Synthesis of α -trinositol related analogues. Structure–activity (analgesic and anti-inflammatory) relationships

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(Received 19 September 1996; accepted 2 December 1996)

Summary — α -Trinositol analogues, including methyl ethers, deoxy, oxa and aza derivatives were prepared. The parent compound possesses weak analgesic and anti-inflammatory properties. Removal of the non-phosphorylated hydroxyls generates a compound devoid of analgesic activity but which retains the anti-inflammatory property of the parent compound. The protection of these hydroxyls as methyl ethers leads to compounds which keep their anti-inflammatory activity, whereas the replacement of the cyclohexane carbone backbone by a tetrahydropyrane or a piperidine ring leads to compounds which increase the pain.

α-trinositol / inositol phosphate / analgesic activity / anti-inflammatory activity / oxa-inositol / aza-inositol

Introduction

 α -Trinositol (D-myo-inositol-1,2,6-tris(phosphate) (Ins-(1,2,6)P₃)) [1, 2] **1** is a regioisomer of the well-known D-myo-inositol-1,4,5-tris(phosphate), Ins(1,4,5)P₃ **2**. It has been demonstrated that Ins(1,4,5)P₃ is an intracellular second messenger [3]. α -Trinositol does not bind to Ins(1,4,5)P₃ receptors, but possesses interesting anti-inflammatory and weak analgesic activities [1, 4–6].

Binding studies on rat membranes showed a marked influence of the pH, demonstrating that the ionization state of either the molecule or the receptor could play an important role in the strength of binding [7].

The behaviour of the protons present on the phosphate groups of inositol phosphates is largely influenced by the number and relative positions of the neighbouring functions and the cations present in the medium, particularly the alkali and alkali-earth cations [8–10]. The α -trinositol protons can be stabilized by two main factors: electrostatic interactions and hydrogen bonding.

As the hydroxyls can participate in this stabilization, it seemed of interest to prepare some analogues of α -trinositol in which the non-phosphorylated hydroxyls were modified, in order to compare the activity with that of the parent compound. The modifications included either removal of these hydroxyl groups, or transformation of one or all of them into methyl ethers. Other modifications concerned the replacement of the carbon ring backbone by an analogue containing an atom of oxygen or of nitrogen.

Herein we report the synthesis of (\pm) - $(1\alpha, 2\alpha, 3\beta)$ trihydroxy-cyclohexane-tris(phosphate) **3**, (\pm) -3,4,5tri-O-methyl *myo*-inositol-1,2,6-tris(phosphate) **4**, and (\pm) -5-O-methyl *myo*-inositol-1,2,6-tris(phosphate) **5** (fig 1). These racemic compounds as well as 1,5anhydro-D-arabinitol-2,3,4-tris(phosphate) **6** [11] and 1,5-dideoxy-1,5-imino-D-arabitol-2,3,4-tris(phosphate) **7** [12] were submitted to a preliminary screening for analgesic and anti-inflammatory properties. Their toxicity was also investigated.

Synthesis

The key intermediate for the synthesis of (\pm) - $(1\alpha,2\alpha,3\beta)$ -trihydroxy-cyclohexane-tris(phosphate) **3** is (\pm) - $(1\alpha,2\alpha,3\beta)$ trihydroxy-cyclohexane **8**. Among the reports published on the preparation of this triol [13–15], we selected a slightly modified procedure of

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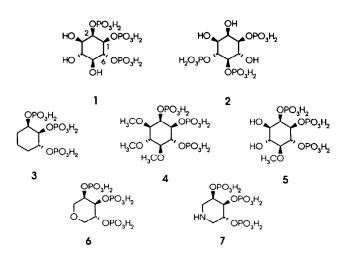
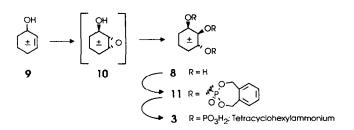


Fig 1. Inositol phosphate and analogues in their acid species.

Fredericks and Guthrie [15]. Thus, treatment of 2-cyclohexenol 9, dissolved in a mixture of water and tetrahydrofuran with m-CPBA furnished the epoxide 10, which was slowly hydrolyzed in situ to give after extraction 45% of crystalline triol 8. NMR studies confirmed the relative orientations of the three hydroxyl functions. The racemic triol was phosphorylated according to the method of Perich and Johns [16]. Thus, the triol was reacted with N,N-diethyl-O-xylylene phosphoramidite in the presence of tetrazole, vielding an intermediate tris(phosphite) which was oxidized to the tris(phosphate) 11 by means of *m*-CPBA. Hydrogenolysis with palladium on charcoal as catalyst removed the phosphate protective groups, giving the expected product 3, which was stabilized as the cyclohexylammonium salt until used for pharmacological studies to avoid elimination and other side reactions (scheme 1).

