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Organoantimony(III) halide complexes with azastibocine framework as potential antitumor agents: Correlation between cytotoxic activity and N→Sb inter-coordination

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# Graphic abstract



1	Organoantimony(III) Halide Complexes with Azastibocine
2	Framework as Potential Antitumor Agents: Correlation between
3	Cytotoxic Activity and N $\rightarrow$ Sb Inter-Coordination
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15	<sup>1</sup> Lei and Liu contributed equally to this work.

# 16 Abstract

17	The relationship between chemical structure and in vitro cytotoxic activities of a
18	series of azastibocine-framework organoantimony(III) halide complexes against
19	cancerous (HepG2, MDA-MB-231, MCF-7 and HeLa) and nonmalignant (HEK-293)
20	cell lines was studied for the first time. A positive correlation between cytotoxic
21	activity and the length of $N \rightarrow Sb$ coordinate bond on azastibocine framework of same
22	nitrogen substituent was observed. By comparison, the organoantimony(III) complex
23	6-cyclohexyl-12-fluoro-5,6,7,12-tetrahydrodibenzo[ $c,f$ ][ $1,5$ ]azastibocine (C4)
24	exhibited the highest selectivity index, giving a $IC_{50}$ (nonmalignant)/ $IC_{50}$ (cancerous)
25	ratio of up to 8.33. The results of cell cycle analysis indicated that the inhibitory effect
26	of C4 on the cellular viability was caused by cell cycle arrest mainly at the S phase.
27	The necrosis induced by C4 was confirmed by the Trypan blue dye exclusion test and
28	the increase of lactic dehydrogenase (LDH) released in the culture medium.
29	Furthermore, evaluation of the levels of intracellular reactive oxygen species (ROS) in
30	MDA-MB-231 cells, by quantifying the relative fluorescence units (RFU) using
31	spectrofluorometer, indicated that cytotoxic activity of C4 is dependent on the
32	production of ROS. This work established the correlation between cytotoxic activity
33	and $N \rightarrow Sb$ inter-coordination, a finding that provided theoretical and experimental
34	basis for in-depth design of antimony-based organometallic complexes as potential
35	anticancer agents.

# 36 Keywords

- 37 Organoantimony(III) complex; Cytotoxic Activity; Structure-activity relationship;
- 38 Necrosis; Reactive oxygen species

# 39 Abbreviations

- 40 CCK-8, Cell Counting Kit-8; IC<sub>50</sub>, half-maximal inhibitory concentration; SI,
- 41 selectivity index; PI, propidium iodide; PS, phosphatidylserine; LDH, lactic
- 42 dehydrogenase; ROS, reactive oxygen species; DCFH-DA,
- 43 5-(and-6)-carboxy-2',7'-dichlorodihydrofluorescein diacetate; NAC, *N*-acetylcysteine

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# 44 **1. Introduction**

The rich diversity of coordination chemistry provides exciting prospects for the design of novel metallodrugs with unique biological activity [1]. Since the landmark report of cisplatin compounds in cancer therapy, continuous efforts have been made in the investigation of new metallic and organometallic complexes for the purpose of developing metal-based chemotherapeutics of low toxicity that are endowed with anticancer activity [2–6].

In the past century, antimony-based drugs have been extensively investigated 51 because of their clinically therapeutic efficacy against leishmaniosis [7,8]. Like 52 53 cancer, the parasite divides rapidly and may share some common pathways that could be targets for chemotherapy [7,8]. Therefore, there is an increasing interest in 54 55 antitumor application of antimony compounds, and significant progress has been made [9,10]. For instance, inorganic trivalent antimonials such as Sb<sub>2</sub>O<sub>3</sub> were found 56 to show biological effects on acute promyelocytic leukemia (APL) that closely 57 resemble those of As<sub>2</sub>O<sub>3</sub> [11]. A number of antimony(III) compounds complexed with 58 polydentate ligands such as carboxylic acids [12–14], hydrazones [15], thioamides 59 [16], and dithiocarbamates [17], were synthesized and assessed as biomedical agents 60 for anticancer purpose. Currently, the most studied antimony(III) compounds in the 61 context of antitumor activity are the ones with at least one antimony-carbon covalent 62 bond, the so-called organoantimony complexes [18–24]. The first example showing 63 the antitumor property of organoantimony(III) species was reported by Silvestru and 64 coworkers, wherein found that diphenylantimony(III) 65 they derivatives of

66 dithiophosphorus ligands exhibited inhibitory effects on the growth of Ehrlich ascites tumor [18]. Meanwhile, the pharmacological applications of antimony(V)-based 67 68 organometallic complexes in cancerous cells proliferation were also reported [25–29]. The potential inhibitory effects of triphenylantimony(V) polyamines on cancer-related 69 cell lines were described by Carraher et al. [25]. Demicheli and coworkers disclosed 70 71 the *in vitro* cytotoxic activity of a series of pentavalent organoantimony complexes against malignant carcinoma cells, such as human chronic myelogenous leukemia 72 (K562) and murine metastatic melanoma (B16F10) [26,27]. Arylantimony(V) 73 exo-7-oxa-bicyclo[2,2,1]heptane (ene)-3-arylamide-2-acid 74 derivatives of and arylhydroxamic acid were introduced by Li and coworkers as potential antitumor 75 agents against a diversity of human neoplastic cell lines [27,28]. Among those 76 77 antimony-based metallic and organometallic complexes, the presence of coordination between antimony center and ligand is relevant to their cellular activity. In most cases, 78 the coordination of antimony by ligands resulted in compounds with antitumor 79 potency greater than that of the uncoordinated ligands [13,15,27–29] as well as their 80 81 antimony-containing precursors [15,28,29]. The strong development in this field motivated us to elucidate the relationship between coordination effect and antitumor 82 83 potency for the purpose of developing organoantimony-based chemotherapeutics in cancer therapy. To our knowledge, relatively few information is available in the 84 literature about the related chemistry between coordination effect and antitumor 85 activity of antimony-based organometallic complexes. 86

87 Previously our group reported the anti-proliferative activity of heterocyclic

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88 hypervalent organoantimony(III) complexes on human alveolar adenocarcinoma cell line (A549) [30]. Although full details are yet to be revealed, preliminary data have 89 90 indicated that anti-proliferative activity detected over these complexes could be attributed to the coordination between the antimony and nitrogen atoms. Very 91 recently, Hadjikakou and coworkers developed a series of antimony(III) halide 92 93 complexes with dithiocarbamate ligands, and they found that the halogen type causes 94 no influence on the antitumor activity [17]. With this in mind, we herein revealed for the first time the correlation between cytotoxicity and coordination effect by studying 95 the *in vitro* structure-activity relationship (SAR) of a series of azastibocine-framework 96 organoantimony(III) halide complexes against different cell lines, including solid 97 cancerous (human liver hepatocellular carcinoma cell line, HepG2; human breast 98 cancer cell line, MDA-MB-231; human breast adenocarcinoma cell line, MCF-7; and 99 human cervical carcinoma cell line HeLa) and nonmalignant (human embryonic 100 kidney cell line, HEK-293) cells. Moreover, considering the selectivity index that is 101 derived from  $IC_{50}$  (nonmalignant)/ $IC_{50}$  (cancerous) ratio, the most potent compound C4 102 103 selected for further mechanistic investigation. Finally, we describe was organoantimony complex C4 driven induction of necrosis in MDA-MB-231 cells 104 105 through intracellular ROS production.

# 106 **2 Results and Discussion**

# 107 *2.1 Chemistry*



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Fig.1. a) Molecular structures of organoantimony(III) halide complexes with azastibocine
framework. b) ORTEP diagram of the X-ray molecular structure of organoantimony(III)
and the transformed C1 C12

111 complexes C1–C12.

Table 1
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Selected bond lengths (Å) and angles (deg) of organoantimony(III) halide complexes C1-
C12.

