



Phosphorus, Sulfur, and Silicon and the Related Elements

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/gpss20

## Ultrasound-promoted, rapid and green synthesis of phosphonamide derivatives under catalyst and solvent-free conditions

Fouzia Bouchareb , Malika Berredjem , Abdeslem Bouzina & Meriem Guerfi

To cite this article: Fouzia Bouchareb , Malika Berredjem , Abdeslem Bouzina & Meriem Guerfi (2020): Ultrasound-promoted, rapid and green synthesis of phosphonamide derivatives under catalyst and solvent-free conditions, Phosphorus, Sulfur, and Silicon and the Related Elements, DOI: 10.1080/10426507.2020.1854254

To link to this article: <u>https://doi.org/10.1080/10426507.2020.1854254</u>



View supplementary material



Published online: 14 Dec 2020.

_	_
Г	
	11.
	<u> </u>
_	

Submit your article to this journal 🖸



View related articles



View Crossmark data 🗹

## Ultrasound-promoted, rapid and green synthesis of phosphonamide derivatives under catalyst and solvent-free conditions

Fouzia Bouchareb<sup>a,b</sup>, Malika Berredjem<sup>a</sup>, Abdeslem Bouzina<sup>a</sup>, and Meriem Guerfi<sup>a</sup>

<sup>a</sup>Laboratory of Applied Organic Chemistry, Synthesis of Biomolecules and Molecular Modelling Group, Faculty of Sciences, Department of Chemistry, Badji-Mokhtar – Annaba University, Annaba, Algeria; <sup>b</sup>Department of Chemistry, Faculty of Sciences and Technology, Chadli Bendjedid – EL Tarf University, El Tarf, Algeria

#### ABSTRACT

We report a rapid, efficient, economic, environmentally benign, and easy to scale-up method for the synthesis of phosphonamide derivatives using ultrasound irradiation, under catalyst and solvent-free conditions starting from the corresponding amine and phenyl phosphonic dichloride. The reaction was achieved in excellent isolated yield in a short reaction time at room temperature. The structures of the synthesized compounds are confirmed by elemental analysis as well as by IR and <sup>1</sup>H, <sup>13</sup>C, <sup>31</sup>P NMR spectroscopic data and mass spectrometry.

#### **ARTICLE HISTORY**

Received 4 August 2020 Accepted 18 November 2020

Taylor & Francis

Check for updates

Taylor & Francis Group

#### **KEYWORDS**

Green chemistry; phenylphosphonic dichloride; phosphonamide; ultrasound irradiation

#### **GRAPHICAL ABSTRACT**



## Introduction

The phosphonamide functional group is a key structural feature present in a number of biologically active natural products such as phosmidosine,<sup>[1]</sup> agrocin 84,<sup>[2]</sup> and microcin.<sup>[3]</sup> Compounds containing the phosphonamide moiety are important in biochemistry because a very large number of the compounds of biological interest belong to this family: DNA, RNA, and nucleotides.<sup>[4]</sup> Many applications have been described and some clinical trials are under development from a chemotherapy perspective as inhibitors of matrix metalloproteinases MMP-1, MMP-3, and MMP-9 1, 2,<sup>[5-8]</sup> inhibitors of tumor necrosis factor- $\alpha$  converting enzyme (TACE) **3**<sup>[9]</sup> (Figure 1).

Phosphonamides are important for the synthesis of various bioactive compounds serving as natural products, phosphates, phosphonopeptides, amino acids analogues,

CONTACT Malika Berredjem imberredjem@yahoo.fr imalika.berredjem@univ-annaba.org imalika.berredjem@univ-annab

© 2020 Taylor & Francis Group, LLC



Figure 1. Examples of bioactive molecules containing phosphonamide moiety.



Figure 2. Structures of phosphonamides prepared by the conventional method.



Scheme 1. Synthesis of phosphonamides under ultrasound irradiation.

prodrugs, pharmacological agents, and as important synthetic precursors.<sup>[10-13]</sup> Despite the importance of phosphonamides, their synthesis requires harsh conditions and/or the use of halides or precious transition metals.<sup>[14]</sup> The quest for green procedures has become of paramount importance for organic chemists in the last decade.<sup>[15]</sup> Discovery of benign, green and atom economic processes is one of the current corner stones for chemists.<sup>[16]</sup> One of the most promising ecofriendly processes is the use ultrasound irradiation to catalyze organic reactions.

One of the main objectives of our research group is the synthesis a new series of organophosphorus compound,<sup>[17-25]</sup> including phosphonamides.

In previous work, we have prepared a series of bisphosphonamides (8) (Figure 2) in good isolated yield by conventional method *via* a one-pot synthetic route, starting from corresponding primary amines with phenyl phosphonic dichloride in dry acetonitrile at  $-5 \,^{\circ}C$ .<sup>[26]</sup> We also have synthesized another derivative (6, 7) (Figure 2) from amines and aminoester or chloroacetamide with phenyl phosphonic dichloride in dry THF at  $0 \,^{\circ}C$  in tow steps.<sup>[27]</sup> Different process parameters such as temperature, solvent, catalyst type, and other factors are utilized by other researchers to prepare these compounds in excellent isolated yield.<sup>[28-35]</sup> But the traditional notion of yield is no longer sufficient to assess the efficiency of chemical processes.

