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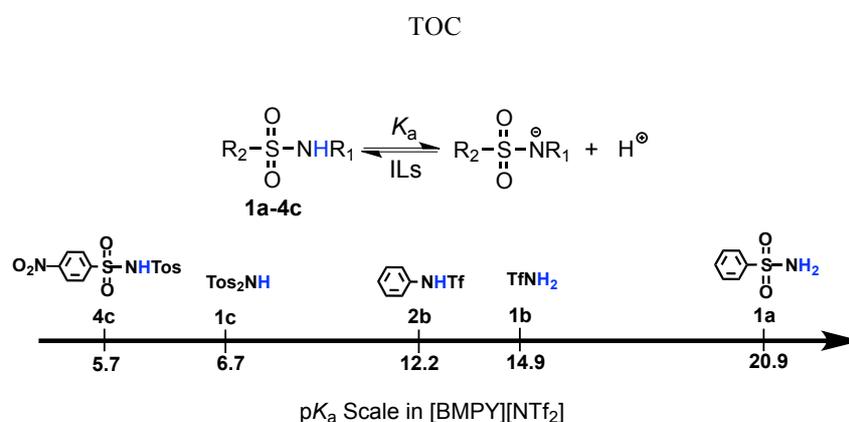
Absolute pK_a s of Sulfonamides in Ionic Liquids: Comparisons to Molecular Solvents

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ABSTRACT

Absolute pK_a s of 25 sulfonamides in four ionic liquids (ILs) were measured spectroscopically with high precision and subsequently compared with these in conventional molecular solvents. It is found that the acidity order of these sulfonamides follows: in water > in DMSO > in ILs > in acetonitrile (ACN). The well-known solvent polarity index ϵ fails to explain the observed stronger bond-weakening effect of ILs than that of ACN whose ϵ is much greater. In addition, the regression analyses show that the pK_a s of sulfonamides determined in ILs linearly correlate with these in molecular solvents of distinct properties, but with various slopes. A rationale and related discussion on the effect of solvation in ILs are presented.

INTRODUCTION

Solvents are ubiquitous, and in most circumstances, are indispensable in chemistry. In a sense, the selection of solvent based on the knowledge of solvation reflects a rational development of chemistry, which, in return, can significantly improve the image of organic reactions regarding the environment

