ORIGINAL ARTICLE

Synthesis of proline-derived dipeptides and their catalytic enantioselective direct aldol reactions: catalyst, solvent, additive and temperature effects

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Received: 7 February 2009/Accepted: 31 March 2009/Published online: 17 April 2009 © Springer-Verlag 2009

Abstract A series of dipeptides of L-proline-L-amino acid and L-proline-D-amino acid were synthesized to evaluate the catalytic effect for asymmetric direct aldol reactions. In the direct aldol reaction, a catalyst of L-proline-L-amino acid achieves better enantioselectivity than the corresponding L-proline-D-amino acid catalyst. Solubility of the dipeptide catalysts in the solvents is a key point for achieving a better yield of the direct aldol reaction, while hydrogen bonding of solvent does not play an important role in attaining better enantioselectivity and yield. Yield and enantioselectivity of the direct aldol reaction in water were improved by NMM and SDS additives, but the results that were done in plain DMSO were even better.

Keywords Dipeptide · Direct aldol reaction · Enantioselective reaction

Introduction

Aldolases efficiently catalyze aldol reactions of nonactivated unprotected polyfunctionalized substrates under mild ambient conditions to form a new carbon–carbon bond in a stereospecific fashion. Last decade, Barbas III, List et al. started to recreate natural aldolase enzymes with synthetic catalysts and found that L-proline and its analogs can work as chiral catalysts in direct aldol reactions (Notz et al. 2004; List 2004). Later on, the concept of small organic molecules as chiral catalysts has received great attention (Noziere and Cordova 2008; Casas et al. 2005; Ibrahem et al. 2006; Pizzarello and Weber 2004; Northrup and MacMillan 2002;

Y.-H. Chen · P.-H. Sung · K. Sung (⊠) Department of Chemistry, National Cheng Kung University, Tainan, Taiwan, ROC e-mail: kssung@mail.ncku.edu.tw Majewski et al. 2006) because metal-free chiral organic catalysts are in high demand in the pharmaceutical industry.

Since aldolases are globular proteins, organic chemists increased the peptide length of the catalyst from a single amino acid to a dipeptide, tripeptide and oligopeptide to see if a catalyst with a longer peptide chain works better (Tang et al. 2004; Lei et al. 2007; Luppi et al. 2005; Zhao and Dodda 2007; Zheng et al. 2006; Shi et al. 2004; Tsogoeva and Jagtap 2004). Recently, some of the dipeptide catalysts have been used to carry out enantioselective direct aldol reaction. Li et al. found that dipeptide 4 catalyzed enantioselective direct aldol reaction of 4-nitrobenzaldehyde with acetone better than dipeptides 1, 2, 3 and 6 in DMSO with NMM base and PGME 5000 surfactant (Shi et al. 2004). Gong et al. reported that dipeptide ester 5 was better than dipeptide esters 8, 9 and 13 for catalyzing enantioselective direct aldol reaction of 4-nitrobenzaldehyde with hydroxyacetone in THF/H₂O (Tang et al. 2004). Tomasini et al. studied enantioselective direct aldol reaction of isatin with acetone catalyzed by dipeptide esters 17, 16, 9, 15, 18 or 19 at -15° C and found that 16 worked the best (Luppi et al. 2005). Fang et al. performed enantioselective direct aldol reaction of cyclohexanone with 4-pyridinecarbaldehyde in water by dipeptides 4, 14, 12, 7, or 20 with NMM base and PEG400 surfactant, and 20 worked the best (Lei et al. 2007). Zhao and Dodda found that dipeptide 11 worked best among proline 22, prolinamide 21, dipeptides 10 and 11, and dipeptide ester 9 in enantioselective direct aldol reaction of ethyl glyoxylate with acetone (Zhao and Dodda 2007).

The dipeptides, which have been used for direct aldol reactions are all L-proline-L-amino acid. Thus, the question arises: will a dipeptide of L-proline-D-amino acid give a better enantioselectivity in direct aldol reactions? In this article, we will address this question.















Materials and methods

Chemicals

N-Cbz-L-proline was prepared according to the literature (Tang et al. 2003). THF was dried with Na/benzophenone and used fresh.

