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SYNTHESIS AND BIOLOGICAL ACTIVITY OF ARTIFICIAL ANALOGS OF MYCALAMIDE A

Hideto Fukui,^a Yoshinori Tsuchiya,^a Keiko Fujita,^a Tadakiyo Nakagawa,^b Hiroyuki Koshino,^b and Tadashi Nakata^{b*}

^aCentral Research Institute, Kaken Pharmaceutical Co., Ltd., Shinomiya, Yamashina, Kyoto 607, Japan ^bThe Institute of Physical and Chemical Research (RIKEN), Wako-shi, Saitama 351-01, Japan

Abstract: Artificial analogs of mycalamide A, a potent antitumor and antiviral compound isolated from a New Zealand marine sponge, were synthesized and their biological activities were tested. © 1997 Published by Elsevier Science Ltd.

Mycalamide A (1) and B were isolated in 1988 from a New Zealand marine sponge of the genus Mycale.¹ Onnamides² and theopederins,³ which are structurally related compounds, have been isolated from a Japanese marine sponge of the genus *Theonella*. Interestingly, the structure of these compounds is strikingly similar to that of pederin, a strong insect poison isolated from *Paederus fuscipes*.⁴ The mycalamides exhibit potent *in vitro* cytotoxicity and *in vivo* antitumor activity as well as potent antiviral activity.^{1,5} In addition, mycalamide A (1) blocks T-cells activation in mice and is 10-fold more potent than FK-506.⁶ Recently, their unique structure and potent biological activity have attracted the attention of synthetic organic chemists.⁷ The structure-activity relationship of simple derivatives prepared from naturally occurring mycalamides has been reported.⁸ We now report the synthesis and biological activity of artificial analogs of mycalamide A (1) based on the synthetic strategy for our total synthesis of mycalamide A (1) and 10-*epi*-mycalamide A (2).^{7c, d} Mycalamide analogs, MA-1 (5) ~ MA-10 (14), were synthesized to investigate the requisite functional groups in the left half **3** and the possibility of replacing the right half **4** by glucose derivatives.



Right Half 4



Chemical Synthesis

Synthesis of the artificial analogs with the modified left half. To investigate the relationship between the functional group of the left half and the biological activity, we synthesized mycalamide analogs having 24, 28 and 31 as the left half. First, the left half 24 that lacks the C3-methyl group was synthesized using the synthetic strategy for the left half 3 in our total synthesis of pederin (Scheme 1).⁹ Michael addition of nitromethane to α , β -unsaturated ester 15,¹⁰ deprotection of the THP group, and lactonization produced δ lactone 16. Successive treatment of 16 with TiCl₃-AcONH₄ and with c. HCl afforded a 1:5 mixture of bicyclic lactones 17 and 18, which was treated with ethanedithiol-BF₃·Et₂O to give δ -lactones 19 and 20. The lactone 20 was converted into the α -keto ester 21 by aldol reaction, methylacetalization, and the Moffatt



Scheme 1. (a) Triton B, MeNO₂, rt (82%); (b) *p*-TsOH, MeOH, rt, then benzene azeotropic (85%, 4α:4β=1:1); (c) TiCl₃, AcONH₄-H₂O, THF, rt; (d) c. HCl, CH₂Cl₂, rt; (e) HSCH₂CH₂SH, BF₃El₂O, CH₂Cl₂, rt (40%, 3 steps); (f) LDA, MeOC(Me)₂OCH₂COMe, THF, -78 °C; (g) CSA, CH(OMe)₃, MeOH, CH₂Cl₂, rt (71%, 2 steps); (h) DMSO, DCC, pyridine, CF₃COOH, El₂O, rt (69%); (i) Zn(BH₄)₂, El₂O, -78 °C; (j) BZCl, DMAP, pyridine, rt (66%, 2 steps); (k) HgO, HgCl₂, H₂O, MeCN, 60 °C; (l) Zn(BH₄)₂, El₂O, 0 °C; (m) PhSeCN, *n*Bu₃P, THF, 0 °C (75%, 3 steps); (n) H₂O₂, THF, rt; El₃N, benzene, reflux; (o) *n*-PrSLi, HMPA (69%, 2 steps).

oxidation. The reduction of 21 with $Zn(BH_4)_2^{11}$ produced 7α -alcohol 22 and its 7β -epimer in a ratio of 6.8:1. After benzoylation of 22, deprotection of thioacetal, reduction with $Zn(BH_4)_2$, and treatment with PhSeCN-*n*-Bu₃P produced phenyl selenide 23. Successive treatment with H₂O₂ and with *n*-PrSLi gave the desired carboxylic acid 24.

