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Activity of 6-aryl-pyrrolo[2,3-d]pyrimidine-4-amines to Tetrahymena

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ABSTRACT

A series 6-aryl-pyrrolo[2,3-*d*]pyrimidine-4-amines (43 compounds), some of which are epidermal growth factor tyrosine kinase inhibitors, were tested for their protozoal toxicity using an environmental *Tetrahymena* strain as model organism. The protozoacidal activity of the analogues was found to be highly dependent on a 4-hydroxyl group at the 6-aryl ring, and a chiral 1-phenylethanamine substituent in position 4. Further, the potency was affected by the aromatic substitution pattern of the phenylethanamine: the unsubstituted, the *meta*-fluoro and the *para*-bromo substituted derivatives had the lowest minimum protozoacidal concentrations (8–16 µg/mL). Surprisingly, both enantiomers were found to have high potency suggesting that this compound class could have several modes of action. No correlation was found between the compounds protozoacidal activity and the *in vitro* epidermal growth factor receptor tyrosine kinase inhibitory potency. This suggests that the observed antimicrobial effects are related to other targets. Testing towards a panel of kinases indicated several alternative modes of action.

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1. Introduction

Diseases caused by parasitic protozoa, as for instance malaria, dysentery, leishmaniasis, and human African trypanosomiasis are major causes of mortality throughout the world, thus, the study of effects of organic compounds on protozoa is important. Therapeutic agents are available [1,2] however, many of the drugs have critical side effects [3,4] and also resistance is emerging [5,6]. Therefore, identification of new lead compounds is required, and inhibition of cellular kinase activity has been recognised as a useful strategy [7–11]. Among others, tyrosine kinase inhibitors such as Erlotinib, Canertinib and Sunitinib designed for cancer chemotherapy have been identified as efficient antiprotozoal agents [11].

Tetrahymena is a genus of ciliated protozoa. Its members are easily grown and relatively safe to handle making them useful model systems for biochemical mechanistic studies in eukaryotes [12]. The motility behaviour of Tetrahymena is conveniently used to monitor bioactivity and cell toxicity of chemicals [13–15]. Compounds such as diphenols, aminophenols, diaminoaromatics, halogenated aromatic nitro compounds, aromatic aldehydes and α -haloketones are generally toxic to Tetrahymena. This is due to their ability to undergo various reactions with biomacromolecules [16–19]. Tetrahymena do not pose a serious threat to human health. However, Legionella in symbiosis with Tetrahymena tropicalis appears more resistant and aggressive [20,21]. In addition,

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infections attributed to members of this genus are a problem in closed fish farming. Low molecular weight compounds such as Menadione (I) [22], and anti-infective agents such as Niclosamide (II) have been proposed as treatment alternatives [23]. Other compounds with *in vitro* activity towards *Tetrahymena* include among others Climacostol (III) [24], known antimicrobial agents as Chloroquine [25] and Chloroamphenicol [26,27] and antineoplastics such as Necodazole [28], Fig. 1.

Tetrahymena are known to have epidermal growth factor (EGF)-like receptors which are involved in cell division [29], and cyst formation [30]. Also other processes such as chemotaxis [31], hormonal imprinting [32], cell division [33,34], stress response [34,35], and GTP signalling [36], are triggered and controlled by kinase activity. Using an environmental *Tetrahymena* isolate as model, we have evaluated the potency of a series of 6-aryl-7*H*-pyrrolo[2,3-*d*]pyrimidine-4-amines as antiprotozoal agents. One goal has been to identify new lead compounds for combating protozoa. Secondly, the study could shed light on the toxicity profile of this compound class since some of the derivatives are efficient inhibitors of the epidermal growth factor receptor tyrosine kinase (EGFR-TK) *in vitro* [37].

2. Materials and methods

2.1. General

¹H and ¹³C NMR spectra were recorded with a Bruker Avance 400 spectrometer operating at 400 MHz and 100 MHz,



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Fig. 1. Structure of compounds I-III with potency towards Tetrahymena and the kinase inhibitors Canertinib (IV) and Sunitinib (V).

respectively. $^{19}{\rm F}$ NMR was performed on a Bruker Avance 600 operating at 564 MHz. The $^{19}{\rm F}$ NMR shift values are relative to hexafluorobenzene. Coupling constants are in Hertz. HPLC (Agilent 110-Series) with a G1379A degasser, G1311A Quatpump, G1313A ALS autosampler and a G1315D Agilent detector (230 nm) was used to determine the purity of the synthesised compounds. Conditions: a Omrisphere 5 C18 (100×3.0 mm) column, flow rate 1.0 mL/min, elution starting with $H_2O + 1\%$ TFA/acetonitrile (98/ 2), linear gradient elution for 15 min. ending at acetonitrile/ water+1% TFA (90/10), then 15 min isocratic elution. The software used with the HPLC was Agilent ChemStation. Accurate mass determination was performed with EI (70 eV) using a Finnigan MAT 95 XL. FTIR spectra were recorded on a Thermo Nicolet Avatar 330 infrared spectrophotometer. All melting points are uncorrected and measured by a Büchi melting point instrument. Optical rotation was measured with a PerkinElmer Instruments Model 341 Polarimeter.