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3 issue.¹ In recent years, as it became more and more crucial, many volatile organic solvents are listed as
4 “unclean”, and various room temperature ionic liquids (RTILs) come into play as their alternatives, in
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7 spite of a severe shortage of the knowledge on their solvation behavior.
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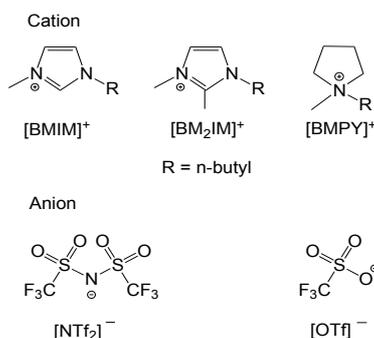
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10 Ionic liquids are composed entirely of ions and exhibit unique properties that are significantly
11 different from these of conventional molecular solvents.² As a rising mainstream medium category, ILs
12 are labelled as green solvent³ which have aroused enormous research interests in recent decade and
13 have also been extensively used in industry.⁴ Compared to the tremendous attentions paid on the
14 development of their synthetic applications, fundamental aspects on the solvation phenomena of ILs
15 are very scarce, however. As it is well known that solvents have strong impacts on the acidity of
16 substrate by virtue of their differential strengths of solvation towards various species in solution, we
17 believe that to study the pK_a s in different media would provide quantitative understandings in this
18 regard.
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28 Some acidity scales in various molecular solvents have been established in past,⁵ which allowed a
29 good understanding of solvation behavior for conventional media. By contrast, acidity studies in neat
30 ILs are far less, and the early works reported *relative* pK_a s in most cases⁶ that are not suitable for the
31 solvation study. Although more recent works showed that the *absolute* acidity in ILs could be obtained
32 electrochemically,⁷ the narrow substrate scope and large uncertainty associated with irreversible
33 electrode data make its use in solvation study problematic. However, we have recently found that the
34 overlapping indicator method, which has been dominantly used in molecular media for most of the
35 authentic pK_a s data known today⁸, can be well adopted to measure the acidities in ILs with high
36 precision⁹ and hence should be able to fulfil the current research need.
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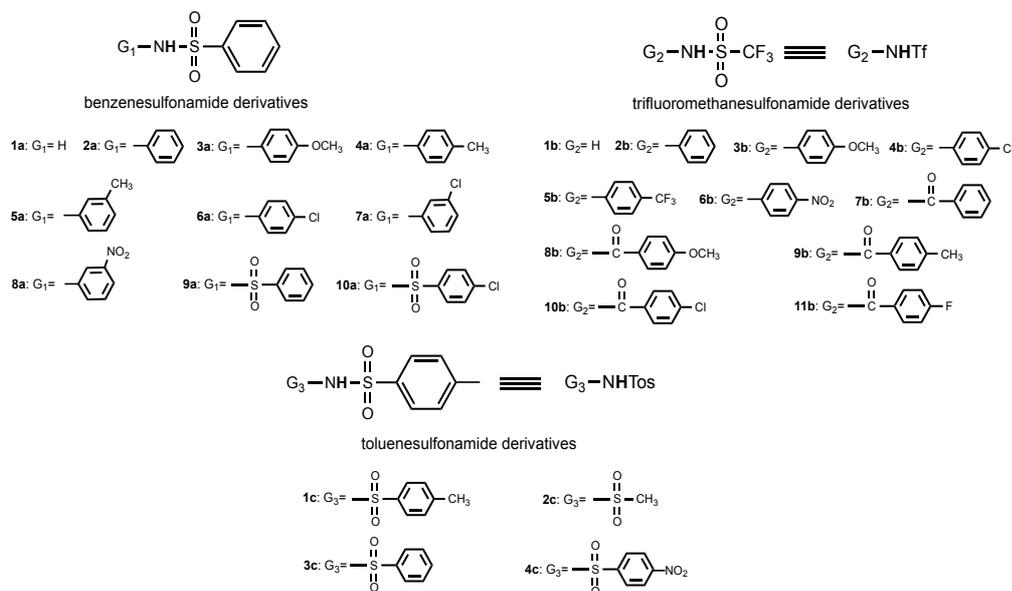
47 Sulfonamides are important compounds in pharmaceutical and agricultural chemicals industry. A
48 large number of drugs, such as antimicrobials and diuretics, and several novel herbicides, bear this
49 sulfonamide moiety. This is mainly due to their unique structure that resembles the tetrahedral
50 intermediate involved in many acyl substitutions and stabilized by proteases and esterases.¹⁰
51 Sulfonamide is known to be more acidic than carboxamides. The knowledge on sulfonamide acidities is
52 important to pharmaceutical industry because it delivers valuable information on the mechanisms of
53 drug actions and metabolisms. The acidities of sulfonamides have been investigated extensively in the
54 conventional molecular solvents.¹¹ However, till present almost no attention has been paid on the
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acidities of N–H bonds in IL. On the other hand, recent seminal study shows that developing IL drugs and use of ILs as a drug delivery medium may bring a revolution to pharmaceutical industry.¹² This, from another angle, points out the need of the information on sulfonamides acidities in ILs, especially on their relations to the solvation strength of different types of media in a more quantitative manner.

Towards this end, here we have measured 68 absolute acidity values of 25 sulfonamides in 4 ILs (Scheme 1) with high precision with the standard deviation (SD) of $\leq \pm 0.05$ pK units. A variety of structurally different sulfonamides (Scheme 2), such as benzenesulfonamides (**1a–8a**), trifluoromethylsulfonamides (**1b–6b**) and several more acidic sulfonamides (**9a–10a**, **7b–11b** and **1c–4c**), were included in this work. The measured pK_a data (Tables 1) are compared with the known acidities in molecular solvents by regression analysis, which revealed insightful and interesting information regarding the change on the solvation strength by ILs and molecular media.



