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A novel berberine-based colorimetric and fluorometric probe for Hg^{2+} detection and its applications in water samples



Shutang Ruan^a, Yan Zhang^a, Suzhen Wu^b, Yu Gao^a, Lijuan Yang^a, Mingxin Li^a, Yiqin Yang^a, Zhonglong Wang^{a,*}, Shifa Wang^{a,*}

^a Co-Innovation Center of Efficient Processing and Utilization of Forest Resources, College of Chemical Engineering, College of Light Industry and Food, Nanjing Forestry University, Nanjing 210037, China

^b School of Basic Medicine, Gannan Medical University, Ganzhou 341000, China

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ABSTRACT

The development of small molecule fluorescent probes for detection of heavy metal ions is highly desirable in fluorescent sensors area, and mercury (Hg^{2+}) is a heavy metal pollutant in environment and food chain processes. Herein, we reported the design and synthesis of a novel berberine-based fluorescent probe **P1**, which can specifically recognize Hg^{2+} in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution by the colorimetric and fluorometric changes. The probe **P1** displayed high selectivity, superior sensitivity and fast response toward Hg^{2+} with a low detection limit and a wide range of pH values. This probe can provide a convenient and effective way for Hg^{2+} detection on test paper strips. Moreover, the probe **P1** can be applied to quantitatively detect Hg^{2+} in real environmental water samples.

1. Introduction

 Hg^{2+} is a high toxic heavy metal pollutant and widely distributes in water and soil [1–3]. Excessive Hg^{2+} in the bodies easily causes some serious diseases, such as kidney damage [4], minamata disease [5], endocrinium and nervous system damage [6], cognitive and motor disorders [7]. It is necessary to control the content of Hg^{2+} in environmental and agricultural products within safe limits [8].

During the past two decades, many chemical analysis methods have been used for detecting toxic heavy metal, such as atomic absorption spectroscopy [9], high resolution ICP-MS [10], spectrophotometry [11] and fluorescent probe technique [12–15]. Among these detection methods, fluorescent probe technique has been widely accepted owing to its on-site detection, real-time analysis, high sensitivity, low cost and fast response. Recently, a number of fluorescent probes have been reported for detecting Hg^{2+} in real samples and living cells. However, some probes were based on "turn-off" mechanism through the spin–orbit coupling effect which cause fluorescence quenching [16–18]. Therefore, the fluorescent probes for detecting Hg^{2+} based on "turn-on" mechanism have attracted more and more attentions [19–22].

Berberine is a kind of typical isoquinoline alkaloids extracted from huanglian (*Rhizoma coptidis*) [23], and its derivatives were widely used

as anti-bacterial agent [24], antiviral agent [25–27], antioxidant and anticancer [28,29]. In addition, it is a valuable researching direction of fluorescence-based theranostics based on berberine for effective diagnosis and therapy in cancer cells [30].

In our previous work [31], a dual-functional fluorescent probe for the detection of both Hg^{2+} and ClO^- has been designed and synthesized from berberine. In order to further expand the application of berberine and detect Hg^{2+} by the naked eye and fluorescence conveniently. Herein, based on the desulfurization reaction of the mercaptal group, a dual-mode berberine-based colorimetric and fluorometric probe was designed and synthesized for the recognization of Hg^{2+} . Meanwhile, the probe **P1** showed "turn-on" fluorescent response toward Hg^{2+} recognition in real water samples. Moreover, from the fluorescence detection experiments, the probe **P1** exhibited fast response, wide pH range, lower detection limit, high selectivity and sensitivity.

2. Experimental

2.1. Instruments and materials

All chemicals and solvents were purchased from commercial sources and used without further purification. $^1\!H$ NMR and $^{13}\!C$ NMR spectra

* Corresponding authors. *E-mail addresses:* wang_zhonglong@njfu.edu.cn (Z. Wang), wangshifa65@163.com (S. Wang).

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Scheme 1. Synthesis of probe P1.

were recorded on a Bruker AV 400 spectrometer in DMSO- d_6 and tetramethylsilane (TMS) as internal standard. High-resolution mass spectra (HRMS) were tested by an America Agilent 5975c mass spectrometer. UV–vis absorption spectra were recorded by a Shimadzu UV-2450 at the room temperature. Fluorescence spectra were carried out on fluorescence spectrophotometer (LS 55, Perkin Elmer).

