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Synthesis and Evaluation of Biaryl Derivatives for Structural

Characterization of Selective Monoamine Oxidase B Inhibitors toward

Parkinson's Disease Therapy

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Abstract

Benzyloxyphenyl moiety is a common structure of highly potent, selective and reversible inhibitors of monoamine oxidase B (MAO-B), safinamide and sembragiline. We synthesized 4-(benzyloxy)phenyl and biphenyl-4-yl derivatives including halogen substituents on the terminal aryl unit. In addition, we modified the carbon linker between amine group and the biaryl linked unit. Among synthesized compounds, **12c** exhibited the most potent and selective MAO-B inhibitory effect (hMAO-B IC₅₀: 8.9 nM; >10,000-fold selectivity over MAO-A) as a competitive inhibitor. In addition, **12c** showed greater MAO-B inhibitory activity and selectivity compared to well-known MAO-B inhibitors such as selegiline, safinamide and sembragiline. In the MPTP-induced mouse model of Parkinson's disease (PD), **12c** significantly protected the tyrosine hydroxylase (TH)immunopositive DAergic neurons and attenuated the PD-associated behavioral deficits. This study suggests characteristic structures as a MAO-B inhibitor that may provide a good insight for the development of therapeutic agents for PD.

Keywords

Benzyloxyphenyl derivatives, MAO-B inhibitor, MPTP mouse model, Parkinson's disease

Introduction

Parkinson's disease (PD) is a progressive neurodegenerative movement disorder characterized by marked motor deficits such as resting tremor, bradykinesia, rigidity and postural instability.¹ These motor symptoms are mostly associated with dopamine (DA) depletion caused by a profound loss of dopaminergic (DAergic) neurons in the substantia nigra pars compacta (SNpc). However, the exact mechanisms causing neurodegeneration are not yet known.²⁻⁴

Recently, several studies have reported that the expression level of monoamine oxidase B (MAO-B) in human brain increases with age, and activity is highly increased in the substantia nigra of PD patients.^{5,6} MAOs are localized on the outer membrane of mitochondria, particularly in the liver and brain, and catalyze the oxidative deamination of monoamine neurotransmitters such as dopamine.⁷ In addition, the reaction catalyzed by MAOs result in the production of hydrogen peroxide (H₂O₂) which causes oxidative stress and neuronal cell death.^{8,9} There are two isoforms, MAO-A and MAO-B. MAO-A is localized in catecholaminergic neurons and selectively inhibited by low concentrations of clorgyline (1), whereas MAO-B is abundant in serotonergic neurons and astrocytes, and selectively inhibited by potent inhibitors selegiline (2) and rasagiline (3) (Fig. 1).¹⁰⁻¹²







Selegiline (2)

Rasagiline (3)

Fig. 1. Chemical structures of irreversible MAO-B inhibitors

Selegiline and rasagiline, selective and potent MAO-B inhibitors, are the most widely used drugs for the therapy of PD by either mono-therapy or in combination with the levodopa, the dopamine

prodrug, resulting in the increase of dopamine.^{13,14} However, selegiline and rasagiline are irreversible MAO-B inhibitors and showed undesirable adverse effects in long term treatment of PD such as hallucinations and headaches. Recent studies have been shown that the irreversibility of MAO-B inhibition may contribute short-lived action.¹⁵ Accordingly, the development of selective and reversible MAO-B inhibitors could reduce undesirable adverse effects and maintain the effectiveness in long-term use for the treatment of neurodegenerative diseases. Recently, safinamide (4, XadagoTM, Fig. 2), a highly selective and reversible MAO-B inhibitor, was approved for the treatment of mildto-late PD patients as an add-on therapy to stable dose of levodopa alone or in combination with other PD medications.¹⁶ However, safinamide may cause disadvantageous effects due to other biological properties such as selective sodium channel blockade, calcium channel modulation, and inhibition of stimulated release of glutamate.¹⁷⁻²⁰ Sembragiline (5, RG-1577, Fig. 2) is another potent reversible MAO-B inhibitor for the treatment of Alzheimer's disease (AD). MAO-B activity is linked to the production of reactive oxygen species (ROS) that can cause neuronal damage. Sembragiline was expected to slow progression of neurodegeneration by reducing oxidative stress. However, it failed to demonstrate benefit of the primary endpoint in clinical trial phase 2.^{21,22} Interestingly, two recent potent and selective MAO-B inhibitors have common moieties, ((3fluorobenzyl)oxy)phenyl group (Fig. 2, red box).

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In this study, we asked if compounds conforming to the biaryl linked unit exhibited MAO-B inhibitory activity. First, we synthesized substituted (4-(benzyloxy)phenyl)-methylammonium chlorides. Next, we modified the carbon linker between amine group and the biaryl linked unit (**Fig.3.**). In addition, we prepared biphenyl-4-yl derivatives wherein the aryl linker (X) is a single bond (**Fig.3.**). We reported that the substituted (4-(benzyloxy)phenyl)-ethylammonium chlorides exhibited potent and selective MAO-B inhibitory activities. One of the compounds with the highest potency was further evaluated for in vivo efficacy in the MPTP-induced mouse model of PD.



Fig. 3. Representative chemical structures of biaryl derivatives.

Recently, some biaryl derivatives in our study have been reported their biological activities such as anticonvulsant activity²³ and trace amine-associated receptor agonist effects²⁴. However, this is the

first report to propose structural features for the excellent activity of MAO-B inhibitors by variously modifying the biaryl linker and the carbon linker. Thus, this study can provide insight into the development of new potential MAO-B inhibitors for PD therapy.

Results and Discussion

2.1. Selection of compounds

According to recent literature, safinamide (**4**) has emerged as a new therapeutic agent for the treatment of PD through MAO-B inhibition. We divided safinamide into two parts (the blue and the red boxes in Fig. 2) to determine the part that affected the MAO-B inhibitory effects. We found that (4-((3-fluorobenzyl)oxy)phenyl)methanaminium chloride (**7f**) corresponding to the red box showed much potent than compound **15** corresponding to blue box (IC₅₀ hMAO-B = 0.926 μ M vs >10 μ M). Sembragiline (**5**, RG-1577), a reversible MAO-B inhibitor, has a ((3-fluorobenzyl)oxy)phenyl group in common with safinamide. We observed that 4-((3-fluorobenzyl)oxy)benzenaminium chloride (**10e**) corresponding to the red box in **5** inhibited MAO-B activity. Accordingly, we prepared 4- (benzyloxy)phenyl and biphenyl-4-yl derivatives which included various substituents on the terminal aryl unit. We also modified the carbon linker between amine group and the biaryl linked unit.

2.2. Synthesis

Compounds **7** and **8** were prepared in two steps (Scheme 1). Using a slightly modified method from the previous method, we prepared the Williamson ether coupled products **6** by treating 4-cyanophenol with the appropriate substituted benzyl bromide and potassium carbonate (K_2CO_3). Lithium aluminum hydride (LiAlH₄) reduction of the nitrile group in **6a-6k** afforded the amines,

which were immediately converted to the corresponding aminium chlorides 7a-7k. The conversion of nitriles (6a-6k) to amides by partial hydrolysis with KOH afforded benzamide derivatives 8a-8f. To prepare 4-nitrophenoxy benzene derivatives 9a-9f, we synthesized 4-nitrophenol and the appropriate substituted benzyl bromide using the similar method above except reaction solvent (DMF) (Scheme 2). 4-nitrophenoxy benzene derivatives 9a-9f were reduced to benzenaminium chlorides 10a-10f by platinum (II) oxide under hydrogen gas (Scheme 2). Compounds 11 were prepared by Williamson ether coupling using tert-butyl (4-hydroxyphenethyl)carbamate and the substituted benzyl bromide. Deprotection of boc-group by 4.0M hydrogen chloride in dioxane gave ethanaminium chlorides 12a-12j (Scheme 3). The (biphenyl-4-yl)ethanaminum chlorides were prepared using Suzuki coupling of N-boc-2-(4-bromophenyl)ethylamine (13) with the appropriate, commercially available substituted aryl boronic acids to give the boc-protected amines, which then were immediately deprotected and converted to their hydrochloride salts 14a-14j using HCl (Scheme

4).



Scheme 1. General Procedure for Substituted (Benzyloxyphenyl)methanaminium Chlorides and Substituted (Benzyloxy)benzamides.



Scheme 3. General Procedure for Substituted (Benzyloxyphenyl)ethanaminium Chlorides.



2.3. Inhibitory Activities of the Synthesized Compounds against Monoamine Oxidase B

The synthesized compounds were evaluated for inhibitory activities of human MAO-A and MAO-B using recombinant enzymes. The enzyme inhibition assay on test compounds was performed using Amplex Red reagents and the inhibitory activity was determined by spectrophotometrically measuring the resorufin formation at 570 nm. The inhibitory potencies (IC_{50} values) for all synthesized compounds are summarized in Table 1, with comparative values of the positive controls, selegiline, safinamide and sembragiline.

First, we prepared 11 (benzyloxyphenyl)methanaminium chlorides with various functional groups on the terminal aryl ring (Table 1, compounds 7). The first set of compounds 7 contained methylamine hydrochloride groups at one end of the biaryl linked unit. Compound 7f, which is the biaryl linked unit of safinamide (the red box on the left in **Fig. 2**), exhibited moderate MAO-B inhibitory activity (IC₅₀ = 0.926 μ M). We introduced a variety of electron-withdrawing groups and electron-donating group into the terminal aryl ring (7a-7k). We observed that the two groups did not exhibit significant activity differences, but most of them showed moderate inhibitory activities (IC₅₀: 0.136 μ M ~ 0.926 μ M).

Next, we changed the number of carbon atoms between the amine group and the biaryl linked unit of compound **7** from one to zero (**10**) and two (**12**). Compound **10e**, which is the biaryl linked unit of sembragiline (the red box on the right in **Fig. 2**), has lower MAO-B inhibitory activity ($IC_{50} = 2.483$ µM) than compound **7f**. All compound **10**, in which no carbon was present between the amine groups and the biaryl linked units, exhibited lower inhibitory activity (IC_{50} : 0.402 µM ~ >10 µM) than compound **7** having one carbon at the corresponding position. We investigated whether adding one carbon to this position of compound **7** improved MAO-B inhibitory activities. Interestingly, this modification resulted in dramatic improvement of inhibitory activity in most compounds (IC_{50} : 0.009 µM ~ 0.396 µM). For the CF₃-substituted compounds **12a-12c**, we found that the 4'-CF₃ derivative

12c was more potent than the corresponding 2'- and 3'-CF₃ derivatives 12a and 12b (12c, IC₅₀ = 0.009 μ M; 12a, IC₅₀ = 1.803 μ M; 12b, IC₅₀ = 0.110 μ M). A similar trend on the inhibitory activities of 2', 3', and 4' regioisomers was observed in the F- or Cl-substituted compounds 12d-12i. In addition, we modified the methanaminum chloride group of compound 7 to an amide group, and found that the substitution with the amide group did not significantly affect MAO-B inhibitory activity.

Finally, we prepared (biphenyl-4-yl)ethanaminium chlorides **14a-14j** wherein the aryl linker is a single bond. In this case, the compounds substituted with bulky electron-withdrawing groups (CF₃ and OCF₃) exhibit far superior inhibitory activity than the F- or Cl-substituted compounds **14c-14h**. In particular, 4'-CF₃ derivatives **14b** showed much better MAO-B inhibitory activities than 4'-F and 4'-Cl derivatives **14e** and **14h** (**14b**, IC₅₀ = 0.041 μ M; **14e**, IC₅₀ = 6.806 μ M; **14h**, IC₅₀ = 0.625 μ M).

Table 1. Inhibitory effects of the synthesized compounds against hMAOs.

$CI H_3N () (R^1 R^2)$																
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Compd	n	\mathbb{R}^1	\mathbb{R}^2	R ³	MAO-B $(IC_{50}, \mu M)^{a}$	MAO-A (IC ₅₀ , μM)	SI ^b	Compd	n	\mathbf{R}^1	R ²	R ³	MAO-B $(IC_{50}, \mu M)^a$	MAO-A (IC ₅₀ , μM)	SI ^b	
7a	1	Н	Н	Н	3.225±0.06	>100	>31	10d	0	F	Н	Н	>10	>30	nd ^e	
7b	1	CF ₃	Н	Н	3.146±0.40	>100	>32	10e	0	Н	F	Н	2.483±0.07	>100	>40	
7c	1	Н	CF ₃	Н	0.387 ± 0.08	>100	>250	10f	0	Н	Н	F	3.324±0.11	>100	>30	
7d	1	Н	Н	CF ₃	0.136±0.01	>100	>730	12a	2	CF ₃	Н	Н	1.803±0.12	>100	>55	
7e	1	F	Н	Н	1.462±0.19	>100	>68	12b	2	Н	CF_3	Н	0.110±0.02	>100	>900	
7 f	1	Н	F	Н	0.926±0.03	>100	>100	12c	2	Н	Н	CF ₃	0.009 ± 0.001	>100	>11000	
7g	1	Н	Н	F	0.452 ± 0.07	>100	>220	12d	2	F	Н	Н	0.396±0.02	>10	>25	
7h	1	Н	Cl	Н	0.257 ± 0.02	>100	>380	12e	2	Η	F	Н	0.193 ± 0.01	>100	>510	
7i	1	Н	Н	Cl	0.440 ± 0.02	>30	>68	12f	2	Η	Н	F	0.109 ± 0.01	>100	>910	
7j	1	Н	OMe	Н	0.175 ± 0.01	>30	>170	12g	2	Cl	Н	Н	2.849 ± 0.07	>10	>3.5	
7k	1	Н	Н	OMe	0.283 ± 0.02	>100	>350	12h	2	Н	Cl	Н	0.062 ± 0.01	>100	>1600	
10a	0	CF ₃	Н	Н	>10	>100	nd ^e	12i	2	Н	Н	Cl	0.059 ± 0.01	>100	>1600	
10b	0	Н	CF ₃	Н	1.507±0.03	>100	>66	12j	2	Н	OMe	Н	0.305±0.03	>10	>33	
10c	0	Н	Н	CF ₃	0.402 ± 0.02	>100	>240									
H_2N R^1 R^2 R^3								$CI H_3N^+$ R^1 R^2 R^3								