2.2. Isolation and characterisation of Tetrahymena

The *Tetrahymena* strain used was originally isolated from pond water in Norway and was identified to the genus level based on its phenotype and on partial sequencing of the 18S rDNA-gene. The sequence had 100% identity with reported sequences for *Tetrahymena iwoffi*, *Tetrahymena tropicalis* and *Tetrahymena furgosoni*. The strain was maintained on non-nutrient agar (CCAP, Scotland) seeded with a thick suspension of pasteurised *Escherichia coli* prior to testing. The strain and further information on the sequencing studies can be made available on request.

2.3. Determination of minimum protozoacidal (MPC) concentrations

Stock solutions of the agents were made in DMSO at a concentration of 5120 μ g/mL. Benzalkonium chloride (stock in water) was included as control. Water was used as dilutant producing doubling concentrations of the agents at 128–4 μ g/mL. These intermediate dilutions (50 μ L) were pipetted in triplicate into a 96-well, Nunc[®] round-bottomed microtiter plate system (Thermo Fischer Scientific, USA). Addition of 50 μ L of the inoculum gave the final tested concentration range (2–64 μ g/mL) and maximally 1.25% DMSO. A positive control (no agent), and a negative control (without Tetrahymena) tests were also included. *Tetrahymena* was grown on NNA seeded with a thick pasteurised suspension of

E. coli for 48 h under a humidified atmosphere in the dark at 22 ± 2 °C. After incubation, protozoa were harvested and washed as previously described [38], and resuspended in pasteurised E. coli (corresponding to a MacFarland 0.5 standard) at 1×10^4 cells/mL. After incubation for 48 h at 22 ± 2 °C, wells were examined for motile cells using an inverted microscope. This approach enabled the whole content of the well to be visualised. The estimated minimum protozoacidal concentration (MPC; 48 h) was the lowest concentration at which no motile cells were seen. After examination in the microscope, the whole content of wells was transferred to culture dishes containing NNA/pasteurised E. coli. Cultures were examined over a 7-day period with an inverted microscope to see if a cell population developed. The MPC value measured (MPC; 7 days) was the lowest concentration that prevented the development of even a single viable cell in the 7day period. Each test was performed in triplicate and the results were averaged to give the MPC value.

2.4. Kinase profiling

Compound (*R*)-**25e** was profiled utilising a panel of 124 protein kinases in the MRC National Centre for Protein Kinase Profiling Service at the University of Dundee (http://www.kinase-screen.mrc.a-c.uk). The compound was tested *in vitro*, in duplicate, at a final concentration of 50 nM. For further details of the methodology see Bain et al. [39].

2.5. Synthesis

Detailed description of the synthesis and characterisation of most of the intermediates and tested compounds can be found elsewhere [37,40]. The synthesis and characterisation of the new chemical entities are given below.

2.5.1. General procedure thermal amination to 20-24

The following is representative: 4-chloro-6-(4-methoxy-phenyl)-7*H*-pyrrolo[2,3-*d*]pyrimidine (**14**) (275 mg, 1.06 mmol) and (*S*)-1-phenylethanamine ((*S*)-**19i**) (0.44 mL, ~3.5 mmol) were added to a dry round bottle flask containing 1-butanol (3.5 mL) under argon atmosphere. The mixture was heated at 145 °C for 24 h. The precipitate formed upon cooling to rt. was isolated by filtration, washed with diethyl ether (25 mL) and dried resulting in a solid.

2.5.1.1. (*S*)-6-(4-*Methoxyphenyl*)-*N*-(1-*phenylethyl*)-7*H*-*pyrrolo*[2,3-*d*]*pyrimidin*-4-*amine* ((*S*)-**20e**). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-methoxyphenyl)-7H-pyrrolo[2,3-*d*]*pyrimidine* (**14**) (159 mg, 0.61 mmol) and (*S*)-1-phenylethanamine (**19e**) (222 mg, 1.84 mmol). This gave 179 mg (0.52 mmol, 85%) of a white solid, mp. 226–228 °C, $[\alpha]_D^{20} = + 289 (c 0.17, DMSO)$. Spectroscopic properties were in correspondence with that reported previously for the (*R*)-enantiomer [40]. ¹H NMR (400 MHz, DMSO-*d*₆) δ : 11.92 (s, 1H, NH, H-7), 8.04 (s, 1H, H-2), 7.73 (m, 3H), 7.43 (m, 2H), 7.30 (m, 2), 7.19 (m, 1H), 7.02 (d, *J* = 8.8, 2H), 6.96 (bs, 1H, H-5), 5.50 (m, 1H), 3.80 (s, 3H), 1.53 (d, *J* = 7.0, 3H). HRMS (EI): 344.1634 (calcd C₂₁H₂₀N₄O, 344.1632, M⁺).

