Utilization of Cyclic Amides as Masked Aldehyde Equivalents in **Reductive Amination Reactions**

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Supporting Information

ABSTRACT: An operationally simple protocol has been discovered that couples primary or secondary amines with Naryl-substituted lactams to deliver differentiated diamines in moderate to high yields. The process allows for the partial reduction of a lactam in the presence of Cp2ZrHCl (Schwartz's reagent), followed by a reductive amination between the resulting hemiaminal and primary or secondary amine. These reactions can be telescoped in a one-pot fashion to significantly simplify the operation. The scope of amines and substituted lactams of various ring sizes was demonstrated



through the formation of a range of differentiated diamine products. Furthermore, this methodology was expanded to include N-aryl pyrrolidinone substrates with an enantiopure ester group at the 5-position, and α -amino piperidinones were prepared with complete retention of stereochemical information. The development of this chemistry has enabled the consideration of lactams as useful synthons.

INTRODUCTION

The propensity of amide groups to participate in resonance stabilization is the fundamental attribute which affords them uncharacteristic durability, as compared to other members of the carboxylic acid family.¹ As such, amide bonds are incredibly robust and require forcing conditions to facilitate their hydrolysis.² The implementation and commercialization within the pharmaceutical industry³ of this resilient chemotype and their applications in the synthesis of high strength materials⁴ serve as testaments to the strength and utility of these bonds.

Due to their sturdiness, utilization of amides as enabling precursors for functional group manipulation in synthetic organic chemistry has been limited in scope, focusing primarily on complete reductions to afford amines.⁵ Recent advances in the use of increasingly mild reducing conditions, and even chemoselective reducing conditions, have expanded the application of reductions of amides.⁶ Additionally, imines, enamines, alcohols, and aldehydes compose a significant minority of amide reduction products.' Even though the aforementioned array of functionalities could be accessed via amide reduction reactions, these transformations require specialized reagents that lack compatibility with a wide range of other functional groups.⁸ Due to this limitation, alternative synthetic strategies are often more desirable for the synthesis of these functional groups over direct amide reductions. Furthermore, syntheses of diamines directly from amides are under-represented in the literature and in all examples, multistep sequences are necessary (vide infra).

However, if a partial reduction product of an amide, for instance, an aminol or other masked aldehyde equivalent, could be readily further functionalized in a coupling reaction, such a protocol would be synthetically attractive, effectively rendering the amide group a useful synthon for further manipulations. Indeed, lactams have been reported to be utilized as starting materials to build molecular complexity.⁵ Substituted N-Boc pyrrolidinones (functionally imides) have been treated with a variety of reducing agents (generally DIBAL-H, LAH, or LiEt₃BH) to afford isolable aminol intermediates, which have been used effectively in a variety of subsequent transformations (Scheme 1, top).^{10,11}

Due to the lack of generality of reaction conditions and the sensitive or incompatible reducing agents employed in prior works, the utilization of an in situ-prepared reactive intermediate of a lactam reduction for further functionalization has not been a widely adopted synthetic strategy, specifically to access differentiated diamines. Herein, we report an investigation of lactams as masked aldehyde equivalents, generated by the treatment of an N-aryl lactam with a chemoselective reducing agent (Cp₂ZrHCl, Schwartz's reagent¹²) under mild conditions $(0 \,^{\circ}C)$ and its subsequent in situ functionalization through a reductive amination sequence. We have telescoped the reaction conditions to enable an operationally simple, onepot procedure to produce a set of differentiated diamines (Scheme 1, bottom). The rationale behind the choice of Narylated lactams as substrates in this study is 2-fold. First, these

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Scheme 1. Original Scope of Research Project



Table 1. Screen of Conditions in Optimizing the One-Pot Coupling Reaction^a

	$ \begin{array}{c} 0 \\ N \\ \hline 0 \\ \hline \hline \hline \hline 0 \\ \hline \hline \hline \hline 0 \\ \hline \hline$	X equiv. reducing agent	X equiv. educing agent MeO Y equiv.		p-OMePh N → NHp-CF ₃ Ph H 3a	
		THF, T ₁ °C, 1 h	2 equiv. NaBH(OAc) ₃ , T ₂ °C, 18 h	N-p-	CF ₃ Ph 4a	
entry	reducing agent (X)	amine (Y)	T_1 (°C)	T_2 (°C)	Conv. (%) ^b	Yield: 3a ; 4a ^{<i>c</i>}
1	DIBAL-H; 1.0 equiv	1.0 equiv	23	23	73	<5%; 30%
2	Cp ₂ ZrHCl; 1.0 equiv	1.0 equiv	23	23	67	15%; 30%
3	Cp ₂ ZrHCl; 2.0 equiv	1.0 equiv	23	23	>98	15%; 30%
4	Cp ₂ ZrHCl; 2.0 equiv	2.0 equiv	23	23	>98	45%; 30%
5	Cp ₂ ZrHCl; 2.0 equiv	3.0 equiv	23	23	>98	43%; 24%
6	Cp ₂ ZrHCl; 2.0 equiv	4.0 equiv	23	23	>98	55%; 15%
7	Cp ₂ ZrHCl; 2.0 equiv	4.0 equiv	23	60	>98	60%; 18%
8	Cp ₂ ZrHCl; 1.5 equiv	4.0 equiv	0	23	>98	63% ^d ; <5%

^{*a*}Average of two runs for each entry. ^{*b*}Conversion refers to the disappearance of lactam starting material. ^{*c*}Determined by liquid chromatography mass spectroscopy (LCMS). ^{*d*}Yield by silica gel chromatography.

lactams can be readily accessed via copper-catalyzed Ullmann couplings between commercially available cyclic carboxamides and various aryl iodides. Second, reactions with these *N*-aryl lactams afford differentiated diamines featuring a versatile aryl amine functionality. Other synthetic strategies for nonvicinal diamines (specifically 1,3-,¹³ 1,4-,¹⁴ and 1,5-diamines) involved multiple steps to selectively functionalize one amine at a time. Additionally, these diamine products could not be selectively formed from the previously disclosed Boc-protected amines through palladium-catalyzed C–N coupling chemistry because of the competing secondary amine present in those molecules.¹⁵

RESULTS AND DISCUSSION

We began our investigation by identifying conditions which would allow both the reduction of the lactam and the reduction of the imine to be carried out in one pot. Our optimization results are summarized in Table 1. We initially evaluated a variety of reducing agents but only saw productive reactions when using DIBAL-H or Schwartz's reagent.¹⁶ Combinations of DIBAL-H or Schwartz's reagent with aluminum or zirconium Lewis acid additives¹⁷ to activate the lactam carbonyl for a more facile reduction afforded no improvement in reactivity or product distribution. When the reducing reagent was omitted in either step of the sequence, the corresponding reactions did not lead to any desired diamine product. Our initial solvent screen showed that tetrahydrofuran (THF) was the most effective solvent¹⁸ for both the partial reduction of 1a and the following utilization of the resulting aldehyde equivalent in a reductive amination reaction with 2a. Therefore, N-aryl-substituted lactam 1a was treated with DIBAL-H in THF and a minimal amount of desired diamine 3a was formed, with mostly over-reduced pyrrolidine 4a identified in the crude mixture (entry 1, Table 1). Switching from DIBAL-H to Schwartz's reagent (Cp₂ZrHCl) in this reductive coupling reaction delivered 15% yield of 3a and 30% 4a with 33% of the remaining lactam 1a (entry 2, Table 1). Subsequently, we observed full consumption of 1a when 2.0 equiv of Cp₂ZrHCl was used but did not note an improvement in yield of desired product 3a (entry 3, Table 1). We surmised that the amine might be

coordinating to the metal center which would render it unavailable to participate in the reductive amination reaction to produce 3a. Hence, we increased the equivalents of amine to 2.0 equiv and obtained 45% yield of diamine 3a with 30% of pyrrolidine 4a (entry 4, Table 1). A comparable yield of diamine 3a and a slight reduction in yield of pyrrolidine 4a were observed when the 3.0 equiv. of amine was investigated (entry 5, Table 1). Further increase to 4.0 equiv. of 2a led to 55% yield of 3a in the reaction mixture and a decreased amount of pyrrolidine 4a (15%, entry 6, Table 1). However, no boost in yield of 3a was noted when more than four equivalents of amine were utilized. Later optimization of reaction temperatures for both sequential steps identified that performing the partial reduction of lactam 1a at 0 °C followed by the standard reductive amination at ambient temperature completely consumed lactam 1a, affording diamine 3a in 63% yield and <5% byproduct 4a (entry 8, Table 1).

With the optimized conditions in hand, we set out to explore the scope of N-aryl-substituted lactam substrates (Table 2). As shown from entries 1-4, para-substituted electron-deficient aromatic substituents on the lactams were capable substrates for this chemistry, giving desired diamines in 35-82% yield with a minimal amount of pyrrolidine byproducts. Electronwithdrawing functional groups on the aryl ring, such as cyano, nitro, and carboxylic ester, were tolerated in this reaction; diamines 3e, 3f, and 3g were obtained in 34-49% yield. However, pyrrolidines 4f and 4g were afforded in 40 and 35%, respectively (entries 6 and 7, Table 2). Moreover, as demonstrated in entry 8 of Table 2, the reductive coupling reaction of disubstituted arene containing lactam **1h** resulted in the formation of diamine 3h in 42% yield and 37% pyrrolidine 4h. An electron neutral phenyl substituent was also tolerated on the pyrrolidinone substrate. Diamine 3i was obtained in 36% yield along with 32% fully reduced pyrrolidine 4i, as shown in entry 9 of Table 2. On the contrary, electrondonating *para*-methoxy substituent containing substrate 1j decomposed to complex reaction mixtures with <5% desired diamines detected by LCMS (entry 10, Table 2). ortho-Methoxyphenyl-substituted lactam 1k, which was presumably less electron donating due to the inductive effect, was examined in hopes of restoring a certain extent of reaction efficiency; however, 1k was proven to be an ineffective coupling partner as well, the reaction with which only led to the formation of pyrrolidine 4k in 35% yield (entry 11).

Next, we investigated steric effects surrounding the lactam functionality. As illustrated in entries 1 and 2 in Table 3, when a methyl group was placed at the 3- or 5-position on the lactam ring, reductive coupling reactions of lactams 11 and 1m delivered methyl-substituted diamines 31 and 3m in 43 and 34% yields, respectively. Unlike the result observed in entry 1 (Table 2), pyrrolidines 4l and 4m were also afforded in high yield (35-45%). In addition to pyrrolidinones, six- and fourmembered N-arylated lactams were examined as well. Upon treatment with the standard reaction conditions, piperidinone 1n was transformed into diamine 3n with a five-carbon linker in between the two nitrogen centers in 72% yield (entry 3, Table 3). Strained azetidinones were more capable coupling partners than either the pyrrolidinones or piperidinones in this sequence of reactions. As shown in entry 4, diamine 30 was obtained in 92% yield. Interestingly, azetidinones with electron-rich aromatic substituents, entries 5 and 6 (Table 3), were effectively converted into diamines 3p and 3q in 30 and 48% yield, respectively. Although some azetidines 4p and



Table 2. Scope of Aryl-substituted Lactams in One-PotReductive Coupling a

^{*a*}Average of at least two runs for each entry. >98% conversion in all cases unless noted otherwise. ^{*b*}95% conversion. ^{*c*}90% conversion. ^{*d*}85% conversion. ^{*c*}Complicated reaction mixtures observed. ^{*f*}Yield by silica gel chromatography. ^{*g*}Determined by liquid chromatography mass spectroscopy (LCMS).

4q accompanied the desired products in these reactions, the results were still very encouraging given that electron-rich arene containing pyrrolidinones were ineffective substrates.

