ChemComm

Chemical Communications

www.rsc.org/chemcomm

Volume 49 | Number 29 | 14 April 2013 | Pages 2945–3060



ISSN 1359-7345

RSCPublishing

COMMUNICATION Ligong Chen, Zhengshuang Xu, Tao Ye *et al.* Total synthesis of padanamides A and B

ChemComm

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Total synthesis of padanamides A and B⁺

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Cite this: Chem. Commun., 2013, 49, 2977

Received 8th January 2013, Accepted 12th February 2013

DOI: 10.1039/c3cc00178d

www.rsc.org/chemcomm

The first total syntheses of padanamides A and B have been achieved, unambiguously confirming their structures.

Many of the marine sediment-derived natural products¹ possess unique structural features rarely or never found among the compounds isolated from the terrestrial sources. It is thus not surprising that their molecular modes of action are sometimes also unique, prompting their investigation as potential targets for total synthesis and drug development. The vast majority of secondary metabolites, especially peptides, are derived from *Streptomyces* sp. obtained from a marine sediment.² We have been interested for some time in marine peptides and view their synthesis as a key route to structural modification and subsequent activity control.³ Here we report our efforts in the total synthesis of two highly modified linear tetrapeptides, padanamides A and B (Scheme 1).

Padanamides A and B were isolated from laboratory cultures of a *Streptomyces* sp. obtained from marine sediment collected near the passage Padana Nahua in Papua New Guinea.⁴ The gross chemical structures of padanamides A and B were established using spectral techniques. Their absolute configurations were elucidated by a combination of spectroscopic, chemical degradation and single-crystal X-ray diffraction analyses. Padanamide B was cytotoxic to Jurkat cells, while padanamide A was suspected to inhibit cysteine and methionine biosynthesis.

Our synthetic approach for padanamides A and B is outlined in Scheme 1. Padanamides A and B contain the same tripeptide unit (3)



Scheme 1 Retrosynthetic analysis of padanamides A (1) and B (2).

and differ only in the last residue at the C-termini. Consequently, a synthetic approach including the late-stage incorporation of either (*S*)-3-amino-2-oxopyrrolidine-1-carboxamide (Aopc) (4) or (*S*)-3-aminopiperidine-2,6-dione (Apd) (5) at the C-termini would potentially access both 1 and 2. We envisaged that the key intermediate 3 would arise from the assembly of acid 6 and amino alcohol 7.

The synthesis of key intermediate **3** commenced with the preparation of 2R,3R-3-hydroxyleucine. Initial experiments for the conversion of azidoester 9^5 to its corresponding acid **11** included protection of the secondary alcohol of **9** as its TBS ether followed by saponification of the ethyl ester with lithium hydroxide, which led to extensively epimerization at the azide group-bearing center. Gratifyingly, reversing the order of the protection and hydrolysis sequence proved to be more successful. Thus, treatment of azidoester **9** with lithium hydroxide afforded carboxylic acid **10**, which was then converted into the corresponding silyl ether **11** in 72% overall yield by reaction with *tert*-butyldimethylsilyl trifluoromethanesulfonate (TBSOTf) in the presence of 2,6-lutidine. No epimerization was observed in any of

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[†] Electronic supplementary information (ESI) available: Full details of experimental procedures for compounds 1–7, 10–12, 14–15, 17–19, 21, 24, 26–28, 30–31 and NMR spectra of compounds 1–7, 10–12, 14, 17–19, 21, 27–28 and 30. See DOI: 10.1039/c3cc00178d



(c) H_2 , Pd/C, MeOH, rt; (d) Fmoc-Cl, NaHCO₃, THF-H₂O, rt.

these two transformations. Reduction of the azide group of **11** followed by protection of the resulting amine with Fmoc–Cl produced acid **12** in 82% overall yield (Scheme 2).