2.5.1.2. (R)-6-(4-Methoxyphenyl)-N-(1-(4-methoxyphenyl)ethyl)-7Hpyrrolo[2,3-d]pyrimidin-4-amine ((R)-20h). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-metoxyphenyl)-7H-pyrrolo-[2,3-d]-pyrimidine (14) (238 mg, (R)-(4-methoxyphenyl)ethanamine 0.92 mmol) and (19h)(288 mg, 1.90 mmol). This gave 220 mg (0.59 mmol, 64%) of an off-white solid, mp. 249–251 °C, $[\alpha]_D^{20} = -330.1$ (c 0.14, DMSO), purity > 99% (by HPLC). ¹H NMR (400 MHz, DMSO- d_6) δ : 11.89 (s, 1H, NH, H-7), 8.04 (s, 1H), 7.71 (d, *J* = 8.9, 2H), 7.64 (d, *J* = 8.4, 1H, NH), 7.34 (d, J = 8.7, 2H), 7.02 (d, J = 8.9, 2H), 6.94 (s, 1H, H-5), 6.86 (d, J = 8.7, 2H), 5.45 (m, 1H), 3.80 (s, 3H), 3.71 (s, 3H), 1.50 (d, J = 7.0, 3H). ¹³C NMR (100 MHz, DMSO- d_6) δ : 158.6, 157.9, 154.8, 151.3 (2C), 137.5, 133.4, 127.2 (2C), 125.9 (2C), 124.5, 114.4 (2C), 113.5 (2C), 103.9, 94.6, 55.2, 55.0, 48.0, 22.9. HRMS (ESI): 375.1814 (calcd C₂₂H₂₂N₄O₂, 375.1816, M + H⁺). IR (neat, cm⁻¹): 3099, 2973, 1588, 1244, 830.

2.5.1.3. (*S*)-*N*-(1-(4-Bromophenyl)ethyl)-6-(4-methoxyphenyl)-7H-pyrrolo[2,3-d]pyrimidin-4-amine ((*S*)-**20i**). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-methoxyphenyl)-7H-pyrrolo-[2,3-d]-pyrimidine (**14**) (190 mg, 0.73 mmol) and (*S*)-1-(4-bromophenyl)ethanamine (**19i**) (439 mg, 2.19 mmol). This gave 245 mg (0.58 mmol, 79%) of a white solid, mp. 274–275 °C, $[\alpha]_D^{20} = +309 (c 0.21, DMSO)$. Spectroscopic properties were in correspondence with that reported previously for the (*R*)-enantiomer [37]. ¹H NMR (400 MHz, DMSO-*d*₆) δ : 11.93 (s, 1H, NH, H-7), 8.03 (s, 1H, H-2), 7.76 (s, 1H, NH), 7.73 (m, 2H), 7.49 (m, 2H), 7.38 (m, 2H), 7.02 (m, 2H), 6.94 (d, *J* = 1.8, 1H, H-5), 5.44 (m, 1H), 3.80 (s, 3H), 1.51 (d, *J* = 7.0, 3H). HRMS (EI): 422.0739 (calcd C₂₁H₁₉BrN₄O, 422.0737, M⁺)

2.5.1.4. 6-(4-Methoxyphenyl)-N-(1-naphthalen-1-ylmethyl)-7H-pyrrolo[2,3-d]pyrimidin-4-amine (**200**). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-methoxyphenyl)-7H-pyrrolo[2,3-d]pyrimidine (**14**) (154 mg, 0.59 mmol) and naphthalen-1-ylmethanamine (**190**) (280 mg, 1.78 mmol). This gave 173 mg (0.45 mmol, 77%) of a white solid, mp 278-281 °C. ¹H NMR (400 MHz, DMSO- d_6) δ : 11.98 (s, 1H, NH, H-7), 8.21–8.19 (m, 1H), 8.14 (s, 1H, H-2), 7.97–7.94 (m, 2H), 7.86– 7.84 (m, 1H), 7.71–7.69 (m, 2H), 7.57–7.52 (m, 3 H), 7.49–7.46 (m, 1H), 7.01–7.00 (m, 2H), 6.89 (s, 1H, H-5), 5.20 (d, *J* = 5.6, 2H), 3.79 (s, 3H). ¹³C NMR (100 MHz, DMSO- d_6), δ : 159.1, 155.8, 151.8 (2C, overlap), 135.7, 134.2, 133.8, 131.5, 128.9, 127.8, 126.6, 126.4 (2C), 126.2, 125.9, 125.7, 124.9, 124.0, 114.9 (2C), 104.4, 95.0, 55.6, 41.9. IR (neat, cm⁻¹): 3152, 1597,1254,769. HRMS (EI): 380.1632 (calcd C₂₄H₂₀N₄O, 380.1632, M⁺).

2.5.1.5. (R)-6-(4-Methoxyphenyl)-N-(1-(naphthalen-1-yl)ethyl)-7Hpyrrolo[2,3-d]pyrimidin-4-amine ((R)-**20p**). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-methoxyphenyl)-7H-pyrrolo[2,3-d]pyrimidine (**14**) (144 mg, 0.55 mmol) and (R)-1-(naphthalen-1-yl)ethanamine (**19p**) (285 mg, 1.66 mmol). This gave 162 mg (0.41 mmol, 74%) of a white solid, mp. 274–276 °C, $[\alpha]_D^{20} = -432$ (*c* 0.37, DMSO). Spectroscopic properties were in correspondence with that reported previously for the racemate [37]. ¹H NMR (400 MHz, DMSO-*d*₆) δ : 11.93 (s, 1H, NH, H-7), 8.24 (d, *J* = 8.2, 1H), 8.04 (s, 1H, H-2), 7.94 (m, 1H), 7.87 (m, 1H), 7.81 (m, 1H), 7.71–7.64 (m, 3H), 7.59–7.45 (m, 3H), 7.01 (m, 2H), 6.97 (s, 1H, H-5), 6.27 (m, 1H), 3.79 (s, 3H), 1.67 (d, *J* = 6.8, 3H). IR (neat, cm⁻¹): 3131, 2962 1624, 1251, 828, 775. HRMS (EI): 394.1783 (calcd C₂₅H₂₂N₄O, 394.1794, M⁺).