The left half **28** without the C3-methyl and C4-*exo*-methylene groups was next synthesized (Scheme 2). Reduction of the double bond in **15** with NaBH₄-NiCl₂, deprotection of the THP group and lactonization produced δ -lactone **25** which was converted into α -keto ester **26**. Zn(BH₄)₂ reduction of **26** produced 7 α -alcohol **27** and its 7 β -isomer (6.8:1). The 7 α -alcohol **27** was converted into the carboxylic acid **28**.

The 7 β -benzoyl carboxylic acid 31, 7-epimer of the left half 3, was synthesized from 29, which is a synthetic intermediate in our total synthesis of pederin,⁹ using the same procedure taken in the synthesis of 24 (Scheme 3).

With the left half analogs 24, 28, and 31 in hand, we then undertook their condensation with the right half amine 4 (Scheme 4). Treatment of 24, 28, and 31 with *p*-TsCl and DMAP in CH₂Cl₂ followed by addition of 4 produced the coupling products $32 \sim 37$ which, after separation, were hydrolyzed to give 5 (MA-1) and its 10-epimer 6 (MA-2), 7 (MA-3) and its 10-epimer 8 (MA-4), and 9 (MA-5) and its 10-epimer 10 (MA-6), respectively.



Scheme 2. (a) NiCl₂, NaBH₄, MeOH, rt (92%); (b) *p*-TsOH, MeOH, rt, then benzene azeotropic (83%); (c) LDA, MeOC(Me)₂OCH₂COOMe, THF, -78 °C; (d) CSA, CH(OMe)₃, MeOH, CH₂Cl₂, rt (60%, 2 steps); (e) DMSO, DCC, pyridine, CF₃COOH, Et₂O, rt (36%); (f) Zn(BH₄)₂, Et₂O, -78 °C; (g) BzCl, DMAP, pyridine, rt; (h) *r*-PrSLi, HMPA, 0 °C (67%, 3 steps).



Scheme 3. (a) HgO, HgCl₂, H₂O, MeCN, 60°C; (b) Zn(BH₄)₂, Et₂O, -78 °C; (c) PhSeCN, *π*-Bu₃P, THF, 0 °C (50%, 3 steps); (d) H₂O₂, THF, rt; Et₃N, benzene, reflux; (e) *π*-PrSLi, HMPA (87%, 2 steps).



Scheme 4. (a) p-TsCl, DMAP, CH₂Cl₂, 0 °C-rt; (b) addition of amine 4 in CH₂Cl₂, rt; separation; (c) LiOH, MeOH, rt (29% for 5, 13% for 6, 36% for 7, 23% for 8, 25% for 9, 10% for 10, each 3 steps)

Synthesis of the artificial analogs with the modified right half. We synthesized mycalamide analogs having 43 and 44 as the right half, which were more readily prepared than the right half 4. After acetylation, 3-O-methyl- α -D-glucopyranose (38) was treated with propargyltrimethylsilane in the presence of TMSOTf and BF₃·Et₂O^{7a} to give 11β-allene 39 exclusively (Scheme 5). Hydrolysis of the triacetate 39 followed by treatment with pivaloyl chloride afforded diol 40, which was converted into azide 41 by successive treatment with O₃, (CH₂O)_n, Ac₂O, and TMSN₃. Alkaline hydrolysis of 41 followed by acetonization produced the tricyclic azide 42. The hydrogenolysis of 41 and 42 gave the right half amines 43 and 44, respectively.