2.2. Synthesis of probe P1

As shown in Scheme 1, berberine was selectively demethylated under vacuum and heating conditions, and then acidized and formylated to get compound 3. The probe P1 was obtained by condensation of compound 3 with 1,3-propanedithiol. The chemical structure of the above compounds and probe P1 was characterized by ¹H NMR and HR-MS (Fig. S1-S7).

2.2.1. Synthesis of compound 2

Berberine chloride (10.0 g, 26.9 mmol) was heated at 180–190 °C for 30–60 min under vacuum to afford dark wine solid, which was washed with MeOH (80 mL) and filtered to afford the red compound. And this crude product was added C₂H₅OH (100 mL) and HCl (3.0 M, 70 mL) to stir for 6 h to obtain yellow material, which was washed with CH₃OH/Et₂O (ν/ν = 3:1) 3 times (3 × 60 mL) to afford compound **2** as a yellow solid, 8.87 g, yield: 92.4%. ¹H NMR (400 MHz, DMSO-*d*₆) δ : 11.31 (s, 1H), 9.97 (s, 1H), 8.88 (s, 1H), 8.12 (d, *J* = 8.9 Hz, 1H), 7.82 (s, 1H), 7.75 (d, *J* = 8.9 Hz, 1H), 7.10 (s, 1H), 6.21 (s, 2H), 4.96 (t, *J* = 6.3 Hz, 2H), 4.09 (s, 3H), 3.25 (t, *J* = 6.3 Hz, 2H);¹³C NMR (100 MHz, DMSO-*d*₆) δ : 149.58, 147.61, 145.70, 145.30, 143.75, 136.52, 132.39, 130.33, 125.43, 120.56, 119.80, 118.03, 117.58, 108.32, 105.29, 101.94, 56.99, 54.89, 26.48;ESI-MS (*m*/*z*): [M–Cl]⁺ : calcd. for C₁₉H₁₆ClNO₄-Cl⁻, 322.1074; found, 322.1074.

2.2.2. Synthesis of compound 3

Compound **3** was obtained according to the reported procedures **[31]**. Compound **2** (1.0 g, 3.1 mmol) and hexamine (1.26 g, 9.0 mmol) were dissolved in trifluoroacetic acid (18 mL), and reacted for 8 h at 85 °C. The reaction system was cooled to room temperature, and 3.0 M HCl (15 mL) was added to react at 85 °C for another 2 h. The reacted mixture was immediately poured into ice water, and extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with distilled water and brine until neutrality, and then evaporated to remove off the solvent. The residue was purified by silica gel chromatography (CH₂Cl₂/CH₃OH, 10:1, *v*/*v*) to give compound **3** as a yellow solid, 0.38 g, yield: 34.6%. ¹H NMR (400 MHz, DMSO-*d*₆) &: 10.19 (s, 1H), 9.28 (s, 1H), 9.08 (s, 1H), 7.72 (s, 1H), 7.56 (s, 1H), 7.02 (s, 1H), 6.15 (s, 2H), 4.60 (t, *J* = 6.3 Hz, 2H), 3.77 (s, 3H), 3.10 (t, *J* = 6.4 Hz, 2H), 1.24 (s,

1H) ;ESI-MS (m/z): $[M-Cl]^+$: calcd. for C₂₀H₁₆ClNO₅-Cl⁻, 350.1023; found, 350.1049.

2.2.3. Synthesis of compound 4

Compound **3** (50 mg, 0.14 mmol) and 1,3-propanedithiol (45.4 mg, 0.42 mmol) were dissolved in CH₂Cl₂ (20 mL), and a catalytic amount of BF₃·Et₂O (0.04 mL, 0.30 mmol) was added, and the mixture was reacted at room temperature for 15 h under N₂ atmosphere. The reacted mixture was evaporated to remove off solvent, and the residue was purified by recrystallization with CH₃OH/Et₂O ($\nu/\nu = 2:1$) to obtain compound **4** as a red solid, 37.5 mg, yield: 57.3%. ¹H NMR (400 MHz, DMSO- d_6) & 9.31 (s, 1H), 8.37 (s, 1H), 7.89 (s, 1H), 7.51 (s, 1H), 7.03 (s, 1H), 6.30 (s, 1H), 6.15 (s, 2H), 4.60 (t, J = 6.2 Hz, 2H), 3.81 (s, 3H), 3.30 (s, 2H), 3.10 (t, J = 6.2 Hz, 2H), 2.90 (dt, J = 14.1, 3.5 Hz, 2H), 1.24 (d, J = 3.3 Hz, 2H) ; ESI-MS (m/z): [M–Cl]⁺ calcd. for C₂₀H₁₆ClNO₅-Cl⁻, 440.0985; found, 440.1019.