Sustainable chemistry requires introducing new concepts that aim to reduce or eliminate the source of hazardous

substances, the use of renewable raw materials, and greater energy efficiency of products. With the awareness of the environmental impact of human activities and the emergence of the concept of sustainable development, chemists strive to put principles into practice by developing methods aimed at minimizing chance during synthesis, and have control over reactivity in order to avoid the production of undesirable compounds and thus limit the quantities of waste the concepts of "Green Chemistry".<sup>[36]</sup>

The objective of this work is to carry out the reactions under ultrasonic irradiation, without solvent. This powerful technique became extremely efficient and attractive in synthetic organic chemistry, and is able to activate many reactions due to cavitational collapse. it greatly contributes to the development of sustainable green chemistry, by responding to scientific challenges and current economic and environmental problems. Indeed, it is capable of inducing a myriad of chemical transformations.<sup>[37]</sup>

In continuation with our research on the synthesis of organophosphorus compounds, we report herein a simple, green, and efficient method for the preparation of these compounds under ultrasound irradiation techniques, solvent and catalyst-free conditions. This technique provides higher yields, shorter reaction times, and milder reaction conditions, nontoxic, environmentally friendly solvent, in a one-step reaction, without isolation of any intermediate thus reducing time, saving money, energy, and raw materials.<sup>[38–47]</sup>



4 🕞 F. BOUCHAREB ET AL.

### Table 1. Continued.



\*[26].

## **Results and discussion**

Herein we studied the synthesis of organophosphorus compounds under green chemical conditions using ultrasound

irradiation. The phosphonamides 8(a-r), were prepared in a one-pot synthetic route, starting from corresponding amine with phenyl phosphonic dichloride in the absence of any

solvent and any catalyst after 5–15 min (Table 1), the reaction was completed with an excellent yield.

Notably, no product was observed when the reaction was realized in the absence of ultrasound irradiation even after 48 h in the same conditions this shows the essential role of ultrasound irradiation. This excellent result encourages us to extend this study to various structurally primary and secondary amines (Scheme 1).

Spectrometric methods confirmed the structures of all synthesized phosphonamides; in the <sup>1</sup>H NMR, the formation of phosphonamide 8 g is confirmed by the appearance of a singlet at 2.40 ppm of the NH group, the phenyl bound to phosphorus atom appears between 7.90 and 7.50 ppm as multiplet. In infrared, we observed an absorption band toward  $3155 \text{ cm}^{-1}$  which corresponds to NH group. The bands of C=C and P=O groups appear, respectively, around 1461 and 1250 cm<sup>-1</sup>. <sup>31</sup>P NMR signals of 8 g were observed at 16 ppm.

## Experimental

#### Materials

The chemicals used in this work were obtained from Fluka and Merck Chemical Company and were used without purification.

#### **Apparatus**

Infrared (IR) spectra were recorded as KBr pellets on a Perkin-Elmer FT-600 spectrometer. Proton nuclear magnetic resonance (<sup>1</sup>H NMR) spectra were recorded in DMSO- $d_6$  or CDCl<sub>3</sub> solvents on a 250 or 300 MHz Bruker spectrometer with tetramethylsilane (TMS) as internal reference. Chemical shifts are reported in  $\delta$  units (ppm). All coupling constants (J) are reported in Hertz. Multiplicity is indicated as s (singlet), d (doublet), t (triplet), m (multiplet), and combination of these signals. Carbon nuclear magnetic resonance (<sup>13</sup>C NMR) spectra were recorded in DMSO- $d_6$  or CDCl<sub>3</sub> solvents at 60 or 100 MHz on a Bruker spectrometer with tetramethylsilane (TMS) as internal reference. Phosphorus nuclear magnetic resonance (<sup>31</sup>P NMR) spectra were recorded on a Brucker instrument at 100 or 121.5 MHz. Chemical Shift (ppm) Relative to 85% H<sub>3</sub>PO<sub>4</sub>. Mass spectra were recorded on a Shimadzu QP 1100 Ex mass spectrometer operating at an ionization potential of 70 eV. All reactions were monitored by thin-layer chromatography TLC on silica Merck h60 F<sub>254</sub> (Art. 5554) percolated aluminum plates and were developed by Spraying with ninhydrin solution. Ultrasound assisted reactions were carried out using a FUNGILAB ultrasonic bath with a frequency of 40 kHz and a nominal power of 250 W. The supplemental materials contain sample <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR and mass spectra for the products 8 (supplementary information, Figures S1-S46).

## Typical experimental procedure for the synthesis of phosphoramidates

In a 10 mL round bottom flask taken a mixture of phenylphosphonic dichloride (1 mmol) with amine (2 mmol) was added. Then reaction mixture was subjected to the ultrasonication for an appropriate time. After completion of the reaction, as indicated by TLC, silica gel; dichloromethane: methanol (9:1). Recrystallization of the crude product in ether afforded pure expected bisphosphonamides 8(a-r) as white solids.

## Bis (ethylamino) phenylphosphine oxide (8a)

Yield: 95%.  $R_f$ =0.65 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3435, 1512, 1200. <sup>1</sup>H NMR (DMSO, 300 MHz): 7.60 (m, 2H, H–Ar); 7.30 (m, 3H, H–Ar); 3.60 (s, 2H, NH); 2.75 (q, *J*=7.16 Hz, 4H, CH<sub>2</sub>); 1.15 (t, *J*=7.30 Hz, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (DMSO, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (DMSO, 100 MHz): 131.0; 130.9; 129.9; 128.2; 127.9; 127.8; 34.4; 12.9. MS ESI+ 30 eV *m/z*: 213.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>10</sub>H<sub>17</sub>N<sub>2</sub>OP] 212. Anal. Calcd for C<sub>10</sub>H<sub>17</sub>N<sub>2</sub>OP: C 56.59, H 8.07, N 13.20. Found: C 56.50, H 8.00, N 13.25%.

