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FIRST SYNTHESIS OF *TRANS*- AND *CIS*-DENDROCHRYSANINES

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Abstract – *Trans*- and *cis*-dendrochrysanines (**1** and **2**), isolated from the stems of *Dendrobium chrysanthum* Wall., were first synthesized from L-proline. The absolute configuration of **1** and **2** was established to be *S* by the syntheses.

INTRODUCTION

Trans- and *cis*-dendrochrysanines (**1** and **2**)¹ were isolated from the stems of *Dendrobium chrysanthum* Wall.² by Z. Wang and co-workers in 2005 and used in traditional Chinese medicine. These structures were identified as the *N-trans*-cinnamoyl-2-oxopropylpyrrolidine (**1**) and the *N-cis*-cinnamoyl-2-oxopropylpyrrolidine (**2**), based on extensive 2D-NMR (¹H-¹H COSY, HMQC, and HMBC) experiments. The absolute configuration of **1** and **2**, however, has been estimated by comparing their optical rotations and CD curve with those of known related compounds, *N-cis*-cinnamoyl-L-proline and *N-cis*-cinnamoyl-L-2-methyl pyrrolidine.³ In the course of studying cysteine protease inhibitors,^{4,5} we have interested in these natural products, since their α,β -unsaturated carbonyl structure is known to be a typical functional group for the thiol group of cysteine proteases. Here, we describe the first synthesis of *trans*- and *cis*-dendrochrysanines (**1** and **2**) as well as the determination of their absolute configurations, prior to medicinal studies of those natural products.

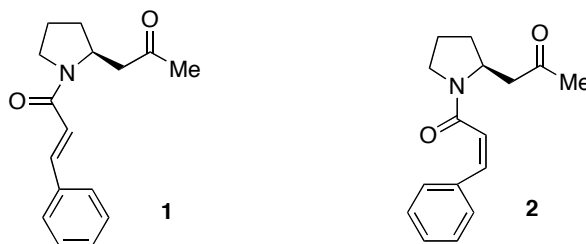
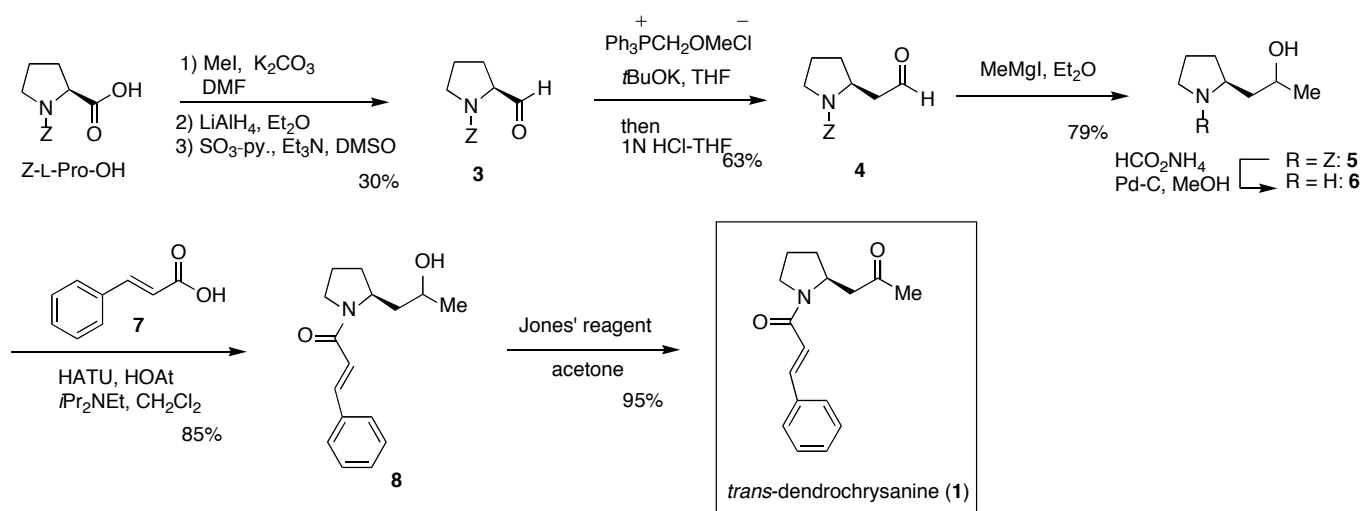


