



# Synthesis and SAR of a series of mGlu<sub>7</sub> NAMs based on an ethyl-8-methoxy-4-(4-phenylpiperazin-1-yl)quinoline carboxylate core

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## ARTICLE INFO

### Keywords:

mGlu<sub>7</sub>  
 Metabotropic glutamate receptor  
 Negative Allosteric modulator (NAM)  
 Structure-activity-relationship (SAR)

## ABSTRACT

A High-Throughput Screening (HTS) campaign identified a fundamentally new mGlu<sub>7</sub> NAM chemotype, based on an ethyl-8-methoxy-4-(4-phenylpiperazin-1-yl)quinolone carboxylate core. The initial hit, VU0226390, was a potent mGlu<sub>7</sub> NAM (IC<sub>50</sub> = 647 nM, 6% L-AP<sub>4</sub> min) with selectivity versus the other group III mGlu receptors (> 30  $\mu$ M vs. mGlu<sub>4</sub> and mGlu<sub>8</sub>). A multi-dimensional optimization effort surveyed all regions of this new chemotype, and found very steep SAR, reminiscent of allosteric modulators, and unexpected piperazine mimetics (whereas classical bioisosteres failed). While mGlu<sub>7</sub> NAM potency could be improved (IC<sub>50</sub>s ~ 350 nM), the necessity of the ethyl ester moiety and poor physiochemical and DMPK properties precluded optimization towards *in vivo* tool compounds or clinical candidates. Still, this hit-to-lead campaign afforded key medicinal chemistry insights and new opportunities.

Recently, metabotropic glutamate receptor subtype 7 (mGlu<sub>7</sub>) has garnered great interest as a viable target for a broad range of CNS disorders (e.g., PTSD, schizophrenia, depression, epilepsy, ADHD) and neurodevelopmental disorders (e.g., Rett syndrome and autism).<sup>1–15</sup> Compared to other mGlu receptors, mGlu<sub>7</sub> is one of the least explored and there is limited validation of its therapeutic potential due to a lack of selective, small molecule probes.<sup>16,17</sup> To study mGlu<sub>7</sub> activation, the field has no selective mGlu<sub>7</sub> positive allosteric modulators (PAMs), but must instead rely on mGlu<sub>7</sub>-preferring PAMs or *pan*-Group III PAMs in combination with selective negative allosteric modulators (NAMs) to isolate mGlu<sub>7</sub> activity.<sup>18</sup> NAMs are more advanced, with essentially four distinct chemotypes (1/2, 3, 4, 5–7, Fig. 1) of subtype selective ligands; however, first generation NAMs were electrophilic and suffered from poor disposition.<sup>19–24</sup> Recently new CNS-penetrant mGlu<sub>7</sub> NAMs, suitable as *in vivo* tool compounds (e.g. 6 and 7) have become available.<sup>25–27</sup>

Based on the pharmacological intricacies of allosteric modulators, we prefer to have multiple chemotypes to probe each mode of pharmacology. Here, we report on the discovery of a fundamentally new mGlu<sub>7</sub> NAM chemotype, based on an ethyl-8-methoxy-4-(4-

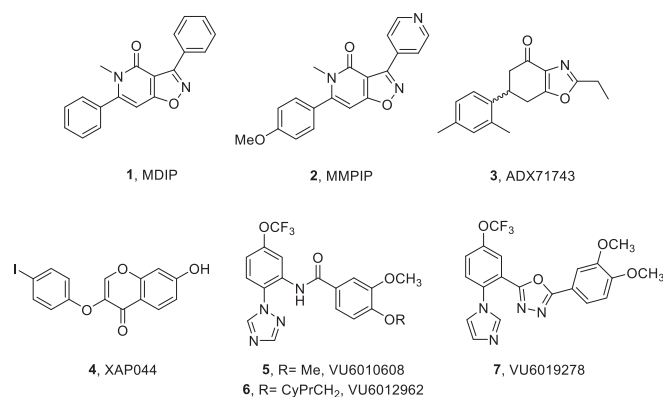
phenylpiperazin-1-yl)quinolone carboxylate core, derived from a functional HTS campaign. While SAR was steep, mGlu<sub>7</sub> potency could be improved, and key medicinal chemistry insights were gained with respect to non-obvious bioisosteres.

