



Synthesis of dipeptides from *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione activated α -amino acids

Shaik Nowshuddin, A. Ram Reddy *

Department of Chemistry, University College of Science, Osmania University Campus, Osmania University, Hyderabad, Andhra Pradesh 500 007, India

ARTICLE INFO

Article history:

Received 27 October 2010

Accepted 23 December 2010

Available online 2 February 2011

ABSTRACT

A simple two step procedure for the synthesis of a dipeptide from *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione (HO-ASUD) activated α -amino acids is described. In presence of DCC, *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione readily esterifies the carboxylic acid group of all the *N*-protected amino acids to yield crystalline *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione activated carboxy ester. The *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione activated carboxy esters of *N*-protected amino acids readily condensed with other amino acids and gave a dipeptide. This new method is effective for the DCC coupling of a variety of chiral amino acids without loss of enantiomeric purity. Synthesis of fifteen dipeptides including the hitherto unreported Fmoc-L-Orn(Boc)-Val-OMe, Fmoc-L-Cys(trt)-Gly-OEt and Boc-L-Tyr-Gly-OEt is presented.

© 2011 Elsevier Ltd. All rights reserved.

1. Introduction

Peptide synthesis is one of the most important and widely studied chemical transformations in organic synthesis. Suitability of the protecting and activating groups^{1–5} is one of the major deterrent problems that necessitate modification of the prevailing peptide methodologies.

There are numerous reports on the use of activated spiro esters in peptide synthesis.^{6–11} The *N*-hydroxy compounds used to activate the *N*-protected amino acids were NOSU (*N*-hydroxy-succinimide)^{12–14} 1-hydroxybenzotriazole¹⁵ (HOBT) and *N*-hydroxyphthalimide (HOPh).¹⁶ Recently, it was observed that HOBT is not safe to use under ambient conditions. In the present investigation, we report a facile two step synthesis of fifteen dipeptides including the hitherto unreported Fmoc-L-Orn(Boc)-Val-OMe, Fmoc-L-Cys(trt)-Gly-OEt and Boc-L-Tyr-Gly-OEt involving *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione (OH-ASUD) activated amino acid esters as intermediates and DCC as a coupling reagent in THF.

2. Results and discussion

The HO-ASUD is a stable reagent. It was prepared starting from cyclohexanone following Stevens et al.¹⁷ A typical procedure for the synthesis of activated Boc-alanine-ASUD ester is shown in Scheme 1. When equimolar amounts of Boc-alanine and HO-ASUD are stirred overnight in THF at room temperature in the presence of

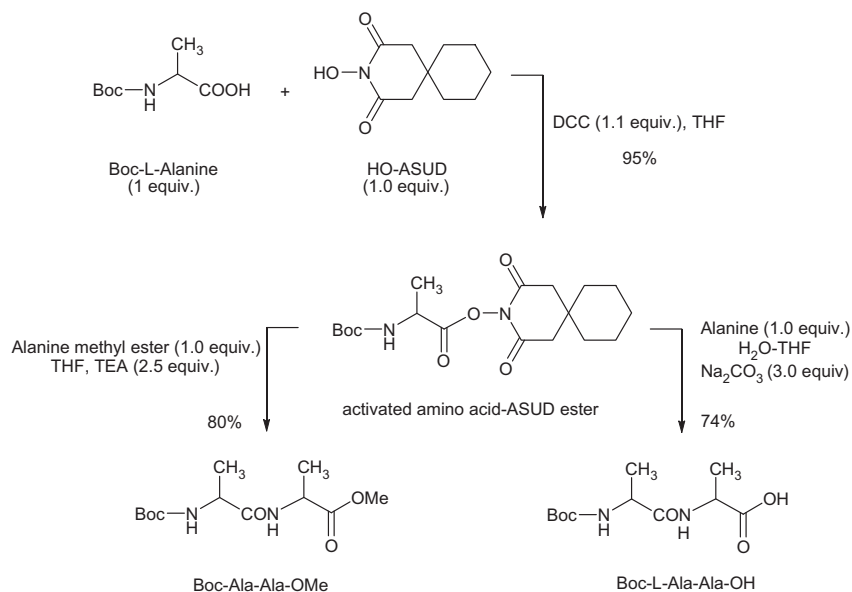
1.1 equiv of DCC, its activated ester, Boc-alanine-ASUD, is obtained. Similarly conversion of Fmoc, Boc and Z-protected amino acids to their respective ASUD esters was carried out by stirring equimolar amounts of the protected amino acids and HO-ASUD overnight at room temperature in the presence of DCC in THF. Similarly the hydroxyl group of HO-ASUD can also readily be esterified with *N*- and side-chain-protected amino acids. Therefore, formation of a protected amino acid-ASUD ester using DCC occurs with all of the amino acids. During the course of the reaction, the reaction mixture developed a white suspension due to the formation of insoluble dicyclohexylurea. The activated amino acid spiro ester is stable and readily crystallizes, during which process the unreacted HO-ASUD was removed. The formation of activated amino acid-ASUD ester can be readily monitored by TLC as well as by the appearance of a new band around 1745 cm^{−1} in the infrared spectrum, due to the spiro ASUD ester.