C1		C2		C3		C4	
Sb(1)-C(1)	2.144(4)	Sb(1)-C(6)	2.143(2)	Sb(1)-C(6)	2.154(3)	Sb(1)-C(6)	2.140(3)
Sb(1)-C(8)	2.134(4)	Sb(1)-C(10)	2.157(2)	Sb(1)-C(7)	2.166(3)	Sb(1)-C(7)	2.145(3)
C(1)-Sb(1)-C(8)	98.2(1)	C(6)-Sb(1)-C(10)	99.23(8)	C(6)-Sb(1)-C(7)	98.8(1)	C(6)-Sb(1)-C(7)	98.5(1)
Cl(1)-Sb(1)-N(1)	162.92(7)	Br(1)-Sb(1)-N(1)	163.53(4)	I(1)-Sb(1)-N(1)	163.34(6)	F(1)-Sb(1)-N(1)	156.42(7)
Sb(1)-N(1)	2.397(2)	Sb(1)-N(1)	2.387(2)	Sb(1)-N(1)	2.400(2)	Sb(1)-N(1)	2.450(2)
Sb(1)-Cl(1)	2.5572(9)	Sb(1)-Br(1)	2.7142(5)	Sb(1)-I(1)	2.9650(4)	Sb(1)-F(1)	2.015(2)
C5		C6		C7		C8	
Sb(1)-C(1)	2.150(2)	Sb(1)-C(1)	2.158(3)	Sb(1)-C(1)	2.166(3)	Sb(1)-C(1)	2.131(3)
Sb(1)-C(8)	2.155(2)	Sb(1)-C(8)	2.156(3)	Sb(1)-C(8)	2.175(3)	Sb(1)-C(8)	2.153(3)
C(1)-Sb(1)-C(8)	100.53(7)	C(1)-Sb(1)-C(8)	100.4(1)	C(1)-Sb(1)-C(8)	96.3(1)	C(1)-Sb(1)-C(8)	91.80(9)
CI(1)-Sb(1)-N(1)	161.98(4)	Br(1)-Sb(1)-N(1)	163.22(6)	I(1)-Sb(1)-N(1)	163.92(6)	F(1)-Sb(1)-N(1)	162.92(7)
Sb(1)-N(1)	2.466(2)	Sb(1)-N(1)	2.469(2)	Sb(1)-N(1)	2.498(3)	Sb(1)-N(1)	2.522(2)
Sb(1)-Cl(1)	2.5173(5)	Sb(1)-Br(1)	2.6620(4)	Sb(1)-I(1)	2.8995(3)	Sb(1)-F(1)	1.998(2)
C9		C10		C11		C12	
Sb(1)-C(1)	2.147(2)	Sb(1)-C(6)	2.153(2)	Sb(1)-C(6)	2.166(3)	Sb(1)-C(6)	2.127(7)
Sb(1)-C(14)	2.155(3)	Sb(1)-C(7)	2.151(2)	Sb(1)-C(7)	2.157(3)	Sb(1)-C(10)	2.135(7)
C(1)-Sb(1)-C(14)	95.60(9)	C(6)-Sb(1)-C(7)	96.18(8)	C(6)-Sb(1)-C(7)	97.6(1)	C(6)-Sb(1)-C(10)	98.2(3)
Cl(1)-Sb(1)-N(1)	161.60(5)	Br(1)-Sb(1)-N(1)	162.99(4)	I(1)-Sb(1)-N(1)	163.59(6)	F(1)-Sb(1)-N(1)	155.9(2)
Sb(1)-N(1)	2.467(2)	Sb(1)-N(1)	2.446(2)	Sb(1)-N(1)	2.462(3)	Sb(1)-N(1)	2.495(5)
Sb(1)-Cl(1)	2.5579(7)	Sb(1)-Br(1)	2.7631(5)	Sb(1)-I(1)	2.9463(3)	Sb(1)-F(1)	2.026(6)

112	The synthesis of organoantimony chlorides, which were labeled C1, C5 and C9,
113	was carried out as previously reported [30]. Organoantimony bromides, iodides, and
114	fluorides could be prepared by mixing the organoantimony chlorides with the
115	corresponding inorganic salts (i.e. potassium bromide, potassium iodide or silver
116	fluoride) in 1:10 or 1:1 molar ratio as specified in the Experimental Section. Fig. 1a
117	depicts the general molecular structures of the complexes of formula
118	$RN(CH_2C_6H_4)SbX$ , where $R = Cy$ (cyclohexyl), Ph (phenyl), or <sup>t</sup> Bu (tertiary butyl)
119	and $X = Cl$ , Br, I, F. These complexes readily recrystallized from the reaction
120	mixtures as colorless crystals in moderate to excellent yields and could be kept in
121	open air for a long-term period without showing any detectable change in <sup>1</sup> H NMR

122 spectra (see Supplementary Materials for details).

The molecular structures of C1-C12 are unambiguously characterized by NMR 123 124 spectroscopy, elemental analysis as well as single crystal X-ray diffraction analysis. ORTEP complexes 125 The diagrams of these revealed that the central antimony-containing part shows analogous pseudo-trigonal bipyramidal structures 126 127 with a butterfly-shaped ligand where the nitrogen and halogen atoms are located at the apical positions and the two carbon atoms adjacent to the antimony atom are at the 128 equatorial positions along with a lone electron pair of antimony (Fig. 1b). The lengths 129 of N $\rightarrow$ Sb coordinate bond in these complexes are within the 2.387(2)–2.522(2) Å 130 range (Table 1), which are slightly longer than the sum of the covalent radii (2.11 Å) 131 but much shorter than the sum of the van der Waals radii (3.74 Å) [31,32]. The results 132 133 evidence the inter-coordination between the antimony and the nitrogen atoms, and the strength of  $N \rightarrow Sb$  coordination on the azastibocine framework could be adjusted by 134 synergistic modulating the property of nitrogen substituents and halogen atoms 135 adjacent to the central antimony atom. The lengths of  $N \rightarrow Sb$  coordinate bond of 136 phenyl derivatives (C5–C8) are obviously longer than those of cyclohexyl derivatives 137 (C1–C4). Given the inverse electronic properties of phenyl and cyclohexyl group, it is 138 apparent that the strength of  $N \rightarrow Sb$  inter-coordination is weakened with the increase 139 of electron-withdrawing ability of the nitrogen substituents. The steric effect of the 140 nitrogen substituents also significantly affect the intramolecular interaction between 141 142 the antimony and nitrogen atoms, as seen in the lengths of  $N \rightarrow Sb$  coordinate bond of 143 tert-butyl derivatives C9-C12. In addition, the coordinate bond lengths of 144 organoantimony(III) fluorides [2.450(2) Å, C4; 2.522(2) Å, C8; 2.495(5) Å, C12] are

145 distinctly longer than those of chlorides, bromides and iodides, owing to the strong

146 electron-withdrawing ability of fluorine atom.

## Table 2

Crystallographic data for organoan	imony(III) halide complexes C1–C12.
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Complex	C1	C2	C3	C4	C5	C6
Empirical formula	C <sub>20</sub> H <sub>23</sub> CINSb	C <sub>20</sub> H <sub>23</sub> BrNSb	C <sub>20</sub> H <sub>23</sub> INSb	C <sub>20</sub> H <sub>23</sub> FNSb	C <sub>20</sub> H <sub>17</sub> CINSb	C <sub>20</sub> H <sub>17</sub> BrNSb
Formula weight	434.59	479.05	526.04	418.14	428.55	473.01
Temperature/K	293	273	100	100	273	298
Crystal system	monoclinic	monoclinic	monoclinic	monoclinic	monoclinic	monoclinic
Space group	P2(1)/c	P2(1)/n	P2(1)/n	P2(1)/n	P2(1)/n	P2(1)/n
a/Å	10.0771(7)	10.2338(5)	11.3664(3)	9.3503(3)	9.3782(4)	9.5744(3)
b/Å	16.2881(12)	16.5885(7)	12.5996(3)	15.9263(5)	10.1833(4)	10.2962(3)
c/Å	12.2040(9)	12.0033(5)	13.0792(4)	11.7239(4)	18.1033(8)	18.1939(6)
αlo	90.00	90.00	90.00	90.00	90.00	90.00
β <b>/</b> °	111.812(10)	113.346(2)	94.703(2)	104.044(3)	102.895(10)	100.795(1)
γ <b>/</b> °	90.00	90.00	90.00	90.00	90.00	90.00
V/Å <sup>3</sup>	1859.7(2)	1870.89(15)	1866.79(9)	1693.69(10)	1685.3(12)	1761.81(10)
Z	4	4	4	4	4	4
No. of reflections collected	10058	30234	12759	10827	9923	10613
No. of unique reflections	3644	4636	3284	2982	4032	3417
R <sub>int</sub>	0.047	0.0363	0.0390	0.0376	0.015	0.038
R <sub>1</sub> (reflections)	0.0324	0.0196	0.0236	0.0256	0.0203	0.0294
wR <sub>2</sub> (reflections)	0.0917	0.0416	0.0561	0.0598	0.0560	0.0709
GOF on <i>F</i> <sup>2</sup>	1.048	1.066	1.022	1.097	1.077	1.041
Complex	C7	C8	C9	C10	C11	C12
Complex Empirical formula	C7 C <sub>20</sub> H <sub>17</sub> INSb	C8 C <sub>20</sub> H <sub>17</sub> FNSb	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb	C10 C <sub>18</sub> H <sub>21</sub> BrNSb	<b>C11</b> C <sub>18</sub> H <sub>21</sub> INSb	<b>C12</b> C <sub>18</sub> H <sub>21</sub> FNSb
Complex Empirical formula Formula weight	<b>C7</b> C <sub>20</sub> H <sub>17</sub> INSb 520.00	<b>C8</b> C <sub>20</sub> H <sub>17</sub> FNSb 412.10	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56	<b>C10</b> C <sub>18</sub> H <sub>21</sub> BrNSb 453.02	<b>C11</b> C <sub>18</sub> H <sub>21</sub> INSb 500.01	<b>C12</b> C <sub>18</sub> H <sub>21</sub> FNSb 392.11
Complex Empirical formula Formula weight Temperature/K	<b>C7</b> C <sub>20</sub> H <sub>17</sub> INSb 520.00 298	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273	<b>C10</b> C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273	<b>C11</b> C <sub>18</sub> H <sub>21</sub> INSb 500.01 100	<b>C12</b> C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273
Complex Empirical formula Formula weight Temperature/K Crystal system	<b>C7</b> C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic	<b>C10</b> C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic	<b>C11</b> C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic	<b>C12</b> C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic
Complex Empirical formula Formula weight Temperature/K Crystal system Space group	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i>	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i>	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i>	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i>	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc
Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8)	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic <i>P2(1)/c</i> 12.3974(4)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9)
Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8)	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12) 15.933(2)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic <i>P2(1)/c</i> 12.3974(4) 9.4136(3)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9) 10.6409(6)
Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å c/Å	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4) 18.0957(7)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8) 15.7161(11)	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12) 15.933(2) 11.4491(14)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5) 11.4078(5)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic <i>P2(1)/c</i> 12.3974(4) 9.4136(3) 14.8803(5)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9) 10.6409(6) 17.8551(10)
Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å c/Å α/°	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4) 18.0957(7) 90.00	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8) 15.7161(11) 90.00	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5) 11.4078(5) 90.00	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic <i>P2(1)/c</i> 12.3974(4) 9.4136(3) 14.8803(5) 90.00	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00
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Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å c/Å c/Å α/° β/° γ/° V/Å <sup>3</sup>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8) 15.7161(11) 90.00 101.559(1) 90.00 1633.6(2)	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1673.5(4)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic <i>P2(1)/c</i> 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3)
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Complex Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å c/Å c/Å α/° β/° γ/° V/Å <sup>3</sup> Z No. of reflections collected No. of unique reflections	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8) 15.7161(11) 90.00 101.559(1) 90.00 1633.6(2) 4 9422 3560	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 monoclinic <i>P2(1)/n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1673.5(4) 4 9317 3889	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232
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ComplexEmpirical formulaFormula weightTemperature/KCrystal systemSpace group $a/Å$ $b/Å$ $c/Å$ $\alpha/°$ $\beta/°$ $\gamma/°$ $\gamma/°$ $V/Å^3$ ZNo. of reflections collectedNo. of unique reflections $R_{int}$ $R_1(reflections)$ $wR_2(reflections)$	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic <i>P2(1)/c</i> 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510 0.055 0.0354 0.0912	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic <i>P2(1)/n</i> 10.2974(8) 10.3031(8) 15.7161(11) 90.00 101.559(1) 90.00 1633.6(2) 4 9422 3560 0.063 0.0323 0.0840	$\begin{array}{c} \hline \textbf{C9} \\ \hline \textbf{C}_{18}\textbf{H}_{21}\text{CINSb} \\ 408.56 \\ 273 \\ \hline \textbf{monoclinic} \\ P2(1)/n \\ 9.7692(12) \\ 15.933(2) \\ 11.4491(14) \\ 90.00 \\ 110.103(2) \\ 90.00 \\ 1673.5(4) \\ 4 \\ 9317 \\ 3889 \\ 0.027 \\ 0.0326 \\ 0.0860 \\ \end{array}$	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic <i>P2(1)/n</i> 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978 0.0437 0.0143 0.0541	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058 0.0353 0.0237 0.0516	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic Cc 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232 0.0229 0.0419 0.1205