Scheme 1. Cations and anions of the ILs used in this study, the ILs are: [BMIM][OTf], [BMIM][NTf₂], [BMPY][NTf₂] and [BM₂IM][NTf₂]



Scheme 2. The structures of sulfonamide derivatives (**1a-4c**) involved in this work

RESULTS AND DISCUSSION

Table 1 shows that the acidity scale of these N–H acids spreads over 15 p*K* units, which covers by far the largest energetic span and substrates collection for one R–H acid family in ILs, and provides with rich information of the effect of structural change on the N–H acidity in ILs. For examples, an additional sulfonyl group causes a dramatic increase of the acidity of **1a** (20.9) by as large as 14 orders of magnitude compared to that of **9a** (6.3), while a carbonyl group makes the acidity of **7b** (6.7) 1.6×10^8 times stronger than **1b**, due to their strong inductive as well as resonance stabilization effect on the sulfonamide nitrogen anion. This is supported by the fact that the carbon acid, PhSO₂CH₂SO₂Ph (p*K*_a = 19.2 in [BMPY][NTf₂]^{9a}) is much less acidic than its analogue **9a** in ILs, apart from the fact that the electronegativity of N is greater than C atom, the large acidity difference ($\Delta pK_a \approx 13$) may also be as a consequence of a significant p-d π -resonance between the anion center and sulfonyl group of sulfonamides.¹⁴

Table 1. p*K*_a Values of sulfonamides (Scheme 2) in ILs and molecular solvents

Sulfonamides ^a	p <i>K</i> _a ^b	p <i>K</i> _a ^b	p <i>K</i> _a ^b	p <i>K</i> _a ^b	p <i>K</i> _a	p <i>K</i> _a	p <i>K</i> _a
	[Bmpy][NTf ₂]	[Bmim][NTf ₂]	[Bm ₂ im][NTf ₂]	[Bmim][OTf]	water	DMSO	ACN
PhSO ₂ NH ₂ (1a)	20.9	20.2 ₅	21.9	18.1	9.4 ^c	15.2 ^d	24.6 ^d
p-MeO-PhNHSO ₂ Ph (3a)	18.7	18.2	19.3	16.1	8.9 ^e	14.2 ^e	22.9 ^e
p-Me-PhNHSO ₂ Ph (4a)	18.4	17.7 ₅	19.1	15.7	8.6 ^e	13.9 ^e	22.6 ^e
m-Me-PhNHSO ₂ Ph (5a)	18.2	17.6	18.9	15.5	8.5 ^e	13.7 ^e	22.7 ^e
PhNHSO ₂ Ph (2a)	18.0	17.4	18.7	15.4	8.2 ^e	13.5 ^e	22.6 ^e
p-Cl-PhNHSO ₂ Ph (6a)	17.1 ₅	16.7	17.7 ₅	14.6	7.9 ^e	12.7 ^e	21.6 ^e
m-Cl-PhNHSO ₂ Ph (7a)	16.7	16.3	17.4	14.3	7.7 ^f	11.7 ^e	
m-NO ₂ -PhNHSO ₂ Ph (8a)	15.7	15.3	16.2	13.3		11.2 ^e	20.4 ^e
TfNH ₂ (1b)	14.9	14.4 ₅	15.5	12.8	6.3 ^g	9.7 ^h	
PhNHTf (2b)	12.2	11.8	12.7	9.9 ₅	4.5 ^g		
p-MeO-PhNHTf (3b)	12.9	12.4	13.3 ₅	10.6	4.9 ^g		
p-Cl-PhNHTf (4b)	11.4	11.0	11.8	9.2	3.9 ^g		
p-CF ₃ -PhNHTf (5b)	10.6	10.2 ₅	11.0	8.5			
p-NO ₂ -PhNHTf (6b)	9.4 ₅	9.0 ₅	9.8	7.6			
p-MeO-PhCONHTf (8b)	7.2	6.7					11.6 ⁱ
TosNHSO ₂ Me (2c)	7.2	6.6					
p-Me-PhCONHTf (9b)	7.0	6.6					11.5 ⁱ
Tos ₂ NH (1c)	6.7	6.2			1.7 ^k		12.0 ^{l,k}
PhCONHTf (7b)	6.7	6.2 ₅					11.1 ⁱ
PhSO ₂ NHTos (3c)	6.5	6.0					
PhSO ₂ NHSO ₂ Ph (9a)	6.3	5.8 (9.7 ^l)			1.4 ₅ ^f		11.3 ^j
p-F-PhCONHTf (11b)	6.3	5.9					10.7 ⁱ
p-Cl-PhCONHTf (10b)	6.3	5.8					10.4 ⁱ
p-Cl-PhSO ₂ NHSO ₂ Ph (10a)	5.9	5.3					
p-NO ₂ -PhSO ₂ NHTos (4c)	5.7	5.0					10.1 ^j

^aThe acidic hydrogens are indicated by boldface type. ^bIn p*K*_a units, SD: ≤ ±0.05 p*K* unit. ^cReference 11e. ^dReference 11f. ^eReference 11g. ^fReference 11b. ^gReference 11a. ^hReference 13. ⁱReference 11m. ^jReference 11j. ^kReference 11i. ^lReference 7c, in [EMIM][NTf₂], where [EMIM]⁺=1-ethyl-3-methylimidazolium, the value is a simulated one based on electrochemical measurement.