The structure of VU0226390 (8), based on an ethyl-8-methoxy-4-(4-phenylpiperazin-1-yl)quinolone carboxylate core, and the corresponding multi-dimensional optimization plan is depicted in Fig. 2. VU0226390 (8) was an attractive hit for several reasons: 1) it was a novel chemotype with selective sub-micromolar potency (mGlu<sub>7</sub> IC<sub>50</sub> = 647 nM, 6% L-AP<sub>4</sub> min, > 30  $\mu$ M vs. mGlu<sub>4</sub> and mGlu<sub>8</sub>), an acceptable DMPK profile ( $f_u$  (r,h) = 0.016, 0.018, CL<sub>hep</sub> = 60 mL/min/kg) and opportunities for a straightforward, modular synthetic plan. However, the ester moiety was a drawback as a potential metabolic liability, as was the inherent DMAP-like core structure of the piperazinyl quinoline. Thus, the optimization plan was to find replacements for these two concerning features, while also exploring alternatives for the distal 2-OMe phenyl ring and the 8-OMe quinolone core.

The synthetic route shown in Scheme 1, and variations thereof, was employed to access analogs 11, surveying multiple dimensions of the hit 8. Aniline 9 was heated with diethyl ethoxymethylenemalonate, in

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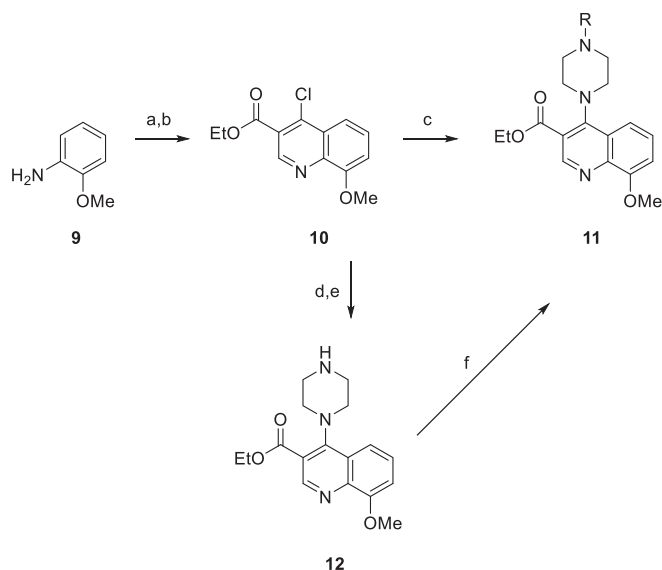


**Fig. 1.** Structures of reported mGlu<sub>7</sub> NAMs **1–7**, note the limited chemical diversity. NAMs **6** and **7** are robust, CNS penetrant *in vivo* tool compounds.

the absence of solvent to provide enamines, which were suspended in diphenyl ether and heated to 250 °C to promote cyclization to the corresponding quinolone in 31% yield. Standard conversion to the 4-chloroquinolone **10** proceeded under POCl<sub>3</sub> conditions in 78% yield. S<sub>N</sub>Ar with various aryl/heteroaryl piperazines afforded analogs **11** in yields ranging from 7 to 66%. Alternatively, *N*-Boc-piperazine could be employed in the S<sub>N</sub>Ar step, such that after deprotection, e.g. **12**, a wide range of aryl/heteroaryl moieties could be installed via Buchwald-Hartwig couplings to also access analogs **11** in good overall yields. The SAR for analogs **11** proved interesting.