Having examined the scope of the *N*-aryl-substituted lactams, we set forth to investigate the range of amines that were able to participate in this class of reductive coupling reactions. As illustrated in Table 4, a variety of primary amines were tested in combination with the substrate 1a. Cyclohexylmethylamine reacted with 1a to furnish diamine 5a in 69% yield with <5% of pyrrolidine 4a present in the crude mixture. Similarly, reductive coupling reactions with cyclohexylamine and nonylamine afforded 5b and 5c in 58 and 63% yield, respectively. Amines with an α -stereogenic center were also tolerated in these reactions, as represented by the formation of diamines 5d and 5e (61 and 50% yield, Table 4). Reductive coupling of 1a with *p*-CF₃ benzylamine 2f resulted in a lower yield of diamine 5f; pyrrolidine 4a, in this

Table 3. Additional Scope of Aryl-substituted Lactams in One-Pot Reductive Coupling a



^{*a*}Average of at least two runs for each entry; Ar = p-OMePh; >98% conversion in all cases unless otherwise noted. ^{*b*}70% conversion. ^{*c*}87% conversion. ^{*d*}Yield by silica gel chromatography. ^{*c*}Determined by liquid chromatography mass spectroscopy (LCMS).

 Table 4. Scope of Primary Amines in One-Pot Reductive

 Coupling Reactions^a



^{*a*}Average of at least two runs for each entry; >98% conversion in all cases unless indicated otherwise; yields for **5** are of isolated diamine products, $Ar = p-CF_3C_6H_4$. ^{*b*}90% conversion of **1a**.

case, was afforded in 50% yield. The data from reactions with benzyl amines 2a and 2f identified the trend of decreased diamine formation as the nucleophilicity of underlying amines decreased; meanwhile, the competing lactam over-reduction process increased. Similarly, the reductive coupling reaction with less nucleophilic aniline 2g furnished only 22% yield of diamine 5g. Furthermore, when a primary alkylamine with a carboxylic ester substituent one carbon away was utilized, only 5% yield of product 5h was obtained along with 80% pyrrolidine 4a, presumably due to the reduced nucleophilicity of the amine group caused by the electron-withdrawing inductive effect from the ester carbonyl.

In addition to reactions with primary amines, we have also investigated the reductive coupling of lactams with secondary amines, which would give a distinct set of diamine products. The results are summarized in Table 5. Treatment of **1a** with

Table 5. Scope of Secondary Amines in One-Pot Reductive Coupling Reactions^a



^{*a*}Average of at least two runs for each entry; >98% conversion and <5% 4 observed in all cases unless Indicated otherwise; yields for 6 are of isolated diamine products. ^{*b*}72% conversion of 1r. ^{*c*}The second portion of this reaction was carried out at 22 °C; >98% conversion of 10.

N-Boc-piperazine under the standard coupling conditions resulted in diamine 6a in 42% yield. Further optimization of this reaction sequence led to the production of 6a in 75% yield, when the temperature of the reductive amination step was raised to 80 °C. With the new conditions in hand, we examined several other substrates with N-Boc-piperazine. Product 6b, derived from the reductive coupling of an Narylated piperidinone, was afforded in 48% yield, and desired diamine 6c, which was generated from an N-arylated azetidinone, was furnished in comparable yield. Similarly, reductive coupling reactions using pyrrolidine as the amine nucleophile resulted in diamines 6e, 6f, and 6g in 56%, 32%, and 59% yield, respectively. Notably, pyrrolidinone 1r, which contains an OCF₃ substituent, was also proven to be a capable coupling partner (i.e., 6f). Additionally, the secondary amine does not need to be cyclic; as exemplified by diamine 6d, such a reaction between N-methyl-p-methoxybenzylamine and lactam 1a delivered the desired product in 30% yield. To further showcase the practicality of this chemistry, we have performed the reductive coupling reaction on gram scale. As illustrated in eq 1, upon treatment of 1.83 g (8.0 mmol) of 1a under standard conditions, the coupling with 2a furnished



diamine 3a in 63% yield (1.77 g; 5.0 mmol) with minimal formation of pyrrolidine 4a. The result demonstrated the amiability to scale up of the current semireduction/reductive amination sequence.

Extensive mechanistic discussions of Schwartz's reagent reductions of amides have suggested that secondary amide reductions proceed through imine intermediates,^{19,20} whereas tertiary amide reductions proceed through metal-stabilized complexes.^{8a} Based on the precedent, the mechanism for this transformation, as proposed in Scheme 2, invokes a zirconium complex I to serve as the masked aldehyde equivalent, as opposed to an iminium intermediate II. Nucleophilic primary amines attack the tetrahedral intermediate, which subsequently collapses to form the imine III or aminal IV. Upon treatment with sodium triacetoxyborohydride (STAB), this imine/aminal is reduced to the corresponding amine, thereby generating an overall diamine product. This mechanism is also invoked when utilizing secondary nucleophilic amines as coupling partners (i.e., the substrates of Table 5).

One benefit of using Schwartz's reagent to reduce the lactam into a masked aldehyde equivalent is the compatibility of these conditions with esters. To take advantage of this tolerability, we investigated the installation of a pendant ester in close proximity to the amide, as indicated in Scheme 3. When treated with Schwartz's reagent, we expected the formation of the masked aldehyde equivalent and upon addition of the amine nucleophile, an ester bearing intermediate diamine was anticipated.²¹⁷ We surmised that this intermediate, when treated with heat, would lead to intramolecular lactam formation between the pendant ester and the newly formed amine, a transformation which would not have been feasible with traditional amide reducing reagents. Standard reagents, such as LAH, LiEt₃BH, lithium borohydride, DIBAL-H, and lithium aminoborohydrides, directly reduced the pendant ester or over-reduced the starting lactam. Moreover, if such a transformation were to retain the stereochemical configuration of the starting material, a class of enantioenriched α -amino

piperidinones could become readily accessible. In Table 6, we summarized the results of reductive coupling reactions with pyrrolidinones that contain a 5-ester substituent, which were prepared in enantiopure form via Ullmann couplings between commercially available ethyl (R)-5-oxopyrrolidine-2-carboxy-late and a range of aryl iodides.

As shown in entry 1, Table 6, upon treatment of 7a with benzylamine 2a under slightly varied conditions, N-arylated piperidinone 8a was identified to be the major product from the reaction mixture, which was isolated in 75% yield and more importantly was obtained with full retention of the stereochemical information. An elevated temperature (80 °C) was needed to induce the cyclization of the benzylic amine center onto the ester after the diamine formation. As established in Tables 2 and 3, a similar trend was observed: electronwithdrawing substrates were generally more capable coupling partners. Piperidinones 8b-g were afforded in good yields, with minimal formation of over-reduced pyrrolidine byproducts (65–77% yield, <5–13% pyrrolidines). Interestingly, electron-rich substrates, which were previously incapable of participation in the coupling reactions (Table 2), were now effective in the formation of piperidinones. For examples, 8h and 8i were furnished in 51 and 55% yield, respectively, albeit a higher production of pyrrolidines was observed (7-20%). Reductive coupling of more hindered and electron-rich 7j did not proceed to completion within 18 h, resulting in 35% yield of 8j. Additionally, 88-100% ee was obtained for N-arylated piperidinones. The complete retention of stereochemistry in this transformation was purported to be derived from two complementary factors: (1) the lack of a basic reducing agent interacting directly with the α -stereocenter and (2) a proposed zirconacycle masked aldehyde equivalent V, as depicted in Scheme 4, which would reinforce the retention of the α stereocenter. Additionally, intermediate V disfavors the collapsed iminium ion VI, which would lead to the pyrrolidine byproduct. This explains our observation of relatively insignificant yields of pyrrolidines in various examples in Table 6 as compared to the amounts of pyrrolidines formed as side products from the diamine formation reactions described in previous sections (see Tables 2-5).

Scheme 2. Proposed Mechanism





Table 6. Examples of Reductive Coupling/Cyclization Sequence of α -Ester Containing *N*-Arylated Pyrrolidinones^{*a*}



"Average of at least two runs for each entry; >98% conversion in all cases unless indicated otherwise; ee = enantiomer excess as determined by chiral supercritical fluid chromatography (SFC) analysis in comparison with an authentic racemic sample; yield by silica gel chromatography; Ar = p-OMePh; pyrrolidine refers to ester containing over-reduction byproducts similar to 4. ^b90% conversion of 7i. ^c60% conversion of 7j.

CONCLUSIONS

In summary, we have established an operationally simple protocol that couples a primary or secondary amine with an Naryl-substituted lactam, delivering a differentiated diamine in 22-91% yield. We have successfully optimized conditions to allow semireduction/reductive amination sequence to be telescoped in a one-pot fashion. Nucleophilic amines and electron-deficient arylated lactams were generally more capable coupling partners in these reactions. Azetidinones, pyrrolidinones, and piperidinones were shown to participate in reductive coupling reactions effectively. This was significant as it allowed for a singular synthetic strategy to access 1,3-, 1,4-, and 1,5-diamines (which, if targeted individually, would require differing synthetic approaches based on the length of the carbon chain). A range of primary and secondary amines were included in the study to diversify the scope of diamine products. This work was further strengthened with a class of α carboxylic ester containing pyrrolidinones and their efficient conversion to enantiomerically pure α -amino piperidinones in 35-77% yield and complete retention of stereochemical information. Through an operationally simple, one-pot procedure to convert N-aryl lactams into masked aldehyde equivalents for use in reductive amination reactions, we have shown that lactams can be considered viable functional group handles potentially for a wider array of further transformations. Additional exploration in the use of lactams in this fashion is currently underway and will be disclosed in due course.

EXPERIMENTAL SECTION

Unless otherwise noted, all reactions were carried out with anhydrous solvents (dichloromethane, toluene, tetrahydrofuran, 1,4-dioxane, acetonitrile) purchased from Fisher Scientific and in standard glass vials and round-bottom flasks. All reagents were purchased from Sigma-Aldrich or Combi-Blocks and used as received, unless noted otherwise. A nitrogen atmosphere was maintained using standard vacuum line technique. The final compounds were analyzed or purified according to one of the analytical or purification methods referred to below unless otherwise described and characterized by ¹H, $^{13}\mathrm{C}$ NMR, and high-resolution mass spectrometry. For LC/MS analysis, a sample is dissolved in a suitable solvent such as MeCN, dimethyl sulfoxide (DMSO), or MeOH and is injected directly into the column using an automated sample handler. The analysis used one of the following methods: (1) acidic method (1.5, 2, 3.5, 4, or 7 min runs, see Supporting Information (SI) for additional details): conducted on a Shimadzu 2010 Series, Shimadzu 2020 Series, or Waters Acquity UPLC BEH. (MS ionization: ESI) instrument equipped with a C18 column (2.1 mm × 30 mm, 3.0 mm or 2.1 mm \times 50 mm, C18, 1.7 μ m), eluting with 1.5 mL/4 L of trifluoroacetic acid (TFA) in water (solvent A) and 0.75 mL/4 L of TFA in acetonitrile (solvent B) or (2) basic method (3, 3.5, 7 min runs, see SI for additional details): conducted on a Shimadzu 2020 Series or Waters Acquity UPLC BEH (MS ionization: ESI) instrument equipped with XBridge Shield RP18, 5um column (2.1 mm \times 30 mm, 3.0 mm i.d.) or 2.1 mm \times 50 mm, C18, 1.7 μ m column, eluting with 2 mL/4 L NH₃·H₂O in water (solvent A) and acetonitrile (solvent B). Silica gel column chromatography was performed using 20-40 μ M (particle size), 250-400 mesh, or 400Scheme 4. Proposed Mechanism for the Synthesis of Enantioenriched α -amino Piperidinones



632 mesh silica gel using either a Teledyne ISCO Combiflash RF or a Grace Reveleris X2 with ELSD purification systems. SFC chiral separation was performed using a Waters UPC2 analytical SFC (SFC-H). Column: ChiralCel OJ, 150 mm \times 4.6 mm i.d., 3 μ m. Mobile phase: A for CO₂ and B for ethanol (0.05% DEA). Gradient: B 40%. Flow rate: 2.5 mL/min. Back pressure: 100 bar. Column temperature: 35° C. Wavelength: 220 nm. Preparative HPLC purifications were performed on a Gilson UV/VIS-156 with UV detection at 220/254 nm, Gilson 281 automatic collection, utilizing acidic, basic and neutral methods (see SI for additional details). The ¹H NMR spectra were recorded on a Bruker Avance III HD 500 MHz, Bruker Avance III 500 MHz, Bruker Avance III 400 MHz, Varian-400 VNMRS, or Varian-400 MR. Chemical shifts are expressed in parts per million (ppm) units and referenced to the residual solvent resonance as noted in the spectra (i.e., $CDCl_3$; 7.27 ppm). Coupling constants (J) are in units of Hertz (Hz). Splitting patterns describe apparent multiplicities and are designated as: s (singlet), d (doublet), t (triplet), q (quartet), quin (quintet), dd (doubled doublet), dt (doubled triplet), dq (doubled quartet), m (multiplet), or br (broad). The ${}^{13}C{}^{1}H$ NMR spectra were recorded on a Bruker Avance III HD 500 MHz, Bruker Avance III 500 MHz, Bruker Avance III 400 MHz, Varian-400 VNMRS, or Varian-400 MR. Chemical shifts are expressed in parts per million (ppm) units and referenced to the residual solvent resonance as noted in the spectra (i.e., CDCl₃; 77.0 ppm). Coupling constants (J) are in units of Hertz (Hz). Splitting patterns describe apparent multiplicities and are designated as: s (singlet), d (doublet), t (triplet), dd (doubled doublet), dt (doubled triplet), dg (doubled quartet), m (multiplet), or br (broad). High-resolution mass spectrometry data were recorded using Sciex Triple TOF 6600 mass spectrometer (column: Waters UPLC Acquity HSS T3 C18 1.8 μ m, 2.1 mm × 50 mm; mobile phase: A = 0.1% formic acid in H₂O; B = 0.1% formic acid in acetonitrile; method: 7 min, A to B gradient 5-65%, positive ionization mode). All masses were reported as the exact masses in the format of $[M + H]^+$, unless noted otherwise.

