

# Synthesis of telluroamino acid derivatives with remarkable GPx like activity†

Antonio L. Braga,\* Eduardo E. Alberto, Letiére C. Soares, João B. T. Rocha, Jéssie H. Sudati and Daniel H. Roos

Received 28th August 2008, Accepted 3rd November 2008

First published as an Advance Article on the web 13th November 2008

DOI: 10.1039/b814990a

A series of modular telluroamino acid derivatives with remarkable GPx-like behavior was prepared in an efficient and short two-step synthesis.

Glutathione peroxidase enzymes (GPx) are excellent catalysts for antioxidant reactions. They protect our body against the potentially damaging effects of reactive oxygen species (ROS), such as hydrogen peroxide and some organic hydroperoxides, formed during aerobic metabolism. Several neurodegenerative diseases, including Alzheimer's and Parkinson's disease are linked to ROS activity.<sup>1</sup> Since the discovery that selenium plays a pivotal role in GPx enzymes, catalyzing the reduction of hydroperoxides at the expense of glutathione (GSH),<sup>2</sup> synthetic developments and design of new chalcogen-based catalytic antioxidants have attracted considerable attention.<sup>3</sup> Synthetic organoselenium and organotellurium compounds have emerged as excellent candidates to act as GPx mimics, due their well-known ability to undergo two-electron redox cycle between chalcogen (II) and (IV) species.<sup>4</sup>

The cyclic selenenamide ebselen **1** (Fig. 1), was the first synthetic compound suggested for hydrogen peroxide inactivating therapy in the presence of glutathione.<sup>5</sup>

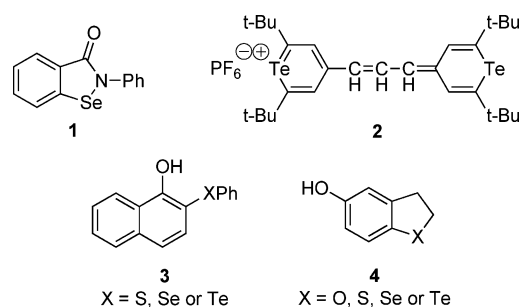


Fig. 1 Representative examples of GPx mimics.

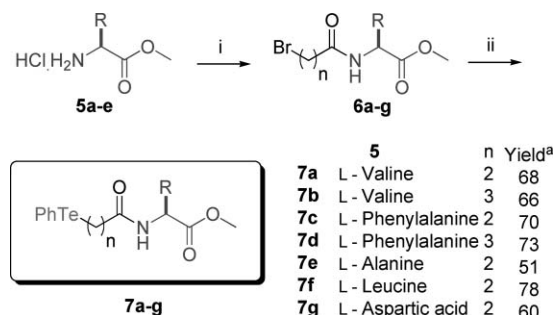
Based on the recognized GPx like activity of ebselen, several papers have appeared describing simple synthetic organoselenium compounds with this property (e.g., benzoselenazinones,<sup>6</sup> benzoselenazolinones,<sup>7</sup> camphor-derived selenenamide,<sup>8</sup> diaryl diselenides<sup>9</sup> and dendrimers with a diselenide core<sup>10</sup>).

On the other hand, the first organotellurium compound **2**, described as a GPx mimic, was reported by Detty.<sup>11</sup> After that, a series of diaryl ditellurides<sup>12</sup> and diaryl tellurides<sup>13</sup> have been reported. Engman and collaborators have developed the synthesis

and studied the antioxidant properties of compounds **3**<sup>14</sup> and **4**.<sup>15</sup> A noteworthy characteristic of these compounds is the much improved antioxidant activity of tellurides, when compared with their selenium and sulfur analogues.

Amino groups that are capable of interacting with selenium, through Se–N nonbonded interactions, are known to play a significant role in modulating the redox proprieties of seleno-based antioxidants.<sup>16</sup> However, to the best of our knowledge, there are just a few reports concerning the synthesis and GPx like evaluations of aminoacid derivatives containing selenium<sup>17</sup> and none about tellurium. As part of our growing interest in aminoacid derivatives containing chalcogen,<sup>3c,18</sup> we report herein the synthesis of novel telluroamino acid derivatives, easily obtained in a simple, modular and efficient two-step synthesis. We envisioned that this modular characteristic would allow us to investigate the structure–activity relationship of these compounds. Their GPx like activity was evaluated, and we promoted the variation of aminoacid residues, the chain length between the chalcogen atom and the aminoacid moiety in order to find a more efficient catalyst.