It was found that esterification of *N*-protected α -amino acids with HO-ASUD can be carried out not only in THF but also in other organic solvents such as acetone, dichloromethane, chloroform, toluene and acetonitrile, but not in *n*-hexane and water.

When the activated amino acid-ASUD ester is charged and stirred overnight with equimolar amounts of another α -amino acid in water-THF (1:1 v/v) and 2.5–3.0 equiv of Na₂CO₃ at room temperature, an *N*-protected dipeptide was obtained. Similarly an *N*-protected dipeptide methyl ester was prepared using 2.5 equiv of triethylamine in place of Na₂CO₃ in THF at room temperature along the lines of Scheme 1. The overall yield of dipeptides obtained is very high ranging between 69% and 86% as shown in Table 1. No premature deblocking of the *N*-protecting and other side chain groups occurred either during esterification or peptide bond

* Corresponding author.

E-mail address: aramreddy2008@yahoo.com (A. Ram Reddy).



Scheme 1. Synthesis of Boc-Ala-Ala-OH and Boc-Ala-Ala-OMe involving *N*-hydroxy-3-azaspiro[5,5]undecane-2,4-dione activated amino acid ester intermediate.

Table 1
Dipeptides prepared involving the activated amino acid-ASUD ester intermediate

S. No.	Product	Obtained yield	Reported yield	Obtained ORD ^b	Reported ORD c 1 $[\alpha]_D^{20}$
1	Z-Gly-Gly-OEt	76	60 ²⁰	—	—
2	Fmoc-Gly-Val-OMe	88	89 ²¹	+18.2	+18.6 (CHCl ₃)
3	Z-L-Pro-Val-OH	80.5	78 ²²	−12.6	−13.4 (DMF)
4	Boc-L-Ala-Ala-OMe	80	66.7 ^{23a}	−50.8	−51.5 (MeOH)
5	Boc-L-Ala-Ala-OH	74	57 ^{23a}	−42.6	−43.1 (MeOH)
6	Boc-L-Phe-Gly-OEt	71	78 ²⁰	−4.7	−4.2 (EtOH)
7	Z-L-Trp-Ala-OH	83	77 ^{23b}	−38.5	−40.1 (DMF)
8	Z-L-Gly-Leu-OH	69	56 ²⁰	−18.3	−18.2 (1 N NaOH)
9	Fmoc-L-Orn(Boc)-Val-OMe ^a	78	New	−29.2 (MeOH) ^c	—
10	Boc-L-Lys(Z)-Gly-OMe	86	83 ¹³	−12.9	−13.1 (MeOH)
11	Boc-L-Pro-Pro-OH	72	74 ^{23c}	−123.2	−121.5 (MeOH)
12	Fmoc-L-Cys(trt)-Gly-OEt ^a	84	New	+34.7 (MeOH) ^d	—
13	Z-L-Glu(OMe)-Dihydroxy-Tyr-OMe	78.6	69 ^{23d}	−8.3	−8.5 (MeOH)
14	Boc-Gly-Pro-OH	76.4	71 ^{23e}	−22.4	−23.9 (EtOH)
15	Boc-L-Tyr-Gly-OEt ^a	75	New	+3.3(MeOH) ^e	—