147 2.2 Biological activity

148 2.2.1 Anticancer/cytotoxic activity and structure-activity relationship (SAR) study

#### Table 3

Cytotoxic effect of organoantimony(III) halide complexes C1–C12, nitrogen-containing precursors P1–P3 and cisplatin on various cancerous and nonmalignant cell lines after 24 h.

Cell lines	HepG2		MDA-MB-231		MCF-7		HeLa		HEK-293
Compound	$\rm IC_{50}\pm SD^a$	Sl <sup>b</sup>	$IC_{50} \pm SD$	SIc	$IC_{50} \pm SD$	Sl <sup>d</sup>	$IC_{50}\pm SD$	Sl <sup>e</sup>	$IC_{50} \pm SD$
C1	$\textbf{1.84} \pm \textbf{0.42}$	5.05	$\textbf{1.40} \pm \textbf{0.32}$	6.64	$\textbf{6.08} \pm \textbf{1.38}$	1.53	$1.63\pm0.49$	5.70	$9.29 \pm 1.16$
C2	$\textbf{4.86} \pm \textbf{0.91}$	2.10	$\textbf{3.63} \pm \textbf{0.87}$	2.82	$\textbf{6.39} \pm \textbf{1.45}$	1.60	$\textbf{3.12} \pm \textbf{0.45}$	3.28	$10.23\pm0.55$
C3	$1.61 \pm 0.33$	5.63	$\textbf{1.28} \pm \textbf{0.54}$	7.08	$5.79 \pm 1.56$	1.56	$1.13\pm0.21$	8.02	$9.06 \pm 1.31$
C4	$1.06\pm0.17$	7.15	$\textbf{0.91} \pm \textbf{0.22}$	8.33	$\textbf{2.14} \pm \textbf{0.93}$	3.54	$\textbf{0.93} \pm \textbf{0.15}$	8.15	$\textbf{7.58} \pm \textbf{0.89}$
C5	$\textbf{2.78} \pm \textbf{0.59}$	3.07	$1.72\pm0.33$	4.97	$\textbf{3.14} \pm \textbf{0.81}$	2.72	$\textbf{3.32} \pm \textbf{0.50}$	2.57	$\textbf{8.54} \pm \textbf{1.88}$
C6	$1.90\pm0.12$	4.10	$1.05\pm0.13$	7.42	$\textbf{2.61} \pm \textbf{0.60}$	2.98	$\textbf{2.34} \pm \textbf{0.32}$	3.33	$7.79 \pm 1.36$
C7	$1.87\pm0.11$	3.33	$1.04\pm0.11$	5.98	$\textbf{2.39} \pm \textbf{1.67}$	2.60	$\textbf{2.06} \pm \textbf{0.29}$	3.02	$\textbf{6.22} \pm \textbf{1.69}$
C8	$\textbf{0.88} \pm \textbf{0.32}$	4.41	$\textbf{0.52} \pm \textbf{0.02}$	7.46	$\textbf{2.31} \pm \textbf{0.32}$	1.68	$1.12 \pm 0.21$	3.46	$3.88 \pm 1.01$
C9	1.41± 0.06	4.03	$\textbf{0.84} \pm \textbf{0.23}$	6.76	$\textbf{3.41} \pm \textbf{0.30}$	1.67	$1.98 \pm 0.54$	2.87	$\textbf{5.68} \pm \textbf{1.32}$
C10	$\textbf{2.74} \pm \textbf{0.63}$	2.27	$\textbf{1.85} \pm \textbf{0.25}$	3.36	$6.34 \pm 1.31$	0.98	$\textbf{4.78} \pm \textbf{0.83}$	1.30	$\textbf{6.22} \pm \textbf{1.07}$
C11	$\textbf{1.84} \pm \textbf{0.28}$	4.05	$\textbf{0.97} \pm \textbf{0.19}$	7.69	$\textbf{4.89} \pm \textbf{1.13}$	1.53	$\textbf{2.09} \pm \textbf{0.62}$	3.57	$\textbf{7.46} \pm \textbf{0.66}$
C12	$0.71 \pm 0.06$	3.51	$\textbf{0.51} \pm \textbf{0.12}$	4.88	$\textbf{0.96} \pm \textbf{0.83}$	2.59	$\textbf{1.08} \pm \textbf{0.47}$	2.31	$\textbf{2.49} \pm \textbf{0.67}$
P1	>50.00	-	>50.00	-	>50.00	-	>50.00	-	>50.00
P2	>50.00	-	>50.00	-	>50.00	-	>50.00	-	>50.00
P3	>50.00	-	>50.00	-	>50.00	-	>50.00	-	>50.00
cisplatin	$13.19 \pm 4.51$	3.71	$12.63 \pm 1.22$	3.88	$\textbf{22.75} \pm \textbf{8.59}$	2.15	$17.35 \pm 3.64$	2.82	$\textbf{48.96} \pm \textbf{8.75}$

Cancerous Cell Lines: HepG2 (human liver hepatocellular carcinoma cell line), MDA-MB-231 (human breast cancer cell line), MCF-7 (human breast adenocarcinoma cell line) and HeLa (human cervical carcinoma cell line). Nonmalignant Cell Line: HEK-293 (human embryonic kidney cell line). IC<sub>50</sub>: concentration that is cytotoxic against 50% of cell lines. SI: selectivity index. <sup>*a*</sup> The IC<sub>50</sub> values were determined through non-linear regression analysis; Each well was triplicated and each experiment was repeated at least three times. IC<sub>50</sub> values quoted are mean  $\pm$  SD ( $\mu$ M). <sup>*b*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MDA-MB-231. <sup>*d*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7. <sup>*e*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7.

149 The anticancer activity of the synthesized organoantimoy(III) halide complexes was

evaluated in a panel of four human solid tumor cell lines derived from hepatocellular carcinoma (HepG2), breast cancer (MDA-MB-231 and MCF-7) and cervical carcinoma (HeLa), together with cisplatin as positive control. Cell viability was assessed using Cell Counting Kit-8 (CCK-8) as described and the results of half-maximal inhibitory concentration (IC<sub>50</sub>) that represents the drug concentration required to inhibit cell growth by 50% are summarized in Table 3. The results

156 demonstrated that all of the organoantimony(III) halide complexes exhibited much higher cytotoxic activity against the selected cancerous cell lines than cisplatin and 157 158 the nitrogen-containing precursors P1-P3. It was also observed that the modulation of 159 the of the nitrogen substituent halogen type and atom on the 160 tetrahydrodibenzo[*c*,*f*][*1*,*5*]azastibocine framework would result in distinct 161 antineoplastic performance towards the same cancerous cell line.