Compd	\mathbf{R}^1	\mathbf{R}^2	\mathbb{R}^3	$\begin{array}{c} \text{MAO-B} \\ (\text{IC}_{50}, \mu\text{M})^a \end{array}$	MAO-A (IC ₅₀ , μM)	SI ^b	Compd	\mathbf{R}^1	R ²	R ³	$\begin{array}{c} \text{MAO-B} \\ (\text{IC}_{50}, \mu\text{M})^a \end{array}$	MAO-A (IC ₅₀ , μM)	SI ^b
8a	CF ₃	Н	Н	2.023±0.20	>100	>49	14a	Н	CF ₃	Н	1.568 ± 0.04	>10	>6.4
8b	Н	CF ₃	Н	1.108 ± 0.17	>100	>90	14b	Н	Н	CF_3	0.041 ± 0.02	>10	>240
8c	Н	Н	CF ₃	0.642 ± 0.09	>100	>150	14c	F	Н	Н	>10	>10	nd ^e
8d	F	Н	Н	1.055 ± 0.07	>100	>95	14d	Н	F	Н	>10	>10	nd ^e
8e	Н	F	Н	1.550 ± 0.05	>100	>65	14e	Н	Н	F	6.806±0.09	>10	>1.5
8f	Н	Н	F	0.271 ± 0.01	>100	>360	14f	Cl	Н	Н	8.340±0.12	>10	>1.2
Clorgyline ^c (1)				>1	0.008 ± 0.001	nd ^e	14g	Н	Cl	H	2.110±0.07	>10	>4.7
Selegiline ^d (2)				0.011 ± 0.001	1.5 ± 0.02	>130	14h	Н	Н	CI	0.625 ± 0.05	>10	>16
Safinamide ^d (4)				0.017 ± 0.002	>100	>5800	14i	Н	OMe	Н	5.003±0.23	>10	>1.9
Sembragiline ^d (5)				0.016 ± 0.001		nd ^e	14j	Н	Н	OMe	5.757±0.19	>10	>1.7
15	H ₂ N	H N MeSO ₃ H	OMe	>10	>10	nd ^e		5	2				

^{*a*}Inhibition data are reported as IC₅₀ (μ M) ± SEM.

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 ${}^{b}SI =$ selectivity index, the selectivity for the MAO-B isoform and is given as the ratio of IC₅₀(MAO-A)/IC₅₀(MAO-B).

^cPositive controls (MAO-A inhibitor); clorgyline: irreversible inhibitor

^dPositive controls (MAO-B inhibitor); selegiline: irreversible inhibitor, (S)-safinamide & sembragiline: reversible inhibitor. ^end: not determined

2.4. Mode of MAO-B inhibition of 12c

To examine the interaction mode of MAO-B inhibition, the most potent compound **12c** was evaluated in substrate-dependent kinetic experiments, and both the corresponding progression curves and the Linewever-Burk plots were generated. The initial rates of the MAO-B-catalyzed oxidation of benzylamine at seven different substrate concentrations in the absence and in the presence of three different concentrations of **12c** were measured. The results are depicted in **Fig. 4**. The Lineweaver-Burk plots for different concentration of **12c** were linear and intersected at the *y*-axis. Using GraphPad Prism, the maximal velocity (V_{max}), the Michaelis constant (K_m) and the inhibition constant (K_i) were calculated ($V_{max} = 5.864e^{+7}$, $K_m = 4.093^{-4}$ and $Ki = 6.88e^{-9}$). These results indicate that **12c** is a competitive MAO-B inhibitor. (**Fig. 4**)



Fig. 4. Mode of **12c** with the binding site of MAO-B. The catalytic rates were measured at different concentrations of benzylamine (0.065, 0.125, 0.25, 0.5, 1, 2 and 4 mM) in the absence and in the presence of different concentrations (0.3, 1 and 3 nM) of **12c**. In the Lineweaver-Burk plot, the reciprocal MAO-B inhibitory activity was plotted against the reciprocal substrate concentration. The maximal velocity (V_{max}), the Michaelis constant (K_m) and the inhibition constant (K_i) were calculated

 $(V_{\text{max}} = 5.864 \text{e}^{+7}, K_{\text{m}} = 4.093^{-4} \text{ and } Ki = 6.88 \text{e}^{-9}) \text{ using Sigma plot}^{\text{\tiny (B)}}$.

2.5. Alleviative effect of compound 12c in MPTP-induced motor impairment

To verify in vivo efficacy of the compound 12c as a MAO-B inhibitor, we conducted an animal model for Parkinson's disease generated by MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, administration. The dyskinesia in Parkinson's disease occurs when MPP⁺, 1-methyl-4phenylpyridinium, is converted by MAO-B in glia cells, cause the death of dopamine(DA)ergic neurons in substantia nigra pars compacta (SNpc). We conducted treatment of compound 12c (30 mg/kg, p.o. injection) for 3 consecutive days including before and after MPTP administration (4 injections of MPTP, 2 h intervals; 20 mg/kg, i.p. injection), and assessed whether treatment can restore motor ability (Fig. 5A). Commonly, transient weight loss is observed in the acute injection method of MPTP.²⁵ As shown in Fig. 5B, MPTP-treated mice showed weight loss compared to saline-treated control mice. In the case of treatment with compound 12c, the weight loss was restored more rapidly. In coat-hanger experiment, MPTP-induced mouse showed a falling or slipping to the first hanging position on the base of coat hanger (score 2.0 ± 0.58). In contrast to the vehicle-treated control mouse climbing to the top of the central vertical handle (score 4.75 \pm 0.25). In comparison, the compound 12c oral administrated mice recovered ascending ability to reach a stable position (score 3.83 ± 0.40) (Fig. 5C). Similarly, we found that mice treated with compound 12c though a vertical grid test showed significant improvement in motor impairment by measuring the total time spent on the test, the time of rotation at the top of the grid and time to go down (Fig. 5D). These results indicate that oral administration of compound 12c can inhibit MAO-B activity in the brain and alleviates the MPTP-induced motor dysfunction.



Fig. 5. Alleviation in MPTP-induced motor impairment by **12c** oral treatment A) The experimental protocol of MPTP treatment (20 mg/kg, *i.p.*) with or without **12c** compound (30 mg/kg, *p.o.*). Score diagram in coat hanger test and schematic diagram of vertical grid test. B) Weight change curve over time by MPTP treatment. C) Coat hanger score (F = 9.094, p = 0.005), D) Total time taken in vertical grid test (left, F = 10.87, p < 0.001), time to turn at the top of the grid (middle, F = 7.384, p = 0.003) and time to climb down (right, F = 15.55, P < 0.001). ***p < 0.001, **p < 0.01, *p < 0.05, and n.s., not significant (One-way ANOVA with *Tukey's* test), *n* refers to the number of mice. Data are means±s.e.m.

2.6. Protection against reduction of MPTP-induced tyrosine hydroxylase (TH)-positive dopamine neurons by oral treatment of compound 12c

To determine whether the recovery of motor ability by compound 12c was due to the protection of

DAergic neurons, we measured the amount of tyrosine hydroxylase (TH) by immunohistochemistry (**Fig. 6A**). TH is an enzyme involved in the synthesis of dopamine in DAergic neurons of SNpc. Thus, TH-positive neurons are considered markers of DAergic neurons. The number of TH-positive DAergic neurons in SNpc was significantly reduced by MPTP, whereas the number of TH-positive DAergic neurons in compound **12c**-treated group was reduced by only 38%. Moreover, although a sharp decrease in TH-immunoreactive fiber density was observed in striatum of MPTP treated mice (7% compared to vehicle-treated mice by density measurement analysis), the decrease was also prevented in mice administered with compound **12c** (**Fig. 6B**).



Fig. 6. Protection in MPTP-induced Tyrosine hydroxylase (TH) positive cell reduction by 12c oral treatment A) Representative images of immunostaining was conducted with TH on SNpc (substantia nigra pars compacta) and striatum sections. B) Left, TH-immuno-positive cells in SNpc were counted (F = 63.31, p < 0.001), right, The TH-immuno-positive fibers in the striatum were y Ai quantitated by optical density (F = 196, p < 0.001). ***p < 0.001 (One-way ANOVA with Tukey's

Conclusion

In this study, we synthesized a series of 4-(benzyloxy)phenyl and biphenyl-4-yl derivatives by modifying the benzyloxyphenyl moiety, a common structure of potent, selective, and reversible MAO-B inhibitors such as safinamide and sembragiline. Significant activity was observed for many of these compounds. Comparison of CF₃ substituted aryl regioisomers (2', 3', and 4') on the terminal aryl ring demonstrated that 4'-CF₃ derivatives exhibited the most potent inhibitory activity against MAO-B. The compound with the highest potency (**12c**) significantly protected the tyrosine hydroxylase (TH)-immunopositive DAergic neurons and attenuated the PD-associated behavioral deficits in the MPTP-induced mouse model of PD. In conclusion, we present a potent and selective MAO-B inhibitor with structural features that can be a good starting point for development of therapeutic agents for PD.

Experimental Section

General Methods. NMR spectra were obtained at either 400 MHz (¹H) and 100 MHz (¹³C) or 300 MHz(¹H) and 75 MHz (¹³C) using TMS as an internal standard. Chemical shifts (δ) are reported in parts per million (ppm) from tetramethylsilane (TMS). Reactions were monitored by analytical thinlayer chromatography (TLC) plates (Merck, Cat # 1.05715) and analyzed with 254 nm light. The reactions were purified by column chromatography using silica gel (Merck, Cat # 1.07734& 1.09385). All chemicals and solvents were reagent grade and used as obtained from commercial sources without further purification. Yields reported are for purified products and were not optimized. Compounds were checked by ¹H and ¹³C NMR.

General procedure for the preparation of (4-(benzyloxy)phenyl)methanaminium chloride derivatives (Method A). To a THF (400 mL) solution of LiAlH₄ (3.0 equiv) was added a THF (30 mL) solution of the 4-(benzyloxy)benzonitrile derivative dropwise at 0 °C. The reaction mixture was stirred at room temperature (16 h). Distilled water (4 mL) was added dropwise at 0 °C followed by an aqueous NaOH solution (2 mL, 15% w/w), then distilled water (4 mL) again. The product mixture was stirred at room temperature (1-2 h) for quenching the reaction, and was filtered through celite. The filtered solution was concentrated in vacuo. The product residue was dissolved in CH₂Cl₂, and 4.0M HCl in dioxane was added dropwise for salt formation. The product mixture was evaporated in vacuo and washed with EtOAc and *n*-hexane.

General procedure for the preparation of 4-(benzyloxy)benzamide derivatives (Method B). To a 4-(benzyloxy)benzonitrile derivative dissolved in *tert*-butanol was added potassium hydroxide and reflux for 2 h. After the reaction is completed, product mixture is extracted with EtOAc and brine

(200 mL x 3). Organic layer is dried with sodium sulfate. The residue is recrystallized.

General procedure for the preparation of 4-(benzyloxy)benzenaminium chloride (Method C). To a 1-(benzyloxy)-4-nitrobenzene derivative dissolved in MeOH was added Platinum (II) oxide. The reaction mixture was stirred for 1-2 h under H_2 gas at room temperature. The product mixture was filtered through celite and evaporated in vacuo. To a product residue dissolved in EtOAc was added 4.0M HCl in dioxane for salt formation. The product is washed with MeOH and ether.

General procedure for the preparation of 2-(4-(benzyloxy)phenyl)ethan-1-aminium chloride (Method D). To a *tert*-butyl (4-(benzyloxy)phenethyl)carbamate derivative dissolved in CH_2Cl_2 was added 4.0M HCl in dioxane and stirred for overnight at room temperature. After the reaction, the product mixture was evaporated in vacuo and washed with EtOAc and *n*-hexane.

General Procedure for the preparation of 2-([1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride derivatives (Method E). A mixture of *N*-boc-2-(4-bromophenyl)ethylamine, the desired arylboronic acid (a-m) (1.2 equiv), tetrakis(triphenylphosphine)palladium(0) (0.04 equiv), Na₂CO₃ (5 equiv) in degassed toluene/H₂O (5/2) was refluxed for 18 h. The reaction mixture was filtered through celite and concentrated in vacuo. The resulting residue was dissolved in in EtOAc (200 mL), washed with H₂O (200 mL× 2) and brine (200 mL). The organic layer was dried with anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on SiO₂.

Preparation of (4-(benzyloxy)phenyl)methanaminium chloride (7a). Using Method A, 6a (0.50 g,

2.4 mmol), LiAlH₄ (0.27 g, 7.2 mmol), and NaOH (0.2 mL) gave **7a** as a white solid (0.60 g, 100%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 3.91 (s, CH₂NH₃), 5.13 (s, OCH₂), 7.03 (d, J = 8.6 Hz, 2 ArH), 7.29-7.35 (m, 1 ArH), 7.36-7.41 (m, 3 ArH), 7.41-7.46 (m, 3 ArH), 8.40 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 41.5 (CH₂NH₂), 69.2 (OCH₂), 114.8, 126.3, 127.7, 128.4, 130.5, 130.7, 137.0 (11 ArC), 158.3 (1 ArC).