The synthesis of protected piperazic alcohol 14 began with the enantiomerically pure (S)-tetrahydropyridazine 13,⁶ a known building block available from proline-catalyzed asymmetric α-hydrazination of 5-bromopentanal, protection of alcohol with TBS-Cl and subsequent NaH-promoted cyclization. Two Cbz protecting groups in 13 were removed under catalytic hydrogenation with 5% Pd-C to give the corresponding cyclic hydrazine. The sterically less-hindered nitrogen atom of the resulting cyclic hydrazine was then reprotected in situ with CbzCl to provide 14 in 86% yield. (S)-Tetrahydropyridazine 13 is also easily converted into the corresponding piperazic ester 15. Thus, removal of the TBS protecting group in 13 resulted in an alcohol, which was converted into the corresponding carboxylic acid via a TEMPO/NaClO/NaClO₂ promoted oxidation process.⁷ After conversion of the carboxylic acid to its methyl ester, it was then elaborated to piperazic ester 15 in 64% overall yield using an identical strategy as described for 14 (Scheme 3). Literature precedent⁸ suggested that N-2 acylation of a piperazic-acid-derivative is a challenging task due to the unusually poor nucleophilicity of the piperazic ester. In order to take advantage of the higher reactivity of the piperazic alcohol, we decided to employ 14 as the N-terminal coupling partner for the synthesis of segment 6.

With 3-hydroxyleucine-derived acids **10–12** and piperazic alcohol **14** in hand, the coupling reactions were investigated. Unfortunately, all attempts to effect condensation of acid **10** with piperazic alcohol **14** under the influence of coupling reagents (such as HATU (entry 1) and PyBOP (entry 2)) or *via* a mixed anhydride (entry 3) did not succeed. The reactions led only to decomposition of the starting material. Gratifyingly, silver cyanide-mediated coupling⁹ of **14** with acid chlorides derived from either **11** or **12** proceeded smoothly, delivering dipeptides **17** and **18** in 85% and 80% yield, respectively (Scheme 4).

The Staudinger reduction of azide **17** with PPh₃ in THF–H₂O was followed by a condensation of the resulting amine with acid chloride **20** to afford tripeptide **21** in 68% overall yield for the two steps. Selective removal of the primary TBS group of **21** with CSA/ MeOH followed by oxidation of the resulting hydroxyl group with NaIO₄ in the presence of catalytic amounts of RuCl₃ furnished carboxylic acid **6** in 68% yield over the two steps (Scheme 5).

Methyl ester of Ahmpp (7) was prepared from the known aldol adduct 24 (ref. 10) through a diastereoselective *syn* aldol addition of



Scheme 3 (a) H_2 , Pd/C; (b) Cbz–Cl, Et₃N, MeOH; (c) TBAF, THF; (d) TEMPO, NaClO/NaClO₂; (e) SOCl₂, MeOH, -20 °C.



Entry	Hleu	Conditions	yield and product
1	10	HATU, HOAt, DIPEA, DMF, rt, overnight	no desired product (16)
2	10	PyBOP, DIPEA, DMF, rt, overnight	no desired product (16)
3	10	CICO ₂ ⁱ Bu, NMM, THF, -10 °C, 0.5h; then 14, rt, overnight	no desired product (16)
4	11	1. (COCI) ₂ , DMF (cat.), CH ₂ CI ₂ , 0 °C 2. 14 , AgCN, Toluene, 80 °C, 1h	85% (17)
5	12	1. (COCI) ₂ , DMF (cat.), CH ₂ CI ₂ , 0 °C 2. 14 , AgCN, Toluene, 80 °C, 1h	80% (18)

Scheme 4 Attempted synthesis of dipeptide fragment. HATU: 2-(7-aza-1*H*-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate; HOAT: 1-hydroxy-7-azabenzotriazole; PyBOP: benzotriazol-1-yl-oxytripyrrolidino-phosphonium hexafluorophosphate.



Scheme 5 (a) PPh₃, THF–H₂O, reflux; (b) Et_3N , CH_2Cl_2 , 0 °C–rt; (c) CSA, MeOH–CH₂Cl₂; (d) NalO₄, RuCl₃, acetone–H₂O.

the boron (*Z*)-enolate derived from (*S*)-4-isopropyl-3-propionyl-2oxazolidinone (22) with *N*-Cbz-L-phenyl-alaninal 23.¹¹ Hydrolysis of the chiral auxiliary of 24 with LiOH–H₂O₂ followed by esterification of the resulting adduct with MeI, and Cbz hydrogenolysis, produced 7 in 71% yield over three steps (Scheme 6). Condensation of acid **6** with amine 7 *via* the mixed carbamic–carbonic anhydride gave rise to 25 in 86% yield. Unexpectedly, attempted hydrogenolysis of the Cbz protecting group in 25 under a variety of reaction conditions resulted in decomposition of the starting material. We then elected to cleave off the Cbz protecting group in **6** prior to coupling with amine 7. Gratifyingly, this revised approach produced the tripeptide unit **3** in 72% yield over two steps (Scheme 7).