The synthesis of (\pm) -3,4,5-tri-O-methyl myo-inositol-1,2,6-tris(phosphate) **4** starts with racemic 3,4-di-



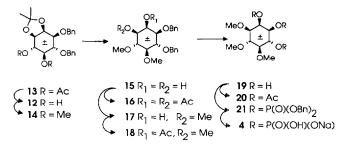
Scheme 1. Synthesis of (\pm) - $(1\alpha, 2\alpha, 3\beta)$ -cyclohexane-triol-tris(phosphate) **3**.

O-benzyl-1,2-O-isopropylidene myo-inositol 12, obtained by saponification of the crystalline diacetate 13 [17]. The diol 12 was converted into the dimethyl ether 14 and the O-isopropylidene group was hydrolyzed to give the diol 15, which yielded a crystalline diacetate 16. Tin-mediated methylation of the equatorial hydroxyl group of 15 gave the trimethyl ether 17 and hydrogenolysis removed the benzyl ether groups, yielding the triol 19 which was characterized as the triacetate **20**. Phosphorylation was as described above; but using N,N-diisopropylamino dibenzyl phosphoramidite instead of N,N-diethyl-O-xylylene phosphoramidite led to the protected final product 21. Hydrogenolysis in glacial acetic acid in the presence of three equivalents of sodium acetate gave the racemic 3,4,5-tri-O-methyl myo-inositol 1,2,6-tris(sodium hydrogen phosphate) 4 as a white powder (scheme 2).

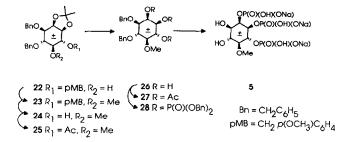
The (\pm) -5-O-methyl myo-inositol 1,2,6-tris(phosphate) **5** was prepared from the *p*-methoxybenzyl ether **22** [17]. The remaining free hydroxyl was transformed into methyl ether **23**. The *p*-methoxybenzyl group was removed from **23** by the action of dichlorodicyanobenzoquinone to give the alcohol **24**, which yielded a crystalline acetate **25**. Hydrolysis of the *O*-isopropylidene group from **24** gave the triol **26**, which was phosphorylated as described above, yielding **28**. Subsequent hydrogenolysis in glacial acetic acid in the presence of three equivalents of sodium acetate gave the expected racemic 5-O-methyl derivative **5** as a trisodium salt (scheme 3).

Pharmacological properties

The compounds 1, 3–7 were tested (3–5 as racemates) for their acute toxicity according to Irwin's test [18]. The anti-inflammatory properties were tested after iv injection (64 mg/kg) in measuring the reduction of carrageenan-induced foot pad oedema [19]. The analgesic effects were tested against the acetic acid-induced writhing using ip injection (64 mg/kg) [20, 21].



Scheme 2. Synthesis of 3,4,5-tri-*O*-methyl *myo*-inositol-1,2,6-tris(phosphate) **4**.



Scheme 3. Synthesis of 5-O-methyl myo-inositol-1,2,6-tris(phosphate) 5.

Discussion

Toxicity appeared only at relatively high doses. All the compounds appeared atoxic at doses up to 256 mg/kg, and toxicity did not seem to be significantly influenced by the substituents (table I).

The anti-inflammatory activity evaluation of the compounds provided various results. The trideoxy derivative **3** significantly reduced the carrageenaninduced oedema. The compound seemed four-fold more potent than aspirin. Masking the non-phosphorylated hydroxyls as methyl ethers led to compounds **4** and **5**, showing relatively similar levels of antiinflammatory activity to **1**. But replacing the cyclo-

Table I. Pharmacological data for the tested compounds.

hexane backbone by a tetrahydropyrane or a piperidine ring seemed to suppress this property (compounds 6 and 7). On the other hand, the latter compounds showed an aggravation in the acetic acid-induced writhing test. The methyl ethers 4 and 5 as well as the deoxy analogue 3 did not possess the analgesic property of the parent compound. However, the analgesic activity of the parent compound remained much lower than that of morphine.