2.3. Spectroscopic measurements of probe P1

The absorption and fluorescence detection experiments were done in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution at the room temperature, and the excitation wavelength is 365 nm. The probe **P1** stock solution (2.5 × 10⁻⁴ M) was prepared in CH₃OH. The test solution was prepared by adding 100 µL stock solution to 10 mL CH₃OH/PBS buffer (8:2, ν/ν , pH = 7.4, 10 mM). The solutions of various analytes (5.0 × 10⁻⁴ M) (including Na⁺, K⁺, Ca²⁺, Mg²⁺, Co²⁺, Al³⁺, Ni²⁺, Sn²⁺, Ag²⁺, Zn²⁺, Cu²⁺, Γ , NO₃⁻, AcO⁻, H₂PO₄²⁻, S²⁻) were prepared in deionized water.

2.4. Visualization of Hg^{2+} in solution by test paper

The filter paper was immersed into the CH₃OH solution of probe **P1** (2.5 × 10⁻⁴ M) and dried in air for 30 min. The treated test papers were immersed in the Hg²⁺ solution with different concentrations (0 μ M-500 μ M) for 30 min at the room temperature. The fluorescence color changes of filter paper were observed under day light and 365 nm UV light.

2.5. Quantitation of Hg^{2+} contamination in real water samples

The real water samples were respectively taken from Zhangjiang river and Poyang lake, and the real water samples were utilized to prepare CH₃OH solutions (80%, 10 mM). Hg²⁺ solutions with different concentrations (0, 50, 100, 150, 200, 250 μ M) were added to the aforesaid two water samples, and the fluorescence intensity of the water samples were measured using a fluorescence spectrophotometer.



Fig. 1. Absorption (a) and fluorescence emission (b) spectra of probe P1 (2.5×10^{-4} M) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution in the presence and absence of Hg²⁺ (5.0×10^{-4} M). $\lambda_{ex} = 365$ nm. Inset: color changes of probe P1 solution after the addition of Hg²⁺ under daylight and 365 nm UV-light.



Fig. 2. (a) Fluorescence intensity of **P1** (2.5×10^{-4} M) added to Hg²⁺ (5.0×10^{-4} M) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution from pH = 2.0 to pH = 12.0 at 550 nm. Inset: color changes of **P1** after the addition of Hg²⁺ from pH = 2.0 to pH = 12.0 under 365 nm UV-light. (b) Time-dependent fluorescence spectra of **P1** (2.5×10^{-4} M) presence and absence Hg²⁺ (5.0×10^{-4} M) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution ($\lambda_{em} = 550$ nm).

3. Results and discuss

3.1. Spectroscopic properties of probe **P1** to Hg^{2+}

The UV-vis absorbance and fluorescence spectroscopic properties of

probe **P1** in the presence and absence of Hg^{2+} at the room temperature were examined. As shown in Fig. 1a, the free probe **P1** showed two absorption peaks at 400 nm and 520 nm. Upon addition of Hg^{2+} , the original maximum absorption peaks were significantly reduced and the new peaks were appeared obviously at 350 nm and 450 nm, and the



Fig. 3. (a) Fluorescence spectra of P1 (2.5×10^{-4} M) in the presence of Hg²⁺ (0–2.0 eqv.) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution. (b) Linear calibration curve between the fluorescence intensity of P1 and Hg²⁺ (0–2.0 eqv.) at 550 nm.