Figure Structure of *trans*-dendrochrysanine (**1**) and *cis*-dendrochrysanine (**2**)

RESULTS AND DISCUSSION

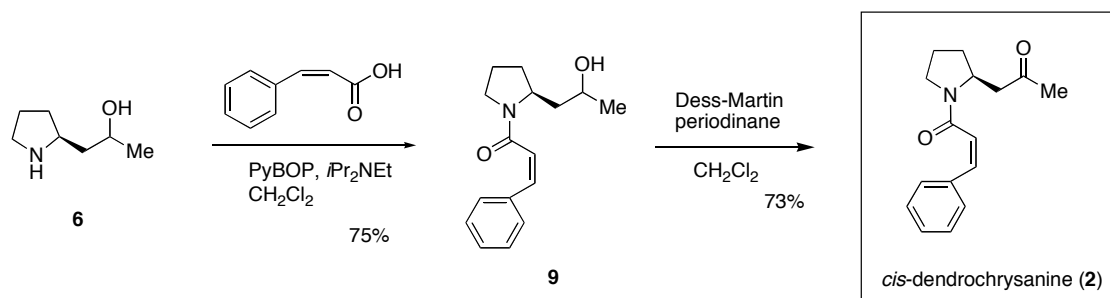
Trans-dendrochrysanine (**1**) was synthesized according to the route shown in Scheme 1. *Z*-L-Proline (*Z*-L-Pro-OH) was first transformed into *Z*-prolinal (**3**) through protection of the carboxyl group with iodomethane, and aldehyde transformation via the alcohol in a overall yield of 30%.⁶ The optical purities of **3** and the intermediate alcohol were determined to be >98%*ee* by HPLC using chiral column. A subsequent Wittig reaction with (methoxymethyl)triphenylphosphonium chloride and potassium *tert*-butoxide in THF gave a one-carbon extended methoxy olefin product, which was then treated with 1N HCl in THF to give the desired aldehyde (**4**) in 63% yield. Enantiomeric excess of aldehyde (**4**) was determined to be 98%*ee* by HPLC using chiral column. A Grignard reaction of aldehyde (**4**) with methylmagnesium iodide in ether afforded two diastereomeric alcohols (**5**) as a 1:1.5 mixture in 79% yield. Deprotection of the *Z* group in the presence of 5% palladium on carbon in a hydrogen atmosphere gave the pyrrolidine (**6**), which was coupled with *trans*-cinnamic acid (**7**) by a reaction with O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HATU)⁹/ 1-hydroxy-7-azabenzotriazole (HOAt)¹⁰/ *N,N*-diisopropylethylamine in CH₂Cl₂ to afford **8** as a diastereoisomeric mixture in 85% yield. Finally, the alcohol (**8**) was oxidized with Jones' reagent to give *trans*-dendrochrysanine (**1**), [α]_D²⁰ -11.8° (*c* 0.15, CHCl₃) [lit.; [α]_D²⁰ -19.2° (*c* 3.4, CHCl₃)] in 95% yield. The ¹H- and ¹³C-NMR spectra of the synthetic **1** were in good agreement with those of natural **1**.