GPx mimics **7a–g** were prepared in two steps, in 51–78 overall yield, from readily available L-aminoesters and the appropriate bromo-carboxylic acid, as shown in Scheme 1.



<sup>a</sup> Overall yields.

**Scheme 1** Synthetic strategy to prepare compounds **7a–g**. Reagents: i) bromo-carboxylic acid, NMM, ethyl chloroformate, CHCl<sub>3</sub>; ii) PhTe<sub>2</sub>, NaBH<sub>4</sub>, THF, EtOH.

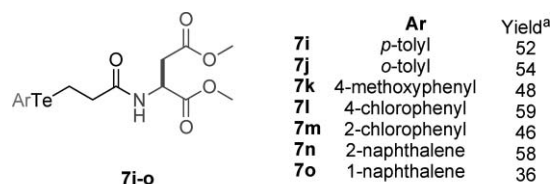
The catalytic activity of tellurides **7a–g**, as a GPx model enzyme, was evaluated according to the Tomoda method<sup>15</sup> using benzenethiol as a glutathione alternative. The reduction of H<sub>2</sub>O<sub>2</sub> was monitored through the UV absorption increase at 305 nm, due to diphenyl disulfide formation.

Our prime concern in the evaluation of these compounds as GPx mimics was the influence of the chain length between the tellurium atom and aminoacid moiety. It should be noted that compounds **7a** (n = 2) and **7b** (n = 3) derived from L-Valine

Departamento de Química, Universidade Federal de Santa Maria, 97.105–900, Santa Maria, RS, Brazil. E-mail: albraga@quimica.ufsm.br

† Electronic supplementary information (ESI) available: General procedures, <sup>1</sup>H and <sup>13</sup>C NMR spectra of selected compounds. See DOI: 10.1039/b814990a

Encouraged by these results, we next explored the effects of substituents in the aryl group attached to tellurium in **7g** derivatives. A new series of telluroaminoacids were prepared with electron donating **7i–k**, electron withdrawing **7l** and **7m**, as well as with steric hindrance substituents **7n** and **7o** (Fig. 3).

<sup>a</sup> Overall yields.

**Fig. 3**

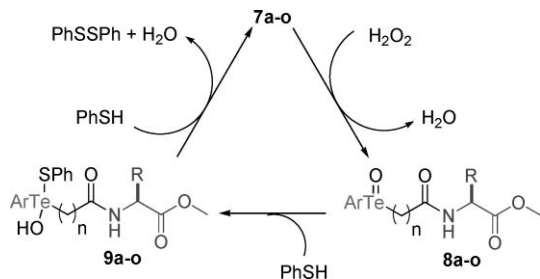
A new set of experiments to screen the GPx activity of these new catalysts was performed with **7i-o** (2 mol%), in a more concentrated solution (Table 2). It was found that the GPx behavior is strongly influenced by steric effects. The  $T_{50}$  of *para* substituted compounds were lower when compared with the *ortho* analogues (Entries 2 and 3, 5 and 6). The changing of electronic environment at the tellurium atom did not produce a pronounced change to the thiol peroxidase activity of these compounds. The  $T_{50}$  of **7g** (Ar = Ph, Entry 1), **7i** (Ar = *p*-tolyl, Entry 2), **7k** (Ar = 4-MeOC<sub>6</sub>H<sub>4</sub>, Entry 4), and **7l** (Ar = 4-ClC<sub>6</sub>H<sub>4</sub>, Entry 5) was between 2.11 and 2.98 min.

