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Directed evolution of a designer enzyme featuring an unnatural catalytic amino acid

Clemens Mayer[a]*, Christopher Dulson[a], Eswar Reddem[a], Andy-Mark W.H. Thunnissen[b] and Gerard Roelfes[a]*

Abstract: The impressive rate accelerations that enzymes display in nature often result from boosting the inherent catalytic activities of side chains by their precise positioning inside a protein binding pocket. Here we show that such fine-tuning is also possible for catalytic *unnatural* amino acids. Specifically, we report the directed evolution of a recently described designer enzyme, which utilizes an aniline side chain to promote a model hydrazone formation reaction. Consecutive rounds of directed evolution identified a number of mutations in the promiscuous binding pocket, in which the unnatural amino acid is embedded in the starting catalyst. When combined, these mutations boost the turnover frequency (k_{cat}) of the designer enzyme by almost 100-fold. Crucially, these gains result from strengthening the catalytic contribution of the unnatural amino acid, as the engineered designer enzymes outperform variants, in which the aniline side chain is replaced with a catalytically inactive tyrosine residue, by >200-fold.

The enviable rates and selectivities with which enzymes catalyze their transformations in nature have fueled efforts to create designer enzymes that can promote new-to-nature reactions with comparable proficiencies.[1–3] Toward this goal, several approaches for enzyme design have been developed over the past decades; those include *de novo* design,[4,5] (computationally-aided) protein redesign,[6,7] or recruitment of (un)natural cofactors to appropriate binding pockets.[8,9] Irrespective of the approach though, the catalytic activities of the resulting designer enzymes pale in comparison to those found in nature.[2] However, one important prospect of installing abiotic activities into proteinaceous scaffolds is the ability to boost low starting activities by mimicking the Darwinian algorithm in the laboratory.[10] Indeed, the iterative cycle of (1) introducing diversity through mutations, (2) identifying improved catalyst, and (3) amplifying more efficient enzyme variants – collectively referred to as directed evolution[11] – has given rise to engineered designer enzymes that display rate accelerations akin to those found in nature.[12–14]

In this report, we demonstrate that directed evolution is also a means to boost the proficiency of a novel class of designer enzymes, those that feature an unnatural amino acid as a catalytic residue.[15–17] Such designer catalysts mimic natural enzymes that employ posttranslational modifications of active site residues to install uniquely reactive functionalities to promote their target reactions (*i.e.* formylglycine in type-I-sulfatases).[18–20] Designer enzymes that make use of *catalytic* unnatural amino acids are also distinct from protein engineering efforts, in which genetic code expansion strategies[21,22] have been used to install non-standard side chains to improve/alter hydrophobic packing or substrate recognition.[23,24]

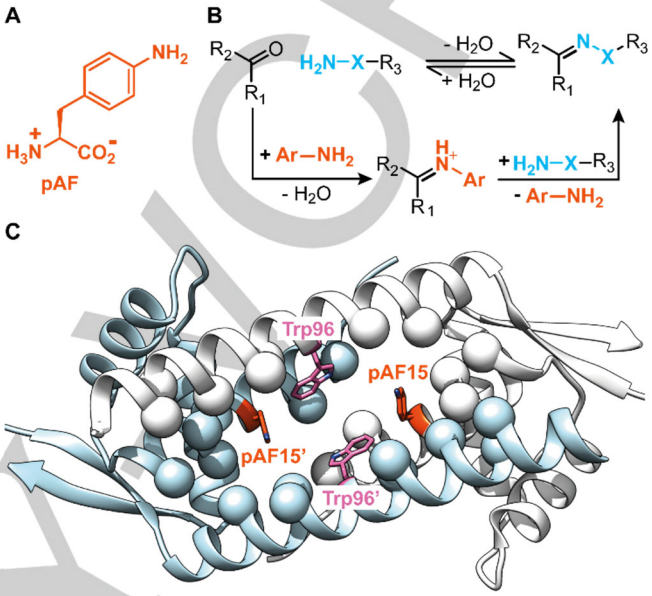

Figure 1. A: Chemical structure of *p*-aminophenylalanine. **B:** Formation of an iminium ion intermediate in presence of anilines accelerates hydrazone (X = NH) and oxime (X = O) formation reactions (for clarity, the reversibility of these reactions is not shown). **C:** Crystal structure of the LmrR_pAF homodimer (PDB: 6I8N). Catalytic aniline side chains (red) and Trp96 (pink) are shown as sticks. The positions of the β-carbons of 13 additional residues that line the binding pocket of LmrR_pAF are shown as spheres. A 3-(N-morpholino)propanesulfonic acid (MOPS) buffer molecule that was found to be sandwiched between the central tryptophans is omitted for clarity (see Supporting Information)