Scheme 1. Synthesis of *trans*-dendrochrysanine (**1**)

Similarly, *cis*-dendrochrysanine (**2**) was synthesized from pyrrolidine (**6**) in 2 steps (Scheme 2). The pyrrolidine (**6**) was coupled with *cis*-cinnamic acid¹¹ using HATU/ HOAt/ *N,N*-diisopropylethylamine in CH₂Cl₂ as above. The condensation product, however, was obtained with only 5% yield probably due to the formation of stable adduct from the *cis*-cinnamic acid and HOAt. Desired alcohol (**9**) was obtained in 75% yield when benzotriazol-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate (PyBOP)¹² was employed instead of HOAt. Finally, oxidation of **11** with Dess-Martin periodinane¹³ in CH₂Cl₂

afforded *cis*-dendrochrysanine (**2**), $[\alpha]_D^{24} -14.5^\circ$ (*c* 0.04, CHCl_3) [lit.; $[\alpha]_D^{20} -17.7^\circ$ (*c* 3.5, CHCl_3)]. The ^1H - and ^{13}C -NMR spectra of the synthetic **2** were in good agreement with those of natural **2**.



Scheme 2. Synthesis of *cis*-dendrochrysanine (**2**)

In conclusion, we have completed the first synthesis of *trans*- and *cis*-dendrochrysanines (**1** and **2**). The absolute structure of these compounds has been established to be (2*S*)-*N*-*trans*-cinnamoly-2-oxopropylpyrrolidine for **1** and (2*R*)-*N*-*cis*-cinnamoly-2-oxopropylpyrrolidine for **2**.

EXPERIMENTAL

General Experimental Procedures. All manipulations were conducted under an inert atmosphere (N_2). All solvents were of reagent grade. THF was distilled from sodium and benzophenone ketyl. CH_2Cl_2 was distilled from CaH_2 . All commercial reagents were of the highest purity available. Analytical TLC was performed on silica gel (60 F-254, Plates 0.25 mm). Column chromatography was carried out on Wakogel 60 (particle size, 0.063-0.200 mm). ^1H -, and ^{13}C -NMR were recorded on a JEOL JNM-EX-400 or Bruker AVANCEII-300. Chemical shifts are expressed in ppm relative to TMS (0 ppm) or CHCl_3 (7.28 ppm for ^1H and 77.1 ppm for ^{13}C). IR were obtained on HORIBA FREEXACT-II FT-710 spectrometer. Optical rotations were recorded on a HORIBA SEPA-200 or SEPA-300 polarimeter at the sodium D line. Low-resolution mass spectra (LRMS) and High-resolution mass spectra (HRMS) were obtained on either a JOEL JMS-DX-303 or a JMS-AX-500 (EI or FAB). Optical purities were determined on a Hitachi LaChrom HPLC instrument equipped with the DAICEL chiral column.

(2*S*)-*N*-Benzyloxycarbonyl-prolinal (3**):** To a solution of *Z*-L-proline (43.4 g, 174 mmol) in DMF (200 mL) were added K_2CO_3 (121 g, 1.75 mmol) and MeI (16.2 mL, 260 mmol) at 0°C and the mixture was stirred for 2 h. To the mixture were added H_2O and Et_2O . The organic layer was washed with brine, dried over MgSO_4 , and evaporated in vacuo. To the residue in THF (300 mL) was added LiAlH_4 (13.2 g, 348 mmol) at 0°C and the mixture was stirred for 1 h. H_2O (15 mL) and 1N NaOH (30 mL) were added and filtered. Evaporation of the solvent provided an oil, which was purified with silica gel column chromatography (hexane:AcOEt = 4:1) to give *Z*-prolinol (24.2 g, 103 mmol, 59%) as a colorless oil: IR (film) $\nu = 3410, 3033, 2954, 1684, 1415, 1358, 1105, 769, 698\text{ cm}^{-1}$; ^1H -NMR (300 MHz, CDCl_3) $\delta =$