Table 2

Entry <sup>a</sup>	Catalyst <sup>b</sup>	Ar	T <sub>50</sub> <sup>c,d</sup>
1	<b>7g</b>	phenyl	2.98 (±0.21)
2	<b>7i</b>	<i>p</i> -tolyl	2.78 (±0.20)
3	<b>7j</b>	<i>o</i> -tolyl	4.78 (±0.12)
4	<b>7k</b>	4-methoxyphenyl	2.11 (±0.24)
5	<b>7l</b>	4-chlorophenyl	2.94 (±0.19)
6	<b>7m</b>	2-chlorophenyl	8.14 (±1.52)
7	<b>7n</b>	2-naphthalene	3.65 (±0.18)
8	<b>7o</b>	1-naphthalene	4.27 (±0.32)

<sup>a</sup> Under this condition addition of H<sub>2</sub>O<sub>2</sub> in the absence of telluride did not produce any significant oxidation of PhSH. <sup>b</sup> MeOH (1 mL); catalyst [0.1 mmol L<sup>-1</sup>]; PhSH [5 mmol L<sup>-1</sup>]; H<sub>2</sub>O<sub>2</sub> [5 mmol L<sup>-1</sup>]. <sup>c</sup>  $T_{50}$  is the time required, in min, to reduce the thiol concentration with 50% after the addition of H<sub>2</sub>O<sub>2</sub>. <sup>d</sup> Data in parentheses: experimental error.

Concerning mechanistic aspects, and in agreement with Detty's study,<sup>19</sup> we believe that initially Te(II) compounds **7a-o** react with H<sub>2</sub>O<sub>2</sub> to form the Te(IV) oxides **8a-o**, and H<sub>2</sub>O (Scheme 2). Addition of one equivalent of PhSH to these compounds generate tellurenyl sulfides **9a-o** which react with another equivalent of PhSH to regenerate **7a-o** to the catalytic cycle and produce PhSSPh and H<sub>2</sub>O.



Scheme 2

In conclusion, we have prepared a series of telluroamino acid derivatives, in a short, modular and efficient synthetic route. These compounds were tested as GPx mimics, catalyzing the reduction of H<sub>2</sub>O<sub>2</sub> to water at expense of thiophenol using a very low amount of catalyst. We found that the time required to reduce the concentration of the PhSH to a half, *T*<sub>50</sub>, is strongly influenced by the amino acid residue, as well as by steric effects. New studies to investigate the influence of amino acid residues of telluroamino acid derivatives have been performed in our lab using glutathione as reducing agent.

## Acknowledgements

The authors thank CNPq, CAPES, and FAPERGS for financial support.

