim$	_	
	{ <b>−</b> ( <b>− N</b>		
81		>30	-
-	2 0011		
	F		
8m	OCH3	<u>&gt;10</u>	72.8
	}_осн₃		
	`		

<sup>a</sup>For SAR determination, calcium mobilization assays with rat  $mGlu_7/G_{\alpha15}/HEK$  cells performed in the presence of an EC<sub>80</sub> fixed concentration of glutamate; values represent (n = 1) independent experiments performed in triplicate.

As shown in Table 1, SAR amongst analogs 8 was very steep, affording few active mGlu7 NAMs. Of the very few active compounds, 8a (mGlu<sub>7</sub> IC<sub>50</sub> = 1.31  $\mu$ M, 13.2% L-AP<sub>4</sub> Min) was a 1,3,4-oxadiazole analog of 6, and a cyclobutyl ether congener 8b of 5 was also of comparable activity (mGlu<sub>7</sub> IC<sub>50</sub> = 1.55  $\mu$ M, 10.4% L-Alternate functionality and/or incorporation of a  $AP_4$  Min). heteroatom (8g, 8j and 8k) all led to inactive or a significant diminution in NAM activity. To compare with 7h, we evaluated the DMPK profile of 8a. NAM 8a displayed very low predicted hepatic clearance in rat ( $CL_{hep} = 2.9 \text{ mL/min/kg}$ ) in combination with favorable plasma protein binding ( $f_u = 0.038$ ). Moreover, a rat plasma:brain level (PBL) study demonstrated that 8a was CNS penetrant ( $K_p = 2.0$ ,  $K_{p,uu} = 0.43$ ). This positive disposition data supported further optimization efforts to enhance mGlu<sub>7</sub> NAM potency, while hopefully, maintaining an attractive DMPK profile.

Based on these data, we elected to hold the 3,4dimethoxyphenyl moiety of **7h** constant and surveyed saturated and unsaturated nitrogen-containing heterocycles as replacements for the 1*H*-1,2,4-triazole in *N*-linked analogs **14**. Once again, an  $S_NAr$  with **11**, and various saturated and unsaturated nitrogencontaining heterocycles **15**, provided *N*-linked derivatives **16**. Hydrolysis of the nitrile delivers acid **17**, which undergoes a T3P®-mediated microwave–assisted condensation with 3,4dimethoxybenzohydrazide to provide putative mGlu<sub>7</sub> NAMs **14**.

Scheme 2. Synthesis of putative mGlu<sub>7</sub> NAM analogs 14.<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) **11**, K<sub>2</sub>CO<sub>3</sub>, DMF, 150 °C, mw, 30 min, 54-72%; (b) KOH, EtOH:H<sub>2</sub>O (1:1), 160 °C, mw, 30 min, 38-51%; (c) 3,4-

dimethoxybenzohydrazide, propylphosphonic anhydride (T3P®), Et\_3N, 150  $^{\circ}C,$  mw, 1h, 22-62%.

Table 2. Structure and mGlu<sub>7</sub> NAM activities of analogs 14.



<sup>a</sup>For SAR determination, calcium mobilization assays with rat  $mGlu_7/G_{\alpha15}/HEK$  cells performed in the presence of an EC<sub>80</sub> fixed concentration of glutamate; values represent (n = 1) independent experiments performed in triplicate.

As shown in Table 2, analogs 14 proved more tractable towards producing mGlu7 NAMs. Saturated azacines, such as pyrrolidine (14a) lacked mGlu<sub>7</sub> NAM activity, but ring expansion to the piperidine derivative (14b) afforded a weak partial NAM, while a morpholine congener (14c) proved as potent (mGlu<sub>7</sub> IC<sub>50</sub> = 1.18  $\mu$ M, 23.1% L-AP<sub>4</sub> Min) as **7h**. In contrast, a basic piperazine analog was inactive. Replacement of the 1,2,4triazole with an imidazole ring (i.e., 14e, VU6019278) was highly successful, affording the most potent and efficacious mGlu<sub>7</sub> NAM (mGlu<sub>7</sub> IC<sub>50</sub> = 571 nM, 6.3% L-AP<sub>4</sub> Min) discovered during this campaign, and more potent than 5. When assayed in triplicate, the potency of 14e further improved (IC<sub>50</sub> = 501 nM,  $pIC_{50} = 6.30\pm0.11$ ,  $6.3\pm1.0$  L-AP<sub>4</sub> min). Moreover, NAM 14e displayed low predicted hepatic clearance in rat (CL<sub>hep</sub> = 27.7 mL/min/kg) in combination with favorable plasma protein binding  $(f_u = 0.10)$  and rat brain homogenate binding  $(f_u = 0.013)$ . Finally, a rat plasma:brain level (PBL) study demonstrated that 14e was highly CNS penetrant ( $K_p = 4.9$ ,  $K_{p,uu} = 0.65$ ), offering improvements over NAM 5.22 Finally, NAM 14e was also selective versus the other seven mGlu receptors (>10  $\mu M$  vs. mGlu<sub>1-6.8</sub>).