The amines 43 and 44 were condensed with the left half 3 to give a mixture of 45 and 46, and 47 and 48, respectively, using the same procedure as mentioned above (Scheme 6). After separation, hydrolysis of 45, 46, 47, and 48 produced 11 (MA-7), 12 (MA-8), 13 (MA-9) and 14 (MA-10), respectively.



Scheme 5. (a) Ac₂O, pyridine, rt; (b) propargyITMS, TMSOTf, BF₃Et₂O, MaCN, 0 °C(68%, 2 steps); (c) K₂CO₃, MeOH, rt; (d) PivCl, pyridine, CH₂Cl₂ (91%, 2 steps); (e) O₃, MeOH, -78 °C; Me₂S, 0 °C; (f) (CH₂O)_n, CSA, CH₂Cl₂, 0 °C; (g) Ac₂O, pyridine, rt (74%, 3 steps); (h) TMSN₃, TMSOTf, MeCN, 0 °C (88%); (i) K₂CO₃, MeOH, rt; (j) Me₂C(OMe)₂, CSA, CH₂Cl₂, rt (85%, 2 steps); (k) H₂, 10% Pd-C, EtOAc, rt.



Scheme 6. (a) p-TsCl, DMAP, CH₂Cl₂, rt; (b) addition of amine 43 or 44 in CH₂Cl₂, rt; separation(c) LiOH, MeOH, rt (16% for 11, 22% for 12, 22% for 13, and 35% for 14, each 3 steps).

Conformation of the right half of the artificial analogs. The right halves in mycalamide A (1) and 10-epi-mycalamide A (2) were reported to have the conformation A and B, respectively.^{1a, 8b} Detailed NMR analysis suggested that the artificial analogs with the same configuration at C10 as that of 1 and their 10-epimers have the same conformation as A and B, respectively. The right half of MA-10 (14) was found to have a chair-boat-chair form C.



The cytotoxicity against HeLa cells and antiviral activity against HSV-1 and VZV of the synthetic compounds *in vitro* were tested along with 5-fluorouracil and acyclovir as the standard, and the results are shown in Table 1.¹²

Compound aga	Cytotoxicity ainst HeLa cells IC ₅₀	Antiviral activity against HSV-1		Antiviral activity against VZV	
		MIC ^b	IC ₅₀ c	MICd	IC ₅₀ e
5-Fluorouracil	3.0				
Acyclovir	-	1.563	50.0	6.25	>50.0
Mycalamide A (1)	< 0.03	< 0.391	< 0.391	< 0.391	<0.391
10-epi-Mycalamide A (2	3.0	12.5	12.5	1.563	12.5
MA-1 (5)	<0.03	<0.391	<0.391	< 0.391	<0.391
MA-2 (6)	3.0	50.0	50.0	1.563	>50.0
MA-3 (7)	0.03	3.125	0.391	< 0.391	1.563
MA-4 (8)	>10.0	50.0	>50.0	3.125	>50.0
MA-5 (9)	>10.0	50.0	12.5	< 0.391	12.5
MA-6 (10)	>10.0	50.0	>50.0	12.5	>50.0
MA-7 (11)	1.0	25.0	25.0	3.125	>50.0
MA-8 (12)	>10.0	>50.0	>50.0	25.0	>50.0
MA-9 (13)	3.0	50.0	>50.0	12.5	>50.0
MA-10 (14)	10.0	50.0	50.0	12.5	>50.0

Table 1. The Biological Activity of Mycalamide A, 10-epi-Mycalamide A and Mycalamide Analogs.^a

a) IC_{50} (µg/ml), MIC (µg/ml), HSV-1 (Herpes simplex virus type 1) and VZV (Varicella-zoster virus) were propagated in vero and HEL cells at 37 °C, respectively. b) against HSV-1. c) against vero cells. d) against VZV. e) against HEL cells.