2.5.1.6. (R)-N-(1-(Naphthalen-1-yl)ethyl)-6-phenyl-7H-pyrrolo[2,3*d*]*pyrimidin-4-amine* ((*R*)-**21***p*). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-phenyl)-7H-pyrrolo[2,3-d]pyrimidine (15) (87 mg, 0.38 mmol) and (R)-1-(naphthalen-1-yl)ethanamine (19p) (195 mg, 1.14 mmol). This gave 107 mg (0.29 mmol, 76%) of a white solid. The solid melted at 155–157 °C, but partly re-solidified to a solid melting at 200 °C, $[\alpha]_D^{20} = -481$ (*c* 1.00, DMSO), purity: 98% (by HPLC). ¹H NMR (400 MHz, DMSO-d₆) δ : 12.07 (s, 1H, NH, H-7), 8.26-8.24 (m, 1H), 8.08 (s, 1H, H-2), 8.01-7.93 (m, 2H), 7.82-7.77 (m, 3H), 7.67-7.66 (m, 1H), 7.59-7.41 (m, 5H), 7.30-7.27 (m, 1H), 7.14 (s, 1H, H-5), 6.32–6.25 (m, 1H), 1.68 (d, J = 6.8, 3H). ¹³C NMR $(100 \text{ MHz}, \text{DMSO-}d_6) \delta$: 155.3, 152.3, 152.0 141.3, 133.92, 133.87, 132.3, 131.1, 129.42 (2C), 129.1, 127.7, 127.6, 126.6, 126.0, 125.9, 124.9 (2C), 123.8, 122.7, 104.4, 60.2, 96.7, 22.3. IR (neat, cm⁻¹): 2976, 1586, 1471, 1311, 774, 749. HRMS (EI): 364.1683 (calcd C24H20N4, 364.1682, M⁺).

2.5.1.7. (*S*)-*N*-(1-(*Naphthalen-1-yl*)*ethyl*)-6-*phenyl*-7*H*-*pyrrolo*[2,3-*d*]*pyrimidin-4-amine* ((*S*)-**21***p*). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-phenyl)-7*H*-pyrrolo[2,3-*d*]*pyrimidine* (**15**) (70 mg, 0.30 mmol) and (*S*)-1-phenylethanamine (**19e**) (156 mg, 0.91 mmol). This gave 84 mg (23 mmol, 77%) of a white solid, mp. 162–166 °C, $[\alpha]_D^{20}$ = +422 (*c* 1.00, DMSO), purity: 98% (by HPLC). The spectroscopic properties corresponded with that reported for (*S*)-**21***p* in Section 2.5.1.6. HRMS (EI): 364.1682 (calcd C₂₄H₂₀N₄, 364.1682, M⁺).

(R)-6-(4-Fluorophenyl)-N-(1-(naphthalen-1-yl)ethyl)-7H-2.5.1.8. pyrrolo[2,3-d]pyrimidin-4-amine ((R)-**22p**). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-fluorophenyl)-7*H*-pyrrolo[2,3-*d*]pyrimidine (16)(64 mg. and (R)-1-(naphthalen-1-yl)ethanamine 0.26 mmol) (19p) (133 mg, 0.78 mmol). This gave 70 mg (0.18 mmol, 70%) of a white solid, mp. 147–150 °C, $[\alpha]_D^{20} = -444$ (*c* 0.40, DMSO), purity: 99% (by HPLC). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 12.07 (s, 1H, NH, H-7), 8.26-8.24 (m, 1H), 8.08 (s, 1H, H-2), 7.99-7.93 (m, 2H), 7.82-7.80 (m, 3H), 7.67-7.65 (m, 1H), 7.58-7.45 (m, 3H), 7.31-7.27 (m, 2H), 7.09 (s, 1H, H-5), 6.31–6.25 (m, 1H), 1.68 (d, J = 6.8, 3H). ¹³C NMR (100 MHz, DMSO- d_6), δ : 161.9 (d, J = 244.5), 155.3, 152.3 (2C), 152.0, 141.3, 139.9, 133.0, 131.1, 129.1, 128.9 (d, *J* = 2.9), 127.6, 126.9 (d, *J* = 7.9), 126.6, 126.0 (d, *J* = 3.5, 2C), 123.8, 122.7, 116.4 (d, J = 21.7, 2C), 104.4, 96.6, 60.2, 22.2. ¹⁹F NMR (564 MHz, DMSO-*d*₆, C₆F₆) δ: -117.1 (m). IR (neat, cm⁻¹): 2985, 1585, 1496, 1312, 1233, 834, 774. HRMS (EI): 382.1585 (calcd C₂₄H₁₉FN₄, 382.1586, M⁺).