## Bis (propylamino) phenylphosphine oxide (8b)

Yield: 98%.  $R_f$ =0.61 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3193, 1465, 1200. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 2.90 (m, 4H, CH<sub>2</sub>–NH); 2.50 (m, 2H, NH); 1.50 (m, 4H, CH<sub>2</sub>–CH<sub>3</sub>); 0.90 (t,  $J_1$ =7.31 Hz,  $J_2$  =7.46 Hz, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 135.2; 131.5; 128.3; 128.3; 42.5; 25.3; 11.3. MS ESI+ 30 eV *m*/z: 241.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>12</sub>H<sub>21</sub>N<sub>2</sub>OP] 240. Anal. Calcd for C<sub>12</sub>H<sub>21</sub>N<sub>2</sub>OP: C 59.98, H 8.81, N 11.66. Found: C 59.90, H 8.75, N 11.70%.

## Bis (butylamino) phenylphosphine oxide (8c)

Yield: 97%.  $R_f$ =0.60 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3428, 1506, 1467, 1269. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.60 (m, 2H, H–Ar); 7.45 (m, 3H, H–Ar); 4.75 (m, 2H, NH); 3.20 (m, 4H, CH<sub>2</sub>–NH); 1.55 (m, 4H, CH<sub>2</sub>); 1.33 (m, 4H, CH<sub>2</sub>–CH<sub>3</sub>); 0.90 (t, *J*=8.0 Hz, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ =16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 135.0; 131.5; 129.6; 128.6; 47.4; 32.1; 20.0; 13.9. MS ESI<sup>+</sup> 30 eV *m/z*: 269.2 [M+H]<sup>+</sup> 100% calcd for [C<sub>14</sub>H<sub>25</sub>N<sub>2</sub>OP] 268. Anal. Calcd for C<sub>14</sub>H<sub>25</sub>N<sub>2</sub>OP: C 62.66, H 9.39, N 10.44. Found: C 62.62, H 9.30, N 10.50%.

### Bis (tert-butylamino) phenylphosphine oxide (8d)

Yield: 98%.  $R_f$ =0.61 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3228, 1436, 1221. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.80 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 2.00 (m, 2H, NH) 1.27 (s, 18H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 135.0; 131.5; 129.6; 128.6; 47.6; 29.1. MS ESI+ 30 eV *m/z*: 269.2 [M+H]<sup>+</sup> 100% calcd for

 $\label{eq:c14} \begin{array}{l} [C_{14}H_{25}N_2OP] \ 268. \ Anal. \ Calcd \ for \ C_{14}H_{25}N_2OP: \ C \ 62.66, \\ H \ 9.39, \ N \ 10.44. \ Found: \ C \ 62.62, \ H \ 9.30, \ N \ 10.50\%. \end{array}$ 

## Bis (piperidine) phenylphosphine oxide (8e)

Yield: 91%.  $R_f$ =0.62 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 1591, 1456, 1211. <sup>1</sup>H NMR (DMSO, 300 MHz): 7.60 (m, 2H, H–Ar); 7.30 (m, 3H, H–Ar); 2.95 (t, *J* = 10.05 Hz, 8H, CH<sub>2</sub>); 1.65 (m, 8H, CH<sub>2</sub>) 1.55 (m, 4H, CH<sub>2</sub>). <sup>31</sup>P NMR (DMSO, 100 MHz)  $\delta$  = 16.2 ppm. <sup>13</sup>C NMR (DMSO, 100 MHz): 131.0; 130.7; 129.9; 128.1; 127.9; 127.8; 43.9; 22.5; 22.1. MS ESI+ 30 eV *m/z*: 293.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>16</sub>H<sub>25</sub>N<sub>2</sub>OP] 292. Anal. Calcd for C<sub>16</sub>H<sub>25</sub>N<sub>2</sub>OP: C 65.73, H 8.62, N 9.58. Found: C 65.70, H 8.59, N 9.60%.

## Bis (morpholine) phenylphosphine oxide (8f)

Yield: 90%.  $R_f$ =0.68 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 1457, 1200. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.65 (m, 3H, H–Ar); 3.67 (t, *J*=8.45 Hz, 8H, **CH**<sub>2</sub>–O); 2.90 (t, *J*=7.30 Hz, 8H, **CH**<sub>2</sub>–N). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 135.0; 131.5; 129.6; 128.6; 72.8; 46.2. MS ESI+ 30 eV *m/z*: 297.2 [M+H]<sup>+</sup> 100% calcd for [C<sub>14</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P] 296. Anal. Calcs for C<sub>14</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P: C 56.75, H 7.14, N 9.45. Found: C 56.70, H 7.10, N 9.50%.

## Bis (cyclohexylamino) phenylphosphine oxide (8g)

Yield: 97%.  $R_f$ =0.62 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3155, 1461, 1250. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 3.15 (m, 2H, CH-cyc); 2.40 (m, 2H, NH); 1.90 (m, 4H, CH<sub>2</sub>-cyc); 1.65 (m, 4H, CH<sub>2</sub>-cyc); 1.15 (m, 12H, CH<sub>2</sub>-cyc). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.5 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 134.1; 132.4; 129.3; 55.2; 35.1; 25.2; 24.8. MS ESI+ 30 eV *m/z*: 321.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>18</sub>H<sub>29</sub>N<sub>2</sub>OP] 320. Anal. Calcd for C<sub>18</sub>H<sub>29</sub>N<sub>2</sub>OP: C 67.47, H 9.12, N 8.74. Found: C 67.50, H 9.20, N 8.70%.