The structure and composition of the ILs in this study have pronounced effect on the N–H acidic dissociation equilibrium. Normally, the anion of IL plays a dominant role on the acidity, due to the relatively large proton solvation energy [$\Delta G_{\text{Solv}}^*(\text{H}^+)$] by anion (-258 kcal mol⁻¹ for [NTf₂]⁻), compared with that by cation on the deprotonated species of various substrates.¹⁵ From Table 1, the acidity of individual sulfonamide decreases in a

sequence of [BMIM][OTf] > [BMIM][NTf₂] > [BMPY][NTf₂] > [BM₂IM][NTf₂], which is consistent with our previous observations for other series of acids.⁹ It is not difficult to understand that the acidity of these sulfonamides are more acidic in [BMIM][OTf] than in [BMIM][NTf₂]. This should be ascribed to a more localized negative charge and smaller size of [OTf]⁻ compared to [NTf₂]⁻, both leading to a stronger solvation of proton. The trend of p*K*_a change in the ILs with [NTf₂]⁻ as the common counter anion could be rationalized on the basis of the extent of charge delocalization and the accessibility of the cation moiety towards the sulfonamide anion. For example, the observed stronger acidity of sulfonamide in [BMIM][NTf₂] than in the other two ILs with [NTf₂]⁻ as a common anion should be due to the less hindered [BMIM]⁺ moiety than the other two and its C₂-H that is capable of providing an H-bonding with the sulfonamide nitranion.

Table 1 also shows that the acidities of sulfonamides in various solvents follow the order, in water > in DMSO > in [BMIM][OTf] > [BMIM][NTf₂] > [BMPY][NTf₂] > [BM₂IM][NTf₂] > in ACN, which actually reflects the order of solvability for these solvents.^{7b} This sequence is beyond one's expectation, however, because AILs are known for their relatively low polarity ($\epsilon = 10\text{--}15$ ¹⁶), which would otherwise suggest a weaker acidity in ILs than in ACN, whose dielectric constant is much greater ($\epsilon = 36.1$ ¹⁷). Nonetheless, as we recently found, the cations and anions in AILs should exist essentially as “free ions” rather than as ion-pairs^{9c} (showing an “ionic liquid effect”^{9c,9e,18,19}), it may not be too big a surprise to see that some classical parameters suitable for describing properties in molecular media (such as ϵ) may not always be so for certain properties in ILs. Thus, the presently observed but yet unanticipated unusually strong N–H bond weakening ability of the low-polarity ILs could be understood by considering that the IL's anion and cation should be able to act separately in stabilizing the proton and the incipient sulfonamide nitranion upon deprotonation by virtue of their respective solvability (via Coulombic, resonance, π - π interactions, etc.) towards ions of opposite charges. This *in situ* situation occurred in a solvation process in IL is not exactly the same as the status of an IL in a dielectric

constant measurement,¹⁶ and thus, it should be the reason behind the unusually strong N–H bond-weakening ability of these superficially weakly-polar ILs.

Next, we carried out a regression study on the sulfonamides acidities measured in the 4 ILs. All the correlations between the pK_a s in any two ILs show excellent linearity with $R^2 = 0.999$ (see SI, Figure S3, one of them is shown in Figure 1). Furthermore, regression analyses between the pK_a s in one representative IL ([BMPY][NTf₂]) and these in molecular solvents were performed. Remarkably, in spite of the tremendous differences in solvent properties of water, DMSO and ACN from ILs, good to excellent linearity is observed in