Cytotoxicity against HeLa cells. Mycalamide A (1), MA-1 (5) and MA-3 (7) showed very potent cytotoxicity against HeLa cells. On the other hand, their corresponding 10-epimers, *i.e.*, **2**, MA-2 (**6**), and MA-4 (**8**), are 100-fold less active than **1**, **5**, and **7**, respectively, although the activity of **2** and **6** is still the same as that of 5-fluorouracil. Thus, the configuration at C10 is essential to show the strong cytotoxicity against HeLa cells. The presence of the C4-*exo*-methylene and C3-methyl groups is not an important factor for the potent cytotoxicity, although lack of the *exo*-methylene group slightly decreased the activity. The 7 β -hydroxy isomers, MA-5 (**9**) and its 10-epimer MA-6 (**10**), decreased the activity, which suggested that the configuration of the C7-hydroxy group is also essential for its potent cytotoxicity, which would be supported by the reported results;⁸ alkylation, acylation, and silylation of the 7-hydroxy group in 1 decreased the activity. It is noteworthy that MA-7 (**11**) and MA-9 (**13**), analogs replaced by glucose derivatives, showed the almost same activity as that of 5-fluorouracil.

Antiviral activity against HSV-1. A compound can be judged to have significant antiviral activity if its therapeutic ratio (TR=IC₅₀/MIC) is higher than that of acyclovir. The antiviral activity of mycalamide A (1), MA-1 (5), and MA-3 (7) against HSV-1 is very strong. However, their cytotoxicity (IC₅₀) against vero cells is also strong: TRs of all synthetic compounds tested are less than 1 (*cf.* TR of acyclovir = 32).

Antiviral activity against VZV. Although mycalamide A (1), MA-1 (5), and MA-3 (7) showed strong activity against VZV, their potent cytotoxicity (IC_{50}) against HEL cells was also observed. Interestingly, several synthetic compounds were found to have significant antiviral activity against VZV.

10-epi-Mycalamide A (2), MA-2 (6), MA-4 (8), MA-5 (9), and MA-7 (11) showed potent antiviral activity against VZV and low cytotoxicity against HEL cells: TRs of 2, 6, 8, 9, and 11 are 8, >32, >16, >32, and >16, respectively (cf. TR of acyclovir = >8). Thus, 7- or 10-epimeric compounds showed significant antiviral activity against VZV. It is noteworthy that MA-7 (11) also showed good antiviral activity against VZV.

The design and synthesis of the artificial analogs of mycalamides are under further investigation.