General method for preparation of 24a-e

To a solution of 23 (498 mg, 2 mmol) in triethylamine (0.3 mL, 2 mmol) and 4 mL of dry THF, ethyl chloroformate (0.2 mL, 2 mmol) was added dropwise over a period of 3 min at 0°C, followed by stirring for 3 h. The reaction mixture was added dropwise to the solution of glycine, D-valine, L-valine, D-phenylalanine or L-phenylalanine (2 mmol) in 4 mL of NaHCO₃ (168 mg, 2 mmol) aqueous solution at 0°C, and then the mixture was stirred for 18 h at room temperature. The final reaction mixture was acidified with dilute HCl aqueous solution, followed by extraction with ethyl acetate. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated by rotary evaporator to get the crude product. The crude product was purified by column chromatography with mobile phase of ethyl acetate and hexane in 1:1 ratio to get the pure product.

24a: ¹H NMR (CDCl₃, 300 MHz): δ 7.19–7.44 (m, 5H, PhH), 5.26 (d, J = 12.3 Hz, 1H, CH), 5.18 (d, J = 12.3 Hz, 1H, CH), 4.45–4.55 (m, 1H, CH), 3.97–4.12 (m, 2H, CH₂), 3.58–3.67 (m, 2H, CH₂), 1.96–2.26 (m, 4H, CH₂CH₂); ¹³C NMR (CDCl₃, 75 MHz): δ 172.31, 171.94, 155.93, 155.60, 135.83, 128.31, 128.00, 127.76, 67.46, 60.67, 60.20, 47.30, 46.93, 41.05, 30.88, 28.68, 24.23, 23.31; IR (CHCl₃): 3,323 (OH, br), 1,682 (C=O) cm⁻¹.

24b: ¹H NMR (CDCl₃, 300 MHz): δ 7.26–7.30 (m, 5H, PhH), 5.12 (s, 2H, CH₂), 4.47–4.53 (m, 1H, CH), 4.38–4.47 (m, 1H, CH), 3.46–3.56 (m, 2H, CH₂), 1.85–2.12 (m, 5H, CH and CH₂CH₂), 0.81–0.90 (m, 6H, 2CH₃); ¹³C NMR (CDCl₃, 75 MHz): δ 174.01, 172.31, 155.65, 135.99, 128.29, 127.96, 127.30, 67.33, 60.78, 60.39, 56.66, 47.33, 46.91, 30.94, 30.72, 29.39, 24.08, 23.22, 18.75, 17.15; IR (CHCl₃): 3,331(OH, br), 1,681(C=O) cm⁻¹.

24c: ¹H NMR (CDCl₃, 300 MHz): δ 7.26–7.31 (m, 5H, PhH), 5.05–5.18 (m, 2H, CH₂), 4.46–4.50 (m, 1H, CH), 4.34–4.45 (m, 1H, CH), 3.44–3.53 (m, 2H, CH₂), 1.88–2.16 (m, 5H, CH and CH₂CH₂), 0.85–0.89 (m, 6H, 2CH₃); ¹³C NMR (CDCl₃, 75 MHz): δ 174.33, 172.48, 172.05, 155.93, 136.02, 128.26, 127.86, 127.67, 67.32, 60.54, 60.18, 57.02, 47.21, 30.78, 28.28, 24.32, 23.33, 18.78, 17.27; IR (CHCl₃): 3,321(OH, br), 1,681(C=O) cm⁻¹.

24d: ¹H NMR (CDCl₃, 300 MHz): δ 6.89–7.32 (m, 10H, PhH), 4.94–5.17 (m, 2H, CH₂), 4.80–4.94 (m, 1H, CH), 4.31–4.37 (m, 1H, CH), 3.4–3.64 (m, 2H, CH₂), 2.95–3.2 (m, 2H, CH₂), 1.79–2.06 (m, 4H, CH₂CH₂); ¹³C NMR (CDCl₃, 75 MHz): δ 173.58, 172.02, 155.51, 135.81, 129.10, 128.33, 128.26, 127.99, 127.74, 126.79, 67.36, 60.66, 60.22, 52.63, 47.02, 46.75, 37.17, 30.87, 29.40, 24.04, 23.08; IR (CHCl₃): 3,315(OH, br), 1,686(C=O) cm⁻¹.