Representative Experimental Procedure for Ullmann Couplings. Compounds 1a–r were Prepared by ChemPartner, Except Where Noted Otherwise. Nonetheless, Many are Commercially Available. A round-bottom flask was charged with aryl iodide or aryl bromide (1 equiv), 2-pyrrolidinone OR 2-piperidinone OR 2azetidinone (1.4 equiv), copper iodide (0.25 equiv), cesium carbonate (2 equiv), and p-dioxane (0.5 M reaction concentration) under an atmosphere of nitrogen. Then, tetramethylethylenediamine (TMEDA) or 1,2-dimethylethylenediamine (DMEDA) (0.5 equiv) was added, and the resulting mixture was heated to 40 °C and stirred at that temperature for 5 h. Additional portions of copper iodide (0.12 equiv) and TMEDA or DMEDA (0.25 equiv) were added (as needed). The reaction mixture continued to stir at 40 °C overnight. The reaction mixture was cooled to ambient temperature and filtered. The filtrate was concentrated, and the crude material was purified by silica gel column chromatography (EtOAc/heptanes, 1:1) to give the desired *N*-aryl azetidinones, *N*-aryl pyrrolidinones, or *N*-aryl piperidinones.

1-(4-(Trifluoromethyl)phenyl)pyrrolidin-2-one (1*a*, White Solid, 2.6 *g*, Yield: 96%).²² ¹H NMR (DMSO- d_6 , 500 MHz): δ 7.89 (2H, br d, *J* = 9.2 Hz), 7.73 (2H, d, *J* = 8.5 Hz), 3.88 (2H, t, *J* = 7.0 Hz), 2.57–2.52 (2H, m), 2.08 (2H, br t, *J* = 7.6 Hz). LRMS (ESI)⁺: 192.2. 1-(Pyridin-4-yl)pyrrolidin-2-one (1b, White Solid, 2.7 *g*, Yield: 72%). ¹H NMR (DMSO- d_6 , 500 MHz): δ 8.52–8.45 (2H, m), 7.71– 7.63 (2H, m), 3.88–3.80 (2H, m), 2.58–2.52 (2H, m), 2.12–2.03 (2H, m). LRMS (ESI)⁺: 163.1.

1-(4-Bromophenyl)pyrrolidin-2-one (1c, White Solid, 2.2 g, yield: 78%).²³ ¹H NMR (DMSO- d_{6} , 500 MHz): δ 7.68–7.61 (2H, m), 7.58–7.52 (2H, m), 3.84–3.78 (2H, m), 2.49–2.44 (2H, m), 2.10– 2.01 (2H, m). LRMS (ESI)⁺: 240.1.

1-(4-Chlorophenyl)pyrrolidin-2-one (1d, White Solid, 2.4 g, Yield: 53%).²⁴ ¹H NMR (DMSO- d_{6} , 500 MHz): δ 7.74–7.64 (2H, m), 7.47–7.39 (2H, m), 3.85–3.78 (2H, m), 2.48 (2H, s), 2.11–2.00 (2H, m). LRMS (ESI)⁺: 196.1.

4-(2-Oxopyrrolidin-1-yl)benzonitrile (1e, White Solid, 2.3 g, Yield: 71%).²⁵ ¹H NMR (DMSO- d_{6} , 500 MHz): δ 7.92–7.80 (4H, m), 3.91–3.84 (2H, m), 2.58–2.53 (2H, m), 2.13–2.04 (2H, m). LRMS (ESI)⁺: 187.1.

1-(4-Nitrophenyl)pyrrolidin-2-one (**1f**, Yellow Solid, 3.2 g, Yield: 67%).²⁶ ¹H NMR (DMSO- d_6 , 500 MHz): δ 8.29–8.24 (2H, m), 7.97–7.91 (2H, m), 3.91 (2H, t, *J* = 7.3 Hz), 2.58 (2H, t, *J* = 8.2 Hz), 2.13–2.05 (2H, m). LRMS (ESI)⁺: 207.1.

Ethyl 4-(2-Oxopyrrolidin-1-yl)benzoate (**1g**, White Solid, 3.6 g, Yield: 67%).²⁷ ¹H NMR (DMSO- d_6 , 500 MHz): δ 8.00–7.93 (2H, m), 7.85–7.80 (2H, m), 4.29 (2H, q, *J* = 7.1 Hz), 3.87 (2H, t, *J* = 7.3 Hz), 2.54 (2H, t, *J* = 8.2 Hz), 2.12–2.03 (2H, m), 1.31 (3H, t, *J* = 7.0 Hz). LRMS (ESI)⁺: 234.2.

1-(3-Chloro-5-fluorophenyl)pyrrolidin-2-one (**1h**, Yellow Solid, 3.5 *g*, Yield: 70%). ¹H NMR (DMSO- d_6 , 500 MHz): δ 7.71–7.63 (1H, m), 7.58 (1H, td, *J* = 11.6 Hz, 2.1 Hz), 7.18 (1H, td, *J* = 8.5 Hz, 2.2 Hz), 3.83 (2H, t, *J* = 7.3 Hz), 2.55–2.52 (2H, m), 2.10–2.01 (2H, m). LRMS (ESI)⁺: 214.1.

1-Phenylpyrrolidin-2-one (1i, White Solid). This material was purchased from Sigma-Aldrich and used as received. ¹H NMR (CDCl₃, 400 MHz): δ 7.65–7.59 (2H, m), 7.42–7.34 (2H, m), 7.19–7.12 (1H, m), 3.88 (2H, t, *J* = 7.0 Hz), 2.63 (2H, t, *J* = 8.0 Hz), 2.23–2.12 (2H, m). LRMS (ESI)⁺: 162.1.

1-(4-Methoxyphenyl)pyrrolidin-2-one (**1***j*, White Solid, 3.2 g, Yield: 72%).²⁸ ¹H NMR (DMSO- d_{6} , 500 MHz): δ 7.59–7.49 (2H, m), 6.99–6.90 (2H, m), 3.78 (2H, t, *J* = 7.0 Hz), 3.74 (3H, s), 2.45 (2H, t, *J* = 8.2 Hz), 2.04 (2H, quin, *J* = 7.5 Hz). LRMS (ESI)⁺: 192.2. 1-(2-Methoxyphenyl)pyrrolidin-2-one (**1***k*, Clear Oil, 2.1 g, Yield: 64%).²⁹ ¹H NMR (CDCl₃, 500 MHz): δ 7.31–7.24 (2H, m), 7.02– 6.94 (2H, m), 3.84 (3H, s), 3.79–3.74 (2H, m), 2.60–2.53 (2H, m), 2.24–2.15 (2H, m). LRMS (ESI)⁺: 192.0.

3-Methyl-1-(4-(trifluoromethyl)phenyl)pyrrolidin-2-one (11, White Solid, 900 mg, Yield: 78%). ¹H NMR (CDCl₃, 400 MHz): δ 7.80 (2H, d, *J* = 8.5 Hz), 7.62 (2H, d, *J* = 8.5 Hz), 3.85–3.78 (2H, m), 2.78–2.64 (1H, m), 2.42 (1H, tdd, *J* = 13.0 Hz, 8.1 Hz, 5.0 Hz), 1.90–1.75 (1H, m), 1.33 (3H, d, *J* = 7.0 Hz). LRMS (ESI)⁺: 244.1.

5-Methyl-1-(4-(trifluoromethyl)phenyl)pyrrolidin-2-one (1m, White Solid, 854 mg, Yield: 87%).³⁰ ¹H NMR (CDCl₃, 500 MHz): δ 7.67–7.63 (2H, m), 7.61–7.57 (2H, m), 4.45–4.36 (1H, m), 2.75– 2.65 (1H, m), 2.63–2.53 (1H, m), 2.46–2.36 (1H, m), 1.86–1.75 (1H, m), 1.27 (3H, d, *J* = 6.1 Hz). LRMS (ESI)⁺: 244.0.

1-(4-(Trifluoromethyl)phenyl)piperidin-2-one (**1n**, White Solid, 2.1 g, Yield: 78%).³¹ ¹H NMR (DMSO- d_6 , 500 MHz): δ 7.74 (2H, d, J = 8.5 Hz), 7.53 (2H, d, J = 8.5 Hz), 3.66 (2H, t, J = 5.5 Hz), 2.43 (2H, t, J = 6.4 Hz), 1.93–1.80 (4H, m). LRMS (ESI)⁺: 244.1.

1-(4-(*Trifluoromethyl*)*phenyl*)*azetidin-2-one* (**10**, *White Solid*). This material was purchased from Sigma-Aldrich and used as received. ¹H NMR (DMSO- d_6 , 500 MHz): δ 7.74 (2H, d, J = 8.5 Hz), 7.51 (2H, d, J = 8.5 Hz), 3.70 (2H, t, J = 4.9 Hz), 3.14 (2H, t, J = 4.9 Hz). LRMS (ESI)⁺: 216.1.

1-(*p*-Tolyl)*azetidin-2-one* (**1***p*, *White Solid*, 1.0 *g*, *Yield*: 44%).³² ¹H NMR (CD₃OD, 500 MHz): δ 7.30–7.24 (2H, m), 7.16 (2H, d, *J* = 7.9 Hz), 3.65 (2H, t, *J* = 4.3 Hz), 3.08 (2H, t, *J* = 4.6 Hz), 2.30 (3H, s). LRMS (ESI)⁺: 162.1.

1-(4-Methoxyphenyl)azetidin-2-one (**1q**, White Solid, 830 mg, Yield: 34%).³² ¹H NMR (CD₃OD, 500 MHz): δ 7.35–7.30 (2H, m), 6.94–6.89 (2H, m), 3.77 (3H, s), 3.64 (2H, t, *J* = 4.3 Hz), 3.07 (2H, t, *J* = 4.6 Hz). LRMS (ESI)⁺: 178.1.

1-(4-(*Trifluoromethoxy*)*phenyl*)*pyrrolidin-2-one* (**1***r*, *White Solid*, 2.2 *g*, *Yield*: 78%). ¹H NMR (DMSO- d_6 , 500 MHz): δ 7.83–7.73 (2H, m), 7.38 (2H, d, *J* = 9.2 Hz), 3.84 (2H, t, *J* = 7.0 Hz), 2.54–2.51 (2H, m), 2.13–2.00 (2H, m). LRMS (ESI)⁺: 246.1.

