Natural Product Synthesis

Enantioselective Total Synthesis of Dysidavarone A, a Novel Sesquiterpenoid Quinone from the Marine Sponge *Dysidea avara*

Yurie Fukui, Koichi Narita, and Tadashi Katoh*^[a]

Dedicated to Professor Philip D. Magnus FRS on the occasion of his 70th birthday

Abstract: Dysidavarone A, a structurally unprecedented sesquiterpenoid quinone, was synthesized in 30% overall yield in a longest liner sequence of 13 steps from commercially available *o*-vanillin. A highly strained and bridged eight-membered carbocyclic core was established by the C7–C21 carbon bond formation through a copper enolate mediated Michael addition to the internal quinone ring.

In 2012, Lin et al. reported the isolation and structural elucidation of four novel sesquiterpenoid quinones, dysidavarones A-D (1-4, Figure 1), from the marine sponge, Dysidea avara, collected along the coast of Yongxing Island in the South China Sea.^[1] These marine natural products were found to have an unprecedented tetracyclic dysidavarane carbon skeleton (see structure 5) with a highly strained and bridged eight-membered carbocycle.^[1] Lin et al. proposed that this unusual tetracyclic carbon skeleton might be produced biogenetically from well-known drimane-type sesquiterpenoid quinones, avarone (6) and neoavarone (7), by the C7–C21 bond formation through an intramolecular Prins-type reaction.^[1] Compounds 1 and 4 have shown inhibitory activity against protein tyrosine phosphatase 1B (PTP1B) with IC $_{50}$ values of 9.98 and 21.6 $\mu \text{M},$ respectively.^[1] PTP1B is a major negative regulator in both insulin and leptin signalling pathways, and it acts as a positive regulator in the tumorigenesis and progression of cancers.^[2] It has also been shown that 1 and 4 exhibit antiproliferative activities against several human cancer cell lines in the micromolar range ($IC_{50} = 11.6-28.8 \mu M$).^[1] More recently, **1** has been demonstrated to show potent inhibitory effects against Gram-positive bacteria, in particular against various Staphylococci $(MIC_{50} = 0.2-9.9 \ \mu g \ mL^{-1})$.^[3] Therefore, we anticipated that dysidavarones may be promising candidates or provide new leads for the treatment of pathogenic states such as diabetes, obesi-

[a]	Y. Fukui, K. Narita, Prof. Dr. T. Katoh
	Laboratory of Synthetic and Medicinal Chemistry
	Faculty of Pharmaceutical Sciences
	Tohoku Pharmaceutical University
	4-4-1 Komatsushima, Aoba-ku, Sendai, 981-8558 (Japan)
	Fax: (+81)22-727-0135
	E-mail: katoh@tohoku-pharm.ac.jp
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Figure 1. Structures of dysidavarones A–D (1–4), dysidavarane skeleton (5), avarone (6) and neoavarone (7).

ty, cancer and infections. However, further biological studies of these natural products are severely restricted by the scarcity of samples from marine sponge *Dysidea* species (0.0002–0.0004% yields upon isolation). Consequently, developing methods for synthesis of **1–4** is desirable and worthwhile from the viewpoint of medicinal chemistry and pharmaceuticals.

Their unique structural features, attractive biological activities and limited availability from natural resources have made 1–4 exceptionally intriguing and timely targets for total synthesis. Recently, while our synthetic studies of 1–4 were in progress, Menche et al. reported the first and elegant total synthesis of 1, which was rendered in a longest liner sequence of 10 steps with 11% overall yield.^[3] Their synthetic route is outlined in Scheme 1, where the strategic Pd-catalyzed intramolecular α -arylation^[4] of appropriately elaborated ketone 10, which was prepared from known (+)-enone 8^[5] and aryl bromide 9, to construct the requisite tetracyclic structure 11 was the key step (10 \rightarrow 11, 66% yield).^[3] Herein, we describe our total synthesis of 1 using an alternative strategy that involves an advantageous intramolecular Michael addition to construct the requisite tetracyclic core structure.

Our retrosynthetic analysis of dysidavarone A (1) is illustrated in Scheme 2. We envisioned that the requisite tetracyclic structure **13** could be established from appropriately functionalized

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Scheme 1. Outline of the total synthesis of dysidavarone A (1) by Menche et al.