To further elucidate the related chemistry between coordination effect and 162 cytotoxic activity of organoantimony complexes, a systematic structure-activity 163 relationship study was explored. Given the different electronic and steric properties of 164 165 the nitrogen substituents as well as based on the results of our preliminary study on antimony-based organometallic antitumor agents [30], a ranking of cytotoxic activity 166 across the organoantimony complexes with the same nitrogen substituent (e.g. 167 cyclohexyl, phenyl and tertiary butyl) and  $N \rightarrow Sb$  coordinate bond length was 168 performed (Fig. 2a-c). In each case, the nonlinear correlations between the lengths of 169  $N \rightarrow Sb$  coordinate bond, which were modulated simply by the halogen type, and the 170 171 cytotoxic activities of the corresponding organoantimony halide complexes were found, respectively. When the complexes were ranked by decreasing coordination 172 173 effect, i.e., with increasing  $N \rightarrow Sb$  coordinate bond lengths, the complexes having the same nitrogen substituent showed increasing inhibitory activities on all four cancerous 174 cell lines. On the other hand, the most potent cytotoxic activities were observed over 175 the organoantimony fluorides (C4, C8 and C12) in each set, albeit there is no trend 176 for the complexes bearing the other halogens (e.g. chloride, bromide and iodide). 177

178 According to this relationship and experimental results, we deduce that the cytotoxic activity of these complexes was determined by the level of exposure of the 179 antimony(III) center. With decreasing extent of N-Sb coordination, the cytotoxic 180 activities of organometal moiety towards cancerous cells were increased. In other 181 words, the nitrogen-containing ligand may serve as an auxiliary moiety to decrease 182 the cytotoxicity of the antimony(III) center. With this in mind, we speculate that there 183 are two potential functions of  $N \rightarrow Sb$  inter-coordination in these organoantimony(III) 184 complexes: (1) maintaining the skeleton structure to improve the air-stability and 185 186 water-tolerance; (2) reducing the level of exposure of the antimony(III) center to decrease the cytotoxicity of trivalent antimonial. 187



Fig. 2. Correlation between N→Sb coordinate bond length in organoantimony complexes
with the same nitrogen substituent (a: cyclohexyl; b: phenyl; c: *tert*-butyl) and corresponding
anticancer activity against human tumor cells: HepG2 (red line), MDA-MB-231 (purple line),
MCF-7 (blue line) and HeLa (green line).

188

To test the effect of these organoantimony(III) halide complexes on nonmalignant cells, we employed human embryonic kidney cells (HEK-293). The study of the effects of these twelve complexes on nonmalignant cells is of particular interest in terms of their potential application in clinical practice. After 24 h incubation, the IC<sub>50</sub> values of complexes C1–C12 on HEK-293 are within the 2.49–10.23  $\mu$ M range. By

198	comparison with the commercial cisplatin, the majority of the synthesized
199	organoantimony(III) complexes exhibited relatively higher selectivity index in the
200	biological tests, especially so with the complex C4
201	6-cyclohexyl-12-fluoro-5,6,7,12-tetrahydrodibenzo[ $c,f$ ][ $1,5$ ]azastibocine, which gives
202	a IC <sub>50</sub> (nonmalignant)/IC <sub>50</sub> (cancerous) ratio of up to 8.33. With such a performance,
203	C4 opens a potential therapeutic window for antimony-based organometallic
204	antitumor drugs, and we focused our investigation on the mechanistic study of C4 in
205	MDA-MB-231 cells.

206 2.2.2 Cell Cycle Analysis

Generally, metal-based chemotherapeutic agents exhibit perturbation effect on cell 207 cycle that is composed of  $G_1$ , S,  $G_2$ , M, and  $G_0$  phases in proliferating cells [33,34]. 208 209 At different stages of the cell cycle, cell nuclei contain different amounts of DNA. Therefore, according to the DNA content detected by propidium iodide (PI) staining, 210 cell cycle is commonly described on the basis of  $G_0/G_1$ , S, and  $G_2/M$  phases. To 211 investigate whether the cytotoxic effect of C4 was caused by cell cycle arrest, PI 212 staining and flow cytometry analysis of cells was performed in MDA-MB-231 cells. 213 As shown in Fig. 3, after independently incubated with C4 at 0.25, 0.5, 1, and 2 214 equipotent concentrations of IC<sub>50</sub> (0.91  $\mu$ M) for 24 h, the MDA-MB-231 cells in 215  $G_0/G_1$  phase showed a significant population decrease from 50.4% (untreated cells) to 216 217 3.7% (1.82  $\mu$ M of C4), accompanied by a dose-dependent increase in the fraction of cells in the S (68.9% compare to 39.8% in untreated cells) and  $G_2/M$  phases (26.8% 218 versus 9.8%). The results confirmed the retardation effect of organoantimony fluoride 219

C4 on the cell cycle progression, and it is reasonable to deduce that the observedinhibitory effect on the cellular viability of MDA-MB-231cells was caused by cell

222 cycle arrest mainly at the S phase, which represents a period of DNA replication.



223

Fig. 3. Effect of C4 on the cell cycle distribution of MDA-MB-231 cells. a) Flow cytometry analysis was performed on MDA-MB-231 cells treated with C4 for 24 h. b) With the percentages of cells in different phases quantified. Data were shown as mean  $\pm$  S.D. (n=3). \**P* < 0.05, \*\**P* < 0.01 compared with the population of cells in the control group.

228 2.2.3 Apoptosis assay

Because prolonged cell cycle arrest may influence cellular viability through processes including anti-proliferation and cell death [35,36], cell apoptosis has been adopted as a vehicle for cancer treatment [37]. We next evaluated the apoptotic

activity of C4-treated MDA-MB-231 cells by means of Annexin-V and PI double 232 233 staining using a FACSCalibur flow cytometer. During apoptosis, phosphatidylserine (PS) is translocated from the cytoplasmic face of the plasma membrane to the cell 234 surface. Thus, apoptotic cells can be identified by the presence of PS on the cell 235 surface. Annexin V has a strong  $Ca^{2+}$ -dependent affinity for PS, and therefore can be 236 237 used as a probe for detecting apoptosis. PI can only pass through the membranes of later apoptotic and necrotic cells, and consequently intercalate into their nucleic acids, 238 since the integrity of their plasma and nuclear membranes was decreased. This allows 239 apoptotic cells (annexin- $V^+/PI^-$  and annexin- $V^+/PI^+$ ) to be distinguished from living 240 241 (annexin- $V^{-}/PI^{-}$ ) and necrotic (annexin- $V^{-}/PI^{+}$ ) cells in flow cytometry analysis.



242

**Fig. 4.** a) Apoptosis of MDA-MB-231 cells induced by the C4 (0,  $0.25 \times IC_{50}$ ,  $0.5 \times IC_{50}$ ,  $1 \times IC_{50}$ 

244	IC <sub>50</sub> , and $1 \times IC_{50}$ ) for 48 h. b) All data were obtained from three independent experiments
245	and presented as the mean $\pm$ S.D. * <i>P</i> < 0.05, ** <i>P</i> < 0.01 compared with control.

246 In MDA-MB-231 cells incubated with compound C4 at a 0.25 equipotent concentration of  $IC_{50}$  for 48 h, the proportion of apoptotic (early + late) cells 247 increased to 2.07%, relatively slight compared to that of DMSO alone (1.51%). In 248 249 contrast, even at a low C4 dosage of 0.23  $\mu$ M, the treatment resulted in 10.43% of the 250 MDA-MB-231 cells becoming necrotic, obviously higher than the 2.81% value of necrotic cells in untreated control (Fig. 4). With the increase of C4 dosage to 0.5, 1, 251 252 and 2 equipotent concentrations of  $IC_{50}$ , there was dose-dependent enhancement of 253 both apoptotic (early + late) and necrotic cells to 8.76% and 38.88%, respectively. It is apparent that the main cause of cell death in human breast adenocarcinoma was the 254 necrosis induced by compound C4. 255

# 256 2.2.4 Lactate dehydrogenase release assay

Lactate dehydrogenase (LDH) is a stable cytoplasmic enzyme that is found in all 257 cells. The leaking of LDH enzyme from the cytosol into the surrounding culture 258 259 medium is generally regarded as an indicator of necrosis, and reflects a loss of membrane integrity in dead cells [38]. Because compound C4 failed to significantly 260 induce apoptosis, while the Annexin V-FITC/propidium iodide staining revealed that 261 C4 induced necrosis robustly, an LDH release assay was performed to further 262 investigate the effects of C4 on the LDH activity of treated MDA-MB-231 cells as 263 specified in the experimental section. After treatment of MDA-MB-231 cells with 264 compound C4 (0.23, 0.46, 0.91, and 1.82 µM for 48 h), there was somewhat elevation 265

of LDH release levels in the culture medium (Fig. 5a). Especially in the cases where the cancerous cells were exposed to C4 at concentration of 0.91 and 1.82  $\mu$ M, the levels of LDH release were almost 3.21 and 3.05 fold (*P* < 0.01) higher than that in non-treated supernatant, respectively. With the increased levels of LDH release, we confirmed that the necrosis of MDA-MB-231 cells was induced by C4.



271

Fig. 5. a) LDH release from MDA-MB-231 cells treated with different concentrations of C4 for 48 h. b) MDA-MB-231 cells treated with various concentrations of C4 for 48 h, and the necrotic cells were counted by Trypan blue exclusion assay. The values are presented as mean  $\pm$  S.D. of three independent experiments. \**P* < 0.05, \*\**P* < 0.01.