We recently reported the design and characterization of a designer enzyme, which utilizes a uniquely reactive *p*-aminophenylalanine (pAF, **Fig. 1A**) residue to promote abiological hydrazone and oxime formation reactions.[25] More specifically, introducing pAF at position 15 in the multidrug resistance regulator from *Lactococcus lactis* (LmrR), resulted in LmrR_pAF (previously assigned as LmrR_V15pAF), which promotes the condensation reactions of aldehydes with hydrazines or hydroxylamines through the formation of an iminium ion intermediate with the unnatural side chain (**Fig. 1B**).[26,27] While the designer enzyme outperformed aniline in solution by a factor ~560, the catalytic contribution of the unnatural side chain remained modest. In fact, an LmrR_pAF variant, which features a structurally similar but catalytically-

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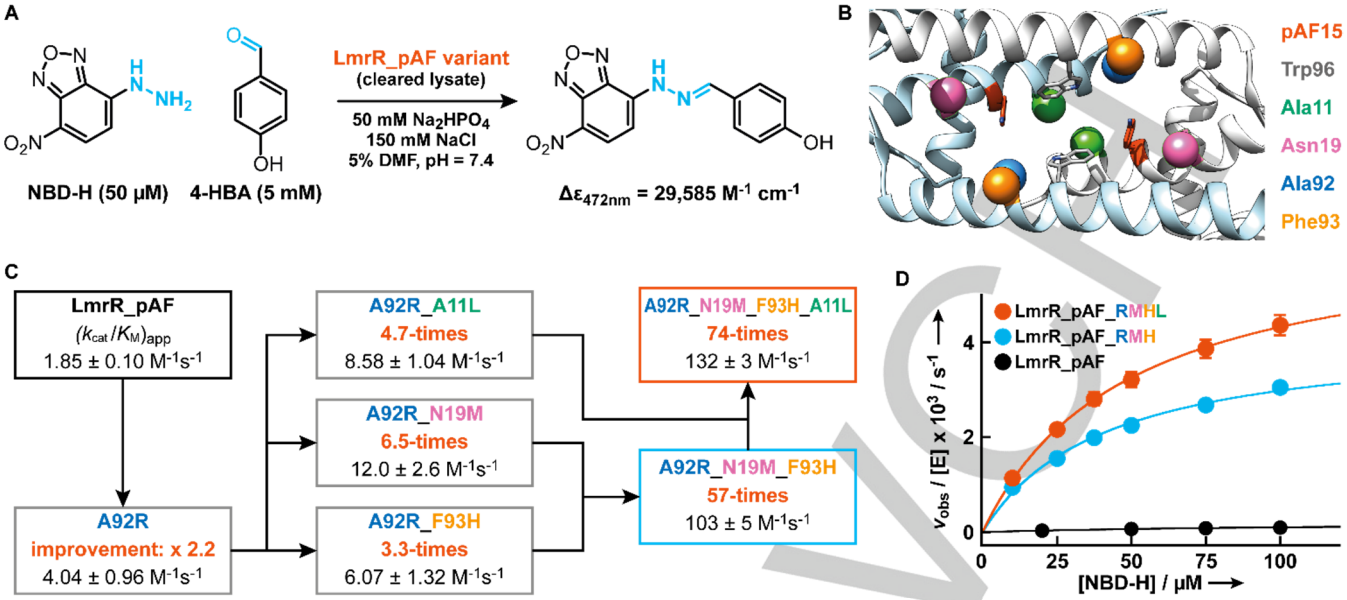