2.5.1.9. (*S*)-6-(4-Fluorophenyl)-*N*-(1-(naphthalen-1-yl)ethyl)-7H-pyrrolo[2,3-d]pyrimidin-4-amine ((*S*)-22p). The compound was prepared as described in Section 2.5.1 starting with 4-chloro-6-(4-fluorophenyl)-7H-pyrrolo[2,3-d]pyrimidine (**16**) (66 mg, 0.27 mmol) and (*S*)-1-phenylethanamine (**19p**) (137 mg, 0.80 mmol). This gave 76 mg (0.20 mmol, 74%) of a white solid, mp. 149–152 °C, $[\alpha]_D^{20} = +380$ (*c* 0.13, DMSO), purity: 98% (by HPLC). The spectroscopic properties corresponded with that reported for (*R*)-**22p** in Section 2.5.1.8.

2.5.2. General procedure demethylation of 20-25

The following is representative: (*S*)-6-(4-methoxyphenyl)-*N*-(1-phenylethyl)-7*H*-pyrrolo[2,3-*d*]pyrimidin-4-amine (**20i**) was dissolved in dry CH₂Cl₂ (2 mL) under argon atmosphere. BBr₃ (0.17 ml, ~1.8 mmol) in dry CH₂Cl₂ (1.5 mL) was added drop wise over 1 h. at 0 °C using a syringe pump. Then the mixture was allowed to react at 20 °C for 24 h. The reaction was quenched by addition of water (10 mL), and the mixture was extracted with EtOAc (3 × 25 mL). The combined organic phase was washed with brine (15 mL), dried over MgSO₄ and concentrated. The resulting residue was purified by precipitation from acetone (0.5 mL). The solid formed was isolated by filtration, washed with diethyl ether (10 mL) and dried.

2.5.2.1. (*S*)-4-(4-(1-Phenylethylamino)-7H-pyrrolo[2,3-d]pyrimidin-6-yl)phenol hydrobromide ((*S*)-**25e**). The compound was synthesised as described in Section 2.5.2 starting from (*S*)-6-(4-methoxyphenyl)-*N*-(1-phenylethyl)-7H-pyrrolo[2,3-d]pyrimidin-4-amine (**20e**) (99 mg, 0.29 mmol). This gave 56 mg, (0.14 mmol, 47%) of a white solid, mp > 300 °C, $[\alpha]_D^{20} = +289 (c 0.17, DMSO)$. purity > 98% (by HPLC) Spectroscopic properties were in correspondence with that reported previously for the (*R*)-enantiomer [37]. ¹H NMR (400 MHz, DMSO-d₆) δ : 12.98 (s, 1H, NH, H-7), 9.83 (br s, 1H, OH), 9.55 (br s, 1H, NH), 8.30 (s, 1H, H-2), 7.66 (m, 2H), 7.50–7.48 (m, 2H), 7.39 (m, 2H), 7.32–7.29 (m, 1H), 7.24 (s, 1H, H-5), 6.89 (m, 2H), 5.37 (m, 1H), 1.66 (d, *J* = 6.5, 3H). HRMS (EI): 330.1475 (calcd C₂₀H₁₈N₄O, 330.1475, M⁺).

2.5.2.2. (S)-4-(4-(1-(4-bromophenyl)ethylamino)-7H-pyrrolo[2,3d]pyrimidin-6-yl)phenol hydrobromide ((S)-**25i**). The compound was synthesised as described in Section 2.5.2 starting from (S)-N-(1-(4-bromophenyl)ethyl)-6-(4-methoxyphenyl)-7H-pyrrolo[2,3d]pyrimidin-4-amine (**20i**) (134 mg, 0.32 mol). This gave 83 mg (0.17 mmol, 52%) of a white solid, mp. 274–276 °C, $[\alpha]_D^{20} = +272$ (*c* 0.28, DMSO), purity > 97% (by HPLC). Spectroscopic properties were in correspondence with that reported previously for the (*R*)-enantiomer [37]. ¹H NMR (400 MHz, DMSO-*d*₆) δ : 12.94 (s, 1H, NH, H-7), 9.82 (br s, 1H, OH), 9.47 (br s, 1H, NH), 8.29 (s, 1H, H-2), 7.65 (m, 2H), 7.58 (m, 2H), 7.44 (m, 2H), 7.16 (s, 1H, H-5), 6.88 (m, 2H), 5.37 (m, 1H), 1.63 (d, *J* = 6.6, 3H. HRMS (EI): 408.0581 (calcd C₂₀H₁₇Br⁷⁹N₄O, 408.0580, M⁺).