Preparation of (4-((2-(trifluoromethyl)benzyl)oxy)phenyl)methanaminium chloride (7b). Using Method A, **6b** (1.50 g, 5.4 mmol), LiAlH₄ (0.62 g, 16.2 mmol), and NaOH (0.6 mL) gave **7b** as a white solid (0.52 g, 34%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.95 (s, CH₂NH₃), 5.25 (s, OCH₂), 7.05 (d, *J* = 8.6 Hz, 2 ArH), 7.42 (d, *J* = 8.6 Hz, 2 ArH), 7.57-7.62 (m, 1 ArH), 7.69-7.78 (m, 2 ArH), 7.78-7.83 (m, 1 ArH), 8.24 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 41.6 (CH₂NH₂), 66.3 (OCH₂), 114.7, 124.3 (CF₃, q, *J*_{C-F} = 275.5 Hz), 126.2, 126.9 (q, *J*_{C-F} = 31.7 Hz), 128.8, 130.5, 130.7, 132.8, 134.8 (11 ArC), 158.1 (1 ArC).

Preparation of (4-((3-(trifluoromethyl)benzyl)oxy)phenyl)methanaminium chloride (7c). Using Method A, **6c** (0.50 g, 1.8 mmol), LiAlH₄ (0.20 g, 5.4 mmol), and NaOH (0.2 mL) gave **7c** as a white solid (0.20 g, 40%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.94 (s, CH₂NH₃), 5.24 (s, OCH₂), 7.07 (d, *J* = 8.7 Hz, 2 ArH), 7.40 (d, *J* = 8.6 Hz, 2 ArH), 7.61-7.67 (m, 1 ArH), 7.68-7.73 (m, 1 ArH), 7.74-7.78 (m, 1 ArH), 7.80-7.82 (m, 1 ArH), 8.13 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 41.6 (CH₂NH₂), 68.3 (OCH₂), 114.8, 124.0, 124.2 (CF₃, q, *J*_{C-F} = 270.5 Hz), 124.6, 126.5, 129.2 (q, *J*_{C-F} = 31.3 Hz), 129.6, 130.6, 131.6, 138.6 (11 ArC), 158.1 (1 ArC).

Preparation of (4-((4-(trifluoromethyl)benzyl)oxy)phenyl)methanaminium chloride (7d). Using

Method A, **6d** (0.3 g, 1.1 mmol), LiAlH₄ (0.12 g, 3.2 mmol), and NaOH (0.12 mL) gave **7d** as a white solid (0.18 g, 59%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 3.93 (s, CH₂NH₃), 5.26 (s, OCH₂), 7.06 (d, J = 8.5 Hz, 2 ArH), 7.41 (d, J = 8.5 Hz, 2 ArH), 7.63-7.79 (m, 4 ArH); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 41.5 (CH₂NH₂), 68.2 (OCH₂), 114.8, 124.2 (CF₃, q, $J_{C-F} = 271.6$ Hz), 125.3 (q, $J_{C-F} = 3.8$ Hz), 126.5, 127.9, 128.3 (q, $J_{C-F} = 31.5$ Hz), 130.6, 141.9 (11 ArC), 158.0 (1 ArC).

Preparation of (4-((2-fluorobenzyl)oxy)phenyl)methanaminium chloride (7e). Using Method A, **6e** (0.50 g, 2.2 mmol), LiAlH₄ (0.25 g, 6.6 mmol), and NaOH (0.2 mL) gave **7e** as a white crystalline (0.08 g, 16%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.92 (s, CH₂NH₃), 5.15 (s, OCH₂), 7.05 (d, *J* = 8.5 Hz, 2 ArH), 7.20-7.30 (m, 2 ArH), 7.38-7.46 (m, 3 ArH), 7.55 (t, *J* = 7.6 Hz, 1 ArH), 8.40 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 41.6 (CH₂NH₂), 63.6 (OCH₂), 114.7, 115.4 (d, *J*_{C-F} = 20.7 Hz), 123.7 (d, *J*_{C-F} = 14.3 Hz), 124.5, 126.7, 130.4, 130.5, 130.6, 130.7 (10 ArC), 158.2 (1 ArC), 160.4 (1 ArC, d, *J*_{C-F} = 244.7 Hz).

Preparation of (4-((3-fluorobenzyl)oxy)phenyl)methanaminium chloride (7f). Using Method A, **6f** (3.00 g, 13.2 mmol), LiAlH₄ (1.50 g, 39.6 mmol), and NaOH (2.4 mL) gave **7f** as a white solid (2.70 g, 89%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.92 (s, CH₂NH₃), 5.16 (s, OCH₂), 7.01-7.07 (m, 2 ArH), 7.11-7.19 (m, 1 ArH), 7.24-7.30 (m, 2 ArH), 7.37-7.47 (m, 3 ArH), 8.13 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 44.6 (CH₂NH₂), 68.3 (OCH₂), 114.1 (d, *J*_{C-F} = 21.7 Hz), 114.3, 114.5, 114.6, 123.3, 123.4, 128.3, 130.4 (d, *J*_{C-F} = 8.2 Hz), 140.2 (10 ArC), 156.8 (1 ArC), 162.2 (1 ArC, d, *J*_{C-F} = 244.7 Hz).

Preparation of (4-((4-fluorobenzyl)oxy)phenyl)methanaminium chloride (7g). Using Method A, 6g (1.00 g, 4.4 mmol), LiAlH₄ (0.50 g, 13.2 mmol), and NaOH (0.40 mL) gave 7g as a white solid (1.0 g, 97%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 300 MHz) δ 3.92 (s, CH₂NH₃), 5.11 (s, OCH₂), 7.00-7.09 (m, 2 ArH), 7.16-7.29 (m, 2 ArH), 7.50-7.65 (m, 4 ArH), 8.45 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 45.0 (CH₂NH₂), 68.4 (OCH₂), 114.5, 115.2 (d, $J_{C-F} = 21.2$ Hz), 128.2, 129.8 (d, $J_{C-F} = 8.2$ Hz), 133.5, 136.5, 156.8 (11 ArC), 161.7 (1 ArC, d, $J_{C-F} = 242.0$ Hz).

Preparation of (4-((3-chlorobenzyl)oxy)phenyl)methanaminium chloride (7h). Using Method A, 6h (0.40 g, 1.6 mmol), LiAlH₄ (0.19 g, 4.9 mmol), and NaOH (0.16 mL) gave 7h as a white solid (0.20 g, 50%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 300 MHz) δ 3.94 (s, CH₂NH₃), 5.26 (s, OCH₂), 7.03-7.10 (m, 2 ArH), 7.36-7.43 (m, 2 ArH), 7.62-7.80 (m, 4 ArH), 8.15 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 42.1 (CH₂NH₂), 68.8 (OCH₂), 115.3, 125.8, 125.8, 126.5, 127.0, 128.2, 128.4, 128.6, 129.0, 129.4, 131.1, 142.4, 158.5 (12 ArC).

Preparation of (4-((4-chlorobenzyl)oxy)phenyl)methanaminium chloride (7i). Using Method A, 6i (0.40 g, 1.6 mmol), LiAlH₄ (0.19 g, 4.9 mmol), and NaOH (0.16 mL) gave 7i as a white solid (0.16 g, 39%): $\mathbf{R}_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 3.93 (s, CH₂NH₃), 5.14 (s, OCH₂), 7.01-7.07 (m, 2 ArH), 7.36-7.42 (m, 2 ArH), 7.44-7.49 (m, 4 ArH), 8.20 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 42.1 (CH₂NH₂), 68.8 (OCH₂), 115.3, 126.9, 128.9, 129.9, 131.0, 132.8, 136.5, 158.6 (12 ArC).

Preparation of (4-((3-methoxybenzyl)oxy)phenyl)methanaminium chloride (7j). Using Method A, 6j (1.00 g, 4.2 mmol), LiAlH₄ (0.48 g, 12.5 mmol), and NaOH (0.4 mL) gave 7j as a white solid

(0.37 g 37%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 3.75 (s, OCH₃), 3.92 (s, CH₂NH₃), 5.10 (s, OCH₂), 6.85-6.91 (m, 1 ArH), 6.97-7.06 (m, 4 ArH), 7.26-7.33 (m, 1 ArH), 7.38-7.44 (m, 2 ArH), 8.35 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 42.0 (CH₂NH₂), 55.1 (OCH₃), 69.0 (OCH₂), 113.1, 113.2, 114.6, 114.8, 119.7, 127.3, 128.3, 129.6, 130.4, 138.6, 158.2, 159.4 (12 ArC).

Preparation of (4-((4-methoxybenzyl)oxy)phenyl)methanaminium chloride (7k). Using Method A, 6k (0.30 g, 1.3 mmol), LiAlH₄ (0.14 g, 3.8 mmol), and NaOH (0.12 mL) gave 7k as a white solid (0.25 g, 71%): R_f = 0.00 (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 3.75 (s, OCH₃), 3.93 (s, CH₂NH₃), 5.09 (s, OCH₂), 6.90-7.06 (m, 4 ArH), 7.32-7.43 (m, 4 ArH), 8.26 ((s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 42.1 (CH₂NH₂), 55.6 (OCH₃), 69.4 (OCH₂), 114.3, 115.3, 126.5, 129.3, 129.9, 131.0, 159.5, 162.3 (12 ArC).

Preparation of 4-((2-(trifluoromethyl)benzyl)oxy)benzamide (8a). Using Method B, **6b** (0.50 g, 1.8 mmol) and potassium hydroxide (0.76 g, 13.5 mmol) gave **8a** as a baige solid (0.47 g, 89%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (CDCl₃, 400 MHz) δ 5.33 (s, OCH₂), 6.98-7.05 (m, 2 ArH), 7.41-7.48 (m, 1 ArH), 7.55-7.61 (m, 1 ArH), 7.69-7.75 (m, 2 ArH), 7.77-7.83 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 66.8 (OCH₂), 114.6, 120.7, 124.8 (CF₃, q, *J*_{C-F} = 272.2 Hz), 126.7 (q, *J*_{C-F} = 5.7 Hz), 127.0, 127.5 (q, *J*_{C-F} = 30.3 Hz), 127.6, 127.9, 128.9, 129.3, 129.9, 131.1, 133.3, 135.0 (11 ArC), 160.8 (1 ArC), 167.8 (C(O)).

Preparation of 4-((3-(trifluoromethyl)benzyl)oxy)benzamide (8b). Using Method B, **6c** (0.20 g, 0.7 mmol) and potassium hydroxide (0.30 g, 5.4 mmol) gave **8b** as a white solid (0.19 g, 91%): $R_f =$

0.00 (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.28 (s, OCH₂), 7.06-7.14 (m, 2 ArH), 7.16-7.27 (m, 1 ArH), 7.62-7.70 (m, 1 ArH), 7.70-7.76 (m, 1 ArH), 7.77-7.82 (m, 1 ArH), 7.82-7.93 (m, 2 ArH, C(O)NH₂); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 68.4 (OCH₂), 114.3, 124.1, 124.1, 124.2 (CF₃, q, *J*_{C-F} = 270.7 Hz), 124.6, 127.0, 129.2 (q, *J*_{C-F} = 29.2 Hz), 129.6, 131.7, 138.3 (11 ArC), 160.4 (1 ArC), 167.4 (C(O)).

Preparation of 4-((4-(trifluoromethyl)benzyl)oxy)benzamide (8c). Using Method B, 6d (0.30 g, 1.1 mmol) and potassium hydroxide (0.45 g, 8.1 mmol) gave 8c as a brown solid (0.29 g, 92%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.30 (s, OCH₂), 7.05-7.14 (m, 2 ArH), 7.16-7.27 (m, 1 ArH), 7.66-7.74 (m, 2 ArH), 7.76-7.82 (m, 2 ArH), 7.82-7.90 (m, 1 ArH, C(O)NH₂); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 68.9 (OCH₂), 114.7, 124.7 (CF₃, q, *J*_{C-F} = 272.0 Hz), 125.8, 125.8, 127.4, 128.5, 128.9 (q, *J*_{C-F} = 31.6 Hz), 129.9, 142.1 (11 ArC), 160.8 (1 ArC), 167.8 (C(O)).

Preparation of 4-((2-fluorobenzyl)oxy)benzamide (8d). Using Method B, **6e** (0.50 g, 2.2 mmol) and potassium hydroxide (0.93 g, 16.5 mmol) gave **8d** as a brown powder (0.50 g, 93%): $R_f = 0.00$ (EtOAc **1** : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.20 (s, OC**H**₂), 7.06-7.11 (m, 2 Ar**H**), 7.18-7.30 (m, 3 Ar**H**), 7.40-7.48 (m, 1 Ar**H**), 7.55-7.61 (m, C(O)N**H**H), 7.83-7.89 (m, 2Ar**H**, C(O)NH**H**); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 63.6 (OCH₂), 114.1, 115.4 (d, *J*_{C-F} = 20.8 Hz), 123.4, 123.6, 124.5, 124.6, 126.9, 129.4, 130.5 (d, *J*_{C-F} = 8.3 Hz), 130.8, 130.8, 160.4 (d, *J*_{C-F} = 244.9 Hz), 160.5 (12 Ar**C**), 167.3 (**C**(O)).

Preparation of 4-((3-fluorobenzyl)oxy)benzamide (8e). Using Method B, 6f (2.0 g, 8.8 mmol) and

potassium hydroxide (3.70 g, 66.0 mmol) gave **8e** as a beige crystalline (1.89 g, 88%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO- d_6 , 400 MHz) δ 5.19 (s, OCH₂), 7.04-7.10 (m, 2 ArH), 7.13-7.26 (m, 2 ArH), 7.27-7.34 (m, 2 ArH), 7.40-7.48 (m, 1 ArH), 7.83-7.90 (m, 1 ArH, C(O)NH₂); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 68.5 (OCH₂), 114.3, 114.4, 114.7 (d, $J_{C-F} = 20.8$ Hz), 123.6, 126.9, 129.4, 130.5 (d, $J_{C-F} = 8.2$ Hz), 139.7, 139.7, 160.4, 162.2 (d, $J_{C-F} = 242.2$ Hz) (12 ArC), 167.38 (C(O)).

