SYNTHESIS OF THE CONJUGATE OF CYTISINE AND KOJIC ACID

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N-(5-Hydroxypyran-4-on-2-ylmethyl)cytisine was synthesized.

Keywords: cytisine, kojic acid, pyran-4-one, N-alkylation.

Chemical modification of biologically active natural compounds is a promising and effective method for new drug discovery. The quinolizidine alkaloid cytisine is especially interesting among such compounds [1, 2]. Kojic acid (5-hydroxy-2-hydroxymethylpyran-4-one) (1) is a biologically important fungal secondary metabolite that is produced by various genera of fungi, e.g., *Aspergillus, Acetobacter*, and *Penicillium*, and possesses various pharmacological properties [3]. Kojic acid contains several reactive centers and is a very convenient synthon for synthesizing various derivatives and heterocyclic compounds [4].



a. SOCl₂, room temp., 2 h, 65%; b. cytisine, Et₃N, MeCN, reflux, 6 h, 76%

In continuation of research on the synthesis of cytisine derivatives [5-8], we synthesized *N*-(5-hydroxypyran-4-on-2-ylmethyl)cytisine (**3**).

Chlorokojic acid (2) was produced in 65% yield via treatment of kojic acid with thionylchloride at room temperature [9, 10]. Considering the structural features of alkylating agent 2, we optimized the conditions for cytisine *N*-alkylation. The bases were NaOH solution, K_2CO_3 , and tertiary amines [Et₃N, *N*-methylmorpholine, 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), and di-isopropylethylamine]. The reaction was carried out in Me₂CO, EtOH, and MeCN. The most satisfactory results were obtained using Et₃N in anhydrous MeCN. Under these conditions, cytisine was monoalkylated on N-3 to give *N*-(5-hydroxypyran-4-on-2-ylmethyl)cytisine (3) in high yield.

Synthesized conjugate **3** was of interest as a key polyfunctional synthon for organic synthesis because it contained an active phenol and pyran-4-one ring. This provided new possibilities for subsequent modification of cytisine derivatives.

EXPERIMENTAL

The course of reactions and purity of products were monitored by TLC on Merck 60 F254 plates. The eluent was CHCl₃–MeOH (9:1 and 95:5). Melting points were determined on a Kofler block. NMR spectra were recorded on Varian VXR-300 and Bruker Avance DRX-500 spectrometers vs. TMS (internal standard). Optical rotation was measured on a PerkinElmer 341 polarimeter. LC-MS spectra were recorded using an Agilent 1100 Series HPLC-MS equipped with a diode-array and Agilent LC\MSD SL mass-selective detectors with chemical ionization at atmospheric pressure (APCI). Elemental analyses of all compounds agreed with those calculated.

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Pharmacopoeial (–)-cytisine { $[\alpha]_D^{20}$ –110° (*c* 0.5%, EtOH)} isolated from *Thermopsis lanceolata* and commercial kojic acid (Chemos GmbH & Co. KG, Germany) were used.

2-(Chloromethyl)-5-hydroxypyran-4-one (2) was prepared in 65% yield by treating kojic acid with thionylchloride at room temperature [9, 10]. Mp 161–162°C (lit. 146–147°C [11–13], 165–166°C [14], 165–168°C [15], 166–167°C [9, 16–24]), C₆H₅ClO₃. ¹H NMR spectrum (300 MHz, DMSO-d₆, δ , ppm): 9.15 (1H, br.s, 5-OH), 8.11 (1H, s, H-6), 6.56 (1H, s, H-3), 4.65 (2H, s, CH₂-2).