2.5.2.3. 4-(4-(*Naphthalen-1-ylmethylamino*)-7*H-pyrrolo*[2,3-*d*]*pyr-imidin-6-yl*)*phenol hydrobromide* (**250**). The compound was synthesised as described in Section 2.5.2 starting from 6-(4-methoxyphenyl)-*N*-(naphthalen-1-ylmethyl)-7*H*-pyrrolo[2,3-*d*]-pyrimidin-4-amine (**200**) (117 mg, 0.31 mmol) and BBr₃ (0.29 mL, 3.1 mmol). This gave 74 mg (0.20 mmol, 66%) of a white solid, mp > 300 °C. purity > 98% (by HPLC). ¹H NMR (400 MHz, DMSO-*d*₆) δ : 13.01 (s, 1H, H-7), 9.82 (s, 1H, OH), 8.36 (s, 1H, H-2), 8.15-8.13 (m, 1H), 8.04–8.02 (m, 1H), 7.97–7.95 (m, 1H), 7.66–7.60 (m, 4H), 7.54–7.7.50 (m, 2H), 7.17 (s, 1H, H-5), 6.87–6.89 (m, 2H), 5.26 (bs, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 158.5, 149.9, 148.6, 142.8, 138.1, 133.9, 132.1, 131.3, 129.1, 128.9, 127.2 (2C), 127.1, 126.7, 126.1, 125.4, 124.1, 121.7, 116.4 (2C), 103.5, 96.8, 43.9. IR (neat, cm⁻¹): 3123, 1643, 1612, 1493, 1178, 757. HRMS (EI): 366.1471 (calcd C₂₃H₁₈N₄O, 366.1475, M⁺).

2.5.2.4. (*R*)-4-(4-(1-(*naphthalen-1-yl*)*ethylamino*)-7*H*-*pyrrolo*[2,3-*d*]*pyrimidin-6-yl*)*phenol* ((*R*)-**25***p*). The compound was synthesised as described in Section 2.5.2 starting (*R*)-6-(4-methoxyphenyl)-*N*-(1-(*naphthalen-1-yl*)*ethyl*)-7*H*-*pyrrolo*[2,3-*d*]*pyrimidin-4-amine* ((*R*)-**20***p*) (113 mg, 0.29 mmol). This gave 58 mg (0.15 mmol, 53%) of a white solid, mp > 300 °C. $[\alpha]_D^{20} = -379$ (*c* 0.26, DMSO) purity > 98% (by HPLC). Spectroscopic properties were in correspon-

dence with that reported previously for the racemate [37]. ¹H NMR (400 MHz, DMSO- d_6) δ : 12.98 (s, 1H, NH, H-7), 9.82 (br s, 1H, OH), 9.55 (br s, 1H, NH), 8.30 (s, 1H), 8.14 (s, 1H, H-2), 8.00 (m, 1H), 7.91 (m, 1H), 7.64–7.57 (m, 5H), 7.54–7.50 (m, 1H), 7.26 (s, 1H, H-5), 6.89–6.87 (m, 2H), 6.09 (m, 1H), 1.78 (d, *J* = 6.5, 3H). HRMS (EI): 380.1635 (calcd C₂₄H₂₀N₄O, 380,1637, M⁺).

2.5.2.5. (S)-4-(4-(1-(Naphthalen-1-yl)ethylamino)-7H-pyrrolo[2,3d]pyrimidin-6-yl)phenol hydrobromide ((S)-**25p**). The compound was synthesised as described in Section 2.5.2 starting (S)-6-(4methoxyphenyl)-*N*-(1-(naphthalen-1-yl)ethyl)-7H-pyrrolo[2,3d]pyrimidin-4-amine ((S)-**20p**) (73 mg, 0.19 mmol). This gave 48 mg (0.13 mmol, 68%) of a white solid, mp > 300 °C. $[\alpha]_D^{20} = +389$ (*c* 0.16, DMSO) purity > 99% (by HPLC). Spectroscopic properties were in correspondence with that reported previously for the racemate [37], and that described for (*R*)-**25p** in Section 2.5.2.4.

3. Result and discussion

3.1. Synthesis

The pyrrolopyrimidines were synthesised as described previously, Scheme 1 [37,41,42].

The first step forming the pyrroles 1–5 gave mediocre yields. We found that formation of 6-8 was the main reason for the loss in yield. The alcohol 6 may be caused by water generated in the pyrrole cyclisation, whereas the esters 7 and 8 most likely originate from fragmentation of the unstable aminoimidate. An increase in yield of **1–5** was seen when using >2 equivalents of the aminoimidate and 3 equivalents of sodium ethoxide. It has previously been suggested that such pyrroles are UV labile [42]. Discolouration of the products was seen on storage in DMSO for 1 day at room temperature. Cyclisation of 1–5 using formamide gave the 4-hydroxypyrrolopyrimidines 9-13 which all were crystalline and easily isolated. Standard chlorination gave 14-18, which also were conveniently isolated and purified if full conversion was obtained in the reaction. In the next step, thermal nucleophilic aromatic substitution on 14-18 was performed using various amines. Compound 14 was reacted with 19a-q giving 20a-q, while the 4-chloropyrrolopyrimidines **15–18** were mainly substituted with 19e-f and 19p, giving the corresponding 4-amino derivatives 21-24. Deprotection of the methoxy derivatives 20 with boron tribromide gave the phenolic compounds 25. Twelve of the compounds reported in this study are new chemical entities.

3.2. Toxicity towards Tetrahymena

The *Tetrahymena* strain used in this study was originally isolated from pond water in Norway.

It was observed that this isolate grew faster and was more vigorous than several of our culture collection *Tetrahymena* strains. These observations were considered important when choosing it as a test strain for the present study. Benzylalkonium chloride was used as a control in the testing showing a MPC value of 8 μ g/mL. We first investigated the effect of compounds having benzylamines, chiral 1-phenylethanamines and 1-naphthylethanamines as substituents in Fragment B (see Table 1), and methoxy, hydrogen, fluoro, bromo and cyano as R in Fragment A.