^a New peptides fully characterized by ¹H NMR, ¹³C NMR, IR and Mass spectra.

^b Observed values are obtained under the identical conditions of the reported literature conditions. ORD = $[\alpha]_D^{20}$ (c 10 mg/ml, CH₃OH).

^c Chiral HPLC: Chiral PACK-AS-H; *n*-Hexane/IPA/TFA (50:50:0.1%); %ee: 98.29.

^d Chiral HPLC: Chiral PACK-AS-H; *n*-Hexane/IPA/TFA (80:20:0.1%); %ee: 99.78.

^e Chiral HPLC: Chiral PACK-AD-H; *n*-Heptane/IPA/TFA (90:10:0.1%); %ee: 99.96.

formation. The co-product HO-ASUD was washed off with 5% Na₂CO₃ solution. A control experiment was performed using the *N*-protected amino acid-*N*-hydroxy succinimide method¹⁸ for TLC analysis and enantiomeric purity. The purity of the *N*-protected dipeptides was monitored by HPLC¹⁹ and compared with that of the respective peptides obtained by the *N*-OSU method. The structure of all the dipeptides prepared was characterized by micro analysis, IR, NMR and Mass spectra and compared with the reported literature. The IR spectrum correlated more than 95% with standard samples. All of the dipeptides prepared by the present method and NOSU method are listed in Table 1. Entries 9, 12 and 15 in the table are new dipeptides, which are hitherto unreported. Their structure is established by mass, ¹H NMR, ¹³C NMR and infra red spectra. The ¹³C NMR has peaks at 169, 165 and 156 ppm, respectively for the newly formed peptide carbonyl carbon, respectively. The proton NMR spectra of Fmoc-L-Orn(Boc)-Val-OMe, Fmoc-L-Cys(trt)-Gly-OEt and Boc-L-Tyr-Gly-OEt exhibited new sig-

nals at 8.3, 9.07 and 9.04 ppm, respectively representing the newly formed peptide proton. Similarly the pseudo molecular ions obtained at 689 (M⁺+H₂O) and 365 (M−1) in the ES Mass confirms the structure of 12 and 15. In the case of entry 9, no molecular ion peak in the mass spectrum was obtained. However the fragment obtained at *m/z* 512 is the ion obtained after the loss of an OMe and the side chain of valine.

The present method of dipeptide synthesis has the following merits. Preparation of HO-ASUD is simple. It has shelf stability at room temperature and is not hygroscopic like *N*-OSU. It is highly soluble in most of the organic solvents employed in peptide synthesis. It retains all the advantage of DCC coupled reactions. HO-ASUD is more reactive than *N*-OSU and gave comparable yields in less time. The overall yield of dipeptide with the present method and with the *N*-OSU as activating group under identical conditions is shown in Table 1. It can be noticed from Table 1 that HOASUD activation is comparable with the *N*-OSU method. The

enantiomeric purity was measured by optical rotatory dispersion (ORD) and compared with standard samples.¹⁸ It has been found that the dipeptide formation occurred with complete preservation of stereochemistry. HO-ASUD activation of protected amino acids is not sensitive to any particular protecting group or amino acid and can be widely employed in peptide synthesis.