Since the *N*-linked southern azacines **14** proved attractive, we elected to explore the viability of C-linked heterocyclic congeners **18** as mGlu<sub>7</sub> NAMs. Analogs **18** were readily prepared in two steps from commercial benzoic acid **19** (Scheme **3**). A T3P®-mediated microwave–assisted condensation with 3,4-dimethoxybenzohydrazide and **19** delivers **20** in acceptable yield (43%). Next, a Suzuki coupling with various heteroaryl boronic acids provides putative mGlu<sub>7</sub> NAMs **18** in yields ranging from 36-60%.

Representative structures and activities for tricyclic analogs **18** are highlighted in **Table 3**. While not as productive as the *N*-

liked congeners **14**, interesting SAR resulted. C-linked 6membered heterocycles, both pyridines (**18a**,**b**) and a pyridazine (**18c**) were inactive. A 4-pyrazole derivative (**18d**) was a modest mGlu<sub>7</sub> NAM (mGlu<sub>7</sub> IC<sub>50</sub> = 2.24  $\mu$ M, 10.2% L-AP<sub>4</sub> Min), whereas the *N*-Me congener (**18e**) lost all mGlu<sub>7</sub> NAM activity. However, an *N*-Me, 5-pyrazole analog (**18f**), a regioisomer of inactive **18e**, proved to be the most potent mGlu<sub>7</sub> NAM in this subseries (mGlu<sub>7</sub> IC<sub>50</sub> = 1.32  $\mu$ M, 20.6% L-AP<sub>4</sub> Min).

Scheme 3. Synthesis of putative mGlu<sub>7</sub> NAM analogs 18.<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) 3,4-dimethoxybenzohydrazide, propylphosphonic anhydride (T3P®),  $Et_3N$ , 150 °C, mw, 1h, 43%; (b) R-B(OH)<sub>2</sub>, Cs<sub>2</sub>CO<sub>3</sub>, Pd(dppf)Cl<sub>2</sub>, 1,4-dioxane:H<sub>2</sub>O (9:1), 90 °C, 16h, 36-60%.

Table 3. Structure and mGlu<sub>7</sub> NAM activities of analogs 18.





Cmpd	R	mGlu <sub>7</sub>	mGlu <sub>7</sub>
		$IC_{50}~(\mu M)^a$	% L-AP <sub>4</sub> Min
18a	5000	>30	-
	N		
18b	~~~	<u>&gt;</u> 10	27.9
18c	~~~	<u>≥</u> 10	27.6
18d	N-NH	2.24	10.3
18e	N-N <sub>CH3</sub>	>30	-
18f	H <sub>3</sub> C <sub>N</sub>	1.32	20.6
18g	N S-	3.53	49.2

<sup>a</sup>For SAR determination, calcium mobilization assays with rat  $mGlu_7/G_{a15}/HEK$  cells performed in the presence of an EC<sub>80</sub> fixed concentration of glutamate; values represent (n = 1) independent experiments performed in triplicate.

In summary, we detailed a diversity-oriented library approach to rapidly assess diverse heterocycles as bioisosteric replacements for a metabolically labile amide moiety within a series of mGlu<sub>7</sub> negative allosteric modulators (NAMs), exemplified by **5**. SAR rapidly identified a 1,3,4-oxadiazole ring system as an effective bioisostere for the amide linker. Further optimization of the southern heterocycle of this new chemotype led to the discovery of VU6019278 (**14e**), a potent mGlu<sub>7</sub> NAM (IC<sub>50</sub> = 501 nM, 6.3% L-AP<sub>4</sub> Min) with favorable plasma protein binding (rat  $f_u$ = 0.10), low predicted hepatic clearance in microsomal incubations (rat  $CL_{hep} = 27.7 \text{ mL/min/kg}$ ) and high CNS penetration (rat  $K_p = 4.9$ ,  $K_{p,uu} = 0.65$ ). Not only was **14e** more potent than **5**, at the 0.2 mg/kg IV PBL dose, **14** achieve total brain levels of 456 nM, comparable to the *in vitro* IC<sub>50</sub> (501 nM), whereas **5** was relegated as an *in vitro* tool due to low total brain concentrations. However, free brain levels for **14e** from the PBL cassette study, based on rat brain homogenate binding, suggest ~6 nM free brain concentration at 0.2 mg/kg (if dose linear, doses of 10-30 mg/kg would afford free brain levels of **14e** above the *in vitro* IC<sub>50</sub>). NAM **14e** represented an improvement over the prototypical NAMs **1-5**, and argued for additional exploration of this subseries. Efforts in this vein are underway, and results will be reported in due course.