1.50-2.10 (m, 4H), 3.30-3.70 (m, 4H), 4.01 (brs, 1H), 4.32 (brs, 1H), 5.15 (s, 2H), 7.36 (m, 5H). Optical purity was determined to be 99%ee by HPLC using CHIRALCEL OD-H column (0.46 cm Φ $\times$ 25 cm); elution with 2% *i*-PrOH in hexane, t_R = 8.11 min for *S* isomer, t_R = 9.91 min for *R* isomer. To a solution of Z-prolinol (2.45 g, 10.5 mmol) in DMSO (30 mL) were added sulfur trioxide pyridine complex (5.00 g, 31.4 mmol) and Et₃N (13.9 mL, 98.8 mmol) and the mixture was stirred for 1 h. To the mixture were added H₂O, and Et₂O, and the organic layer was washed with brine, dried over MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 6:1) to give Z-prolinal (**3**) (1.22 g, 5.25 mmol, 51%) as a colorless oil: $[\alpha]_D^{25}$ -211.5° (c 0.10, CHCl₃); IR (film) ν = 3033, 2956, 2702, 1732, 1695, 1413, 1357, 1101, 769, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.89 (m, 2H), 2.06 (m, 2H), 3.58 (m, 2H), 4.22 (m, 0.5H), 4.33 (m, 0.5H), 5.12 (m, 2H), 7.35 (m, 5H), 9.51 (d, 0.5H, *J* = 2.4 Hz), 9.62 (d, 0.5H, *J* = 1.5 Hz); ¹³C-NMR (75 MHz, CDCl₃) δ = 23.9, 24.6, 26.7, 27.9, 46.8, 47.4, 65.0, 65.4, 67.4, 128.1, 128.2, 128.62, 136.6, 200.1; MS: *m/z* = 234 (MH⁺, 40%), 154 (70%), 91 (100%). HRMS Calcd. for C₁₃H₁₆NO₃: 234.1130. Found: 234.1125. Optical purity was determined to be 98%ee by HPLC using CHIRALCEL OJ-H column (0.46 cm Φ $\times$ 25 cm); elution with 0.5% *i*-PrOH in hexane, t_R = 22.95 min for *S* isomer, t_R = 28.64 min for *R* isomer.

(2S)-N-Benzoyloxycarbonyl-2-pyrrolidinylethanal (4): To a solution of (methoxymethyl)triphenylphosphonium chloride (2.92 g, 8.50 mmol) and potassium *tert*-butoxide (960 mg, 8.50 mmol) in THF (20 mL) was added Z-prolinal (1.00 g, 4.29 mmol), and the mixture was stirred at room temperature for 30 min. To the mixture were added H₂O and Et₂O, and the organic layer was washed with brine, and dried over MgSO₄, and evaporated in vacuo. The product was stirred in 1N HCl-THF (1:5 mixture, 10 mL) for 30 min. The mixture was extracted with Et₂O, and the organic layer was washed with brine, dried over MgSO₄. The solvent was removed, and the product was purified with silica gel column chromatography (hexane:AcOEt = 8:1) to give **4** (707 mg, 2.71 mmol, 63%) as a colorless oil, $[\alpha]_D^{25}$ +4.9° (c 1.22, CHCl₃); IR (film) ν = 3033, 2956, 2727, 1718, 1699, 1415, 1358, 1105, 769, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.89 (m, 2H), 2.16 (m, 1H), 2.53 (m, 1H), 2.97 (m, 1H), 3.46 (m, 2H), 4.34 (m, 1H), 5.14 (s, 2H), 7.36 (m, 5H), 9.69 (brs, 0.4H), 9.82 (brs, 0.6H); ¹³C-NMR (75 MHz, CDCl₃) δ = 22.8, 23.6, 31.1, 31.9, 46.3, 46.6, 48.4, 49.1, 52.1, 52.9, 66.6, 127.7, 127.9, 128.0, 128.6, 128.8, 136.7, 154.7, 200.6; MS: *m/z* = 248 (MH⁺, 45%), 154 (45%), 91 (100%). HRMS Calcd. for C₁₄H₁₈NO₃: 248.1281. Found: 248.1288. Optical purity was determined to be 98%ee by HPLC using chiral CHIRALCEL AD-H column (0.46 cm Φ $\times$ 25 cm); elution with 2% *i*-PrOH in hexane, t_R = 21.70 min for *S* isomer, t_R = 26.26 min for *R* isomer.