3. Conclusion

Synthesis of N-protected dipeptides and N-protected dipeptide methyl esters via N-protected L-amino acid activated ASUD ester intermediates is simple and can be carried out not only in THF but also in other organic solvents such as acetone, dichloromethane, toluene and acetonitrile. The overall yield of the dipeptides obtained is high and comparable to that of N-OSU.¹² We have prepared three new dipeptides involving these activated ASUD ester intermediates.

4. Experimental

4.1. Preparation of Boc-L-alanine-ASUD ester

Boc-L-alanine 1.0 g (0.0052 mol), OH-ASUD 1.04 g (0.00529 mol), DCC 1.19 g (0.0058 mol) and THF (25 ml) were stirred in a 100 ml RB flask at room temperature for 12 h. The dicyclohexylurea was filtered and the filtrate was concentrated to give the Boc-L-alanine-ASUD ester. Yield 1.9 g (100%).

4.2. Preparation of Boc-L-Ala-ala-OH 5

At first, L-alanine 0.45 g (0.00516 mol) and Na₂CO₃ 1.64 g (0.01548 mol) were dissolved in 16 ml of water at room temperature in a 100 ml three necked round bottomed flask. Boc-L-alanine-ASUD ester 1.9 g (0.00516 mol) dissolved in 16 ml of THF was added to the above reaction mixture at room temperature over a period of 5 min. After the addition was complete, the reaction mixture was stirred for 10 h at room temperature. The reaction mixture was acidified with 5% KHSO₄ to reach a pH of 2–3. The product was extracted into ethyl acetate. The ethyl acetate extract was washed with 5% Na₂CO₃ solution, water, brine solution and dried over anhydrous Na₂SO₄. The ethyl acetate was removed at reduced pressure. The oily crude product obtained was crystallized using ethyl acetate and *n*-hexane. The yield of white crystalline Boc-L-Ala-Ala-OH is 0.98 g (74%).

4.3. Preparation of Boc-L-Ala-Ala-OMe 4

At first, Boc-L-alanine-ASUD ester 1.9 g (0.00516 mol), L-alanine methyl ester hydrochloride 0.71 g (0.00516 mol), triethylamine 1.30 g (0.0129 mol) and THF 20 ml were charged into a 100 ml three necked round bottomed flask at room temperature. The reaction mixture was stirred over night at room temperature. The triethylamine hydrochloride salt was filtered and the filtrate was concentrated under vacuum to give an oily crude. The oily crude was dissolved in ethyl acetate and washed with 0.5% NaHCO₃ solution and water. The ethyl acetate layer was dried over Na₂SO₄ and distilled under vacuum to give an oily crude product. The yield of Boc-L-Ala-Ala-OMe is 1.1 g (80%).

4.4. Preparation of Fmoc-L-Orn(Boc)-ASUD ester

At first, Fmoc-L-Orn(Boc)-OH 1.0 g (0.0022 mol), OH-ASUD 0.43 g (0.0022 mol), DCC 0.45 g (0.0022 mol) and THF 10 ml were stirred in a 50 ml three necked round bottomed flask at room temperature for 12 h. The dicyclohexylurea was filtered and concen-

trated to give a colourless viscous oily liquid of Fmoc-L-Orn(Boc)-ASUD ester. Yield 1.3 g (100%).