(1*R*,5*S*)-3-(5-Hydroxy-4-oxo-4*H*-pyran-2-ylmethyl)-1,2,3,4,5,6-hexahydro-8*H*-1,5-methanopyrido[1,2*a*][1,5]diazocin-8-one (3). A solution of chlorokojic acid (2, 0.80 g, 5 mmol), cytisine (0.95 g, 5 mmol), and Et₃N (0.2 mL) in anhydrous MeCN (5 mL) was refluxed under N₂ for 6 h (course of reaction monitored by TLC) and cooled. The solvent was distilled off at reduced pressure in a rotary evaporator. The oily residue was treated with distilled H₂O (10 mL). The resulting precipitate was filtered, rinsed with H₂O, and crystallized from EtOH. Yield 1.19 g (76%), mp 182–183°C, C₁₇H₁₈N₂O₄, $[\alpha]_D^{20}$ -206.5° (*c* 1%, EtOH). ¹H NMR spectrum (500 MHz, DMSO-d₆, δ, ppm, J/Hz): 9.00 (1H, br.s, 5'-OH), 7.93 (1H, s, H-6'), 7.33 (1H, dd, J = 6.5, 9.0, H-10), 6.24 (1H, d, J = 9.0, H-9), 6.08 (1H, d, J = 6.5, H-11), 5.94 (1H, s, H-3'), 3.82 (1H, d, J = 15.5, H-6_{endo}), 3.70 (1H, dd, J = 15.9, 6.5, H-6_{exo}), 3.31 and 3.35 (2H, two d, J = 15.5, CH₂-2'), 2.98–3.07 (1H, m, H-1), 2.90 (1H, d, J = 10.0, H-2_{endo}), 2.79 (1H, d, J = 10.0, H-4_{endo}), 2.34–2.45 (3H, m, H-5, H-2_{exo}, 4_{exo}), 1.80 (1H, d, J = 12.5, H-13_{syn}), 1.69 (1H, d, J = 12.5, H-13_{anti}). ¹³C NMR spectrum (126 MHz, DMSO-d₆, δ, ppm): 173.99 (C-4'), 165.12 (C-2'), 162.67 (C-8), 152.18 (C-12), 146.14 (C-5'), 139.89 (C-6'), 139.30 (C-10), 115.91 (C-9), 112.39 (C-3'), 104.40 (C-11), 60.05 (C-2), 59.65 (C-4), 58.17 (C-2a'), 49.97 (C-6), 34.92 (C-1), 27.79 (C-5), 25.29 (C-13). LC-MS: 315.2 [MH]⁺ (100%).

REFERENCES

- 1. E. G. Perez, C. Mendez-Galvez, and B. K. Cassels, Nat. Prod. Rep., 29, 55 (2012).
- 2. J. Rouden, M. C. Lasne, J. Blanchet, and J. Baudoux, Chem. Rev., 114, 712 (2014).
- 3. J. Brtko, L. Rondahl, M. Fickova, D. Hudecova, V. Eybl, and M. Uher, Cent. Eur. J. Public Health, 12, S16 (2004).
- 4. M. Zirak and B. Eftekhari-Sis, *Turk. J. Chem.*, **39**, 439 (2015).
- 5. I. P. Dubovik, M. M. Garazd, V. I. Vinogradova, and V. P. Khilya, Chem. Nat. Compd., 42, 133 (2006).
- I. V. Nagorichna, A. S. Ogorodniichuk, M. M. Garazd, V. I. Vinogradova, and V. P. Khilya, *Chem. Nat. Compd.*, 43, 10 (2007).
- 7. M. V. Veselovskaya, M. M. Garazd, V. I. Vinogradova, and V. P. Khilya, Chem. Nat. Compd., 42, 277 (2006).
- 8. A. V. Yazlovitskii, M. M. Garazd, and V. G. Kartsev, Chem. Nat. Compd., 52, 272 (2016).
- 9. T. Yabuta, J. Chem. Soc., **125**, 575 (1924).
- 10. K. Sander, T. Kottke, L. Weizel, and H. Stark, Chem. Pharm. Bull., 58, 1353 (2010).
- 11. B. Berk, D. Us, S. Oktem, Z. T. Kocagoz, B. Caglayan, I. A. Kurnaz, and D. D. Erol, Turk, J. Chem., 35, 317 (2011).
- 12. G. Oeztuerk, D. D. Erol, M. D. Aytemir, and T. Uzbay, Eur. J. Med. Chem., 37, 829 (2002).
- 13. D. D. Erol and N. Yulug, *Eur. J. Med. Chem.*, **29**, 893 (1994).
- 14. C. A. Kingsbury, M. Cliffton, and J. H. Looker, J. Org. Chem., 41, 2777 (1976).
- 15. S. Wei and Z. Li, Chem. Nat. Compd., 52, 123 (2016).
- 16. M. D. Aytemir, U. Calis, and M. Ozalp, Arch. Pharm., 337, 281 (2004).
- 17. M. D. Aytemir, E. Septioglu, and U. Calis, Arzneim. Forsch., 60, 22 (2010).
- 18. M. D. Aytemir and U. Calis, Arch. Pharm., 343, 173 (2010).
- 19. M. D. Aytemir and B. Ozcelik, Eur. J. Med. Chem., 45, 4089 (2010).
- 20. M. D. Aytemir and B. Ozcelik, Med. Chem. Res., 20, 443 (2010).
- 21. G. Karakaya, M. D. Aytemir, B. Ozcelik, and U. Calis, J. Enzyme Inhib. Med. Chem., 28, 627 (2013).
- 22. M. Y. Moridani, G. S. Tilbrook, H. H. Khodr, and R. C. Hider, J. Pharm. Pharmacol., 54, 349 (2002).
- 23. K. Hryniewicz, K. Stadnicka, and A. Pattek-Janczyk, J. Mol. Struct., 919, 255 (2009).
- 24. M. D. Aytemir, B. Ozcelik, and G. Karakaya, Bioorg. Med. Chem. Lett., 23, 3646 (2013).