Fig. 4. (a) Fluorescent spectra of P1 (2.5×10^{-4} M) with different competing ions (5.0×10^{-4} M) (including Na⁺, K⁺, Ca²⁺, Mg²⁺, Co²⁺, Al³⁺, Ni²⁺, Sn²⁺, Ag²⁺, Zn²⁺, Cu²⁺, Γ , NO₃⁻, OAc⁻, H₂PO₄²⁻, S²⁻, Hg²⁺) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution (λ_{ex} = 365 nm). (b) The fluorescence intensity of P1 (2.5×10^{-4} M) with Hg²⁺ (5.0×10^{-4} M) in the presence of different competitive anions (5.0×10^{-4} M) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution (λ_{ex} = 365 nm). (c) Photographs of P1 (2.5×10^{-4} M) with different competing ions (5.0×10^{-4} M) in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution under day-light and 365 nm UV light.

colors of the solution changed from light red to pale yellow under daylight.

The fluorescence properties of probe **P1** were investigated. As shown in Fig. 1b, the free probe **P1** displayed two weak fluorescence emissions at 450 nm and 550 nm with excitation at 365 nm. However, upon addition of Hg²⁺, the fluorescence intensity at around 550 nm was significantly enhanced, and the color of the solution changed from pale blue to yellow green. These results showed that the probe **P1** can serve as a colorimetric and fluorometric probe for Hg²⁺ detection.

3.2. Response time and effect of pH values of probe P1 to Hg^{2+}

Influence of pH from 2.0 to 12.0 on the fluorescence responses of probe **P1** with and without Hg^{2+} were investigated as shown in Fig. 2a, It was known from the results of fluorescent experiments that the fluorescence emission intensities were stable in a wide pH range from 3.0 to 10.0, which indicated that the probe **P1** was suitable for detecting Hg^{2+} under the real water samples in a wide pH range. In addition, the time-dependent fluorescence results demonstrated that the probe **P1** could remain stable and recognize Hg^{2+} with a short response time (Fig. 2b).

3.3. Sensitivity of probe **P1** to Hg^{2+}

The fluorescence responses of probe **P1** to Hg^{2+} were investigated by titration experiments in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution, and the results were shown in Fig. 3a. With the increasing of

Hg²⁺ concentration from 0 to 2.0 equiv., the emission intensities of probe **P1** solution enhanced at 550 nm and quenched at 450 nm, which may be attributed to an ICT (intra-molecular charge transfer) process [32]. Furthermore, the emission intensities at 550 nm showed a good linear relationship over the concentration range from 0 μ M to 500 μ M (y = 0.5335x + 184.0493, R² = 0.9950). The detection limit for Hg²⁺ was calculated to be 5.4 μ M based on 3 σ /k method (Fig. 3b).

3.4. Selectivity and competivity of probe P1 to various analytes

The selectivity experiments were carried out with the addition of various analytes. As shown in Fig. 4a, the fluorescence spectral changes of probe P1 (2.5×10^{-4} M) were examined in CH₃OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution upon addition of different various analytes (5.0×10^{-4} M, including Na⁺, K⁺, Ca²⁺, Mg²⁺, Co²⁺, Al³⁺, Ni²⁺, Sn²⁺, Ag²⁺, Zn²⁺, Cu²⁺, Γ , NO₃⁻, AcO⁻, H₂PO₄²⁻, S²⁻). Upon addition of these ions, there were no significant fluorescence intensity changes, while the fluorescence intensity in the presence of Hg²⁺ (5.0×10^{-4} M) obviously increased at 550 nm with the color changes from pale blue to yellow green. Meanwhile, the other competing species could hardly interfere the detection of Hg²⁺ in a complex environment (Fig. 4b). Under the day-light and 365 UV-light, the competing species did not produce any dramatic color changes and "turn-on" fluorescent emission except Hg²⁺ (Fig. 4c). The results confirmed that the probe P1 is a specific "turn-on" fluorescent probe for detecting Hg²⁺.



Scheme 2. Sensing mechanism of P1.



Fig. 5. The optimized geometric structures and corresponding LUMO and HOMO of P1 and P1-CHO by DFT calculation.

3.5. Sensing mechanisms of probe P1 to Hg^{2+}

Based on the fluorescence intensities of probe **P1** in the presence or absence of Hg^{2+} , the detection mechanism was speculated that the desulfurization reaction of probe **P1** was promoted by Hg^{2+} . The molecular ion peaks of probe **P1** before and after adding Hg^{2+} were 440.1019 [Fig. S7, P1-Cl, Exact Mass: 440.0985] and 350.1030 [Fig. S8, P1-CHO-Cl, Exact Mass: 350.1023]. In addition, ¹H NMR titration of probe **P1** with Hg^{2+} can be carried out, and these two methods indicated that detection mechanism of probe **P1** was the same with the postulated shown in Scheme 2 [33–36].