(2S)-N-Benzylloxycarbonyl-2-hydroxypropyrrolidine (5): To a solution of **4** (271 mg, 1.04 mmol) in THF (5 mL) was added methylmagnesium iodide (2M in Et₂O, 1.50 mL) at -20°C and mixture was stirred for 20 min. To the mixture were added saturated *aq.* NH₄Cl and Et₂O, and the organic layer was washed with brine, dried over MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 2:1) to give **5** (205 mg, 779 μmol, 75%) as a diastereomeric mixtures. Low polar **5** as a colorless oil, $[\alpha]_D^{25}$ -6.4° (*c* 0.94, CHCl₃); IR (film) ν = 3438, 3033, 2966, 1699, 1684, 1356, 1105, 769, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.21 (d, 3H, *J* = 5.7 Hz), 1.48 (m, 1H), 1.72 (m 1H), 1.96 (m, 4H), 3.43 (t, 2H, *J* = 5.7 Hz), 3.88 (brs, 1H), 4.01 (m, 1H), 5.14 (s, 2H), 7.36 (m, 5H); ¹³C-NMR (75 MHz, CDCl₃) δ = 23.0, 23.8, 24.2, 31.1, 32.0, 45.2, 46.3, 54.6, 56.1, 66.4, 66.9, 127.9, 128.0, 128.5, 136.9, 155.6; MS: *m/z* = 264 (MH⁺, 95%), 154 (70%), 91 (100%). HRMS Calcd. for C₁₅H₂₂NO₃: 264.1600. Found: 264.1598. High polar **5** as a colorless oil, $[\alpha]_D^{25}$ -4.2° (*c* 0.72, CHCl₃); IR (film) ν = 3431, 3032, 2966, 1700, 1684, 1414, 1358, 1109, 769, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.19 (d, 3H, *J* = 6.0 Hz), 1.47 (m, 2H), 1.63 (brs, 1H), 1.93 (m, 3H), 3.43 (m, 2H), 3.76 (m, 1H), 4.23 (m, 1H), 5.16 (s, 2H), 7.38 (m, 5H); ¹³C-NMR (75 MHz, CDCl₃) δ = 22.7, 23.7, 31.2, 45.6, 46.4, 54.8, 63.8, 67.2, 127.9, 128.1, 128.6, 129.0, 136.8, 157.0; MS: *m/z* = 264 (MH⁺, 50%), 154 (100%), 91 (45%). HRMS Calcd. for C₁₅H₂₂NO₃: 264.1600. Found: 264.1600.

(2S)-N-trans-Cinnamoyl-2-hydroxypropyrrolidine (8): To a solution of **5** (27 mg, 102 μmol) in MeOH (5 mL) were added HCO₂NH₄ (64 mg, 1.0 mmol) and Pd-C (20 mg). The mixture was stirred for 40 min at 50°C and the solvent was removed in vacuo. The residue was added to H₂O, extracted with AcOEt, dried over MgSO₄, and evaporated in vacuo. To the residue in CH₂Cl₂ (2 mL) were added *trans*-cinnamic acid (20 mg, 134 μmol), HATU (58 mg, 204 μmol), HOAt (28 mg, 204 μmol), and *i*Pr₂NEt (50 μL, 703 μmol), and the mixture was stirred at room temperature for 10 h. To the mixture were added H₂O and CH₂Cl₂. The organic layer was washed with brine, dried over MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 1:1) to give **8** (20 mg, 77 μmol, 75%) as a colorless oil, IR (film) ν = 3410, 2966, 1647, 1456, 1435, 1151, 843 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.18 (d, 2H, *J* = 6.1 Hz), 1.26 (d, 1H, *J* = 6.1 Hz), 1.55-2.10 (m, 6H), 3.62 (m, 2H), 3.80 (m, 1H), 4.40 (m, 1H), 6.72 (d, 1H, *J* = 15.4 Hz), 7.36 (m, 3H), 7.51 (m, 2H), 7.71 (d, 1H, *J* = 15.4 Hz); MS: *m/z* = 260 (MH⁺, 20%), 282 (M⁺+Na, 100%). HRMS Calcd. for C₁₆H₂₂NO₂: 260.1651. Found: 260.1653.