276 2.2.5 Trypan blue exclusion assay

The Trypan blue dye exclusion test was used to determine the number of viable cells present in a cell suspension. It is based on the principle that live cells will exclude membrane-impermeable dyes such as trypan blue, whereas trypan blue can penetrate inside necrotic cells and stain them [39,40]. In the present study, after independent treatment with **C4** at concentrations of 0.23, 0.46, 0.91, and 1.82  $\mu$ M for 48 h, the MDA-MB-231 cells were trypsinized and stained with 0.2% Trypan blue

solution at 1:1 dilution for 5 min, and, thereafter, loaded into a hemocytometer for separate counting of stained (necrotic) and unstained (viable) cells. As shown in Fig. 5b, complex C4 dramatically induced necrosis in a dose-dependent manner, and almost 50% of MDA-MB-231 cells was identified with necrosis after C4 (1.82  $\mu$ M) incubation of 48 h, which is in a agreement with the results presented in Fig. 4 and Fig. 5a. With such notation, the necrosis induced by antimonial C4 has been repeatedly proven.

### 290 2.2.6 Reactive oxygen species (ROS) assay

Reactive oxygen species (ROS) are radicals, ions or molecules that have a single 291 unpaired electron in their outermost shell of electrons. Recent evidence has shown 292 that ROS play a key role as a messenger for signal transduction and cell cycling in 293 294 normal cells. However, unnecessary increases of ROS could result in cell death [41]. 295 Previous works revealed that metal-based chemotherapeutic agents (e.g. cisplatin) induced apoptosis and/or necrosis through ROS generation in several cancer cell lines 296 297 [34,42]. It is widely accepted that the anticancer effect of these chemotherapeutics is due to the induction of oxidative stress and ROS-mediated cell injury in cancer. To 298 explore whether cell necrosis induced by C4 was dependent on the level of ROS, we 299 300 assessed the production of intracellular ROS in C4-treated MDA-MB-231 cells by using 5-(and-6)-carboxy-2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA) as 301 fluorescence probe and H<sub>2</sub>O<sub>2</sub> as a positive control. Intracellular ROS levels were 302 303 measured and quantified by change in relative fluorescence units (RFU). As shown in

304 Fig. 6a, compared to the negative untreated control, treatment with C4 at graded concentrations induced a significant elevation of ROS production in a dose-dependent 305 306 manner. When the exposure was 1.82 µM for 48 h, C4 promoted an increase of 4.61 fold (P < 0.01) of ROS production compared to that of untreated control. The 307 excessive increase of ROS production in cells might be attributed to the cytotoxic 308 309 activity of compound C4 through the activation of mitochondria-initiated events. Furthermore, we examined the effect of N-acetylcysteine (NAC), a well-known 310 antioxidant, on the necrotic activity of C4-treated MDA-MB-231 cells by means of 311 Trypan blue dye exclusion test, as shown in Fig 6b. After pretreatment with NAC (5 312 µM) for 2 h, the population of necrotic MDA-MB-231 cells induced by C4 was 313 significantly decreased from 30.16% to 19.79%, evidencing the retarding effect of 314 315 NAC. It is consistent with the notion that the C4-driven induction of necrosis in MDA-MB-231 cells is dependent on the production of intracellular ROS. 316



317

**Fig. 6.** a) ROS generation in C4 treated MDA-MB-231 cells. Relative fluorescence units (RFU) of DCF was measured using a spectrofluorometer with excitation at 485 nm and emission at 530 nm. b) Cells were pretreated with NAC ( $5 \mu$ M) for 2 h, followed by treatment

- 321 with an IC<sub>50</sub> concentration of C4 (0.91  $\mu$ M) for 48 h before determination of cell death by
- 322 Trypan blue dye exclusion assay. The results are presented as mean  $\pm$  S.D. (\*P < 0.05, \*\*P <
- **323** 0.01).

# **3. Conclusion**

325	In the present study, a series of organoantimony(III) halide complexes with
326	azastibocine framework were synthesized, characterized and evaluated for in vitro
327	cytotoxic activity on human liver hepatocellular carcinoma cell line (HepG2), human
328	breast cancer cell line (MDA-MB-231), human breast adenocarcinoma cell line
329	(MCF-7), human cervical carcinoma cell line (HeLa) and human embryonic kidney
330	cell line (HEK-293). The results reveal a positive correlation between cytotoxic
331	activity and the N $\rightarrow$ Sb coordinate bond lengths in the complexes with same nitrogen
332	substituent. Among all tested trivalent organoantimony complexes, the complex
333	6-cyclohexyl-12-fluoro-5,6,7,12-tetrahydrodibenzo[ $c,f$ ][ $1,5$ ]azastibocine (C4)
334	exhibited the highest selectivity index. Further mechanistic investigation indicated
335	that C4 causes cell cycle arrest mainly at the S phase, and consequently results in
336	necrosis of MDA-MB-231 cells through the production of intracellular reactive
337	oxygen species. The results indicate that the azastibocine-framework organoantimony
338	halide complexes could be promising candidates for the development of new drugs in
339	cancer therapy. Specifically, the elucidation of correlation between cytotoxicity and
340	intermolecular interaction has provided theoretical and experimental basis for
341	in-depth design of antimony-based antineoplastic metallodrugs.

# 342 **4. Experimental**

#### 343 *4.1. Chemistry*

355

The commercially available starting materials were purchased from Adamas-beta, 344 and were used as received unless otherwise noted. The preparation of N-containing 345 precursors (P1-P3) and organoantimony(III) chlorides (C1, C4 and C9) were 346 347 prepared according to literature procedures [30,43]. Melting points were determined over a XT-4 micro melting point apparatus (Beijing Tech Instrument Co., Ltd.). 348 349 Nuclear magnetic resonance (NMR) data were obtained on a Bruker-400 spectrometer (400 MHz for <sup>1</sup>H, 100 MHz for <sup>13</sup>C and 376 MHz for <sup>19</sup>F spectroscopy). Chemicals 350 shifts are reported in ppm ( $\delta$ ) relative to internal tetramethylsilane (TMS). In this 351 paper, data are reported as follows: Chemical shift, multiplicity (s = singlet, d =352 doublet, t = triplet, q = quartet, m = multiplet). Coupling constants (J) are reported in 353 hertz. Elemental analyses were performed using a VARIO EL III instrument. 354



**Scheme 1.** Synthesis of organoantimony(III) halide complexes C1–C12. (a) <sup>*n*</sup>BuLi (2.0 equiv.), Et<sub>2</sub>O, -60  $\Box$  to room temperature. (b) SbCl<sub>3</sub> (1.1 equiv.), Et<sub>2</sub>O, -78  $\Box$  to room temperature; C1, 72%; C5, 78%; C9, 80%. (c) KBr or KI (10.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O, room temperature; C2, 92%; C3, 86%; C6, 87%; C7, 85%; C10, 91%; C11, 81%. (d) AgF (1.0 equiv.), CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O, room temperature, dark; C4, 93%; C8, 95%; C12, 76%.

### 361 4.1.1. General procedure for the synthesis of organoantimony(III) chlorides

362	To a solution of N-containing precursors (25 mmol) in anhydrous ether (100 mL),
363	n-BuLi in hexane (2.5 M, 20 mL) was added dropwise under nitrogen with vigorous
364	stirring at -30 °C, and the mixture thus obtained was warmed to room temperature for
365	3 h. Then a solution of SbCl <sub>3</sub> (5.8 g, 25.5 mmol) in anhydrous ether (60 mL) was
366	added slowly at -78 °C within 30 min. After subject to stirring at room temperature
367	overnight, the solution was subject to evaporation under reduced pressure. The
368	residue was extracted with dichloromethane (100 mL) and washed with a solution of
369	NH <sub>4</sub> Cl (1M) in H <sub>2</sub> O. The organic layer was washed with deionized water (3 $\times$ 50
370	mL), dried over anhydrous Na <sub>2</sub> SO <sub>4</sub> , subject to filtration, and concentrated in vacuo.
371	The residue was purified by recrystallization in CH <sub>2</sub> Cl <sub>2</sub> /n-hexane mixture to afford
372	the corresponding organoantimony chloride as colorless crystals.

373 4.1.2. General procedure for the synthesis of organoantimony(III) bromides

To a solution of organoantimony chloride (5.0 mmol) in  $CH_2Cl_2$  (30 mL), a solution of KBr (6.0 g, 50 mmol) in deionized water (25 mL) was added under open-flask conditions. After being stirred at room temperature overnight, the solution was extracted with  $CH_2Cl_2$  (3 × 30 mL). The combined organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and subject to filtration. The filtrate was mixed with *n*-hexane and kept at room temperature for 24 h to afford the corresponding organoantimony bromide as colorless crystals.

381 4.1.3. General procedure for the synthesis of organoantimony(III) iodides

To a solution of organoantimony chloride (5.0 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL), a solution of KI (8.3 g, 50 mmol) in deionized water (25 mL) was added under open-flask

384 conditions. After subject to stirring at room temperature overnight, the solution was 385 extracted with  $CH_2Cl_2$  (3 × 30 mL). The combined organic layer was dried over 386 anhydrous  $Na_2SO_4$ , and subject to filtration. The filtrate was mixed with *n*-hexane and 387 kept at room temperature for 24 h to afford the corresponding organoantimony iodide 388 as colorless crystals.

