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An efficient Cu-catalyzed azide–alkyne cycloaddition (CuAAC) reaction in aqueous medium with a zwitterionic ligand, betaine†

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Cu-catalyzed azide–alkyne cycloaddition reaction in aqueous medium was dramatically accelerated by a simple zwitterionic additive, betaine, at ambient temperature. In the presence of betaine, 1,4-disubstituted-

1,2,3-triazoles were obtained from azides and terminal alkynes in excellent yields under 2.5-200 ppm levels

of Cu^(I) in water. This method resulted in the remarkable reduction of the Cu^(I) concentration down to ppm levels. The advantages of this protocol include the use of a conventional Cu catalytic system with commer-

cially available betaine, which is generally non-toxic for biological application. Therefore, this efficient cata-

lytic system should have broad applications in life science research and materials science.

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Introduction

Azide-alkyne cycloaddition reaction, first discovered by Huisgen,¹ has received enormous attention as a powerful tool to form 1,4-disubstituted-1,2,3-triazoles, since Sharpless and Meldal developed the Cu^(I)-catalyzed azide-alkyne 1,3cycloaddition (CuAAC) reaction.² The CuAAC reaction is easy, highly efficient, regioselective and compatible with various functional groups. Therefore, it has been extensively utilized in organic synthesis, biochemistry, medicinal chemistry, chemical biology and materials science.³ For biological application, since it is desirable for the reaction to be compatible with living systems, especially in aqueous medium and live cells, highly efficient Cu^(I) catalysts have been developed for the synthesis of triazoles to minimize the amount of Cu^(I) catalyst used. To improve the activity of the catalysts, various ligands such as polysaccharide-supported nanoparticles,⁴ polymer-embedded catalysts,⁵ and particularly catalysts with nitrogen ligands such as N-heterocyclic carbenes,⁶ polytriazoles,⁷ and tri(aminoalkyl)amine derivatives⁸ have been developed.

To further reduce the cellular toxicity of the catalysts, a dendritic nanoreactor containing intradendritic triazoles,⁹ poly(*N*-isopropylacrylamide-*co-N*-vinylimidazole) polymers,¹⁰ and tris(triazolyl)-polyethylene glycol¹¹ promoted CuAAC, which reduced the amount of Cu⁽¹⁾ catalyst for the reaction to the part per million (ppm) level. Even though these endeav-

ours led to great advances in reducing the amount of Cu catalyst, they still have shortcomings.

The major limitation of the aqueous medium of biological systems is the mixing problem with organic materials including the ligands of CuAAC catalysts, and thus even the reactive ligands work less efficiently. To overcome this mixing problem in general, various surfactants and phase transfer catalysts have been employed.¹² Recently, Lipshutz reported a CuAAC reaction using TPGS-750-M as a non-ionic surfactant in water,¹³ and Scarso also reported that organic bromide was reacted with sodium azide and a terminal alkyne in the presence of a surfactant to give 1,4-disubstituted-1,2,3-triazoles in water.¹⁴ In the same context, we have successfully introduced β -cyclodextrin as the nanoreactor and the carrier of substrates to the aqueous phase to accelerate the CuAAC reaction.¹⁵

As a part of our continuing studies on the development of readily applicable methods for triazole synthesis in water, we tested the CuAAC reaction using different surfactants such as nonionic, cationic, anionic, and zwitterionic types, with anticipation of increasing the reaction efficiency through micellar effects^{12*a*} and the on-water effect.¹⁶ Herein we report a synthetic method for triazoles using ppm levels of a Cu⁽¹⁾ catalyst in the presence of betaine, which is a readily available and biocompatible zwitterionic surfactant. With amounts of 0.0003 mol% (2.5 ppm) to 0.025 mol% (200 ppm) of Cu⁽¹⁾, the CuAAC reaction was carried out at 30 °C, efficiently affording excellent yields of triazoles.

Results and discussion

Initially with anticipation of micellar effects, we carried out CuAAC reactions with various additives such as β -cyclodextrin (the phase transfer catalyst), SDS, aliquat 128, Triton X-100,



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and betaine (anionic, cationic, non-ionic, and zwitterionic surfactants, respectively) in water. The reaction of benzyl azide **1a** with phenyl acetylene **2a** in the presence of 200 ppm of $Cu^{(1)}$ and 5 mol% of surfactant in water at 30 °C gave the corresponding triazole **3aa**, and the results are listed in the Table **1**. Interestingly, different reaction rates for triazole formation were observed depending on the surfactant type. Among them, in the case of using the zwitterionic surfactant betaine, the reaction showed a remarkable improvement in the reactivity to produce **3aa** in 98% conversion in **1** h (entry 6).

Through these experiments, we decided to investigate the betaine-Cu catalytic system in detail. To verify the effects of betaine, reactions were performed in various amounts of Cu^(I) with 5 mol% of betaine (Table 2). No reaction occurred in the absence of Cu^(I) regardless of the presence of betaine for 24 h (entry 1-2). Without betaine, the reaction also proceeded sluggishly using 100 ppm of Cu^(I) (entry 3). On the other hand, in the presence of betaine, 3aa was obtained in 97% conversion even with only 10 ppm of Cu^(I) in 5 h (entry 4). Reduction of the Cu^(I) concentration to 1 ppm resulted in 77% conversion, leading to TONs of up to 616 000 and TOFs of up to 123 200 h⁻¹ (entry 5). Surprisingly, when the reaction temperature was increased to 40 °C, 1a was converted to 3aa in 24 h cleanly even at a 25 ppb level of Cu^(I) (entry 6). Even though the best results were obtained at 40 °C, considering a wide range of applications including chemical biology and bioorthogonal chemistry, all the reactions were carried out at 30 °C. These results implied that both Cu and betaine play important roles in the CuAAC reaction in water.

