Synthesis, Characterization, and Comparative Study of Some Heterocyclic Compounds Containing Isoniazid and Nicotinic Acid Hydrazide Moieties

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Abstract—Some new derivatives of six-membered heterocyclic compounds containing isoniazid and nicotinic acid hydrazide fragments have been synthesized according to green procedures with excellent yields. The structures of the synthesized compounds were confirmed by ¹H and ¹³C NMR, IR, and mass spectra and elemental analyses. The compounds were screened for their in vitro antibacterial and antifungal activities. The results showed that the isoniazid derivatives are more active than their analogs containing a nicotinic hydrazide moiety.

Keywords: isoniazid, nicotinic hydrazide, antimicrobial activity

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INTRODUCTION

Investigation of heterocyclic compounds is an interesting topic from both theoretical and practical points of view. There are a huge number of pharmacologically active heterocyclic compounds, many of which are in regular clinical use. Nitrogen heterocycles are most copious in nature than those containing oxygen or sulfur. Heterocyclic compounds are crucial to life in different ways. Most of sugars and their derivatives, including vitamins and some members of vitamin B group, have heterorings with a nitrogen atom. Analysis of the US FDA-approved drugs database shows that 59% of small-molecule drugs contain nitrogen heteroatom in a heterocycle [1]. Nitrogen-containing sixmembered heterocyclic compounds have acquired enormous significance in the field of drug discovery. In this work we report the synthesis and comparative study of antimicrobial activities of two types of sixmembered heterocyclic compounds derived from isoniazid and nicotinic acid hydrazide. Isoniazid is isonicotinic acid hydrazide which plays a vital role in the manufacture of various drugs such as anticancer, antitubercular, antifungal, antibacterial, and antiviral agents [2-5]. Nicotinic acid hydrazide which is isomeric to isoniazid has multiple applications and has long been known as biologically active compound. Pyridine derivatives have remarkable pharmaceutical importance because of their biological activity as anti-HIV [6], antitubercular, antimicrobial [7], antidiabetic [8], anti-inflammatory [9], antiplasmodial [10], and anticancer [11] activities. Our goal was to synthesize and evaluate biological activity of some isoniazid and nicotinic acid hydrazide derivatives.

RESULTS AND DISCUSSION

In this study, new series of bipyrazole derivatives bearing isoniazid and nicotinic acid hydrazide moieties were synthesized as shown in Scheme 1. The starting compounds, 1*H*-pyrazole-4-carbaldehydes **4a** and **4b** were prepared by the Vilsmeier–Haack reaction according to literature procedure [12]. The key intermediates, chalcones **6a–6d** were synthesized in good to excellent yields by base-catalyzed Claisen–Schmidt condensation of aldehydes **4a** and **4b** with 2-acetylthiophene (**5a**) and 2-acetylfuran (**5b**) in the presence of NaOH (5 mol %) in aqueous ethanol at room temperature. Finally, cyclization of **6a–6d** with isoniazid and nicotinic acid hydrazide using NaOH as catalyst in ethanol





R = H (a, b), MeO (c, d); X = S (a, c), O (b, d).

at room temperature afforded target compounds **8a–8d** and **9a–9d**, respectively (Scheme 1).

The structure of **8a–8d** and **9a–9d** was confirmed by elemental analyses and FT-IR, ¹H and ¹³C NMR, and mass spectra. In the IR spectra of **8a–8d** and **9a– 9d**, the C–H stretching band was observed at 2925– 3025 cm⁻¹, and the C=N and C=O stretchings were observed at 1587–1610 and 1685–1710 cm⁻¹, respectively. In the ¹H NMR spectra of these compounds, the 4-H protons of the dihydropyrazole ring resonated as multiplets at δ 3.29–3.45 and 4.05-4.17 ppm. The 5-H proton of the same ring appeared as a multiplet at δ 5.82–5.99 ppm due to vicinal coupling with the two magnetically nonequivalent protons on C⁴. The mass spectra of all compounds **8** and **9** showed the molecular ion peak in conformity with the assigned structure.