Representative Experimental Procedure for Reductive Coupling of Nonester Containing Aryl Lactams with Primary and Secondary Amines (for Tables 1-5). To a 2-dram vial equipped with a magnetic stir bar was added Schwartz's reagent (155 mg, 0.6 mmol, 1.5 equiv; preferably less than 1 month old) as a solid in the air. The vial was then sealed with a red open top cap (with a solvent-resistant Teflon cover), and a N_2 flow ran through the sealed vial for 5 min. The N_2 line was then removed, and THF (2 mL) was added through a syringe under positive pressure. The reaction mixture was cooled to 0 °C in an ice-water cooling bath and subjected to vigorous stirring, followed by the injection of a solution of the substrate (0.4 mmol in 1 mL of THF). After 1 h at 0 °C, to the milky white suspension was added neat amine coupling partner (1.6 mmol, 4 equiv). The vial was then removed from the ice-water bath, and STAB (170 mg, 0.8 mmol, 2 equiv) was added in one portion as a solid to the reaction mixture. The vial was capped and stirred at 23 °C for 18 h. A sample was taken from the vial to be analyzed by LCMS. Upon reaction completion, water (1 mL) was added to the vial. The mixture was stirred for 10 min before the addition of EtOAc (2 mL). The mixture was filtered through a plug of Celite into a 30 mL vial. The filtrate was dried by a V10 rotovap. The residue was purified by silica gel column chromatography (4g of silica gel column) with an automatic purification system (grading from 0% MeOH/EtOAc to 50% MeOH/EtOAc over 13 min). The fractions were collected and dried to give the desired diamine products as clear films.

*N*¹-(4-*M*ethoxybenzyl)-*N*⁴-(4-(trīfluoromethyl)phenyl)butane-1,4diamine (**3a**; 88.7 mg, 63% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 9.33 (1H, br s), 7.38 (2H, br d, *J* = 8.1 Hz), 7.25 (2H, d, *J* = 8.0 Hz), 6.83 (2H, br d, *J* = 7.8 Hz), 6.61 (2H, br d, *J* = 8.3 Hz), 4.56 (1H, br s), 3.86 (2H, br s), 3.70 (3H, s), 3.06 (2H, br t, *J* = 5.6 Hz), 2.85 (2H, br s), 1.73 (2H, br s), 1.61 (2H, br s). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 160.4, 150.0, 131.4, 126.6 (q, *J* = 4 Hz), 124.9 (q, *J* = 271 Hz), 122.0, 119.3 (q, *J* = 32 Hz), 114.4, 112.2, 55.2, 50.7, 46.0, 42.8, 25.6, 23.4. HRMS (ESI⁺): calcd for C₁₉H₂₃F₃N₂O [M + H]⁺: 353.1835; found: 353.1828.

 N^{1} -(4-Methoxybenzyl)- N^{4} -(pyridin-4-yl)butane-1,4-diamine (**3b**; 67.3 mg, 59% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.97 (1H, br s), 8.70 (1H, br s), 8.24 (1H, br d, *J* = 6.8 Hz), 8.08 (1H, br d, *J* = 6.8 Hz), 7.40 (2H, d, *J* = 8.8 Hz), 6.98 (2H, d, *J* = 8.6 Hz), 6.87 (2H, br t, *J* = 5.7 Hz), 4.07 (2H, br s), 3.76 (3H, s), 3.28 (2H, q, *J* = 6.3 Hz), 2.92 (2H, br s), 1.74-1.54 (4H, m). $^{13}C{}^{1}H$ NMR (DMSO- d_{6} , 100 MHz): δ 159.7, 157.9, 140.8, 138.5, 131.5, 123.7, 114.0, 109.8, 104.7, 55.2, 49.5, 45.8, 41.5, 25.0, 22.8. HRMS (ESI⁺): calcd for C₁₇H₂₄N₃O [M + H]⁺: 286.1914; found: 286.1908.

N¹-(4-Bromophenyl)-N⁴-(4-methoxybenzyl)butane-1,4-diamine (**3***c*; 118.7 mg, 82% Yield). ¹H NMR (DMSO- d_6 , 400 MHz): δ 8.81 (2H, br s), 7.40 (2H, d, J = 8.6 Hz), 7.21 (2H, d, J = 8.6 Hz), 6.98 (2H, d, J = 8.6 Hz), 6.57–6.50 (2H, m), 4.07 (2H, br t, J = 5.4 Hz), 3.76 (3H, s), 2.99 (2H, t, J = 6.7 Hz), 2.94–2.85 (2H, m), 1.68 (2H, quin, J = 7.5 Hz), 1.61–1.49 (2H, m). ¹³C{¹H} NMR (DMSO- d_6 , 100 MHz): δ 159.7, 147.7, 131.5, 131.4, 123.7, 117.5, 114.2, 114.0, 55.2, 49.5, 46.0, 42.3, 25.4, 23.1. HRMS (ESI⁺): calcd for C₁₈H₂₃BrN₂O [M + H]⁺: 363.1067; found: 363.1066.

¹⁸ ¹ ²(4-Chlorophenyl)-N⁴-(4-methoxybenzyl)butane-1,4-diamine (**3d**; 44.5 mg, 35% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 8.51 (1H, br s), 7.30 (2H, br d, *J* = 8.1 Hz), 7.07 (2H, d, *J* = 8.6 Hz), 6.84 (2H, br d, *J* = 7.8 Hz), 6.46 (2H, d, *J* = 8.6 Hz), 5.42 (1H, br s), 3.83 (2H, br s), 3.72 (3H, s), 2.98 (2H, br t, *J* = 6.0 Hz), 2.77 (2H, t, *J* = 5.9 Hz), 1.77 (2H, br s), 1.64–1.52 (2H, m). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 160.0, 146.8, 131.3, 128.9, 123.4, 121.5, 114.2, 113.6, 55.2, 50.3, 45.8, 43.0, 26.1, 23.8. HRMS (ESI⁺): calcd for C₁₈H₂₃ClN₂O [M + H]⁺: 319.1572; found: 319.1569.

4-(*i*4-(*i*4-Methoxybenzyl)amino)butyl)amino)benzonitrile (**3e**; 60.5 mg, 49% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 9.30 (1H, br s), 7.35 (2H, br d, *J* = 8.6 Hz), 7.25 (2H, br d, *J* = 8.1 Hz), 6.83 (2H, br d, *J* = 8.1 Hz), 6.48 (2H, br d, *J* = 8.3 Hz), 4.59 (1H, br s), 3.85 (2H, br s), 3.69 (3H, s), 3.05 (2H, br t, *J* = 6.2 Hz), 2.85 (2H, br s), 1.71 (2H, m), 1.64–1.52 (2H, m). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 160.4, 151.2, 133.6, 131.3, 121.9, 120.4, 114.4, 112.0, 98.3, 55.2, 50.7, 46.0, 42.0, 25.5, 23.3. HRMS (ESI⁺): calcd for C₁₉H₂₃N₃O [M + H]⁺: 310.1914; found: 310.1908.

N¹-(4-Methoxybenzyl)-N⁴-(4-nitrophenyl)butane-1,4-diamine (**3f**; 51.3 mg, 39% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 8.02 (2H, d, J = 8.0 Hz), 7.27 (2H, d, J = 8.0 Hz), 6.90 (2H, d, J = 8.0 Hz), 6.59 (2H, d, J = 8.0 Hz), 3.80 (2H, s), 3.78 (3H, s), 3.24–3.19 (2H, m), 2.75–2.70 (2H, m), 1.71–1.65 (4H, m). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 161.0, 156.2, 138.0, 131.4, 130.7, 127.5, 115.1, 111.8, 55.9, 53.5, 43.8, 27.7, 27.2. One carbon signal under solvent multiplet. HRMS (ESI⁺): calcd for C₁₈H₂₃N₃O₃ [M + H]⁺: 330.1812; found: 330.1806.

Ethyl 4-((4-(Methoxybenzyl)amino)butyl)amino)benzoate (**3g**; 48.4 mg, 34% Yield). ¹H NMR (DMSO- d_6 , 400 MHz): δ 7.67 (2H, d, J = 8.8 Hz), 7.42 (2H, d, J = 8.6 Hz), 6.93 (2H, d, J = 8.6 Hz), 6.64–6.60 (1H, m), 6.57 (2H, d, J = 8.6 Hz), 4.19 (2H, q, J = 7.1 Hz), 3.92 (2H, s), 3.75 (3H, s), 3.11–3.02 (2H, m), 2.75 (2H, br t, J = 7.3 Hz), 1.76–1.63 (2H, m), 1.62–1.51 (2H, m), 1.26 (4H, t, J = 7.1 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 100 MHz): δ 165.9, 159.2, 152.8, 131.0, 130.9, 126.2, 115.8, 113.8, 110.8, 59.5, 55.1, 50.1, 46.4, 41.8, 25.7, 24.0, 14.4. HRMS (ESI⁺): calcd for C₂₁H₂₈N₂O₃ [M + H]⁺: 357.2173; found: 357.2168.

*N*¹-(3-Chloro-5-fluorophenyl)-*N*⁴-(4-methoxybenzyl)butane-1,4diamine (**3h**; 56.4 mg, 42% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.93 (2H, br s), 7.42 (2H, br d, *J* = 8.1 Hz), 6.99 (2H, br d, *J* = 7.8 Hz), 6.43 (1H, s), 6.42–6.37 (1H, m), 6.35–6.28 (1H, m), 4.08 (2H, br t, *J* = 5.4 Hz), 3.77 (3H, s), 3.02 (2H, t, *J* = 6.8 Hz), 2.92 (2H, br s), 1.69 (2H, br s), 1.60–1.50 (2H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 163.4 (d, *J* = 242 Hz), 159.7, 151.4 (d, *J* = 13 Hz), 134.2 (d, *J* = 14 Hz), 131.5, 123.8, 114.0, 107.7, 101.9 (d, *J* = 26 Hz), 96.8 (d, *J* = 25 Hz), 55.2, 49.5, 46.0, 41.9, 25.4, 23.1. HRMS (ESI⁺): calcd for C₁₈H₂₂CIFN₂O [M + H]⁺: 337.1477; found: 337.1474.

 N^{1} -(4-Methoxybenzyl)- N^{4} -phenylbutane-1,4-diamine (**3i**; 40.9 mg, 36% Yield). ¹H NMR (CD₃OD, 500 MHz): δ 7.60−7.52 (2H, m), 7.52−7.46 (3H, m), 7.43−7.38 (2H, m), 7.01−6.95 (2H, m), 4.12 (2H, s), 3.80 (3H, s), 3.49−3.39 (2H, m), 3.10−3.02 (2H, m), 1.88−1.74 (4H, m). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 162.3, 137.5, 132.7, 131.6, 130.5, 124.4, 123.5, 115.7, 56.0, 52.0, 47.6, 24.4, 24.3. One carbon signal under solvent multiplet. HRMS (ESI⁺): calcd for C₁₈H₂₄N₂O [M + H]⁺: 285.1961; found: 285.1967.

 N^{1} -(4-Methoxybenzyl)-2-methyl- N^{4} -(4-(trifluoromethyl)phenyl)butane-1,4-diamine (**3**]; 62.9 mg, 43% Yield). ¹H NMR (CDCl₃, 500 MHz): δ 7.38 (2H, d, *J* = 8.5 Hz), 7.25 (2H, d, *J* = 8.5 Hz), 6.88 (2H, d, *J* = 8.5 Hz), 6.54 (2H, d, *J* = 8.5 Hz), 4.53 (1H, br s), 3.83– 3.79 (3H, m), 3.74 (2H, s), 3.24–3.16 (1H, m), 3.15–3.07 (1H, m), 2.55 (2H, d, *J* = 6.1 Hz), 1.84–1.70 (2H, m), 1.61–1.49 (2H, m), 0.99 (3H, d, *J* = 6.7 Hz). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 158.7, 150.9, 132.3, 129.3, 126.5 (q, *J* = 4 Hz), 125.1 (d, *J* = 270 Hz), 118.2 (q, *J* = 33 Hz), 113.8, 111.5, 55.3, 55.3, 53.5, 41.3, 34.2, 31.6, 18.6. HRMS (ESI⁺): calcd for C₂₀H₂₅F₃N₂O [M + H]⁺: 367.1992; found: 367.1985.