4.5. Preparation of Fmoc-L-Orn(Boc)-Val-OMe 9

L-Valine methyl ester 0.27 g (0.002 mol) and Na₂CO₃ 0.63 g (0.006 mol) were dissolved in 16 ml of water at room temperature in a 100 ml three necked round bottomed flask. Next, Fmoc-L-Orn(Boc)-ASUD ester 1.3 g (0.002 mol) dissolved in 16 ml of THF was added to the above reaction mixture at room temperature over a period of 5 min. After the addition was complete, the reaction mixture was stirred for 10 h at room temperature. The reaction mixture was acidified with 5% KHSO₄ to reach a pH of 2–3. The product was extracted into ethyl acetate. The ethyl acetate extract was washed with 5% Na₂CO₃ solution, water, brine solution and dried over anhydrous Na₂SO₄. The ethyl acetate was removed at reduced pressure. The oily crude product obtained was crystallized using ethyl acetate and *n*-hexane. The yield of pale yellow crystalline Fmoc-L-Orn(Boc)-Val-OMe is 0.9 g (78%). Mp: 155–157 °C, ¹H NMR (300 MHz, CDCl₃-d): δ 7.1–7.7 (m, 8H), δ 5.92 (d, 1H), δ 5.5 (dd, 1H), δ 4.1–4.8 (m, 2H), δ 3.37 (s, 3H), δ 3.0 (d, 1H), δ 2.5–2.8 (m, 2H), δ 1.9 (m, 2H), δ 1.7 (m, 2H), δ 1.50–1.59 (s, 9H), δ 0.77–0.98 (dd, 6H); ¹³C NMR (75 MHz, CDCl₃-d): δ 168.93, 168.69, 157.45, 156.43, 144.04, 141.22, 127.69, 127.08, 125.17, 119.9, 79.1, 77.5, 76.66, 67.2, 54.6, 49.2, 47.1, 44.0, 39.8, 36.6, 35.8, 33.7, 29.8, 28, 26.9, 18.1; IR (cm⁻¹): 3332.56, 2930.63, 2853.36, 164.71, 1627.82; MS *m/z* 512.2 (M-OMe and side chain of valine). [α]_D²⁰ = -29.2 (c 1, CH₃OH); Chiral HPLC: Chiral PACK-AS-H; *n*-Hexane/IPA/TFA (50:50:0.1%); %ee: 98.29; Elemental analysis; observed (theoretical): C = 65.13 (65.59); H = 7.21 (7.28); N = 7.35 (7.4).

4.6. Preparation of Fmoc-L-Cys(trt)-ASUD ester

Fmoc-L-Cys(trt)-OH 1.5 g (0.0025 mol), OH-ASUD 0.5 g (0.0025 mol), DCC 0.52 g (0.0025 mol) and THF (15 ml) were stirred in a 50 ml three neck round bottomed flask at room temperature for 12 h. The dicyclohexylurea was filtered and the filtrate concentrated to give Fmoc-L-Cys(trt)-ASUD ester. Yield 1.71 g (100%).

4.7. Preparation of Fmoc-L-Cys(trt)-Gly-OEt 12

Glycine ethyl ester 0.21 g (0.0022 mol) and Na₂CO₃ 0.7 g (0.0066 mol) were dissolved in 20 ml of water at room temperature in a 100 ml three neck round bottomed flask. Fmoc-L-Cys(trt)-ASUD ester 1.71 g (0.0022 mol) dissolved in 20 ml of THF was added to the above reaction mixture at room temperature over a period of 5 min. After the addition was complete, the reaction mixture was stirred for 10 h at room temperature. The reaction mixture was acidified with 5% KHSO₄ to reach a pH to 2–3. The product was extracted into ethyl acetate. The ethyl acetate extract was washed with 5% Na₂CO₃ solution, water, brine solution and dried over anhydrous Na₂SO₄. The ethyl acetate was removed at reduced pressure. The oily crude product obtained was crystallized using ethyl acetate and *n*-hexane. The yield of white crystalline Fmoc-L-Cys(trt)-Gly-OEt is 1.25 g (84%). Mp: 78–82 °C, ¹H NMR (300 MHz, CDCl₃-d): 7.1–7.4 (m, 24H), δ 4.6 (s, 1H), δ 4.2 (s, 2H), δ 4.1 (d, 1H), δ 3.5 (m, 4H), δ 2.8–3.1 (dd, 2H), δ 1.7 (s, 3H); ¹³C NMR (75 MHz, CDCl₃-d): δ 178.0, 165.47, 156.99, 144.49, 144.10, 129.49, 128.16, 127.98, 127.89, 127.05, 126.87, 77.5, 77.10, 76.67, 67.53, 53.19, 49.05, 36.1, 29.03, 26.4, 14.1; IR (cm⁻¹): 3327.94, 3056.65, 2928.03, 2851.32, 1681.75, 1627.14; MS *m/z* 689 (M+H₂O). [α]_D²⁰ = +34.7 (c 1, CH₃OH); Chiral HPLC: Chiral PACK-AS-H; *n*-Hexane/IPA/TFA (80:20:0.1%); %ee: 99.78; Elemental

analysis; observed (theoretical): C = 72.97 (73.41); H = 5.64 (5.71); N = 4.15 (4.18).