All compounds **8a–8d** and **9a–9d** were screened for their antibacterial and antifungal activities using the broth dilution method recommended by the National Committee for Clinical Laboratory Standards (NCCLS) [13, 14]. Mueller–Hinton broth was used as the nutrient medium for the test bacteria, and Sabouraud dextrose broth was used for fungi. The inoculum size for the test strains was adjusted to 10⁸ CFU per milliliter according to the McFarland turbidity standards. Each compound was dissolved in DMSO to a stock concentration of 2000 µg/mL. Primary and secondary screenings were performed. All compounds were screened for their antibacterial activity against Bacillus subtilis (MTCC 441), Clostridium tetani (MTCC 449), Staphylococcus aureus (MTCC 96), Escherichia coli (MTCC 443), Salmonella typhi (MTCC 98), Candida albicans (MTCC 227), and Trichophyton rubrum (MTCC 296) at concentrations of 1000, 500, and 250 µg/mL for primary screening. Stock solutions were diluted with DMSO to get the desired concentration. The compounds that showed antimicrobial activity in the primary screening were further screened at concentrations of 200, 100, 62.5, 50, and 25 µg/mL. A 10-µL portion of each solution was inoculated in a 96-well plate, and microbial growth was noted after 24 and 48 h. The lowest concentration at which no visible growth (turbidity) was observed was considered as the mini-

Compd. no.	Gram-positive bacteria			Gram-negative bacteria			Fungi	
	B. subtilis	C. tetani	S. aureus	E. coli	S. typhi	V. cholerae	C. albicans	T. rubrum
8a	250	250	500	200	200	250	>1000	>1000
8b	100	50	500	500	500	100	500	500
8c	250	100	125	200	250	200	1000	500
8d	200	250	62.5	250	250	200	1000	1000
9a	500	250	250	250	250	500	1000	1000
9b	500	200	200	200	200	500	500	500
9c	250	200	250	200	250	250	1000	>1000
9d	125	500	500	125	100	100	250	100
Ampicillin	100	250	100	100	100	250	-	_
Chloramphenicol	50	50	50	50	50	50	-	_
Ciprofloxacin	50	100	25	25	25	50	-	_
Norfloxacin	10	50	10	10	10	100	-	_
Nystatin	_	—	_	_	—	_	100	100
Griseofulfin	_	_	_	_	—	_	500	100

Table 1. In vitro antimicrobial activity of compounds 8a-8d and 9a-9d (MIC, µg/mL)

mum inhibitory concentration (MIC). Ampicillin, chloramphenicol, ciprofloxacin, and norfloxacin were used as standard antibacterial drugs, and nystatin and griseofulvin were used as standard antifungal drugs. The results are summarized in Table 1.

Isoniazid derivatives 8a-8d showed a good antibacterial activity compared to nicotinic acid hydrazide derivatives 9a-9d. Compound 8b was found to be equipotent to ampicillin against gram-positive B. subtilis (MIC 100 µg/mL), and its activity against C. tetani (MIC 50 µg/mL) was higher than those of ampicillin (MIC 250 µg/mL) and ciprofloxacin (MIC 100 μ g/mL) and was comparable to that of norfloxacin. The activity of 8c against C. tetani (MIC 100 µg/mL) was similar to that of ciprofloxacin. Compound 8d was more active than ampicillin against S. aureus (MIC 62.5 and 100 µg/mL, respectively), whereas its activity against gram-negative E. coli was comparable to the activity of ampicillin (MIC 100 µg/mL). The synthesized isoniazid derivatives showed moderate to good antifungal activity, whereas the nicotinic acid hydrazide derivatives showed moderate to low antifungal activity. Thus, isoniazid derivatives 8a-8d proved to be more effective against all microorganisms than nicotinic hydrazide derivatives 9a–9d.

EXPERIMENTAL

All reagents (analytical grade) were purchased from Sigma–Aldrich and were used without further purification. The progress of reactions was monitored by thinlayer chromatography (TLC) on aluminum plates coated with silica gel 60 F_{254} (Merck); layer thickness 0.25 mm. The components were visualized by exposure to UV light or iodine vapor. The melting points were measured in open capillary tubes on an Electrothermal melting point apparatus. The IR spectra were recorded with a FTIR MB 3000 spectrophotometer using ZnSe optics (490–8500 cm⁻¹). The mass spectra were recorded on a Shimadzu LCMS 2010 spectrometer (Japan). The ¹H and ¹³C NMR spectra were recorded in DMSO- d_6 on a Bruker Avance 400 spectrometer at 400 and 100 MHz, respectively, using the residual proton and carbon signals of the solvent as internal standard. The elemental analysis was done with a Perkin Elmer 2400 Series II elemental analyzer (USA).