Preparation of 4-((4-fluorobenzyl)oxy)benzamide (8f). Using Method B, **6g** (0.3 g, 1.3 mmol) and potassium hydroxide (0.56 g, 9.9 mmol) gave **8f** as a white crystalline (0.32 g, 98%): $\mathbf{R}_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.14 (s, OCH₂), 7.02-7.08 (m, 2 ArH), 7.15-7.26 (m, 3 ArH), 7.47-7.55 (m, 2 ArH), 7.80-7.87 (m, 1 ArH, C(O)NH₂); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 69.0 (OCH₂), 114.7, 115.7 (d, *J*_{C-F} = 21.2 Hz), 127.2, 129.8, 130.5 (d, *J*_{C-F} = 8.2 Hz), 133.4, 161.0, 162.3 (d, *J*_{C-F} = 242.3 Hz) (12 ArC), 167.8 (C(O)).

Preparation of 4-((2-(trifluoromethyl)benzyl)oxy)benzenaminium chloride (10a). Using Method C, 9a (0.20 g, 0.7 mmol), platinum oxide, (0.016 g, 0.7 mmol), and 4.0M hydrogen chloride gave 10a as a white powder (0.11 g, 53%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.25 (s, OCH₂), 7.09-7.14 (m, 2 ArH), 7.29-7.34 (m, 2 ArH), 7.57-7.63 (m, 1 ArH), 7.70-7.79 (m, 2 ArH), 7.79-7.83 (m, 1 ArH), 10.02 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 67.0 (OCH₂), 116.1, 124.7 (CF₃, q, *J*_{C-F} = 272.1 Hz), 125.0, 125.4, 126.6, 126.7, 127.4 (q, *J*_{C-F} = 30.3 Hz), 129.3, 131.1, 133.3, 135.0, 157.9 (12 ArC).

Preparation of 4-((3-(trifluoromethyl)benzyl)oxy)benzenaminium chloride (10b). Using Method

C, **9b** (0.20 g, 0.7 mmol), platinum oxide, (0.016 g, 0.7 mmol), and 4.0M hydrogen chloride gave **10b** as a white powder (0.13 g, 63%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.24 (s, OCH₂), 7.12-7.17 (m, 2 ArH), 7.30-7.36 (m, 2 ArH), 7.62-7.84 (m, 4 ArH), 10.13 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.6 (OCH₂), 115.8, 124.2 (CF₃, q, *J*_{C-F} = 270.9 Hz), 124.1, 124.5, 124.6, 124.7, 129.2 (q, *J*_{C-F} = 31.4 Hz), 129.6, 131.7, 138.3, 157.5 (12 ArC).

Preparation of 4-((4-(trifluoromethyl)benzyl)oxy)benzenaminium chloride (10c). Using Method C, **9c** (0.20 g, 0.7 mmol), platinum oxide, (0.016 g, 0.7 mmol), and 4.0M hydrogen chloride gave **10c** as a white powder (0.15 g, 72%): $R_f = 0.00$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (CDCl₃, 400 MHz) δ 5.26 (s, OCH₂), 7.11-7.16 (m, 2 ArH), 7.30-7.35 (m, 2 ArH), 7.65-7.70 (m, 2 ArH), 7.75-7.80 (m, 2 ArH), 10.15 (s, NH₃); ¹³C NMR (DMSO-*d*₆.75 MHz) δ 69.1 (OCH₂), 116.2, 124,7 (CF₃, q, *J*_{C-F} = 270.5 Hz), 125.0, 125.3, 125.8 (q, *J*_{C-F} = 3.8 Hz), 128.5, 128.8 (q, *J*_{C-F} = 31.4 Hz), 142.1, 157.9 (12 ArC).

Preparation of 4-((2-fluorobenzyl)oxy)benzenaminium chloride (10d). Using Method C, **9d** (0.15 g, 0.6 mmol), platinum oxide, (0.012 g, 0.06 mmol), and 4.0M hydrogen chloride gave **10d** as a white solid (0.04 g, 30%): $R_f = 0.59$ (5% CH₂Cl₂/MeOH): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.10 (s, OCH₂), 7.08-7.14 (m, 2 ArH), 7.19-7.26 (m, 2 ArH), 7.29-7.35 (m, 2 ArH), 7.47-7.54 (m, 2 ArH) 10.19 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.8 (OCH₂), 115.3 (d, *J*_{C-F} = 21.3 Hz), 115.3, 115.5, 115.7, 115.7, 123.4, 123.6, 124.5, 124.6, 124.7, 130.0 (d, *J*_{C-F} = 8.2 Hz), 130.5, 130.6, 130.7, 130.8, 133.0, 157.6, 157.6, 160.4 (d, *J*_{C-F} = 245.0 Hz) (12 ArC).

Preparation of 4-((3-fluorobenzyl)oxy)benzenaminium chloride (10e). Using Method C, 9e (0.10

g, 0.4 mmol) and platinum oxide, (0.008 g, 0.04 mmol), and 4.0M hydrogen chloride gave **10e** as a white solid (0.38 g, 71%): $R_f = 0.33$ (EtOAc 1 : *n*-Hexane 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.15 (s, OCH₂), 7.09-7.20 (m, 3 ArH), 7.26-7.34 (m, 4 ArH), 7.41-7.48 (m, 1ArH), 10.06 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.6 (OCH₂), 114.2 (d, *J*_{C-F} = 21.8 Hz), 114.7 (d, *J*_{C-F} = 20.8 Hz), 115.7, 123.5, 123.5, 124.5, 124.7, 130.5 (d, *J*_{C-F} = 8.3 Hz), 139.6, 139.7, 161.9, 162.2 (d, *J*_{C-F} = 242.2 Hz) (12 ArC).

Preparation of 4-((4-fluorobenzyl)oxy)benzenaminium chloride (10f). Using Method C, **9f** (0.10 g, 0.4 mmol), platinum oxide, (0.008 g, 0.04 mmol), and 4.0M hydrogen chloride gave **10f** as a white solid (0.04 g, 30%): $R_f = 0.59$ (5% CH₂Cl₂/MeOH): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.10 (s, OCH₂), 7.08-7.13 (m, 2 ArH), 7.19-7.26 (m, 2 ArH), 7.26-7.31 (m, 2ArH), 7.47-7.53 (m, 2 ArH), 9.93 (s, NH₃) ; ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.8 (OCH₂), 115.3 (d, *J*_{C-F} = 21.3 Hz), 115.7, 124.4, 124.6, 130.0 (d, *J*_{C-F} = 8.2 Hz), 132.9, 157.6, 161.8 (d, *J*_{C-F} = 242.4 Hz) (12 ArC).

Preparation of 2-(4-((2-(trifluoromethyl)benzyl)oxy)phenyl)ethan-1-aminium chloride (12a). Using Method D, 11a (0.50 g, 1.3 mmol) and 4.0M HCl in dioxane (2.3 g, 63.0 mmol) gave 12a as a white cotton form (0.38 g, 90%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.77-2.84 (m, NH₃CH₂CH₂), 2.96-3.03 (m, NH₃CH₂CH₂), 5.20 (s, OCH₂), 6.98-7.04 (m, 2 ArH), 7.18-7.23 (m, 2 ArH), 7.62-7.83 (m, 4 ArH), 7.86 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.5 (NCH₂CH₂), 66.6 (OCH₂), 115.3, 124.8 (CF₃, q, $J_{C-F} = 272.9$ Hz), 126.5, 126.7, 127.3 (q, $J_{C-F} = 30.3$ Hz), 129.2, 130.3, 130.5, 130.8, 133.3, 135.5, 157.3 (12 ArC).

Preparation of 2-(4-((3-(trifluoromethyl)benzyl)oxy)phenyl)ethan-1-aminium chloride (12b). Using Method D, **11b** (0.50 g, 1.3 mmol) and 4.0M HCl in dioxane (2.3 g, 63.0 mmol) gave **12b** as a

white crystalline (0.35 g, 84%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.73-2.83 (m, NH₃CH₂CH₂), 2.91-3.02 (m, NH₃CH₂CH₂), 5.20 (s, OCH₂), 6.90-7.02 (m, 2 ArH), 7.13-7.21 (m, 2 ArH), 7.60-7.82 (m, 4 ArH), 7.86 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.2 (NCH₂CH₂), 68.4 (OCH₂), 115.1, 124.0, 124.3 (CF₃, q, $J_{C-F} = 270.5$ Hz), 124.6, 129.3 (q, $J_{C-F} = 31.6$ Hz), 129.7, 129.9, 131.7, 138.8, 157.0 (12 ArC).

Preparation of 2-(4-((4-(trifluoromethyl)benzyl)oxy)phenyl)ethan-1-aminium chloride (12c). Using Method D, 11c (0.50 g, 1.3 mmol) and 4.0M HCl in dioxane (2.3 g, 63.0 mmol) gave 12c as a white cotton form (0.35 g, 84%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.75-2.83 (m, NH₃CH₂CH₂), 2.93-3.01(m, NH₃CH₂CH₂), 5.22 (s, OCH₂), 6.93-7.01 (m, 2 ArH), 7.14-7.21 (m, 2 ArH), 7.61-7.69 (m, 2 ArH), 7.73-7.79 (m, 2 ArH), 7.90 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 32.5 (NCH₂CH₂), 68.7 (OCH₂), 115.4, 120.6, 124.7 (CF₃, q, $J_{C-F} = 270.4$ Hz), 125.7, 125.8, 128.3, 128.4, 128.6, 128.8, 129.0 (q, $J_{C-F} = 31.5$ Hz), 130.2, 130.3, 142.6, 157.3 (12 ArC); HPLC purity: 10.6 min, > 98% at 254 & 280 nm; LRMS (M + H)⁺ (ESI⁺) 296.1 [M + H]⁺ (calcd for C₁₆H₁₆F₃NOH⁺ 296.1).

Preparation of 2-(4-((2-fluorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12d). Using Method D, 11d (0.90 g, 2.6 mmol) and 4.0M HCl in dioxane (3.91 mL, 15.6 mmol) gave 12d as a white solid (0.73 g, 99%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.84-3.00 (m, NH₃CH₂CH₂), 5.12 (s, OCH₂), 7.00 (d, J = 8.5 Hz, 2 ArH), 7.53-7.57 (m, 6 ArH), 8.24 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.5 (NCH₂CH₂), 64.0 (OCH₂), 115.3, 115.8 (d, $J_{C-F} = 21.0$ Hz), 124.4 (d, $J_{C-F} = 14.5$ Hz), 125.0 (d, $J_{C-F} = 3.4$ Hz), 130.2. 130.3, 130.8 (d, $J_{C-F} = 8.2$ Hz), 131.1 (d, $J_{C-F} = 4.0$ Hz), 157.5, 160.8 (d, $J_{C-F} = 244.5$ Hz) (12 ArC).

Preparation of 2-(4-((3-fluorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12e). Using Method D, **11e** (0.20 g, 0.6 mmol) and 4.0M HCl in dioxane (1.01 mL, 29.0 mmol) gave **12e** as a white powder (0.16 g, 100%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.77-3.02 (m, NH₃CH₂CH₂), 5.12 (s, OCH₂), 6.89-7.01 (m, 2 ArH), 7.07-7.22 (m, 3 ArH), 7.23-7.31 (m, 2 ArH), 7.38-7.49 (m, 1 ArH), 8.04 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.1 (NCH₂CH₂), 68.3 (OCH₂), 114.1 (d, $J_{C-F} = 21.7$ Hz), 114.5 (d, $J_{C-F} = 20.7$ Hz), 114.9, 123.4, 129.7, 130.5 (d, $J_{C-F} = 8.2$ Hz), 140.1, 140.2, 156.9, 162.2 (d, $J_{C-F} = 242.0$ Hz) (12 ArC).

Preparation of 2-(4-((4-fluorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12f). Using Method D, **11f** (0.50 g, 1.5 mmol) and 4.0M HCl in dioxane (2.50 mL, 72.4 mmol) gave **12f** as a white cotton form (0.40 g, 98%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.80-3.01 (m, NH₃CH₂CH₂), 5.06 (s, OCH₂), 6.90-7.00 (m, 2 ArH), 7.10-7.30 (m, 4 ArH), 7.44-7.55 (m, 2 ArH), 8.22 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.5 (NHCH₂CH₂), 68.9 (OCH₂), 115.4, 115.7 (d, $J_{C-F} = 21.2$ Hz), 130.1, 130.2, 130.3 (d, $J_{C-F} = 8.2$ Hz), 133.8, 133.9, 157.5, 162.2 (d, $J_{C-F} = 241.8$ Hz) (12 ArC).

Preparation of 2-(4-((2-chlorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12g). Using Method D, 11g (1.15 g, 3.2 mmol) and 4.0M HCl in dioxane (4.77 mL, 19.1 mmol) gave 12g as a white solid (0.95 g, 99%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.85-3.00 (m, NH₃CH₂CH₂), 5.14 (s, OCH₂), 7.00 (d, J = 8.3 Hz, 2 ArH), 7.21 (d, J = 8.2 Hz, 2 ArH), 7.38-7.40 (m, 2 ArH), 7.50-7.60 (m, 2 ArH), 8.26 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.5 (NCH₂CH₂), 67.3 (OCH₂), 115.3, 127.8, 128.9, 129.8, 130.2, 130.3, 130.4, 130.5, 133.0, 134.8,

157.5 (12 ArC).

Preparation of 2-(4-((3-chlorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12h). Using Method D, 11h (0.50 g, 1.4 mmol) and 4.0M HCl in dioxane (2.4 mL, 69.1 mmol) gave 12h as a white powder (0.39 g, 95%): $R_f = 0.25$ (15% CH₂Cl₂/MeOH): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.79-3.01 (m, NH₃CH₂CH₂), 5.11 (s, OCH₂), 6.93-7.00 (m, 2 ArH), 7.15-7.23 (m, 2 ArH), 7.33-7.48 (m, 3 ArH), 7.49-7.54 (m, 1 ArH), 8.17 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.5 (NHCH₂CH₂), 68.7 (OCH₂), 115.4, 126.5, 127.6, 128.1, 130.2, 130.3, 130.8, 133.6, 140.3, 157.4 (12 ArC).