For this preliminary screening we used racemates for compound 3-5. It should be borne in mind that the two enantiomers may behave differently.

The parent compound 1 possesses both anti-inflammatory and weak analgesic properties. The structural variations around the vicinal tris(phosphate) separate the effects, and confer to the molecule only the antiinflammatory effect (compound 3).

Experimental protocols

Melting points were measured on a Mettler PF 62 apparatus and are uncorrected. NMR spectra were recorded either on a Bruker AC 200 spectrometer or on a Jeol FX90Q instrument using the δ scale. Coupling constants are given in Hz. Synthesis of 1,5-anhydro-D-arabinitol-2,3,4-tris(phosphate) **6** and 1,5-dideoxy-1,5-imino-D-arabitol-2,3,4-tris(phosphate) **7** have been described in [11] and [12] respectively. Male rats of NMRI strain, mean weight 22–26 g were used. Aspirin (Synthélabo) and morphine (Coopération Pharmaceutique Française) were dissolved in saline. The tests were carried out at 20–21 °C under artificial lighting between 7.00 and 19.00 h.

| Compound No | Toxicity | | Carrageenan- induced oedema | Writhing |
|----------------|-----------|-------------|--------------------------------|-----------------------------|
| | 256 mg/kg | 512 mg/kg | (% antagonism) ^a | (% protection) ^a |
| 1 | Atox | 3/3 < 1 min | 32 ± 3 | 63 ± 5 |
| 3 | Atox | 3/3 < 1 min | 50 ± 3 | ns |
| 4 | Atox | 3/3 < 1 min | 26 ± 3 | ns |
| 5 | Atox | 2/3 < 1 min | 25 ± 3 | ns |
| 6 | Atox | 2/3 < 1 min | ns | -83 ± 9 |
| 7 | Atox | 3/3 < 1 min | ns | -61 ± 5 |
| Aspirin | | | 54 ± 2 ^b | |
| Morphine | | | | 99 ± 1° |

Atox: atoxic; ns: not significant; atested dose 64 mg/kg iv; b256 mg/kg; c4 mg/kg; a,b, and cgroups of 10 mice; mean value ± standard deviation.

(\pm) -I(R),2,3(R)-Trihydroxy-cyclohexane 8

In a 50 mL flask, *m*-CPBA (0.75 g, 4.35 mmol) was dissolved in 5 mL of a water/THF mixture (5:1). The solution was heated at 40 °C and 2-cyclohexenol **9** (0.4 mL, 0.4 g, 4.07 mmol) was slowly added and heating maintained for 3 h. The flask was then kept overnight at 5 °C. The precipitate of *m*-chlorobenzoic acid was filtered off and the filtrate neutralized by means of a saturated solution of sodium hydrogen carbonate. The solid residue was dissolved in a mixture of ethanol/acetone (1:1). After filtration, the solvents were evaporated giving 0.39 g of a yellow oil, which was crystallized and recrystallized from ethyl acetate. Triol **8** (0.36 g, 37%) was obtained as white crystals, mp = 126.5 °C (lit 124 °C, Beilstein, E III, 6, 6250). ¹H-NMR ((CD₃)₂CO): 3.90 (dt, 1H, *J* = 3.1 Hz, *J* = 2.6 Hz, *H*₂), 3.69 (ddd, 1H, *J* = 9.0 Hz, *J* = 7.7 Hz, *J* = 4.3 Hz, *H*₃), 3.6–3.4 (m, 1H, OH), 3.30 (dd, 1H, *J* = 7.7 Hz, *J* = 3.1 Hz, *H*₂), 3.1–2.7 (m, 2H, (OH)₂), 1.0–1.9 (m, 6H, (CH₂) ring protons).