The ethyl ester moiety of **8** emerged as a necessary pharmacophore for mGlu<sub>7</sub> NAM activity, and proved stable under our cell-based assay conditions. Replacement with a methyl ester lost ~6-fold in activity (IC<sub>50</sub> = 4 μM), but this was likely due to hydrolysis (the methyl ester partially hydrolyzed in cell buffer) to the corresponding acid, which was inactive (IC<sub>50</sub> > 30 μM). Similarly, the primary carboxamide congener, as well as diverse secondary and tertiary amides, all proved devoid of mGlu<sub>7</sub> NAM activity. Small alkyl, cycloalkyl and ether replacements for the ethyl ester were uniformly inactive. Thus, we elected to hold the ethyl ester constant, optimize activity elsewhere, and then return to find a more suitable replacement.

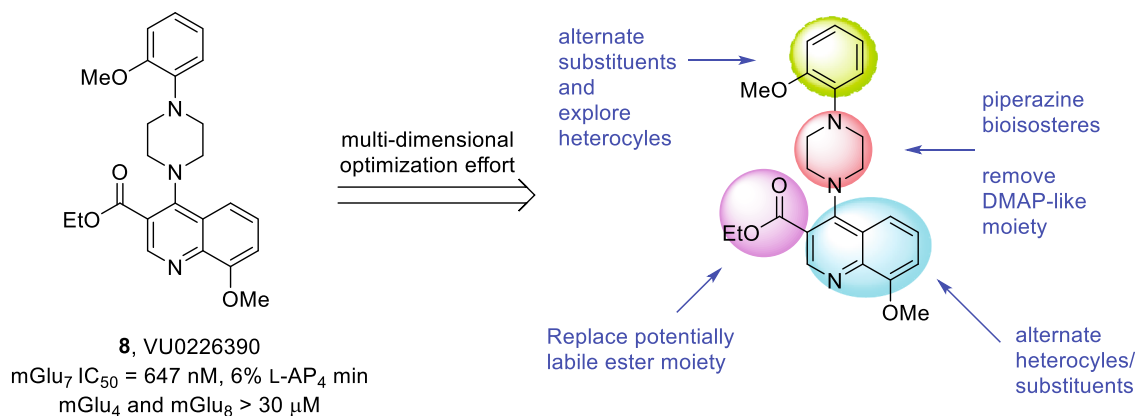
We surveyed a diverse array of substituted aryl and heteroaryl moieties to replace the 2-OMe phenyl moiety in **8** (Table 1) in an attempt to improve mGlu<sub>7</sub> NAM potency and/or disposition with analogs **11**. SAR was steep, with substitution only tolerated in the 2-position on the distal phenyl ring, e.g., 3-OMe and 4-OMe phenyl congeners were



**Scheme 1.** Synthesis of analogs **11** of the mGlu<sub>7</sub> NAM hit **8**<sup>a</sup>. <sup>a</sup>Reagents and conditions: (a) DEEMM, 140 °C, neat, 45 min, then Ph<sub>2</sub>O, 250 °C, 30 min, 31%; (b) POCl<sub>3</sub>, neat, 100 °C, 20 min, 78%; (c) aryl/heteroaryl piperazine, DIEA, DMF, 60 °C, 4 h, 7–66%; (d) *N*-Boc-piperazine, DIEA, DMF, 60 °C, 4 h, 74%; (e) 1:1 TFA:DCM, rt, 1 h, 98%; (f) Ar/Het-Br (I), Pd<sub>2</sub>dba<sub>3</sub>, P<sup>t</sup>Bu<sub>3</sub>, NaO<sup>t</sup>Bu, toluene, mw, 115 °C, 1 h, 4–60%.