(2S)-N-trans-Cinnamoyl-2-oxoxypropyrrolidine (trans-dendrochrysanine; 1): To a solution of **8** (20 mg, 77 μmol) in acetone (2 mL) was added Jones' reagent at 0°C and the mixture was stirred for 20 min. To the mixture were added isopropanol and Et₂O, and the organic layer was washed with brine, dried over

MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 1:1) to give **1** (19 mg, 73 μmol, 95%) as a colorless oil, $[\alpha]_D^{25}$ -11.8° (*c* 0.15, CHCl₃); IR (film) ν = 2930, 1711, 1649, 1601, 1419, 1149, 980, 766 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ = 1.78 (m, 1H), 2.00 (m, 2H), 2.14 (m, 2H), 2.21 (s, 3H), 2.49 (dd, 1H, *J* = 16.6, 9.5 Hz), 3.28 (dd, 1H, *J* = 12.0, 7.8 Hz), 3.67 (m, 2H), 4.54 (m, 1H), 6.73 (d, 1H, *J* = 15.6 Hz), 7.39 (m, 3H), 7.54 (m, 2H), 7.70 (d, 1H, *J* = 15.4 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ = 23.7, 29.9, 30.0, 46.7, 46.8, 53.6, 118.7, 127.6, 128.5, 129.4, 134.9, 141.7, 164.5; MS: *m/z* = 131 (100%), 257 (M⁺, 30%). HRMS Calcd. for C₁₆H₁₉NO₂: 257.1416. Found: 257.1414.

(2S)-N-cis-Cinnamoyl-2-hydroxypropyrrolidine (9): To a solution of **6** (15 mg, 116 μmol) in CH₂Cl₂ (1 mL) were added *cis*-cinnamic acid (18 mg, 121 μmol), PyBOP (60 mg, 115 μmol), and *i*Pr₂NEt (50 μL, 703 μmol). The mixture was stirred at room temperature for 10 h. To the mixture were added H₂O and CH₂Cl₂, and the organic layer was washed with brine, dried over MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 1:1) to give **11** (23 mg, 87 μmol, 75%) as a colorless oil, $[\alpha]_D^{25}$ -20.0° (*c* 0.20, CHCl₃); IR (film) ν = 2923, 1712, 1612, 1434, 1371, 1153, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.56 (m, 1H), 1.73 (m, 2H), 2.07 (m, 1H), 2.19 (s, 3H), 2.38 (dd, 1H, *J* = 16.5, 9.6 Hz), 3.12 (td, 1H, *J* = 10.8, 6.6 Hz), 3.31 (m, 2H), 4.43 (m, 1H), 6.03 (d, 1H, *J* = 12.6 Hz), 6.65 (d, 1H, *J* = 12.6 Hz), 7.30 (m, 3H), 7.40 (m, 2H); ¹³C-NMR (75 MHz, CDCl₃) δ = 24.0, 30.2, 30.7, 46.6, 47.4, 53.3, 124.3, 128.38, 128.44, 128.6, 123.9, 135.7, 162.4, 207.0; MS: *m/z* = 260 (MH⁺, 20%), 282 (M⁺+Na, 100%). HRMS Calcd. for C₁₆H₂₂NO₂: 260.1651. Found: 260.1653.