(±)-Cyclohexane-1(R),2,3(R)-tri-O-(orthoxylylene)phosphate

The triol 8 (84.1 mg, 0.5 mmol) and dried tetrazole (0.28 g, 4.0 mmol) were dissolved in anhydrous THF (10 mL). The flask was placed under an argon atmosphere and diethylamino-1,5-dihydro-2,4,3-benzo-dioxaphosphepine (N,N-diethyl-O-xylylene phosphoramidite) (0.6 g, 2.4 mmol) in 10 mL dried THF was added and the mixture stirred overnight at room tempera-ture. The flask was cooled at 0 $^{\circ}$ C and *m*-CPBA (0.55 g, 3.2 mmol) in 10 mL CH₂Cl₂ was added. After 1 h of stirring at room temperature, the solvents were removed under vacuum and the residue redissolved in a mixture of ether/CH₂Cl₂ (2:1) and washed with a 10% solution of $Na_2S_2O_5$ (2 x 30 mL), washed again with a 5% NaHCO₃ (2 x 30 mL) and finally with a saturated solution of NaCl. After drying over Na₂SO₄, the solvents were removed under reduced pressure to give a brownish paste which was redissolved in a minimum of CH₂Cl₂ and precipitated in ether. The tris(phosphate) **11** was obtained as a white powder (158 mg, 47.3%) mp = 92–93 °C. ¹H-NMR (CDCl₃): 7.4–7.2 (m, 12H, (C₆H₄)₃), 5.5–5.0 (m, 15H, (OCH)₃ + $(OCH_2)_6$, 1.8–1.6 (m, 6H, CH₂ cycle).

(±)-Cyclohexane-1(R),2,3(R)-tris(phosphate) 3 as its tetracyclohexylammonium salt

The protected tris(phosphate) **11** (158 mg, 0.24 mmol) dissolved in 30 mL of a mixture of ethanol/CH₂Cl₂ (2:1) was hydrogenolyzed under atmospheric hydrogen pressure in the presence of 10% Pd/C (20 mg) for 12 h at 20 °C. The Pd/C was filtered off and the filtrate was evaporated to dryness and redissolved in water (1 mL). Cyclohexylamine (0.5 mL) was slowly added at 0 °C. The salt was precipitated by means of acetone. After decantation the supernatant was removed. The precipitate was washed twice with acetone and finally dried under vacuum. The final product was obtained as a white salt (173 mg, 95%). Anal C₃₀H₆₇N₄O₁₂P₃: (C, H, N, P). ¹H-NMR (D₂O): 4.6–4.5 (m, 1H, H_3), 4.4–4.3 (m, 1H, H_1), 4.0–4.1 (m, 1H, H_2), 3.7–3.3 (m, 4H, [H₂-C(H)C₅H₁₀]₄), 2.5–1.3 (m, 46H [H₂N-C(H)C₅H₁₀]₄, (H₄)₂, (H₅)₂, (H₆)₂).

(\pm) -1,6-di-O-Benzyl-4,5-di-O-methyl myo-inositol 15 and the diacetate 16

The diol **12** (3.0 g, 7.5 mmol), prepared by saponification of the crystalline diacetate **13** [17] was treated with an excess of sodium hydride and methyl iodide in DMF and the product isolated in the usual way. The crude product was purified by column chromatography on silica gel eluted with a 1:1 ether/light petroleum mixture, yielding **14** (2.34 g, 73%) as an oil. This was treated under reflux for 30 min with 1 M HCl/

MeOH (1:9) (50 mL). An excess of NaHCO₃ was added to the cooled solution and the solvents were evaporated. The product was extracted from the residue with CH₂Cl₂ and recrystallization from 1:10 ethyl acetate/light petroleum gave the diol **15** (1.5 g, 71%) as crystals, mp = 77–79 °C. Anal $C_{22}H_{28}O_6$ (C, H).

A portion was treated with acetic anhydride-pyridine at 50 °C for 8 h and the product isolated in the usual way to give the diacetate **16**, which was recrystallized from 1:10 ethyl acetate/light petroleum, mp = 98–100 °C. Anal C₂₆H₃₂O₈: (C, H). ¹H-NMR (CDCl₃): 7.3–7.2 (m, 10H (C₆H₅)₂), 5.67 (t, J = 2.7 Hz, 1H, H_2), 4.9–4.3 (m, 5H, H_3 (CH₂C₆H₅)₂), 3.9–3.0 (m, 4H, H_1 , H_4 , H_5 , H_6), 3.66 and 3.56 (2 s, each 3H, (OCH₃)₂), 2.15 and 2.06 (2 s, each 3H, (H₃CCO)₂).