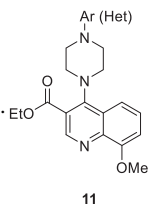
inactive. Ethers in the 2-position larger than methyl (**11a–c**) lost activity, as did 2-Cl (**11d**), 2-Me (**11e**), 2-CN (**11f**) and 2-F (**11h**) substitutions. From this survey, only the 2-SMePh derivative (**11g**) displayed improved potency (IC<sub>50</sub> = 510 nM) over **8**, with a slight improvement in rat predicted hepatic clearance (CL<sub>hep</sub> = 51.3 mL/min/kg). However, the corresponding sulfoxide and sulfone of **11g** proved devoid of mGlu<sub>7</sub> NAM activity. While di-OMe substituted analogs, such as **11i** and **11j**, retained mGlu<sub>7</sub> NAM potency, their disposition (as expected) was inferior to **8**. Direct heterocyclic congeners of **8**, such as the pyridine **11l** or pyrazine **11m** displayed modest potency. However, all 5-membered heterocyclic analogs (**11n–p**) proved inactive.

In parallel, we evaluated replacements for the quinolone core of **8** (Fig. 3). Truncation to a monocyclic pyridine, **13a**, resulted in a ~6.5-fold loss of potency, as did both regioisomeric thienopyridines **13b** and **13c**, highlighting the lack of bioisosterism with these substitutions.



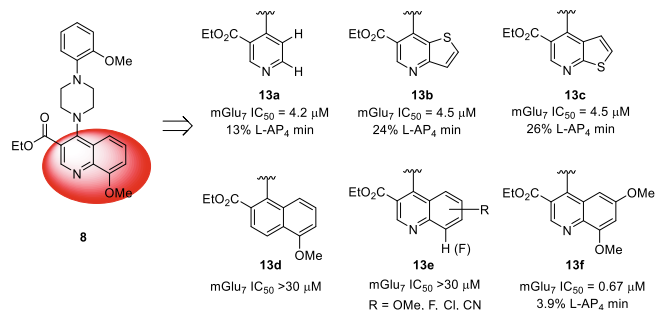
**Fig. 2.** Multi-dimensional optimization plan for the mGlu<sub>7</sub> NAM hit **8**, VU0226390.

Table 1

Structure and mGlu<sub>7</sub> NAM activities of analogs **11**.

Cmpd	Ar/Het	mGlu <sub>7</sub> IC <sub>50</sub> (μM) <sup>a</sup>	mGlu <sub>7</sub> % L-AP <sub>4</sub> Min
<b>8</b>	2-OMePh	0.65	5.6
<b>11a</b>	2-OEtPh	5.1	8.7
<b>11b</b>	2-O <sup>i</sup> PrPh	2.3	6.5
<b>11c</b>	2-OCF <sub>3</sub> Ph	> 30	—
<b>11d</b>	2-ClPh	> 30	—
<b>11e</b>	2-MePh	> 10	41
<b>11f</b>	2-CNPh	4.2	14
<b>11g</b>	2-SMePh	0.51	4.7
<b>11h</b>	2-FPh	6.1	12
<b>11i</b>	2,4-diOMePh	1.9	9.0
<b>11j</b>	2,6-diOMePh	0.80	11
<b>11k</b>	2-OMe-5-FPh	2.1	15
<b>11l</b>		1.5	10
<b>11m</b>		3.0	11
<b>11n</b>		> 30	—
<b>11o</b>		> 30	—
<b>11p</b>		> 30	—

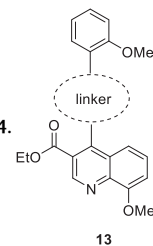
<sup>a</sup>For SAR determination, calcium mobilization assays with rat mGlu<sub>7</sub>/G<sub>α15</sub>/HEK cells were performed in the presence of a fixed EC<sub>80</sub> concentration of L-AP<sub>4</sub>; values represent (n = 1) independent experiments performed in triplicate.<sup>25</sup>

Fig. 3. SAR efforts to replace the quinolone core of mGlu<sub>7</sub> NAM **8**.