General procedure for the synthesis of 1-phenyl-2-[1-(4-R-phenyl)ethylidene]hydrazines 3a and 3b. Phenylhydrazine (0.1 mol) and 3-4 drops of acetic acid were added to a solution of acetophenone or *p*-methoxyacetophenone (0.1 mol) in anhydrous ethanol (20 mL), and the mixture was stirred at 70–80°C for 1 h. The mixture was then left to stand at room temperature, and the solid product was filtered off, washed with ethanol, and dried. Compounds **3a** and **3b** were sufficiently pure and were used without further purification.

General procedure for the synthesis of 3-(4-R-phenyl)-1-phenyl-1*H*-pyrazole-4-carbaldehydes 4a and 4b. Cold DMF (0.4 mol) was added dropwise with stirring over a period of 30 min to phosphoryl chloride (0.4 mol), and the mixture was stirred for further 45 min at 0°C. Compound **3a** or **3b** (0.08 mol) was then added, and the mixture was allowed to warm up to room temperature, heated at 90°C for 4 h, cooled, and poured into a mixture of crushed ice and water. The solid product was filtered off, dried, and recrystallized from chloroform-methanol (1:1).

General procedure for the synthesis of (E)-3-[1-phenyl-3-(4-R-phenyl)-1H-pyrazol-4-yl]-1-[thiophen(or furan)-2-yl]prop-2-en-1-ones 6a-6d. Equimolar amounts of compound 4a or 4b and 2-acetylthiophene (5a) or 2-acetylfuran (5b) were added to a solution of sodium hydroxide (5 mol %) in a waterethanol mixture (1:2, 30 mL). The mixture was stirred at room temperature for 1-2 h, and the solid product was filtered off, and recrystallized from methanol.

(*E*)-3-(1,3-Diphenyl-1*H*-pyrazol-4-yl)-1-(thiophen-2-yl)prop-2-en-1-one (6a). Yield 92%, mp 139–141°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1694 (C=O), 1567 (C=C). ¹H NMR spectrum, δ , ppm: 7.10–8.18 m (15H, H_{arom}, CH=CH), 9.35 s (1H, 5-H). ¹³C NMR spectrum, δ_C , ppm: 118.5, 119.4, 121.5, 122.2, 122.6, 126.1, 126.6, 127.3, 128.4, 128.6, 128.9, 129.5, 130.4, 131.5, 132.2, 133.5, 134.3, 139.5, 150.1, 150.4, 162.9, 186.4.

(*E*)-3-(1,3-Diphenyl-1*H*-pyrazol-4-yl)-1-(furan-2-yl)prop-2-en-1-one (6b). Yield 90%, mp 142–144°C. IR spectrum, v, cm⁻¹: 3022 (C–H_{arom}), 1690 (C=O), 1560 (C=C). ¹H NMR spectrum, δ , ppm: 7.09–8.17 m (15H, H_{arom}, CH=CH), 9.35 s (1H, 5-H). ¹³C NMR spectrum, δ_C , ppm: 118.2, 120.2, 121.9, 122.8, 123.3, 126.6, 126.8, 127.2, 128.3, 128.5, 128.8, 129.6, 130.4, 131.6, 132.4, 133.3, 134.6, 139.2, 150.4, 151.8, 163.2, 186.6.

(*E*)-3-[3-(4-Methoxyphenyl)-1-phenyl-1*H*-pyrazol-4-yl]-1-(thiophen-2-yl)prop-2-en-1-one (6c). Yield 92%, mp 137–139°C. IR spectrum, v, cm⁻¹: 3015 (C–H_{arom}), 1694 (C=O), 1549 (C=C). ¹H NMR spectrum: 3.85 s (3H, OCH₃), 7.12–8.13 m (14H, H_{arom}, CH=CH), 9.36 s (1H, 5-H). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 56.6, 118.6, 119.7, 121.7, 122.3, 122.8, 126.2, 126.8, 127.5, 128.2, 128.5, 128.6, 129.9, 130.6, 131.7, 132.5, 133.6, 134.4, 139.7, 150.3, 150.6, 163.1, 186.5.

(*E*)-1-(Furan-2-yl)-3-[3-(4-methoxyphenyl)-1phenyl-1*H*-pyrazol-4-yl]prop-2-en-1-one (6d). Yield 92%, mp 141–143°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1688 (C=O), 1550 (C=C). ¹H NMR spectrum, δ , ppm: 3.85 s (3H, OCH₃), 7.12–8.14 m (14H, H_{arom}, CH=CH), 9.35 s (1H, 5-H). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 56.5, 119.2, 119.8, 121.6, 122.5, 123.1, 126.4, 126.7, 127.4, 128.2, 128.6, 129.1, 129.8, 130.4, 131.5, 132.2, 133.8, 134.6, 139.4, 150.0, 150.8, 163.3, 186.4

General procedure for the synthesis of compounds 8a–8d and 9a–9d [15, 16]. A mixture of compound 6a–6d, isoniazid (7a) or nicotinic acid hydrazide (7b), aqueous sodium hydroxide, and ethanol (10 mL) was stirred at room temperature for 8–9 min. After completion of the reaction, the solid product was filtered off and recrystallized from methanol.