*N*¹-(4-Methoxybenzyl)-*N*⁴-(4-(trifluoromethyl)phenyl)pentane-1,4-diamine (**3m**; 49.2 mg, 34% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 8.82 (2H, br s), 7.39 (2H, d, *J* = 8.8 Hz), 7.34 (2H, d, *J* = 8.8 Hz), 6.97 (2H, d, *J* = 8.6 Hz), 6.65 (2H, d, *J* = 8.6 Hz), 4.06 (2H, br t, *J* = 5.6 Hz), 3.76 (3H, s), 3.54–3.40 (1H, m), 2.94–2.81 (2H, m), 1.77–1.60 (2H, m), 1.57–1.41 (2H, m), 1.11 (3H, d, *J* = 6.4 Hz). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 159.0, 151.2, 130.8, 126.7, 126.2 (q, *J* = 3 Hz), 125.4 (q, *J* = 270 Hz), 114.5 (q, *J* = 32 Hz), 113.7, 111.4, 55.1, 50.2, 46.8, 46.7, 33.1, 23.4, 20.1. HRMS (ESI⁺): calcd for C₂₀H₂₅F₃N₂O [M + H]⁺: 367.1992; found: 367.1985.

N¹-(4-Methoxybenzyl)-N⁵-(4-(trifluoromethyl)phenyl)pentane-1,5-diamine (**3n**; 105.4 mg, 72% Yield). ¹H NMR (DMSO- d_{6} , 400 MHz): δ 8.81 (2H, br s), 7.41 (2H, d, *J* = 7.5 Hz), 7.35 (2H, d, *J* = 8.5 Hz), 6.99 (2H, d, *J* = 7.7 Hz), 6.64 (2H, d, *J* = 8.6 Hz), 4.08 (2H, t, *J* = 5.6 Hz), 3.76 (3H, s), 3.04 (2H, t, *J* = 6.8 Hz), 2.93–2.83 (2H, m), 1.64 (2H, br quin, *J* = 7.7 Hz), 1.55 (2H, br quin, *J* = 7.1 Hz), 1.38 (2H, br quin, *J* = 7.6 Hz). ¹³C{¹H} NMR (DMSO- d_{6} , 100 MHz): δ 159.7, 151.8, 131.5, 126.2 (q, *J* = 4 Hz), 125.4 (q, *J* = 270 Hz), 123.8, 114.9 (q, *J* = 32 Hz), 114.0, 111.1, 55.2, 49.5, 46.1, 42.0, 27.8, 25.1, 23.5. HRMS (ESI⁺): calcd for C₂₀H₂₅F₃N₂O [M + H]⁺: 367.1992; found: 367.1985.

*N*¹-(4-*Methoxybenzyl*)-*N*³-(4-(*trifluoromethyl*)*phenyl*)*propane*-1,3-*diamine* (**3***o*; 123.0 mg, 91% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.87 (2H, br s), 7.40 (2H, d, *J* = 8.6 Hz), 7.38 (2H, d, *J* = 8.6 Hz), 6.98 (2H, d, *J* = 7.7 Hz), 6.66 (2H, d, *J* = 8.6 Hz), 4.09 (2H, t, *J* = 5.5 Hz), 3.76 (3H, s), 3.15 (2H, t, *J* = 6.8 Hz), 3.05−2.94 (2H, m), 1.88 (2H, quint, *J* = 7.1 Hz). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 160.8, 151.2, 131.1, 126.0 (q, *J* = 4 Hz), 125.2 (q, *J* = 268 Hz), 122.8, 117.7 (q, *J* = 32 Hz), 114.2, 111.4, 54.4, 50.5, 44.9, 39.7, 26.2. HRMS (ESI⁺): calcd for C₁₈H₂₂F₃N₂O [M + H]⁺: 339.1679; found: 339.1673.

*N*¹-(4-Methoxybenzyl)-*N*³-(*p*-tolyl)propane-1,3-diamine (**3***p*; 34.1 mg, 30% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.31−7.23 (2H, m), 7.05−6.98 (2H, m), 6.94−6.86 (2H, m), 6.60−6.52 (2H, m), 3.82 (3H, s), 3.76 (2H, s), 3.21 (2H, t, *J* = 6.6 Hz), 2.79 (2H, t, *J* = 6.6 Hz), 2.27 (3H, s), 1.83 (2H, quin, *J* = 6.6 Hz). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 158.7, 146.4, 132.5, 129.8, 129.3, 126.3, 113.9, 113.0, 55.3, 53.5, 47.7, 43.3, 29.6, 20.4. HRMS (ESI⁺): calcd for C₁₈H₂₄N₂O [M + H]⁺: 285.1961; found: 285.1967.

*N*¹-(4-Methoxybenzyl)-*N*³-(4-methoxyphenyl)propane-1,3-diamine (**3q**; 57.6 mg, 48% Yield). ¹H NMR (CDCl₃, 500 MHz): δ 7.20−7.13 (2H, m), 6.79 (2H, br d, *J* = 7.9 Hz), 6.70 (2H, br d, *J* = 9.2 Hz), 6.49 (2H, br d, *J* = 9.2 Hz), 3.73 (3H, s), 3.68−3.64 (5H, m), 3.08 (2H, t, *J* = 6.4 Hz), 2.68 (2H, t, *J* = 6.4 Hz), 1.73 (2H, quin, *J* = 6.6 Hz). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 158.6, 151.9, 142.9, 132.5, 129.2, 114.9, 114.0, 113.8, 55.8, 55.3, 53.5, 47.7, 43.8, 29.6. HRMS (ESI⁺): calcd for C₁₈H₂₄N₂O₂ [M + H]⁺: 301.1911; found: 301.1924.

*N*¹-(*Cyclohexylmethyl*)-*N*⁴-(4-(*trifluoromethyl*)*phenyl*)*butane*-1,4-*diamine* (*5a*; 90.5 mg, 69% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.35 (2H, br s), 7.36 (2H, d, *J* = 8.6 Hz), 6.65 (2H, d, *J* = 8.6 Hz), 3.08 (2H, t, *J* = 6.6 Hz), 2.99–2.86 (2H, m), 2.81–2.69 (2H, m), 1.81–1.49 (10H, m), 1.34–1.08 (3H, m), 1.03–0.82 (2H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 151.7, 126.2 (q, *J* = 4 Hz), 125.4 (q, *J* = 270 Hz), 115.0 (q, *J* = 32 Hz), 111.2, 52.6, 47.2, 41.7, 34.4, 29.9, 25.5, 25.4, 25.0, 23.0. HRMS (ESI⁺): calcd for C₁₈H₂₇F₃N₂ [M + H]⁺: 329.2199; found: 329.2191.

N¹-Cyclohexyl-N⁴-(4-(trifluoromethyl)phenyl)butane-1,4-diamine (**5b**; 72.8 mg, 58% Yield). ¹H NMR (DMSO- d_{64} 400 MHz): δ 8.65 (1H, br s), 7.36 (2H, br d, J = 8.3 Hz), 6.66 (2H, br d, J = 8.6 Hz), 6.46 (1H, br t, J = 5.4 Hz), 3.08 (2H, q, J = 6.4 Hz), 2.99–2.82 (3H, m), 2.02 (2H, br d, J = 10.3 Hz), 1.79–1.64 (4H, m), 1.62–1.53 (3H, m), 1.34–1.15 (4H, m), 1.15–1.02 (1H, m). $^{13}C{}^{1}H$ NMR (DMSO- d_6 , 100 MHz): δ 151.8, 126.2 (q, J = 4 Hz), 125.4 (q, J = 270 Hz), 114.8 (q, J = 32 Hz), 111.1, 55.8, 43.2, 41.7, 28.4, 25.5, 24.8, 23.9, 23.3. HRMS (ESI⁺): calcd for $C_{17}H_{25}F_3N_2$ [M + H]⁺: 315.2043; found: 315.2037.

N¹-Nonyl-N⁴-(4-(trifluoromethyl)phenyl)butane-1,4-diamine (**5***c*; 90.2 mg, 63% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.71 (1H, br s), 7.35 (2H, d, *J* = 8.6 Hz), 6.66 (2H, d, *J* = 8.6 Hz), 6.47 (1H, br s), 3.07 (2H, br d, *J* = 5.4 Hz), 2.90–2.71 (4H, m), 1.69 (2H, quin, *J* = 7.4 Hz), 1.64–1.52 (4H, m), 1.25 (12H, br s), 0.85 (3H, br t, *J* = 6.6 Hz). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 151.7, 126.1 (d, *J* = 4 Hz), 125.4 (q, *J* = 269 Hz), 114.8 (q, *J* = 32 Hz), 111.1, 46.6, 46.3, 41.6, 31.2, 28.8, 28.6, 28.5, 26.0, 25.4, 25.3, 23.1, 22.1, 13.9. HRMS (ESI⁺): calcd for C₂₀H₃₃F₃N₂ [M + H]⁺: 359.2669; found: 359.2661.

(*S*)-*N*¹-(1-*Methoxypropan*-2-*y*])-*N*⁴-(4-(trifluoromethyl)phenyl)butane-1,4-diamine (*5d*; 74.1 mg, 61% Yield). ¹H NMR (DMSO- d_6 , 400 MHz): δ 8.78 (1H, br s), 8.65 (1H, br s), 7.36 (2H, d, *J* = 8.6 Hz), 6.66 (2H, m, *J* = 8.6 Hz), 6.45 (1H, br t, *J* = 5.4 Hz), 3.58–3.43 (2H, m), 3.31 (3H, s), 3.08 (2H, q, *J* = 6.3 Hz), 2.99–2.82 (2H, m), 1.77–1.66 (2H, m), 1.66–1.51 (2H, m), 1.21 (3H, d, *J* = 6.6 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 100 MHz): δ 151.8, 126.2 (q, *J* = 4 Hz), 125.5 (q, *J* = 269 Hz), 114.9 (q, *J* = 32 Hz), 111.2, 71.8, 58.4, 52.4, 44.0, 41.7, 25.5, 23.3, 13.6. HRMS (ESI⁺): calcd for C₁₅H₂₃F₃N₂O [M + H]⁺: 305.1835; found: 305.1829.

(*S*)-*N*¹-(1-Phenylethyl)-*N*⁴-(4-(trifluoromethyl)phenyl)butane-1,4diamine (**5e**; 67.2 mg, 50% Yield). ¹H NMR (DMSO- d_6 , 400 MHz): δ 8.92 (1H, br s), 8.78 (1H, br s), 7.55-7.39 (5H, m), 7.36 (2H, d, *J* = 8.6 Hz), 6.63 (2H, d, *J* = 8.6 Hz), 4.42–4.29 (1H, m), 3.04 (2H, t, *J* = 6.7 Hz), 2.88 (1H, br s), 2.74–2.59 (1H, m), 1.71–1.59 (2H, m), 1.58–1.46 (5H, m). ¹³C{¹H} NMR (DMSO- d_6 , 100 MHz): δ 151.7, 137.5, 128.8, 128.7, 127.9, 126.2, 125.4 (q, *J* = 269 Hz), 114.9 (q, *J* = 32 Hz), 111.1, 57.2, 44.7, 41.7, 25.5, 23.2, 19.8. HRMS (ESI⁺): calcd for C₁₉H₂₃F₃N₂ [M + H]⁺: 337.1886; found: 337.1878.

*N*¹-(4-(Trifluoromethyl)benzyl)-*N*⁴-(4-(trifluoromethyl)phenyl)butane-1,4-diamine (**5f**; 49.9 mg, 32% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 9.13 (2H, br s), 7.82 (2H, d, *J* = 8.1 Hz), 7.72 (2H, d, *J* = 8.3 Hz), 7.36 (2H, d, *J* = 8.6 Hz), 6.65 (2H, d, *J* = 8.6 Hz), 4.26 (2H, br t, *J* = 5.5 Hz), 3.08 (2H, t, *J* = 6.7 Hz), 2.98 (2H, br s), 1.78– 1.65 (2H, m), 1.65–1.53 (2H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 151.8, 136.7, 130.9, 129.6 (q, *J* = 32 Hz), 126.4 (q, *J* = 4 Hz), 125.7 (q, *J* = 4 Hz), 125.5 (q, *J* = 269 Hz), 124.2 (q, *J* = 272 Hz), 115.0 (q, *J* = 32 Hz), 111.4, 49.6, 46.8, 41.8, 25.5, 23.3. HRMS (ESI⁺): calcd for C₁₉H₂₀F₆N₂ [M + H]⁺: 391.1603; found: 391.1595.