Then, to gain insight into the role of betaine in the Cu catalytic mechanism, we tested other additives, such as choline chloride, sodium acetate, and sodium propionate, with respect to their potential to accelerate triazole formation (Table 3). In the case of choline chloride, which has only an ammonium part in the molecule, the reaction gave a moderate conversion of 66% in the presence of 200 ppm of Cu⁽¹⁾ for 1 h at 30 °C (entry 1). This result indicates that the ammonium part of choline chloride contributes to the catalytic sys-

Table 1 Surfactant effects on the Cu-catalyzed [2 + 3] cycloaddition of azide 1a with alkyne $2a^{\alpha}$

Ta	[^] N _{3 +}	} 2a −	Surfactant Cu ^(I) 200 ppm Na ascorbate H ₂ O, 30 °C	. 07	NNN 3aa
Entry	Surfactant	Ту	ре	Time	Conv. (%)
1	_			5 h	49
2^{b}	β-CD	_		1 h	61
3	SDS	An	ionic	1 h	68
4	Aliquat 12	8 Ca	tionic	1 h	33
5	Triton X-10	00 No	n-ionic	1 h	42
6	Betaine	Zw	itterionic	1 h	98

^{*a*} All the reactions were carried out with benzyl azide 1a (0.75 mmol), phenyl acetylene 2a (1.05 equiv.), CuSO₄·5H₂O (0.025 mol%, 200 ppm based on Cu), Na ascorbate (0.15 equiv.) and a surfactant (0.05 equiv.) in H₂O (1 mL) at 30 °C. ^{*b*} β -CD, the phase transfer catalyst.

tem. Sodium acetate and sodium propionate, having a carboxylate group in their molecules, showed 46% (entry 2) and 74% (entry 3) conversions, respectively. Remarkably, betaine, which has both a positively charged quaternary ammonium group and a negatively charged carboxylate group, existing in the zwitterionic form, showed an excellent conversion under the same reaction conditions (entry 4). These results showed clearly that both the ammonium part and the anionic part play important roles in this catalytic system. It is well known that betaine coordinates easily with Cu to stabilize and/or activate it.¹⁷ Therefore, we could speculate that betaine improves the solubility of the reactants and reagents in water, facilitates deprotonation of the terminal alkyne, and stabilizes the Cu⁽¹⁾ generated in the catalytic cycle.¹⁸

Next, the recycling of the CuAAC reaction between 1a and 2a in the presence of 200 ppm of Cu^(I) and betaine (5 mol%) was performed; a longer reaction time was required to observe the complete conversion with successive reuse of the betaine–Cu system (1st cycle: 1 h, 2nd cycle: 3.5 h, 3rd cycle: 9 h). The decrease in catalytic activity was ascribed to the well anticipated loss of Cu in the aqueous medium during workup.

To find the minimum concentration of Cu, we carried out experiments to optimize the reaction conditions for the CuAAC reaction depending on various substrates. As shown in Table 4, **1a** was reacted with various alkynes **2a**–**2n** in the presence of 2.5–200 ppm of Cu at 30 °C, affording the desired triazoles **3aa–3an**. Although most reactions were completed within several hours, the reaction time was set to 24 h to maximize the conversion and to compare the results with the recently reported CuAAC reaction results using ppm levels of Cu⁽¹⁾.^{9–11} In the case of phenyl acetylene (**2a**), the desired triazole **3aa** was obtained quantitatively with only 2.5 ppm of Cu⁽¹⁾ in water at 30 °C for 24 h. Moreover, a large-scale reaction (**1a**, **1g** scale) under these conditions was also successfully conducted to afford an excellent isolated yield (95%)

Table 2 CuAAC reaction between 1a and 2a using various concentrations of the Cu catalyst^a

		,					
			Betain	ie (5 mol%	b)		
	1a + 2a	CuSO ₄ :	CuSO ₄ ·5H ₂ O , Na ascorbate (15 mol%)				
			H ₂ O, 30 °C				
Entry	Cu ^(I)	Betaine (mol%)	Time (h)	Conv. (%)	TON	${\mathop{\rm TOF}}{\left(h^{-1} ight)}$	
1	_	_	24	<1	_	_	
2	—	5	24	<1			
3	100 ppm (0.0125 mol%)	—	5	48	3840	768	
4	10 ppm	5	5	97	77 600	15520	
5	1 ppm	5	5	77	616000	123 200	
6 ^b	25 ppb	5	24	100	32 000 000	1 333 333	
7 ^b	10 ppb	5	24	19	15200000	633 333	

^{*a*} All the reactions were carried out with 1a (0.75 mmol), 2a (1.05 equiv.), $CuSO_4$ ·5H₂O (25 ppb – 100 ppm based on Cu), Na ascorbate (0.15 equiv.) and betaine (0.05 equiv.) in H₂O (1 mL) at 30 °C. ^{*b*} The reaction was carried out at 40 °C.