Figure 1. A: Reaction conditions for the model hydrazone formation between NBD-H and 4-HBA in cleared lysates. **B:** Close-up of the hydrophobic pore in LmrR_pAF. Trp96s and pAF15s shown as sticks; β-carbons of positions that gave rise to improved variants in round 1 and 2 are shown as spheres (color code as indicated). **C:** Evolutionary optimization of LmrR_pAF. Apparent catalytic efficiencies ($k_{cat}/K_m)_{app}$) of selected variants and their improvement with respect to LmrR_pAF (see **Table S1** and **Fig S3** for a more detailed analysis); errors are standard deviations of at least 3 independent experiments; color code as in **Fig. 2B**. **D:** Comparison of saturation kinetics at a 4-HBA concentration of 5 mM for LmrR_pAF and the best variants obtained after two rounds of directed evolution. Errors are standard deviations of at least 3 independent experiments.

inactive tyrosine residue instead of pAF (LmrR_Y), was only about 10-times less efficient the parent designer enzyme. Given the power of the evolutionary algorithm, we surmised that the role pAF for catalysis could be optimized by identifying beneficial mutations of residues surrounding the unnatural side chain.

Since our initial report of LmrR_pAF, we were able to elucidate its structure (**Fig. 1C**, see Supporting Information for details). As anticipated, the uniquely reactive aniline side chains are embedded in the hydrophobic pore at the homodimer interface of LmrR_pAF. Moreover, pAF residues flank the central tryptophans (W96s), which are key for recruiting planar, aromatic (substrate) molecules.[28] Upon inspection of this rudimentary active site, we identified 13 additional residues that line the hydrophobic pore and, as a result, are in proximity to the aniline side chain (**Fig. 1C**). We anticipated that targeting these residues by site-directed mutagenesis could give rise to more proficient LmrR_pAF variants.

In order to rapidly identify beneficial mutations in the LmrR_pAF binding pocket, we aimed to establish a medium-throughput screening assay that would allow for the parallel evaluation of LmrR_pAF variants. For this, we first optimized production of LmrR_pAF and a catalytically-inactive control protein (RamR)[29] in 96-well format and prepared cleared lysates. Incorporation of pAF was achieved by first introducing *p*-azidophenylalanine in response to a stop codon at position 15 (using the helper plasmid pEVOL-pAzF)[30] and subsequently reducing the azido group with tris(2-carboxyethyl)phosphine in cell lysates (see Supporting Information). To assess the catalytic activity of the resulting LmrR_pAF in this complex mixture, we took advantage of the chromogenic hydrazone formation reaction between 4-hydrazino-7-nitro-2,1,3-benzoxadiazole (NBD-H) and 4-hydroxybenzaldehyde (4-HBA, **Fig. 2A**).[31] These two substrates (final concentrations: 50 µM NBD-H and 5 mM 4-HBA) were added to the cleared lysates and product formation was followed at 472 nm in a 96-well plate reader for 2 hours. Indeed, lysates containing LmrR_pAF displayed 30% higher product formation rates than those containing RamR (**Fig. S1**). Moreover, consistent with its lower activity, LmrR_Y (the variant containing tyrosine instead of pAF) did not provide significant rate acceleration with respect to RamR (**Fig. S1**). Taken together, these results suggest that it should be feasible to accurately assess the activity of LmrR_pAF variants in parallel.

To identify beneficial mutations, we initially targeted 8 residues (**Fig S2**), which are either in close proximity to pAF15 or have previously been identified to improve catalytic parameters of unrelated, LmrR-based designer enzymes.[32–34] Libraries targeting each position were constructed using degenerate primers (NNK codons, which allow for all 20 canonical amino acids) and ~400 library members were evaluated following the previously established screen. Two variants LmrR_pAF_A92R and LmrR_pAF_L18K, which gave large improvements of 2.46 ± 0.13 and 2.04 ± 0.11 when compared to LmrR_pAF, were subsequently produced and purified (**Fig. S2**, see Supporting Information). Upon determining their catalytic parameters, both variants displayed improved apparent catalytic efficiencies ($(k_{cat}/K_m)_{app}$), when compared to the parent designer enzyme (2.2-times for A92R and 1.5-times for L18K, **Fig. 2A**, **Fig. S3** and **Table S1**). Combined, these results attest that reproducible gains in the screen translate into improved catalytic parameters for purified LmrR_pAF variants.