389 4.1.4. General procedure for the synthesis of organoantimony(III) fluorides

To a solution of organoantimony chloride (5.0 mmol) in  $CH_2Cl_2$  (30 mL), a solution of AgF (635 mg, 5.0 mmol) in deionized water (25 mL) was added under open-flask conditions. After being stirred in the dark at room temperature for 4 h, the mixture was subject to filtration. The filtrate was mixed with *n*-hexane and kept at room temperature for 24 h to afford the corresponding organoantimony fluoride as colorless crystals.

### 396 4.1.5. Analytical data for the synthesized organoantimony halide complexes

12-chloro-6-cyclohexyl-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (**C1**). 397 Yield: 72% (7.8 g). Melting point: 254–256 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS):  $\delta$ 398 8.30 (2H, d, J = 7.4 Hz), 7.39–7.27 (4H, m), 7.14 (2H, d, J = 7.4 Hz), 4.15 (4H, dd, J 399 = 61.9, 15.1 Hz), 3.09 (1H, t, J = 11.3 Hz), 2.05 (2H, d, J = 11.7 Hz), 1.86 (2H, d, J = 400 12.5 Hz), 1.69 (d, J = 13.0 Hz), 1.46–1.27 (m, 4H), 1.18–1.08 (m, 1H); <sup>13</sup>C NMR 401 (100 MHz,CDCl<sub>3</sub>, TMS): δ 144.0, 140.1, 134.9, 128.8, 128.7, 124.7, 65.4, 57.8, 29.5, 402 25.6, 25.4. Anal. Calc. for C<sub>20</sub>H<sub>23</sub>ClNSb: C, 55.27; H, 5.33; N, 3.22. Found: C, 55.39; 403 H, 5.25; N, 3.14. 404

405 *12-bromo-6-cyclohexyl-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine* (C2).

406	Yield: 92% (2.2 g). Melting point: 245–247 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> , TMS): $\delta$
407	8.40–8.38 (2H, m), 7.37–7.27 (4H, m), 7.13 (2H, d, J = 6.6 Hz), 4.15 (4H, dd, J =
408	58.0, 15.1 Hz), 3.13–3.08 (1H, m), 2.04 (2H, d, J = 10.9 Hz), 1.85 (2H, d, J = 11.8
409	Hz), 1.68 (1H, d, J = 11.8 Hz), 1.45–1.27 (4H, m), 1.17–1.09 (1H, m); <sup>13</sup> C NMR (100
410	MHz,CDCl <sub>3</sub> , TMS): δ 144.0, 138.0, 136.3, 128.9, 128.7, 124.7, 65.5, 57.8, 29.6, 25.6,
411	25.4. Anal. Calc. for C <sub>20</sub> H <sub>23</sub> BrNSb: C, 50.14; H, 4.84; N, 2.92. Found: C, 50.29; H,
412	4.95; N, 2.99.
413	6-cyclohexyl-12-iodo-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C3). Yield:
414	86% (2.3 g). Melting point: 253–255 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> , TMS): $\delta$ 8.52
415	(2H, d, J = 7.0 Hz), 7.39–7.30 (4H, m), 7.12 (2H, d, J = 6.9 Hz), 4.14 (4H, dd, J =
416	52.4, 15.0 Hz), 3.12 (1H, t, J = 11.4 Hz), 2.07 (2H, d, J = 11.2 Hz), 1.88 (2H, d, J =
417	12.4 Hz), 1.71 (1H, d, <i>J</i> = 13.2 Hz), 1.47–1.31 (4H, m), 1.19–1.13 (1H, m); <sup>13</sup> C NMR
418	(100 MHz,CDCl <sub>3</sub> , TMS): $\delta$ 143.8, 139.4, 134.5, 129.2, 129.0, 124.7, 65.6, 57.6, 29.7,
419	25.7, 25.4. Anal. Calc. for C <sub>20</sub> H <sub>23</sub> INSb: C, 45.66; H, 4.41; N, 2.66. Found: C, 45.74;
420	H, 4.54; N, 2.79.
421	6-cyclohexyl-12-fluoro-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C4).

422 Yield: 93% (1.9 g). Melting point: 237–239 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>,

423 TMS):  $\delta$  7.97 (2H, d, J = 7.3 Hz), 7.40–7.25 (4H, m), 7.14 (2H, t, J = 7.4 Hz),

424 4.09 (4H, dd, *J* = 71.4, 15.2 Hz), 3.03 (1H, t, *J* = 10.8 Hz), 2.00 (2H, d, *J* = 11.4

425 Hz), 1.86 (2H, d, J = 12.5 Hz), 1.69 (2H, d, J = 12.7 Hz), 1.45–1.26 (4H, m),

426 1.18–1.09 (1H, m); <sup>13</sup>C NMR (100 MHz,CDCl<sub>3</sub>, TMS):  $\delta$  144.1, 144.0 (d, J =

427 6.4 Hz), 133.4 (d, J = 6.0 Hz), 128.5, 128.3, 124.7, 65.2, 57.7 (d, J = 2.1 Hz),

428	29.5, 25.7, 25.5; <sup>19</sup> F NMR (376 MHz, CDCl <sub>3</sub> ): $\delta$ -185.45. Anal. Calc. for
429	C <sub>20</sub> H <sub>23</sub> FNSb: C, 57.45; H, 5.54; N, 3.35. Found: C, 57.53; H, 5.59; N, 3.44. FT-IR
430	(KBr, cm <sup>-1</sup> ): v 2935, 2857, 1455, 1440, 1268, 1204, 1100, 975, 950, 934, 896, 758.
431	12-chloro-6-phenyl-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C5). Yield:
432	76% (8.1 g). Melting point: 222–224 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> , TMS): $\delta$ 8.40–
433	8.28 (2H, m), 7.47–7.42 (2H, m), 7.38–7.34 (4H, m), 7.30–7.19 (5H, m), 4.66 (4H, dd,
434	$J = 70.8$ , 14.9 Hz); <sup>13</sup> C NMR (100 MHz,CDCl <sub>3</sub> , TMS): $\delta$ 147.9, 143.1, 140.5, 135.2,
435	129.5, 129.2, 129.2, 125.4, 125.3, 119.7, 61.2. Anal. Calc. for C <sub>20</sub> H <sub>17</sub> ClNSb: C, 56.05;
436	H, 4.00; N, 3.27. Found: C, 55.73; H, 4.12; N, 3.35.
437	12-bromo-6-phenyl-5,6,7,12-tetrahydrodibenzo $[c,f]$ [1,5]azastibocine (C6). Yield:
438	87% (2.0 g). Melting point: 233–235 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> , TMS): $\delta$ 8.41–
439	8.29 (2H, m), 7.47–7.43 (2H, m), 7.40–7.35 (4H, m), 7.31–7.29 (2H, m), 7.25–7.21
440	(3H, m), 4.67 (4H, dd, $J = 65.7$ ,14.9 Hz); <sup>13</sup> C NMR (100 MHz,CDCl <sub>3</sub> , TMS) $\delta$ 147.8,
441	143.0, 138.2, 136.8, 129.6, 129.4, 129.3, 125.5, 125.3, 119.8, 61.2. Anal. Calc. for
442	C <sub>20</sub> H <sub>17</sub> BrNSb: C, 50.78; H, 3.62; N, 2.96. Found: C, 50.89; H, 3.69; N, 3.07.
443	12-iodo-6-phenyl-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C7). Yield:
444	85% (2.2 g). Melting point: 249–251 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> , TMS): $\delta$
445	8.47-8.45 (2H, m), 7.39-7.30 (6H, m), 7.26-7.23 (2H, m), 7.19-7.16 (3H, m),
446	4.60 (4H, dd, $J = 66.6$ , 14.9 Hz); <sup>13</sup> C NMR (100 MHz,CDCl <sub>3</sub> , TMS): $\delta$ 147.6,
447	142.9, 140.0, 134.1, 129.6, 129.5, 129.4, 125.5, 125.3, 119.8, 60.9. Anal. Calc.
448	for C <sub>20</sub> H <sub>17</sub> INSb: C, 46.19; H, 3.30; N, 2.69. Found: C, 46.27; H, 3.33; N, 2.76.