{**1'**,**3'**-**Diphenyl-5-(thiophen-2-yl)-3,4-dihydro-1'***H*,**2***H*-[**3**,**4'-bipyrazol**]-**2-yl**}(**pyridin-4-yl**)**methanone (8a).** Yield 86%, mp 226–228°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1695 (C=N), 1560 (C=C). ¹H NMR spectrum, δ, ppm: 3.36 d.d (1H, 4-H), 4.07 d.d (1H, 4-H), 5.82 d.d (1H, 5-H), 7.10–8.76 m (18H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 40.6, 61.5, 117.2, 119.9, 121.7, 123.1, 124.4, 125.8, 126.2, 127.2, 129.2, 129.3, 140.8, 149.7, 155.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 475.15 (100.0), 476.15 (30.3), 477.14 (4.5), 477.15 (2.7), 476.14 (1.8), 477.15 (1.7), 478.15 (1.4). Calculated for C₂₈H₂₁N₅OS: *M* 475.57.

{5-(Furan-2-yl)-1',3'-diphenyl-3,4-dihydro-1'*H*,2*H*-[3,4'-bipyrazol]-2-yl}(pyridin-4-yl)methanone (8b). Yield 88%, mp 235–238°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1696 (C=N), 1563 (C=C). ¹H NMR spectrum, δ, ppm: 3.44 d.d (1H, 4-H), 4.16 d.d (1H, 4-H), 5.92 d.d (1H, 5-H), 7.04–8.77 m (18H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 38.1, 55.8, 59.1, 109.4, 109.9, 114.2, 117.2, 119.9, 121.7, 123.2, 126.2, 128.5, 129.3, 139.7, 140.8, 141.7, 142.1, 149.7, 149.9, 155.6, 160.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 459.17 (100.0), 460.17 (30.3), 461.18 (2.7), 460.17 (1.8), 461.18 (1.7). Calculated for C₂₈H₂₁N₅O₂: *M* 459.51.

{3'-(4-Methoxyphenyl)-1'-phenyl-5-(thiophen-2yl)-3,4-dihydro-1'H,2H-[3,4'-bipyrazol]-2-yl}(pyridin-4-yl)methanone (8c). Yield 85%, mp 237–239°C. IR spectrum, v, cm⁻¹: 3015 (C–H_{arom}), 1601 (C=N), 1567 (C=C). ¹H NMR spectrum, δ , ppm: 3.35 d.d (1H, 4-H), 3.84 s (3H, OCH3), 4.12 d.d (1H, 4-H), 5.89 d.d (1H, 5-H), 7.03–8.77 m (17H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 40.6, 55.8, 61.5, 114.8, 117.2, 119.9, 121.7, 123.2, 124.4, 125.3, 125.8, 126.2, 127.4, 128.5, 129.3, 139.7, 149.7, 155.6, 167.2. Mass spectrum (ESI), *m/z* ($I_{\rm rel}$, %): 505.16 (100.0), 506.16 (31.4), 507.15 (4.5), 507.16 (2.7), 507.16 (2.0), 506.15 (1.8), 508.16 (1.4). Calculated for C₂₉H₂₃N₅O₂S: *M* 505.60.

{5-(Furan-2-yl)-3'-(4-methoxyphenyl)-1'-phenyl-3,4-dihydro-1'*H*,2*H*-[3,4'-bipyrazol]-2-yl}(pyridin-4-yl)methanone (8d). Yield 82%, mp 226–228°C. IR spectrum, v, cm⁻¹: 3012 (C–H_{arom}), 1605 (C=N), 1561 (C=C). ¹H NMR spectrum, δ , ppm: 3.37 d.d (1H, 4-H), 3.82 s (3H, OCH₃), 4.12 d.d (1H, 4-H), 5.84 d.d (1H, 5-H), 7.03–8.76 m (17H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 38.1, 56.5, 109.4, 109.9, 117.2, 119.9, 121.7, 123.2, 126.2, 127.5, 128.7, 129.2, 133.2, 139.7, 140.8, 141.7, 142.1, 149.7, 149.9, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 459.17 (100.0), 460.17 (30.3), 461.18 (2.7), 460.17 (1.8), 461.18 (1.7). Calculated for C₂₉H₂₃N₅O₃: *M* 459.54.