## Bis (phenylamino) phenylphosphine oxide (8h)

Yield: 94%.  $R_f$ =0.67 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3223, 1599, 1499, 1293. <sup>1</sup>H NMR (DMSO, 300 MHz): 7.95 (m, 2H, H–Ar); 7.85 (m, 3H, H–Ar); 7.50 (m, 4H, H–Ar); 7.20 (m, 2H, H–Ar); 6.80 (m, 4H, H–Ar). <sup>31</sup>P NMR (DMSO, 100 MHz)  $\delta$  = 17.8 ppm. <sup>13</sup>C NMR (DMSO, 100 MHz): 142.6; 132.1; 132.0; 129.2; 128.9; 128.8; 120.7; 118.2; 118.1. MS ESI+ 30 eV *m/z*: 309.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>18</sub>H<sub>17</sub>N<sub>2</sub>OP] 308. Anal. Calcd for C<sub>18</sub>H<sub>17</sub>N<sub>2</sub>OP: C 70.12, H 5.56, N 9.09. Found: C 70.10, H 5.51, N 9.19%.

## Bis (3-fluorophenylamino) phenylphosphine oxide (8i)

Yield: 90%.  $R_f$ =0.67 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3415, 1529, 1494, 1279. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 6.99 (m, 2H, H–Ar); 6.33 (m,

2H, H–Ar); 6.23 (m, 2H, H–Ar); 6.17 (m, 2H, H–Ar); 4.00 (m, 2H, NH). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.1 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 162.9; 148.3;135.0; 131.5; 130.9; 129.6; 128.6; 110.7; 105.5. MS ESI+ 30 eV *m/z*: 345.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>18</sub>H<sub>15</sub> F<sub>2</sub> N<sub>2</sub>OP] 344. Anal. Calcd for C<sub>18</sub>H<sub>15</sub>F<sub>2</sub>N<sub>2</sub>OP: C 62.79, H 4.39, N 8.14. Found: C 62.75, H 4.35, N 8.24%.

## Bis (4-chlorophenylamino) phenylphosphine oxide (8j)

Yield: 92%.  $R_f$ =0.68 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3264, 1597, 1492, 1275. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 7.02 (m, 4H, H–Ar); 6.40 (m, 4H, H–Ar); 4.00 (m, 2H, NH). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.4 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 144.8; 135.0; 131.5; 129.7; 129.6; 128.6; 123.8; 116.5. MS ESI+ 30 eV *m*/*z*: 377.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>18</sub>H<sub>15</sub>Cl<sub>2</sub>N<sub>2</sub>OP] 376. Anal. Calcd for C<sub>18</sub>H<sub>15</sub>Cl<sub>2</sub>N<sub>2</sub>OP: C 57.32, H 4.01, N 7.43. Found: C 57.39, H 4.10, N 7.40%.

### Bis (4-methylphenylamino) phenylphosphine oxide (8k)

Yield: 90%.  $R_f$ =0.66 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3418, 1513,1436, 1279. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.95 (m, 2H, H–Ar); 7.55 (m, 3H, H–Ar); 6.81 (m, 4H, H–Ar); 6.34 (m, 4H, H–Ar); 4.00 (m, 2H, NH); 2.35 (s, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.4 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 143.7; 135.0; 131.5; 130.0; 129.6; 128.6; 127.7; 115.0; 20.9. MS ESI+ 30 eV *m/z*: 337.2 [M+H]<sup>+</sup> 100% calcd for [C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>OP] 336. Anal. Calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>OP: C 71.41, H 6.29, N 8.33. Found: C 71.35, H 6.20, N 8.39%.

## Bis (4-methoxyphenylamino) phenylphosphine oxide (81)

Yield: 95%.  $R_f=0.67$  (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3427, 1598, 1442, 1258. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 6.52 (m, 4H, H–Ar); 6.35 (m, 4H, H–Ar); 4.00 (m, 2H, NH) 3.73 (s, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 16.4$  ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 152.0; 139.0; 135.0; 131.5; 129.6; 128.6; 116.1; 114.9; 56.0. MS ESI+ 30 eV *m*/*z*: 369.2 [M+H]<sup>+</sup> 100% calcd for [C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P] 368. Anal. Calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P: C 65.21, H 5.75, N 7.60. Found: C 65.25, H 5.80, N 7.57%.

# *Bis (2-methoxyphenylamino) phenylphosphine oxide (8m)*

Yield: 92%.  $R_f$ =0.66 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3414, 1599, 1498, 1256. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.95 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 6.57 (m, 2H, H–Ar); 6.52 (m, 2H, H–Ar); 6.51 (m, 2H, H–Ar); 6.35 (m, 2H, H–Ar); 4.00 (m, 2H, NH) 3.73 (s, 6H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.4 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 148.6; 135.0; 132.3; 131.5; 129.6; 128.6; 121.6; 119.5; 116.1; 114.9; 56.0. MS ESI+ 30 eV *m*/*z*: 369.2 [M+H]<sup>+</sup> 100% calcd for [C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P]368. Anal. Calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>P: C 65.21, H 5.75, N 7.60. Found: C 65.25, H 5.80, N 7.57%.