(±)-1,6-di-O-Benzyl-3,4,5-tri-O-methyl myo-inositol 17

A mixture of the diol **15** (1.4 g, 3.6 mmol), dibutyl tin oxide (1.0 g, 4.0 mmol), tetrabutylammonium bromide (1.28 g, 4.0 mmol), acetonitrile (50 mL) and methyl iodide (10 mL, 160 mmol) was heated under reflux with a Soxhlet apparatus containing 3 Å molecular sieves for 24 h when TLC (ether) showed a major product ($R_{\rm f}$ 0.50). The solvents were evaporated from the cooled solution and the residue was stirred with ether (100 mL) and saturated NaHCO₃ (100 mL) for 2 h to precipitate the tin derivative. The mixture was filtered through a Celite pad and the ether layer was separated, dried (K₂CO₃) and concentrated. Column chromatography on silica gel eluted with ether gave the alcohol **17** (1.37 g, 94%), mp = 82–83 °C. Anal C₂₃H₃₀O₆: (C, H). ¹H-NMR (CDCl₃): 7.33 (s, 101, (C₆H₃)₂), 4.84 and 4.72 (2 s, each 2H, (CH₂C₆H₅)₂), 4.24 (t, J = 2.5 Hz, 1H, H_2), 4.0–2.9 (m, 5H, H_1 , H_3 , H_4 , H_5 , H_6), 3.64, 3.62 and, 3.49 (3 s, each 3H, (OCH₃)₃).

A portion was acetylated with acetic anhydride-pyridine at 60 °C for 8 h to give the acetate **18** as a syrup. ¹H-NMR (CDCl₃): 7.4–7.3 (m, 10H (C₆H₅)₂), 5.77 (t, J = 2.5 Hz, <u>1H</u>, H₂), 4.81 (AB, $J_{AB} = 12.0$ Hz, $\Delta\delta$ 0.14, 2H, CH₂C₆H₅), 4.62 (AB, $J_{AB} = 12.0$ Hz, $\Delta\delta$ 0.16 2H, CH₂C₆H₅), 3.8–3.0 (m, 5H, H₁, H₃, H₄, H₅, H₆), 3.66, 3.62 and 3.42 (3 s, each 3H (OCH₃)₃), 2.14 (s, 3H, H₃CCO).

(±)-1,2,6-tri-O-Acetyl-3,4,5-tri-O-methyl myo-inositol 20

A solution of the alcohol **17** (1.2 g, 2.98 mmol) in glacial acetic acid (20 mL) with 10% Pd/C (270 mg) was stirred under hydrogen at atmospheric pressure and room temperature for 36 h. The catalyst was removed by filtration, washed with acetic acid and the eluate was concentrated. Recrystallization of the crystalline product from EtOH (or EtOAc) gave the triol **19** (600 mg, 90%), mp = 172–174 °C. This was acetylated with acetic anhydride-pyridine at 60 °C for 9 h to give the triacetate **20**, mp = 106–108 °C. Anal C₁₅H₂₄O₉: (C, H). ¹H-NMR (CDCl₃): 5.63 (t, *J* = 2.7 Hz, 1H, H₂), 5.34 (t, *J* = 9.2 Hz, 1H, H_6), 4.82 (dd, *J* = 10.4 Hz, *J* = 2.4 Hz, 1H, H_1), 3.6–3.0 (m, 3H, H_3 , H_4 , H_5), 3.61, 3.55, 3.41 (3s, each 3H, (OCH₃)₃), 2.15, 2.08, 1.99 (3s, each 3H (H_3 CCO)₃).

(±)-3,4,5-tri-O-Methyl myo-inositol-1,2,6-tris(dibenzylphosphate) 21

The triol **19** (460 mg, 2.07 mmol) was dissolved in warm CH₃CN (30 mL). Toluene (100 mL) was added and the solvents were evaporated to azeotrope any water from crystallization. The residual product was dissolved in CH₃CN (30 mL) and CH₂Cl₂ (30 mL) and tetrazole (870 mg, 12.4 mmol) and a solution of *N*,*N*-diisopropylamino-dibenzylphosphoramidite [22] (4.3 g, 12.4 mmol) in CH₂Cl₂ (70 mL) was added and the solution stirred at room temperature for 2 h, then cooled to 0 °C and a solution of technical *m*-chloroperbenzoic acid (4.9 g) in CH₂Cl₂ (40 mL) was added and the mixture stirred

for 1 h. The solution was concentrated to remove the CH_3CN and the residue diluted with CH_2Cl_2 (50 mL).