N¹-(4-Methoxyphenyl)-N⁴-(4-(trifluoromethyl)phenyl)butane-1,4-diamine (**5g**; 29.7 mg, 22% Yield). ¹H NMR (DMSO-*d*₆, 400 MHz): δ 7.35 (2H, d, *J* = 8.6 Hz), 6.70 (2H, d, *J* = 8.8 Hz), 6.65 (2H, d, *J* = 8.6 Hz), 6.52 (2H, d, *J* = 8.8 Hz), 6.36 (1H, br t, *J* = 5.1 Hz), 5.09 (1H, br s), 3.63 (3H, s), 3.13−3.04 (2H, m), 2.97 (2H, br s), 1.72−1.56 (4H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 151.9, 150.5, 143.4, 126.2 (q, *J* = 4 Hz), 125.4 (q, *J* = 270 Hz), 114.8 (q, *J* = 32 Hz), 114.6, 112.9, 111.1, 55.3, 43.5, 42.2, 26.5, 26.2. HRMS (ESI⁺): calcd for C₁₈H₂₁F₃N₂O [M + H]⁺: 339.1679; found: 339.1670.

tert-Butyl 4-(4-((4-(Trifluoromethyl)phenyl)amino)butyl)piperazine-1-carboxylate (**6a**; 120.3 mg, 75% Yield). ¹H NMR (CD₃OD, 500 MHz): δ 7.32 (2H, d, *J* = 9.2 Hz), 6.64 (2H, d, *J* = 8.5 Hz), 3.42 (4H, br, s), 3.15 (2H, br t, *J* = 6.4 Hz), 2.47–2.35 (6H, m), 1.71–1.56 (4H, m), 1.45 (9H, s). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 156.5, 153.4, 127.4 (q, *J* = 4 Hz), 126.9 (q, *J* = 269 Hz), 118.4 (q, *J* = 32 Hz), 112.6, 81.4, 59.5, 54.1, 44.5 (br d, rotameric quaternary carbon), 44.0, 28.8, 28.1, 25.2. HRMS (ESI⁺): calcd for C₂₀H₃₀F₃N₃O₂ [M + H]⁺: 402.2363; found: 402.2370.

tert-Butyl 4-(5-((4-(Trifluoromethyl)phenyl)amino)pentyl)piperazine-1-carboxylate (**6b**; 79.7 mg, 48% Yield). ¹H NMR (CDCl₃, 500 MHz): δ 7.39 (2H, d, J = 8.5 Hz), 6.58 (2H, d, J = 8.5 Hz), 3.97 (1H, br s), 3.49–3.38 (4H, m), 3.19–3.11 (2H, m) 2.42– 2.31 (6H, m), 1.73–1.61 (2H, m), 1.59–1.51 (2H, m), 1.48–1.41 (11H, m). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 154.7, 150.7, 126.6 (q, *J* = 4 Hz), 125.0 (q, *J* = 271 Hz), 118.5 (q, *J* = 33 Hz), 111.6, 79.6, 58.5, 53.1, 43.7 (br d, rotameric quaternary carbon), 43.4, 29.2, 28.4, 26.6, 24.9. HRMS (ESI⁺): calcd for C₂₁H₃₂F₃N₃O₂ [M + H]⁺: 416.2519; found: 416.2522.

tert-Butyl 4-(3-((4-(Trifluoromethyl)phenyl)amino)propyl)piperazine-1-carboxylate (**6c**; 69.7 mg, 45% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.39 (2H, d, *J* = 8.3 Hz), 6.58 (2H, d, *J* = 8.3 Hz), 5.13 (1H, br s), 3.52–3.42 (4H, m), 3.24 (2H, q, *J* = 6.0 Hz), 2.51 (2H, t, *J* = 6.4 Hz), 2.42 (4H, t, *J* = 5.0 Hz), 1.82 (2H, quin, *J* = 6.4 Hz), 1.48 (9H, s). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 154.7, 151.0, 126.6 (q, *J* = 4 Hz), 125.0 (q, *J* = 270 Hz), 118.4 (q, *J* = 32 Hz), 111.6, 79.8, 57.2, 53.1, 43.7 (br d, rotameric quaternary carbon), 43.0, 28.4, 25.3. HRMS (ESI⁺): calcd for C₁₉H₂₈F₃N₃O₂ [M + H]⁺: 388.2206; found: 388.2208.

 N^{1} -(4-*Methoxybenzyl*)- N^{1} -*methyl*- N^{4} -(4-(*trifluoromethyl*)*phenyl*)butane-1,4-diamine (**6d**; 43.9 mg, 30% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 7.35−7.31 (2H, m), 7.31−7.26 (2H, m), 6.90 (2H, d, *J* = 7.8 Hz), 6.64 (2H, d, *J* = 8.5 Hz), 3.78 (3H, s), 3.75 (2H, s), 3.14 (2H, t, *J* = 6.7 Hz), 2.72−2.62 (2H, m), 2.40 (3H, s), 1.78−1.68 (2H, m) 1.68−1.59 (2H, m). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 159.0, 150.8, 130.7, 128.9, 126.5 (q, *J* = 4 Hz), 125.0 (q, *J* = 270 Hz), 118.1 (q, *J* = 32 Hz), 113.7, 111.5, 60.9, 56.0, 55.2, 43.1, 41.2, 26.7, 24.3. HRMS (ESI⁺): calcd for C₂₀H₂₅F₃N₂O [M + H]⁺: 367.1992; found: 367.2008.

N-(4-(*Pyrrolidin*-1-*yl*)*butyl*)-4-(*trifluoromethyl*)*aniline* (*6e*; 64.0 mg, 56% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 7.34 (2H, d, J = 8.5 Hz), 6.68 (2H, d, J = 8.5 Hz), 3.68–3.59 (2H, m), 3.24–3.17 (4H, m), 3.11–3.00 (2H, m), 2.19–2.07 (2H, m), 2.07–1.95 (2H, m), 1.88–1.79 (2H, m), 1.75–1.66 (2H, m). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 150.2, 126.5 (q, J = 4 Hz), 124.9 (q, J = 268 Hz), 118.7 (q, J = 33 Hz), 112.0, 55.0, 53.8, 42.4, 25.6, 23.2, 23.1. HRMS (ESI⁺): calcd for C₁₅H₂₁F₃N₂ [M + H]⁺: 287.1730; found: 287.1732.

N-(4-(*Pyrrolidin*-1-*yl*)*butyl*)-4-(*trifluoromethoxy*)*aniline* (**6***f*; 38.7 mg, 32% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 7.12 (2H, d, *J* = 8.6 Hz), 6.87 (2H, d, *J* = 7.8 Hz), 3.64 (2H, br s), 3.26–3.18 (4H, m), 3.14–2.99 (2H, m), 2.20–2.08 (2H, m), 2.08–1.95 (2H, m), 1.88–1.78 (2H, m), 1.78–1.68 (2H, m). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 144.8, 140.8, 122.4, 120.4 (q, *J* = 257 Hz), 118.7, 54.8, 53.8, 47.2, 24.3, 23.0, 22.9. HRMS (ESI⁺): calcd for C₁₅H₂₁F₃N₂O [M + H]⁺: 303.1679; found: 303.1683.

N-(3-(*Pyrrolidin*-1-*yl*)*propyl*)-4-(*trifluoromethyl*)*aniline* (*6g*; 64.2 mg, 59% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.39 (2H, d, *J* = 8.3 Hz), 6.57 (2H, d, *J* = 8.5 Hz), 5.14 (1H, br s), 3.26–3.21 (2H, m), 2.61 (2H, t, *J* = 6.7 Hz), 2.56–2.49 (4H, m), 1.87–1.78 (6H, m). ¹³C{¹H} NMR (CDCl₃, 125 MHz): δ 151.2, 126.5 (q, *J* = 4 Hz), 122.9 (q, *J* = 271 Hz), 118.1 (q, *J* = 32 Hz), 111.6, 54.9, 54.2, 43.1, 27.6, 23.5. HRMS (ESI⁺): calcd for C₁₄H₁₉F₃N₂ [M + H]⁺: 273.1573; found: 273.1581.

Representative Experimental Procedure for Ullmann Couplings. Compounds 7*a*–*j* were Prepared by ChemDiscover LLC. A round-bottom flask was charged with aryl iodide or aryl bromide (1 equiv), ethyl (*R*)-5-oxopyrrolidine-2-carboxylate (1.4 equiv), copper iodide (0.25 equiv), cesium carbonate (2 equiv), and *p*-dioxane (0.5 M reaction concentration) under an atmosphere of nitrogen. Then, TMEDA or DMEDA (0.5 equiv) was added, and the resulting mixture was heated to 40 °C and stirred at that temperature for 5 h. Additional portions of copper iodide (0.12 equiv) and TMEDA or DMEDA (0.25 equiv) were added (as needed). The reaction mixture continued to stir at 40 °C overnight. The reaction mixture was cooled to ambient temperature and filtered. The filtrate was concentrated, and the crude material was purified by silica gel column chromatography (EtOAc/petroleum ether, 1:2) to give the desired *N*-aryl pyrrolidinones.

Ethyl (R)-5-Oxo-1-(4-(trifluoromethyl)phenyl)pyrrolidine-2-carboxylate (**7a**, Yellow Oil, 18.2 g, Yield: 86%, 99.5% ee). ¹H NMR (DMSO- d_6 , 500 MHz) δ : 7.82–7.67 (4H, m), 5.10 (1H, dd, J = 8.9 Hz, 2.7 Hz), 4.21–4.07 (2H, m), 2.66–2.53 (3H, m), 2.17–2.07 (1H, m), 1.15 (3H, t, J = 7.0 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 125 MHz) δ : 175.0, 171.9, 142.5, 126.5 (q, J = 4 Hz), 124.9 (q, J = 32 Hz), 124.7 $(q, J = 270 \text{ Hz}), 120.7, 61.8, 60.6, 31.1, 22.7, 14.4. HRMS (ESI⁺): calcd for C₁₄H₁₄F₃NO₃ <math>[M + H]^+$: 302.0999; found: 302.1003.

Ethyl (R)-5-Oxo-1-(3-(trifluoromethyl)phenyl)pyrrolidine-2-carboxylate (**7b**, Yellow Oil, 10.0 g, Yield: 80%, 97.5% ee). ¹H NMR (DMSO- d_6 , 500 MHz) δ: 7.98 (1H, s), 7.71–7.65 (1H, m), 7.65–7.60 (1H, m), 7.52 (1H, d, J = 7.9 Hz), 5.15–5.10 (1H, m), 4.17–4.09 (2H, m), 2.66–2.53 (2H, m), 2.49–2.44 (1H, m), 2.17–2.07 (1H, m), 1.13 (3H, t, J = 7.3 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 125 MHz) δ: 174.4, 171.5, 139.2, 130.1, 129.4 (q, J = 32 Hz), 124.3, 124.0 (q, J = 272 Hz), 121.1 (q, J = 4 Hz), 117.1 (q, J = 4 Hz), 61.3, 60.3, 30.5, 22.2, 13.8. HRMS (ESI⁺): calcd for C₁₄H₁₄F₃NO₃ [M + H]⁺: 302.0999; found: 302.1005.

Ethyl (R)-1-(3-Fluoro-5-(trifluoromethyl)phenyl)-5-oxopyrrolidine-2-carboxylate (**7c**, Yellow Oil, 3.2 g, Yield: 58%, 97.0% ee). ¹H NMR (DMSO- d_6 , 500 MHz) δ : 7.77 (1H, s), 7.73 (1H, dd, J = 11.3 Hz, 2.1 Hz), 7.49 (1H, br d, J = 8.5 Hz), 5.21 (1H, dd, J = 8.9 Hz, 2.7 Hz), 4.14 (2H, q, J = 7.1 Hz), 2.68–2.53 (2H, m), 2.50–2.44 (1H, m), 2.18–2.10 (1H, m), 1.14 (3H, t, J = 7.0 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 125 MHz) δ : 174.7, 171.2, 162.0 (d, J = 245 Hz), 141.1 (d, J = 11 Hz), 131.1 (dq, J = 33 Hz, 10 Hz), 123.2 (dq, J = 272 Hz, 4 Hz), 112.8 (q, J = 4 Hz), 111.1 (d, J = 26 Hz), 108.5 (dq, J = 25 Hz, 4 Hz), 61.4, 60.2, 30.6, 22.1, 13.8. HRMS (ESI⁺): calcd for C₁₄H₁₃F₄NO₃ [M + H]⁺: 320.0904; found: 320.0910.