Most of the compounds were synthesised as their (R)-enantiomers, but the (S)-enantiomers of the methoxy substituted compound **20e**, and **20p**, **21p** and **22p** having a 1-naphthylethanamine substituent at C-4 were included to investigate the importance of stereochemistry. Testing revealed that the (R)-naphthyl derivative **20p** had an MPC-value of 32 (Table 1, entry 7), while other derivatives with R = OMe, H, F, Br and CN did not possess protozoacidal



Scheme 1. Synthesis of 20–25 using the amines 19a-q.

Table 1 Activity of the pyrrolopyrimidines **20–24** and **250-p** towards *Tetrahymena*.



Entry	Substance	R	R_1	Ar/R_2	MPC µg/mL (48 h) ^a
1	20a	OMe	Н	Н	>64
2	20b	OMe	Н	p-F	>64
3	(R)- 20e	OMe	CH ₃	Н	>64
4	(S)- 20e	OMe	CH ₃	Н	>64
5	(R)- 20h	OMe	CH ₃	<i>p</i> -MeO	>64
6	200	OMe	Н	C10H7	>64
7	(R)- 20p	OMe	CH ₃	C ₁₀ H ₇	32 ^b
8	(S)- 20p	OMe	CH ₃	C ₁₀ H ₇	>64
9	(R)- 20r	OMe	Et	Н	>64
10	21c	Н	Н	<i>m</i> -F	>64
11	21e	Н	CH ₃	Н	>64
12	(R)- 21f	Н	CH ₃	p-F	>64
13	(R) -21p	Н	CH ₃	C ₁₀ H ₇	>64
14	(S) -21p	Н	CH ₃	C ₁₀ H ₇	>64
15	22c	F	Н	<i>m</i> -F	>64
16	(R)- 22e	F	CH ₃	Н	>64
17	(R)- 22f	F	CH ₃	p-F	>64
18	(R) -22p	F	CH ₃	C ₁₀ H ₇	>64
19	(S) -22p	F	CH ₃	C ₁₀ H ₇	>64
20	(R)- 23e	Br	CH ₃	Н	>64
21	(R)- 23f	Br	CH ₃	p-F	>64
22	(R)- 24e	CN	CH ₃	Н	>64
23	(R)- 24f	CN	CH ₃	p-F	>64
24	250	OH	Н	C ₁₀ H ₇	64
25	(rac)- 25p	OH	CH ₃	C ₁₀ H ₇	64
26	(R)- 25 p	OH	CH ₃	C ₁₀ H ₇	32 ^a
27	(S)- 25p	ОН	CH ₃	C ₁₀ H ₇	64

^a The MPC values were determined by averaging three parallel measurements.

^b MPC = 32 also after 7 days.

activity with respect to Tetrahymena in the lower $\mu g/mL$ concentration range.

Solubility challenges were encountered when some of these compounds were tested, the investigation was continued with

the naphthylic compounds **250-p** which had a hydrophilic phenolic group in fragment A, (Table 1, entries 24–27). However, no drastic improvement in potency was observed. The (R)-enantiomer of **25p** (Table 1, entry 26) was the most potent and gave a similar

Table 2 Activity of compounds 25 towards Tetrahymena.



Entry	Substance	R	<i>R</i> ₂	MPC µg/ml (48 h) ^a	MPC µg/ml (7 days) ^a
1	25a	Н	Н	>64	>64
2	25b	Н	p-F	>64	>64
3	25c	Н	<i>m</i> -F	>64	>64
4	25d	Н	0-F	>64	>64
5	(R)- 25e	CH_3	Н	8/16 ^b	16/16 ^b
6	(R)- 25f	CH_3	p-F	32	32
7	(R)- 25g	CH_3	$p-CH_3$	>64	>64
8	(R)- 25i	CH_3	p-Br	8/8 ^b	8/8 ^b
9	(rac)- 25j	CH ₃	p-CF ₃	32	32
10	(R)- 25k	CH ₃	0-F	64	>64
11	(Rac)- 251	CH ₃	o-CH ₃	64	>64
12	(R)- 25m	CH ₃	m-F	16	16
13	(Rac)- 25n	CH ₃	m-CH ₃	32	32
14	(R)- 25q	Et	Н	32	32
15	(S)- 25e	CH ₃	Н	8/16 ^b	8/16 ^b
16	(S)- 25i	CH_3	Br	8	16

^a The MPC values were determined by averaging three parallel measurements. ^b Values given represent a second triplicate ground of testing.

MPC value to that of the methoxy derivative (*R*)-**20p** (Table 1, entry 7).

Then we decided to evaluate the effect of the substitution pattern in fragment B by varying R_1 (hydrogen, methyl and ethyl) and including mono *ortho*, *meta* or *para* R_2 -groups, while keeping the phenolic unit in fragment A. The compounds tested and their potencies towards *Tetrahymena* are compiled in Table 2.