The solution was washed successively with aqueous sodium bisulfite solution and saturated NaHCO₃, dried (MgSO₄) and concentrated. Column chromatography on silica gel eluted with a 1:1 mixture of ether/EtOAc removed the non-polar byproducts, and further elution with EtOAc gave the tris: (dibenzylphosphate) derivative **21** (1.76 g, 85%) as a syrup. Anal C₅₁H₃₇O₁₅P₃: (C, H, P). ¹H-NMR (CDCl₃): 5.4–4.7 (m, 15H, (CH₂, C₆H₅)₆, and 3H) 4.30 (t, J = 8.3 Hz, 1H), 3.53, 3.51, 3.45 (3s, each 3H, (OCH₃)₃), 3.10 (t, J = 9.2 Hz, 2H). ³¹P-NMR (CDCl₃): -2.42, -1.28, -0.87

(±)-3,4,5-tri-O-Methyl myo-inositol-1,2,6-tris(sodium hydrogenphosphate) **4**

A solution of the tris(dibenzylphosphate) **21** (1.71 g, 1.7 mmol) in glacial acetic acid (25 mL) containing sodium acetate (420 mg, 5.12 mmol, 3 equiv) and 10% Pd/C (500 mg) was stirred at atmospheric hydrogen pressure and at room temperature for 20 h. The catalyst was removed by filtration through a Celite pad and the residue was washed with acetic acid and water. The filtrate was concentrated and toluene (3 x 50 mL) was evaporated from the residue to remove traces of acetic acid. Evaporation of EtOH from the residue then left the solid sodium salt of the expected compound **4** (900 mg, 100%). ³¹P-NMR (D₂O): 0.0 (2P), 2.56 (1P).

(±)-1,6-di-O-Benzyl-2,3-O-isopropylidene-4-O-p-methoxybenzyl-5-O-methyl myo-inositol 23

A stored sample of the alcohol **22** (prepared as described in [17], where it was obtained as a syrup), had crystallized, and recrystallization from 1:20 EtOAc/light petroleum gave a crystalline product, mp = 68-70 °C. Anal C₃₁H₃₆O₇: (C, H). ¹H-NMR (CDCl₃): 7.4–6.8 (m, 14H, (C₆H₅)₂, and C₆H₄), 5.0–4.5 (m, 6H, (CH₂C₆H₅)₂ and CH₂C₆H₄), 4.4–3.0 (m, 2H, ring protons), 3.9–3.4 (m, 4H, ring protons), 3.79 (s, 3H, OCH₃), 2.61 (broad s. 1H, OH), 1.51, 1.35 (2s, each 3H, C(CH₃)₂). The alcohol **22** (1.0 g, 2.3 mmol) was treated with sodium hydride and methyl iodide in DMF, and the product was isolated in the usual way. Column chromatography on silica gel eluted with 1:2 ether/light petroleum mixture gave **23** (950 mg, 77%) as a syrup. Anal C₃₂H₃₈O₇: (C, H). ¹H-NMR (CDCl₃): 7.4–6.8 (m, 14H, (C₆H₅)₂ and C₆H₄), 4.9–4.6 (m, 6H, (CH₂C₆H₅)₂ and CH₂C₆H₄), 4.3–1.0 (m, 2H, ring protons), 3.7–3.6 (m, 3H, ring protons), 3.79, 3.58 (2s, each 3H, C(CH₃)₂), 3.13 (t, *J* = 8.5 Hz, 1H), 1.51, 1.34 (2s, each 3H, C(CH₃)₂).