Table 3Comparative screening of other additives, similar types of beta-ine, for the CuAAC reactions a

		A	dditive (5 mol%)	
1a	a + 2a	Cu ^(I) 200 ppm	, Na ascorbate (15 mol%	%) → 3aa
			H ₂ O, 30 ^o C, 1 h	ouu
Entry	Addit	ive	Structure	Conv. (%)
1	Choli	ne chloride	N OH	66
2	Sodiu	im acetate	O_ Na ⁺	46
3	Sodiu	im propionate	O O Na ⁺	74
4	Betai	ne	N N N O	98

^{*a*} All the reactions were carried out with benzyl azide **1a** (0.75 mmol), phenyl acetylene **2a** (1.05 equiv.), $CuSO_4$ ·5H₂O (0.025 mol%, 200 ppm based on Cu), Na ascorbate (0.15 equiv.) and an additive (0.05 equiv.) in H₂O (1 mL) at 30 °C for 1 h.

close to the conversion yield.¹⁹ For aryl alkynes containing electron-donating and electron-withdrawing groups, the reactions were completed in 24 h, with excellent isolated yields only at less than 25 ppm $Cu^{(I)}$ levels (**3ab-3ag**). Alkyne **3ah** bearing the sterically demanding adamantane group required a higher amount of the catalyst (100 ppm) to form the corresponding triazole. Also, for alkynes containing a hydroxyl group, more $Cu^{(I)}$ was required for the completion of the reaction (100 ppm for **3ai** and 200 ppm for **3aj**), probably due to the compromised hydrophobic effect of the ligand. Moreover, aliphatic alkynes **3ak**, **3al**, and **3am** efficiently reacted with only 75–100 ppm of $Cu^{(I)}$. Interestingly, ethynylferrocene (**2n**) also reacted with **2a** without any difficulty, with only 25 ppm of $Cu^{(I)}$, to produce the desired product **3an** in 94% isolated yield.

To further demonstrate the general applicability and versatility of this catalytic system, diverse azides **1b–1h** were also allowed to react with alkynes **2a–2l** in the presence of the betaine–Cu system (Table 5). Most aryl or aliphatic azides and alkynes afforded good yields of the corresponding triazoles using low Cu^(I) levels of 25–75 ppm (**4ba**, **4bl**, **4ca**, **4da**, **4ee**, **4ea**, **4el**, **4fa**, **4fl**, and **4ga**). An azide containing a carboxyl group and alkynes containing a hydroxyl group required 125 and 150 ppm of Cu^(I) to produce the desired products **4ha** and **4bi** in high yields, respectively.

Conclusions

A simple betaine–Cu catalytic system with remarkably small amounts of Cu, which provided facile and convenient access to triazoles, was developed. The use of this catalytic system in water at 30 °C efficiently produced a wide range of triazoles in good to excellent yields. Furthermore, workup was easy, requiring only simple suction filtration or solvent extraction. In addition, this method resulted in the remarkable reduction of the $Cu^{(1)}$ concentration down to ppm levels during the reaction, which demonstrates easy applicability since specially designed ligands were not required for the activation and stabilization of $Cu^{(1)}$. The advantages of this protocol include the use of a conventional Cu catalytic mixture that is easily generated from $CuSO_4$ -Na ascorbate and betaine, which is an easily obtained, commercially available, biodegradable, and biocompatible surfactant. Therefore, this efficient catalytic system is believed to have broad applications in biological and materials sciences.

Experimental section

General information

¹H-NMR and ¹³C-NMR spectra were recorded on a Bruker AVANCE 400 spectrometer. Chemical shifts are given with respect to residual CHCl₃ (7.28 ppm) in CDCl₃. IR spectra of all compounds were recorded on a Frontier FT-IR spectrometer, and HRMS analyses were carried out using a Bruker micrOTOF-Q instrument with electrospray ionization operating in positive mode. The melting temperature was determined with an IA 9200 melting point apparatus from Electrothermal Thermo Scientific. The mass fractions of residual Cu in the products were determined by inductively coupled plasma mass spectrometry (ICP-MS, Thermo Scientific Neptune Plus) with the external calibration method.

Typical procedure for CuAAC

In a vial fitted with a screw cap, to a solution of benzyl azide 1a (100 mg, 0.75 mmol) and phenyl acetylene 2a (83 mg, 1.05 equiv.) was added sodium ascorbate (23 mg, 0.15 equiv.), betaine (4.5 mg, 0.05 equiv.), and copper(II) sulfate pentahydrate in H₂O (1 mL of stock solution, 2.5 ppm based on Cu) at 30 °C. The reaction mixture was stirred for 24 h at 30 °C. The resulting product was extracted with CH₂Cl₂ (3 mL) three times. The combined organic layer was concentrated *in vacuo*. The crude compound was washed with pentane to give 172 mg of 3aa (97%).

1-Benzyl-4-phenyl-1H-1,2,3-triazole (3aa)

Ivory needles (97%); mp = 131 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, *J* = 7.1 Hz, 2H), 7.67 (s, 1H), 7.44–7.33 (m, 7H), 5.60 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.1, 134.6, 130.5 (2C), 129.0 (2C), 128.7, 128.6, 128.0, 127.9 (2C), 125.6 (2C), 119.5, 54.1.