The unsubstituted benzylamine derivative **25a** and three fluoro substituted benzylamine derivatives, **25b-d**, were all inactive in

the concentration range tested (Table 2, entries 1-4). However, to our satisfaction (R)-25e, PKI-166 [43], having a para-hydroxyphenyl at C-6 and a 1-phenylethanamine substituent as C-4, proved to be potent (MPC: $8-16 \mu g/mL$ Entry 5), indicating the importance of a chiral centre. Keeping the chiral R₁ group as methyl, and introducing rather conservative variations in the para position in terms of size and electronic properties, gave an MPC = $32 \mu g/mL$ for the fluoro derivative **25f**, while the methyl analogue **25g** was inactive. By changing the para substituent to bromo, (*R*)-25i, a MPC value of 8 µg/mL was obtained. Compared to the activity of the fluoro containing compound **25f** this indicates that a combination of both increased size and polarisability might be beneficial for achieving good potency. Also, the racemic trifluoromethyl derivative (*rac*)-**25***j* showed activity (MPC = $32 \mu g/mL$). Ortho and meta substitution by fluoro or a methyl substituent lowered the toxicity, but the *meta*-fluoro derivative **25m** showed appreciable protozoacidal activity (MPC: 16 µg/mL, entry 13). It was further investigated how the potency was affected by extending the chain length of R₁, but the result for **25q** containing a 1phenylpropanamine substituent at C-4 (MPC = $32 \mu g/mL$, entry 14) did not encourage further evaluation. To verify the importance of stereochemistry for the toxicity profile of these compounds we also synthesised and analysed for the effect of the (S)-enantiomers of the 1-phenylethanamine containing 25e and its para-bromo substituted derivative 25i. Both were found to be highly potent (Table 2, entries 15-16).

Some of the compounds evaluated in this study are efficient inhibitors of EGFR-TK in vitro [37]. These kinases depends on activation from the epidermal growth factor (EGF), which is a known signalling polypeptide in Tetrahymena [44]. However, we did not find any correlation between the in vitro activity towards EGFR-TK and the MPC values. This might indicate that receptors found in Tetrahymena are structurally different to the human version. To investigate if other kinases might be the target, the parahydroxyphenyl derivative 25e was evaluated against a panel of 124 kinases. Low inhibitory potency was observed in most cases (data not shown). Kinases which were inhibited to a degree of 25% or more at 50 nM are compiled in Fig. 2. Of these kinases, protein kinase B [45-47], calmodulin dependant enzymes [48,49], protein kinase C [9,50], and ERK1 type proteins [51] are found in protozoa and represent possible sites of action for the pyrrolopyrimidines.



Fig. 2. Effect of **25e** (50 nM) on the activity of a selection of kinases. PKCα: Protein kinase C alpha; JNK2: c-Jun N-terminal kinase/mitogen-activated protein kinase, TrkA: Neurotrophic tyrosine kinase receptor type 1, ERK1: extracellular-signal-regulated kinase; IKKe: inhibitory κB kinase; MAPKAP-K3: MAPK-activated protein kinase 3; CAMK 1: calmodulin-dependent kinase: MINK1: misshapen-like kinase 1 VEGFR: vascular endothelial growth factor receptor, Yes 1: Yamaguchi sarcoma viral oncogene homologue. PKBb: protein kinase B.

The toxicity/potency of the compounds presented in this study against Tetrahymena crucially depended on the presence of a para phenolic group in fragment A. This might be due to better solubility and bioavailability than for 20-24, or that the hydroxyl function engages in critical bonding interaction. In Tetrahymena pyriformis there is evidence for a transmembrane efflux pump as a detoxification mechanism, preferably excluding lipophilic compounds [52]. This might be an alternative explanation for the observed toxicity differences seen on introduction of the 4-hydroxyl group in fragment A. As Tetrahymena has only limited CYP-450 dependent metabolism activity [53], it is less likely that the toxicity is due to oxidation of the para-hydroxyphenyl unit of 25 leading to aromatic 1,2-dienones, which are typical Michael acceptors in reaction with bio macromolecules [16,18]. The low level of CYP-450 enzymes should also exclude the possible formation of active compounds from putative precursors such as the C-6 phenyl substituted derivative 21. Furthermore, the MPC values were affected by the substitution pattern and the presence of a chiral centre in fragment B. All the above indicates that there is a specific mode of action involving a defined 3-dimentional receptor target. However, the fact that both enantiomers showed toxicity is suggesting that the pyrrolopyrimidines could have multiple modes of action, or that the target is rather flexible.

As a model for unspecific toxicity the study shows that introduction of a 4-hydroxyl group in fragment A could be problematic for EGFR-TK inhibitors, but also that the toxicity profile could be modulated by the substitution pattern both in fragment A and B. Human toxicity issues have in fact been seen for **25e** [54].

4. Conclusion

A series of pyrrolopyrimidines have been tested for their protozoacidal activity against *Tetrahymena*. Five compounds were found to be highly active (MPC 8–16 μ g/mL). The identified compounds do not contain the typical groups which trigger non-specific toxicity effects. The presence of a *para* phenolic group in position 6 (fragment A), and a chiral centre in the 4-benzylamine (fragment B) enhanced the potency considerably. *Ortho*-substitution in fragment B, and an electron donating methyl group in the *para* position reduced the toxicity. There is no evidence that EGFR-TK kinases are targets for these compounds in *Tetrahymena*, however, kinase profiling identified other potential sites of action. The detailed mechanism will be investigated in continuing work. The presented structure–activity relationships could be used as guidelines for targeting other, medically more important protozoa.

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