## Novel polyoxazole-based cyclopeptides from *Streptomyces sp.* Total synthesis of the cyclopeptide YM-216391 and synthetic studies towards telomestatin<sup>†</sup>

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Received 13th February 2008, Accepted 6th March 2008
First published as an Advance Article on the web 11th April 2008
DOI: 10.1039/b802477d

A convergent, complementary, synthetic approach to the contiguously linked tris-oxazole units 10, 11 and 12 in telomestatin (1) and YM-216391 (2) is described. The route involves coupling reactions between oxazole 4-carboxylic acids, viz 16a, 16c, 16d and oxazole 2-substituted methylamines, viz 16b, 16e, 17, leading to the amides 18 and 21, followed by cyclodehydrations to the corresponding bis-oxazole oxazolines, e.g. 19, and oxidations of the latter using well-established protocols. The tris-oxazoles 11 and 12 were next converted stepwise into the hexa-oxazole bis-macrolactams 33. Although the bis-macrolactams 33 (cf. 28) could be converted into the corresponding oxazoline-hexa-oxazoles 34 and to the enamides 35, neither of these intermediates could be elaborated to the hepta-oxazole 30 en route to telomestatin 1. Likewise, neither the hexa-oxazole 47 or application of an intramolecular Hantzsch oxazole ring-forming reaction from 44b allowed access to the advanced polyoxazole-macrolactam intermediates 48 and 30a, respectively, towards telomestatin. Combination of the tris-oxazole based methylamine 70 with the dipeptide carboxylic acid 71 derived from D-valine and L-isoleucine, leads to the corresponding amide which, in two straightforward steps, is converted into the ω-amino acid 78. Macrolactamisation of 78, using HATU, next produces the cyclopeptide 79 which is then elaborated to the thiazole and oxazole based cyclopeptide YM-216391 (2). The synthetic cyclopeptide 2 is shown to be the enantiomer of the natural product isolated from Streptomyces nobilis

#### Introduction

Telomestatin (1)¹ and YM-216391 (2)² are members of an ever-expanding family of novel macrocycles, containing contiguously-linked 2,4-disubstituted oxazoles and thiazoles, to be isolated from nature. Other members include ulapualide A (3),³ diazonamide A (4)⁴ and IB-01211 (5).⁵ Closely related structures are those cyclic peptides which contain oxazoles, thiazoles, oxazolines and thiazolines linked by several amide bonds, e.g. dendroamide A (6),⁶ patellamide B (7)² and ascidiacyclamide 8,⁶ and the oxazole-pyridine-thiazole linked cyclopeptide antibiotics, represented by sulfomycin II (9).⁰

The contiguously-linked oxazoles **3** and **4**, together with the azole-based cyclic peptides **6**, **7** and **8** are mainly of marine origin, whereas those macrocycles **1**, **2**, **5**, and **9** incorporating thiazole/thiazoline rings have, so far, been isolated from microorganisms, *e.g. Streptomyces* bacteria. Telomestatin **1** is a potent inhibitor of telomerase, interacting specifically with the G-quadruplex, and not affecting DNA polymerases or reverse transcriptases; <sup>10</sup> it is showing promise in cancer therapy. The cyclic polyoxazole IB-01211 (**5**) is toxic against tumour cell lines and, like YM-216391 (**3**) shares a structural homology with telomestatin. Indeed, all of the cyclic polyazole based secondary metabolites **1**-**9** show a diverse array of interesting biological

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† Electronic supplementary information (ESI) available: Further experimental details for synthesised compounds. See DOI: 10.1039/b802477d

activities, 12 which has made them attractive targets to synthetic chemists in recent years. 13

In this paper we describe our synthetic studies towards a variety of contiguously linked tris-, penta- and hepta-azole units present in telomestatin and YM-216391, which have culminated in a total synthesis of YM-216391 (2)<sup>14</sup> and the synthesis of several polyoxazole acyclic precursors and macrocyclic intermediates related to telomestatin 1.<sup>15</sup>

#### Results and discussion

### Synthetic strategy, and studies towards arrays of contiguous polyoxazole macrocycles

At the outset of our synthetic studies with telomestatin and YM-216391 only the tris-oxazole based macrolides ulapualide A (3) and mycalolide A had succumbed to total synthesis. <sup>16,17</sup> In these syntheses the tris-oxazole units were either elaborated from amide serine precursors by cyclodehydration to oxazoline intermediates followed by oxidation, <sup>18</sup> or by applying iterative Hantzsch oxazolering syntheses. <sup>19</sup> Both of these approaches were reliable, robust and high yielding overall. However, the prospect of applying a similar linear approach to the polyazole units in telomestatin or YM-216391 was prohibitive. Other synthetic approaches that have been used to elaborate consecutively linked azoles include: sequential [3 + 2] cycloaddition reactions between rhodium carbenoids and nitriles; Pd(0) catalysed cross coupling reactions; rearrangements of tertiary amides; and base-catalysed cyclisations

of alkynyl glycine derivatives.<sup>20,21</sup> To a large degree based on our contemporaneous synthetic studies towards ulapualide A (3), 16,18 where we had used a biogenetically-patterned approach to the tris-oxazole unit, i.e. making the central oxazole by an initial macrolactamisation from an ω-amino acid precursor, we designed a variety of convergent synthetic approaches to the polyoxazole arrays in 1 and 2, involving macrolactamisations of bis-, tris- and higher polyoxazoles.

The contiguous tris-oxazole unit 10, i.e. rings A, B, C in YM-216391 (2) is directly related to the EFG ring system 11 in telomestatin (1). The same ABC tris-oxazole unit 10 in 2 is also analogous to the CBA ring system 12 in telomestatin, except that rings A and B in the latter structure each contain an additional C-2-methyl substituent, *i.e.* the oxazole rings are derived in nature from threonine rather than serine. In our complementary synthetic investigations towards telomestatin and YM-216391, therefore, the closely related substituted tris-oxazoles 10, 11 and 12 became central synthetic precursors. After examining a range of synthetic strategies we followed a similar convergent approach to each of the tris-oxazoles 10, 11 and 12, involving amide coupling reactions between pre-formed oxazole-4-carboxylic acids, e.g. 16a/16c, and oxazole-4-substituted amines, e.g. 16b/17c (Scheme 1). The bisoxazole amides 18 and 21 were then elaborated to the trisoxazoles 20 and 22 respectively following cyclodehydrations to the corresponding oxazolines e.g. 19 and oxidations of the latter using the procedures and reagents presented in Scheme 2.22 Saponifications of the methyl ester groups in 22 and 20 then gave the carboxylic acids 23 and 26 respectively. Likewise, removal of the N-protecting groups in 20 and 22, led to the corresponding amines 24 and 25 respectively (Scheme 3).

#### Synthetic studies towards telomestatin 1

With the tris-oxazole-based, and suitably protected, amino acids 23, 24, 25 and 26 available, in quantity, via the aforementioned synthetic investigations, a number of tactics were considered for their combination, en route to telomestatin 1. In order to allow the greatest conformational flexibility in key macrocyclisation steps, however, we decided to first combine the acids 23/26 with the amines 24/25, leading to the respective amides 27/29, and then to effect macrolactamisations from 27/29 leading to the corresponding bis-macrolactam 28 as a key intermediate (Scheme 4). Subsequent manipulation of the hydroxyl protecting groups in 28, accompanied by sequential oxazole/thiazoline ringforming reactions, via intermediates 30/31, would then complete a complementary, convergent synthetic approach to telomestatin. Intuitively, we felt that the tactic of leaving the formation of the thiazoline ring in telomestatin to last i.e. proceeding via the intermediate 30 was the most prudent. We also felt that the synthesis of 30 starting from 27 would give us the greater number of options to determine at which stage we would introduce a thio ether and/or thioamide unit for thiazoline-ring formation in the natural product.

Thus, a coupling reaction between the amine 24a and the carboxylic acid 23a, using EDC-HOBt-NMM23 first gave the amide 27 in 68% yield. Removal of the Z-carbamate group in

27 by hydrogenolysis, followed by saponification of the resulting amino ester 32a next gave the amino acid salt 32b (Scheme 5). Macrolactamisation of 32b in the presence of DPPA-HOBt then gave an excellent yield of the hexa-oxazole-bis-macrolactam 28. Removal of the silvl ether protecting group in 28 followed by treating the resulting alcohol 33a with DAST<sup>24</sup> led to the (ring D) oxazoline 34a in 90% yield. We had hoped that oxidation of the oxazoline ring in 34a to the corresponding heptaoxazole 30a, followed by manipulation of the remaining amide unit in 30a would then provide an expeditious synthesis of telomestatin 1. How wrong we were! We examined a variety of oxidising agents and conditions, including nickel peroxide, 25 DDQ, 26 and BrCCl<sub>3</sub>-DBU22 to convert 34a into 30a but all to no avail; on each occasion either starting material was recovered unchanged or intractable material resulted. Furthermore, treatment of the alcohol 33a with PDC failed to give any of the corresponding aldehyde;<sup>27</sup> hence we were not able to investigate a more direct Robinson Gabriel oxazole-ring forming reaction approach from 33a to 30a (Scheme 5). Interestingly, when 33a was treated with Dess-Martin periodinane followed by immediate treatment with PPh<sub>3</sub>, 1,2dibromotetrachloroethane, di-tert-butylpyridine and then with DBU, work-up gave the enamide 35a resulting from overall dehydration of 33a. Unfortunately we were not able to convert the enamide 35a into the corresponding oxazole 30a following the usual treatment with Br<sub>2</sub>-Et<sub>3</sub>N, leading to the intermediate vinyl bromide 35b, and cyclisation of the latter in the presence of Cs<sub>2</sub>CO<sub>3</sub>.18,28

We surmised that a large part of our problem in completing a synthesis of telomestatin 1 from the intermediate bis-macrolactam 28 had its origins in the very poor solubility of 28 and of compounds produced from it, making characterisation inconclusive. In anticipation therefore we changed the protecting groups in the trisoxazole precursors 20 and 22 and made the alternative silyl/benzyl ether protected bis-macrolactam 37, cf. 28 (Scheme 6). Much to our chagrin, although we could convert 37 into the oxazolinehexa-oxazole 34b, we simply could not oxidise this compound to the corresponding hepta-oxazole 30b and therefore complete a synthesis of telomestatin by this route.

