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ARTICLE TYPE

A highly selective phenothiazine-based fluorescence 'turn-on' indicator based on cyanide-promoted novel protection/deprotection mechanism[†]

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A cyanide anion (CN⁻)-triggered deprotection of NH-protected phenothiazine, (*E*)-10-(*10H*-phenothiazin-3'-yl)propenal, has been discovered as a novel mechanism for the highly selective fluorescence detection of CN⁻ under ambient conditions. The ¹⁰ present protocol may pave the way for its broad applications in organic synthesis in near future.

Cyanide anion (CN⁻) is highly toxic to living organisms that damage by absorption through the lungs, gastrointestinal track, or skin and can kill mammals upon binding to a heme unit.¹ Despite ¹⁵ this, the industrial use of mass quantities of cyanide is one of the most routine operations in many chemical processes, essential to a healthy economy, including gold mining, electroplating, and plastic production.² Nevertheless, this mechanization or industrialization, certainly, has resulted 'an unsweet smell' of ²⁰ growing threat of cyanide exposure into the environment. The third world is particularly at risk, because the flooding can result in a rapid rise in the concentration of cyanide in drinking water. Besides, the cyanide has also been used as a chemical warfare reagent and even as a terror material.³ The importance of both these ²⁵ areas to health and security has made a cogent need for the development of cyanide-selective sensors or indicators.

Traditionally, hydrogen bonding or supramolecular interactions have been used for the design of CN⁻ sensors.⁴ These approaches, however, usually display poor selectivities over other common

- ³⁰ anions. The shift towards an emphasis on selectivity issue has resulted in extensive research efforts, particularly, in recent years, devoted to the development of reaction-based indicators that take advantage of nucleophilic property of CN⁻ to effect chemical changes in a chemodosimetric fashion.⁵ In this milieu, endeavours
- ³⁵ have been made using C-B bond formation,⁶ C-S bond formation,⁷ and predominantly C-C bond forming reactions,⁸ utilizing systems based on benzyl,^{8a,b} calix[4]pyrrole,^{8c} acridinium salt,^{8d} coumarin,^{8e} BODIPY,^{8f} oxazine,^{8g} squaraine,^{8h} phenothiazine,⁸ⁱ 2,4,6-triphenylthiopyrylium,^{8j} and triazolopyridinium salt,^{8k} to
- ⁴⁰ name but a few. Effective as these systems are, it would be advantageous to ripen a new cyanide-driven reaction that (*i*) has a vibrant mechanistic signature, (*ii*) exhibits high selectivity, (*iii*) operates at room temperature, and (*iv*) could be exploited for the development of off-the-shelf optical sensors for CN⁻.
- ⁴⁵ With these realizations as well as based on our experience in developing optical probes or method for detection of biologically important species, ^{8i,9} herein, we report a CN⁻-selective indicator, namely, (*E*)-10-(*10H*-phenothiazin-3'-yl)propenal (**PTZ-AL**), a



⁵⁰ Scheme 1 Synthesis of (*E*)-10-(*10H*-phenothiazin-3'-yl)propenal (PTZ-AL) and its CN⁻-promoted deprotection reaction.

NH-protected phenothiazine (**PTZ**). The **PTZ-AL** contains an α , β -unsaturated aldehyde group as a putative CN⁻-triggered deprotection subunit and thus works in a chemodosimetric fashion.

- PTZ-AL was synthesized following the route as outlined in 55 Scheme 1. Briefly, a Vilsmeier-Hacck formylation reaction of PTZ employing 3-(dimethylamino)propenal as an amide in dichloroethane (DCE) solution followed by hydrolysis of the resulting intermediate in the presence of NaHCO₃ solution 60 afforded PTZ-AL in appreciable yield. The structure and purity of PTZ-AL was confirmed using NMR spectroscopy and mass spectrometry (see ESI[†]). The unequivocal proof about the structure of PTZ-AL was accessed by X-ray diffraction analysis. Single crystals suitable for X-ray structure analysis were grown by 65 layering *n*-hexane onto a THF solution followed by slow evaporation of solutions at room temperature over a period of 2 days. The PTZ-AL crystallizes in the monoclinic system, space group P 1 21/c 1, where the crystal structure shows a typical transconfiguration with respect to the C1'-C2' bond (see ESI⁺).
- ⁷⁰ In order to evaluate the nature of the interaction of CN⁻ with **PTZ-AL**, we commenced our investigations by first monitoring the ¹H NMR spectral changes produced *via* the incremental addition of tetra-*n*-butylammonium cyanide (TBACN) to a solution of **PTZ-AL** in (CD₃)₂SO) at room temperature.
- ⁷⁵ As shown in Fig. 1, upon the addition of ~ 1.5 equiv. of TBACN, the aldehyde, vinylic and aromatic protons of **PTZ-AL** at $\delta = 9.37$ (H₁[•]), 8.01, 5.72 (H₃[•], H₂[•]), and 7.35-7.62 ppm (H₁₋₄ and H₆₋₉), respectively, were almost completely disappeared. Concomitantly, however, new resonances at $\delta = 6.57-6.94$ ppm emerged. On ⁸⁰ careful examination, the newly appeared resonances were found identical with those of the aromatic protons of standard compound **PTZ**, suggesting that, indeed, **PTZ-AL** was transformed into **PTZ** (Scheme 1) rather than the typical addition of CN⁻ to the α,β unsaturated carbonyl moiety as reported previously.¹⁰