27

- 449 12-fluoro-6-phenyl-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C8). Yield: 95% (2.0 g). Melting point: 216–218 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS):  $\delta$ 450 451 8.00 (2H, d, J = 7.3 Hz), 7.46 (2H, t, J = 7.3 Hz), 7.35–7.34 (4H, m), 7.20–7.25 (4H,m), 7.20 (1H, t, J = 6.8 Hz), 4.63 (4H, dd, J = 85.1, 15.0 Hz); <sup>13</sup>C NMR 452 (100 MHz,CDCl<sub>3</sub>, TMS):  $\delta$  148.5, 144.7 (d, J = 6.6 Hz), 143.1, 133.4 (d, J =453 6.2 Hz), 129.5, 128.9, 128.8, 125.2, 125.0, 119.4, 61.2 (d, J = 1.8 Hz); <sup>19</sup>F 454 NMR (376 MHz, CDCl<sub>3</sub>): δ -198.97. Anal. Calc. for C<sub>20</sub>H<sub>17</sub>FNSb: C, 58.29; H, 455 4.16; N, 3.40. Found: C, 58.39; H, 4.24; N, 3.58. 456
- 457 *6-(tert-butyl)-12-chloro-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine* (**C9**).
- 458 Yield: 80% (8.1 g). Melting point: 213–215 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS):  $\delta$
- 459 8.32 (2H, d, *J* = 7.3 Hz), 7.37 (2H, t, *J* = 7.2 Hz), 7.31–7.28 (2H, m), 7.15 (2H, d, *J* =
- 460 7.5 Hz), 4.23 (4H, dd, *J*= 161.2, 15.4 Hz), 1.38 (9H, s); <sup>13</sup>C NMR (100 MHz,CDCl<sub>3</sub>,
- 461 TMS):  $\delta$  145.1, 139.9, 134.9, 128.9, 128.4, 124.5, 60.4, 57.2, 27.0. Anal. Calc. for
- 462 C<sub>18</sub>H<sub>21</sub>ClNSb: C, 52.91; H, 5.18; N, 3.43. Found: C, 52.78; H, 5.26; N, 3.52.
- 463 *12-bromo-6-(tert-butyl)-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine* (C10).
- 464 Yield: 91% (2.0 g). Melting point: 244–246 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS):
- 465  $\delta$  8.41 (2H, d, J= 7.2 Hz), 7.36–7.27 (4H, m), 7.15 (2H, d, J= 6.8 Hz), 4.22 (4H, dd,
- 466 J= 152.9, 15.4 Hz, 1.37 (9H, s); <sup>13</sup>C NMR (100 MHz,CDCl<sub>3</sub>, TMS):  $\delta$  145.0, 137.8,
- 467 136.4, 129.0, 128.5, 124.5, 60.6, 57.2, 27.1. Anal. Calc. for C<sub>18</sub>H<sub>21</sub>BrNSb: C, 47.72; H,
- 468 4.67; N, 3.09. Found: C, 47.82; H, 4.75; N, 3.16.
- $469 \qquad 6-(tert-butyl)-12-iodo-5, 6, 7, 12-tetrahydrodibenzo[c, f][1,5]azastibocine \qquad (C11).$
- 470 Yield: 81% (2.0 g). Melting point: 228–230 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>,

4/1	TMS): $\delta$ 8.54–8.51 (2H, m), 7.34–7.30 (4H, m), 7.12–7.10 (2H, m), 4.20 (4H,
472	dd, J= 151.4, 15.3 Hz), 1.38 (9H, s); $^{13}$ C NMR (100 MHz,CDCl <sub>3</sub> , TMS): $\delta$
473	144.9, 139.6, 134.3, 129.2, 128.8, 124.5, 60.8, 57.1, 27.2. Anal. Calc. for
474	C <sub>18</sub> H <sub>21</sub> INSb: C, 43.24; H, 4.23; N, 2.80. Found: C, 43.33; H, 4.35; N, 2.96.
475	6-(tert-butyl)-12-fluoro-5,6,7,12-tetrahydrodibenzo[c,f][1,5]azastibocine (C12).
476	Yield: 76% (1.5 g). Melting point: 206–208 °C. <sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> ,
477	TMS): δ 7.80 (2H, d, J = 7.3 Hz), 7.16 (2H, t, J = 7.3 Hz), 7.07 (2H, t, J = 7.3
478	Hz), 6.96 (2H, d, J = 7.7 Hz), 3.95 (4H, dd, J= 171.1, 15.4 Hz), 1.13 (9H, s);
479	<sup>13</sup> C NMR (100 MHz,CDCl <sub>3</sub> , TMS): $\delta$ 145.1, 143.8 (d, $J$ = 5.8 Hz,), 133.2 (d, $J$
480	= 6.5 Hz), 128.5, 128.0, 124.5, 59.7, 57.1, 26.9; <sup>19</sup> F NMR (376 MHz, CDCl <sub>3</sub> ): $\delta$
481	-185.16. Anal. Calc. for C <sub>18</sub> H <sub>21</sub> FNSb: C, 55.13; H, 5.40; N, 3.57. Found: C, 52.25; H,
482	5.46; N, 3.62.

483 4.1.6. X-ray crystal structural determination of organoantimony(III) halide complexes
484 C1–C12

X-ray crystal structural determination of complexes C1, C5, and C9 have described 485 486 previously [30]. X-ray single crystal diffraction analysis of these complexes was performed at Hunan University using a Bruker SMART APEX diffractometer. The 487 CDCC number is 1864560 (C2), 1859257 (C3), 1859365 (C4), 945750 (C6), 945752 488 489 (C7), 769426 (C8), 1861311 (C10), 1859366 (C11), 1864561 (C12), respectively. In all cases, the diffraction data were collected using graphite monochromated Mo-Ka 490 radiation ( $\lambda = 0.71073$  Å). The collected frames were processed with SAINT+ 491 492 software and the collected reflections were subject to absorption correction

493 (SADABS) [44]. The structure was solved by the Direct method (SHELXTL) in 494 conjunction with standard difference Fourier techniques and subsequently refined by 495 full-matrix least-squares analyses on  $F^2$  [45]. The hydrogen atoms were generated in 496 their idealized positions and all non-hydrogen atoms were refined anisotropically (see 497 Table 2).

- 498 *4.2. Biological evaluation*
- 499 *4.2.1. Cell lines and cell culture*

500 Human cancerous cell lines: HepG2 (human liver hepatocellular carcinoma cell line), MDA-MB-231 (human breast cancer cell line), MCF-7 (human breast 501 adenocarcinoma cell line), HeLa (human cervical carcinoma cell line) and 502 nonmalignant cell lines: HEK-293 (human embryonic kidney cell line) used in this 503 504 study were supplied by the Cancer Research Institute, Central South University (Changsha, Hunan, PR China). The cells were cultivated in Dulbecco's modified 505 Eagle's medium (DMEM, Gibco) or RPMI (HyClone), supplemented with 10% fetal 506 507 bovine serum containing L-glutamine and 1% penicillin-streptomycin (HyClone) and maintained in a humidified atmosphere of 95% air, 5% CO<sub>2</sub> at 37°C. 508

509 4.2.2. Cell viability

510 The cell viability of the organometallic complexes in cells was determined by the 511 CCK-8 assay (Dojindo Molecular Technologies, Shanghai, China) using a modified 512 method as previously described [46]. Each of the tested organoantimony complexes 513 was completely dissolved in DMSO to a solution of 10 mM. Briefly,  $5 \times 10^3$  cells

were seeded in each well of the 96-well plates with fresh medium. After complete adhesion, the cells were continuously exposed to test complexes of 0.01, 0.3, 1, 3 and  $10 \mu$ M for 24 h. A CCK-8 solution ( $10 \mu$ L) was added and the plates were incubated at 37 °C for a further 2 h. Subsequently, the absorbance of formazan yellow formed in the cells was measured at 450 nm, using an iMark microplate absorbance reader (Bio-Rad, USA). The IC<sub>50</sub> values were determined by the four-parameter logistic method. Mean IC<sub>50</sub> values were obtained from at least three independent experiments.

521 *4.2.3. Cell cycle analysis* 

Complex C4 on the cell cycle perturbation in MDA-MB-231 cells was examined 522 by flow cytometry analysis. Briefly,  $1.0 \times 10^6$  per well were seeded in a six-well 523 plate. After treated with  $0.25 \times IC_{50}$ ,  $0.5 \times IC_{50}$ ,  $1 \times IC_{50}$  and  $2 \times IC_{50}$  of C4 for 24 h, 524 525 the supernatants were removed and cells were washed with PBS. The cells were subsequently harvested by trypsinization, then fixed and stained with PI (Cat. no. 526 537059, Calbiochem, San Diego, CA, USA). This was followed by the measurement 527 of propidium-iodide-mediated fluorescence with flow cytometry (FACSCalibur BD, 528 Bedford, MA, USA) by using excitation of DNA-bound PI at 488 nm, with emission 529 at 585 nm. The cell cycle distribution is shown as the percentage of cells containing 530  $G_0/G_1$ , S and  $G_2/M$  DNA as identified by PI staining. 531

532 4.2.4. Apoptosis Analysis

533 For apoptosis assay, cells were stained with Annexin V-FITC and PI Kit (Cat. no.

- 534 LOT9104010102, Dinguo Bio. Inc., Beijing, China) as previously described [30].
- 535 Briefly, MDA-MB-231 cells were incubated with complexes as indicated in the

536 figures for 48 h. At the end of incubation, the cells were harvested and washed twice in PBS, then re-suspended in 500  $\mu$ L of binding buffer and incubated with 5  $\mu$ L of 537 538 annexin V-FITC and 5 µL of PI for 15 min at room temperature in the dark. Flow cytometry was performed on a FACS CaliburTM flow cytometer and the collected 539 data were analyzed using FlowJo software (Becton, Dickinson & Company). The 540 541 results were interpreted as follows: cells in the lower left quadrant (annexin-V<sup>-</sup>/PI<sup>-</sup>) were considered as living cells, in the lower right quadrant (annexin- $V^+/PI^-$ ) as early 542 apoptotic cells, in the upper right quadrant (annexin- $V^+/PI^+$ ) as late apoptotic cells, 543 and in the upper left quadrant (annexin- $V/PI^+$ ) as necrotic cells. The total apoptotic 544 rate was the rate of cells in the lower right quadrant (annexin- $V^+/PI^-$ ) plus that in the 545 upper right quadrant (annexin- $V^+/PI^+$ ). 546

547 4.2.5. Lactate dehydrogenase (LDH) release assay

The LDH activity of treated MDA-MB-231 cells was monitored according to the 548 manufacturer's instructions (Cat. no. C0016, Beyotime, Haimen, China). Briefly, 549 treated cells with diverse concentrations of complex C4 were incubated for 48 hours. 550 Next, the medium was centrifuged at 2000 rpm for 5 min to obtain the supernatant. 551 The supernatant was transferred to a new 96-well plate. Then 100 µl of the LDH 552 reaction was added to each well and was incubated for 30 min at room temperature 553 before absorbance measurement using an iMark microplate absorbance reader 554 (Bio-Rad, USA) at 490 nm. 555

556 4.2.6. Trypan blue dye exclusion assay

557 MDA-MB-231 cells (  $5 \times 10^4$  ) were seeded in 12-well plates and exposed to

complex C4 at the indicated concentrations as illustrated in the figures for 48 h. The
cells were trypsinized and stained with 0.2% Trypan blue solution (Cat. no. SBJ-0245,
Beyotime, China) at 1:1 dilution for 5 min, and were then loaded into a
hemocytometer for separate counting of stained (necrotic) and unstained (viable)
cells.