Ethyl (R)-1-(3-Chloro-5-fluorophenyl)-5-oxopyrrolidine-2-carboxylate (**7d**, Yellow Oil, 3.1 g, Yield: 55%, 95.5% ee). ¹H NMR (DMSO- d_6 , 500 MHz) δ: 7.48 (1H, s), 7.43 (1H, td, *J* = 11.1 Hz, 2.1 Hz), 7.24 (1H, td, *J* = 8.5 Hz, 2.1 Hz), 5.12 (1H, dd, *J* = 9.2 Hz, 3.1 Hz), 4.15 (2H, q, *J* = 7.3 Hz), 2.66–2.52 (2H, m), 2.48–2.40 (1H, m), 2.14–2.07 (1H, m), 1.15 (3H, t, *J* = 7.0 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 125 MHz) δ: 174.5, 171.2, 162.1 (d, *J* = 245 Hz), 141.0 (d, *J* = 13 Hz), 134.1 (d, *J* = 13 Hz), 116.1 (d, *J* = 4 Hz), 111.8 (d, *J* = 25 Hz), 106.2 (d, *J* = 26 Hz), 61.2, 60.2, 30.6, 22.1, 13.9. HRMS (ESI⁺): calcd for C₁₃H₁₃ClF₃NO₃ [M + H]⁺: 286.0641; found: 286.0643.

Ethyl (R)-5-Oxo-1-(pyridin-4-yl)pyrrolidine-2-carboxylate (**7e**, White Solid, 12.5 g, Yield: 76%, 97.4% ee). ¹H NMR (DMSO- d_6 , 500 MHz) δ : 8.54–8.44 (2H, m), 7.57–7.49 (2H, m), 5.07 (1H, dd, J = 9.2 Hz, 2.4 Hz), 4.21–4.08 (2H, m), 2.70–2.54 (2H, m), 2.49– 2.42 (1H, m), 2.17–2.08 (1H, m), 1.18 (3H, t, J = 7.3 Hz). ¹³C{¹H} NMR (DMSO- d_6 , 125 MHz) δ : 175.1, 171.2, 150.4, 145.1, 113.1, 61.5, 59.3, 30.8, 22.2, 13.9. HRMS (ESI⁺): calcd for C₁₂H₁₄N₂O₃ [M + H]⁺: 235.1077; found: 235.1080.

Ethyl (R)-5-Oxo-1-(pyridin-3-yl)pyrrolidine-2-carboxylate (**7f**, Yellow Oil, 10.6 g, Yield: 91%, 99.3% ee). ¹H NMR (DMSO- d_{60} , 500 MHz) δ: 8.69 (1H, d, J = 2.4 Hz), 8.36 (1H, dd, J = 4.6 Hz, 1.5 Hz), 7.96–7.89 (1H, m), 7.45–7.39 (1H, m), 5.12–5.05 (1H, m), 4.18–4.07 (2H, m), 2.63–2.52 (3H, m), 2.19–2.08 (1H, m), 1.14 (3H, t, J = 7.0 Hz). ¹³C{¹H} NMR (DMSO- d_{60} , 125 MHz) δ: 174.4, 171.4, 145.6, 142.3, 135.1, 128.2, 123.6, 61.3, 59.9, 30.1, 22.5, 13.9. HRMS (ESI⁺): calcd for C₁₂H₁₄N₂O₃ [M + H]⁺: 235.1077; found: 235.1081.

Ethyl (*R*)-5-Oxo-1-(*pyridin-2-yl*)*pyrrolidine-2-carboxylate* (**7***g*, Yellow Oil, 11.1 g, Yield: 95%, 98.6% ee). ¹H NMR (DMSO- d_{cb} , 500 MHz) δ: 8.34–8.22 (2H, m), 7.89–7.77 (1H, m), 7.13 (1H, ddd, *J* = 7.3 Hz, 4.9 Hz, 1.2 Hz), 4.99 (1H, dd, *J* = 9.8 Hz, 3.1 Hz), 4.18–4.08 (2H, m), 2.71–2.54 (2H, m), 2.43 (1H, qd, *J* = 13.0 Hz, 9.5 Hz), 2.06–1.97 (1H, m), 1.16 (3H, t, *J* = 7.0 Hz). ¹³C{¹H} NMR (DMSO- d_{cb} , 125 MHz) δ: 174.4, 172.0, 150.7, 147.4, 138.0, 119.7, 113.5, 60.7, 59.1, 31.5, 21.2, 13.9. HRMS (ESI⁺): calcd for C₁₂H₁₄N₂O₃ [M + H]⁺: 235.1077; found: 235.1079.

Ethyl (*R*)-5-Oxo-1-phenylpyrrolidine-2-carboxylate (**7h**, 15.0 g, Yield: 87%, 95.7% ee). ¹H NMR (DMSO- $d_{6^{j}}$ 500 MHz) δ: 7.47 (2H, br d, *J* = 7.3 Hz), 7.36 (2H, t, *J* = 7.9 Hz), 7.20-7.11 (1H, m), 4.99–4.93 (1H, m), 4.19–4.06 (2H, m), 2.63–2.52 (2H, m), 2.48–2.42 (1H, m), 2.12–2.01 (1H, m), 1.13 (3H, t, *J* = 7.3 Hz). ¹³C{¹H} NMR (DMSO- d_{6} , 125 MHz) δ: 173.9, 171.8, 138.5, 128.8, 124.8, 120.9, 61.1, 60.6, 30.5, 22.4, 13.9. HRMS (ESI⁺): calcd for C₁₃H₁₅NO₃ [M + H]⁺: 234.1125; found: 234.1132.

Ethyl (R)-1-(3-Methoxyphenyl)-5-oxopyrrolidine-2-carboxylate (**7i**, Yellow Oil, 14.6 g, Yield: 93%, 99.8% ee). ¹H NMR (DMSO- d_{6} , 500 MHz) δ: 7.27 (1H, t, J = 8.2 Hz), 7.20–7.13 (1H, m), 6.98

(1H, dd, *J* = 8.2 Hz, 1.5 Hz), 6.75 (1H, dd, *J* = 8.2 Hz, 2.1 Hz), 4.95 (1H, dd, *J* = 8.9 Hz, 2.7 Hz), 4.12 (2H, q, *J* = 6.9 Hz), 3.73 (3H, s), 2.62–2.52 (1H, m), 2.49–2.40 (2H, m), 2.10–2.02 (1H, m), 1.21–1.11 (3H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 125 MHz) δ : 174.0, 171.8, 159.4, 139.6, 129.6, 112.8, 110.0, 107.0, 61.2, 60.7, 55.1, 30.7, 22.3, 13.9. HRMS (ESI⁺): calcd for C₁₄H₁₇NO₄ [M + H]⁺: 264.1230; found: 264.1232.

Ethyl (*R*)-1-(2-*Methoxyphenyl*)-5-oxopyrrolidine-2-carboxylate (*7j*, Yellow Oil, 14.8 g, Yield: 80%, 99.7% ee). ¹H NMR (DMSO $d_{6^{i}}$ 500 MHz) δ: 7.33–7.26 (1H, m), 7.21 (1H, dd, *J* = 7.6 Hz, 1.5 Hz), 7.09 (1H, d, *J* = 7.9 Hz), 6.95 (1H, dt, *J* = 7.6 Hz, 1.2 Hz), 4.68– 4.60 (1H, m), 4.04 (2H, qq, *J* = 10.9 Hz, 7.0 Hz), 3.78 (3H, s), 2.50– 2.34 (3H, m), 2.15–2.05 (1H, m), 1.09 (3H, t, *J* = 7.0 Hz). ¹³C{¹H} NMR (DMSO- $d_{6^{i}}$ 125 MHz) δ: 174.3, 171.6, 154.3, 129.5, 128.7, 125.6, 120.2, 112.2, 60.9, 55.6, 29.1, 22.9, 13.9. One carbon signal under solvent multiplet. HRMS (ESI⁺): calcd for C₁₄H₁₇NO₄ [M + H]⁺: 264.1230; found: 264.1234.

Representative Experimental Procedure for Reductive Coupling/ Cyclization of Ester Containing Aryl Lactams with Primary Amines (for Table 6). To a 2-dram vial equipped with a magnetic stir bar was added Schwartz's reagent (155 mg, 0.6 mmol, 1.5 equiv; preferably less than 1 month old) as a solid in the air. The vial was then sealed with a red open top cap (with a solvent-resistant Teflon cover), and a N_2 flow ran through the sealed vial for 5 min. The N_2 line was then removed, and THF (2 mL) was added through a syringe under positive pressure. The reaction mixture was cooled to 0 °C in an icewater cooling bath and subjected to vigorous stirring, followed by the injection of a solution of the substrate (0.4 mmol in 1 mL of THF). After 1 h at 0 °C, to the milky white suspension was added neat amine coupling partner (1.6 mmol, 4 equiv). The vial was then removed from the ice-water bath, and STAB (254 mg, 1.2 mmol, 3 equiv) was added in one portion as a solid to the reaction mixture. The vial was capped and stirred at 80 °C for 18 h. A sample was taken from the vial to be analyzed by LCMS. Upon reaction completion, water (1 mL) was added to the vial. The mixture was stirred for 10 min before the addition of EtOAc (2 mL). The layers were separated, and the aqueous phase was extracted with additional portions of EtOAc (3 mL \times 3 mL). The combined organic extracts were dried (MgSO₄) and filtered. The filtrate was concentrated. The residue was purified by silica gel column chromatography (12g silica gel column) with an automatic purification system (grading from 0% EtOAc/heptanes to 30% EtOAc/heptanes over 18 min). The fractions were collected and dried to give the desired lactam products as white solids.

(*R*)-1-(4-Methoxybenzyl)-3-((4-(trifluoromethyl)phenyl)amino)piperidin-2-one (**8***a*; 113.9 mg, 75% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 7.36 (2H, br d, *J* = 8.3 Hz), 7.29–7.17 (2H, m), 6.92–6.86 (2H, m), 6.77 (2H, br d, *J* = 8.5 Hz), 4.53 (2H, s), 4.16 (1H, br dd, *J* = 10.2 Hz, 5.9 Hz), 3.79–3.76 (3H, m), 3.36–3.33 (2H, m), 2.23 (1H, br dd, *J* = 12.7 Hz, 5.1 Hz), 1.96–1.85 (2H, m), 1.84–1.68 (1H, m). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 172.3, 160.8, 152.3, 130.7, 130.3, 127.4 (q, *J* = 4 Hz), 126.8 (q, *J* = 269 Hz), 119.5 (q, *J* = 32 Hz), 115.2, 113.8, 55.8, 54.8, 51.1, 48.1, 28.7, 21.8. HRMS (ESI⁺): calcd for C₂₀H₂₂N₂F₃O₂ [M + H]⁺: 379.1628; found: 379.1623. Product enantiomer excess: 100% ee.

(*R*)-1-(4-Methoxybenzyl)-3-((3-(trifluoromethyl)phenyl)amino)piperidin-2-one (**8b**; 105.9 mg, 70% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.27 (1H, t, *J* = 8.0 Hz), 7.21 (2H, d, *J* = 8.3 Hz), 6.97 (1H, d, *J* = 7.6 Hz), 6.90-6.80 (4H, m), 4.64-4.50 (2H, m), 3.89 (1H, dd, *J* = 11.3 Hz, 5.4 Hz), 3.81 (3H, s), 3.38-3.23 (2H, m), 2.52-2.42 (1H, m), 1.99-1.90 (2H, m), 1.70-1.58 (1H, m). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 170.0, 159.1, 147.7, 131.5 (q, *J* = 32 Hz), 129.6, 129.5, 128.8, 124.3 (q, *J* = 273 Hz), 116.7, 114.3 (q, *J* = 4 Hz), 114.0, 109.3 (q, *J* = 4 Hz), 55.2, 54.3, 50.2, 46.3, 27.5, 20.5. HRMS (ESI⁺): calcd for C₂₀H₂₂N₂F₃O₂ [M + H]⁺: 379.1628; found: 379.1623. Product enantiomer excess: 100% ee.