4.8. Preparation of Boc-L-Tyr- ASUD ester

Boc-L-tyrosine 2.0 g (0.0071 mol), OH-ASUD 1.4 g (0.0071 mol), DCC 1.46 g (0.0071 mol) and THF 25 ml were stirred in a 100 ml three neck round bottomed flask at room temperature for 12 h. The dicyclohexylurea was filtered and the filtrate concentrated to give Boc-L-tyrosine-ASUD ester. Yield 3.2 g (100%).

4.9. Preparation of Boc-L-Tyr-Gly-OEt 15

Glycine ethyl ester 0.71 g (0.0069 mol) and Na₂CO₃ 2.2 g (0.01548 mol) were dissolved in 25 ml of water at room temperature in a 100 ml three neck round bottomed flask. Boc-L-tyrosine-ASUD ester 3.2 g (0.00516 mol) dissolved in 25 ml of THF was added to the above reaction mixture at room temperature over a period of 5 min. After the addition was complete, the reaction mixture was stirred for 10 h at room temperature. The reaction mixture was acidified with 5% KHSO₄ to reach a pH of 2–3. The product was extracted into ethyl acetate. The ethyl acetate extract was washed with 5% Na₂CO₃ solution, water, brine solution and dried over anhydrous Na₂SO₄. The ethyl acetate was removed at reduced pressure. The oily crude product obtained was crystallized using ethyl acetate and *n*-hexane. The yield of off-white crystalline Boc-L-Tyr-Gly-OEt is 1.9 g (75%). Mp: 122–125 °C, ¹H NMR (300 MHz, CDCl₃-d): δ 7.04–7.06 (dd, 2H), δ 6.74–6.77 (dd, 2H), δ 6.51–6.54 (t, 1H), δ 5.1 (m, 1H), δ 4.4 (d, 1H), δ 4.17–4.24 (q, 2H), δ 3.95–4.02 (d, 2H), 3.00–3.02 (d, 2H), δ 1.38–1.46 (s, 9H), δ 1.26–1.28 (t, 3H); ¹³C NMR (75 MHz, CDCl₃-d): δ 172.28, 169.62, 157.40, 155.69, 155.53, 130.32, 127.48, 115.64, 80.44, 77.5, 77.0, 76.65, 61.60, 55.85, 49.18, 41.35, 37.61, 33.82, 28.24, 25.53, 24.56, 14.061; IR (cm⁻¹): 3335.50, 2982.98, 2928.95, 1743.08, 1690.05, 1638.92, 1595.19; MS *m/z* 365.13 (M–1). [α]_D²⁰ = +3.3 (c 1, CH₃OH); Chiral HPLC: Chiral PACK-AD-H; *n*-Heptane/IPA/TFA (90:10:0.1%); %ee: 99.96; Elemental analysis; observed (theoretical): C = 58.82 (59.0); H = 7.11 (7.15); N = 7.60 (7.65).

Acknowledgements

Shaik Nowshuddin would like to thank Dr. Murali K. Divi, Chairman and Managing Director, Divis Laboratories Limited, Dr. P. Gundu Rao and Dr. M. N. A. Rao for their encouragement.