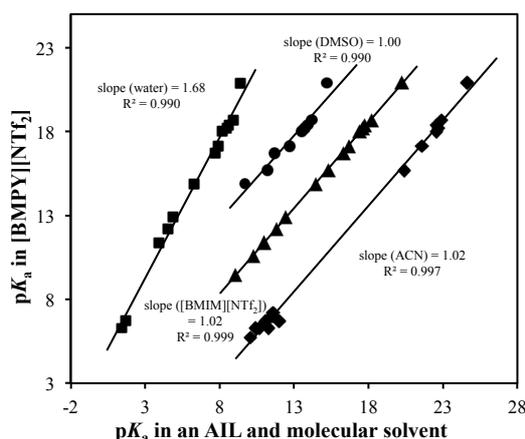


Figure 1. pK_a s of sulfonamides in [BMPY][NTf₂] against these in [BMIM][NTf₂] and molecular solvent; filled square, circle, triangle and diamond represents the pK_a s in water, DMSO, [BMIM][NTf₂] and ACN, respectively.

all the correlations (Figure 1). It is noteworthy that the slope of the correlation between the pK_a s in [BMPY][NTf₂] and in water is significantly greater than unity (slope = 1.68), indicating a much lower sensitivity of sulfonamide acidity towards structure variation in water than in AILs. This is understandable, because the negative charge of the sulfonamide anion can be better dispersed in water than in IL due to the greater solvability of water caused mainly by its strong hydrogen-bondings. This reduces the charge density on nitrogen and weakens the influence of a structural change more significantly than the situation in ILs. On the other hand, the slopes of the correlations between the acidities in ILs (taking [BMPY][NTf₂] as example) vs. DMSO and vs. ACN are all observed to be around unity (Figure 1, slope = 1.00 and 1.02, respectively). Though this seeming coincidence in the slope values cannot be fully understood based on our present knowledge on solvation, these correlation Equations 1-3 (Scheme 3) together with those in ILs mentioned above can provide a practically very useful way for estimating unknown pK_a s in these molecular and ionic solvents for relevant compounds.

$$pK_a([\text{BMPY}][\text{NTf}_2]) = 1.68 pK_a(\text{water}) + 4.24 \quad \text{Eqn. 1}$$

$$pK_a([\text{BMPY}][\text{NTf}_2]) = pK_a(\text{DMSO}) + 4.76 \quad \text{Eqn. 2}$$

$$pK_a(\text{ACN}) \approx pK_a([\text{BMPY}][\text{NTf}_2]) + 4.81 \quad \text{Eqn. 3}$$

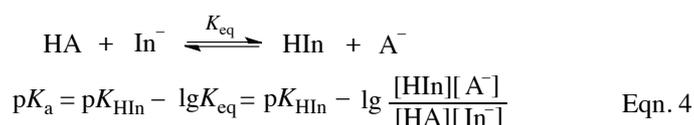
Scheme 3. The acidity correlation equations between pK_a s in IL and molecular media

In summary, the absolute acidities of 25 N–H acids (altogether 68 pK_a values) were accurately measured in four ILs. The wide energetic range (>15 pK units) of the obtained data enabled regression analyses among the pK_a s scales in both ILs and molecular media, which exhibit excellent linear correlations of the acidity scales both in ILs and in between the ILs and molecular solvents (water, DMSO and ACN). Such relationships allow reasonable estimates for the unknown acidities of sulfonamides in these solvents. A detailed comparison among the acidity scales in these ionic and molecular media finds an interesting trend that the ILs of low ϵ value can show an unusually strong ability (vs. ACN, $\epsilon = 36.1$) in facilitating deprotonation. The absolute acidities and the solvation insights disclosed in this work may serve either as benchmark values or reliable references for the development of appropriate theoretical solvation models and for mechanistic analyses in ILs.

EXPERIMENTAL

General

ILs (Scheme 1)²⁰, carbon indicator acids (fluorene derivatives, Table S1) and the base used in this work were synthesized and purified as described in previous work.⁹ In addition, ILs were dried in vacuum at 70 °C for 5 hours before use and were stored in desiccators under argon. The water contents of ILs are less than 10 ppm, which was determined by the Karl-Fischer titration. Sulfonamides, except otherwise noted, were commercially available and were carefully recrystallized and dried before used as substrates. The sulfonamides and indicator acids were kept in a glove box. The principle of acidity measurement by indicator overlapping method^{11c, 13} is introduced as Equation 4:



Where HIn and HA denotes the indicator and substrate acids respectively. It is worth noting that all the acidities of indicator acids were referenced to an anchor compound whose acidity was measured by self-dissociation (SI), so the pK_a s of these indicator acids are absolute values in essence. All manipulations were

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3 carried out under dry argon using standard Schlenk techniques. The pK_a measurement started by adding 1.5
4 ml IL solution of base ($\sim 5 \times 10^{-4}$ M) into an empty UV cell with a 3-way valve (Figure S1), and then the
5 spectrum for baseline was recorded on a UV instrument. Next an IL solution of an appropriate indicator acid
6 (HIn) with known pK_a was added dropwisely until the UV absorbance was not increasing with the addition.
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8 Then the IL solution (10^{-5} to 10^{-4} M) of substrate acid of interest (HA) was added in several portions. The
9 weight of the UV cell and the corresponding spectrum were also recorded upon each addition. The pK_a of HA
10 was then calculated from the change of absorbance and the amount of acid added.
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17 **Synthesis of picric derivatives as indicators**²¹

18 a. 3-chloropicric acid (Scheme S1, C):

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21 To a 100 ml 3-neck round bottom flask charged with 2.5 g (19.5 mmol) 3-chlorophenol, under an acetone/dry
22 ice bath, 10 ml concentrated sulfuric acids (98% w/w, 0.183 mol) was added slowly, then the temperature of
23 reaction mixture was raised cautiously to 90 °C and kept at this temperature for 40 mins before the
24 temperature was cooled down to 0 °C by a water and ice bath (NaCl/icewater). Again under the acetone/dry
25 ice bath, with cautions 10 ml fuming nitric acid (90% w/w, 0.214 mol) was added dropwisely, then the reaction
26 temperature was allowed to rise to 80 °C and was held at this temperature for 50 mins. After system was
27 cooled down to ambient temperature, the reaction mixture was then poured into ice water, the brown
28 yellowish precipitates were collected and recrystallized from water once and CCl_4 twice to give a yellowish
29 solid 2.3g. Yield: 45%. 1H NMR (400 MHz, $DMSO-d_6$) δ 8.77 (s, 1H); ^{13}C NMR (100 MHz, $DMSO-d_6$) δ 127.9.
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42 b. 3, 5-dichloropicric acid (Scheme S1, D):

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45 The starting material was 3,5-dichlorophenol, synthetic procedures were similar to these for 3-chloropicric
46 acid (C) describe above, which gave a yellow solid with a yield of 38%. 1H NMR (400 MHz, $DMSO-d_6$) δ 14.58
47 (s, 1H); ^{13}C NMR (100 MHz, $DMSO-d_6$) δ 155.6, 143.1, 127.7, 121.0.
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51 **Synthesis of sulfonamides**