With the persistent problem in attempting to elaborate the ring D oxazole in structure 30 via the intermediates 28, 34

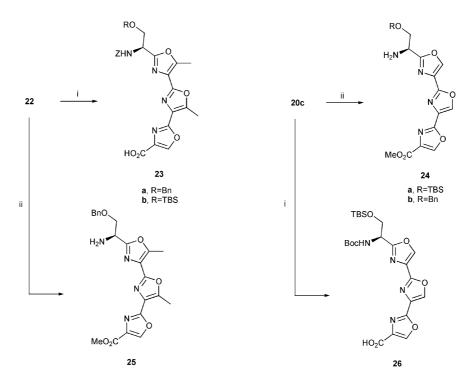
RO 
$$\stackrel{\text{HO}}{\underset{\text{NHR}^1}{\text{NHR}^2}}$$
  $\stackrel{\text{RO}}{\underset{\text{NHR}^2}{\text{NO}_2\text{Me}}}$   $\stackrel{\text{ii}}{\underset{\text{R}^1\text{HN}}{\text{RO}_2\text{Me}}}$   $\stackrel$ 

a, R=Bn; R<sup>1</sup>=Boc; R<sup>2</sup>=H; b, R=TBS; R<sup>1</sup>=Z; R<sup>2</sup>=H; c, R=Bn; R<sup>1</sup>=Z; R<sup>2</sup>=Me; d, R=TBS; R<sup>1</sup>=Z; R<sup>2</sup>=Me; e, R=TBS; R<sup>1</sup>=Z; R<sup>2</sup>=Ph

- **a**, R=Bn; R<sup>1</sup>=Boc; R<sup>2</sup>=R<sup>3</sup>=H; **b**, R=R<sup>1</sup>=R<sup>2</sup>=H; R<sup>3</sup>=Me
- **c**, R=Bn; R<sup>1</sup>=Z; R<sup>2</sup>=Me; R<sup>3</sup>=H; **d**, R=R<sup>2</sup>=R<sup>3</sup>=H; R<sup>1</sup>=Z;
- e, R=TBS; R<sup>1</sup>=R<sup>2</sup>=H; R<sup>3</sup>=Me; f, R=TBS; R<sup>1</sup>=Z; R<sup>2</sup>=Me; R<sup>3</sup>=H;
- g, R=TBS; R1=Z; R2=R3=H; h, R=TBS; R1=Z; R2=H; R3=Me
- - a, R=TMS, R'=Z, R2=H
  - b, R=H, R<sup>1</sup>=Z, R<sup>2</sup>=H
  - c. R=R1=R2=H

Scheme 1 Reagents and conditions: i, DAST, DCM, -78 °C, 85–95%; ii, BrCCl<sub>3</sub>, DBU, DCM, 0-25 °C, 60–70%; iii, sequential and selective deprotections of OR, NHR1, CO2Me.

Scheme 2 Reagents and conditions: i, EDC, NMM, HOBt, DCM, 50–70%; ii, DAST, DCM, -78 °C, 85–95%; iii, BrCCl<sub>3</sub>, DBU, DCM, 0–25 °C, 60–70%.



Scheme 3 Reagents and conditions: i, NaOH, H<sub>2</sub>O, THF, 93–98%; ii, 10% Pd/C in MeOH, ~95% (for 24a and 25); HCl-dioxane, 98% (for 24b).

Scheme 4

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and 35, we decided to examine a more direct macrocyclisation from corresponding  $\alpha$ -bromoketone-amide intermediates, *viz.* 44b, using the classical Hantzsch reaction. The methyl ketone precursor 44a to 44b was synthesised in a relatively straightforward manner starting from the tris-oxazole carboxylic acid 41 produced from the mono-oxazole 38 and the oxazole amino-alcohol 16b *via* the amide 39 and the oxazoline 40 (Scheme 7). A coupling

reaction between the tris-oxazole carboxylic acid 41b and the secondary amine 42 derived from the previously synthesised tris-oxazole intermediate 25 next led to the double amide 43a, which, in two straightforward steps, was converted into the methyl ketone 44a. Frustratingly, we were not able to convert the methyl ketone 44a into the corresponding bromide 44b as a prelude to investigating the subsequent Hantzsch cyclisation to 30. The

Scheme 5 Reagents and conditions: i, EDC, HOBt, NMM, DCM, 0-25 °C, 68%; ii, Pd/C, MeOH-EtOH; iii, NaOH-H<sub>2</sub>O, THF, 88% over 2 steps; iv, DPPA, NMM, DMF, 86%; v, HCl-dioxane; vi, DAST, DCM, -78 °C, 90% over 2 steps; vii, Dess-Martin periodinane, DCM, rt, then PPh<sub>3</sub>, 1,2-dibromotetrachloroethane, di-tert-butyl pyridine, 0 °C-rt, then DBU, rt, 74% over two steps.

b R=Br

methyl ketone 44a remained unchanged under all but the harshest conditions for bromination, when extensive decomposition ensued.

At this point in our studies we threw caution to the wind, and planned to make the isomeric hexa-oxazole 47, corresponding to telomestatin, and study its macrolactamisation to the precursor 48 to the natural product. Thus, following procedures that had already been established in our work, the bis-oxazole amide 45b was first prepared from the carboxylic acid 15d and the amine 16d. A coupling reaction between 45b and 23 next led to the hexa-oxazole 46 which, in two straightforward steps, was then converted into 47a (Scheme 8). Manipulation of the protecting groups in 47a next led to the sodium salt 47b of the  $\omega$ -amino acid

but, frustratingly, all attempts to form the hexa-oxazole-based macrolactam 48 from 47b met with failure.

Coincident with the abandonment of our synthetic studies towards telomestatin, Takahashi et al.29 described the first and, so far, only reported total synthesis of this novel telomerase inhibitor. Interestingly, for all intents and purposes, their route was strategically identical with the route we had followed ourselves. Thus, Takahashi et al. first prepared the tris-oxazole based carboxylic acid 49 and the amine 50, and next coupled them to form the amide 51 (Scheme 9). Manipulation of the protecting groups in 51, followed by macrolactamistation of the resulting ωamino acid then provided the bis-macrolactam 52 (cf. structure 28). The alcohol residue in 52 was next dehydrated to the enamide

Scheme 6 Reagents and conditions: EDC, NMM, HOBt, DCM, 87%; ii, H<sub>2</sub>, Pd/C, MeOH, THF then NaOH–H<sub>2</sub>O, 76%; iii, HATU, NMM, DMF, DCM, 0–25 °C, 80%; iv, H<sub>2</sub>, Pd(OH)<sub>2</sub>/C, MeOH, THF, then DAST, DCM, –78 °C, 91%.

Scheme 7 Reagents and conditions: i, EDC, NMM, HOBt, 0–25 °C, 67% for **39**; 76% for **43a**; ii, DAST, DCM, –78 °C; iii, BrCCl<sub>3</sub>, DBU, DCM, 0–25 °C, 44% over 2 steps; iv, NaOH, THF, H<sub>2</sub>O, 25 °C, 85%; v, TBAF, THF, 0–25 °C, 87%; vi, PDC, DCM, 25 °C, 98%.

**53** (*cf.* structure **35a**), which was then converted into the oxazoline **54** (*cf.* structure **34**) using NBS in MeOH, followed by treatment of the intermediate α-methyl-β-bromo cyclic peptide with  $K_2CO_3$ . Elimination of methanol from **54** using camphorosulfonic acid produced the hepta-oxazole **55** which was finally converted into telomestatin, following simultaneous deprotection of the *t*-Bu thio ether and cyclodehydration using  $Ph_3PO-Tf_2O$  in  $CH_2Cl_2^{30}$  at room temperature.

Comparison of the routes to telomestatin followed by Takahashi et al. and by ourselves revealed a number of similarities

and also some subtle differences in reaction conditions. Thus, whereas we had been able to smoothly convert the hydroxymethyl amide intermediate 33 into the corresponding oxazoline 34 using DAST,<sup>24</sup> Takahashi *et al.* found that treatment of their analogous intermediate 52 with DAST (or Burgess reagent)<sup>31</sup> instead gave the enamide 53. Takahashi *et al.* were then able to convert the enamide 53 into the hepta-oxazole 55 *via* the oxazoline intermediate 54, whereas we were not able to convert our enamide 35, or oxidise the oxazoline 34 into the corresponding hepta-oxazole 30 using a range of conditions. It does seem likely therefore

Scheme 8 Reagents and conditions: i, EDC, NMM, HOBt, DCM, 34%; ii, H2, Pd/C, MeOH-EtOAc, 55%; iii, 23, EDC, NMM, HOBt, DCM, 50%; iv, DAST, DCM, -78 °C, then BrCCl<sub>3</sub>, DBU, 43%; v, H<sub>2</sub>, Pd/C, MeOH, 99%.