1-Benzyl-4-(p-tolyl)-1H-1,2,3-triazole (3ab)

White needles (91%); mp = 154–155 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.71 (d, J = 7.4 Hz, 2H), 7.64 (s, 1H), 7.42–7.38 (m, 3H), 7.35–7.31 (m, 2H), 7.23 (d, J = 7.6 Hz, 2H), 5.58 (s, 2H), 2.38 (s, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.3, 138.0, 134.7, 129.4 (2C), 129.1 (2C), 128.7, 128.0, 127.7 (2C), 125.6 (2C), 119.1, 54.2, 21.2.

1-Benzyl-4-(3-fluorophenyl)-1H-1,2,3-triazole (3ac)

Ivory needles (100%); mp = 109–111 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.72 (s, 1H), 7.57–7.53 (m, 2H), 7.38–7.30 (m, 6H),

Table 4Results of the CuAAC reaction between 1a and various alkynes2a-2n using betaine in water^a

	N _{3 +} R ₁ 1a 2a - 2n	Betaine CuSO ₄ 5H ₂ O, Na ascort H ₂ O, 30 °C, 24 h	bate	N-N N 3an R ₁
Entry	Alkyne	Product	Cu ^(I) (ppm)	Isolated
1	$\equiv -\sqrt{2a}$	3aa	2.5	97
2		3ab	25	97
3	= $2c$	3ac	25	88
4	=	3ad ^b	25	98
5		3ae	25	97
7	= $2f$	3af	25	95
8	=-{~_2g	3ag	25	96
9	≡2h	3ah	100	83
10	=он2і	3ai	100	98
11	⊚∽он 2 ј	3aj	200	87
12	≫∕∕∕ 2k	3ak	100	93
13	≫∽2l	3al	75	97
14	۶۰۰۰۰ 2m	3am	100	76
15	Fe 2n	3an	25	94

^{*a*} All reactions were performed with azide 1a (0.75 mmol), alkynes 2a–2n (1.05 equiv.), CuSO₄·5H₂O (2.5–200 ppm based on Cu), Na ascorbate (0.15 equiv.), and betaine (0.05 equiv.) in water (1 mL) for 24 h at 30 °C. The yields shown represent those obtained for isolated products. ^{*b*} The concentration of residual Cu in the product was 14 ppm.

7.00 (t, J = 8.4 Hz, 1H), 5.56 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 163.0 (d, J = 245.6 Hz), 147.0, 134.4, 132.6 (d, J = 8.6 Hz), 130.3 (d, J = 8.4 Hz), 129.1 (2C), 128.7, 127.9 (2C), 121.2 (d, J = 2.8 Hz), 119.9, 114.7 (d, J = 21.2 Hz), 112.4 (d, J = 23.0 Hz), 54.1.

1-Benzyl-4-(2,4-difluorophenyl)-1H-1,2,3-triazole (3ad)

Tan needles (98%); mp = 124–126 °C; ¹H-NMR (400 MHz, CDCl₃) δ 8.29 (dd, J = 15.3, 7.2 Hz, 1H), 7.84 (d, J = 3.0 Hz, 1H), 7.42–7.31 (m, 5H), 7.00 (t, J = 8.4 Hz, 1H), 6.87 (t, J = 10.8 Hz, 1H), 5.60 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 163.7 (d, J = 12.2 Hz), 160.8 (dd, J = 83.9, 11.9 Hz), 157.9 (d, J = 11.7 Hz), 140.9, 134.6, 129.1 (2C), 128.9–128.6 (m), 127.9 (2C), 122.1 (d, J = 12.2 Hz), 115.0 (dd, J = 13.2, 3.8 Hz), 111.9 (dd, J = 21.3, 3.4 Hz), 104.0 (t, J = 25.7 Hz), 54.2.

1-Benzyl-4-(3-(trifluoromethyl)phenyl)-1*H*-1,2,3-triazole (3ae)

White amorphous solid (97%); mp = 70 °C; ¹H-NMR (400 MHz, CDCl₃) δ 8.07 (s, 1H), 8.00 (d, J = 7.6 Hz, 1H), 7.79 (s, 1H), 7.57–7.48 (m, 2H), 7.42–7.31 (m, 5H), 5.59 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 146.8, 134.4, 131.4, 131.4 (q, J = 32.4 Hz), 129.3, 129.2 (2C), 128.9, 128.8, 128.1 (2C), 124.7 (q, J = 3.7 Hz), 124.0 (d, J = 272.3 Hz), 122.4 (q, J = 3.9 Hz), 120.0, 54.3.