Preparation of 2-(4-((4-chlorobenzyl)oxy)phenyl)ethan-1-aminium chloride (12i). Using Method D, **11i** (0.50 g, 1.4 mmol) and 4.0M HCl in dioxane (2.4 mL, 69.1 mmol) gave **12i** as a white powder (0.39 g, 94%): $R_f = 0.25$ (15% CH₂Cl₂/MeOH): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.80-3.00 (m, NH₃CH₂CH₂), 5.08 (s, OCH₂), 6.93-6.99 (m, 2 ArH), 7.15-7.21 (m, 2 ArH), 7.42-7.50 (m, 4 ArH), 8.18 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.0 (NHCH₂CH₂), 68.3 (OCH₂), 114.9, 128.4, 129.4, 129.7, 132.3, 136.2, 156.9, 161.8 (12 ArC).

Preparation of 2-(4-((3-methoxybenzyl)oxy)phenyl)ethan-1-aminium chloride (12j). Using Method D, 11j (1.20 g, 3.4 mmol) and 4.0M HCl in dioxane (5.04 mL, 20.2 mmol) gave 12j as a white solid (0.92 g, 93%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.81-2.96 (m, NH₃CH₂CH₂), 3.75 (OCH₃), 5.06 (s, OCH₂), 6.87-7.01 (m, 5 ArH), 7.16-7.19 (m, 2 ArH), 7.27-7.33 (m, 1 ArH), 8.16 (s, NH₃); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 32.1 (NCH₂CH₂), 55.0 (OCH₃), 69.0 (OCH₂), 113.1, 114.9, 119.6, 129.5, 129.7, 138.7, 157.1, 159.3 (12 ArC).

Preparation of 2-(3'-(trifluoromethyl)-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14a). Using Method E, 13 (1.00 g, 3.3 mmol), 3-(trifluoromethyl)phenylboronic acid (0.76 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave 14a as a white solid (0.57 g, 56%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.98-3.10 (m, NH₃CH₂CH₂), 7.42 (d, J = 8.2 Hz, 2 ArH), 7.69-7.74 (m, 4 ArH), 7.95-8.00 (m, 2 ArH), 8.31 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 33.0 (NCH₂CH₂), 123.4 (q, $J_{C-F} = 3.9$ Hz), 124.4 (q, $J_{C-F} = 3.7$ Hz), 124.7 (CF₃, q, $J_{C-F} = 270.9$ Hz), 127.6, 129.9, 130.2 (q, $J_{C-F} = 31.4$ Hz), 130.5, 131.1, 137.4, 138.1, 141.4 (12 ArC).

Preparation of 2-(4'-(trifluoromethyl)-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14b). Using Method E, **13** (0.50 g, 1.7 mmol), 4-(trifluoromethyl)phenylboronic acid (0.38 g, 2.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.08 g, 0.1 mmol), Na₂CO₃ (0.89 g, 8.3 mmol) in toluene/H₂O (16 ml/6.6 ml), followed by 4.0 M HCl in dioxane (1.25 ml, 5.0 mmol) gave **14b** as a white solid (0.28 g, 56%): $\mathbf{R}_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.01-3.11 (m, NH₃CH₂CH₂), 7.44 (d, *J* = 8.1 Hz, 2 ArH), 7.71-7.91 (m, 6 ArH), 8.37 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 33.0 (NCH₂CH₂), 124.8 (CF₃, q, *J*_{C-F} = 270.1 Hz), 126.2 (q, *J*_{C-F} = 3.8 Hz), 127.6, 127.7, 128.2 (q, *J*_{C-F} = 31.7 Hz), 129.9, 137.4, 138.4, 144.3 (12 ArC).

Preparation of 2-(2'-fluoro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14c). Using Method E, 13 (1.00 g, 3.3 mmol), 2-fluorophenylboronic acid (0.56 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol) and Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave 14c as a white solid (0.36 g, 43%); $R_f = 0.00$ (EtOAc 9 : acetone 1); ¹H NMR (DMSO- d_6 , 400 MHz) δ 2.89-

3.09 (m, NH₃CH₂CH₂), 7.24-7.52 (m, 8 ArH), 8.32 (s, NH₃); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 33.1 (NCH₂CH₂), 116.6 (d, $J_{C-F} = 22.4$ Hz), 120.3, 125.4 (d, $J_{C-F} = 3.5$ Hz), 129.4, 129.5, 129.9 (d, $J_{C-F} = 8.2$ Hz), 131.1 (d, $J_{C-F} = 3.3$ Hz), 131.5, 131.9, 137.5 (d, $J_{C-F} = 21.9$ Hz), 159.6 (d, $J_{C-F} = 244.2$ Hz) (12 ArC).

Preparation of 2-(3'-fluoro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14d). Using Method E, 13 (1.00 g, 3.3 mmol), 3-fluorophenylboronic acid (0.56 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave 14d as a white solid (0.52 g, 62%); $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.98-3.09 (m, NH₃CH₂CH₂), 7.17-7.52 (m, 6 ArH), 7.68 (d, *J* = 8.0 Hz, 2 ArH), 8.34 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 33.0 (NCH₂CH₂), 113.6 (d, *J*_{C-F} = 21.8 Hz), 114.5 (d, *J*_{C-F} = 20.9 Hz), 123.0 (d, *J*_{C-F} = 2.4 Hz), 127.4, 129.8, 131.3 (d, *J*_{C-F} = 8.5 Hz), 137.6 (d, *J*_{C-F} = 2.1 Hz), 137.9, 142.8 (d, *J*_{C-F} = 7.7 Hz), 163.2 (d, *J*_{C-F} = 241.7 Hz) (12 ArC).

Preparation of 2-(4'-fluoro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14e). Using Method **E**, **13** (1.00 g, 3.3 mmol), 4-fluorophenylboronic acid (0.56 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave **14e** as a white solid (0.84 g, 67%); $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.96-3.08 (m, NH₃CH₂CH₂), 7.27-7.31 (m, 2 ArH), 7.37 (d, *J* = 8.1 Hz, 2 ArH), 7.60-7.71 (m, 4 ArH), 8.30 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 33.0 (NCH₂CH₂), 116.2 (d, *J*_{C-F} = 21.2 Hz), 127.3, 129.0 (d, *J*_{C-F} = 8.1 Hz), 129.7, 136.8 (d, *J*_{C-F} = 3.1 Hz), 137.2, 138.0, 162.3 (d, *J*_{C-F} = 242.7 Hz) (12 Ar**C**).
Preparation of 2-(2'-chloro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14f). Using Method E, 13 (1.00 g, 3.3 mmol), 2-chlorophenylboronic acid (0.63 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol) and Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave 14f as a white solid (0.47 g, 53%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.90-3.11 (m, NH₃CH₂CH₂), 7.24-7.64 (m, 8 ArH), 8.33 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 33.0 (NCH₂CH₂), 129.0, 129.7, 136.8, 137.2, 138.0, 162.3 (12 ArC).

Preparation of 2-(3'-chloro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14g). Using Method E, 13 (1.00 g, 3.3 mmol), 3-chlorophenylboronic acid (0.63 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave 14g as a white solid (0.84 g, 67%): $\mathbf{R}_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.93-3.09 (m, NH₃CH₂CH₂), 7.25-7.82 (m, 8 ArH), 8.33 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 33.1 (NCH₂CH₂), 128.0, 129.0, 129.4, 129.6, 129.7, 129.9, 130.3, 131.5, 131.7, 131.9, 137.6, 140.0 (12 ArC).

Preparation of 2-(4'-chloro-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14h). Using Method E, **13** (1.50 g, 5.0 mmol), 4-chlorophenylboronic acid (0.94 g, 6.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.23 g, 0.2 mmol), Na₂CO₃ (2.65 g, 25.0 mmol) in toluene/H₂O (50 ml/20 ml), followed by 4.0 M HCl in dioxane (3.75 ml, 15.0 mmol) gave **14h** as a white solid (0.94 g, 69%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.99-

3.06 (m, NH₃CH₂CH₂), 7.37-7.77 (m, 8 ArH), 8.31 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 33.0 (NCH₂CH₂), 127.3, 128.7, 129.3, 129.8, 132.7, 137.6, 137.7, 139.1 (12 ArC).

Preparation of 2-(3'-methoxy-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14i). Using Method E, **13** (1.00 g, 3.3 mmol), 3-methoxyphenylboronic acid (0.61 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave **4i** as a white solid (0.47 g, 53%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 2.93-3.07 (m, NH₃CH₂CH₂), 3.79 (OCH₃), 7.01 (d, *J* = 8.7 Hz, 1 ArH), 7.33 (d, *J* = 8.1 Hz, 2 ArH), 7.56-7.60 (m, 4 ArH), 8.27 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 33.0 (NCH₂CH₂), 55.6 (CH₃), 114.8, 126.8, 128.1, 129.7, 132.7, 136.3, 138.7, 159.3 (OArC) (12 ArC).

Preparation of 2-(4'-methoxy-[1,1'-biphenyl]-4-yl)ethan-1-amine hydrochloride (14j). Using Method E, **13** (1.00 g, 3.3 mmol), 4-methoxyphenylboronic acid (0.61 g, 4.0 mmol), tetrakis(triphenylphosphine)palladium(0) (0.15 g, 0.1 mmol), Na₂CO₃ (1.77 g, 16.7 mmol) in toluene/H₂O (33 ml/13.3 ml), followed by 4.0 M HCl in dioxane (2.50 ml, 10.0 mmol) gave **14j** as a white solid (0.52 g, 58%): $R_f = 0.00$ (EtOAc 9 : acetone 1): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.02-3.05 (m, NH₃CH₂CH₂), 3.82 (OCH₃), 6.93 (d, *J* = 8.1 Hz, 1 ArH), 7.18-7.23 (m, 2 ArH), 7.35-7.40 (m, 3 ArH), 7.64 (d, *J* = 6.8 Hz, 2 ArH), 8.37 (s, NH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 33.0 (NCH₂CH₂), 55.6 (CH₃), 112.5, 113.4, 119.3, 127.4, 129.7, 130.4, 137.4, 138.9, 141.9, 160.2 (OArC) (12 ArC).

Monoamine oxidase (MAO) enzyme assay

The MAO enzyme inhibitory activity of the compounds was evaluated according to the method described previously.⁶ The MAO activity was carried out with a fluorescence-based method by measuring the effects of the compounds on the production of H₂O₂ from substrate (MAO-A: ptyramine, MAO-B: benzylamine, Sigma-Aldrich, Co., Louis, USA) using 10-acetyl-3,7dihydroxyphenoxazine (Amplex[®] Red, Molecular Probes, Eugene, Oregon, USA). The test compounds in final concentrations ranged from 10⁻⁵ µM to 10 µM with recombinant hMAO-A or hMAO-B (Sigma-Aldrich, Co., Louis, USA) in 100 µL of aqueous 50 mM of sodium phosphate buffer (pH 7.4) were preincubated for 15 min 37 °C in a black-clear bottom 96 well assay plate (Costar[®], Corning, NY, USA). The reaction was performed by adding 200 µM Amplex red, 2U/mL horseradish peroxidase (Sigma-Aldrich, Co., Louis, USA) and 2 mM p-tyramine for hMAO-A or 2 mM benzylamine (Sigma-Aldrich, Co., Louis, USA) for hMAO-B in 100 µL of aqueous 50 mM of sodium phosphate buffer (pH 7.4) (final volume of 200 µL). After 20 min of incubation at 37 °C, the assay was quantified with a multi-mode microplate reader (SpectraMax[®]i3, Molecular Device, CA, USA) based on the generated fluorescence of resorufin (excitation: 545 nm, emission: 590 nm). The IC_{50} values of the compounds were determined in quadruplicate from the dose response inhibition curves using SigmaPlot software version 13.0 as the mean \pm S.E.M.

Kinetic studies of MAO-B inhibition

To investigate the interaction mode of compound 12c, the type of enzyme inhibition was determined by Michaelis-Menten kinetic experiment. The catalytic rates of human MAO-B enzyme were measured at seven different concentrations of substrate benzylamine (0.065, 0.125, 0.25, 0.5, 1, 2, and 4 mM) in the absence and in the presence of three different concentrations (0.3, 1, and 3 nM) of compound 12c. The corresponding progression curves and the Lineweaver-Burk plots were generated using GraphPad Prism (GraphPad software). In addition, the maximal velocity (V_{max}), the

Michaelis constant (K_m) and the inhibition constant (K_i) were calculated using SigmaPlot software version 13.0.

Generation of the mouse model of Parkinson's disease induced by MPTP

All mice were housed in a temperature and humidity controlled environment with a 12 h light-dark cycle and were free to consume food and water. All animal care and handling was performed in accordance with the instructions of the KIST Institutional Animal Care and Use Committee (Seoul, Korea). For animal experiments, 10-week-old male C57Bl/6 mice weighing 23–25 g were used. All MPTP experiments used acute administration consisting of intraperitoneal injection of MPTP-HCl (M0896, Sigma-Aldrich, 2 mg/mL in saline, 20 mg/kg for one injection) at 2 h intervals. The mice (n = 5-8 per group) were orally administrated with compound **12c** suspended in 12.5% of mixture Castor oil with ethanol in the same ratio in distilled water at 30 mg/kg body weight per day for 24 h per 3 consecutive days. See also **Fig. 4A**.