Scheme 9 Reagents and conditions: i, PyBroP, DIEA, DCM, DMF, rt, 85%; ii, 4 M HCl, 1,4-dioxane, rt; iii, LiOH, MeOH–THF–H<sub>2</sub>0 (3:3:1); iv, DPPA, HOBt, DIEA, DMAP, DMF, DCM, rt, 48%; v, MsCl, DBU, DCM, 0 °C, 92%; vi, NBS, MS 4 Å, DCM, MeOH, 0 °C, 90%; vii, K2CO<sub>3</sub>, 1,4-dioxane, 60 °C, 79%; viii, camphorsulfonic acid, toluene, MS 5 Å, 70 °C; ix, Ph<sub>3</sub>P(O)–Tf<sub>2</sub>O, anisole, DCM, rt, 20% isolated yield.

that we were simply unfortunate, at the time, with our choice of reaction conditions attempting to elaborate the hepta-oxazole 30 from the bis-macrolactam intermediate 33, on the way to telomestatin.

#### Total synthesis of (-)-YM-216391 (2)

The structure of YM-216391 (2), isolated from Streptomyces nobilis,<sup>2</sup> consists of a continuum of five azoles which have their origins in serine, cysteine and phenylalanine, linked via a glycine-valine-isoleucine tripeptide tether. At the outset of our studies the stereochemical assignment of the valine and isoleucine stereocentres in the natural product had not been established. We therefore chose a synthetic strategy to 2 that would allow us to introduce these two stereocentres as late as possible in the synthesis, and involving a penta-azole intermediate 58 and a valine-isoleucine dipeptide fragment, i.e. 57. Furthermore, we initially decided to effect the key macrolactamistion from the  $\omega$ amino acid 56, i.e. using the sterically less encumbered glycine amine-valine carboxylic acid amide bond connection (Scheme 10). The penta-azole based  $\omega$ -amino acid **56**, derived from an L-(natural) isoleucine-valine dipeptide fragment therefore became our first macrolactamisation precursor.

Thus, the 2,4-disubstituted oxazoles 59 and 60 and the trisubstituted oxazole 61 were first elaborated from their constituent amino acids using methodologies which are well precedented in this paper and in the literature. A coupling reaction between 60 and 59b in the presence of EDC-HOBt-NMMO23 led to the amide 62a which was next converted into the tris-oxazole 63a via 62a, in three straightforward steps (Scheme 11). Saponification of 63a, followed by coupling the resulting carboxylic acid 63b with the amine 61b, then led to tetra-oxazole amide 64. Treatment of 64 with Lawesson's reagent<sup>32</sup> gave the thioamide 65 which, after deprotection and reaction with DAST,24 was cyclised to the thiazoline 66 (Scheme 12). Finally, when the thiazoline 66 was reacted with BrCCl<sub>3</sub>-DBU<sup>22</sup> it was converted into the corresponding thiazole 67a which could be saponified to the carboxylic acid 67b. A coupling reaction between the carboxylic acid 67b and the L-isoleucine-L-valine dipeptide 68 next gave the substituted penta-azole 69 which, after double deprotection of the ester and *N*-BOC groups gave the  $\omega$ -amino acid **56**. Attempts to effect macrolactamisation of **56**, using a variety of traditional reagents, however, led to intractable mixtures of polymeric material and none of the anticipated YM-216391 (2).

We reasoned that the failure to obtain macrolactamisation from the intermediate ω-amino acid 56 had its origins in poor conformational flexibility, i.e. the penta-azole ring system in 56 was too rigid. We therefore decided to examine the thioamide intermediate 73 analogous to 56 in the key macrolactamisation step, with a view to introducing the thiazole ring in the target as the

$$\begin{array}{c} & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ &$$

Scheme 11 Reagents and conditions: i, EDC, HOBt, NMM, DCM, 56%; ii, H<sub>2</sub>, Pd(OH)<sub>2</sub>/C, MeOH-THF, 81%; iii, DAST, DCM, -78 °C; iv, BrCCl<sub>3</sub>, DCM, 57% over two steps; v, NaOH, THF, H<sub>2</sub>O, 88%.

Scheme 12 Reagents and conditions: i, EDC, HOBt, NMM, DCM, 87%; ii, Lawesson's reagent, THF, 50%; iii, TBAF, THF, 95%, then DAST, DCM, 59%; iv, BrCCl<sub>3</sub>, DBU, DCM; v, NaOH, THF–H<sub>2</sub>O; vi, EDC, HOBt, NMM, DCM; vii, 4 M HCl in dioxane; then NaOH, THF, H<sub>2</sub>O.

final step in the synthesis. The previously synthesised thioamide 65 was therefore first deprotected and the resulting amine 70 was then coupled to the protected dipeptide 71 leading to the intermediate 72. The methyl ester and the N-BOC protecting groups in 72 were next removed, in sequence, producing the  $\omega$ amino acid 73, which underwent smooth macrolactamisation in the presence of HATU<sup>33</sup> leading to the cyclopeptide 74 in 88% yield (Scheme 13). Finally, the thioamide unit in 74 was converted into the corresponding thioazoline 75, using DAST, which then underwent oxidation, in the presence of MnO<sub>2</sub> to produce the YM-216391 structure with the stereochemistry shown in formula 76. When we compared the <sup>1</sup>H NMR spectroscopic data for 76 with those recorded for natural YM-216391, although there were close similarities, the two structures were not identical. We surmised that the two structures differed in their stereochemistry at the valine residue and that the natural product was most likely derived from D- rather than L-valine. This situation is perhaps not too surprising, given the prevalence of cyclopeptides such as ascidiacylamide (8)8 and dendroamide A (6),6 which are made up from nonproteinogenic D-amino acids. We therefore re-synthesised the dipeptide 77 using D-valine and L-isoleucine, and then reproduced the sequence of reactions shown in Scheme 13, producing the cyclopeptide intermediate 79 (Scheme 14). The cyclopeptide 79 differs from structure 74 only according to its stereochemistry at the isopropyl bearing carbon centre. Following conversion of the thioamide unit in 79 into the thiazoline 80, oxidation with MnO<sub>2</sub> in  $\text{CH}_2\text{Cl}_2$  finally gave the cyclopeptide **81**  $[a]_D^{20}$  –56  $(c = 0.5, \text{CHCl}_3)$ whose <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data matched very closely those reported for naturally derived YM-216391. Following the completion of our synthesis, which allowed us to establish the relative stereochemistry shown in structure 81 for YM-216391, Sohda et al.34 published details of their investigations

of the structure of YM216391 from *Streptomyces nobilis*. The studies of Sohda *et al.* led to the same assignment of relative stereochemistry for YM216391 as we had determined by synthesis, but the natural product showed  $[a]_D^{25}$  +48 (c = 0.1, CH<sub>3</sub>CN), *i.e.* equal and opposite in value to that recorded for our synthetic material. The structural and synthetic studies therefore complemented each other, and established that YM-216391 isolated from *S. nobilis* has the absolute stereochemistry shown in structure 82.

#### **Summary and Conclusions**

A range of robust and efficient methods are available to quickly assemble a variety of amino acid based contiguously linked 2,4-substituted tris-oxazoles, *viz.* 23–26, and to effect their coupling and cyclisation to hexa-oxazole bis-macrolactam intermediates, *i.e.* 33 and 37, *en route* to telomestatin 1. However, the elaboration of these advanced structures to the hepta-oxazole-thiazoline ring systems in telomestatin is thwarted by problems. These problems were associated with the poor solubilities of rotameric mixtures of several intermediates, making characterisations extremely difficult by NMR spectroscopy, and by the important need for proper choices of protecting groups and specific reaction conditions at an advanced stage in the overall synthesis.

By contrast, elaboration of the tris-oxazole based protected amino acid 63 to the cyclic thioamide 74, followed by its conversion into the naturally occurring thiazole-tetra-oxazole based cyclic peptide YM-216391 (81) proved to be trouble-free, and the total synthesis enabled the establishment of the absolute stereochemistry, *i.e.* 82, of this new metabolite isolated from *Streptomyces nobilis*.

Scheme 13 Reagents and conditions: i, 4 M HCl, dioxane, 91%; ii, EDC, HOBt, NMM, DCM, 57%; iii, NaOH, THF, H<sub>2</sub>O; then 4 M HCl in dioxane, 77% over two steps; iv, HATU, NMM, DCM, DMF, 88%; v, DAST, DCM, 50%; vi, MnO<sub>2</sub>, DCM, 27%.