{1,3'-Diphenyl-5-(thiophen-2-yl)-3,4-dihydro-1'*H*,2*H*-[3,4'-bipyrazol]-2-yl}(pyridin-3-yl)methanone (9a). Yield 87%, mp 233–235°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1695 (C=N), 1550 (C=C). ¹H NMR spectrum, δ , ppm: 3.36 d.d (1H, 4-H), 4.07 d.d (1H, 4-H), 5.82 d.d (1H, 5-H), 6.93–9.02 m (18H, H_{arom}). ¹³C NMR spectrum, δ_{C} , ppm: 40.6, 61.1, 117.2, 119.9, 123.2, 124.4, 125.1, 125.8, 126.2, 127.2, 127.4, 127.5, 128.7, 129.2, 129.3, 129.9, 130.7, 133.2, 135.7, 139.7, 148.1, 148.7, 149.9, 155.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 475.15 (100.0), 476.15 (30.3), 477.14 (4.5), 477.15 (2.7), 476.14 (1.8), 477.15 (1.7), 478.15 (1.4). Calculated for C₂₈H₂₁N₅OS: *M* 475.57.

{5-(Furan-2-yl)-1',3'-diphenyl-3,4-dihydro-1'*H*,2*H*-[3,4'-bipyrazol]-2-yl}(pyridin-3-yl)methanone (9b). Yield 84%, mp 226–228°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1695 (C=N), 1550 (C=C). ¹H NMR spectrum, δ, ppm: 3.44 d.d (1H, 4-H), 4.16 d.d (1H, 4-H), 5.92 d.d (1H, 5-H), 7.10–9.02 m (18H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 38.1, 55.8, 58.7, 109.4, 109.4, 109.9, 114.8, 117.2, 119.9, 121.7, 123.2, 125.3, 126.2, 128.5, 129.3, 130.7, 135.3, 139.7, 140.8, 141.7, 142.1, 148.1, 148.7, 155.6, 160.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 459.17 (100.0), 460.17 (30.3), 461.18 (2.7), 460.17 (1.8), 461.18 (1.7). Calculated for C₂₈H₂₁N₅O₂: *M* 459.51,