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Table 1. Steady-state parameters of parent and engineered designer enzymes.

catalyst [a]	k_{cat} x 10^2 (s^{-1})	K_{NBD-H} (μM)	K_{4-HBA} (mM)	k_{cat}/K_{NBD-H} K_{4-HBA} (M^{-2} s^{-1})	EM[a] (M)	1 / $K_{TS}^{‡}$ (M^{-1})[b]	vs aniline[c]
LmrR_pAF	0.05 (0.002)	100 (7)	7.92 (0.49)	630 (60)	1.26	1.6 x 10^6	560
LmrR_pAF_RMH	4.53 (0.33)	48 (4)	46.4 (4.3)	20,500 (2,500)	115	5.2 x 10^7	18,400
LmrR_pAF_RMHL	2.76 (0.11)	49 (2)	18.9 (1.0)	29,500 (2,000)	69.8	7.5 x 10^7	26,500

Determined at 25 °C in phosphate buffer (50 mM) containing NaCl (150 mM) and 5% (v/v) DMF at pH 7.4. The estimated errors reflect the standard deviations of at least three independent experiments. Under the same conditions k_{uncat} = 3.95 x 10^{-4} M^{-1} s^{-1} and $K_{aniline}$ = 1.12 M^{-2} s^{-1}.[16] [a] effective molarity (EM = k_{cat}/k_{uncat}) [b] chemical proficiency ($1/K_{TS}^{‡}$ = [$k_{cat}/(K_{NBD-H} \cdot K_{4-HBA})$]/$k_{uncat}$ [c] vs aniline = ([$k_{cat}/(K_{NBD-H} \cdot K_{4-HBA})$] / $K_{aniline}$) comparison of apparent third order rate constants of designer enzymes and aniline.

Encouraged by these results, we chose LmrR_pAF_R (containing the A92R mutation) as the template for a new round of mutagenesis. In total, 12 additional positions were randomized and 84 library members per targeted position screened (~1000 variants in total, **Fig. S2**). We identified another three mutations (A11L, N19M, and F93H, but not L18K), which independently led to significant rate accelerations with respect to LmrR_pAF_R (**Fig. 2B-C**). To identify potential synergistic effects between these three mutations we constructed LmrR_pAF_R variants that combined
two or all three of these mutations. Strikingly, the combination of N19M and F93H (LmrR_pAF_RMH from here onward) provided an exceptionally large improvement compared to either of the other variants tested (**Figs. 2C-D**, **Fig. S3** and **Table S1**). In fact, when comparing the apparent catalytic efficiencies, LmrR_pAF_RMH was 57-times more efficient than the parent designer enzyme. The variant that featured the additional A11L mutation (LmrR_pAF_RMHL) provided another ~30% improvement and, thus, outperformed LmrR_pAF by 74-times (**Fig. 2C-D**).

Higher efficiencies for these two LmrR_pAF variants result predominantly from an increase in the apparent turnover frequency ($k_{cat,app}$, **Table S1**) and, point toward the desired fine-tuning of the inherent catalytic activity of the unnatural amino acid. Consistent with such a scenario, mutation of pAF15 to tyrosine in LmrR_pAF_RMH and LmrR_pAF_RMHL proved crippling, resulting in an efficiency loss of 99.7% for the former and 99.5% for the latter. Moreover, when compared to the apparent catalytic efficiency determined for LmrR_Y, the addition of the three or four identified mutations only led to modest improvements of 2.0 and 3.9-times, respectively (**Fig. S3** and **Table S1**). These results further underscore that the identified mutations tailor the catalytic contribution of pAF.