(2S)-N-cis-Cinnamoyl-2-oxoxypropyrrolidine (cis-dendrochrysanine; 2): To a solution of **9** (11 mg, 42 μmol) in CH₂Cl₂ (1 mL) was added Dess-Martin periodinane (0.2 M in CH₂Cl₂, 200 μL) at 0°C and the mixture was stirred for 20 min at 0°C. To the mixture were added H₂O and CH₂Cl₂, and the organic layer was washed with brine, dried over MgSO₄, and evaporated in vacuo. The product was purified with silica gel column chromatography (hexane:AcOEt = 1:1) to give **2** (8 mg, 31 μmol, 73%) as a colorless oil, $[\alpha]_D^{25}$ -14.5° (*c* 0.04, CHCl₃); IR (film) ν = 2923, 1712, 1612, 1434, 1371, 1153, 698 cm⁻¹; ¹H-NMR (300 MHz, CDCl₃) δ = 1.56 (m, 1H), 1.73 (m, 2H), 2.07 (m, 1H), 2.19 (s, 3H), 2.38 (dd, 1H, *J* = 16.5, 9.6 Hz), 3.12 (td, 1H, *J* = 10.8, 6.6 Hz), 3.31 (m, 2H), 4.43 (m, 1H), 6.03 (d, 1H, *J* = 12.6 Hz), 6.65 (d, 1H, *J* = 12.6 Hz), 7.30 (m, 3H), 7.40 (m, 2H); ¹³C-NMR (75 MHz, CDCl₃) δ = 24.0, 30.2, 30.7, 46.6, 47.4, 53.3, 124.3, 128.38, 128.44, 128.6, 123.9, 135.7, 162.4, 207.0; FABMS: *m/z* = 258 (MH⁺, 45%), 69 (100%). HRMS Calcd. for C₁₆H₂₀NO₂: 258.1494. Found: 258.1498.

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REFERENCES AND NOTES

1. L. Yang, C. Zheng, H. Yang, M. Zhang, Z. Wang, and L. Xu, *Heterocycles*, 2005, **65**, 633.
2. L. Yang, Y. Wang, Z. M. Bi, P. Lin, Z. T. Wang, and L. S. Xu, *Chin. J. Nat. Med.*, 2004, **2**, 280 and references therein.
3. U. Ekevag, M. Elander, L. Gawell, K. Leander, and B. Luning, *Acta Chem. Scand.*, 1973, **27**, 1982.
4. J. C. Powers, J. L. Asgian, O. D. Ekici, and K. E. James, *Chem. Rev.*, 2002, **102**, 4639.
5. H. Konno, K. Kubo, H. Makabe, N. Fujii, K. Nosaka, and K. Akaji, *Peptide Sciences*, 2005, 2006, 47.
6. Although Z-prolinal (**3**) has already been prepared by several groups,^{7,8} detailed spectral data including the optical purity of **3** were not shown in these literatures.
7. A. Fleurant, J. P. Celerier, and G. Lhomme, *Tetrahedron: Asymmetry*, 1992, **3**, 695.
8. Y. Sugino and J. A. Katzenellenbogen, *Bioorg. Med. Chem. Lett.*, 1996, **6**, 361.
9. L. A. Carpino, *J. Am. Chem. Soc.*, 1993, **115**, 4397.
10. L. A. Carpino, H. Imazumi, A. El-Faham, F. J. Ferrer, C. Zhang, Y. Lee, B. M. Foxman, P. Henklein, C. Hanay, C. Mügge, H. Wenschuh, J. Klose, M. Beyerman, and M. Bienert, *Angew. Chem. Int. Ed.*, 2002, **41**, 442.
11. A. B. Concepcion, K. Maruoka, and H. Yamamoto, *Tetrahedron*, 1995, **51**, 4011.
12. R. von Eggelkraut-Gottanka, A. Klose, A. G. Beck-Sickinger, and M. Beyermann, *Tetrahedron Lett.*, 2003, **44**, 3551.
13. R. E. Ireland and L. Liu, *J. Org. Chem.*, 1993, **58**, 2899.