24e: ¹H NMR (CDCl₃, 300 MHz): δ 7.12–7.35 (m, 10H, PhH), 4.90–5.18 (m, 2H, CH₂), 4.85 (m, 1H, CH), 4.30–4.36 (m, 1H, CH), 3.20–3.40 (m, 2H, CH₂), 3.00 (m, 2H, CH₂), 1.6–2.18 (m, 4H, CH₂CH₂); ¹³C NMR (CDCl₃, 75 MHz): δ 173.64, 171.98, 171.61, 155.94, 155.31, 135.86, 135.75, 129.10, 128.32, 127.95, 127.74, 126.80, 67.37, 67.33, 60.23, 52.68, 47.13, 46.80, 37.15, 30.58, 29.46, 28.10, 24.04, 23.03; IR (CHCl₃): 3,317(OH, br), 1,682(C=O) cm⁻¹.

General method for preparation of 25, 26, 10, 27 or 4

To each of the solution of **24a–e** (200 mg) in 2 mL of methanol, 10% Pd/C (20 mg) was added, followed by stirring for 3 h under a hydrogen balloon. The reaction mixture was filtered and washed with methanol, and the filtrate was concentrated by a rotary evaporator. The crude product was passed through a short column of silica with water to remove residual metal, then concentrated and dried to get the pure product.

25: $[\alpha]_D^{24^\circ C} = -23.8$ (c = 0.44 g/100 mL, H₂O); ¹H NMR (D₂O, 300 MHz): δ 4.45 (t, J = 8.3 Hz, 1H, CH), 3.86 (q, J = 17.3 Hz, 2H, CH₂), 3.40–3.50 (m, 2H, CH₂), 2.45–2.53 (m, 1H,), 2.06–2.18 (m, 4H, CH₂); ¹³C NMR (D₂O, 75 MHz): δ 175.62, 169.25, 59.63, 46.16, 42.97, 29.22, 23.55.

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26: $[\alpha]_{D}^{24^{\circ}C} = -27.9$ (c = 0.34 g/100 mL, H₂O); ¹H NMR (D₂O, 300 MHz): δ 4.47 (t, J = 6.7 Hz, 1H, CH), 4.17 (d, J = 4.3 Hz, 1H, CH), 3.40–3.49 (m, 2H, CH₂), 2.4–2.6 (m, 1H, CH), 2.09–2.24 (m, 4H, CH₂), 0.94 (t, J = 7.3 Hz, 6H, CH₃); ¹³C NMR (D₂O, 75 MHz): δ 177.55, 169.21, 60.52, 59.62, 46.28, 29.96, 29.95, 23.55, 18.55, 16.76.

10: $[\alpha]_D^{24^\circ C} = -30.9$ (c = 0.34 g/100 mL, H₂O); ¹H NMR (D₂O, 300 MHz): δ 4.44 (t, J = 8.4 Hz, 1H, CH), 4.05 (d, J = 5.9 Hz, 1H, CH), 3.39–3.49 (m, 2H, CH₂), 2.4–2.6 (m, 1H, CH), 2.05–2.17 (m, 4H, CH₂), 0.94 (t, J = 7.1 Hz, 6H, CH₃); ¹³C NMR (D₂O, 75 MHz): δ 177.88, 168.77, 61.40, 59.45,46.24, 29.79, 29.42, 23.47,18.51, 17.15.

27: $[\alpha]_D^{24^\circ C} = -40$ (c = 0.25 g/100 mL aq. HCl); ¹H NMR (D₂O, 300 MHz): δ 7.29–7.41 (m, 5H, PhH), 4.63 (dd, J = 4.2, 10.1 Hz, 1H, CH), 4.30 (t, J = 7.6 Hz, 1H, CH), 3.28–3.38 (m, 3H, CH and CH₂), 2.90 (t, J = 10 Hz, 1H, CH), 2.18–2.25 (m, 1H, CH), 1.89–1.96 (m, 1H, CH), 1.72–1.79 (m, 1H, CH), 1.50–1.57 (m, 1H, CH); ¹³C NMR (D₂O, 75 MHz): δ 177.02, 168.53, 137.34, 128.93, 128.37, 126.62, 59.44, 55.70, 46.08, 37.48, 29.64, 23.28.