The effect of the beneficial mutations in LmrR_pAF_RMH and LmrR_pAF_RMHL on the kinetics was studied in more detail. By measuring the dependence of the reaction velocity on NBD-H concentration (10-100 μM) at several fixed 4-HBA concentrations (3-20 mM), we obtained steady-state kinetic parameters for both variants (**Table 1** and **Fig. S4**). Notably, the two engineered designer enzymes have significantly higher K_m values for 4-HBA than LmrR_pAF, an intriguing result, as increasing the affinity for this substrate could have provided a straightforward means to accelerate the bimolecular hydrazone formation. Instead, higher catalytic efficiencies are the result of higher turnover frequencies (k_{cat}), which, again, is consistent with boosting the performance of pAF for catalysis. Notably, under saturation conditions LmrR_pAF_RMH and LmrR_pAF_RMHL display k_{cat} values 91 and 55 times higher than the parent designer enzyme. Moreover, the engineered variants outperform aniline in solution by more than 4 orders of magnitude (**Table 1**). Thus, while milimolar concentrations of aniline are required to observe appreciable rate acceleration in our model hydrazone formation reaction,[25] engineered LmrR_pAF variants give rise to the same increases at a concentration of <1 μM.

Notably, fine-tuning the inherent catalytic potential of the aniline side chain proved also beneficial for hydrazone formations with aldehydes different than 4-HBA. In fact, both LmrR_pAF_RMH and LmrR_pAF_RMHL significantly accelerated hydrazone formation in presence of a total of 7 aldehydes (**Fig. S5**, see Supporting Information for details). A minor specialization for 4-HBA (the screening substrate), in the engineered variants is apparent, as the evolved variants displayed a slight preference for this aldehyde, while LmrR_pAF did not. Another notable difference between the parent and the engineered designer enzymes was the ability of the latter to significantly accelerate the hydrazone formation in presence of 2-formylbenzoic acid; LmrR_pAF itself did not provide appreciable levels of activity, under which all other aldehydes did. Thus, these results indicate that the directed evolution of unnatural amino acid containing designer enzymes can generate highly active as well as versatile catalysts.

In absence of available structural information, it is difficult to rationalize how the identified mutations boost the catalytic potential of the aniline side chain. Both A11L and N19M are located one helical turn away from pAF15 (**Fig. 2B**) and are likely involved in positioning the unnatural side chain in a productive conformation. A92R and F93H are located opposite of pAF15. The introduction of a permanent positive charge by A92R is somewhat puzzling, as it could negatively impact the population of the crucial iminium ion (**Fig. 1A**), due to charge-charge repulsion.[35] However, in the formation of this covalent, transient intermediate, negative species are formed, which in turn could be stabilized by the guanidinium side chain.[31,36] The F93H mutation, in combination with N19M, appears critical for the boosts

observed in LmrR_pAF_RMH and LmrR_pAF_RMHL, as demonstrated by the drastic loss of activity in the respective F93 or N19 reversion variants (**Fig. 2C**, **Table S1**). It is, therefore, tempting to suggest that histidine aides in the formation of the iminium ion and/or transimination through proton shuffling or positioning of ordered water molecules.[26,31,36,37]

In conclusion, our work demonstrates that designer enzymes featuring an unnatural amino acid as a catalytic residue can be privileged starting points for directed evolution campaigns. Enhancing the inherent catalytic activity of an unnatural side chain is feasible by identifying beneficial mutations in a protein scaffold. Future efforts will focus on the structural and computational analysis of tailored LmrR_pAF variants in order to pinpoint the exact mechanisms by which these engineered designer enzymes can display an almost 100-times higher turnover frequency when compared to the parent variant. Lastly, we surmise that the impressive improvements observed for LmrR_pAF are not limited to an aniline side chain. Instead, we suggest that the introduction and fine tuning of other organocatalysts[38–40] – which are versatile yet notoriously slow – through genetic code expansion will result in proficient designer enzymes for a wide variety of new-to-nature reactions. Ultimately, such efforts could provide a promising route for developing efficient protein catalysts for synthetically relevant transformations.