#### **Experimental**

#### General details

Melting points were determined on a Stuart Scientific SMP3 melting point apparatus and are uncorrected. Optical rotations were recorded in spectroscopic grade chloroform or ethanol on a Jasco DIP-370 polarimeter at ambient temperature. [a]<sub>D</sub> values are recorded in units of 10<sup>-1</sup> deg cm<sup>2</sup> g<sup>-1</sup>. Infrared spectra were obtained using a Perkin-Elmer 1600 series FT-IR instrument as dilute solutions in spectroscopic grade chloroform. Proton (1H) NMR spectra were recorded on either a Bruker DPX 360 (360 MHz) or a Bruker DRX 500 (500 MHz) spectrometer as dilute solutions in deuterochloroform, unless stated otherwise. The chemical shifts are quoted in parts per million (ppm) relative to residual chloroform ( $\delta$  7.27) as internal standard. The multiplicity of each signal is designated by the following abbreviations: s, singlet; d, doublet; t, triplet; q, quartet; qn, quintet; m, multiplet; br, broad; app, apparent; obs, obscured. All coupling constants are quoted in Hertz. Carbon-13 (13C) NMR spectra were recorded on either a Bruker DPX 360 (90 MHz) or a Bruker DRX 500 (125 MHz) spectrometer as dilute solutions in deuterochloroform, unless otherwise stated. The chemical shifts are quoted in parts per

million (ppm) relative to internal chloroform standard ( $\delta$  77.0) on a broad band decoupled mode. The multiplicities were determined using a DEPT sequence, and are designated by the following abbreviations: s, quaternary; d, tertiary methine; t, secondary methylene; q, primary methyl. Mass spectra were recorded on a VG Autospec spectrometer using fast atom bombardment (FAB) or chemical ionisation (CI) techniques; a MM-701CF spectrometer using chemical ionisation (CI); or a Micromass LCT spectrometer using electrospray ionistation (ESI). Microanalytical data were obtained on a Perkin-Elmer 240B elemental analyser.

Flash chromatography was performed on Merck silica gel 60 as the stationary phase and the solvents used were of analytical grade or were distilled before use. All reactions were monitored by thin layer chromatography (TLC) using Merck silica gel 60 F<sub>254</sub> precoated aluminium backed plates which were visualised with ultraviolet light and then developed with either iodine on silica, acidic ninhydrin solution, acidic alcoholic vanillin solution, basic potassium permanganate solution or ethanolic phosphomolybdic acid solution.

Routinely, dry organic solvents were stored under nitrogen and/or over sodium wire. When necessary, commonly used organic solvents were dried prior to use. Tetrahydrofuran (THF),

Scheme 14 Reagents and conditions: i, EDC, HOBt, NMM, DCM, 82%; ii, NaOH, THF, H<sub>2</sub>O, then 4 M HCl in dioxane, 70% over two steps; iii, HATU, NMM, DCM, DMF, 75%; iv, DAST, DCM, 89%; v, MnO<sub>2</sub>, DCM, 55%.

(+) - (natural) YM-216391

diethyl ether, benzene and toluene were distilled from sodium benzophenone ketyl or dried by passing through towers of activated alumina. Dichloromethane was distilled from calcium hydride. Anhydrous *N*,*N*-dimethylformamide was obtained from Aldrich. Other organic solvents and reagents were purified by the accepted literature procedures and organic extracts were dried as stated. Petrol refers to petroleum ether (bp 40–60 °C). Solvents were removed on a Büchi rotary evaporator connected to a water pump. Where necessary, reactions requiring anhydrous conditions were performed in flame-dried apparatus under a nitrogen atmosphere.

## 2-((S)-1-{[2-((S)-2-Benzyloxy-1-tert-butoxycarbonylaminoethyl)-oxazole-4-carbonyll-amino}-2-hydroxyethyl)-oxazole-4-carboxylic acid methyl ester (18a)

4-Methylmorpholine (0.40 mL, 3.6 mmol) was added to a stirred suspension of the acid **16a** (0.58 g, 1.6 mmol) and 1-hydroxybenzotriazole (0.24 g, 1.8 mmol) in dry dichloromethane

(40 mL) at 0 °C under a nitrogen atmosphere. 1-[3-(Dimethylamino)propyll-3-ethylcarbodiimide hydrochloride (0.35 g, 1.8 mmol) was added and the mixture was then stirred at 0 °C for 10 min. The hydrochloride salt of amine **16b** (0.56 g, 2.5 mmol) was added in one portion, and the mixture was then allowed to warm to room temperature overnight. The reaction was quenched with water (30 mL) and the separated organic layer was washed with 10% aqueous citric acid ( $3 \times 30$  mL). The combined aqueous extracts were extracted with dichloromethane (3 × 30 mL) and the combined organic extracts were then washed with saturated sodium bicarbonate solution (30 mL), dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica gel using ethyl acetate as eluent to give the amide (1.53 g, 62%) as a colourless foaming solid;  $[a]_{p}^{22}$  -36  $(c = 1.0, \text{CHCl}_3); \nu_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1} 3444, 3401, 2954, 1714, 1678$ and 1598;  $\delta_{\rm H}(360 \text{ MHz}, \text{CDCl}_3)$  8.20 (1H, s, oxazole-H), 8.10 (1H, s, oxazole-H), 8.04 (1H, d, J 8.6 Hz, CONH), 7.35–7.18  $(5H, m, 5 \times aryl-H), 5.70 (1H, d, J 9.2 Hz, BocNH), 5.54-5.48$  (1H, m, CONHC*H*), 5.09–5.02 (1H, m, BocNHC*H*), 4.53 (1H, d, *J* 12.1 Hz, OCH<sub>a</sub>H<sub>b</sub>Ph), 4.49 (1H, d, *J* 12.1 Hz, OCH<sub>a</sub>H<sub>b</sub>Ph), 4.28 (1H, dd, *J* 11.7 and 4.6 Hz, C*H*<sub>a</sub>H<sub>b</sub>OH), 4.05 (1H, dd, *J* 11.7 and 4.2 Hz, CH<sub>a</sub>H<sub>b</sub>OH), 3.88 (3H, s, CO<sub>2</sub>C*H*<sub>3</sub>), 3.79 (1H, dd, *J* 9.7 and 4.4 Hz, C*H*<sub>a</sub>H<sub>b</sub>OBn), 3.73 (1H, dd, *J* 9.7 and 4.6 Hz, CH<sub>a</sub>H<sub>b</sub>OBn), 2.62 (1H, br s, O*H*) and 1.46 (9H, s, OC(C*H*<sub>3</sub>)<sub>3</sub>) ppm;  $\delta$ <sub>C</sub>(90 MHz, CDCl<sub>3</sub>) 163.1 (s), 162.3 (s), 161.4 (s), 155.2 (s), 150.8 (s), 144.3 (d), 141.7 (d), 137.3 (s), 135.4 (s), 133.0 (s), 128.4 (d), 127.8 (d), 127.5 (d), 80.3 (s), 73.0 (t), 69.8 (t), 62.8 (t), 52.2 (q), 49.3 (d) and 28.2 (q) ppm; *m/z* (ESI) found: 553.1865, C<sub>25</sub>H<sub>30</sub>N<sub>4</sub>O<sub>9</sub>Na [(M + Na)<sup>+</sup>] requires 553.1910.

### 2"-((S)-2-Benzyloxy-1-*tert*-butoxycarbonylaminoethyl)-[2,4';2',4"|teroxazole-4-carboxylic acid methyl ester (20a)

(Diethylamino)sulfur trifluoride (0.48 mL, 3.6 mmol) was added dropwise over 1 min to a stirred solution of the bis-oxazole 18a (1.6 g, 3.0 mmol) in dry dichloromethane (30 mL) at -78 °C under a nitrogen atmosphere. The mixture was stirred at -78 °C for 1 h and then allowed to warm to room temperature. The mixture was quenched with saturated sodium bicarbonate solution (20 mL) and the separated organic layer was then dried (MgSO<sub>4</sub>) and concentrated *in vacuo* to leave the crude oxazoline 19a, which was used immediately without further purification.

Bromotrichloromethane (0.87 mL, 9.0 mmol) was added to a stirred solution of the crude oxazoline in dry dichloromethane (30 mL) at 0 °C under a nitrogen atmosphere and the mixture was stirred at 0 °C for 5 min. 1,8-Diazabicyclo[5.4.0]undec-7-ene (1.4 mL, 9.0 mmol) was added dropwise over 2 min and the mixture was allowed to warm to room temperature overnight. The reaction was quenched with 10% aqueous citric acid (20 mL) and the separated organic extract was then washed with saturated sodium bicarbonate solution (20 mL), dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica gel using dichloromethane-diethyl ether (4:1) as eluent to give the tris-oxazole (1.1 g, 72%) as a colourless solid; mp 202–203 °C (from dichloromethane–petrol);  $[a]_D^{21}$  –25 (c = 1.0, CHCl<sub>3</sub>);  $v_{\text{max}}$ (CHCl<sub>3</sub>)/cm<sup>-1</sup> 3441, 3170, 2980, 1715, 1654 and 1579;  $\delta_{\rm H}$ (360 MHz, CDCl<sub>3</sub>) 8.43 (1H, s, oxazole-H), 8.33 (1H, s, oxazole-H), 8.32 (1H, s, oxazole-H), 7.35–7.19 (5H, m, 5  $\times$ aryl-H), 5.57 (1H, d, J 8.2 Hz, BocNH), 5.22–5.13 (1H, m, BocNHCH), 4.55 (1H, d, J 12.1 Hz, OCH<sub>a</sub>H<sub>b</sub>Ph), 4.51 (1H, d, J 12.1 Hz, OCH<sub>a</sub> $H_b$ Ph), 4.00-3.93 (1H, m, C $H_a$ H<sub>b</sub>OBn), 3.97 (3H, s,  $CO_2CH_3$ ), 3.85 (1H, dd, J 9.6 and 4.4 Hz,  $CH_aH_bOBn$ ) and 1.46 (9H, s,  $OC(CH_3)_3$ ) ppm;  $\delta_C$ (90 MHz,  $CDCl_3$ ) 164.0 (s), 161.2 (s), 156.0 (s), 155.3 (s), 155.1 (s), 143.8 (d), 139.6 (d), 139.3 (d), 137.2 (s), 134.3 (s), 130.7 (s), 129.8 (s), 128.3 (d), 127.8 (d), 127.5 (d), 80.3 (s), 73.2 (t), 70.3 (t), 52.3 (q), 49.4 (d) and 28.2 (q) ppm; m/z (ESI) found: 533.1658,  $C_{25}H_{26}N_4O_8Na$  [(M + Na)<sup>+</sup>] requires 533.1648.