Behavior test

The vertical grid test was performed as described in the previous study.⁴ A mouse was gently placed inside of the device 3 cm away from the top facing upward. All mice were habituated twice a day for two consecutive days prior to MPTP administration. This test was performed 6 days after MPTP administration, and all experimental sessions were video recorded for time measurement. For the coat-hanger test, the triangular-shaped coat-hanger consisted of a horizontal steel wire (diameter: 3 mm, length: 40 cm, height: 20 cm) with two diagonal side-bars (length: 20 cm; inclination: 35° from the horizontal axis). The mice were placed in a dangling posture in the middle of the horizontal wire. The motor coordination was rated (0) If the mouse fell off the hanger, (1) lifted both limbs, (2) reached one of the ends of the hanger, (3) rose above the diagonal edge, (4) reached the top corner of

triangle, and (5) reached the handle of coat-hanger. See also **Fig. 4A.** This assessment is a single test that lasts for 3 minutes or when the mouse falls off the wire or reach the hanger handle.

Immunostaining for TH

For histological analysis, the mice were anesthetized with 2% avertin (20 μ g/g, *i.p.*) and perfused with 0.9% saline, and fixed by perfusion with 4% paraformaldehyde (PFA). The isolated brain postfixed with 4% PFA for overnight at 4 °C and immersed in 30% sucrose solution for over 48 h. Cornal sections for striatum and SNpc were cut at 25 µm in a cryostat and kept in tissue stock solution Glycerol, ethelene glycol, DW and 0.2 M PB (3:3:3:1, v/v/v/v). After further washing with PBS, the sections were immunostained with DAB staining kit (TL-060-QHD, Thermo). The sections were incubated for 10 min in a 4% hydrogen peroxide block, washed three times with PBS, incubated for 10 min in an Protein Block Serum-Free (X0909, DAKO), then washed again. The sample was then immunostained with a mixture of primary antibodies against TH (1:500, p40101-0, Pel-freez) in a blocking solution (0.3% Triton-X, 2% Protein Block Serum-Free in 0.1 M PBS) overnight at 4 °C on a shaker. After washing three times with PBS, the section were incubated in HRP Polymer Quanto for 1 h and washed 4 times with PBS. And the sections were immersed in the mixture of DAB+ substrate buffer (K3468, DAKO) at 1:10 ratio. Finally, the sections were mounted. A series of bright field images were obtained by Olympus microscope with 4X magnification. The optical density of the TH stained striatum was determined in the equivalent frame range, with optical density in the corpus callosum used as a reference. The results were expressed as the average of all sections. Cell counting with the 20 X magnification images were performed using the cell counting tool in Image J.

Data analysis

Data is expressed as mean ± S.E.M. Comparisons of three or more groups were analyzed through

one-way ANOVA and *Turkey's* multiple comparison tests. Statistical analysis was carried out using PRISM (GraphPad Software). p < 0.05 was considered statistically significant.

Abbreviations

AD, Alzheimer's disease; DA, dopamine; DAergic: dopaminergic; hMAO-A, human monoamine oxidase A; hMAO-B, human monoamine oxidase B; HRP, horseradish peroxidase; IC_{50} : the half maximal inhibitory concentration; i.p., intraperitoneal; K_i , inhibition constant; K_m , Michaelis constant; MPP+, 1-methyl-4-phenylpyridinium; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; PBS, phosphate buffer saline; PD, Parkinson's disease; PFA, paraformaldehyde; *p.o.*, peroral; ROS, reactive oxygen species; S.E.M, standard error of mean; SNpc, substantia nigra pars compacta; TH, tyrosine hydroxylase; V_{max} , maximal velocity.

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Supplementary data

Supplementary data (experimental procedures for intermediates and ¹H and ¹³C NMR spectra for all final compounds (7, 8, 10, 12, 14) associated with this article can be found, in the

online version, at http://dx.doi.org

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A

Supplementary Data

Synthesis and Evaluation of Biaryl Derivatives for Structural Characterization of Selective Monoamine Oxidase B Inhibitors toward Parkinson's Disease Therapy

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Supplementary Experiments

General procedure for the preparation of 4-(benzyloxy)benzonitrile derivatives (Method A). A solution mixture of benzylbromide derivative, 4-cyanophenol (1.0–1.2 equiv) and K₂CO₃ (4.0 equiv) was heated in acetone (200–400 mL) at 50-60 °C of reflux (4 h). The reaction mixture was evaporated in vacuo, and the resulted residue was diluted in CH₂Cl₂ (300 mL), then extracted with distilled H₂O (300 mL \times 2). The combined organic layer was dried (Na₂SO₄) and evaporated in vacuo.

General procedure for the preparation of 1-(benzyloxy)-4-nitrobenzene derivatives (Method B).

To a 4-nitrophenol dissolved in DMF was added potassium carbonate and benzyl bromide derivative. The reaction mixture was stirred for overnight at room temperature. After the reaction, the product mixture was extracted by EtOAc and distilled water. Organic layer was dried over sodium sulfate and recrystallized.

General procedure for the preparation of *tert*-butyl (4-(benzyloxy)phenethyl)carbamate derivatives (Method C). To a *N*-Boc-tyramine dissolved in acetone was added benzyl bromide derivative and potassium carbonate. The reaction mixture was refluxed at 50-60 °C for 4 h. After the reaction, the reaction mixture was filtered, and filtered solution was evaporated in vacuo. The product residue was purified by column chromatography.

Preparation of 4-(benzyloxy)benzonitrile (6a). Using Method A, 4-cyanophenol (2.00 g, 16.8 mmol), benzyl bromide (2.89 mL, 25.7 mmol), and K₂CO₃ (9.29 g, 67.2 mmol) gave **6a** as a white solid (3.27 g, 93%): $R_f = 0.31$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.21 (s, OCH₂), 7.19 (d, J = 8.8 Hz, 2 ArH), 7.32-7.38 (m, 1 ArH), 7.38-7.43 (m, 2 ArH), 7.44-7.48 (m, 2 ArH), 7.78 (d, J = 8.8 Hz, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 69.7 (OCH₂), 103.0 (ArC-CN), 115.9 (2 ArC), 119.1 (CN), 127.9, 128.1, 128.5, 134.5, 136.1 (8 ArC), 161.8 (OArC).

Preparation of 4-((2-(trifluoromethyl)benzyl)oxy)benzonitrile (6b). Using Method A, 4cyanophenol (1.00 g, 8.4 mmol), 2-(trifluoromethyl)benzyl bromide (1.28 mL, 8.4 mmol), and K₂CO₃ (4.64 g, 33.6 mmol) gave **6b** as a white solid (1.63 g, 70%): $R_f = 0.39$ (EtOAc 1 : *n*-hexane 5): mp 67-68 °C; ¹H NMR (CDCl₃, 400 MHz) δ 5.33 (s, OCH₂), 7.20 (d, J = 8.8 Hz, 2 ArH), 7.59-7.64 (m, 1 ArH), 7.71-7.77 (m, 2 ArH), 7.78-7.84 (m, 3 ArH); ¹³C NMR (CDCl₃, 100 MHz) δ 66.5

(OCH₂), 104.8 (ArC-CN), 115.6, 119.1 (CN), 124.4 (q, *J*_{C-F} = 272.0 Hz), 126.3 (CF₃), 127.8, 127.6 (q, *J*_{C-F} = 30.9 Hz), 128.4, 128.8, 132.4, 134.2, 134.3 (8 ArC), 161.6 (OArC).

Preparation of 4-((**3-**(**trifluoromethyl**)**benzyl**)**oxy**)**benzonitrile** (**6c**). Using Method A, 4-cyanophenol (0.30 g, 2.5 mmol), 3-(trifluoromethyl)benzyl bromide (0.40 mL, 2.5 mmol), and K₂CO₃ (1.39 g, 10.1 mmol) gave **6c** as a white solid (0.64 g, 91%): $\mathbf{R}_f = 0.31$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (CDCl₃, 400 MHz) δ 5.32 (s, OCH₂), 7.21 (d, J = 8.8 Hz, 2 Ar**H**), 7.62-7.68 (m, 1 Ar**H**), 7.70-7.75 (m, 1 Ar**H**), 7.76-7.82 (m, 3 Ar**H**), 7.82-7.86 (m, 1 Ar**H**); ¹³C NMR (CDCl₃, 100 MHz) δ 69.5 (OCH₂), 104.8 (Ar**C**-CN), 115.7, 119.1 (CN), 124.0 (q, $J_{C-F} = 265.3$ Hz), 124.2 (CF₃), 124.3, 130.7, 131.3 (q, $J_{C-F} = 32.3$ Hz), 134.2 (10 Ar**C**), 161.7 (OAr**C**).

Preparation of 4-((**4-**(**trifluoromethyl**)**benzyl**)**oxy**)**benzonitrile** (**6d**). Using Method A, 4cyanophenol (0.30 g, 2.5 mmol), 4-(trifluoromethyl)benzyl bromide (0.40 mL, 2.5 mmol), and K₂CO₃ (1.39 g, 10.1 mmol) gave **6d** as a white solid (0.40, 57%): $R_f = 0.27$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (CDCl₃, 400 MHz) δ 5.33 (s, OCH₂), 7.20 (d, J = 8.8 Hz, 2 Ar**H**), 7.65-7.70 (m, 2 Ar**H**), 7.75-7.82 (m, 4 Ar**H**); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 69.2 (OCH₂), 103.8 (Ar**C**-CN), 116.4 (1 Ar**C**), 119.5 (**C**N), 124.8 (q, $J_{C-F} = 270.5$ Hz), 125.8 (q, $J_{C-F} = 3.7$ Hz), 125.9, 128.6, 129.0 (q, $J_{C-F} =$ 31.5 Hz), 134.7, 141.5 (10 Ar**C**), 162.0 (OAr**C**).

Preparation of 4-((**2-fluorobenzyl)oxy)benzonitrile** (**6e**). Using Method A, 4-cyanophenol (1.00 g, 8.4 mmol), 2-fluorobenzyl bromide (1.90 g, 10.1 mmol), and K₂CO₃ (4.64 g, 33.6 mmol) gave **6e** as a white powder (1.61 g, 84%): $R_f = 0.50$ (EtOAc 1 : *n*-hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.24 (s, OCH₂), 7.19-7.31 (m, 4 ArH), 7.41-7.49 (m, 1 ArH), 7.54-7.60 (m, 1 ArH), 7.77-7.82 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 64.1 (OCH₂), 103.3 (ArC-CN), 115.5 (d, *J*_{C-F} = 20.8 Hz), 115.8, 119.1 (CN), 122.9, 123.1, 124.6, 130.8 (d, *J*_{C-F} = 8.0 Hz), 130.9, 134.3 (9 ArC), 160.5 (d, *J*_{C-F} = 244.2 Hz), 161.6 (OArC).

Preparation of 4-((**3-fluorobenzyl)oxy**)**benzonitrile (6f).** Using Method A, 4-cyanophenol (4.00 g, 33.6 mmol), 3-fluorobenzyl bromide (4.18 mL, 33.6 mmol), and K₂CO₃ (18.60 g, 134.3 mmol) gave **6f** as a white solid (7.25 g, 95%): $\mathbf{R}_f = 0.47$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.23 (s, OCH₂),7.14-7.22 (m, 3 ArH), 7.28-7.32 (m, 2 ArH), 7.45 (q, *J* = 7.6 Hz, 14.1 Hz, 1 ArH), 7.79 (d, *J* = 8.8 Hz, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.8 (OCH₂), 103.2 (ArC-CN), 114.4,

114.6, 114.9 (d, $J_{C-F} = 20.9$ Hz), 115.9, 119.1 (CN), 123.8, 130.6 (d, $J_{C-F} = 8.3$ Hz), 134.3, 139.1 (10 ArC), 161.6 (OArC), 162.2 (d, $J_{C-F} = 242.4$ Hz).

Preparation of 4-((**4-fluorobenzyl**)**oxy**)**benzonitrile (6g).** Using Method A, 4-cyanophenol (1.00 g, 8.39 mmol), 4-fluorobenzyl bromide (1.05 mL, 8.39 mmol), and K₂CO₃ (4.64 g, 33.56 mmol) gave **6g** as a white solid (1.86 g, 97%): $R_f = 0.26$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.19 (s, OCH₂), 7.16-7.27 (m, 4 ArH), 7.49-7.55 (m, 2 ArH), 7.78 (d, *J* = 8.8 Hz, 2 ArH); ¹³C NMR (CDCl₃, 100 MHz) δ 68.9 (OCH₂), 103.1 (1 ArC), 115.4 (d, *J*_{C-F} = 21.4 Hz), 119.1 (CN), 130.3 (d, *J*_{C-F} = 8.3 Hz), 132.4, 134.2 (4 ArC), 161.7 (1 ArC), 161.9 (d, *J*_{C-F} = 242.7 Hz).

Preparation of 4-((3-chlorobenzyl)oxy)benzonitrile (6h). Using Method A, 4-cyanophenol (1.00 g, 8.4 mmol), 3-chlorobenzyl bromide (2.07 g, 10.1 mmol), and K₂CO₃ (4.64 g, 33.6 mmol) gave **6h** as a white crystalline (1.63 g, 80%): R_f = 0.33 (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 5.22 (s, OCH₂), 7.16-7.22 (m, 2 ArH), 7.40-7.45 (m, 3 ArH), 7.52-7.55 (m, 1 ArH), 7.75-7.81 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 68.7 (OCH₂), 103.2 (ArC-CN), 115.9, 119.1 (CN), 126.3, 127.5, 128.0, 130.4, 133.2, 134.2, 138.7 (10 ArC), 161.5 (OArC).

Preparation of Preparation of 4-((4-chlorobenzyl)oxy)benzonitrile (6i). Using Method A, 4cyanophenol (1.00 g, 8.4 mmol), 4-chlorobenzyl bromide (1.72 g, 8.4 mmol), and K₂CO₃ (4.64 g, 33.6 mmol) gave 6i as a white solid (1.96 g, 96%): R_f = 0.20 (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ5.21 (s, OCH₂), 7.18 (d, *J* = 8.9 Hz, 2 ArH), 7.45-7.52 (m, 4 ArH), 7.78 (d, *J* = 8.8 Hz, 2 ArH); ¹³C NMR (CDCl₃, 100 MHz) δ68.8 (OCH₂), 103.2 (ArC-CN), 115.9, 119.1 (CN), 128.5, 129.7, 132.7, 134.2 (ArC-Cl), 135.2 (10 ArC), 161.6 (OArC).