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[1] V. Nanda, R. L. Koder, *Nat. Chem.* **2010**, *2*, 15–24.
[2] D. Hilvert, *Annu. Rev. Biochem.* **2013**, *82*, 447–70.
[3] S. C. Hammer, A. M. Knight, F. H. Arnold, *Curr. Opin. Green Sustain. Chem.* **2017**, *7*, 23–30.
[4] D. N. Woolfson, G. J. Bartlett, A. J. Burton, J. W. Heal, A. Niitsu, A. R. Thomson, C. W. Wood, *Curr. Opin. Struct. Biol.* **2015**, *33*, 16–26.
[5] P.-S. Huang, S. E. Boyken, D. Baker, *Nature* **2016**, *537*, 320–327.
[6] G. Kiss, N. Çelebi-Ölçüm, R. Moretti, D. Baker, K. N. Houk, *Angew. Chem. Int. Ed.* **2013**, *52*, 5700–5725. *Angew. Chem.*. **2013**, *125*, 5810–5836.
[7] M. P. Frushicheva, M. J. L. Mills, P. Schopf, M. K. Singh, R. B. Prasad, A. Warshel, *Curr. Opin. Chem. Biol.* **2014**, *21*, 56–62.
[8] O. F. Brandenberg, R. Fasan, F. H. Arnold, *Curr. Opin. Biotechnol.* **2017**, *47*, 102–111.
[9] F. Schwizer, Y. Okamoto, T. Heinisch, Y. Gu, M. M. Pellizzoni, V. Lebrun, R. Reuter, V. Köhler, J. C. Lewis, T. R. Ward, *Chem. Rev.* **2018**, *118*, 142–231.
[10] H. Renata, Z. J. Wang, F. H. Arnold, *Angew. Chem. Int. Ed.* **2015**, *54*, 3351–3367. *Angew. Chem.* **2015**, *127*, 3408–3426.
[11] M. S. Packer, D. R. Liu, *Nat. Rev. Genet.* **2015**, *16*, 379–394.
[12] C. Zeymer, D. Hilvert, *Annu. Rev. Biochem.* **2018**, *87*, 131–157.
[13] R. Obexer, A. Godina, X. Garrabou, P. R. E. E. Mittl, D. Baker, A. D. Griffiths, D. Hilvert, *Nat. Chem.* **2017**, *9*, 50–56.
[14] R. Blomberg, H. Kries, D. M. Pinkas, P. R. E. Mittl, M. G. Grütter, H. K. Privett, S. L. Mayo, D. Hilvert, *Nature* **2013**, *503*, 418–421.
[15] M. Pott, T. Hayashi, T. Mori, P. R. E. Mittl, A. P. Green, D. Hilvert, *J. Am. Chem. Soc.* **2018**, *140*, 1535–1543.
[16] T. Hayashi, D. Hilvert, A. P. Green, *Chem. Eur. J.* **2018**, *24*, 11821–11830.
[17] H. Yang, A. M. Swartz, H. J. Park, P. Srivastava, K. Ellis-Guardiola, D. M. Upp, G. Lee, K. Belsare, Y. Gu, C. Zhang, et al., *Nat. Chem.* **2018**, *10*, 318–324.
[18] N. M. Okeley, W. a van der Donk, *Chem. Biol.* **2000**, *7*, 159-71.
[19] M. J. Appel, C. R. Bertozzi, *ACS Chem. Biol.* **2015**, *10*, 72–84.
[20] H. A. Cooke, C. V. Christianson, S. D. Bruner, *Curr. Opin. Chem. Biol.* **2009**, *13*, 453–461.
[21] D. D. Young, P. G. Schultz, *ACS Chem. Biol.* **2018**, *13*, 854−870.
[22] A. Dumas, L. Lercher, C. D. Spicer, B. G. Davis, *Chem. Sci.* **2015**, *6*, 50–69.
[23] C. L. Windle, K. J. Simmons, J. R. Ault, C. H. Trinh, A. Nelson, A. R. Pearson, A. Berry, *Proc. Natl. Acad. Sci.* **2017**, *114*, 2610–2615.
[24] F. Agostini, J. S. Völler, B. Koksch, C. G. Acevedo-Rocha, V. Kubyshkin, N. Budisa, *Angew. Chem. Int. Ed.* **2017**, *56*, 9680–9703. *Angew. Chem.* **2017**, 129, 9810-9835