4: $[\alpha]_D^{24^{\circ}C} = -15.4$ (c = 0.13 g/100 mL aq. HCl); ¹H NMR (D₂O, 300 MHz): δ 7.31–7.40 (m, 10H, PhH), 4.60 (dd, J = 4.5, 10.3 Hz, 1H, CH), 4.49 (dd, J = 5.1, 9.2 Hz, 1H, CH), 4.25–4.34 (m, 2H, 2CH), 3.22–3.44 (m, 6H, CH₂), 2.90–3.24 (m, 2H, CH₂), 1.5–2.5 (m, 8H, CH₂); ¹³C NMR (D₂O, 75 MHz): δ 177.44, 177.17, 168.46, 168.33, 137.47, 128.91, 128.37, 128.34, 126.60, 126.56, 59.44, 59.39, 56.70, 56.01, 46.17, 46.07, 37.58, 37.04, 29.64, 29.24, 23.44, 23.27.

General method for direct aldol reactions

To the solution of catalyst **25**, **26**, **10**, **27** or **4** (0.09 mmol) in 0.8 mL of solvent and 0.2 mL of acetone or cyclohexanone, *p*-nitrobenzaldehyde (45 mg, 0.3 mmol) was added, followed by stirring at room temperature or 0°C for some time. Then the reaction mixture was quenched with 20 mL of ammonium chloride aqueous solution, and the mixture was extracted with ethyl acetate. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated by a rotary evaporator to get the aldol product.

4-Hydroxy-4-(4-nitrophenyl)butan-2-one

¹H NMR (CDCl₃, 300 MHz): δ 8.21 (d, J = 8.7 Hz, 2H, PhH), 7.54 (d, J = 8.7 Hz, 2H, PhH), 5.23–5.28 (m, 1H, CH), 3.57 (d, J = 3.4 Hz, 1H, OH), 2.82–2.90 (m, 2H, CH₂), 2.22 (s, 3H, CH₃). The ee value was determined by HPLC analysis of the acetylated product, which was carried out by reaction with triethylamine and acetyl chloride

at room temperature. The HPLC conditions and analytic data are: chiral column, chiralcel OD-RH; mobile phase, water/acetonitrile = 7:3; flow rate, 0.4 mL/min; UV wavelength of detector, 271 nm; retention time for *R*-isomer (major) and *S*-isomer (minor), 37, 40.8 min.

(2S)-2-[(R)-Hydroxy(4-nitrophenyl)methyl]cyclohexanone (anti-isomer)

¹H NMR (CDCl₃, 300 MHz): δ 8.21 (d, J = 8.6 Hz, 2H, PhH), 7.50 (d, J = 8.6 Hz, 2H, PhH), 4.89 (dd, J = 3.0, 8.4 Hz, 1H, CH), 4.06 (d, J = 3.0 Hz, 1H, OH), 1.3–2.6 (m, 9H, CH & CH₂). The ee value was determined by GC analysis of the silylated product, which was carried out by reaction with triethylamine and chlorotrimethylsilane at room temperature. The GC conditions and analytic data are: chiral column, CP-Chirasil-Dex CB; temperature, 170°C; retention time for (*R*,*S*)-isomer (minor), (*S*,*R*)-isomer (major): 35.1, 36.2 min.