{3'-(4-Methoxyphenyl)-1'-phenyl-5-(thiophen-2yl)-3,4-dihydro-1'*H*,2*H*-[3,4'-bipyrazol]-2-yl}(pyridin-3-yl)methanone (9c). Yield 86%, mp 237–239°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1695 (C=N), 1550 (C=C). ¹H NMR spectrum, δ , ppm: 3.36 d.d (1H, 4-H), 3.84 s (3H, OCH₃), 4.13 d.d (1H, 4-H), 5.84 d.d (1H, 5-H), 7.03–8.76 m (18H, H_{arom}). ¹³C NMR spectrum, δ_{C} , ppm: 38.1, 55.8, 58.7, 109.4, 109.4, 109.9, 114.8, 117.2, 119.9, 121.7, 123.2, 125.3, 126.2, 128.5, 129.3, 130.7, 135.3, 139.7, 140.8, 141.7, 142.1, 148.1, 148.7, 155.6, 160.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 505.16 (100.0), 506.16 (31.4), 507.15 (4.5), 507.16 (2.7), 507.16 (2.0), 506.15 (1.8), 508.16 (1.4). Calculated for C₂₉H₂₃N₅O₂S: *M* 505.60. **{5-(Furan-2-yl)-3'-(4-methoxyphenyl)-1'-phenyl-3,4-dihydro-1'***H***,2***H***-[3,4'-bipyrazol]-2-yl}(pyridin-3-yl)methanone (9d).** Yield 82%, mp 236–238°C. IR spectrum, v, cm⁻¹: 3017 (C–H_{arom}), 1696 (C=N), 1552 (C=C). ¹H NMR spectrum, δ, ppm: 3.44 d.d (1H, 4-H), 3.84 s (3H, OCH₃), 4.16 d.d (1H, 4-H), 5.92 d.d (1H, 5-H), 7.04–8.76 m (17H, H_{arom}). ¹³C NMR spectrum, $\delta_{\rm C}$, ppm: 38.1, 55.8, 58.7, 109.4, 109.4, 109.9, 114.8, 117.2, 119.9, 121.7, 123.2, 125.3, 126.2, 128.5, 129.3, 130.7, 135.3, 139.7, 140.8, 141.7, 142.1, 148.1, 148.7, 155.6, 160.6, 167.2. Mass spectrum (ESI), *m/z* (*I*_{rel}, %): 489.18 (100.0), 490.18 (31.4), 491.19 (4.7), 490.18 (1.8). Calculated for C₂₉H₂₃N₅O₃: *M* 489.18.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Vitaku, E., Smith, D., and Njardarson, J., J. Med. Chem., 2014, vol. 57, p. 10257. https://doi.org/10.1021/jm501100b
- Sinha, N., Jain, S., Tilekar, A., Upadhayaya, R.S., Kishore, N., Jana, G.H., and Arora, S.K., *Bioorg. Med. Chem. Lett.*, 2005, vol. 15, p. 1573. https://doi.org/10.1016/j.bmcl.2005.01.073
- Bayrak, H., Demirbas, A., Demirbas, N., and Karaoglu, S.A., *Eur. J. Med. Chem.*, 2009, vol. 44, p. 4362. https://doi.org/10.1016/j.ejmech.2009.05.022
- 4. Jaju, S., Palkar, M., Maddi, V., Ronad, P., Mamledesai, S., Satyanarayana, D., and Ghatole, M., Arch. Pharm. (Weinheim, Ger.), 2009, vol. 342, p. 723. https://doi.org/10.1002/ardp.200900001
- Judge, V., Narasimhan, B., Ahuja, M., Sriram, D., Yogeeswari, P., de Clercq, E., Pannecouque, C., and Balzarini, J., *Med. Chem. Res.*, 2011, vol. 21, p. 1451. https://doi.org/10.1007/s00044-011-9662-9
- Desai, N.C., Kotadiya, G.M., and Trivedi, A.R., *Bioorg. Med. Chem. Lett.*, 2014, vol. 24, p. 3126. https://doi.org/10.1016/j.bmcl.2014.05.002
- Barakat, A., Soliman, M., Al-Majid, A.M., Lotfy, G., Ghabbour, H.A., Fun, H.-K., Yousuf, S., Choudhary, M.I., and Wadood, A., *J. Mol. Struct.*, 2015, vol. 1098, p. 365. https://doi.org/10.1016/j.molstruc.2015.06.037
- 8. Hanna, M.M., *Eur. J. Med. Chem.*, 2012, vol. 55, p. 12. https://doi.org/10.1016/j.ejmech.2012.06.048

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- Kaur, H., Machado, M., Kock, C., Smith, P., Chibale, K., Prudêncio, M., and Singh, K., *Eur. J. Med. Chem.*, 2015, vol. 101, p. 266. https://doi.org/10.1016/j.ejmech.2015.06.045
- Yadlapalli, R.K., Chourasia, O.P., Vemuri, K., Sritharan, M., and Perali, R.S., *Bioorg. Med. Chem. Lett.*, 2012, vol. 22, p. 2708. https://doi.org/10.1016/j.bmcl.2012.02.1
- Bariwal, J.J., Malhotra, M., Molnar, J., Jain, K.S., Shah, A.K., and Bariwal, J.B., *Med. Chem. Res.*, 2012, vol. 21, p. 4002. https://doi.org/10.1007/s00044-011-9925-5
- Kira, M.A., Abdel-Rahman, M.O., and Gadalla, K.Z., *Tetrahedron Lett.*, 1969, vol. 10, p. 109. https://doi.org/10.1016/S0040-4039(01)88217-4

- Performance Standards for Antimicrobial Susceptibility Testing: Twelfth Informational Supplement (2002) NCCLS (National Committee for Clinical Laboratory Standards). ISBN 1-56238454-6. M100-S12 (M7)
- 14. Patel, H.B., Gohil, J.D., and Patel, M.P., *Monatsh. Chem.*, 2017, vol. 148, p. 1057. https://doi.org/10.1007/s00706-016-1875-7
- Kalaria, P.N., Makawana, J.A., Satasia, S.P., Raval, D.K., and Zhu, H.-L., *Med. Chem. Commun.*, 2014, vol. 5, p. 1555. https://doi.org/10.1039/C4MD00238E
- Sharma, P.K., Kumar, S., Kumar, P., Kaushik, P., Kaushik, D., Dhingra, Y., and Aneja, K.R., *Eur. J. Med. Chem.*, 2010, vol. 45, p. 2650. https://doi.org/10.1016/j.ejmech.2010.01.059