## Bis (4-hydroxyphenylamino) phenylphosphine oxide (8n)

Yield: 93%.  $R_f$ =0.65 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3175, 1511, 1468, 1247. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.85 (m, 2H, H–Ar); 7.45 (m, 3H, H–Ar); 6.48 (m, 4H, H–Ar); 6.29 (m, 4H, H–Ar); 5.10 (s, 1H, OH); 4.00 (m, 2H, NH). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 147.3; 139.3; 135.0; 131.5; 129.6; 128.6; 116.5. MS ESI+ 30 eV *m*/*z*: 341.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>18</sub>H<sub>17</sub>N<sub>2</sub>O<sub>3</sub>P] 340. Anal. Calcd for C<sub>18</sub>H<sub>17</sub>N<sub>2</sub>O<sub>3</sub>P: C 63.53, H 5.04, N 8.23. Found: C 63.50, H 5.00, N 8.27%.

#### Bis (benzylamino) phenylphosphine oxide (80)

Yield: 95%.  $R_f$ =0.60 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 3163,1461, 1253. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.50 (m, 3H, H–Ar); 7.25 (m, 10H, H–Ar); 4.15 (m, 4H, CH<sub>2</sub>–N); 3.0 (m, 2H, NH). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 140.0; 139.9; 133.4; 131.9; 131.8; 131.7; 131.4; 128.7; 128.5; 127.7; 127.5; 127.3; 44.8. MS ESI+ 30 eV *m*/*z*: 337.2 [M + H]<sup>+</sup> 100% calcd for [C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>OP] 336. Anal. Calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub>OP: C 71.41, H 6.29, N 8.33. Found: C 71.47, H 6.33, N 8.30%.

## Bis (phenylethylamino) phenylphosphine oxide (8p)

Yield: 94%.  $R_f$ =0.63 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>1</sup>): 3209, 1450, 1191. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.75 (m, 2H, H–Ar); 7.40 (m, 3H, H-Ar); 7.35–7.00 (m, 10H, H–Ar); 4.60 (m, 1H,\*CH); 4.35 (m, 1H,\*CH); 2.75 (s, 2H, NH); 1.5 (d, J=6.25 Hz, 3H, CH<sub>3</sub>); 1.30 (d, J=6.77 Hz, 3H, CH<sub>3</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ =16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 145.2; 134.2; 132.1; 130.1; 128.8; 128.5; 126.9; 126.7; 55.4; 24.1. MS ESI+ 30 eV *m/z*: 365.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>OP] 364. Anal. Calcd for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>OP: C 72.51, H 6.91, N 7.69. Found: C 72.50, H 6.90, N 7.70%.

### Bis (diphenylamino) phenylphosphine oxide (8q)

Yield: 97%.  $R_f$ =0.69 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 1594, 1494, 1199. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.98 (m, 2H, H–Ar); 7.80 (m, 3H, H–Ar); 7.50 (m, 8H, H–Ar); 7.20 (m, 4H, H–Ar); 6.80 (m, 8H, H–Ar).<sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 142.6; 132.1; 132.0; 129.2; 128.9; 128.8; 120.7; 118.2; 118.1. MS ESI+ 30 eV *m/z*: 461.1 [M+H]<sup>+</sup> 100% calcd for [C<sub>30</sub>H<sub>25</sub>N<sub>2</sub>OP] 460. Anal. Calcd for C<sub>30</sub>H<sub>25</sub>N<sub>2</sub>OP: C 78.24, H 5.47, N 6.08. Found: C 78.20, H 5.40, N 6.10%.

### Bis (phenylpiperazine) phenylphosphine oxide (8r)

Yield: 89%.  $R_f$ =0.67 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH). IR (KBr, cm<sup>-1</sup>): 1512, 1405, 1222. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz): 7.90 (m, 2H, H–Ar); 7.60 (m, 3H, H–Ar); 7.08 (m, 4H, H–Ar); 6.60 (m, 2H, H–Ar); 6.59 (m, 4H, H–Ar); 3.47 (t, *J* = 6.85, 8H, CH<sub>2</sub>); 2.78 (t, *J* = 6.95, 8H, CH<sub>2</sub>). <sup>31</sup>P NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 16.3 ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): 144.5; 135.0; 131.5; 129.6; 129.4; 128.6; 118.0; 113.1; 58.9; 44.8. MS ESI+ 30 eV m/z: 447.1 [M + H] + 100% calcd for [C<sub>26</sub>H<sub>31</sub>N<sub>4</sub>OP] 446. Anal. Calcd for C<sub>26</sub>H<sub>31</sub>N<sub>4</sub>OP: C 69.94, H 7.00, N 12.55. Found: C 69.99, H 7.09, N 12.50%.

#### Conclusion

We have successfully developed a simple, efficient, and environmentally benign methodology toward the synthesis of phosphonamides under ultrasonic techniques, but solvent and catalyst-free conditions at room temperature. We achieved the synthesis of phenyl phosphonamides derived from amines with phenyl phosphonic dichloride in one step. This methodology was established with many advantages, including mild reaction conditions, short reaction times, excellent yields, simple work-up procedures.

#### **Disclosure statement**

No potential conflict of interest was reported by the author(s).

### Funding

This work was supported financially by The General Directorate for Scientific Research and Technological Development (DG-RSDT), Algerian Ministry of Scientific Research, Applied Organic Chemistry Laboratory (FNR 2000).