#### Acknowledgments

We thank the Warren Family and Foundation for establishing the William K. Warren, Jr. Chair in Medicine (C.W.L.). The authors also acknowledge funding by CDMRP Grant W81XWH-17-1-0266 (to C.M.N.).

#### AUTHOR INFORMATION

Corresponding Authors

\*C.M.N.: phone, 615-343-4303; fax, 615-343-3088; e-mail, colleen.niswender@vanderbilt.edu.

\*C.W.L.: phone, 615-322-8700; fax, 615-343-3088; e-mail, craig.lindsley@vanderbilt.edu.

ORCID 279 Craig W. Lindsley: 0000-0003-0168-1445

#### References

- N.M. Fisher, M. Seto, C.W. Lindsley, et al. Front. Mol. Neurosci. 11 (2018) article 387.
- (2) G. Sansig, T.J. Bushell, V.R. Clarke, et al. J. Neurosci. 21 (2001) 8734.
- (3) H. Goddyn, Z. Callaerts-Vegh, S. Stroobants, et al. Neurobiol. Learn. Mem. 90 (2008)103.
- (4) A. Palucha, K. Klak, P. Branski, et al. *Psychopharmacology* 194 (2007), 555.
- (5) Z. Callaerts-Vegh, T. Beckers, S.M. Ball, et al. J. Neurosci. 26 (2006) 6573.
- (6) K. Mitsukawa, C. Mombereau, E. Lötscher, E.; et al. *Neuropsychopharmacology* 31 (2006) 1112.
- (7) C. Hölscher, S. Schmid, P.K. Pilz, et al. Learn. Mem 12 (2005), 450.
- (8) C. Hölscher, S. Schmid, P.K. Pilz, et al. Behav. Brain Res. 154 (2004), 473.
- (9) T.J. Bushell, G. Sansig, V.J. Collett, et al. ScientificWorldJournal 2 (2002), 730.
- (10) M. Masugi, M. Yokoi, R. Shigemoto, et al. J. Neurosci. 19 (1999) 955.
- (11) N.M. Fisher, R.G. Gogliotti, S.A.D. Vermudez, et al. ACS Chem. Neurosci. 9 (2018) 2210.
- (12) R.G. Gogliotti, R.K. Senter, N.M. Fisher, et al. *Sci. Trans. Med.* 9 (2017) eaai7459.
- (13) N. Jalan-Sakrikar, J.R. Field, R. Klar, et al. ACS Chem. Neurosci. 5 (2014) 1221.
- (14) C.M. Niswender and P.J. Conn. Ann. Rev. Pharmacol. Toxicol. 50 (2010) 295.
- (15) C.W. Lindsley, K.A. Emmitte, C.R. Hopkins, et al. Chem. Rev. 116 (2016) 6707.
- (16) M. Abe, M. Seto, R.G. Gogliotti, et al. ACS Med. Chem. Lett. 8 (2017) 1110.
- (17) G. Suzuki, N. Tsukamoto, H. Fushiki, et al. J. Pharmacol. Exp. Ther. 232 (2007) 147.
- (18) M. Nakamura, H. Kurihara, G. Suzuki, et al. *Bioorg. Med. Chem. Lett.* 20 (2010) 726.
- (19) M. Kalinichev, M. Rouillier, F. Girard, et al. J. Pharmacol. Exp. Ther. 344 (2013) 624.

- (20) C.E. Gee, D. Peterlik, C. Neuhäuser J. Biol. Chem. 289 (2014) 10975.
- (21) C.M. Niswender, K.A. Johnson, N.R. Miller, et al. Mol. Pharmacol. 77 (2010) 77, 459.
- (22) C.W. Reed, K.M. McGowan, P.K. et al. ACS Med. Chem. Lett. 8 (2017) 1326.
- Accepting (23) C.W. Reed, S. E. Yohn, J.P. Washecheck, et al. J. Med. Chem. doi: 10.1021/acs.jmedchem.8b018810
- (24) J.M. Rook, J.L. Bertron, H.P. Cho, et al. ACS Chem. Neurosci. 9 (2018)

#### **HIGHLIGHTS**

- Novel series of selective and CNS penetrant mGlu7 NAMs
- Identified a 1,3,4-oxadiazole as • biosiostere for a labile amide linker
- Acception • Iterative libraries quickly optimized this