# 2"-[(S)-1-{[2"-((S)-2-Benzyloxy-1-benzyloxycarbonylaminoethyl)-5',5"-dimethyl-[2,4';2',4"]teroxazole-4-carbonyl]-amino}-2-(tert-butyldimethylsilanyloxy)-ethyl]-[2,4';2',4"]teroxazole-4-carboxylic acid methyl ester (27)

-Methylmorpholine (0.11 mL, 1.0 mmol) was added to a stirred suspension of the carboxylic acid **23a** (0.28 g,

0.5 mmol) and 1-hydroxybenzotriazole (0.14 g, 1.0 mmol) in dry dichloromethane (3 mL) at 0 °C under a nitrogen atmosphere. 1-[3-(Dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (0.13 g, 0.65 mmol) was added and the mixture was then stirred at 0°C for 2 min. A solution of the amine **24a** (0.17 g, 0.4 mmol) in dry dichloromethane (4 mL) was added dropwise over 2 min at 0 °C, and the mixture was then allowed to warm to room temperature overnight. The reaction was quenched with water (5 mL) and the separated organic layer was then washed with 10% aqueous citric acid (2  $\times$  5 mL). The combined aqueous extracts were extracted with dichloromethane  $(2 \times 10 \text{ mL})$  and the combined organic extracts were then washed with saturated sodium bicarbonate solution (10 mL), dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica gel using ethyl acetate as eluent to give the amide (0.26 g, 68%) as a colourless solid; mp 170–172 °C (from ethyl acetate);  $[a]_D^{21}$  –32 (c = 1.0, CHCl<sub>3</sub>); found: C, 59.0; H, 5.2; N, 11.3%. C<sub>48</sub>H<sub>50</sub>N<sub>8</sub>O<sub>13</sub>Si requires C, 59.1; H, 5.2; N, 11.5%;  $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$  3412, 2931, 1724, 1674 and 1596;  $\delta_{\rm H}(360~{\rm MHz},~{\rm CDCl_3})~8.45~(1{\rm H},~{\rm s},~{\rm oxazole\text{-}H}),$ 8.36 (1H, s, oxazole-H), 8.35 (1H, s, oxazole-H), 8.33 (1H, s, oxazole-H), 7.89 (1H, d, J 8.7 Hz, CONH), 7.43-7.22 (10H, m,  $10 \times \text{aryl-H}$ ), 5.82 (1H, d, J 8.5 Hz, ZNH), 5.61–5.55 (1H, m, CONHCH), 5.23–5.12 (3H, m, CO<sub>2</sub>CH<sub>2</sub>Ph, ZNHCH), 4.58 (1H, d, J 12.2 Hz, CH<sub>2</sub>OCH<sub>a</sub>H<sub>b</sub>Ph), 4.52 (1H, d, J 12.2 Hz,  $CH_2OCH_aH_bPh$ ), 4.31 (1H, dd, J 10.2 and 3.7 Hz,  $CH_aH_bOTBS$ ), 4.13 (1H, dd, J 10.2 and 4.4 Hz, CH<sub>a</sub>H<sub>b</sub>OTBS), 4.01–3.93 (4H, m,  $CO_2CH_3$ ,  $CH_aH_bOBn$ ), 3.87 (1H, dd, J 9.5 and 4.1 Hz,  $CH_aH_bOBn$ ), 2.82 (3H, s, oxazole- $CH_3$ ), 2.74 (3H, s, oxazole- $CH_3$ ), 0.87 (9H, s,  $SiC(CH_3)_3$ ), 0.08 (3H, s,  $SiCH_3(CH_3)$ ) and 0.02  $(3H, s, SiCH_3(CH_3))$  ppm;  $\delta_C(90 \text{ MHz}, CDCl_3)$  163.6 (s), 161.3 (s), 160.9 (s), 156.0 (s), 155.9 (s), 155.4 (s), 154.8 (s), 150.6 (s), 143.8 (d), 141.0 (d), 139.7 (d), 139.3 (d), 137.3 (s), 136.5 (s), 134.4 (s), 130.8 (s), 130.0 (s), 128.5–127.6 (Ar s and d), 125.5 (s), 124.8 (s), 73.2 (t), 70.2 (t), 67.2 (t), 64.1 (t), 52.3 (q), 49.7 (d), 49.2 (d), 25.6 (q), 18.0 (s), 11.8 (q), 11.8 (q), -5.5 (q) and -5.7 (q) ppm; m/z (ESI) found: 992.3419,  $C_{48}H_{52}N_8O_{14}Si[(M + H_2O)^+]$  requires 992.3372.

#### Macrocycle (28)

4-Methylmorpholine (0.10 mL, 0.9 mmol) was added to a stirred suspension of the amino acid 32b (45 mg, 0.053 mmol) in dry N,Ndimethylformamide (9 mL) at room temperature under a nitrogen atmosphere. Diphenylphosphoryl azide (46 µL, 0.21 mmol) was added dropwise over 2 min and the mixture was stirred at room temperature for 18 h. The mixture was evaporated to dryness in vacuo and the residue was then partitioned between dichloromethane (20 mL) and saturated sodium bicarbonate solution (15 mL). The organic layer was washed with saturated sodium bicarbonate solution (15 mL) and saturated sodium chloride solution (20 mL), then dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue partially purified by recrystallisation from dichloromethane-ether-petrol to give the macrolactam (37 mg, 86%) as a colourless solid;  $\delta_{\rm H}$ (360 MHz, CDCl<sub>3</sub>) 8.65–8.49 (2H, m,  $2 \times CONH$ ), 8.32–8.40 (4H, m,  $4 \times oxazole-H$ ), 7.41–7.00  $(5H, m, 5 \times aryl-H), 5.65-5.30 (2H, m, 2 \times CONHCH), 4.60-4.40$  $(2H, m, CH_2OCH_2Ph), 4.83-4.38 (4H, m, CH_2OTBS, CH_2OBn),$ 2.75 (3H, s, oxazole-C $H_3$ ), 2.65 (3H, s, oxazole-C $H_3$ ), 0.80 (9H, s,  $SiC(CH_3)_3$ ) and 0.13 to -0.13 (6H, m,  $SiCH_3(CH_3)$ ) ppm.

#### Desilylated macrocycle (33a)

Hydrogen chloride (4.0 M solution in dioxane) (5 mL) was added to the ether **28** (37 mg, 0.046 mmol) and the mixture was stirred at room temperature for 10 min under a nitrogen atmosphere. The volatiles were evaporated to leave the *alcohol* (32 mg, 100%) as a colourless solid, which was used directly in the next reaction without further purification;  $\delta_{\rm H}(360~{\rm MHz}, {\rm CDCl_3-CD_3OD}$  (1 : 1)) 8.39–8.00 (4H, m, 4 × oxazole-H), 7.12–6.79 (5H, m, 5 × aryl-H), 5.35–5.10 (2H, m, 2 × CONHCH), 4.40–4.18 (2H, m, CH<sub>2</sub>OCH<sub>2</sub>Ph), 3.99–3.60 (4H, m, CH<sub>2</sub>OH, CH<sub>2</sub>OBn), 2.50 (3H, s, oxazole-CH<sub>3</sub>) and 2.42 (3H, s, oxazole-CH<sub>3</sub>) ppm.

#### Oxazole macrocycle 35a

Dess-Martin periodinane (4 mg, 0.008 mmol) was added in one portion to a stirred solution of the macrocycle 33a (3 mg, 0.004 mmol) in anhydrous dichloromethane (0.5 mL) at room temperature under an atmosphere of nitrogen. The solution was stirred at room temperature for 1.5 h and the volatiles were then removed in vacuo. Triphenylphosphine (5 mg, 0.02 mmol), 1,2-dibromotetrachloroethane (6.5 mg, 0.02 mmol) and di-tertbutyl pyridine (0.01 mL, 0.04 mmol) were added to the residue in dichloromethane (0.1 mL) at 0 °C under an atmosphere of nitrogen. The mixture was stirred and allowed to gradually warm to room temperature overnight. DBU (0.015 mL, 0.1 mmol) was added and the mixture was stirred at room temperature for a further 3.5 h. Water (2 mL) was added and the separated organic layer was washed with 10% aqueous citric acid solution  $(3 \times 10 \text{ mL})$  and sodium bicarbonate solution  $(3 \times 10 \text{ mL})$ , then dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica using dichloromethane to 4% methanol in dichloromethane as eluent to give the enamide (2 mg, 74%) as a colourless solid;  $\delta_{H}(500 \text{ MHz}, \text{CDCl}_3) 9.65 (1 \text{H}, \text{s},$  $NHC=CH_2$ ), 8.58 (1H, d, J 7.4 Hz, NHCH), 8.34 (1H, S, oxazole-H), 8.23 (1H, S, oxazole-H), 8.22 (1H, S, oxazole-H), 8.19 (1H, S, oxazole-H), 5.40-5.45 (1H, m, CHNH), 4.55 (1H, d, J 12.5 Hz, CH<sub>2</sub>OCH<sub>a</sub>H<sub>b</sub>Ph), 4.51 (1H, d, J 12.5 Hz, CH<sub>2</sub>OCH<sub>a</sub>H<sub>b</sub>Ph), 3.99 (1H, dd, J 11.1 and 4.5 Hz, CHCH<sub>a</sub>H<sub>b</sub>OBn), 3.95 (1H, dd, J 11.1 and 5.6 Hz, CHCH<sub>a</sub> $H_b$ OBn), 2.79 (3H, s, oxazole-C $H_3$ ), 2.69 (3H, s, oxazole- $CH_3$ ).