#### References

- Phillips, D. R.; Uramoto, M.; Isono, K.; McCloskey, A. Structure of the Antifungal Nucleotide Antibiotic Phosmidosine. J. Org. Chem. 1993, 58, 854–859. DOI: 10.1021/ jo00056a017.
- [2] (a) Roberts, W. P.; Tate, M. E.; Kerr, A. Agrocin 84 is a 6-N-Phosphoramidate of an Adenine Nucleotide Analogue. *Nature*. 1977, 265, 379–381. DOI: 10.1038/265379a0; (b) Tate, M. E.; Murphy, P. J.; Roberts, W. P.; Kerr, A. Adenine N<sup>6</sup>-Substituent of Agrocin 84 Determines its Bacteriocin-like Specificity. *Nature*. 1979, 280, 697–699. DOI: 10.1038/280697a0.
- [3] Guijarro, J. I.; González-Pastor, J. E.; Baleux, F.; Millán, J. L. S.; Castilla, M. A.; Rico, M.; Moreno, F.; Delepierre, M. Chemical Structure and Translation Inhibition Studies of the Antibiotic Microcin C7. J. Biol. Chem. 1995, 270, 23520–23532. DOI: 10. 1074/jbc.270.40.23520.
- [4] Dousson, C. B. Current and Future Use of Nucleo(s)Tide Prodrugs in the Treatment of Hepatitis C Virus Infection. *Antivir. Chem. Chemother.* 2018, 26, 1–8. DOI: 10.1177/ 2040206618756430.
- [5] Sørensen, M. D.; Blaehr, L. K. A.; Christensen, M. K.; Høyer, T.; Latini, S.; Hjarnaa, P.-J. V.; Björkling, F. Cyclic Phosphinamides and Phosphonamides, Novel Series of Potent Matrix Metalloproteinase Inhibitors with Antitumour Activity. *Bioorg. Med. Chem.* 2003, 11, 5461–5484. DOI: 10.1016/j.bmc. 2003.09.015.
- [6] Pikul, S.; McDow Dunham, K. L.; Almstead, N. G.; De, B.; Natchus, M. G.; Anastasio, M. V.; McPhail, S. J.; Snider, C. E.; Taiwo, Y. O.; Chen, L.; et al. Design and Synthesis of Phosphinamide-Based Hydroxamic Acids as Inhibitors of Matrix Metalloproteinases. *J. Med. Chem.* 1999, 42, 87–94. DOI: 10.1021/jm980142s.
- [7] Sawa, M.; Kiyoi, T.; Kurokawa, K.; Kumihara, H.; Yamamoto, M.; Miyasaka, T.; Ito, Y.; Hirayama, R.; Inoue, T.; Kirii, Y.; et al. New Type of Metalloproteinase Inhibitor: Design and

#### 8 🕞 F. BOUCHAREB ET AL.

Synthesis of New Phosphonamide-Based Hydroxamic Acids. J. Med. Chem. 2002, 45, 919–929. DOI: 10.1021/jm0103211.