1-Benzyl-4-(3,5-dimethoxyphenyl)-1H-1,2,3-triazole (3af)

Tan needles (95%); mp = 119–121 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H), 7.37–7.29 (m, 5H), 6.99 (s, 2H), 6.44 (s, 1H), 5.55 (s, 2H), 3.82 (s, 6H); ¹³C-NMR (100 MHz, CDCl₃) δ 161.1 (2C), 147.9, 134.6, 132.3, 129.0 (2C), 128.6, 127.9 (2C), 119.8, 103.5 (2C), 100.5, 55.3 (2C), 54.1; FT-IR (KBr, cm⁻¹) 3119 (m), 3089 (w), 2998 (w), 2961 (w), 2936 (w), 2837 (w), 1595 (vs), 1551 (w), 1497 (w), 1474 (m), 1453 (m), 1417 (m), 1359 (m), 1274 (w), 1229 (m), 1153 (s), 1179 (m), 1200 (vs), 1092 (m), 1065 (m), 1044 (m), 1001 (m), 925 (m), 849 (m), 833 (s), 716 (s); HRMS [M + Na]⁺ calcd. For C₁₇H₁₇N₃O₂Na 318.1218, found 318.1226.

3-(1-Benzyl-1H-1,2,3-triazol-4-yl)aniline (3ag)

Brown needles (96%); mp = 135–137 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.63 (s, 1H), 7.41–7.38 (m, 3H), 7.38–7.31 (m, 6H), 7.30–7.28 (m, 2H),7.25 (s, 1H), 7.18 (t, *J* = 7.7 Hz, 1H), 7,12–7.11 (m, 1H), 6.66–6.64 (m, 1H), 5.57 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.3, 146.9, 134.7, 131.4, 129.7 (2C), 129.1, 128.7, 128.0 (2C), 119.5, 115.9, 114.9, 112.2, 54.1.

4-((Adamantan-1-ylmethoxy)methyl)-1-benzyl-1*H*-1,2,3-triazole (3ah)

Yellow solid (94%); mp =125-127 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.44 (s, 1H), 7.40-7.37 (m, 2H), 7.30-7.28 (m, 3H), 5.55 (s, 2H), 4.60 (s, 2H), 3.07 (s, 2H), 1.95 (s, 3H), 1.66 (dd, J = 32.0, 12.2 Hz, 6H), 1.51-1.50 (m, 6H); ¹³C-NMR (100 MHz, CDCl₃) δ 146.5, 134.7, 129.0 (2C), 128.7, 128.0 (2C), 122.1, 81.5, 65.1, 54.1, 39.7 (3C), 37.1 (3C), 34.0, 28.2 (3C); FT-IR (KBr, cm⁻¹) 3141 (w), 2901 (vs), 2847 (s), 1722 (w), 1638 (w), 1497 (w), 1454 (m), 1362 (w), 1344 (w), 1224 (m), 1156 (w), 1089 (m), 1048 (m), 812 (w), 723 (m), 695 (w); HRMS [M + Na]⁺ calcd. For C₂₁H₂₇N₃ONa 360.2052, found 360.2038.

1-(1-Benzyl-1H-1,2,3-triazol-4-yl)cyclohexan-1-ol (3ai)

Ivory needles (98%); mp = 123–125 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.40–7.32 (m, 4H), 7.27–7.25 (m, 2H), 5.47 (s, 2H), 2.92 (brs, 1H, OH), 1.93–1.25 (m, 10H); ¹³C-NMR (100 MHz, CDCl₃) δ 156.0, 134.6, 128.9 (2C), 128.7, 128.1 (2C), 119.6, 69.4, 54.0, 37.9 (2C), 25.2, 21.8 (2C).

Table 5Results of the CuAAC reaction of different azides 1b-1h with al-
kynes 2a-2l using betaine in water^a

	R2-N3		Betaine CuSO ₄ •5H ₂ O, Na asco	orbate R ₂ N	NN
	1b - 1h	2a - 2l	H ₂ O, 30 °C, 24 h	4ba -	4ha ^R 1
Entry	Azide	Alky	ne Produ	ct Cu ^(I)	Isolated
1	\bigcirc	_N ₃	4ba	(ppm) 25	yield (%) 95
	2b	2a			
2	2b	~	4bl ^b	75	96
		21			
3	2b	=	on 4bi	150	96
		2i			
4	\bigcirc	∕_ _{N3} 2a	4ca	25	100
	2 c				
5		_{N3} 2a	4da	25	94
	2d				
6	1 N3		4ee	25	94
	Ze	2e	CF3		
7	2e	2e 2a	4ea	75	92
8	2e	21	4el	75	97
9		. _{N3} 2a	4fa	25	92
	2 f				
10	2 f	21	4fl	75	92
11		_ ^{№3} 2a	4ga	50	94
	2g				
12	но	~_ ^{N3} ^{2a}	4ha	125	90
	2h				

^{*a*} All reactions were performed with azides **1b-1h** (0.75 mmol), alkynes **2a-2l** (1.05 equiv.), CuSO₄·5H₂O (25–150 ppm based on Cu), Na ascorbate (0.15 equiv.), and betaine (0.05 equiv.) in water (1 mL) for 24 h at 30 °C. The yields shown represent those obtained for isolated products. ^{*b*} The concentration of residual Cu in the product was 16 ppm.

3-(1-Benzyl-1H-1,2,3-triazol-4-yl)propan-1-ol (3aj)

Yellow oil (87%); ¹H-NMR (400 MHz, CDCl₃) δ 7.40–7.35 (m, 3H), 7.28–7.25 (m, 3H), 5.50 (s, 2H), 3.70 (t, *J* = 5.8 Hz, 2H), 2.82 (t, *J* = 7.2 Hz, 2H), 2.70–2.60 (brs, 1H, OH), 1.92 (quintet, *J* = 6.6 Hz, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.0, 134.6, 128.8 (2C), 128.4, 127.7 (2C), 121.0, 61.1, 53.8, 31.8, 21.7.