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54 The commercial available (**1a** and **1b**) and prepared sulfonamides (**2a-8a**, **2b-4c**) were carefully recrystallized
55 at least 3 times and stored in glove box before used in pK_a measurement. The synthetic procedures for the
56 sulfonamides used in this work can be found elsewhere.²²
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3 PhNHSO₂Ph (**2a**), yield: 72%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.37 (s, 1H), 7.80 (d, *J* = 7.2 Hz, 2H), 7.60–7.48
4 (m, 3H), 7.22 (t, *J* = 7.8 Hz, 2H), 7.14 (d, *J* = 7.7 Hz, 2H), 7.00 (t, *J* = 7.2 Hz, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆)
5 δ 140.0, 138.2, 133.3, 129.7, 129.6, 127.1, 124.5, 120.5.^{22b}
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10 p-MeO-PhNHSO₂Ph (**3a**), yield: 58%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.94 (s, 1H), 7.69 (d, *J* = 7.1 Hz, 2H),
11 7.60–7.45 (m, 3H), 6.98 (d, *J* = 7.2 Hz, 2H), 6.80 (d, *J* = 8.6 Hz, 2H), 3.66 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆)
12 δ 157.0, 139.9, 133.2, 130.5, 129.6, 127.1, 123.9, 114.7, 55.5.^{22b}
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16 p-Me-PhNHSO₂Ph (**4a**), yield: 68%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.53 (s, 1H), 7.80 (d, *J* = 7.1 Hz, 2H),
17 7.60–7.48 (m, 3H), 7.28 (d, *J* = 8.7 Hz, 2H), 7.15 (d, *J* = 8.7 Hz, 2H), 10.14 (s, 1H), 7.73 (d, *J* = 7.1 Hz, 2H), 7.58–
18 7.49 (m, 3H), 7.04–6.95 (m, 4H), 2.17 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 139.9, 135.4, 133.9, 133.3, 130.0,
19 129.7, 127.1, 121.1, 20.8.^{22b}
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25 m-Me-PhNHSO₂Ph (**5a**), yield: 69%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.29 (s, 1H), 7.80 (d, *J* = 7.2 Hz, 2H),
26 7.60–7.50 (m, 3H), 7.09 (t, *J* = 8.1 Hz, 1H), 7.16 (s, 1H), 6.98–6.88 (m, 2H), 6.82 (d, *J* = 7.9 Hz, 1H), 2.16 (s, 3H);
27 ¹³C NMR (100 MHz, DMSO-*d*₆) δ 140.0, 138.9, 138.1, 133.3, 129.7, 129.4, 127.1, 125.3, 121.0, 117.5, 21.5.^{22b}
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32 p-Cl-PhNHSO₂Ph (**6a**), yield: 63%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.53 (s, 1H), 7.80 (d, *J* = 7.1 Hz, 2H),
33 7.60–7.48 (m, 3H), 7.28 (d, *J* = 8.7 Hz, 2H), 7.15 (d, *J* = 8.7 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 139.6, 137.1,
34 133.5, 129.7, 129.6, 128.7, 127.1, 122.0.^{22b}
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39 m-Cl-PhNHSO₂Ph (**7a**), yield: 65%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.66 (s, 1H), 7.82 (d, *J* = 7.2 Hz, 2H),
40 7.63–7.50 (m, 3H), 7.24 (t, *J* = 8.1 Hz, 1H), 7.16 (s, 1H), 7.11 (d, *J* = 8.2 Hz, 1H), 7.05 (d, *J* = 7.9 Hz, 1H); ¹³C NMR
41 (100 MHz, DMSO-*d*₆) δ 139.8, 139.6, 133.9, 133.6, 131.3, 129.8, 127.1, 124.2, 119.5, 118.4.^{22b}
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46 m-NO₂-PhNHSO₂Ph (**8a**), yield: 48%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.98 (s, 1H), 7.99 (s, 1H), 7.85 (d, *J* =
47 7.2 Hz, 2H), 7.81 (d, *J* = 7.9 Hz, 1H), 7.60–7.51 (m, 4H), 7.48 (t, *J* = 8.1 Hz, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ
48 148.6, 139.6, 139.4, 133.7, 131.1, 129.9, 127.1, 125.7, 118.8, 113.9.^{22b}
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53 PhSO₂NHSO₂Ph (**9a**), yield: 33%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.87 (s, 1H), 7.71 (d, *J* = 7.4 Hz, 4H), 7.51 (t, *J*
54 = 7.2 Hz, 2H), 7.44 (t, *J* = 7.4 Hz, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 144.3, 131.9, 128.9, 126.8.^{22c}
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58 p-Cl-PhSO₂NHSO₂Ph (**10a**), yield: 31%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.65 (s, 1H), 7.74–7.68 (m, 4H),
59 7.50–7.45 (m, 3H), 7.41 (t, *J* = 7.2 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 144.4, 143.5, 136.5, 131.8, 128.9, 128.8,
60 126.8.^{22c}

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3 PhNHTf (**2b**), yield: 67%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.91 (s, 1H), 7.42 (t, *J* = 7.7 Hz, 2H), 7.29 (d, *J* = 6.8
4 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 135.3, 130.0, 127.1, 123.3, 120.2 (¹*J*_{CF} = 322 Hz).^{22b}

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7
8 p-MeO-PhNHTf (**3b**), yield: 59%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.60 (s, 1H), 7.21 (t, *J* = 8.3 Hz, 2H), 6.97 (d,
9 *J* = 8.8 Hz, 2H), 3.75 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 158.6, 127.3, 126.5, 120.3 (¹*J*_{CF} = 323 Hz), 55.7.^{22b}

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13 p-Cl-PhNHTf (**4b**), yield: 72%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.02 (s, 1H), 7.49 (d, *J* = 8.8 Hz, 2H), 7.29 (d, *J*
14 = 8.3 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 134.4, 131.3, 130.0, 124.8, 120.1 (¹*J*_{CF} = 321 Hz).^{22b}