Scheme 2. Retrosynthetic analysis of dysidavarone A (1).

decalone 14 by the C7–C21 bond formation through a Michael addition of the C8 carbonyl derived enolate to the internal quinone ring,^[6] followed by in situ air oxidation of the resulting hydroquinone. To the best of our knowledge, this type of cyclization to construct a highly strained and bridged eight-membered carbocycle fused with a quinone ring has not been previously reported; thus, this approach posed a considerable challenge from a synthetic viewpoint.^[7] The cyclization product 13 could be converted into the target molecule 1 via the advanced intermediate 12 by deprotection and functional-group manipulation or vice versa. The cyclization precursor 14, in turn, was to be prepared by the stereocontrolled reductive alkylation of known optically active enone 8^[5] and benzyl bromide 15, which is accessible from commercially available inexpensive o-vanillin (16), under Birch conditions. This devised scheme involves the substitution reaction of the alkoxy group in the final stage of the synthesis (see $12 \rightarrow 1$); therefore, this method is applicable to the synthesis of structural analogues of 1 possessing various alkoxy groups on the guinone ring.

We first pursued the synthesis of intermediate **15**, the coupling partner of the decalin portion (see structure **8**), starting



Scheme 3. Synthesis of intermediate 15. Reaction conditions: a) TBSCl, imidazole, CH_2Cl_2 , RT, 12 h, 97%; b) NaBH₄, THF/H₂O, 0 °C, 30 min, 99%; c) PBr₃, CH_2Cl_2 , 0 °C to RT, 5 min, 98%. TBS = *tert*-butyldimethylsilyl.

from *o*-vanillin (**16**) (Scheme 3). After protection of the hydroxy group in **16** (97% yield), the formyl group in the resulting *tert*-butyldimethylsilyl (TBS) ether **17** was reduced with NaBH₄ to provide benzylic alcohol **18** in quantitative yield. Subsequent bromination of **18** (PBr₃, CH₂Cl₂, 0°C to RT) afforded the desired intermediate **15** in 98% yield.

Having synthesized intermediate **15**, we approached the synthesis of intermediate **14**, a precursor of the key cyclization reaction, as shown in Scheme 4. The critical reductive alkyla-



Scheme 4. Synthesis of intermediate 14. Reaction conditions: a) Li, NH₃/THF, -78 to -30 °C, 45 min; isoprene, 15, -78 to -30 °C, 2 h, 81%; b) TBAF, THF, RT, 30 min, 98%; c) O₂ (1 atm), salcomine, MeCN, RT, 15 h, 86%. TBAF = tetra-*n*-butylammonium fluoride, salcomine = *N*,*N'*-bis(salicylidene)ethylenediaminocobalt(II).

tion of enone **8**^[5] (>99% *ee*) with **15** under Birch conditions^[8] proceeded smoothly and cleanly to give the expected coupling product **19** in 81% yield as the single diastereomer. Deprotection of the TBS group in **19** resulted in the formation of hemiacetal **20** in 98% yield. The stereochemistry at the C8 position in **20** was confirmed by NOESY experiment (see the Supporting Information). To construct the quinone system directly, **20** was allowed to react with molecular oxygen (O₂ balloon) in the presence of salcomine, that is, *N*,*N'*-bis(salicylidene)ethylenediaminocobalt(II),^[9] in acetonitrile at ambient temperature for 15 h, thus producing the desired quinone **14** in high yield (86%).

After obtaining intermediate **14**, we next investigated the key cyclization reaction to construct the requisite dysidavarane skeleton ($14 \rightarrow I \rightarrow II \rightarrow 13$; see scheme above Table 1). The sequence involved Michael addition of the metallic enolate I^[10,11] to the internal quinone ring and subsequent air oxidation of





the resulting intermediate II^[12] during the cyclization reaction. To achieve cyclization, initial screening was carried out using several metal amide reagents, such as LiNiPr₂, LiN(SiMe₃)₂, NaN-(SiMe₃)₂ and KN(SiMe₃)₂ in a THF solvent (entries 1-4, Table 1).^[10,11] Among these reagents, LiN(SiMe₃)₂ proved to be superior from the viewpoint of reaction cleanliness (entry 2, Table 1); however, the yield of the desired cyclized product 13 was not satisfactory (28%). In the cases of LiNiPr2 and KN-(SiMe₃)₂, the reaction was not clean, and the complete decomposition of material was observed (entries 1 and 4, Table 1). When NaN(SiMe₃)₂ was used, only a trace amount of 13 (~5% yield) was produced and a large amount of the starting material 14 (90-95% yield) was recovered (entry 3, Table 1). After repeated attempts, we found that the addition of CuBr·SMe₂ was extremely effective and reliable for this cyclization event (entry 5, Table 1). Thus, reaction of 14 with LiN(SiMe₃)₂ (5 equiv) in the presence of CuBr·SMe₂ (2 equiv) in a dilute THF solution (1 mm) from -40 °C to RT over the course of 48 h, resulted in the formation of 13 in high yield (84%). In this reaction, we believed that copper enolate I (M = Cu) was generated in situ^[10] and that this intermediate species promoted the intramolecular Michael addition reaction. Thus, the expected cyclization reaction proceeded in a clean and efficient way. To the best of our knowledge, this is the first successful example of