1-Benzyl-4-butyl-1H-1,2,3-triazole (3ak)

Ivory amorphous solid (93%); mp = 60–62 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.42–7.36 (m, 3H), 7.28–7.26 (m, 2H), 7.19 (s, 1H), 5.51 (s, 2H), 2.71 (t, *J* = 7.6 Hz, 2H), 1.68–1.61 (m, 3H), 1.38 (sextet, *J* = 7.5 Hz, 2H), 0.93 (t, *J* = 7.6 Hz, 2H); ¹³C-NMR

(100 MHz, CDCl₃) δ 149.0, 135.0, 129.0 (2C), 128.6, 127.9 (2C), 120.4, 54.0, 31.5, 25.4, 22.3, 13.8.

1-Benzyl-4-hexyl-1H-1,2,3-triazole (3al)

Ivory amorphous solid (97%); mp = 52–54 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.45 (m, 3H), 7.37–7.35 (m, 3H), 5.60 (s, 2H), 2.79 (t, *J* = 7.7 Hz, 2H), 1.77–1.71 (m, 2H), 1.46–1.39 (m, 6H), 0.98–0.95 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 149.0, 135.0, 129.0 (2C), 128.6, 127.9 (2C), 120.4, 54.0, 31.5, 29.3, 28.9, 25.7, 22.5, 14.0.

1,5-Bis(1-benzyl-1H-1,2,3-triazol-4-yl)pentane (3am)

Ivory amorphous solid (98%); mp = 125–127 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.42–7.36 (m, 6H), 7.30–7.28 (m, 4H), 7.21 (s, 2H), 5.52 (s, 4H), 2.71 (t, *J* = 7.4 Hz, 4H), 1.74–1.67 (m, 4H), 1.46–1.40 (m, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.6 (2C), 135.0 (2C), 129.0 (4C), 128.6 (2C), 127.9 (4C), 120.5 (2C), 54.0 (2C), 29.0 (2C), 28.6 (2C), 25.5.

1-Benzyl-4-ferrocenyl-1H-1,2,3-triazole (3an)

Red needles (94%); mp = 153–155 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.40–7.36 (m, 4H), 7.29–7.28 (m, 2H), 5.54 (s, 2H), 4.71 (s, 2H), 4.23 (s, 2H), 4.07 (s, 5H); ¹³C-NMR (100 MHz, CDCl₃) δ 147.1, 134.8, 129.0 (2C), 128.6, 127.8 (2C), 118.7, 75.3, 69.4 (5C), 68.5 (2C), 66.5 (2C), 53.9.

1-Phenethyl-4-phenyl-1*H*-1,2,3-triazole (4ba)

White amorphous solid (95%); mp = 140 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.79 (d, J = 7.8 Hz, 2H), 7.49 (s, 1H), 7.43 (t, J = 7.6 Hz, 2H), 7.36–7.26 (m, 2H), 7.17–7.15 (m, 4H), 4.66 (t, J = 7.0 Hz, 2H), 3.28 (t, J = 7.2 Hz, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 147.5, 137.1, 130.7, 128.8 (2C), 128.8 (2C) 128.7 (2C), 128.1, 127.1, 125.7 (2C), 119.9, 51.7, 36.8.

4-Hexyl-1-phenethyl-1H-1,2,3-triazole (4bl)

Yellow oil (96%); ¹H-NMR (400 MHz, CDCl₃) δ 7.33–7.25 (m, 3H), 7.11 (d, *J* = 7.2 Hz, 2H), 7.00 (s, 1H), 4.55 (t, *J* = 7.2 Hz, 2H), 3.20 (t, *J* = 7.2 Hz, 2H), 2.67 (t, *J* = 7.5 Hz, 2H), 1.64–1.59 (m, 2H), 1.33–1.29 (m, 6H), 0.91–0.88 (m, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.1, 137.2, 128.7 (2C), 128.6 (2C), 126.9, 120.9, 51.4, 36.8, 31.5, 29.4, 28.8, 25.5, 22.5, 14.0.

1-(1-Phenethyl-1H-1,2,3-triazol-4-yl)cyclohexan-1-ol (4bi)

Ivory needles (96%); mp = 109 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.29–7.17 (m, 4H), 7.09 (d, *J* = 7.0 Hz, 2H), 4.55 (t, *J* = 7.0 Hz, 2H), 3.19 (t, *J* = 7.0 Hz, 2H), 2.77 (brs, 1H, OH), 1.96–1.70 (m, 6H), 1.59–1.32 (m, 4H); ¹³C-NMR (100 MHz, CDCl₃) δ 155.1, 137.0, 128.7 (2C), 128.6 (2C), 127.0, 120.1, 69.3, 51.5, 38.0, 36.7, 25.3, 22.0.