(±)-1,6-di-O-Benzy1-2,3-O-isopropylidene-5-O-methyl myoinositol 24

A mixture of the *p*-methoxybenzyl ether **23** (1.02 g, 1.9 mmol), dichlorodicyanoquinone (500 mg, 2.2 mmol), CH_2Cl_2 (30 mL) and water (2 mL) was stirred at room temperature for 1.5 h when TLC (1:1, ether/light petroleum) showed conversion of **23** (R_f 0.6) into a product (R_f 0.15) and *p*-methoxybenzaldehyde (R_f 0.65). The mixture was diluted with CH_2Cl_2 and the organic layer washed with saturated NaHCO₃, dried over K₂CO₃ and concentrated. Column chromatography on silica gel eluted with a 1:1 ether/light petroleum removed the aldehyde, and elution with ether gave the expected alcohol **24** (691 mg, 76%) as a syrup. Anal $C_{24}H_{30}O_6$: (C, H). ¹H-NMR (CDCl₃): 7.33 (s, 10H, (C_6H_5)₂), 4.75 (s, 2H, $CH_2C_6H_5$), 4.3-4.2 (m, 1H, ring proton), 4.0–3.6 (m, 4H, ring protons), 3.58 (s, 3H, OCH₃), 2.96 (t, J = 8.5 Hz, 1H, ring proton), 2.69 (broad s, 1H, OH), 1.54, 1.34 (2s, each 3H, C(CH₃)₂). This gave a crystalline acetate **25**; mp = 77–79 °C (from light petroleum), ¹H-NMR (CDCl₃): 7.34 (s, 10H, (C₆ H_5)₂), 5.23 (dd, J = 9.8 Hz, J = 7.3 Hz, 1H, H_4), 4.82 (s, 2H, $CH_2C_6H_5$), 4.79 (AB, $\Delta\delta$ 0.14, $J_{AB} = 10.5$ Hz, 2H, $CH_2C_6H_5$), 4.24 (t, J = 4.3 Hz, 1H, ring proton), 4.1–3.8 (m, 2H, ring protons), 3.65 (dd, J = 8.8 Hz, J = 3.9 Hz, 1H, ring proton), 3.50 (s, 3H, OCH₃), 3.08 (t, J = 8.5 Hz, 1H, ring proton), 2.09 (s, 3H, OCCH₃), 1.59, 1.33 (2s, each 3H, C(CH₃)₂).

(\pm)-1,6-di-O-Benzyl-5-O-methyl myo-inositol **26** and the triacetate **27**

The alcohol **24** (620 mg, 1.5 mmol) was treated at reflux for 30 min with 1 M HCl/MeOH (1:9, 30 mL). An excess of NaHCO₃ was added, the mixture was concentrated and the product extracted with CH₂Cl₂. Column chromatography on silica gel eluted with EtOAc gave the triol **26** (520 mg, 91%) as crystals melting at 95–97 °C (from MeOH/water or EtOAc/light petroleum). Anal C₂₁H₂₆O₆, 0.25 H₂O: (C, H). A portion was acetylated as usual at 60 °C for 12 h and the product isolated in the usual way to give a crystalline triacetate **27**; mp = 99–101 °C (from EtOAc/light petroleum). Anal C₂₇H₃₂O₉: (C, H). ¹H-NMR (CDCl₃): 7.30 (broad s, 10H, (C₆H₅)₂), 5.70 (t, J = 2.4 Hz, 1H, H_2), 5.39 (t, J = 10.0 Hz, 1H, H_4), 4.9–4.4 (m, 5H, (CH₂C₆H₅)₂, and H_3), 3.6–3.5 (m, 1H, H_1), 3.56 (s, 3H, OCH₃), 3.24 (t, J = 9.2 Hz, 1H, H_5), 2.16, 2.08, 2.00 (3s, each 3H, (OCCH₃)).

(±)-3,4-di-O-Benzyl-5-O-methyl myo-inositol-1,2,6-tris(dibenzyl-phosphate) 28

The triol 26 (900 mg, 2.37 mmol) was taken up in CH₂Cl₂ (20 mL), toluene (100 mL) was added and the solution was concentrated to remove the water from crystallization present in the triol 26. The residue was taken up into CH_2Cl_2 (50 mL) and N,N-diisopropylamino-dibenzylphosphoramidite (5.0 g, 14.5 mmol) and a solution of tetrazole (1.03 g, 14.7 mmol) in CH₃CN (25 mL) was added and after 1 h stirring at room temperature, TLC (EtOAc) showed conversion of the triol $(R_{\rm f} 0.25)$ into a product $(R_{\rm f} 0.95)$. The solution was cooled at 0 °C and t-butylhydroperoxide (2.0 mL) added. After 30 min, a 10% solution of sodium metabisulfite (10 mL) was added and the solution concentrated to remove the CH₃CN. The residue was diluted with CH2Cl2 (50 mL) and the solution washed with a 10% solution of sodium metabisulfite and a saturated solution of NaHCO₃, dried over MgSO₄ and concentrated. Column chromatography on silica gel eluted with 1:1 ether/light petroleum removed the apolar byproducts and elution with ether gave compound **28** (1.50 g, 55%) as a syrup. Anal $C_{63}H_{65}O_{15}P_3$: (C, H, P). ³¹P-NMR (CDCl₃): -2.29, -1.28, -0.81.