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Fig. 1 Partial ¹H NMR (600 MHz, (CD₃)₂SO) spectral changes upon the addition of cyanide anion (CN⁻) to PTZ-AL (10.0 mM). In red: signals disappearing. In blue: signals appearing during titration.

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In order to evaluate the CN--selective nature towards deprotection of PTZ-AL, the ¹H NMR titration was also performed with F⁻ which often obscures the detection of CN⁻, especially, in organic solvents acting as a strong competitor of CN^{-.8i} In marked contrast to what was seen with CN⁻, all the 10 resonance lines shown within 5-10 ppm were totally unchanged after the addition of 5 equiv. of F⁻ (Fig. S6⁺). Furthermore, the ¹H NMR spectrum of resulting mixture did not show any noticeable changes even on keeping the solution for more than two weeks, indicating that F⁻ is completely silent for this particular reaction. 15 All other studied anions ranging from Cl⁻, Br⁻, I⁻, HSO₄⁻, NO₃⁻ to H₂PO₄⁻, and AcO⁻ gave similar responses as that of F⁻,

authenticating PTZ-AL to be exceptionally selective for CN⁻. It is worth mentioning here that the deprotection mechanism of PTZ-AL fairly works even in aqueous solution or with inorganic CN-²⁰ (NaCN) as evidenced by ¹H NMR spectroscopy (Fig. S7[†]).

With these intriguing results in hand, the quantitative analytical behaviour of CN--triggered transformation of PTZ-AL into PTZ was further examined by fluorescence emission spectroscopy. In this context, it should be noted that whereas PTZ exhibits blue flu-



Fig. 2 Fluorescence titration (emission) spectra of PTZ-AL $(5.0 \times 10^{-5} \text{ M})$ in acetonitrile solution upon incremental addition of TBACN ($0 \rightarrow 5$ equiv.) Inset (right): Changes in the emission intensity of PTZ-AL vs. equiv. of CN⁻ ($\lambda_{ex} = 320$ nm and $\lambda_{em} = 443$ nm).

30 -orescence, PTZ-AL appears as a transparent and non-emissive compound in solution, presumably, due to the protection of 10N-Hgroup. Therefore, a fluorescence change should ensue due to the CN--triggered transformation of 10N-C=C-CHO into 10N-H group as confirmed above by ¹H NMR titrations. Upon incremental 35 addition of CN⁻, the acetonitrile solution of PTZ-AL resulted in a significant enhancement of emission intensity positioned at $\lambda =$ 443 nm (Fig. 2). When 5 equiv. of CN- was added, the emission intensity reached its maximum with ca. 43-fold enhancement factor validating the results of ¹H NMR spectroscopy. Notably, 40 there was a linear dependence of the fluorescence intensity on the CN^- concentration in the range of 2.3×10^{-5} M to 2.3×10^{-4} M (R^2 = 0.9856) with a detection limit of 1.3×10^{-5} M. In line with this, upon gradual introduction of CN⁻, a significant increase (relatively weak than that of only in acetonitrile) in the fluorescence emission 45 was also observed in a mixture of acetonitrile-water (2:3, v/v; LOD = 3.0×10^{-5} M, $\lambda_{em} = 443$ nm) as well as acetonitrile-PBS buffer (7:3, v/v, pH 7.4; LOD = 2.7×10^{-5} M, $\lambda_{em} = 439$ nm) solutions, which demonstrate, that PTZ-AL can be effectively used in more realistic environments (Fig. S8 and S9[†]).