Preparation of 4-((3-methoxybenzyl)oxy)benzonitrile (6j). Using Method A, 4-cyanophenol (1.00 g, 8.4 mmol), 3-methoxybenzyl bromide (1.18 mL, 8.4 mmol), and K₂CO₃ (4.64 g, 36.6 mmol) gave **6j** as a white solid (2.00 g, 100%): $R_f = 0.32$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 3.76 (s, OCH₃), 5.18 (s, OCH₂), 6.89-6.94 (m, 1 ArH), 7.00-7.04 (m, 2 ArH), 7.18 (d, *J* = 8.8 Hz, 2 ArH), 7.31 (t, *J* = 8.1 Hz, 1 ArH), 7.77 (d, *J* = 8.8 Hz, 2 ArH); ¹³C NMR (CDCl₃, 100 MHz) δ 55.1 (OCH₃), 69.5 (OCH₂), 103.0 (ArC-CN), 113.3, 113.5, 115.9, 119.1 (CN), 119.9, 129.7, 134.2, 137.7 (9 ArC), 159.4 (ArC-OCH₂), 161.7 (ArC-OCH₃).

Preparation of 4-((**4-methoxybenzyl)oxy)benzonitrile** (**6k**). Using Method A, 4-cyanophenol (0.50 g, 4.2 mmol), 4-methoxybenzyl bromide (0.43 mL, 4.2 mmol), and K₂CO₃ (2.32 g, 16.8 mmol) gave **6k** as a white solid (0.65 g, 70%): $R_f = 0.27$ (EtOAc 1 : *n*-hexane 3): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 3.76 (s, OCH₃), 5.11 (s, OCH₂), 6.91-6.99 (m, 2 ArH), 7.13-7.20 (m, 2 ArH), 7.36-7.43 (m, 2 ArH), 7.73-7.80 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 55.6 (OCH₃), 70.0 (OCH₂), 103.3 (ArC-CN), 114.4, 116.4, 119.6 (CN), 128.5, 130.2, 134.6, 159.7 (ArC-OCH₂), 162.3 (ArC-OCH₃).

Preparation of 1-((4-nitrophenoxy)methyl)-2-(trifluoromethyl)benzene (9a). Using Method B, 4nitrophenol (0.50 g, 3.6 mmol), 2-(trifluoromethyl)benzyl bromide (1.03 g, 4.3 mmol), and potassium carbonate (1.98 g, 14.4 mmol) gave **9a** as a white powder (0.58 g, 54%): $R_f = 0.26$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.38 (s, OCH₂), 7.22-7.28 (m, 2 ArH), 7.60-7.66 (m, 1 ArH), 7.72-7.85 (m, 3 ArH), 8.21-8.27 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 67.2 (OCH₂), 115.3, 124.3 (q, *J*_{C-F} = 275.1 Hz), 126.0, 126.3, 126.4, 127.2 ((q, *J*_{C-F} = 30.3 Hz), 129.2, 131.0, 132.9, 133.7, 141.3 (10 ArC), 163.2 (OArC).

Preparation of 1-((**4-nitrophenoxy**)**methyl**)-**3-**(**trifluoromethyl**)**benzene** (**9b**). Using Method B, 4nitrophenol (0.50 g, 3.6 mmol), 3-(trifluoromethyl)benzyl bromide (1.03 g, 4.3 mmol), and potassium carbonate (1.98 g, 14.4 mmol) gave **9b** as a white powder (0.98 g, 91%): $R_f = 0.27$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.38 (s, OCH₂), 7.24-7.30 (m, 2 ArH), 7.65-7.71 (m, 1 ArH), 7.73-7.77 (m, 1 ArH), 7.79-7.83 (m, 1 ArH), 7.86-7.89 (m, 1 ArH), 8.22-8.28 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 69.2 (OCH₂), 115.4, 124.1 (q, *J*_{C-F} = 268.2 Hz), 124.3, 124.3, 124.9, 124.9, 129.3 (q, *J*_{C-F} = 31.5 Hz), 129.7, 131.9, 137.5, 141.1 (10 ArC), 163.3 (OArC).

Preparation of 1-((4-nitrophenoxy)methyl)-4-(trifluoromethyl)benzene (9c). Using Method B, 4nitrophenol (0.50 g, 3.6 mmol), 4-(trifluoromethyl)benzyl bromide (1.03 g, 4.3 mmol), and potassium carbonate (1.98 g, 14.4 mmol) gave **9c** as a white powder (0.61 g, 57%): $R_f = 0.27$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.39 (s, OCH₂), 7.22-7.28 (m, 2 ArH), 7.68-7.72 (m, 2 ArH), 7.76-7.81 (m, 2 ArH), 8.21-8.26 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 69.2 (OCH₂), 115.4, 124.1 (q, *J*_{C-F} = 265.8 Hz), 125.4, 125.5, 128.2, 128.7 (q, *J*_{C-F} = 31.7 Hz), 140.9 (10 ArC), 141.2 (ArC-NO₂), 163.3 (OArC).

Preparation of 1-fluoro-2-((4-nitrophenoxy)methyl)benzene (9d). Using Method B, 4-nitrophenol (0.30 g, 2.2 mmol), 2-fluorobenzyl bromide (0.32 mL, 2.6 mmol), and potassium carbonate (1.19 g,

8.6 mmol) gave **9d** as a white solid (0.38 g, 71%): $R_f = 0.40$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (DMSO- d_6 , 400 MHz) δ 5.31 (s, OCH₂), 7.23-7.32 (m, 4 ArH), 7.46 (q, J = 13.8 Hz, 1 ArH), 7.59 (t, J = 7.5 Hz, 1 ArH), 8.23 (d, J = 9.1 Hz, 2 ArH); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 64.6 (OCH₂), 115.3, 115.5 (d, $J_{C-F} = 20.8$ Hz), 122.7, 122.9, 124.7, 125.9, 130.9 (d, $J_{C-F} = 8.1$ Hz), 141.1 (11 ArC), 160.5 (d, $J_{C-F} = 245.0$ Hz), 163.4 (OArC).

Preparation of 1-fluoro-3-((**4-nitrophenoxy**)**methyl**)**benzene** (**9e**). Using Method B, 4-nitrophenol (0.20 g, 1.4 mmol), 3-fluorobenzyl bromide (0.21 mL, 1.7 mmol), and potassium carbonate (0.80 g, 5.8 mmol) gave **9e** as a white solid (0.31 g, 89%): $R_f = 0.43$ (EtOAc 1 : *n*-hexane 5): ¹H NMR (CDCl₃, 400 MHz) δ 5.39 (s, OCH₂), 7.23-7.34 (m, 3 ArH), 7.35-7.48 (m, 2 ArH), 7.57-7.68 (m, 1 ArH), 8.40-8.50 (m, 2 ArH); ¹³C NMR (CDCl₃, 100 MHz) δ 69.9 (OCH₂), 114.3, 114.5, 115.0, 115.5 (d, $J_{C-F} = 21.0$ Hz), 122.9, 126.1, 130.5 (d, $J_{C-F} = 8.1$ Hz), 138.2, 138.2, 142.0 (11 ArC), 163.1 (d, $J_{C-F} = 245.4$ Hz), 163.5 (OArC).

Preparation of 1-fluoro-4-((**4-nitrophenoxy)methyl)benzene** (**9f**). Using Method B, 4-nitrophenol (0.30 g, 2.2 mmol), 4-fluorobenzyl bromide (0.32 mL, 2.6 mmol), and potassium carbonate (1.19 g, 8.6 mmol) gave **9f** as a pale yellow solid (0.38 g, 71%): $R_f = 0.33$ (EtOAc 1 : *n*-hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 5.29 (s, OCH₂), 7.16-7.26 (m, 3 ArH), 7.30-7.35 (m, 2 ArH), 7.42-7.50 (m, 1 ArH), 8.22 (d, *J* = 9.2 Hz, 2 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 69.4 (OCH₂), 115.3, 115.4 (d, *J*_{C-F} = 21.2 Hz), 125.9, 130.3 (d, *J*_{C-F} = 8.3 Hz), 132.2, 141.0 (11 ArC), 162.0 (d, *J*_{C-F} = 242.7 Hz), 163.5 (OArC).

Preparation of *tert*-butyl (4-((2-(trifluoromethyl)benzyl)oxy)phenethyl)carbamate (11a). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 2-(trifluoromethyl)benzyl bromide (0.60 g, 2.5 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11a** as a white solid (0.79 g, 94%): $R_f = 0.47$ (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 1.36 (s, C(CH₃)₃), 2.56-2.66 (m, NHCH₂CH₂), 3.03-3.13 (m, NHCH₂CH₂), 5.20 (s, OCH₂), 6.82-6.95 (m, 3 ArH), 7.07-7.15 (m, 2 ArH), 7.52-7.61 (m, 1 ArH), 7.67-7.82 (m, 3 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 28.2 (C(CH₃)₃), 34.7 (NHCH₂CH₂), 41.7 (NHCH₂CH₂), 66.1 (OCH₂), 77.5 (C(CH₃)₃), 114.6, 124.3 (q, *J*_{C-F} = 272.2 Hz), 126.1, 126.8 (q, *J*_{C-F} = 30.3 Hz), 128.6, 129.7, 130.3, 12.1, 132.8, 135.2, 155.5 (13 ArC), 156.5 (C(O)C(CH₃)₃).

Preparation of *tert*-butyl (4-((3-(trifluoromethyl)benzyl)oxy)phenethyl)carbamate (11b). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 3-(trifluoromethyl)benzyl bromide (0.60 g, 2.5 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11b** as a white solid (0.84 g, 100%): R_f = 0.47 (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 1.36 (s, C(CH₃)₃), 2.59-2.64 (m, NHCH₂CH₂), 3.05-3.12 (m, NHCH₂CH₂), 5.18 (s, OCH₂), 6.85 (t, *J* = 5.5 Hz, 1 ArH), 6.92-6.97 (m, 2 ArH), 7.09-7.14 (m, 2 ArH), 7.61-7.82 (m, 4 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 28.2 (C(CH₃)₃), 34.6 (NHCH₂CH₂), 41.7 (NHCH₂CH₂), 68.3 (OCH₂), 77.4 (C(CH₃)₃), 114.7, 124.2 (q, *J*_{C-F} = 270.2 Hz), 125.3, 127.9, 128.2 (q, *J*_{C-F} = 31.4 Hz), 129.6, 131.9, 142.2, 155.5 (13 ArC), 156.4 (C(O)C(CH₃)₃).

Preparation of *tert*-butyl (4-((4-(trifluoromethyl)benzyl)oxy)phenethyl)carbamate (11c). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 4-(trifluoromethyl)benzyl bromide (0.60 g, 2.5 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11c** as a white solid (0.85 g, 100%): $\mathbf{R}_f =$ 0.38 (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 1.35 (s, C(CH₃)₃), 2.61 (t, *J* = 7.5 Hz, NHCH₂CH₂), 3.08 (q, *J* = 7.9 Hz, NHCH₂CH₂), 5.19 (s, OCH₂), 6.85 (t, *J* = 5.4 Hz, 1 ArH), 6.91-6.96 (m, 2 ArH), 7.08-7.14 (m, 2 ArH), 7.63-7.78 (m, 4 ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 28.2 (C(CH₃)₃), 34.2 (NHCH₂CH₂), 41.7 (NHCH₂CH₂), 68.2 (OCH₂), 77.4 (C(CH₃)₃), 114.7, 124.2 (q, *J*_{C-F} = 270.2 Hz), 125.3, 127.9, 128.2 (q, *J*_{C-F} = 31.4 Hz), 129.6, 131.9, 142.2, 155.5 (13 ArC), 156.4 (C(O)C(CH₃)₃).

Preparation of tert-butyl (4-((2-fluorobenzyl)oxy)phenethyl)carbamate (11d). Using Method C, *N*-Boc-tyramine (1.00 g, 4.2 mmol), 2-fluorobenzyl bromide (0.53 ml, 4.4 mmol) and potassium carbonate (2.33 g, 16.8 mmol) gave **11d** as a white solid (0.97 g, 66%); $R_f = 0.48$ (EtOAc 1: *n*-hexane 3); ¹H NMR (400 MHz, CDCl₃) δ 1.43 (s, C(CH₃)₃), 2.73 (t, J = 6.8 Hz, NHCH₂CH₂), 7.06-7.17 (m, 4 ArH), 7.28-7.33 (m, 1 ArH), 7.50 (t, J = 7.2 Hz, 1 ArH); ¹³C NMR (75 MHz, CDCl₃) δ 28.4 (C(CH₃)₃), 35.4 (NHCH₂CH₂), 42.0 (NHCH₂CH₂), 63.8 (OCH₂), 79.2 (C(CH₃)₃), 115.0, 115.2, 115.5, 124.2, 115.0, 115.3 (d, $J_{C-F} = 21.1$ Hz), 124.2 (d, $J_{C-F} = 3.6$ Hz), 124.3 (d, $J_{C-F} = 14.1$ Hz), 129.6 (d, $J_{C-F} = 5.8$ Hz), 129.7 (d, $J_{C-F} = 1.4$ Hz), 129.8, 131.6, 155.9 (C(O)), 158.8 (OArC) (12 ArC).