With the key intermediate 3 in hand, we next turned our attention to the synthesis of (S)-3-amino-2-oxopyrrolidine-1-carboxamide (Aopc) (4) or (S)-3-aminopiperidine-2,6-dione (Apd) (5). Thus, treatment of the known lactam 27 with sodium hydride and





Scheme 7 (a) $CICO_2^{i}Bu$, NMM, THF; then 7, $-20 \ ^{\circ}C-rt$; (b) H₂, Pd/C, MeOH.



Scheme 8 (a) NaH, tert-butylisocyanate, THF, 0 °C; (b) TFA, anisole, ⊿ 16 h.



Scheme 9 (a) DCC, HOSu, THF–DMF, $-78~^\circ\text{C-rt};$ then $\text{CHCl}_3,$ <code>d</code> 3 h; (b) H₂, Pd/C (10%), MeOH, rt, 2 h.

tert-butylisocyanate in THF afforded urea **28** in 60% yield. Acidic cleavage of both the *tert*-butyl and Cbz groups in **28** gave rise to the required intermediate **4** in 80% yield (Scheme 8). (*S*)-*N*-Cbz- α -aminoimide **5** was obtained in 67% yield in a two-step sequence including a DCC-mediated intramolecular cyclization of *N*-Cbz- μ -glutamine leading to adduct **30**, and subsequent hydrogenolytic removal of the carboxybenzyl group (Scheme 9).

At this juncture, the time had arrived to assemble the key intermediate 3 with Aopc (4) or Apd (5) leading to padanamides A and B, respectively. Thus, saponification of the methyl ester of 3 followed by coupling with (*S*)-3-amino-2-oxopyrrolidine-1-carbox-amide (Aopc) (4) and cleavage of TBS ether provided padanamide A **1** in 35% yield over three steps. Under identical conditions, padanamide B was obtained in 45% overall yield from the condensation of **3** and **5** (Scheme 10). The spectral data for synthetic **1** and 2 (¹H, ¹³C NMR and HMRS) were identical with those published for the natural products, and the optical rotation of our products ($[\alpha]_D^{25} - 11.4$, *c* 0.2, MeOH, for padanamide A; $[\alpha]_D^{25} - 20.7$, *c* 0.2, MeOH, for padanamide B) corresponded well with the literature value (lit. $[\alpha]_D^{25} - 10.7$, *c* 5.2, MeOH, for



Scheme 10 (a) LiOH, THF–MeOH–H₂O; (b) BOPCI, 4 or 5, HOAT, NMM, THF, 0 °C–rt, 18 h; (c) 40% HF, MeCN, 0 °C, 3 h. BOPCI: bis(2-oxo-3-oxazolidinyl)phosphinic chloride.

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padanamide A; $[\alpha]_{D}^{25}$ –21.5, *c* 7.3, MeOH, for padanamide B), which led us to conclude that synthetic **1** and **2** were of the same absolute stereochemistry as natural padanamides A and B.

In summary, we have accomplished the total synthesis of padanamides A and B from the known azidoester **9** in 7.1 and 8.3% overall yield, respectively, with the longest linear sequence of 12 steps. This synthesis confirmed the structures of padanamides A and B. The extension of this chemistry toward the synthesis of padanamide analogues for further biological evaluation is underway and will be reported in due course.

We acknowledge financial support from the Hong Kong Research Grants Council (Projects: PolyU 5040/10P; PolyU 5037/ 11P, PolyU 5020/12P); Fong Shu Fook Tong Foundation and Joyce M. Kuok Foundation; The Hong Kong Polytechnic University (PolyU 5636/08M; PolyU 5634/09M); The National Science Foundation of China (21072007, 21133002 & 21272011); The Shenzhen Bureau of Science, Technology & Information (JC200903160367A, JC201005260102A, JC201005260220A, ZYC201105170351A and ZD200806170044A).

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