(2*R*)-2-[(*R*)-Hydroxy(4-nitrophenyl)methyl]cyclohexanone (syn-isomer)

¹H NMR (CDCl₃, 300 MHz): δ 8.21 (d, J = 8.7 Hz, 2H, PhH), 7.49 (d, J = 8.7 Hz, 2H, PhH), 5.48 (s, 1H, CH), 3.16 (d, J = 3.2 Hz, 1H, OH), 1.3–2.6 (m, 9H, CH & CH₂). The ee value was determined by GC analysis of the silylated product, which was carried out by reaction with triethylamine and chlorotrimethylsilane at room temperature. The GC conditions and analytic data are: chiral column, CP-Chirasil-Dex CB; temperature, 170°C; retention time for (*S*,*S*)-isomer (minor), (*R*,*R*)-isomer (major): 39.2, 41.4 min.

Results and discussion

Because L-proline gives good enantioselectivity in direct aldol reactions (Notz et al. 2004; List 2004), we synthesized dipeptides from L-proline, and the second amino acids in the dipeptides were glycine, L-valine, D-valine, L-phenylanaline or D-phenylanaline, respectively. As shown in Scheme 1 for the preparation of the dipeptides, we protected the secondary amine group of L-proline (**22**) with a carbobenzyloxy (cbz) group by reaction with benzyl chloroformate in aqueous NaHCO₃. Then the carboxylic acid group of the protected L-proline was activated by reaction with ethyl chloroformate in aqueous NaHCO₃, followed by treatment with the second amino acid. Afterwards, hydrogenation of the protected dipeptides on Pd/C eliminated the cbz protecting group from the dipeptides.

Direct aldol reaction of acetone with p-nitrobenzaldehyde in the presence of 30 mol% L-Pro-Gly 25 in DMSO



was optimized by reaction time and temperature. As shown in Table 1, a better enantioselectivity was achieved at 0°C, and a better yield was attained at a longer reaction time (19 h) and 0°C at the expense of enantioselectivity. Thus, the optimized condition of 0°C and 19 h was applied to the direct aldol reaction catalyzed by 26, 10, 27 or 4. As shown in Table 2, L-Pro-L-Phe 4 accomplished the best enantioselectivity and yield in the direct aldol reaction. In addition, it was found that L-Pro-L-Phe 4 achieved better enantioselectivity than L-Pro-D-Phe 27, and L-Pro-L-Val 10 attained better enantioselectivity than L-Pro-D-Val 26. According to these two examples, the L-proline-L-amino acids achieve better enantioselectivity than the L-proline-Damino acid. A strange result appears in Table 2: a poor vield of the direct aldol reaction was caused by L-Pro-L-Val **10**. A possible explanation for this repeatable result is poor solubility of 10 in DMSO.



Because L-Pro-L-Phe **4** worked the best as a catalyst in this direct aldol reaction, the direct aldol reaction catalyzed by 30 mol% of **4** at 0°C for 19 h was used for solvent optimization. Solvents with various degrees of hydrogen bonding were tested. As shown in Table 3, methanol worked the best among alcohols, and alcohols of higher hydrophobicity reduced yields of the direct aldol reaction.

Table 1 Yields and ee values for the direct aldol reaction of acetonewith p-nitro-benzaldehyde in the presence of 30 mol% of L-Pro-Gly**25** in DMSO

Catalyst	Reaction temperature (°C)	Reaction time (h)	Yield (ee value)
L-Pro-Gly 25	25	2	53% (59%)
L-Pro-Gly 25	25	5	80% (65%)
L-Pro-Gly 25	0	5	25% (80%)
L-Pro-Gly 25	0	19	88% (73%)

Table 2 Yields and ee values for the direct aldol reaction of acetone with *p*-nitrobenzaldehyde in the presence of 30 mol% of dipeptide in DMSO at 0°C for 19 h

Catalyst	Vield	ee value (%)	
Catalyst	(%)		
L-Pro-D-Val 26	89	60	
L-Pro-L-Val 10	32	71	
L-Pro-D-Phe 27	95	58	
L-Pro-L-Phe 4	98	82	

Highly hydrophilic water with hydrogen bonding capability, however, made both yield and enantioselectivity of the direct aldol reaction even worse, and so did water with saturated NaCl. Besides, mixed alcohols or aqueous alcohols attained intermediate yield and enantioselectivity of the direct aldol reaction. Conversely, dipolar aprotic DMSO, which is not a hydrogen bonding donor, achieved much better yield and enantioselectivity of the direct aldol reaction. According to the preceding results of solvent