Replacement with a naphthalene (**13d**) proved inactive, as did replacement of the 8-OMe moiety with either a hydrogen or fluoride atom (**13e**). Here, only a 6,8-diOMe congener, **13f**, proved equipotent to **8**, but once again displayed a poor disposition profile. Thus, moving forward, we would maintain the 8-OMe quinolone core and the 2-OMe phenyl moiety of **8** as we surveyed the piperazine linker (Table 2).

SAR around the piperazine core of **8** (Table 2), with analogs **13**, proved interesting. The addition of a racemic methyl group to either the 3-position (**14a**) or the 2-position (**14b**), led to an ~2-fold loss in mGlu<sub>7</sub> NAM potency. Cleavage of the piperazine ring, as in **14c**, lost ~6-fold, while deletion of the proximal piperazine nitrogen proved inactive.

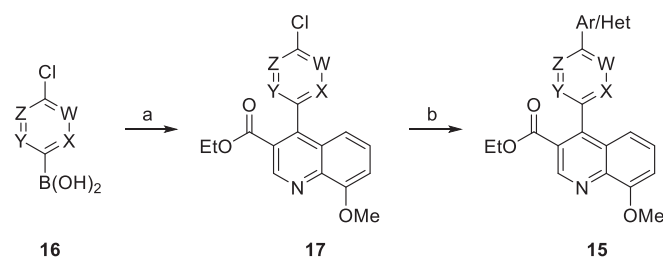
Table 2

Structure and mGlu<sub>7</sub> NAM activities of analogs **14**.

Cmpd	Linker	mGlu <sub>7</sub> IC <sub>50</sub> (μM) <sup>a</sup>	mGlu <sub>7</sub> % L-AP <sub>4</sub> Min
<b>8</b>		0.65	5.6
<b>14a</b>		1.3	8.6
<b>14b</b>		1.5	6.1
<b>14c</b>		4.4	9.6
<b>14d</b>		> 10	41
<b>14e</b>		3.6	9.6
<b>14f</b>		> 30	—
<b>14g</b>		> 10	41
<b>14h</b>		> 10	57
<b>14i</b>		> 30	—
<b>14j</b>		2.9	4.2

<sup>a</sup>For SAR determination, calcium mobilization assays with rat mGlu<sub>7</sub>/G<sub>α15</sub>/HEK cells were performed in the presence of a fixed EC<sub>80</sub> concentration of glutamate; values represent (n = 1) independent experiments performed in triplicate.<sup>25</sup>

Incorporating unsaturation, as with **14e**, regained potency (mGlu<sub>7</sub> IC<sub>50</sub> = 3.5 μM), while cyclic constraints, e.g. **14f** and **14g**, proved inactive. Classical piperazine bioisosteres, such as [3.3.0] system (**14h**)



**Scheme 2.** Synthesis of heteroaryl linker analogs **15**<sup>a</sup>. <sup>a</sup>Reagents and conditions: (a) **10**, 10 mol% Pd(dppf)Cl<sub>2</sub>, Cs<sub>2</sub>CO<sub>3</sub>, 4:1 dioxane:water, mw, 110 °C, 30 min, 45–57%; (b) Ar/Het-B(OH)<sub>2</sub>, 10 mol% Pd(dppf)Cl<sub>2</sub>, Cs<sub>2</sub>CO<sub>3</sub>, 4:1 dioxane:water, mw, 110 °C, 30 min, 25–69%.