Scheme 5. Completion of the total synthesis of dysidavarone A (1). Reaction conditions: a) 3 \mbox{M} HCl, THF, RT, 62%; b) Ph₃P⁺CH₃Br⁻, tBuOK, benzene, RT to reflux, decomposition; c) 30% Na₂S₂O₄, Me₂SO₄, 30% KOH, *n*Bu₄NBr, THF, RT, 80%; d) 3 \mbox{M} HCl, THF, 40 °C, 98%; e) Ph₃P⁺CH₃Br⁻, tBuOK, benzene, reflux, 95%; f) 3 \mbox{M} HCl, THF, reflux, 87%; g) CAN, MeCN/H₂O, -7 °C, 97%; h) DBU, EtOH, RT, 90%. CAN = ceric ammonium nitrate, DBU = 1,8-diazabicyclo-[5.4.0]undec-7-ene.

Michael addition reaction of a copper enolate with a quinone to construct a highly strained medium-sized carbocycle.

The final route that led to the completion of the total synthesis of 1 is shown in Scheme 5. Initial attempts to realize the direct conversion of diketone 21, which was prepared from 13 by deprotection of the ethylene acetal moiety, into bis-exoolefin 22 under Wittig methylenation conditions^[13] (Ph₃P⁺ CH₃Br⁻, tBuOK, benzene, RT to reflux) were unsuccessful and resulted in decomposition of material. This result is likely due to the presence of the sensitive guinone function in 21. Therefore, we decided to mask the reactive guinone system in the form of a *p*-dimethoxybenzene derivative.^[14] Thus, methoxyguinone 13 was converted into the corresponding trimethoxybenzene 23 (80% yield) by reductive methylation (30% Na₂S₂O₄, Me₂SO₄, 30% KOH, nBu₄NBr, THF, RT). After deprotection of the ethylene acetal moiety in 23 (98% yield), the resulting diketone 24 was subjected to two-fold Wittig methylenation to produce bis-exo-olefin 25 in high yield (95%). The subsequent site-selective isomerization of the C4 exo olefinic double bond in 25 (3 M HCl, THF, reflux)^[3] furnished the corresponding endo olefin 26 in 87% yield. The guinone system was then regenerated efficiently by treating 26 with ceric ammonium nitrate (CAN) in acetonitrile/water at low temperature (-7° C), giving rise to the desired methoxyquinone 12 in 97% yield. Finally, exposure of 12 to ethanol in the presence of 1,8-diazabicyclo-[5.4.0]undec-7-ene (DBU) at ambient temperature resulted in the formation of the targeted dysidavarone A (1) in 90% yield.^[15] The spectroscopic properties (IR, ¹H, and ¹³C NMR spectroscopy, and high-resolution mass spectrometry) of synthetic 1 were identical to those of natural 1.^[1] The optical rotation of

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synthetic **1**, $[\alpha]_{D}^{25} = +105.6$ (c = 1.00, MeOH), matched that reported previously, natural **1**: $[\alpha]_{D}^{25} = +30$ (c = 1.00, MeOH),^[1, 16] synthetic **1**: $[\alpha]_{D}^{25} = +125$ (c = 1.00, MeOH).^[3]

In summary, we have accomplished the enantioselective total synthesis of dysidavarone A (1) in 30% overall yield in 13 steps from the starting material *o*-vanillin (16). The most crucial step of the synthesis involved the formation of the eight-membered carbocyclic core by the strategic intramolecular Michael addition/in situ air oxidation to establish the tetracyclic dysidavarane skeleton ($14 \rightarrow 13$, Table 1). On the basis of this study, we are currently synthesizing additional analogues of 1 (for example, analogues possessing a variety of alkoxy groups on the quinone ring) with the aim of exploring its structure–activity relationships. In addition, further investigations to identify additional action mechanisms of 1 using the synthetic sample are in progress in our laboratories.

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