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17
18 p-CF₃-PhNHTf (**5b**), yield: 64%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.33 (s, 1H), 7.79 (d, *J* = 8.4 Hz, 2H), 7.46 (d,
19 *J* = 8.3 Hz, 2H).^{22b}

20
21
22
23 p-NO₂-PhNHTf (**6b**), yield: 68%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.12 (s, 1H), 8.26 (d, *J* = 9.0 Hz, 2H), 7.46 (d,
24 *J* = 9.0 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 144.5, 143.1, 125.6, 121.4, 120.1 (¹*J*_{CF} = 322 Hz).^{22b}

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27
28 PhCONHTf (**7b**), yield: 59%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.16 (s, 1H), 7.93 (d, *J* = 7.3 Hz, 2H), 7.47 (d, *J* =
29 7.3 Hz, 1H), 7.40 (t, *J* = 7.2 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 169.9, 131.6, 129.0, 128.3.^{22d}

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33 p-MeO-PhCONHTf (**8b**), yield: 62%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.10 (s, 1H), 7.91 (d, *J* = 8.6 Hz, 2H),
34 6.98 (d, *J* = 8.7 Hz, 2H), 3.79 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 167.8, 163.1, 131.4, 127.0, 120.3 (¹*J*_{CF} = 322
35 Hz), 113.9, 55.8.^{22d}

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39 p-Me-PhCONHTf (**9b**), yield: 66%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.23 (s, 1H), 7.83 (d, *J* = 7.9 Hz, 2H), 7.23
40 (d, *J* = 7.9 Hz, 2H), 2.32 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 168.9, 142.5, 133.0, 129.3, 129.1, 120.5 (¹*J*_{CF} = 322
41 Hz), 21.5.^{22d}

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45 p-Cl-PhCONHTf (**10b**), yield: 61%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.30 (s, 1H), 7.92 (d, *J* = 8.3 Hz, 2H), 7.43
46 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 168.9, 136.7, 135.8, 130.9, 128.5, 120.6 (¹*J*_{CF} = 322 Hz).^{22d}

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50 p-F-PhCONHTf (**11b**), yield: 49%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.43 (s, 1H), 7.98 (dd, *J* = 8.5 Hz, ⁴*J*_{HF} = 5.9
51 Hz, 2H), 7.19 (t, *J* = 8.8 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 168.8, 164.6 (¹*J*_{CF} = 248 Hz), 133.5, 131.7 (³*J*_{CF} =
52 9 Hz), 120.6 (¹*J*_{CF} = 323 Hz), 115.3 (²*J*_{CF} = 21 Hz).^{22d}

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57 MeSO₂NHTos (**2c**), yield: 48%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.09 (s, 1H), 7.75 (d, *J* = 8.2 Hz, 2H), 7.39 (d, *J*
58 = 8.1 Hz, 2H), 3.15 (s, 3H), 2.38 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 143.6, 138.5, 129.8, 127.3, 43.8, 21.5.^{22c}

PhSO₂NHTos (**3c**), yield: 28%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.80 (s, 1H), 7.75 (d, *J* = 7.4 Hz, 2H), 7.63 (d, *J* = 8.2 Hz, 2H), 7.56 (t, *J* = 7.3 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 2H), 7.27 (d, *J* = 8.1 Hz, 2H), 2.33 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 143.1, 142.8, 139.9, 132.7, 129.6, 129.2, 127.1, 127.0, 21.4.^{22c}

p-NO₂-PhSO₂NHTos (**4c**), yield: 25%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.76 (s, 1H), 8.21 (d, *J* = 8.7 Hz, 2H), 7.88 (d, *J* = 8.6 Hz, 2H), 7.54 (d, *J* = 8.0 Hz, 2H), 7.15 (d, *J* = 7.9 Hz, 2H), 2.28 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 151.6, 148.7, 142.6, 141.1, 129.0, 128.3, 126.8, 124.0, 21.3.^{22c}

ASSOCIATED CONTENT

Supporting Information.

The Supporting Information is available free of charge on the ACS Publications website. Picture of the UV-cell; expansion of the indicator acidity scale; indicators used in this work and their corresponding p*K*_as in ILs; UV-vis spectra for representative measurements; regression analysis; NMR spectra.

NOTES

The authors declare no competing financial interest.

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