#### Oxazoline macrocycle (34a)

(Diethylamino)sulfur trifluoride (24 µL, 0.18 mmol) was added to a stirred solution of the alcohol 33a (32 mg, 0.046 mmol) in dry dichloromethane (2 mL) at -78 °C under a nitrogen atmosphere. The mixture was stirred at -78 °C for 3 h and was then allowed to warm to room temperature. The mixture was quenched with saturated sodium bicarbonate solution (2 mL) and the separated aqueous layer was then extracted with dichloromethane (2 × 5 mL). The combined organic extracts were dried (MgSO<sub>4</sub>) and then concentrated *in vacuo*. The residue was partially purified by recrystallisation from dichloromethane-ethyl acetate-ether to give the oxazoline (28 mg, 90%) as a colourless solid;  $\delta_{\rm H}$ (360 MHz,  $CDCl_3$ ) 9.29–9.14 (1H, m, CONH), 8.61–8.23 (4H, m, 4 × oxazole-H), 7.34–7.05 (5H, m, 5 × aryl-H), 5.83–5.32 (2H, m, CONHCH, oxazoline-CH), 4.99-4.62 (2H, m, oxazoline-CH<sub>2</sub>), 4.59-4.43 (2H, m,  $CH_2OCH_2Ph$ ), 4.15–3.76 (2H, m,  $CH_2OBn$ ), 2.75 (3H, s, oxazole- $CH_3$ ) and 2.68 (3H, s, oxazole- $CH_3$ ) ppm.

2"-[(S)-2-Benzyloxy-1-({2"-[(S)-1-benzyloxycarbonylamino-2-(tert-butyldimethylsilanyloxy)-ethyl]-5',5"-dimethyl-[2,4';2',4"]-teroxazole-4-carbonyl}-amino)-ethyl]-[2,4';2',4"]teroxazole-4-carboxylic acid methyl ester (36a)

4-Methylmorpholine (0.28 mL, 2.6 mmol) was added to a stirred suspension of the carboxylic acid 23b (0.37 g, 0.64 mmol) and 1-hydroxybenzotriazole (0.26 g, 1.9 mmol) in dry dichloromethane (10 mL) at 0 °C under a nitrogen atmosphere. 1-[3-(Dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (0.25 g, 1.3 mmol) was added and the mixture was then stirred at 0 °C for 10 min. The amine 24b (0.35 g, 0.78 mmol) was added in one portion, and the mixture was then allowed to warm to room temperature overnight. Dichloromethane (50 mL) was added and the reaction was quenched with water (30 mL). The separated organic layer was washed with 10% aqueous citric acid  $(2 \times 30 \text{ mL})$  and saturated sodium bicarbonate solution (30 mL), then dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica gel using dichloromethaneethyl acetate (2:1) eluent to give the amide (0.54 g, 87%) as a colourless solid; mp 204–205 °C;  $[a]_D^{21}$  +14 (c = 1.0, CHCl<sub>3</sub>); found: C, 58.6; H, 5.1; N, 11.3%. C<sub>48</sub>H<sub>50</sub>N<sub>8</sub>O<sub>13</sub>Si requires C, 59.1; H, 5.2; N, 11.5%;  $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$  3412, 2930, 1723, 1674 and 1595;  $\delta_{\rm H}$ (360 MHz, CDCl<sub>3</sub>) 8.43 (1H, s, oxazole-H), 8.34 (1H, s, oxazole-H), 8.33 (1H, s, oxazole-H), 8.30 (1H, s, oxazole-H), 7.86 (1H, d, J 8.8 Hz, CONH), 7.42–7.22 (10H, m,  $10 \times \text{aryl-H}$ ), 5.77 (1H, d, J 8.5 Hz, ZNH), 5.71–5.64 (1H, m, CONHCH), 5.18 (1H, m, CO<sub>2</sub>CH<sub>a</sub>H<sub>b</sub>Ph), 5.13 (1H, d, J 12.3 Hz, CO<sub>2</sub>CH<sub>a</sub>H<sub>b</sub>Ph), 5.11-5.03 (1H, m, ZNHCH), 4.60 (2H, s, CH<sub>2</sub>OCH<sub>2</sub>Ph), 4.17–3.93 (4H, m, CH<sub>2</sub>OTBS, CH<sub>2</sub>OBn), 3.96 (3H, s, CO<sub>2</sub>CH<sub>3</sub>), 2.79 (3H, s, oxazole- $CH_3$ ), 2.72 (3H, s, oxazole- $CH_3$ ), 0.81 (9H, s,  $SiC(CH_3)_3$ ), -0.01 (3H, s, SiCH<sub>3</sub>(CH<sub>3</sub>)) and -0.04 (3H, s, SiCH<sub>3</sub>(CH<sub>3</sub>)) ppm;  $\delta_{\rm C}(90 \text{ MHz}, {\rm CDCl_3}) 163.0 \text{ (s)}, 161.1 \text{ (s)}, 161.1 \text{ (s)}, 160.1 \text{ (s)}, 155.8$ (s), 155.8 (s), 155.7 (s), 155.2 (s), 154.8 (s), 150.7 (s), 150.6 (s), 143.8 (d), 141.0 (d), 139.7 (d), 139.2 (d), 137.1 (s), 136.3 (s), 136.0 (s), 134.2 (s), 130.6 (s), 129.8 (s), 128.3 (d), 128.3 (d), 128.0 (d), 128.0 (d), 127.7 (d), 127.5 (d), 125.3 (s), 124.5 (s), 73.1 (t), 69.6 (t), 67.0 (t), 64.3 (t), 52.2 (q), 51.4 (d), 47.2 (d), 25.5 (q), 17.9 (s), 11.7 (q), 11.6 (q), -5.7 (q) and -5.8 (q) ppm; m/z (ESI) found: 997.3126,  $C_{48}H_{50}N_8O_{13}SiNa$  [(M + Na)<sup>+</sup>] requires 997.3164.

#### Macrocycle (37)

4-Methylmorpholine (0.44 mL, 3.8 mmol) was added to a stirred suspension of the amino acid 36b (0.17 g, 0.19 mmol) in dry dichloromethane (52 mL) and dry N,N-dimethylformamide (26 mL) at 0 °C under a nitrogen atmosphere. O-(7-Azabenzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate (0.15 g, 0.39 mmol) was added and the mixture was stirred at 0 °C for 30 min and then allowed to warm to room temperature and stirred for 18 h. Dichloromethane (60 mL) was added and the reaction was quenched with water (60 mL). The separated organic layer was washed with water (60 mL) and saturated sodium chloride solution (60 mL), then dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue partially purified by trituration with ether to give the macrolactam (0.12 mg, 80%) as a colourless solid;  $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$  3384, 2928, 1671 and 1597;  $\delta_{\rm H}(500\,{\rm MHz},{\rm CDCl_3})\,8.32\,(1{\rm H},{\rm s},{\rm oxazole-H}),\,8.30\,(1{\rm H},{\rm s},{\rm oxazole-H})$ H), 8.26 (1H, s, oxazole-H), 8.26 (1H, s, oxazole-H), 7.95 (1H, d,

J 8.8 Hz, CONH), 7.89 (1H, d, J 8.7 Hz, CONH), 7.37–7.26 (5H, m, 5 × aryl-H), 5.78-5.71 (1H, m, CONHCH), 5.62-5.55 (1H, m, CONHCH), 4.62 (1H, d, J 12.3 Hz, CH<sub>2</sub>OCH<sub>a</sub>H<sub>b</sub>Ph),4.58 (1H, d, J 12.3 Hz, CH<sub>2</sub>OCH<sub>a</sub>H<sub>b</sub>Ph), 4.30–3.93 (4H, m,  $CH_2OTBS$ ,  $CH_2OBn$ ), 2.81 (3H, s, oxazole- $CH_3$ ), 2.74 (3H, s, oxazole- $CH_3$ ), 0.90 (9H, s,  $SiC(CH_3)_3$ ), -0.08 (3H, s,  $SiCH_3(CH_3)$ ) and -0.03 (3H, s, SiCH<sub>3</sub>(CH<sub>3</sub>)) ppm; m/z (ESI) found: 831.2527,  $C_{39}H_{40}N_8O_{10}SiNa$  [(M + Na)<sup>+</sup>] requires 831.2534.