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References and Notes

- (a) Perry, N. B.; Blunt, J. W.; Munro, M. H. G.; Pannell, L. K. J. Am. Chem. Soc. 1988, 110, 4850. (b) Perry, N. B.; Blunt, J. W.; Munro, M. H. G.; Thompson, A. M. J. Org. Chem. 1990, 55, 223.
- (a) Sakemi, S.; Ichiba, T.; Kohmoto, S.; Saucy, G.; Higa, T. J. Am. Chem. Soc. 1988, 110, 4851. (b) Matsunaga, S.; Fusetani, N.; Nakao, Y. Tetrahedron 1992, 48, 8369. (c) Kobayashi, J.; Itagaki, F.; Shigemori, H., Sasaki, T. J. Nat. Prod. 1993, 56, 976.
- 3. Fusetani, N.; Sugawara, T.; Matsunaga, S. J. Org. Chem. 1992, 57, 3828.
- (a) Cardani, C.; Ghiringhelli, D.; Mondelli, R.; Quilico, A. Tetrahedron Lett. 1965, 2537. (b) Matsumoto, T.; Yanagiya, M.; Maeno, S.; Yasuda, S. Tetrahedron Lett. 1968, 6297. (c) Furusaki, A.; Watanabe, T.; Matsumoto, T.; Yanagiya, M. Tetrahedron Lett. 1968, 6301.
- 5. (a) Burres, N. S.; Clement, J. J. Cancer Res. **1989**, 49, 2935. (b) Ogawara, H.; Higashi, K.; Uchino, K.; Perry, N. B. Chem. Pharm. Bull. **1991**, 39, 2152.
- 6. Galvin, F.; Freeman, G. J.; Razi-Wolf, Z.; Benacerraf, B.; Nadler, L.; Reiser, H. Eur. J. Immunol. **1993**, 23, 283.
- (a) Hong, C. Y.; Kishi, Y. J. Org. Chem. 1990, 55, 4242. (b) Hong, C. Y.; Kishi, Y. J. Am. Chem. Soc. 1991, 113, 9693. (c) Nakata, T.; Matsukura, H.; Jian, D. L.; Nagashima, H. Tetrahedron Lett. 1994, 35, 8229. (d) Nakata, T.; Fukui, H.; Nakagawa, T.; Matsukura, H. Heterocycles 1996, 42, 159. (e) Roush, W. R.; Marron, T. G. Tetrahedron Lett. 1993, 34, 5421. (f) Marron, T. G.; Roush, W. R. Tetrahedron Lett. 1995, 36, 1581. (g) Roush, W. R.; Marron, T. G.; Pfeifer, L. A. J. Org. Chem. 1997, 62, 474. (h) Hoffmann, R. W.; Schlapbach, A. Tetrahedron Lett. 1993, 34, 7903. (i) Hoffmann, R. W.; Breitfelder, S.; Schlapbach, A. Helv. Chim. Acta 1996, 79, 346. (j) Davis, J.; Kocienski, P. J.; Boyle, T. Pol. J. Chem. 1994, 68, 2249. (k) Kocienski, P.; Raubo, P.; Davis, J. K.; Boyle, F. T.; Davies, D. E.; Richter, A. J. Chem. Soc. Perkin Trans. 1 1996, 1797.
- (a) Thompson, A. M.; Blunt, J. W.; Munro, M. H. G.; Perry, N. B.; Pannel, L. K. J. Chem. Soc. Perkin Trans. 1 1992, 1335. (b) Thompson, A. M.; Blunt, J. W.; Munro, M. H. G; Clark, B. M. J. Chem. Soc. Perkin Trans. 1 1994, 1025. (c) Thompson, A. M.; Blunt, J. W.; Munro, M. H. G; Perry N. B. J. Chem. Soc. Perkin Trans. 1 1995, 1233.
- (a) Nakata, T.; Nagao, S.; Mori, N.; Oishi, T. Tetrahedron Lett. 1985, 26, 6461. (b) Nakata, T.; Nagao, S.; Oishi, T. Tetrahedron Lett. 1985, 26, 6465.
- The ester 15 was prepared from methyl (R)-3-hydroxy-butanoate in 4 steps: (1) DHP, p-TsOH, CH₂Cl₂, rt; (2) LiAlH₄, Et₂O, rt; (3) (COCl)₂, DMSO, CH₂Cl₂, -78 °C; Et₃N, rt; (4) Ph₃P=CHCOOMe, benzene, reflux.
- 11. Oishi, T.; Nakata, T. Acc. Chem. Res. 1984, 17, 338.
- 12. Assay for antiviral activity against HSV-1. Confluent monolayer of vero (African green monkey kidney) cells was infected with 10 TCID₅₀ of HSV-1 (Miyama GC⁺ strain) in a well of a 96-well microtestplate in the presence of serial 2-fold dilution of each drug (in MEM medium supplemented with 1% fetal calf serum, total volume; 200 μl), and cultured for 3 days in 5% CO₂ and 95% humidified air at 37 °C. The minimum drug concentration which inhibits 50% of the viral cytopathic effect (cpe) was designated as MIC.

Assay for antiviral activity against VZV. Confluent monolayer of HEL (human embryonic lung) cells was infected with 10 TCID₅₀ of VZV (Kawaguchi strain) in the presence of serial 2-fold dilution of each drug and cultured for 7 days. All other culture conditions were the same as above. The minimum drug concentration which inhibits 100% of the viral cpe was designated as MIC.

Determination of IC50. Confluent monolayer of HEL cells, vero cells, or HeLa cells (human cervical cancer cell line) was cultured in the presence of serial 2-fold dilution of each drug for 3 days in virus-free conditions. The viability was determined by MTT method.¹³ The drug concentration whose absorbance at 540 nm indicates 50% of the drug-free control was designated as IC50.

13. Tada, H.; Shiho, O.; Kuroshima, K.; Koyama, M.; Tsukamoto, K. J. Immunol. Method 1986, 93, 157.