4-Phenyl-1-(3-phenylpropyl)-1H-1,2,3-triazole (4ca)

Ivory amorphous solid (100%); mp = 81–82 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 7.8 Hz, 2H), 7.74 (s, 1H), 7.45 (t, *J*

= 7.6 Hz, 2H), 7.38–7.32 (m, 3H), 7.27–7.21 (m, 3H), 4.43 (t, J= 7.0 Hz, 2H), 2.72 (t, J = 7.3 Hz, 2H), 2.33 (quintet, J = 7.3 Hz, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 147.8, 140.1, 130.7, 128.8, 128.6, 128.5, 128.1, 126.4, 125.7, 119.5, 49.6, 32.5, 31.7.

4-Phenyl-1-(8-phenyloctyl)-1H-1,2,3-triazole (4da)

White amorphous solid (94%); mp = 70–73 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.86 (d, J = 7.8 Hz, 2H), 7.75 (s, 1H), 7.45 (t, J = 7.6 Hz, 2H), 7.38–7.34 (m, 1H), 7.32–7.26 (m, 2H), 7.20–7.18 (m, 3H), 4.41 (t, J = 7.1 Hz, 2H), 2.61 (t, J = 7.4 Hz, 2H), 2.00–1.93 (m, 2H), 1.63–1.60 (m, 2H), 1.38–1.34 (m, 8H); ¹³C-NMR (100 MHz, CDCl₃) δ 147.7, 142.7, 130.7, 128.8 (2C), 128.4 (2C), 128.2 (2C), 128.1, 125.7(2C), 125.6, 119.3, 50.4, 35.9, 31.4, 30.3, 29.2, 29.1, 28.9, 26.5; FT-IR (KBr, cm⁻¹) 3081 (m), 3031 (w), 2917 (vs), 2849 (vs), 1609 (w), 1462 (m), 1384 (w), 1352 (w), 1222 (w), 1202 (w), 1180 (w), 1085 (w), 1051 (w), 1027 (w), 912 (w), 861 (w), 766 (s), 697 (s), 517 (w); HRMS [M + Na]⁺ calcd. For C₂₂H₂₇N₃Na 356.2103, found 356.2088.

1-Octyl-4-(3-(trifluoromethyl)phenyl)-1H-1,2,3-triazole (4ee)

Yellow oil (94%); ¹H-NMR (400 MHz, CDCl₃) δ 8.09 (s, 1H), 8.03 (d, *J* = 7.2 Hz, 1H), 7.87 (s, 1H), 7.58–7.50 (m, 2H), 4.40 (t, *J* = 7.1 Hz, 2H), 1.95 (s, 2H), 1.36–1.23 (m, 10H), 0.87 (s, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 146.3, 131.6, 131.4 (q, *J* = 32.4 Hz), 129.2, 128.0, 124.5 (q, *J* = 3.7 Hz), 124.0 (q, *J* = 272.3 Hz), 122.3 (q, *J* = 3.9 Hz), 119.9, 50.5, 31.6, 30.2, 28.9, 28.8, 26.4, 22.5, 13.9; FT-IR (KBr, cm⁻¹) 3132 (w), 3096 (w), 2930 (vs), 2858 (s), 1622 (w), 1596 (w), 1557 (w), 1490 (w), 1457 (m), 1420 (m), 1353 (m), 1323 (vs), 1228 (s), 1167 (vs), 1128 (s), 1096 (s), 988 (w), 911 (m), 804 (s), 710 (m), 698 (s); HRMS [M + Na]⁺ calcd. For C₁₇H₂₂F₃N₃Na 348.1664, found 348.1642.

1-Octyl-4-phenyl-1H-1,2,3-triazole (4ea)

White amorphous solid (92%); mp = 74–76 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.85 (d, J = 7.7 Hz, 2H), 7.75 (s, 1H), 7.44 (t, J = 7.5 Hz, 2H), 7.36–7.33 (m, 1H), 4.41 (t, J = 7.2 Hz, 2H), 1.99–1.94 (m, 2H), 1.38–1.28 (m, 10H), 0.91–0.88 (m, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 147.7, 130.8, 128.8 (2C), 128.0, 125.7 (2C), 119.3, 50.4, 31.7, 30.3, 29.0, 28.9, 26.5, 22.6, 14.0.

4-Hexyl-1-octyl-1H-1,2,3-triazole (4el)

White oil (97%); ¹H-NMR (400 MHz, CDCl₃) δ 7.28 (s, 1H), 4.30 (t, *J* = 7.1 Hz, 2H), 2.70 (t, *J* = 7.5 Hz, 2H), 1.89–1.85 (m, 2H), 1.68–1.64 (m, 2H), 1.35–1.25 (m, 16H), 0.87 (s, 6H); ¹³C-NMR (100 MHz, CDCl₃) δ 148.1, 120.2, 49.9, 31.5, 31.4, 30.1, 29.3, 28.8, 28.8, 28.7, 26.3, 25.5, 22.4, 22.3, 13.8 (2C).