and the spiroazetidone (**14i**), were not productive in this context. Surprisingly, a biphenyl monomer we had ‘in house’ proved intriguing and demonstrated that a simple phenyl ring (**14j**) could replace the piperazine linker with reasonable potency (IC<sub>50</sub> = 2.9 μM). However, the DMPK profile of **14j** was poor ( $f_u$  (r, h) < 0.001, CL<sub>hep</sub> (rat) = 66 mL/min/kg), leading the team to consider replacement of the phenyl ring with polar/basic heterocycles, e.g., analogs **15**. Analogs **15** were readily prepared according to Scheme 2. Utilizing commercial heteroaryl chloro-boronic acids **16**, in a chemoselective Suzuki coupling with intermediate **10**, provided derivatives **17** in yields ranging from 45 to 57%. A second Suzuki coupling with aryl boronic acids delivered final compounds **15** in good overall yields. As shown in Table 3, SAR within this sub-series was more robust than in the preceding saturated systems. Replacement of the phenyl linker with a 6-pyridyl congener, **15a**, proved to be equipotent (IC<sub>50</sub> = 2.8 μM), but the 5-pyridyl regioisomer, **15b**, was a potent and efficacious mGlu<sub>7</sub> NAM (IC<sub>50</sub> = 0.60 μM, 6.5% L-AP<sub>4</sub> min). A slight increase in potency (IC<sub>50</sub> = 0.37 μM, 6.3% L-AP<sub>4</sub> min and IC<sub>50</sub> = 0.41 μM, 5.6% L-AP<sub>4</sub> min) was noted with both the 2-OEtPh analog (**15c**) and the 2-SMePh derivative (**15d**), respectively. A pyrimidine congener lost (**15f**) activity (IC<sub>50</sub> = 2.4 μM, 12.0% L-AP<sub>4</sub> min) while a pyrazine maintained good mGlu<sub>7</sub> NAM activity (IC<sub>50</sub> = 0.54 μM, 5.1% L-AP<sub>4</sub> min). The addition of a pendant methoxy moiety to the pyridine ring of **15b**, affording **15h**, led to a more potent mGlu<sub>7</sub> NAM (IC<sub>50</sub> = 0.35 μM, 3.6% L-AP<sub>4</sub> min). Even 5-membered heterocycles, such as pyrrole **15j**, was a competent surrogate for the phenyl linker. Finally, as with **13f**, the incorporation of an additional methoxy group in the 6-position of the quinoline ring system, e.g., **18** and **19** (Fig. 4), led to sub-micromolar mGlu<sub>7</sub> NAMs.

While none of these new mGlu<sub>7</sub> NAMs possessed *in vitro* or *in vivo* DMPK profiles (e.g. **15h**, low fraction unbound:  $f_u$  (rat, mouse) = 0.01, 0.01, BHB  $f_u$  (rat, mouse) = 0.04, 0.04, high predicted hepatic clearance (CL<sub>hep</sub> (rat, mouse) = 65.2 and 66.1 mL/min/kg) and high clearance *in vivo*, CL<sub>p</sub> = 70 mL/min/kg) worthy of advancement as *in vivo* tools, they are unique *in vitro* pharmacological tools with selectivity against the other mGlu receptors. Upon closer inspection of analogs **15**, these resemble classical terphenyl α-helical mimetic protein–protein inhibitor (PPI) chemotype **20** (Fig. 5), with clear overlap of the prototypical i, i + 3/4 and i + 7 moieties in **15h**.<sup>28,29</sup> As **15h** is structurally distinct from previously reported mGlu<sub>7</sub> NAMs, and due to the resemblance to α-helical mimetic protein–protein inhibitor (PPI) chemotypes, the exciting prospect exists that this family of NAMs has a unique mode of pharmacological inhibition of mGlu<sub>7</sub>.

In summary, we have reported on the multi-dimensional optimization of a fundamentally new mGlu<sub>7</sub> NAM chemotype, based on an ethyl-8-methoxy-4-(4-phenylpiperazin-1-yl)quinoline carboxylate core; moreover, we found that aromatic heterocycles, such as pyridines, are an unlikely bioisostere for the central piperazine motif. SAR led to the discovery of a unique terphenyl chemotype, reminiscent of α-helical

**Table 3**

Structure and mGlu<sub>7</sub> NAM activities of analogs **15**.