- [8] Sawa, M.; Kondo, H.; Nishimura, S. Encounter with Unexpected Collagenase-1 Selective Inhibitor: Switchover of Inhibitor Binding Pocket Induced by Fluorine Atom. *Bioorg. Med. Chem. Lett.* 2002, *12*, 581–584. DOI: 10.1016/s0960-894x(01)00796-x.
- [9] Sawa, M.; Kurokawa, K.; Inoue, Y.; Kondo, H.; Yoshino, K. Discovery of Selective Phosphonamide-Based Inhibitors of Tumor Necrosis Factor-Alpha Converting Enzyme (TACE). *Bioorg. Med. Chem. Lett.* 2003, 13, 2021–2024. DOI: 10.1016/ s0960-894x(03)00292-0.
- [10] Hanson, P. R.; Stoianova, D. Ring Closing Metathesis Reactions on Phosphonamide and Phosphonate Templates. *Phosphorus Sulfur Silicon Relat. Elem.* 1999, 147, 107–108. DOI: 10.1080/ 10426509908053534.
- [11] Focken, T.; Hanessian, S. Application of Cyclic Phosphonamide Reagents in the Total Synthesis of Natural Products and Biologically Active Molecules. *Beilstein J. Org. Chem.* 2014, 10, 1848–1877. DOI: 10.3762/bjoc.10.195.
- [12] Hanessian, S. The Enterprise of Synthesis: From Concept to Practice. J. Org. Chem. 2012, 77, 6657–6688. DOI: 10.1021/ jo300902m.
- [13] Hanessian, S.; Delorme, D.; Beaudoin, S.; Leblanc, Y. Design and Reactivity of Topologically Unique, Chiral Phosphonamides. Remarkable Diastereofacial Selectivity in Asymmetric Olefination and Alkylation. J. Am. Chem. Soc. 1984, 106, 5754–5756. DOI: 10.1021/ja00331a070.
- [14] Du Bois, J. Rhodium-Catalyzed C-H Amination An Enabling Method for Chemical Synthesis. Org. Process Res. Dev. 2011, 15, 758–762. DOI: 10.1021/op200046v.
- [15] Clark, J. H. Green Chemistry: Today (and Tomorrow). Green Chem. 2006, 8, 17–21. DOI: 10.1039/B516637N.
- [16] Trost, B. The Atom Economy-a Search for Synthetic Efficiency. Science 1991, 254, 1471–1477. DOI: 10.1126/science.1962206.
- [17] Otmane Rachedi, K.; Tan-Sothea, O.; Bahadi, R.; Bouzina, A.; Djouad, S. E.; Bechlem, K.; Zerrouki, R.; Ben Hadda, T.; Almalki, F.; Berredjem, M. Synthesis, DFT and POM Analyses of Cytotoxicity Activity of α-Amidophosphonates Derivatives: Identification of Potential Antiviral O,O-Pharmacophore site. J. Mol. Struct. 2019, 1197, 196–203. DOI: 10.1016/j.molstruc.2019. 07.053.
- [18] Belhani, B.; Bechlem, K.; Grib, I.; Cheloufi, H.; Berredjem, M. A Green, One-Pot, Three-Component and Microwave Assisted Synthesis of α-Sulfamidophosphonates. J. Mater. Environ. Sci. 2018, 9, 613–618. DOI: 10.26872/jmes.2018.9.2.67.
- [19] (a) Bouzina, A.; Belhani, B.; Aouf, N. E. Berredjem, M. A Novel, Rapid and Green Method of Phosphorylation Under Ultrasound Irradiation and Catalyst Free Conditions. *RSC Adv.* **2015**, 5, 46272–46278. DOI: 10.1039/C5RA06380A;(b) Bouzina, A.; Berredjem, M.; Bouacida, S.; Merazig, H.; Aouf, N. E. A Greener Procedure for the Synthesis of Aureidophosphonates Under Ultrasound Irradiation. An X-ray Crystallographic Study. *RSC Adv.* **2015**, 5, 99775–99780. DOI: 10.1039/c5ra19886k.
- [20] Belhani, B.; Bouzina, A.; Berredjem, M.; Aouf, N. E. One-Pot Synthesis of Novel Oxazaphosphinanes under Ultrasound Irradiation and Solvent-Free Conditions. *Monatsh. Chem.* 2015, 146, 1871–1875. DOI: 10.1007/s00706-015-1461-4.
- [21] Bouzina, A.; Aouf, N. E.; Berredjem, M. Ultrasound Assisted Green Synthesis of  $\alpha$ -Hydroxyphosphonates under Solvent-Free Conditions. *Res. Chem. Intermed.* **2016**, *42*, 5993–6002. DOI: 10.1007/s11164-015-2420-8.
- [22] Belhani, B.; Berredjem, M.; Le Borgne, M.; Bouaziz, Z.; Lebreton, J.; Aouf, N. E. A One-Pot Three-Component Synthesis of Novel  $\alpha$ -Sulfamidophosphonates under Ultrasound Irradiation and Catalyst-Free Conditions. *RSC Adv.* **2015**, *5*, 39324–39329. DOI: 10.1039/C5RA03473F.
- [23] Boufas, W.; Cheloufi, H.; Bouchareb, F.; Berredjem, M.; Aouf, N. E. Convenient Synthesis of Novel N-Acylsulfonamides Containing Phosphonate Moiety. *Phosphorus Sulfur Silicon*

Relat. Elem. 2015, 190, 103-111. DOI: 10.1080/10426507.2014. 931398.