3.6. Density functional theory (DFT) calculations

To further understand the photophysical responses of probe **P1** to Hg^{2+} , DFT calculations were carried out with the Gaussian 09 software package. The optimized geometric structures and corresponding LUMO (the lowest unoccupied molecular orbitals) and HOMO (the highest occupied molecular orbitals) were presented. As shown in Fig. 5, the HOMO of probe **P1** was mainly localized on the hydroxyl group and part of the D ring. However, its LUMO was evenly distributed in the C and D rings, which indicated that there was a weak ICT effect in this molecule. Upon addition of Hg^{2+} , the HOMO of compound **P1-CHO** was localized on the hydroxyl group. However, the LOMO was distributed in the aldehyde group and D rings, indicating a strong ICT effect. In addition,



Fig. 6. Test-paper experiments of P1 with the different concentrations of Hg $^{2+}$ (0 μ M to 500 μ M).

the energy gap between HOMO and LUMO of compound **P1-CHO** was calculated to be 3.14 eV, which is lower than the probe **P1** (3.17 eV). The result also indicated that the sensing mechanism was shown in Scheme 2.

3.7. Visualization of Hg^{2+} in solution by test paper

To investigate the practical application of probe **P1** as an on-situ detection kit for the detection of Hg^{2+} , the test paper experiments were explored. As shown in Fig. 6, as the concentrations of Hg^{2+}

increased from 0 μ M to 500 μ M, the fluorescence emission of test paper strips gradually changed from blue to faint-yellow. Therefore, the probe **P1** could successfully applied to detect Hg²⁺ on test paper.

3.8. Quantitation of Hg^{2+} contamination in real water samples

Probe **P1** could also be used to detect Hg²⁺ in two real water samples collected from Zhangjiang river and Poyang lake. The water samples were mixed with CH₃OH (80%) solutions containing the probe **P1** (2.5 \times 10⁻⁴ M). The various concentrations of Hg²⁺ solution (0, 50, 100, 150, 200, 250 μ M) were added into experimental water samples respectively. The fluorescence intensity of the experimental samples was measured using a fluorescence spectrophotometer. As shown in Fig. 7, two good linear relationships were obtained, and the detection limit for Hg²⁺ was calculated to be 6.3 μ M and 4.4 μ M, indicating that the probe **P1** can be applied to detect Hg²⁺ in real samples.

4. Conclusion

In summary, the probe **P1** for detecting Hg^{2+} was synthesized from berberine. This probe could identify Hg^{2+} in CH_3OH/PBS (8:2, ν/ν , pH = 7.4, 10 mM) solution by the obvious color changes at daylight and the perfect fluorescence changes under 365 nm UV lamp. The probe **P1** displayed high selectivity, superior sensitivity, and fast response toward Hg^{2+} with a low detection limit and a wide pH range. The probe **P1** can provide a convenient and effective way for Hg^{2+} detection by virtue of test paper strips. Moreover, this probe can be also applied to monitor Hg^{2+} in real environmental water samples.



Fig. 7. (a) The linear relationship of fluorescence intensity of P1 and different concentrations of Hg^{2+} in Zhangjiang river ($\lambda_{em} = 550$ nm). (b) The linear relationship of fluorescence intensity of P1 and different concentrations of Hg^{2+} in Poyang lake ($\lambda_{em} = 550$ nm). (c) The fluorescence intensity of P1 (2.5×10^{-4} M) towards different concentration of Hg^{2+} (0, 50, 100, 150, 200, 250 μ M) ($\lambda_{em} = 550$ nm).

CRediT authorship contribution statement

Shutang Ruan: Conceptualization, Data curation, Methodology, Writing – original draft. Yan Zhang: Formal analysis, Resources. Suzhen Wu: Investigation. Yu Gao: Investigation. Lijuan Yang: Resources. Mingxin Li: Software. Yiqin Yang: Supervision. Zhonglong Wang: Writing – review & editing. Shifa Wang: Funding acquisition, Project administration.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary material

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