(±)-5-O-Methyl myo-inositol-1,2,6-tris(sodium hydrogenphosphate) **5** A solution of the tris(dibenzylphosphate) **28** (1.46 g, 1.26 mmol) in glacial acetic acid (30 mL) containing sodium acetate (311 mg, 3.79 mmol, 3 equiv) and 10% Pd/C (600 mg) was stirred at atmospheric pressure at room temperature for 20 h. The product **5** (630 mg, 100%) was isolated as described for compound **4** and obtained as a white solid. ³¹P-NMR (D₂O): -0.2 (1P), + 0.67 (2P).

Irwin's test [18]

Mice were administered the test substance (three animals per tested dose) and were observed in comparison with a control group (saline). Mortality, sedation, excitation, aggressiveness, Straub, writhes, convulsions, tremor exophthalmos, salivation, lacrimation, piloerection, defaecation, fear, traction, reactivity to touch, loss of righting reflex, sleep, motor uncoordination, muscle tone, stereotypies, catalepsy, grasping, ptosis, difficulty in respiration, corneal reflex, analgaesia, gait, rectal temperature and pupil diameter were observed 15, 30, 60, 120, 180 min and 24 h after administration.

Carrageenan oedema test [19]

Mice (groups of 10 animals) were injected with a carrageenan solution into the lower surface of both hind-paws (0.75 mg per paw in 0.05 mL). The animals were sacrificed 3.5 h later by a blow to the cervical vertebrae and the hind-paws sectioned and weighed.

Acetic acid-induced writhing test [20, 21]

Mice (groups of ten animals) were injected ip with 0.5% acetic acid. The number of writhes were counted for 10 min beginning 3 min after acetic acid injection. The tested compounds were administered iv 5 min before acetic acid.

Acknowledgments

We thank E Krempp for her assistance with the NMR experiments.

References

- 1 Sirén M (1986) EPO 179440
- 2 Sirén M, Linné L, Persson L (1991) In: Inositol-Phosphates and Derivatives (Reitz AB, ed) Am Chem Soc Symp Ser 463, Washington, 103-110

- 3 Berridge MJ, Irvine RF (1984) Nature 312, 315-321
- 4 Claxson A, Morris C, Blake D, Siren M, Halliwell B, Gustafsson T, Löfkvist B, Bergelin I (1990) Agents Actions 29, 68-70
- 5 Wahlestedt C, Reis DJ, Yoo H, Adamsson A, Edvinsson L (1992) Neurosci Lett 143, 123-136
- 6 Lund T, Reed RK (1994) J Trauma 36, 761-765
- 7 Yoo H, Fallgren B, Lindhal A, Wahlestedt C (1994) Eur J Pharmacol 268, 55–63
- 8 Bieth H, Schlewer G, Spiess B (1991) J Inorg Biochem 41, 37-44
- 9 Shvets VI, Stepanov AE, Schmitt L, Spiess B, Schlewer G (1991) In: Inositol-Phosphates and Derivatives (Reitz AB, ed) Am Chem Soc Symp Ser 463, Washington, 156–171
- 10 Mernissi Arifi K, Wehrer C, Schlewer G, Spiess B (1994) J Inorg Biochem 55, 263-277
- 11 Regeling H, Zwanburg B, Chittenden GJF, Rehnberg N (1993) Carbohydr Res 244, 187–190
- 12 Malmberg M, Rehnberg N (1996) Synlett 361-362
- 13 Gogek CJ, Moir RY, McRae JA, Purves BCB (1951) Can J Chem 29, 938-945
- 14 Bongini A, Cardillo G, Orena M, Porzi G, Sandri S (1982) J Org Chem 47, 4626–4633
- 15 Fredericks PM, Guthrie RD (1975) Aust J Chem 28, 1385-1387
- 16 Perich JW, Johns RB (1987) Tetrahedron Lett 28, 101-102
- 17 Desai T, Fernandez-Mayoralas A, Gigg J, Gigg R, Payne S (1990) Carbohydr Res 205, 105–123
- 18 Irwin S (1968) Psychopharmacologica 13, 222-257
- 19 Winter CA, Risley GA, Nuss GW (1962) Proc Soc Exp Biol Med 111, 544– 547
- 20 Collier HOJ, Dinneen LC, Johnson CA, Schneider C (1968) Br J Pharmacol Chemother 32, 294–310
- 21 Charpentier J (1961) CR Soc Biol 155, 727-732
- 22 Desai T, Gigg J, Gigg R, Payne S (1992) Carbohydr Res 228, 65-79