(*R*)-3-((3-Fluoro-5-(trifluoromethyl)phenyl)amino)-1-(4methoxybenzyl)piperidin-2-one (**8***c*; 104.5 mg, 66% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.21 (2H, d, *J* = 8.3 Hz), 6.88 (2H, d, *J* = 8.6 Hz), 6.68-6.62 (2H, m), 6.48 (1H, d, *J* = 11.0 Hz), 5.39 (1H, br s), 4.56 (2H, d, *J* = 14.4 Hz), 3.86 (1H, br dd, *J* = 11.2 Hz, 5.1 Hz), 3.81 (3H, s), 3.36–3.24 (2H, m), 2.51–2.39 (1H, m), 1.99–1.88 (2H, m), 1.70–1.55 (1H, m). ${}^{13}C{}^{1}H$ NMR (CDCl₃, 100 MHz): δ 169.5, 164.6 (d, *J* = 244 Hz), 159.1, 149.6 (d, *J* = 11 Hz), 132.7 (dq, *J* = 33 Hz, 11 Hz), 129.4, 128.6, 123.5 (dq, *J* = 272 Hz, 4 Hz), 114.0, 105.9 (t, *J* = 3 Hz), 102.6 (d, *J* = 26 Hz), 101.1 (dd, *J* = 26 Hz, 4 Hz), 55.2, 54.1, 50.1, 46.1, 27.2, 20.4. HRMS (ESI⁺): calcd for C₂₀H₂₁N₂F₄O₂ [M + H]⁺: 397.1534; found: 397.1528. Product enantiomer excess: 100% ee.

(*R*)-3-((3-Chloro-5-fluorophenyl)amino)-1-(4-methoxybenzyl)piperidin-2-one (**8d**; 105.7 mg, 73% Yield). ¹H NMR (DMSO- d_6 , 400 MHz): δ 7.19 (2H, d, *J* = 8.3 Hz), 6.89 (2H, d, *J* = 8.6 Hz), 6.56 (1H, s), 6.46–6.40 (3H, m), 4.49–4.38 (2H, m), 4.15–4.09 (1H, m), 3.73 (3H, s), 3.27–3.15 (2H, m), 2.12–2.06 (1H, m), 1.84–1.77 (2H, m), 1.63–1.57 (1H, m). ¹³C{¹H} NMR (CD₃OD, 125 MHz): δ 172.2, 166.3, 164.4, 160.7, 152.3 (d, *J* = 13 Hz), 136.4 (d, *J* = 14 Hz), 130.5 (d, *J* = 45 Hz), 115.2, 110.1 (d, *J* = 2 Hz), 104.6 (d, *J* = 25 Hz), 99.2 (d, *J* = 25 Hz), 55.8, 54.8, 51.0, 48.0, 28.8, 21.7. HRMS (ESI⁺): calcd for C₁₉H₂₁CIFN₂O₂ [M + H]⁺: 363.1270; found: 363.1265. Product enantiomer excess: 94% ee.

(*R*)-1-(4-Methoxybenzyl)-3-(pyridin-4-ylamino)piperidin-2-one (**8e**; 90.8 mg, 73% Yield). ¹H NMR (DMSO- $d_{6^{+}}$ 400 MHz): δ 8.71 (1H, d, *J* = 8.3 Hz), 8.24 (1H, br d, *J* = 6.8 Hz), 8.12 (1H, br d, *J* = 6.8 Hz), 7.20 (2H, d, *J* = 7.2 Hz), 7.08-6.99 (1H, m), 6.97-6.93 (1H, m), 6.92-6.87 (2H, m), 4.57 (1H, ddd, *J* = 10.6 Hz, 8.3 Hz, 6.0 Hz), 4.51-4.40 (2H, m, 2H), 3.74 (3H, s), 3.31-3.19 (2H, m), 2.13 (1H, br dd, *J* = 12.5 Hz, 5.6 Hz), 1.92-1.83 (2H, m), 1.82-1.70 (1H, m). ¹³C{¹H} NMR (DMSO- $d_{6^{+}}$ 100 MHz): δ 167.5, 158.6, 158.5, 140.3, 138.7, 129.2, 129.0, 113.9, 109.9, 106.2, 55.1, 52.2, 48.9, 46.4, 26.9, 20.4. HRMS (ESI⁺): calcd for C₁₈H₂₁N₃O₂ [M + H]⁺: 312.1707; found: 312.1700. Product enantiomer excess: 100% ee.

(*R*)-1-(4-*Methoxybenzyl*)-3-(*pyridin-3-ylamino*)*piperidin-2-one* (*8f*, 95.8 mg, 77% Yield). ¹H NMR (DMSO- $d_{6^{j}}$ 400 MHz): δ 8.04 (1H, d, *J* = 2.4 Hz), 7.75 (1H, d, *J* = 4.4 Hz), 7.20 (2H, d, *J* = 8.3 Hz), 7.08–6.97 (2H, m), 6.90 (2H, d, *J* = 8.3 Hz), 6.02 (1H, d, *J* = 7.1 Hz), 4.50–4.39 (2H, m), 4.08 (1H, dt, *J* = 10.4 Hz, 6.4 Hz), 3.74 (3H, s), 3.30–3.16 (2H, m), 2.11 (1H, br d, *J* = 12.7 Hz, 5.1 Hz), 1.89–1.80 (2H, m), 1.69–1.58 (1H, m). ¹³C{¹H} NMR (DMSO- $d_{6^{j}}$ 100 MHz): δ 169.6, 158.4, 144.5, 136.8, 135.8, 129.3, 129.1, 123.3, 118.1, 113.8, 55.0, 52.3, 48.8, 46.3, 27.4, 20.3. HRMS (ESI⁺): calcd for C₁₈H₂₁N₃O₂ [M + H]⁺: 312.1707; found: 312.1700. Product enantiomer excess: 98% ee.

(*R*)-1-(4-Methoxybenzyl)-3-(pyridin-2-ylamino)piperidin-2-one (**8***g*; 81.0 mg, 65% Yield). ¹H NMR (CD₃OD, 500 MHz): δ 7.95–7.92 (1H, m), 7.44 (1H, ddd, *J* = 8.5 Hz, 6.7 Hz, 1.8 Hz), 7.24 (2H, d, *J* = 8.6 Hz), 6.91–6.87 (2H, m), 6.61 (1H, d, *J* = 8.5 Hz), 6.59–6.56 (1H, m), 4.60–4.49 (m, 2H), 4.48 (1H, ddd, *J* = 10.4 Hz, 6.1 Hz), 3.78 (3H, s), 3.35–3.32 (2H, m), 2.29–2.21 (1H, m), 1.96–1.89 (2H, m), 1.86–1.76 (1H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 125 MHz): δ 169.8, 158.4, 158.3, 147.1, 136.5, 129.4, 129.0, 113.8, 111.8, 109.0, 55.0, 51.1, 48.9, 46.5, 27.8, 20.8. HRMS (ESI⁺): calcd for C₁₈H₂₁N₃O₂ [M + H]⁺: 312.1707; found: 312.1701. Product enantiomer excess: 88% ee.

(*R*)-1-(4-Methoxybenzyl)-3-(phenylamino)piperidin-2-one (**8**h; 63.2 mg, 51% Yield). ¹H NMR (CD₃OD, 400 MHz): δ 7.20 (2H, d, *J* = 8.6 Hz), 7.07 (2H, t, *J* = 7.7 Hz), 6.90 (2H, d, *J* = 8.3 Hz), 6.66 (2H, d, *J* = 8.1 Hz), 6.55 (1H, t, *J* = 7.2 Hz), 5.70 (1H, d, *J* = 6.6 Hz), 4.53-4.40 (2H, m), 4.04-3.94 (1H, m), 3.74 (3H, s), 3.27-3.16 (2H, m), 2.17-2.10 (1H, m), 1.86-1.79 (2H, m), 1.65-1.57 (1H, m). ¹³C{¹H} NMR (DMSO-*d*₆, 100 MHz): δ 169.8, 158.4, 148.2, 129.4, 129.1, 128.7, 115.9, 113.8, 112.6, 55.0, 52.9, 48.9, 46.2, 27.5, 20.2. HRMS (ESI⁺): calcd for C₁₉H₂₂N₂O₂ [M + H]⁺: 311.1754; found: 311.1747. Product enantiomer excess: 100% ee.

(*R*)-1-(4-Methoxybenzyl)-3-((3-methoxyphenyl)amino)piperidin-2-one (**8***i*; 74.8 mg, 55% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.21 (2H, d, *J* = 8.6 Hz), 7.11 (1H, t, *J* = 8.2 Hz), 6.87 (2H, d, *J* = 8.3 Hz), 6.33-6.31 (2H, m), 6.24 (1H, s), 4.60-4.52 (2H, m), 3.85 (1H, dd, *J* = 11.3 Hz, 5.6 Hz), 3.81 (3H, s), 3.78 (3H, s), 3.33-3.22 (2H, m), 2.53-2.46 (1H, m), 1.95-1.88 (2H, m), 1.65-1.59 (1H, m). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 170.4, 160.7, 159.1, 148.9, 129.9, 129.5, 128.9, 114.0, 106.8, 102.9, 99.7, 55.3, 55.1, 54.7, 50.2, 46.3, 27.8, 20.5. HRMS (ESI⁺): calcd for $C_{20}H_{25}N_2O_3\ [M + H]^+:$ 341.1860; found: 341.1853. Product enantiomer excess: 100% ee.

(*R*)-1-(4-Methoxybenzyl)-3-((2-methoxyphenyl)amino)piperidin-2-one (**8***j*: 47.6 mg, 35% Yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.23 (2H, d, *J* = 8.6 Hz), 6.90–6.86 (3H, m), 6.80 (1H, d, *J* = 7.6 Hz), 6.73 (1H, t, *J* = 7.6 Hz), 6.62 (1H, d, *J* = 7.8 Hz), 4.61–4.53 (2H, m), 3.88 (3H, s), 3.87–3.83 (1H, m), 3.81 (3H, s), 3.34–3.23 (2H, m), 2.52–2.43 (1H, m), 1.98–1.89 (2H, m), 1.78–1.66 (1H, m). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 170.3, 159.0, 147.6, 137.4, 129.6, 129.0, 120.9, 117.4, 114.0, 110.2, 109.5, 55.4, 55.3, 54.7, 50.1, 46.5, 28.0, 20.6. HRMS (ESI⁺): calcd for C₂₀H₂₅N₂O₃ [M + H]⁺: 341.1860; found: 341.1853. Product enantiomer excess: 100% ee.

Preparation and Isolation of Compound 9. The compound was prepared following the same general procedure that was used for the preparation of compound 8a except that the reaction mixture was stirred at 22 °C instead of 80 °C after the addition of STAB. After keeping the reaction mixture at 22 °C overnight, 9 was present in the reaction mixture as a major component. The reaction was worked-up following the same protocol, and the crude was purified by massdirected prep-HPLC (SunFire C18 column, 60 mL/min flow rate, 10% acetonitrile in water to 70% acetonitrile in water, with 0.1% TFA modifier) to give ethyl (R)-5-((4-methoxybenzyl)amino)-2-((4-(trifluoromethyl)phenyl)-amino)pentanoate as a colorless oil. Yield was not determined since it was isolated just for characterization.

Ethyl (*R*)-5-((4-Methoxybenzyl)amino)-2-((4-(trifluoromethyl)phenyl)amino)pentanoate (**9**). ¹H NMR (CD₃OD, 400 MHz): δ 7.41–7.36 (4H, m), 6.98–6.94 (2H, m), 6.71 (2H, br d, *J* = 12.0 Hz), 4.20–4.08 (5H, m), 3.79 (3H, s), 3.07–3.03 (2H, m), 2.02–1.82 (4H, m), 1.22 (3H, t, *J* = 8.0 Hz). ¹³C{¹H} NMR (CD₃OD, 100 MHz): δ 174.9, 162.3, 152.1, 132.6, 127.6 (q, *J* = 3 Hz), 126.6 (q, *J* = 267 Hz), 124.3, 120.2 (q, *J* = 36 Hz), 115.7, 113.6, 62.6, 56.9, 55.9, 52.0, 47.8, 30.4, 24.0, 14.6. HRMS (ESI⁺): calcd for C₂₂H₂₇N₂F₃O₃ [M + H]⁺: 425.2047; found: 425.2051.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.9b00816.

Descriptions of LCMS and HPLC methods; chiral SFC traces of the chiral piperidinone products in Table 6; and ¹H and ¹³C NMR spectra of the products (PDF)

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