Preparation of *tert*-butyl (4-((3-fluorobenzyl)oxy)phenethyl)carbamate (11e). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 3-fluorobenzyl bromide (0.39 mL, 3.2 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11e** as a white solid (0.29 g, 40%): $R_f = 0.34$ (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 1.36 (s, C(CH₃)₃), 2.57-2.65 (m, NHCH₂CH₂), 3.03-3.13 (m,

NHCH₂CH₂), 5.09 (s, OCH₂), 6.84 (t, J = 5.3 Hz, NH), 6.89-6.96 (m, 2 ArH), 7.07-7.19 (m, 3 ArH), 7.23-7.30 (m, 2 ArH), 7.39-7.48 (m, 1 ArH); ¹³C NMR (DMSO- d_6 , 75 MHz) δ 28.7 (C(CH₃)₃), 35.1 (NHCH₂CH₂), 42.2 (NHCH₂CH₂), 68.7 (OCH₂), 77.9 (C(CH₃)₃), 114.5 (d, $J_{C-F} = 21.6$ Hz), 114.8, 115.1, 123.8, 123.8, 130.1, 130.9 (d, $J_{C-F} = 8.3$ Hz), 132.3, 140.7, 140.8 (10 ArC), 156.0 (C(O)), 157.0 (OArC), 162.7 (d, $J_{C-F} = 242.1$ Hz).

Preparation of *tert*-butyl (4-((4-fluorobenzyl)oxy)phenethyl)carbamate (11f). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 4-fluorobenzyl bromide (0.31 mL, 2.5 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11f** as a white solid (0.71 g, 97%): $R_f = 0.23$ (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 1.36 (s, C(CH₃)₃), 2.58-2.66 (m, NHCH₂CH₂), 3.04-3.14 (m, NHCH₂CH₂), 5.04 (s, OCH₂), 6.84 (t, *J* = 5.4 Hz, NH), 6.89-6.96 (m, 2 ArH), 7.07-7.13 (m, 2 ArH), 7.16-7.26 (m, 2 ArH), 7.45-7.52 (m, 2 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 28.2 (C(CH₃)₃), 34.6 (NHCH₂CH₂), 41.7 (NHCH₂CH₂), 68.4 (OCH₂), 77.4 (C(CH₃)₃), 114.6, 115.2 (d, *J*_{C-F} = 21.2 Hz), 129.5, 129.8 (d, *J*_{C-F} = 8.2 Hz), 131.7, 133.4, 133.5 (11 ArC), 155.5 (C(O)), 156.6 (OArC), 161.7 (d, *J*_{C-F} = 242.1 Hz).

Preparation of tert-butyl (4-((2-chlorobenzyl)oxy)phenethyl)carbamate (11g). Using Method C, *N*-Boc-tyramine (1.00 g, 4.2 mmol), 2-chlorobenzyl bromide (0.57 ml, 4.4 mmol) and potassium carbonate (2.33 g, 16.9 mmol) gave **11g** as a white solid (1.19 g, 78%); $R_f = 0.47$ (EtOAc 1: *n*-Hexane 3); ¹H NMR (400 MHz, CDCl₃) δ 1.43 (s, C(CH₃)₃), 2.73 (t, J = 6.9 Hz, NHCH₂CH₂), 3.33-3.35 (m, NHCH₂CH₂), 4.55 (br s, BocNH), 5.14 (s, OCH₂), 6.93 (d, J = 8.3 Hz, 2 ArH), 7.11 (d, J = 8.3 Hz, 2 ArH), 7.23-7.30 (m, 2 ArH), 7.38-7.40 (m, 1 ArH), 7.54-7.56 (m, 1 ArH); ¹³C NMR (75 MHz, CDCl₃) δ 28.4 (C(CH₃)₃), 35.4 (NHCH₂CH₂), 42.0 (NHCH₂CH₂), 67.2 (OCH₂), 79.2 (C(CH₃)₃), 127.0, 128.8, 129.0, 129.4, 129.8, 131.6, 132.2, 132.6, 134.9, 155.9 (C(O)), 157.2 (OArC) (12 ArC).

Preparation of *tert*-butyl (4-((3-chlorobenzyl)oxy)phenethyl)carbamate (11h). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 3-chlorobenzyl bromide (0.41 mL, 3.2 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11h** as a white solid (0.68 g, 93%): $\mathbf{R}_f = 0.40$ (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 300 MHz) δ 1.36 (s, C(CH₃)₃), 2.58-2.66 (m, NHCH₂CH₂), 3.03-3.13 (m, NHCH₂CH₂), 5.09 (s, OCH₂), 6.84 (t, *J* = 5.4 Hz, NH), 6.89-6.96 (m, 2 ArH), 7.07-7.14 (m, 2 ArH), 7.35-7.44 (m, 3 ArH), 7.48-7.52 (m, 1 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 28.7 (C(CH₃)₃), 35.1

(NHCH₂CH₂), 42.2 (NHCH₂CH₂), 68.7 (OCH₂), 77.9 (C(CH₃)₃), 115.1, 126.5, 127.6, 128.1, 130.1, 130.8, 132.3, 133.6 (ArC-Cl), 140.4 (10 ArC), 156.0 (C(O)), 156.9 (OArC).

Preparation of *tert*-butyl (4-((4-chlorobenzyl)oxy)phenethyl)carbamate (11i). Using Method C, *N*-Boc-tyramine (0.50 g, 2.1 mmol), 4-chlorobenzyl bromide (0.41 mL, 3.2 mmol), and potassium carbonate (1.17 g, 8.4 mmol) gave **11i** as a white solid (0.70 g, 96%): $R_f = 0.34$ (EtOAc 1: *n*-Hexane 3): ¹H NMR (DMSO-*d*₆, 400 MHz) δ 1.36 (s, C(CH₃)₃), 2.58-2.66 (m, NHCH₂CH₂), 3.04-3.14 (m, NHCH₂CH₂), 5.06 (s, OCH₂), 6.84 (t, *J* = 5.4 Hz, NH), 6.88-6.95 (m, 2 ArH), 7.07-7.13 (m, 2 ArH), 7.41-7.50 (m, 4 ArH); ¹³C NMR (DMSO-*d*₆, 75 MHz) δ 28.2 (C(CH₃)₃), 34.6 (NHCH₂CH₂), 41.7 (NHCH₂CH₂), 68.3 (OCH₂), 77.4 (C(CH₃)₃), 114.6, 128.4, 129.3, 129.56, 131.7, 132.3 (ArC-Cl), 136.3 (10 ArC), 155.5 (C(O)), 156.5 (OArC).

Preparation of tert-butyl (4-((3-methoxybenzyl)oxy)phenethyl)carbamate (11j). Using Method C, *N*-Boc-tyramine (1.00 g, 4.2 mmol), 3-methoxybenzyl bromide (0.71 ml, 5.1 mmol) and potassium carbonate (2.33 g, 16.9 mmol) gave **11j** as a white solid (1.31 g, 87%); $R_f = 0.40$ (EtOAc 1: *n*-Hexane 3); ¹H NMR (400 MHz, CDCl₃) δ 1.43 (s, C(CH₃)₃), 2.72 (t, *J* = 6.9 Hz, NHCH₂CH₂), 3.32-3.34 (m, NHCH₂CH₂), 3.81 (s, OCH₃), 4.56 (br s, BocNH), 5.01 (s, OCH₂), 6.84-6.93 (m, 3ArH), 7.00 (d, *J* = 7.3 Hz, 2ArH), 7.10 (d, *J* = 8.4 Hz, 2ArH), 7.25-7.32 (m, 1ArH); ¹³C NMR (75 MHz, CDCl₃) δ 28.4 (C(CH₃)₃), 35.4 (NHCH₂CH₂), 42.0 (NHCH₂CH₂), 55.3 (OCH₃), 70.0 (OCH₂), 79.2 (C(CH₃)₃), 112.9 (12ArC), 113.5, 115.0, 119.6, 129.7, 129.8, 131.4, 138.8, 155.9 (C(O)), 157.5(OArC), 159.9 (CH₃OArC) (12 ArC).

of *N*-boc-2-(4-bromophenyl)ethylamine **Preparation** (13). solution То a of 4bromophenethylamine (5.00 g, 25.0 mmol), potassium carbonate (5.18 g, 37.5 mmol) in CH₂Cl₂ (125 ml) was stirred for 15 min at room temperature, and boc anhydride was added (6.04 mL, 26.3 mmol). After 16 h, the reaction mixture was washed with H_2O (125 mL \times 3) and brine (75 mL). The organic layer was dried with anhydrous Na₂SO₄ and concentrated in vacuo. The residue was washed with *n*hexane and gave 13 as a white solid (6.63 g, 88%); $R_f = 0.50$ (EtOAc 1: *n*-Hexane 3); ¹H NMR (300) MHz, DMSO- d_6) δ 1.43 (s, C(CH₃)₃), 2.75 (t, J = 6.6 Hz, NHCH₂CH₂), 3.34-3.35(m, NHCH₂CH₂), 7.06 (d, J = 8.0 Hz, 2 Ar**H**), 7.42 (d, J = 8.2 Hz, 2 Ar**H**); ¹³C NMR (75 MHz, DMSO- d_6) δ 28.4 (C(CH₃)₃), 35.7 (NHCH₂CH₂), 41.6 (NHCH₂CH₂), 79.3 (C(CH₃)₃), 120.2, 130.6, 131.6, 138.0 (6 ArC), 155.8 (C(O)).

$$H \xrightarrow{O} OMe \xrightarrow{NH_2} HCI \xrightarrow{I. MeOH, TEA, 2.5 h, r.t.} H_2N \xrightarrow{O} H \xrightarrow{OMe} \underbrace{MeSO_3H}_{EA, r.t., 1h} \xrightarrow{O} H \xrightarrow{N} OMe \xrightarrow{MeSO_3H} H_2N \xrightarrow{O} H \xrightarrow{N} OMe \xrightarrow{MeSO_3H} H_2N \xrightarrow{O} H \xrightarrow{N} OMe \xrightarrow{N}$$

Scheme S1. Synthetic Procedure for Compound 15.

Preparation of (S)-2-((4-methoxybenzyl)amino)propanamide methanesulfonate (15).

To MeOH solution (4 mL) of L-alanine amide hydrochloride (0.69 g, 5.5 mmol), trimethylamine (0.90 mL, 6.4 mmol) was added. The reaction mixture was stirred for 15 min at room temperature and then treated with p-anisaldehyde (0.45 mL, 3.7 mmol) at room temperature. After 2.5 h, the reaction mixture was concentrated in vacuo. The resulting residue was dissolved in EtOAc (20 mL) and washed (10 mL \times 2) with brine (10 mL \times 2). The organic layer was dried with anhydrous Na₂SO₄ and concentrated in vacuo. To crude mixture, MeOH (4 mL) solution and sodium cyanoborohydride (0.92 g, 14.7 mmol) were added. The reaction mixture was stirred at room temperature (2 h). The reaction mixture was concentrated in vacuo. The resulting residue was dissolved in EtOAc (20 mL) and washed (10 mL \times 2) with brine (10 mL \times 2). The organic layer was dried with anhydrous Na₂SO₄ and then concentrated in vacuo. The residue was purified by column chromatography with SiO₂ and gave (S)-2-((4-methoxybenzyl)amino)propanamide as a white solid (0.31 g, 40%); $R_f = 0.12$ (only EtOAc); mp: 111-112 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 1.11 (d, J = 6.88 Hz, CH₃), 2.31 (br s, NH), 2.98 (q, J = 6.88 Hz, CH₃CH), 3.47 (d, J = 13.1 Hz, CH₂), 3.60 (d, J = 13.1 Hz, CH₂), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH₂), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH₂), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH₂), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH_2), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH_2), 3.72 (s, OCH₃), 6.86 (d, J = 8.56 Hz, 2 ArH), 6.97 (br s, C(O)NHH), 7.23 (d, J = 13.1 Hz, CH_2), 3.72 (d, J = 13.1 Hz, 3.1 Hz, 3.18.52 Hz, 2 ArH), 7.30 (br s, C(O)NHH); ¹³C NMR (400 MHz, CDCl₃) δ 19.7 (CH₂), 52.0, 55.3, 57.6, 114.0, 129.2, 131.7, 158.8 (6 ArC), 178.4 (CO).

Methanesulfonic acid (0.08 mL, 1.2 mmol) was added to a solution of (*S*)-2-((4-methoxybenzyl)amino)propanamide (0.20 g, 1.0 mmol) in EtOAc (2 mL) at 60 °C. After 1 h, the reaction mixture was cooled until room temperature and then filtered in vacuo and washed with EtOAc. The filtercake was dried without further purification and gave **15** as a white solid (0.28 g, 95%); mp: 202–203 °C ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.42 (d, *J* = 6.92 Hz, CH₃SO₃⁻), 2.39 (s, CH₃), 3.72–3.82 (m, CH₃CH, OCH₃), 4.03 (s, CH₂), 6.99 (*J* = 8.44 Hz, 2ArH), 7.41 (*J* = 8.48 Hz, 2ArH), 7.62 (s, C(O)NHH), 7.93 (s, C(O)NHH), 9.03 (br s, NH₂⁺); ¹³C NMR (75 MHz, DMSO-*d*₆) 16.4, 48.5, 54.7, 55.7, 114.5, 123.9, 132.1, 160.2, 171.0 (CO, 6 ArC, CH₂, CH₃SO₃H).






























































































Purity of Compound 12c

Analytical HPLC was performed using a Waters e2695 system equipped with the following columns: Capcell Pak[®] colulmn (4.6 × 75 mm; 3μ m). HPLC data were recorded using following methods: H₂O/MeCN, 90/10 \rightarrow 50/50 in 15 min, + 50/50 \rightarrow 0/100 in 5 min + 10 min isocratic, flow rate of 0.75 mL/min, $\lambda = 254$, 280 nm.

HPLC purity: 10.6 min, > 98% at 254 & 280 nm



The low-resolution mass spectrum was performed on an API 3200 LC/MS/MS system (SCIEX).

LRMS (M + H)+ (ESI+) 296.1 [M + H]+ (calcd for C16H16F3NOH+ 296.1).