1-(3-Methoxyphenethyl)-4-phenyl-1H-1,2,3-triazole (4fa)

Ivory amorphous solid (92%); mp = 77–78 °C; ¹H-NMR (400 MHz, CDCl₃) δ 7.79 (d, J = 7.6 Hz, 2H), 7.51 (s, 1H), 7.43 (t, J = 7.6 Hz, 2H), 7.34 (t, J = 7.5 Hz, 1H), 7.24 (t, J = 7.8 Hz, 1H) 6.82 (d, J = 8.1 Hz, 1H), 6.75 (d, J = 7.5 Hz, 1H), 6.68 (s, 1H), 4.65 (t, J = 7.1 Hz, 2H), 3.77 (s, 3H), 3.25 (t, J = 7.1 Hz, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 159.9, 147.5, 138.6, 130.6,

129.9, 128.8 (2C), 128.1, 125.7 (2C), 121.0, 119.9, 114.3, 112.6, 55.2, 51.6, 36.8; FT-IR (KBr, cm⁻¹) 3120 (w), 3005 (w), 2951 (w), 2841 (w), 1602 (s), 1593 (w), 1494 (m), 1480 (m), 1466 (s), 1435 (w), 1358 (w), 1319 (m), 1276 (m), 1262 (s), 1225 (m), 1193 (w), 1169 (s), 1081 (m), 1041 (s), 1015 (m), 972 (w), 926 (w), 913 (w), 857 (m), 820 (m), 785 (m), 765 (vs), 721 (m), 691 (vs), 559 (w), 510 (w); HRMS $[M + Na]^+$ calcd. For $C_{17}H_{17}N_3ONa$ 302.1269, found 302.1260.

4-Hexyl-1-(3-methoxyphenethyl)-1H-1,2,3-triazole (4fl)

Yellow oil (92%); ¹H-NMR (400 MHz, CDCl₃) δ 7.18 (t, *J* = 7.8 Hz, 1H), 7.02 (s, 1H), 6.77 (d, *J* = 8.1 Hz, 1H), 6.67 (d, *J* = 7.4 Hz, 1H), 6.61 (s, 1H), 4.52 (t, *J* = 7.2 Hz, 2H), 3.74 (s, 3H), 3.14 (t, *J* = 7.1 Hz, 2H), 2.65 (t, *J* = 7.4 Hz, 2H), 1.62–1.56 (m, 2H), 1.28 (s, 6H), 0.88–0.84 (m, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 159.7, 148.0, 138.7, 129.6, 120.9, 114.2, 112.3, 55.0, 51.3, 36.7, 31.4, 29.3, 28.7, 25.4, 22.4, 13.9; FT-IR (KBr, cm⁻¹) 3134 (w), 2930 (vs), 2857 (s), 1603 (s), 1586 (s), 1551 (w), 1492 (s), 1456 (s), 1437 (s), 1293 (m), 1263 (C–O, vs), 1216 (m), 1154 (m), 1127 (w), 1050 (C–O, s), 874 (m), 782 (m), 697 (m); HRMS [M + Na]⁺ calcd. For C₁₇H₂₅N₃ONa 310.1895, found 310.1879.

1-(Naphthalene-2-yl)-2-(4-phenyl-1*H*-1,2,3-triazol-1-yl)ethan-1one (4ga)

Yellow amorphous solid (94%); mp = 183–185 °C; ¹H-NMR (400 MHz, CDCl₃); 8.60 (s, 1H), 8.08–7.90 (m, 7H), 7.72–7.64 (m, 2H), 7.46 (t, *J* = 7.4 Hz, 2H), 7.28 (t, *J* = 7.5 Hz, 1H), 6.05 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 190.2, 148.3, 136.2, 132.4, 131.3, 130.5, 130.3, 129.8, 129.4, 129.3, 128.8, 128.2, 128.0, 127.4, 125.8, 123.3, 121.4, 55.5.

5-(4-Phenyl-1H-1,2,3-triazol-1-yl)pentanoic acid (4ha)

Ivory amorphous solid (90%); mp = 130–131 °C; ¹H-NMR (400 MHz, DMSO-d₆) δ 8.58 (s, 1H), 7.84 (d, J = 7.4 Hz, 2H), 7.44 (t, J = 7.3 Hz, 2H), 7.33 (t, J = 7.3 Hz, 1H), 4.40 (t, J = 6.7 Hz, 2H), 2.27 (t, J = 7.0 Hz, 2H), 1.93–1.83 (m, 2H), 1.52–1.45 (m, 2H); ¹³C-NMR (100 MHz, DMSO-d₆) δ 174.6, 146.7, 131.3, 129.3 (2C), 128.2, 125.6 (2C), 121.8, 49.7, 33.4, 29.5, 21.9; FT-IR (KBr, cm⁻¹) 3121 (m), 2952 (w), 2869 (w), 2815 (w), 1709 (vs), 1694 (C=O, vs), 1464 (m), 1441 (w), 1332 (w), 1273 (w), 1215 (m), 1190 (w), 1132 (w), 1080 (m), 1052 (w), 977 (w), 839 (w), 763 (s), 696 (s), 629 (w), 526 (w); HRMS [M + Na]⁺ calcd. For C₁₃H₁₅N₃O₂Na 268.1062, found 268.1055.

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- 19 Procedure for a gram-scale synthesis of 3aa: In a round bottom flask, to a solution of benzyl azide 1a (1 g, 7.5 mmol) and 2a (1.05 equiv.) was added sodium ascorbate (0.15 equiv.), betaine (0.05 equiv.), and copper(II) sulfate pentahydrate in H_2O (10 mL of stock solution, 2.5 ppm based on Cu). The reaction mixture was stirred for 24 h at 30 °C. The resulting mixture was filtered, washed several times with water, and dried. Product 3aa was obtained (1.68 g) as a white solid in 95% isolated yields.