[25] I. Drienovská, C. Mayer, C. Dulson, G. Roelfes, *Nat. Chem.* **2018**, *10*, 946–952.
[26] E. H. Cordes, W. P. Jencks, *J. Am. Chem. Soc.* **1962**, *84*, 826–831.
[27] A. Dirksen, S. Dirksen, T. M. Hackeng, P. E. Dawson, *J. Am. Chem. Soc.* **2006**, *128*, 15602–15603.
[28] P. K. Madoori, H. Agustiandari, A. J. M. Driessen, A.-M. M. W. H. Thunnissen, *EMBO J.* **2009**, *28*, 156–166.
[29] S. Yamasaki, E. Nikaido, R. Nakashima, K. Sakurai, D. Fujiwara, I. Fujii, K. Nishino, *Nat. Commun.* **2013**, *4*, 1–7.
[30] J. W. Chin, S. W. Santoro, A. B. Martin, D. S. King, L. Wang, P. G. Schultz, *J. Am. Chem. Soc.* **2002**, *124*, 9026–9027.
[31] P. Crisalli, E. T. Kool, *J. Org. Chem.* **2013**, *78*, 1184–1189.
[32] I. Drienovská, A. Rioz-Martínez, A. Draksharapu, G. Roelfes, *Chem. Sci.* **2015**, *6*, 770–776.
[33] I. Drienovská, L. Alonso-Cotchico, P. Vidossich, A. Lledós, J. D. Maréchal, G. Roelfes, *Chem. Sci.* **2017**, *8*, 7228–7235.
[34] L. Villarino, K. E. Splan, E. Reddem, L. Alonso-Cotchico, C. Gutiérrez de Souza, A. Lledós, J. D. Maréchal, A. M. W. H. Thunnissen, G. Roelfes, *Angew. Chem. Int. Ed.* **2018**, *57*, 7785–7789. *Angew. Chem.* **2018**, *130*, 7911–7915.
[35] E. T. Kool, D. Park, P. Crisalli, *J. Am. Chem. Soc.* **2013**, *135*, 17663–17666.
[36] P. Crisalli, E. T. Kool, *Org. Lett.* **2013**, *15*, 1646–1649.
[37] A. Dirksen, T. M. Hackeng, P. E. Dawson, *Angew. Chem. Int. Ed.* **2006**, *45*, 7581–7584. *Angew. Chem.* **2006**, *118*, 7743–7746.
[38] E. Zandvoort, E. M. Geertsema, B. J. Baas, W. J. Quax, G. J. Poelarends, *Angew. Chem. Int. Ed.* **2012**, *51*, 1240–1243. *Angew. Chem.* **2012**, *124*, 1266-1269.
[39] Y. Miao, M. Rahimi, E. M. Geertsema, G. J. Poelarends, *Curr. Opin. Chem. Biol.* **2015**, *25*, 115–123.
[40] A. R. Nödling, K. Świderek, R. Castillo, J. W. Hall, A. Angelastro, L. C. Morrill, Y. Jin, Y. H. Tsai, V. Moliner, L. Y. P. Luk, *Angew. Chem. Int. Ed.* **2018**, *57*, 12478–12482. *Angew. Chem.* **2018**, *130*, 12658–12662.

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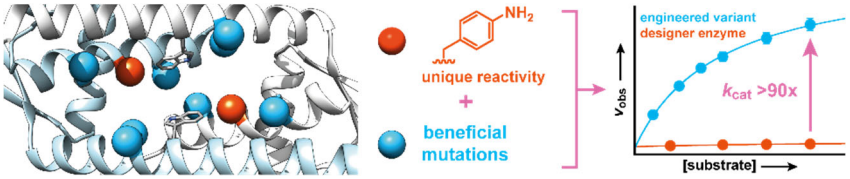

The directed evolution of a designer enzyme featuring a uniquely reactive aniline side chain as catalytic residue (in red) is reported. Multiple beneficial mutations were identified (blue), which when combined increase the turnover frequency (k_{cat}) of the designer enzyme by >90 times. Engineered variants also outperform aniline by >4 orders of magnitude. Critically, this drastic increase is achieved by boosting the inherent catalytic activity of the aniline side chain.

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