- [24] Saib, A.; Berrebbah, H.; Djebar, M. R.; Berredjem, M. Cytotoxic Study of Three Derivatives Amidophosphonates on Alternative Cellular Model: *Paramecium tetraurelia*. *Toxicol. Res.* 2014, 3, 395. DOI: 10.1039/C4TX00033A.
- [25] Hessainia, S.; Berredjem, M.; Ouarna, S.; Cheraiet, Z.; Aouf, N. E. Efficient Synthesis of Modified Sulfamides and Cyclosulfamides Containing Phosphonate Moieties. *Phosphorus Sulfur Silicon Relat. Elem.* **2013**, *188*, 719–725. DOI: 10.1080/ 10426507.2012.700356.
- [26] Bouchareb, F.; Hessainia, S.; Berredjem, M.; Benbouzid, H.; Djebbar, H.; Aouf, N. E. Efficient Method for the Synthesis of Diazaphospholidines: Toxicological Evaluation. *Asian J. Org. Chem.* 2012, *2*, 14–17. DOI: 10.5923/j.ajoc.20120201.03.
- Bouchareb, F.; Berredjem, M.; Aouf, N. E. Synthesis and Spectroscopic Study of New Substituted Phosphoramidates and 1, 3, 2-Diazaphospholidine-2,5-Diones. *Der PharmaChemica* 2015, 7, 90–94.
- [28] Tye, H.; Eldred, C.; Wills, M. An Efficient Method for the Synthesis of N,N'-Dimethyl-1,2-Diamines. *Tetrahedron Lett.* 2002, 43, 155–158. DOI: 10.1016/S0040-4039(01)02054-8.
- [29] Ali, H. M.; Mostafa, A. A.; El-Zohry, M. F. Synthesis and Bioactivity of O-Ethyl Phosphorodiamidates Derived from Quinazolin-4-Ones and Either Amino Acid Esters or Fatty Amines. *Heteroatom Chem.* **1999**, *10*, 455–460. DOI: 10.1002/ (SICI)1098-1071(1999)10:6<455::AID-HC3>3.0.CO;2-9.
- [30] Gholivand, K.; Della Vedova, C. O.; Erben, M. F.; Mahzouni, H. R.; Shariatinia, Z.; Amiri, S. Synthesis, Spectroscopic Study, X-Ray Crystallography and ab Initio Calculations of the Two New Phosphoramidates: C<sub>6</sub>H<sub>5</sub>OP(O)(NHC<sub>6</sub>H11)<sub>2</sub> and [N(CH<sub>3</sub>)(C<sub>6</sub>H<sub>11</sub>)]P(O)(2-C<sub>5</sub>H<sub>4</sub>N-NH)<sub>2</sub>. J. Mol. Struct. 2008, 874, 178–186. DOI: 10.1016/j.molstruc.2007.03.047.
- [31] Tomoskozi, I.; Gacs-Bait, E.; Otvos, L. Stereospecific Conversion of H-Phosphonates into Phosphoramidates. The Use of Vicinal Carbon-Phosphorus Couplings for Configurational Determination of Phosphorus. *Tetrahedron* 1995, 51, 6797–6804. DOI: 10.1016/0040-4020(95)00313-w.
- [32] Nikolaides, N.; Ganem, B. New Chemistry of Amines. 2. A Convenient Synthesis of Phosphotriesters from Phosphoramidates. *Tetrahedron Lett.* **1990**, *31*, 1113–1116. DOI: 10.1016/S0040-4039(00)88739-0.
- [33] Gholivand, K.; Hariatinia, Z.; Pourayoubi, M. Syntheses, Spectroscopic Characterization and Crystal Structures of Some New Phosphoramidates and an Organotin(IV) Complex of N-(4-Fluorobenzoyl)-N',N"-Bis(Piperidinyl)Phosphoric Triamide. Polyhedron 2006, 25, 711–721. DOI: 10.1016/j.poly.2005.07.035.
- [34] Gholivand, K.; Vedova, C. O. D.; Firooz, A. A.; Alizadehgan, A. M.; Michelini, M. C.; Diez, R. P. Pis Diezb, R. Syntheses, Crystal Structure and ab Initio Calculations of Two New Phosphoric Triamides. J. Mol. Struct. 2005, 750, 64–71. DOI: 10.1016/j.molstruc.2005.04.010.
- [35] Fu, N.; Zhang, Q.; Duan, L.; Xu, J. Facile Synthesis of Phosphonamidate- and Phosphonate-Linked Phosphonopeptides. J. Pept. Sci. 2006, 12, 303–309. DOI: 10.1002/psc.727.
- [36] Anastas, P. T.; Warner, J. C. Green Chemistry: Theory and Practice, Edition Oxford University Press: New York, 1998; pp 30.
- [37] Nasir Baig, R. B.; Varma, R. S. Alternative Energy Input: Mechanochemical, Microwave and Ultrasound-Assisted Organic Synthesis. *Chem. Soc. Rev.* 2012, 41, 1559–1584. DOI: 10.1039/ c1cs15204a.
- [38] Mansouri, R.; Aouf, Z.; Lakrout, S.; Berredjem, M.; Aouf, N. E. Greener, Efficient and Catalyst-Free Ultrasonic-Assisted Protocol for the N-Fmoc Protection of Amines. J. Braz. Chem. Soc. 2016, 27, 546–550. DOI: 10.5935/0103-5053.20150286.
- [39] Matveeva, E. V.; Odinets, I. L.; Kozlov, V. A.; Shaplov, A. S.; Mastryukova, T. A. Ionic-Liquid-Promoted Michaelis-Arbuzov Rearrangement. *Tetrahedron Lett.* 2006, 47, 7645–7648. DOI: 10.1016/j.tetlet.2006.08.050.

- [40] Dmitriev, M. E.; Ragulin, V. V. Arbuzov-Type Reaction of Acylphosphonites and N-Alkoxycarbonylimine Cations Generated in Situ with Trifluoroacetic Anhydride. *Tetrahedron Lett.* 2012, 53, 1634–1636. DOI: 10.1016/j.tetlet.2012.01.094.
- [41] Yang, G.; Shen, C.; Zhang, L.; Zhang, W. Nickel-Catalyzed Arbuzov Reactions of Aryl Triflates with Triethyl Phosphite. *Tetrahedron Lett.* 2011, 52, 5032–5035. DOI: 10.1016/j.tetlet. 2011.07.077.
- [42] Azizi, K.; Karimi, M.; Shaterian, H.-R.; Heydari, A. Ultrasound Irradiation for the Green Synthesis of Chromenes Using l-Arginine-Functionalized Magnetic Nanoparticles as a Recyclable Organocatalyst. RSC Adv. 2014, 4, 42220–42225. DOI: 10.1039/ C4RA06198E.
- [43] Dandia, A.; Gupta, S.; Parewa, V. An Efficient Ultrasound-Assisted One-Pot Chemoselective Synthesis of Pyrazolo[3,4-

b] Pyridine-5-Carbonitriles in Aqueous Medium Using NaCl as a Catalyst. *RSC Adv.* **2014**, *4*, 6908. DOI: 10.1039/c3ra47231k.

- [44] Jenck, J. F.; Agterberg, F.; Droescher, M. J. Products and Processes for a Sustainable Chemical Industry: A Review of Achievements and Prospects. *Green Chem.* 2004, 6, 544. DOI: 10.1039/b406854h.
- [45] Mason, T. J.; Peters, D. Practical Sonochemistry, Ellis Horwood: New York, 1991.
- [46] Luche, J. L. Synthetic Organic Sonochemistry, Plenum: New York, 1998; pp 167.
- [47] Mason, T. J. Sonochemistry and the Environment Providing a "Green" Link Between Chemistry, Physics and Engineering. *Ultrason. Sonochem.* 2007, 14, 476–483. DOI: 10.1016/j.ultsonch.2006.10.008.