#### $2-[(S)-1-\{[2''-(tert-Butoxycarbonylaminomethyl)-[2,4';2',4'']$ teroxazole-4-carbonyl|-amino}-2-(tert-butyldimethylsilanyloxy)ethyl|-5-phenyloxazole-4-carboxylic acid methyl ester (64)

4-Methylmorpholine (0.77 mL, 7.0 mmol) was added to a stirred suspension of the acid 63b (0.26 g, 0.70 mmol) and 1hydroxybenzotriazole (0.47 g, 3.5 mmol) in dry dichloromethane (10 mL) and N,N-dimethylformamide (5 mL) at 0 °C under a nitrogen atmosphere. 1-[3-(Dimethylamino)propyl]-3ethylcarbodiimide hydrochloride (0.27 g, 1.4 mmol) was added and the mixture was then stirred at 0 °C for 10 min. A solution of the amine 60 (0.28 g, 0.74 mmol) in dry dichloromethane (5 mL) was added dropwise over 3 min at 0 °C, and the mixture was allowed to warm to room temperature overnight and then quenched with water (20 mL). Dichloromethane (30 mL) was added and the separated organic layer was washed with 10% aqueous citric acid ( $2 \times 15$  mL) and saturated sodium bicarbonate solution (15 mL), then dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by chromatography on silica gel using 1:1 dichloromethane-ethyl acetate as eluent to give the tetraoxazole amide (0.45 g, 87%) as a colourless solid; mp 96–98 °C (from dichloromethane–petrol);  $[a]_D^{27}$  +54 (c = 1.0, CHCl<sub>3</sub>); found: C, 56.9; H, 5.8; N, 11.1%. C<sub>35</sub>H<sub>42</sub>N<sub>6</sub>O<sub>10</sub>Si requires C, 57.2; H, 5.8; N, 11.4%;  $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$  3696, 3605, 3506, 2929, 1719, 1680 and 1595;  $\delta_{\rm H}$ (360 MHz, CDCl<sub>3</sub>) 8.35 (1H, s, oxazole-H), 8.32 (1H, s, oxazole-H), 8.30 (1H, s, oxazole-H), 8.04–8.00 (2H, m, 2 × phenyloxazole-H), 7.83 (1H, d, J 8.7 Hz, CONH), 7.49–7.43 (3H, m, 3 × phenyloxazole-H), 5.59-5.52 (1H, m, CONHCH), 5.27 (1H, br s, BocNH), 4.56 (2H, d, J 5.8 Hz, BocNHCH<sub>2</sub>), 4.23 (1H, dd, J 10.1 and 4.6 Hz, CH<sub>a</sub>H<sub>b</sub>OTBS), 4.10 (1H, dd, J 10.1 and 5.2 Hz, CH<sub>a</sub>H<sub>b</sub>OTBS), 3.94 (3H, s, CO<sub>2</sub>CH<sub>3</sub>), 1.47 (9H, s,  $OC(CH_3)_3$ , 0.84 (9H, s,  $SiC(CH_3)_3$ ), 0.04 (3H, s,  $SiCH_3(CH_3)$ ) and 0.03 (3H, s, SiCH<sub>3</sub>(CH<sub>3</sub>)) ppm;  $\delta_{\rm C}$ (90 MHz, CDCl<sub>3</sub>) 162.8 (s), 162.3 (s), 160.1 (s), 159.8 (s), 156.0 (s), 155.7 (s), 155.4 (s), 154.3 (d), 141.5 (d), 139.7 (d), 139.0 (d), 136.7 (s), 130.8 (s), 130.3 (d), 129.6 (s), 128.3 (d), 128.2 (d), 126.7 (s), 126.5 (s), 80.2 (s), 64.1 (t), 52.1 (q), 49.1 (d), 37.8 (t), 28.1 (q), 25.6 (q), 18.1 (s), -5.6 (q) and -5.6 (q) ppm; m/z (ESI) found: 757.2639,  $C_{35}H_{42}N_6O_{10}SiNa$  $[(M + Na)^{+}]$  requires 757.2629.

#### 2-[(S)-1-{[2"-(tert-Butoxycarbonylaminomethyl)-[2,4';2',4"]teroxazole-4-carbothioyl|-amino}-2-(tert-butyldimethylsilanyloxy)ethyl]-5-phenyloxazole-4-carboxylic acid methyl ester (65)

Lawesson's reagent (0.45 g, 1.1 mmol) was added in one portion to a stirred solution of the amide 64 (0.45 g, 0.61 mmol) in dry tetrahydrofuran (10 mL) at room temperature under a nitrogen atmosphere. The mixture was stirred at reflux for 21 h and then cooled to room temperature. The mixture was concentrated in vacuo and the residue was then partitioned between dichloromethane (80 mL) and saturated sodium bicarbonate solution (80 mL). The aqueous extract was re-extracted with dichloromethane (80 mL) and the combined organic extracts were dried (MgSO<sub>4</sub>) and then concentrated in vacuo. The solid yellow residue was purified by chromatography on silica gel using 6:1 dichloromethane-ether as eluent to give the thioamide (0.23 g, 50%) as a colourless solid; mp 151 °C (decomp.) (from dichloromethane–petrol);  $[a]_D^{22}$  +74  $(c = 1.0, \text{CHCl}_3); \nu_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1} 3449, 3345, 2930, 1719, 1653$ and 1585;  $\delta_{H}$ (360 MHz, CDCl<sub>3</sub>) 9.46 (1H, d, J 8.1 Hz, CSNH), 8.43 (1H, s, oxazole-H), 8.34 (1H, s, oxazole-H), 8.32 (1H, s, oxazole-H), 8.03-7.99 (2H, m, 2 × phenyloxazole-H), 7.48-7.42  $(3H, m, 3 \times phenyloxazole-H), 6.14-6.09 (1H, m, CSNHCH),$ 5.30 (1H, br s, BocNH), 4.55 (2H, d, J 5.9 Hz, BocNHCH<sub>2</sub>), 4.33 (1H, dd, J 10.2 and 4.1 Hz, CH<sub>a</sub>H<sub>b</sub>OTBS), 4.19 (1H, dd, J 10.2 and 5.0 Hz, CH<sub>a</sub>H<sub>b</sub>OTBS), 3.93 (3H, s, CO<sub>2</sub>CH<sub>3</sub>), 1.46 (9H, s,  $OC(CH_3)_3$ , 0.86 (9H, s,  $SiC(CH_3)_3$ ), 0.06 (3H, s,  $SiCH_3(CH_3)$ ) and 0.03 (3H, s, SiCH<sub>3</sub>(CH<sub>3</sub>)) ppm;  $\delta_{\rm C}$ (90 MHz, CDCl<sub>3</sub>) 185.3 (s), 162.8 (s), 162.3 (s), 159.3 (s), 156.1 (s), 155.9 (s), 154.1 (s), 142.8 (d), 142.0 (s), 139.7 (d), 139.1 (d), 130.9 (s), 130.4 (d), 129.7 (s), 128.4 (d), 128.4 (d), 126.9 (s), 126.6 (s), 80.4 (s), 63.4 (t), 53.7 (d), 52.3 (q), 37.9 (t), 28.2 (q), 25.6 (q), 18.1 (s) and -5.5 (q) ppm; m/z(ESI) found: 773.2461,  $C_{35}H_{42}N_6O_9SSiNa$  [(M + Na)+] requires 773.2401.

#### 4-[(S)-2-Hydroxy-1-(4-methoxycarbonyl-5-phenyloxazol-2-yl)ethylthiocarbamoyl]-[2,4';2',4"]teroxazol-2"-ylmethylammonium chloride (70)

Hydrogen chloride (4.0 M solution in dioxane) (3 mL) was added to the carbamate 65 (0.23 g, 0.3 mmol) and the mixture was stirred at room temperature overnight under a nitrogen atmosphere. The mixture was evaporated to leave the amine hydrochloride salt (0.16 g, 91%) as a colourless solid; mp 191 °C (decomp.);  $[a]_D^{26}$  +80  $(c = 0.2, CH_2Cl_2-MeOH (3:1)); \nu_{max}(solid)/cm^{-1} 3350, 1727, 1659$ and 1581;  $\delta_{\rm H}(500 \text{ MHz}, \text{DMSO-}d_6)$  10.33 (1H, br s, CSNH), 9.18 (1H, s, oxazole-H), 9.15 (1H, s, oxazole-H), 8.94 (1H, s, oxazole-H), 8.92 (3H, br s, N $H_3$ ), 7.99–7.95 (2H, m, 2 × phenyloxazole-H), 7.58-7.52 (3H, m, 3 × phenyloxazole-H), 5.97 (1H, br s, OH), 5.59(1H, t, J 5.9 Hz, CSNHCH), 4.41 (2H, s, NH<sub>3</sub>CH<sub>2</sub>), 4.15–4.07 (2H, m, C $H_2$ OH) and 3.82 (3H, s, CO<sub>2</sub>C $H_3$ ) ppm;  $\delta_c$ (125 MHz, DMSO-*d*<sub>6</sub>) 186.6 (s), 165.4 (s), 163.0 (s), 162.9 (s), 158.8 (s), 158.2 (s), 157.7 (s), 147.7 (d), 145.5 (d), 145.1 (s), 144.9 (d), 134.0 (d), 133.4 (s), 132.6 (s), 132.1 (d), 131.7 (d), 130.0 (s), 129.9 (s), 64.5 (t), 58.0 (d), 55.5 (q) and 38.9 (t) ppm; *m/z* (ESI) found: 537.1161,  $C_{24}H_{20}N_6O_7S$  [M<sup>+</sup>] requires 537.1192.