Compd	Ar/Het	R	15	
			mGlu <sub>7</sub> IC <sub>50</sub> (μM) <sup>a</sup>	mGlu <sub>7</sub> % L-AP <sub>4</sub> Min
<b>15a</b>		2-OMe	2.8	6.8
<b>15b</b>		2-OMe	0.60	6.5
<b>15c</b>		2-OEt	0.37	6.3
<b>15d</b>		2-SMe	0.41	5.6
<b>15e</b>		2-O'Pr	1.5	5.8
<b>15f</b>		2-OMe	2.4	12
<b>15g</b>		2-OMe	0.54	5.1
<b>15h</b>		2-OMe	0.35	3.6
<b>15i</b>		2-OMe	2.1	24
<b>15j</b>		2-OMe	1.2	6.5

<sup>a</sup>For SAR determination, calcium mobilization assays with rat mGlu<sub>7</sub>/G<sub>α15</sub>/HEK cells were performed in the presence of a fixed EC<sub>80</sub> concentration of glutamate; values represent (n = 1) independent experiments performed in triplicate.<sup>25</sup>

mimetic PPIs, a first in the mGlu allosteric ligand world. Studies are underway to evaluate the mechanism of inhibition of mGlu<sub>7</sub> by **15h** in comparison to chemically distinct mGlu<sub>7</sub> NAM **6**. These more laborious studies are underway and will be reported in due course.

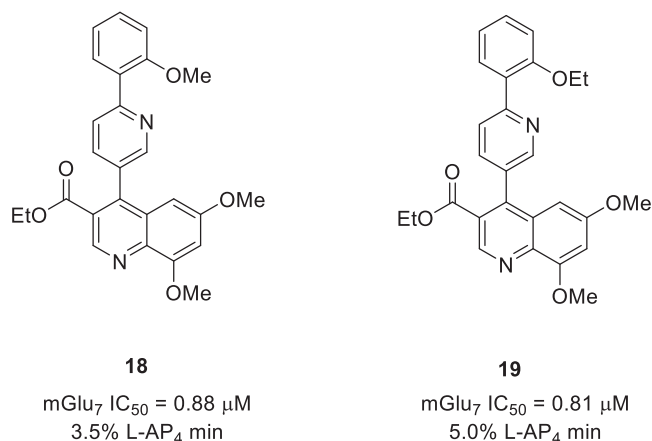


Fig. 4. 6,8-DiOMe substituted terphenyl mGlu<sub>7</sub> NAMs **18** and **19**.

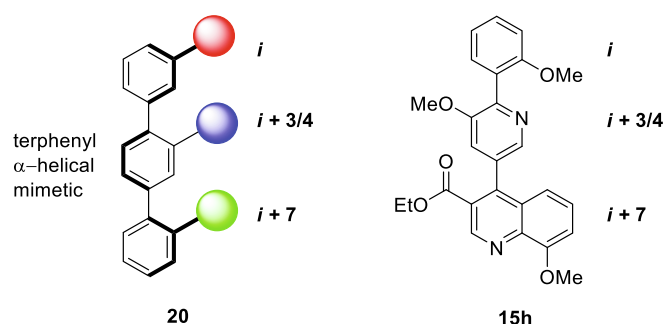


Fig. 5. The classical terphenyl α-helical mimetic PPI chemotype **20**, and the close overlay with terphenyl-based mGlu<sub>7</sub> NAMs, such as **15h**.

#### Declaration of Competing Interest

The authors declare that they have no known competing financial

interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Acknowledgments

We thank the Warren Family and Foundation for establishing the William K. Warren, Jr. Chair in Medicine (C.W.L.) and endowing the Warren Center for Neuroscience Drug Discovery. The authors also acknowledge funding by CDMRP Grant W81XWH-17-1-0266 and MH104158 (to C.M.N.).

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