Fig. 3 Fluorescence emission intensity spectra of PTZ-AL $(5.0 \times 10^{-5} \text{ M})$ in acetonitrile solution recorded after the addition of 5 equiv. of the tetrabutylammonium salts of various anions (CN⁻, AcO⁻, F⁻, Cl⁻, Br⁻, I⁻, HSO4⁻, NO3⁻ and H2PO4⁻. Inset: Corresponding visual fluorescence responses of PTZ-AL to different anions at room temperature.



Scheme 2 Proposed mechanism for CN-triggered transformation of PTZ-AL into PTZ.

To further evaluate the selectivity of **PTZ-AL** towards CN⁻, we also measured fluorescence intensity changes upon addition of a variety of anions. Specifically, 5 equiv. of each anion were added to an acetonitrile solution of **PTZ-AL** (5.0×10^{-5} M) and incubated for 10 min at room temperature, before being subjected to spectral analysis. The fluorescence emission intensity of **PTZ-AL** ¹⁰ enhanced remarkably only after the addition of CN⁻. The other studied anions (AcO⁻, F⁻, Cl⁻, Br⁻, I⁻, HSO₄⁻, NO₃⁻, and H₂PO₄⁻), inorganic sulphide, and (bio)thiols did not cause any remarkable changes in the fluorescence emission intensity (Fig. 3a, S10-17†). The fact that **PTZ** exhibits blue fluorescence also allowed the CN⁻-¹⁵ driven transformation of **PTZ-AL** to be followed visually by making the use of a short UV hand lamp (UV-GL 25; UV-254/365 nm) (Fig. 3b).

The mechanism of interaction of CN^- with **PTZ-AL** is proposed in Scheme 2, which involves initial nucleophilic attack of CN^- on ²⁰ β -position of the aldehyde group (C₃') resulting in the formation of intermediate **A**. Under experimental conditions, intermediate A seems to be unstable and readily undergoes rearrangement, releasing 4-oxobut-2-enenitrile to produce **PTZ**.

In order to get an unequivocal proof that CN⁻ signalling is *via* ²⁵ selective CN⁻-induced deprotection of **PTZ-AL** (Scheme 2), the chemical reaction between **PTZ-AL** and TBACN (1.2 equiv.) was performed in acetonitrile solution under ambient conditions (see ESI[†]). The progress of the reaction could be easily followed by TLC. After completing the reaction, the solvent was evaporated

³⁰ and crude was column chromatographed. As expected, the isolated product turned out to be **PTZ** as confirmed by ¹H NMR and ¹³C NMR spectroscopy.

In summary, a PTZ scaffold having an α , β -unsaturated aldehyde group (**PTZ-AL**) as a putative reacting subunit towards CN⁻ was

- ³⁵ synthesized. In the presence of CN⁻, **PTZ-AL** revealed a highly selective 'turn-on' type fluorogenic signalling behaviour based on the selective and efficient cleavage of N-C bond (CN⁻-triggered deprotection), which was established by ¹H NMR titrations as well as the isolated product from a chemical reaction between **PTZ-AL**
- ⁴⁰ and TBACN. Effective as it is, however, it must be stressed that **PTZ-AL** does not serve as a typical signalling probe but can be classified as a chemodosimeter due to the irreversible nature of as-proposed chemical reaction to produce fluorescence response. Although several chemodosimetric probes for CN⁻ detection have
- ⁴⁵ been reported over the years, to the best of our knowledge, this is the first example enabling the use of a protection/deprotection mechanism for selective CN⁻ detection. The present approach holds great promises in organic synthesis and may be further developed as N–H protection/deprotection strategy. Further
- ⁵⁰ exploration of this approach, as is a search for other heterocyclic systems that might allow an exceptional CN[−] recognition, are currently underway in our laboratory.

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Notes and references

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† Electronic Supplementary Information (ESI) available: [Synthesis and characterization data of **PTZ-AL**, spectra of ¹H NMR, ¹³C NMR, and HRMS, ¹H NMR titrations and single crystal X-ray data for **PTZ-AL**. ⁶⁵ CCDC 1049885]. See DOI: 10.1039/b000000x/