Table 3 Solvent optimization for the direct aldol reaction of acetone with *p*-nitrobenzaldehyde in the presence of 30 mol% of L-Pro-L-Phe **4** in DMSO at 0° C for 19 h

Solvent	Yield	ee value
	()	
DMSO	98	82%
MeOH	86	78%
EtOH	22	83%
i-PrOH	2	n.d.
Water	12	38%
Sat. NaCl _(aq)	11	46%
1:1 MeOH/water	23	48%
1:1 MeOH/ <i>i</i> -PrOH	30	81%

Table 4 Influence of a tertiary amine or surfactant on the direct aldol reaction of acetone with *p*-nitrobenzaldehyde in the presence of 30 mol% of L-Pro-L-Phe **4** in water at 0°C for 19 h

Tertiary amine	Surfactant	Yield (%)	ee value
100 mol% NEt3	_	88	23%
_	28 mol% PEG400	0.8	n.d.
100 mol% NEt3	28 mol% PEG400	85	26%
_	30 mol% SDS	4.1	n.d.
100 mol% NEt3	30 mol% SDS	95	22%
100 mol% NEt3	40 mol% PEG400	56	46%
35 mol% NMM	40 mol% PEG400	46	68%

optimization, hydrogen bonding does not play an important role in achieving better enantioselectivity and yield of the direct aldol reaction, but solubility of the dipeptide catalysts in the solvents is a key point of enantioselectivity and yield of the direct aldol reaction.

Because lower yield of the direct aldol reaction probably results from poor solubility of **4** in solvents, a tertiary amine or surfactant was added to the reaction system in order to improve solubility of **4**. As shown in Table 4, a surfactant of PEG400 or SDS did not improve the yield of the direct aldol reaction, but a tertiary amine



Conclusion

The L-proline-L-amino acid achieves better enantioselectivity of the direct aldol reaction than the corresponding L-proline-D-amino acid. Hydrogen bonding of solvent does not play an important role in achieving better enantioselectivity and yield of the direct aldol reaction, while solubility of the dipeptide catalysts in the solvents is a key point in the yield of the direct aldol reaction. Yield and enantioselectivity of the direct aldol reaction are much better in plain DMSO than those in other solvents even with addition of NMM and SDS into aqueous solutions.

Table 5 Influence of a solvent,
tertiary amine or surfactant on
the direct aldol reaction of
cyclohexanone with p-
nitrobenzaldehyde in the
presence of 30 mol% of L-Pro-
L-Phe 4 at 25°C for 19 h

- ^a Cyclohexanone:DMSO = 1:4
- ^b 4 Å molecular sieve
- ^c Cyclohexanone:DMSO = 4:1

Solvent	Tertiary amine	Surfactant	Yield (anti/syn)	ee (%) for anti/syn
DMSO ^a	_	_	97% (75/25)	89/14
DMSO ^b	-	-	96% (72/28)	91/13
DMSO ^c	-	-	82% (80/20)	89/17
Water	-	-	Trace	n.d.
Water	30 mol% NMM	-	17% (92/8)	41/10
Water	30 mol% NMM	36 mol% PEG400	12% (91/9)	40/12
Water	30 mol% NMM	30 mol% SDS	41% (89/11)	72/42

did dramatically increase the yield of the direct aldol reaction even though its enantioselectivity was limitedly improved.

As shown in Table 5, enantioselectivity dramatically increased when cyclohexanone replaced acetone in the direct aldol reaction in the presence of 30 mol% of L-Pro-L-Phe 4 in DMSO at 25°C for 19 h. Replacement of some DMSO with cyclohexanone in the direct aldol reaction resulted in a decreased yield. When this direct aldol reaction was run in water, conversion was very low. Addition of 30 mol% of NMM and SDS into the reaction system in water did increase both yield and enantioselectivity of the direct aldol reaction, but its result is still worse than that carried out in plain DMSO. Acknowledgment This work was supported by NSC of Taiwan, ROC (NSC 96-2113-M-006-003)

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