#### $2 - \{(S) - 1 - [(2'' - \{[(R) - 1 - ((2S, 3S) - 2 - tert - Butoxycarbonylamino - 3 - ((2S, 3S) - 2 - tert - Butoxycarbonylamino - ((2S, 3S) - ((2S, 3S) - 2 - tert - Butoxycarbonylamino - ((2S, 3S) - 2 - tert - Butoxycarbonylamino$ methylpentanoylamino)-2-methylpropylamino|-methyl}-[2,4';2',4"]teroxazole-4-carbothioyl)-amino[-2-hydroxyethyl}-5phenyloxazole-4-carboxylic acid methyl ester (78a)

4-Methylmorpholine (59 μL, 0.54 mmol) was added to a stirred suspension of the carboxylic acid 77 (90 mg, 0.27 mmol) and 1-hydroxybenzotriazole (0.11 g, 0.81 mmol) in dry dichloromethane (15 mL) at 0 °C under a nitrogen atmosphere. 1-[3-(Dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (0.10 g, 0.54 mmol) was added and the mixture was then stirred at 0 °C for 15 min. A pre-cooled (0 °C) solution of the amine 70 (0.16 g, 0.27 mmol) and 4-methylmorpholine (59 µL, 0.54 mmol) in dry dichloromethane (5 mL) was added dropwise over 5 min, and the mixture was then allowed to warm to room temperature overnight. The mixture was concentrated in vacuo and the residue was then triturated with methanol to leave the amide (0.19 g, 82%) as a colourless solid; mp 258 °C (decomp.);  $[a]_D^{25}$  +56  $(c = 1.0, \text{CHCl}_3\text{-MeOH } (4 : 1)); \nu_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1} 3343, 2964)$ and 1692;  $\delta_{\rm H}(360~{\rm MHz},~{\rm DMSO-}d_6)$  10.34 (1H, d, J 7.8 Hz, CSNH), 9.08 (1H, s, oxazole-H), 8.96 (1H, s, oxazole-H), 8.91 (1H, s, oxazole-H), 8.70 (1H, t, CONHCH<sub>2</sub>), 8.00–7.91 (3H, m, 2  $\times$  phenyloxazole-H, CON*H*-(val)), 7.58–7.49 (3H, m, 3  $\times$ phenyloxazole-H), 6.87 (1H, d, J 8.1 Hz, BocNH), 6.01–5.94 (1H, m, CONHCH-(val)), 5.52 (1H, t, J 5.9 Hz, CSNHCH), 4.59-4.43 (2H, m, gly-CH<sub>2</sub>), 4.29–4.21 (1H, m, BocNHCH), 4.18–4.07 (2H, m, CH<sub>2</sub>OH), 3.92 (1H, t, J 7.9 Hz, CH<sub>2</sub>OH), 3.82 (3H, s,  $CO_2CH_3$ ), 2.14–2.02 (1H, m, CH), 1.76–1.63 (1H, m, CH), 1.37  $(9H, s, OC(CH_3)_3)$ , 1.16–1.03 (2H, m,  $CH_2$ ) and 0.91–0.75 (12H, m,  $3 \times CH_3$ ) ppm;  $\delta_C(90 \text{ MHz}, DMSO-d_6)$  188.6 (s), 175.4 (s), 174.9 (s), 174.8 (s), 166.1 (s), 165.4 (s), 163.0 (s), 159.2 (s), 158.1 (s), 157.8 (s), 147.6 (d), 145.1 (s), 144.6 (d), 144.6 (d), 134.0 (d), 133.3 (s), 132.4 (s), 132.1 (d), 131.7 (d), 129.9 (s), 129.9 (s), 81.7 (s), 64.5 (t), 62.5 (d), 60.9 (d), 58.0 (d), 55.4 (q), 39.6 (d), 39.4 (t), 33.5 (d), 31.6 (q), 27.9 (t), 22.7 (q), 21.2 (q), 18.9 (q) and 14.4 (q) ppm; m/z (ESI) found: 871.3130,  $C_{40}H_{48}N_8O_{11}SNa$  [(M + Na)<sup>+</sup>] requires 871.3061.

# $(1S,2S)-1-[(R)-1-(\{4-[(S)-1-(4-Carboxy-5-phenyloxazol-2-yl)-2-hydroxyethylthiocarbamoyl]-[2,4';2',4''] teroxazol-2''-ylmethyl\}-carbamoyl)-2-methylpropylcarbamoyl]-2-methylbutylammonium chloride (78b)$

A solution of sodium hydroxide (87 mg, 2.2 mmol) in water (5 mL) was added in one portion to a stirred solution of the tetra-oxazole substituted methyl ester **78a** (0.19 g, 0.22 mmol) in tetrahydrofuran (10 mL), and the mixture was stirred at room temperature overnight. Water (20 mL) was added and the mixture was concentrated slowly *in vacuo* to a volume of 20 mL. The aqueous suspension was extracted with dichloromethane—methanol (4:1) (5 × 30 mL) and dichloromethane (3 × 30 mL) and the combined organic extracts were then evaporated *in vacuo* to leave the *sodium carboxylate* (0.17 g, 86%) as a colourless solid, which was used directly in the next reaction without further purification.

Hydrogen chloride (4.0 M solution in dioxane) (5 mL) was added to the *sodium carboxylate* (0.17 g, 0.19 mmol) and the mixture was stirred at room temperature overnight under a nitrogen atmosphere. The volatiles were evaporated to leave the  $\omega$ -amino acid hydrochloride salt (0.14 g, 100%) as a colourless solid, which was used without further purification.

#### (-)-YM-216391 (81)

4-Methylmorpholine (0.28 mL, 2.6 mmol) was added to a stirred suspension of the ω-amino acid (0.14 g, 0.19 mmol) in dry dichloromethane (42 mL) and dry N,N-dimethylformamide (21 mL) at 0 °C under a nitrogen atmosphere. O-(7-Azabenzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate (0.15 g, 0.38 mmol) was added and the mixture was stirred at 0 °C for 5 min and then allowed to warm to room temperature and stirred for 48 h. Dichloromethane (50 mL) was added and the reaction was quenched with water (50 mL). The

separated organic layer was washed with 10% aqueous citric acid (40 mL) and saturated sodium bicarbonate solution (40 mL), then dried (MgSO<sub>4</sub>) and concentrated *in vacuo*. The residue was partially purified by trituration with ether to give the *macrolactam* **79** (95 mg, 75%) as a colourless solid.

(Diethylamino)sulfur trifluoride (35  $\mu$ L, 0.27 mmol) was added to a stirred solution of the thioamide cyclopeptide **79** (95 mg, 0.13 mmol) in dry dichloromethane (3 mL) at -78 °C under a nitrogen atmosphere. The mixture was stirred at -78 °C for 4 h and then allowed to warm to room temperature. The mixture was quenched with saturated sodium bicarbonate solution (3 mL) and the separated aqueous layer was then extracted with dichloromethane (5 mL). The combined organic extracts were dried (MgSO<sub>4</sub>) and then concentrated *in vacuo* to leave the corresponding thiazoline **80** (82 mg, 92%) as a yellow solid, which was used directly in the next reaction without further purification.

Activated manganese(IV) oxide (0.2 g, 2.3 mmol) was added to a stirred solution of the thiazoline 80 (82 mg, 0.12 mmol) in dry dichloromethane (10 mL) at room temperature under a nitrogen atmosphere. The mixture was stirred for 48 h and was then filtered through a pad of celite and eluted with dichloromethanemethanol (1:1) (100 mL). The filtrate was concentrated in vacuo to leave a yellow residue, which was partially purified by trituration with ether to leave the thiazole (55 mg, 66%) as a colourless solid. A portion of this material was further purified by reverse phase HPLC to give the thiazole (10 mg) as a colourless solid; mp 240 °C (decomp.);  $[a]_D^{20}$  –56 (c = 0.50, CHCl<sub>3</sub>);  $\delta_H$ (500 MHz, DMSO- $d_6$ ) 9.12 (1H, s), 9.02 (1H, s), 8.93 (1H, s), 8.71 (1H, dd, J 9.1 and 2.5 Hz), 8.68 (1H, s), 8.58 (1H, d, J 9.0 Hz), 8.39 (2H, d, J 7.3 Hz), 8.26 (1H, d, J 7.2 Hz), 7.61–7.47 (3H, m), 5.04 (1H, dd, J 16.6 and 9.1 Hz), 4.78 (1H, dd, J 7.2 and 4.4 Hz), 4.61 (1H, dd, J 9.0 and 4.6 Hz), 4.22 (1H, d, J 16.6 Hz), 2.18–2.07 (2H, m), 1.73–1.64 (1H, m), 1.18–1.09 (1H, m) and 0.98–0.76 (12H, m) ppm;  $\delta_c$  (125 MHz, DMSO-*d*<sub>6</sub>) 171.3, 170.3, 163.5, 160.6, 158.0, 155.9, 155.5, 154.6, 151.3, 141.5, 140.2, 140.0, 139.8, 136.1, 131.2, 130.6, 130.4, 129.6,  $129.1(\times 2)$ ,  $127.9(\times 2)$ , 127.2, 122.9, 58.2, 57.8, 38.5, 35.6, 32.1, 25.3, 20.3, 17.9, 15.5 and 12.4 ppm; m/z (ESI) found: 719.2043,  $C_{34}H_{32}N_8O_7SNa$  [(M + Na)<sup>+</sup>] requires 719.2012.

#### Acknowledgements

We thank the EPSRC for a studentship (to JD) and Drs K. Sohda and Y. Takebayashi (Yamanouchi Pharmaceutical Co. Ltd. Tokyo, Japan) for helpful correspondence and for supplying relevant NMR spectroscopic data on natural YM-216391. We also thank Celia Ribes Vidal and David Evans for their contributions to these projects.

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