References

- Lippert, J. W., III *Arkivoc* **2005**, 87–95.
- Savrdá, J.; Veyrat, D. H. A. *Tetrahedron Lett.* **1968**, 60, 6253–6254.
- Iorga, B.; Campagne, J. M. *Synlett* **2004**, 1826–1828.
- Venkataraman, K.; Wagle, D. R. *Tetrahedron Lett.* **1979**, 32, 3037–3040.
- Ogura, H.; Kobayashi, T.; Shimizu, K.; Kawabe, K.; Takeda, K. *Tetrahedron Lett.* **1979**, 49, 4745–4746.
- Sureshbabu, V. V.; Venkataramanarao, R. *Indian J. Chem., Sect. B* **2008**, 47, 910–919.
- Mizuno, M.; Goto, K.; Miura, T.; Matsuura, T.; Inazu, T. *Tetrahedron Lett.* **2004**, 45, 3425–3428.
- Montalbetti, C. A. G. N.; Falque, V. *Tetrahedron* **2005**, 61, 10827–10852.
- Davey, J. M.; Laird, A. H.; Morley, J. S. *J. Chem. Soc. (C)* **1966**, 555–566.
- Sameiro, M.; Goncalves, T.; Maia, H. L. S. *Org. Biomol. Chem.* **2003**, 1, 1480–1485.
- Gross, H.; Bilk, L. *Tetrahedron* **1968**, 24, 6935–6939.
- Miyazawa, T.; Otomatsu, T.; Fukui, Y.; Yamada, T.; Kuwata, S. *J. Chem. Soc., Chem. Commun.* **1988**, 419–420.
- Hayashi, I.; Shimizu, K. *Bull. Chem. Soc. Jpn.* **1983**, 56, 3197–3198.
- Rebek, J.; Feitler, D. J. *Am. Chem. Soc.* **1973**, 95, 4052–4053.
- Katritzky, A. R.; Todadze, E.; Angrish, P.; Draghici, B. *J. Org. Chem.* **2007**, 72, 5794–5801.
- Sureshbabu, V. V.; Narendra, N.; Kantharaju *Indian J. Chem., Sect. B* **2008**, 47, 920–926.
- Warner–Lambert pharmaceutical company, GB Patent. 898,692, 1962.
- (a) Anderson, G. W.; Zimmerman, J. E.; Callahan, F. M. *J. Am. Chem. Soc.* **1964**, 1839–1842; (b) Miyazawa, T.; Otomatsu, T.; Fukui, Y.; Yamada, T.; Kuwata, S. *Int. J. Pept. Protein Res.* **1992**, 39, 308–314; (c) Nowshuddin, S.; Rao, M. N. A.; Ram Reddy, A. *Synth. Commun.* **2009**, 39, 2022–2031.
- HPLC method: column: Purosphere Star RP-18e, 55 × 4.0 mm, injection volume, 20 µl, flow rate: 0.05 ml/min (gradient).
- Paul, R.; Anderson, G. W. *J. Am. Chem. Soc.* **1960**, 82, 4596–4600.
- (a) Kiso, Y.; Fujiwara, Y.; Kimura, T.; Nishitani, A.; Akaji, K. *Int. J. Pept. Protein Res.* **1992**, 40, 308–314; (b) Hosahudya, N. G.; Suresh Babu, V. V. *Tetrahedron Lett.* **1998**, 39, 9769–9772.
- Anantharamaiah, G. M.; Sivanandaiah, K. M. *Indian J. Chem., Sect. B* **1978**, 16, 797–802.
- (a) Terashima, S.; Wagatsuma, M.; Yamada, S. *Tetrahedron Lett.* **1973**, 29, 1487–1496; (b) Smith, E. L. *J. Biol. Chem.* **1948**, 39–47; (c) Fukui, H.; Kanehisa, H.; Ishibashi, N.; Miyake, I.; Okai, H. *Bull. Chem. Soc. Jpn.* **1983**, 56, 766–769; (d) Casagronde; cesare US Pat., 5013 753, 1991.; (e) Penke, B.; Pallai, P.; Kovacs, K.; Balaspiri, L. *Pept. Proc. Eur. Pept. Symp.* **1976**, 14, 101–107.