## Notes and references

- 1 *Free Radicals in Biology*, ed. L. Flohe and W. A. Pryor, Academic Press, New York, 1982.
- 2 (a) J. T. Rotruck, A. L. Pope, H. E. Ganther, A. B. Swanson, D. G. Hafeman, W. G. Hoekstra, *Science* 1973, 179, 588; (b) L. Flohé, E. A. Günzler and H. H. Sock, *FEBS Lett.*, 1973, 32, 132; (c) *Selenium in Biology and Human Health*, ed. R. F. Burk, Springer-Verlag, New York, 1994.
- 3 (a) G. Mugesh and H. Singh, *Chem. Soc. Rev.*, 2000, 29, 347; (b) G. Mugesh, W. W. -du Mont and H. Sies, *Chem. Rev.*, 2001, 101, 2125; (c) C. W. Nogueira, G. Zeni and J. B. T. Rocha, *Chem. Rev.*, 2004, 104, 6255; (d) B. K. Sarma and G. Mugesh, *Org. Biomol. Chem.*, 2008, 6, 965.
- 4 (a) H. J. Reich, *Acc. Chem. Res.*, 1979, 12, 22; (b) K. B. Sharpless, K. M. Gordon, R. F. Lauer, D. W. Patrick, S. P. Singer and M. W. Young, *Chem. Scr.*, 1975, 8A, 9; (c) M. R. Detty, *Organometallics*, 1991, 10, 702; (d) M. R. Detty, P. B. Merkel and S. K. Powers, *J. Am. Chem. Soc.*, 1988, 110, 5920.
- 5 (a) A. Müller, E. Cadenas, P. Graf and H. Sies, *Biochem. Pharmacol.*, 1984, 33, 3235; (b) A. Wendel, M. Fausel, H. Safayhi, G. Tiegs and R. Otter, *Biochem. Pharmacol.*, 1984, 33, 3241; (c) M. J. Parnham and S. Kindt, *Biochem. Pharmacol.*, 1984, 33, 3247.
- 6 P. V. Jacquemin, L. E. Christiaens and M. J. Renson, *Tetrahedron Lett.*, 1992, 33, 3663.
- 7 V. Galet, J. L. Bernier, J. P. Hénichart, D. Lesieur, C. Abadie, L. Rochette, A. Lindenbaum, J. Chalas, J. F. R. de la Faverie, B. Pfeiffer and P. Renard, *J. Med. Chem.*, 1994, 37, 2903.
- 8 T. G. Back and B. P. Dick, *J. Am. Chem. Soc.*, 1997, 119, 2079.
- 9 (a) S. R. Wilson, P. A. Zucker, R. R. C. Huang and A. Spector, *J. Am. Chem. Soc.*, 1989, 111, 5936; (b) T. Wirth, *Molecules*, 1998, 3, 164; (c) G. Mugesh, A. Panda, H. B. Singh, N. S. Puneekar and R. Butcher, *J. Chem. Commun.*, 1998, 2227; (d) G. Mugesh, A. Panda, H. B. Singh, N. S. Puneekar and R. J. Butcher, *J. Am. Chem. Soc.*, 2001, 123, 839.
- 10 X. Zhang, H. Xu, Z. Dong, Y. Wang, J. Liu and J. Shen, *J. Am. Chem. Soc.*, 2004, 124, 10556.
- 11 M. R. Detty and S. L. Gibson, *Organometallics*, 1992, 11, 2147.
- 12 L. Engman, D. Stern, I. A. Cotgreave and C. M. Andersson, *J. Am. Chem. Soc.*, 1992, 114, 9737.
- 13 (a) L. Engman, D. Stern, M. Pelcman and C. M. Andersson, *J. Org. Chem.*, 1994, 59, 1973; (b) K. Vessman, K. Ekström, M. Berglund, C. M. Andersson and L. Engman, *J. Org. Chem.*, 1995, 60, 4461.
- 14 L. Engman, D. Stern, H. Frisell, K. Vessman, M. Berglund, B. Ek and C. M. Andersson, *Bioorg. Med. Chem.*, 1995, 3, 1255.
- 15 (a) L. Engman, M. Laws, J. Malmström, C. H. Schiesser and L. M. Zugaro, *J. Org. Chem.*, 1999, 64, 6764; (b) J. Malmström, M. Jonsson, I. A. Cotgreave, L. Hammarström, M. Sjödin and L. Engman, *J. Am. Chem. Soc.*, 2001, 123, 3434.
- 16 M. Iwaoka and S. Tomoda, *J. Am. Chem. Soc.*, 1994, 116, 2557.
- 17 (a) P. P. Phadnis and G. Mugesh, *Org. Biomol. Chem.*, 2005, 3, 2476; (b) K. P. Bhabak and G. Mugesh, *Chem. Eur. J.*, 2007, 13, 4594.
- 18 (a) A. L. Braga, J. A. Sehnem, F. Vargas and R. C. Braga, *J. Org. Chem.*, 2005, 70, 9021; (b) A. L. Braga, D. S. Lüdtke, M. W. Paixão, E. E. Alberto, H. A. Stefani and L. Juliano, *Eur. J. Org. Chem.*, 2005, 4260; (c) A. L. Braga, L. Wessjohann, M. W. Paixão, O. E. D. Rodrigues, A. Schneider and H. R. Appelt, *Tetrahedron Lett.*, 2006, 47, 1019; (d) A. L. Braga, D. P. Bottega, M. W. Paixão, A. M. Deobald, C. Peppe and P. H. Schneider, *J. Org. Chem.*, 2006, 71, 4305.
- 19 Y. You, K. Ahsan and M. R. Detty, *J. Am. Chem. Soc.*, 2003, 125, 4918.