- (a) S. I. Baskin and T. G. Brewer, in *Medical Aspects of Chemical and Biological Warfare*, ed. F. Sidell, E. T. Takafuji and D. R. Franz, TMM Publication, Washington DC, 1997, ch. 10, pp. 271; (b) K. W. Kulig, *Cyanide Toxicity*, U. S. Department of Health and Human Services, Atlanta, 1991.
- 2 G. C. Miller and C. A. Pritsos, Cyanide: Soc., Ind., Econ. Aspects, Proc. Symp. Annu. Meet., 2001, 73.
- 3 J. D Johnson, T. L. Meisenheimer and G. E. Isom, *Toxicol. Appl. Pharmacol.*, 1986, 84, 464A.
- 75 4 (a) M. E. Moragues, R. Martínez-Máñez and F. Sancenón, *Chem. Soc. Rev.*, 2011, **40**, 2593; (b) M. Wenzel, J. R. Hiscock, and P. A. Gale, *Chem. Soc. Rev.*, 2012, **41**, 480.
- 5 For reviews, see: (a) Y. Yang, Q. Zhao, W. Feng and F. Li, *Chem. Rev.*, 2013, **113**, 192; (b) L. E. Santos-Figueroa, M. E. Moragues, E.
- ⁸⁰ Climent, A. Agostini, R. Martínez-Máñez and F. Sancenón, *Chem. Soc. Rev.*, 2013, **42**, 3489; (c) P. A. Gale, N. Busschaert, C. J. E. Haynes, L. E. Karagiannidis and I. L. Kirby, *Chem. Soc. Rev.*, 2014, **43**, 205; (d) F. Wang, L. Wang, X. Chen and J. Yoon, *Chem. Soc. Rev.*, 2014, **43**, 4312; (e) Z. Xu, X. Chen, H. N. Kim and J. Yoon, *Chem. Soc. Rev.*, 2010, **39**, 127.; others: (f) L. Peng, M. Wang, G. Zhang, D.
- Soc. Rev., 2010, 39, 127., 00085. (1) L. Feng, M. wang, O. Zhang, D. Zhang and D. Zhu, Org. Lett., 2009, 11, 1943; (g) X. Huang, X. Gu, G. Zhang and D. Zhang, Chem. Commun., 2012, 48, 12195.
- 6 Y. Kim, H. S. Huh, M. H. Lee, I. L. Lenov, H. Y. Zhao and F. P. Gabbaï, *Chem.-Eur. J.*, 2011, **17**, 2057.
- ⁹⁰ 7 G. Qian, X. Li and Z. Y. Wang, *J. Mater. Chem.*, 2009, **19**, 522.
 ⁸ (a) J. L. Sessler and D. G. Cho, *Org. Lett.*, 2008, **10**, 73; (b) D. G. Cho, J. H. Kim and J. L. Sessler, *J. Am. Chem. Soc.*, 2008, **130**, 12163; (c)
 ⁸ S.-J. Hong, J. Yoo, S.-H. Kim, J.-S. Kim, J. Yoon and C.-H. Lee, *Chem. Commun.*, 2009, **189**; (d) Y. K. Yang and J. Tae, *Org. Lett.*,
- ⁹⁵ 2006, **8**, 5721; (e) G-J. Kim and H.-J. Kim, *Tetrahedron Lett.*, 2010,
 51, 185; (f) C.-H. Lee, H.-J. Yoon, J.-S. Shim and W.-D. Jang, *Chem. Eur. J.*, 2012, **18**, 4513; (g) M. Tomasulo and F. M. Raymo, *Org. Lett.*,
 2005, **7**, 4633; (h) J. V. Ros-Lis, R. Martínez-Máñez, and J. Soto,
 Chem. Commun., 2002, 2248; (i) B. Garg, L. Yan, T. Bisht, C. Zhu and
 Y.-C. Ling, *RSC Advances*, 2014, **4**, 36344-36349; (j) T. Ábalos, S.
 - Royo, R. Martínez-Máñez, F. Sancenón, J. Soto, A. M. Costero, S. Gil and M. Parra, *New. J. Chem.*, 2009, **33**, 1641; (k) T. F. Robbins, H. Qian, X. Su, R. P. Hughes and I. Aprahamian, *Org. Lett.*, 2013, **15**, 2386.
- (a) B. Garg and Y.-C. Ling, *RSC Adv.*, 2013, 3, 10150; (b) B. Garg, S.-L. Lei, S.-C. Liu, T. Bisht, J.-Y. Liu and Y.-C. Ling, *Anal. Chim. Acta*, 2012, 757, 48.
 - 10 G-J. Kim and H.-J. Kim, Tetrahedron Lett., 2010, 51, 2914.

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Graphical Abstract

A highly selective phenothiazine-based fluorescence 'turn-on' indicator based on cyanide-promoted novel protection/deprotection mechanism

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A CN⁻-triggered selective cleavage of N-C bond is described using (*E*)-10-(10*H*)-phenothiazin-3'-yl)propenal to produce selective fluorescence 'turn-on' response for CN⁻.