563 4.2.7. Reactive oxygen species (ROS) assay

The production of intracellular ROS in C4-treated MDA-MB-231 cells was 564 assessed using 2',7'dichlorofluorescein diacetate (DCFH-DA) assay. The assay used 565 the cell-permeable fluorogenic probe DCFH-DA, which upon diffusion into cells is 566 oxidized by cellular ROS to form highly fluorescent 2',7'-dichlorodihydrofluorescein 567 (DCF).  $H_2O_2$  was used as a positive control. The effects of C4 on intracellular ROS 568 production can be measured in terms of relative fluorescence units (RFU). Briefly, 569 MDA-MB-231 cells were seeded in 6-well microplates, treated with complex C4 as 570 indicated in the figures for 48 h. After 48 h of C4 treatment, the cells were washed 571 twice with PBS and then incubated with the DCFH-DA probe (10 µM) at 37 °C for 30 572 min, and then immediately washed three times with PBS. Fluorescence was measured 573 with a BioTek ELx800 microplate reader at an excitation wavelength of 485 nm and 574 an emission wavelength of 528 nm. 575

576 *4.2.8. Statistical analysis* 

577 Data were obtained from at least three separate experiments and the results were 578 expressed as mean ± S.D. The data were analyzed for statistical significance by 579 one-way ANOVA using GraphPad Prism version 7.0.5 for Windows, GraphPad

- 580 Software, San Diego, CA, USA., and p < 0.05 was considered statistically significant
- 581 (notation:  ${}^{*}P < 0.05$ ,  ${}^{**}P < 0.01$ ).

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Table 1

Selected bond lengths (Å) and angles (deg) of organoantimony(III) halide complexes C1-C12.											
C1		C2		C3		C4					
Sb(1)–C(1)	2.144(4)	Sb(1)–C(6)	2.143(2)	Sb(1)–C(6)	2.154(3)	Sb(1)–C(6)	2.140(3)				
Sb(1)–C(8)	2.134(4)	Sb(1)-C(10)	2.157(2)	Sb(1)–C(7)	2.166(3)	Sb(1)–C(7)	2.145(3)				
C(1)–Sb(1)–C(8)	98.2(1)	C(6)-Sb(1)-C(10)	99.23(8)	C(6)-Sb(1)-C(7)	98.8(1)	C(6)–Sb(1)–C(7)	98.5(1)				
Cl(1)-Sb(1)-N(1)	162.92(7)	Br(1)-Sb(1)-N(1)	163.53(4)	I(1)-Sb(1)-N(1)	163.34(6)	F(1)-Sb(1)-N(1)	156.42(7)				
Sb(1)–N(1)	2.397(2)	Sb(1)–N(1)	2.387(2)	Sb(1)–N(1)	2.400(2)	Sb(1)–N(1)	2.450(2)				
Sb(1)-Cl(1)	2.5572(9)	Sb(1)–Br(1)	2.7142(5)	Sb(1)–I(1)	2.9650(4)	Sb(1)–F(1)	2.015(2)				
C5		C6		C7		C8					
Sb(1)–C(1)	2.150(2)	Sb(1)–C(1)	2.158(3)	Sb(1)–C(1)	2.166(3)	Sb(1)–C(1)	2.131(3)				
Sb(1)–C(8)	2.155(2)	Sb(1)–C(8)	2.156(3)	Sb(1)-C(8)	2.175(3)	Sb(1)–C(8)	2.153(3)				
C(1)–Sb(1)–C(8)	100.53(7)	C(1)-Sb(1)-C(8)	100.4(1)	C(1)–Sb(1)–C(8)	96.3(1)	C(1)-Sb(1)-C(8)	91.80(9)				
Cl(1)-Sb(1)-N(1)	161.98(4)	Br(1)-Sb(1)-N(1)	163.22(6)	I(1)-Sb(1)-N(1)	163.92(6)	F(1)-Sb(1)-N(1)	162.92(7)				
Sb(1)–N(1)	2.466(2)	Sb(1)–N(1)	2.469(2)	Sb(1)–N(1)	2.498(3)	Sb(1)-N(1)	2.522(2)				
Sb(1)–Cl(1)	2.5173(5)	Sb(1)–Br(1)	2.6620(4)	Sb(1)–I(1)	2.8995(3)	Sb(1)–F(1)	1.998(2)				
С9		C10		C11		C12					
Sb(1)–C(1)	2.147(2)	Sb(1)–C(6)	2.153(2)	Sb(1)–C(6)	2.166(3)	Sb(1)–C(6)	2.127(7)				
Sb(1)–C(14)	2.155(3)	Sb(1)–C(7)	2.151(2)	Sb(1)–C(7)	2.157(3)	Sb(1)-C(10)	2.135(7)				
C(1)–Sb(1)–C(14)	95.60(9)	C(6)–Sb(1)–C(7)	96.18(8)	C(6)–Sb(1)–C(7)	97.6(1)	C(6)–Sb(1)–C(10)	98.2(3)				
Cl(1)-Sb(1)-N(1)	161.60(5)	Br(1)-Sb(1)-N(1)	162.99(4)	I(1)-Sb(1)-N(1)	163.59(6)	F(1)-Sb(1)-N(1)	155.9(2)				
Sb(1)–N(1)	2.467(2)	Sb(1)–N(1)	2.446(2)	Sb(1)-N(1)	2.462(3)	Sb(1)–N(1)	2.495(5)				
Sb(1)-Cl(1)	2.5579(7)	Sb(1)-Br(1)	2.7631(5)	Sb(1)–I(1)	2.9463(3)	Sb(1)–F(1)	2.026(6)				

2.467(2, 2.5579(7) Sb(1)-Br(1,

Table	2
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Crystallographic data for organoantimony(III) halide complexes C1-C12.

Compound	C1	C2	C3	C4	C5	C6
Empirical formula	C20H23CINSb	C20H23BrNSb	C <sub>20</sub> H <sub>23</sub> INSb	C <sub>20</sub> H <sub>23</sub> FNSb	C <sub>20</sub> H <sub>17</sub> ClNSb	C <sub>20</sub> H <sub>17</sub> BrNSb
Formula weight	434.59	479.05	526.04	418.14	428.55	473.01
Temperature/K	293	273	100	100	273	298
Crystal system	Monoclinic	monoclinic	monoclinic	monoclinic	Monoclinic	monoclinic
Space group	$P_2(1)/c$	P2(1)/n	P2(1)/n	P2(1)/n	$P_2(1)/n$	P2(1)/n
a/Å	10.0771(7)	10.2338(5)	11.3664(3)	9.3503(3)	9.3782(4)	9.5744(3)
b/Å	16.2881(12)	16.5885(7)	12.5996(3)	15.9263(5)	10.1833(4)	10.2962(3)
$c/\AA$	12.2040(9)	12.0033(5)	13.0792(4)	11.7239(4)	18.1033(8)	18.1939(6)
α/°	90.00	90.00	90.00	90.00	90.00	90.00
β/°	111.812(10)	113.346(2)	94.703(2)	104.044(3)	102.895(10)	100.795(1)
$\gamma/^{\circ}$	90.00	90.00	90.00	90.00	90.00	90.00
$V/\text{\AA}^3$	1859.7(2)	1870.89(15)	1866.79(9)	1693.69(10)	1685.3(12)	1761.81(10)
Ζ	4	4	4	4	4	4
No. of reflections collected	10058	30234	12759	10827	9923	10613
No. of unique reflections	3644	4636	3284	2982	4032	3417
R <sub>int</sub>	0.047	0.0363	0.0390	0.0376	0.015	0.038
$R_{I}$ (reflections)	0.0324	0.0196	0.0236	0.0256	0.0203	0.0294
$wR_2(reflections)$	0.0917	0.0416	0.0561	0.0598	0.0560	0.0709
GOF on $F^2$	1.048	1.066	1.022	1.097	1.077	1.041
Compound	C7	C8	С9	C10	C11	C12
Compound Empirical formula	C7 C <sub>20</sub> H <sub>17</sub> INSb	C8 C <sub>20</sub> H <sub>17</sub> FNSb	C9 C <sub>18</sub> H <sub>21</sub> ClNSb	C10 C <sub>18</sub> H <sub>21</sub> BrNSb	C11 C <sub>18</sub> H <sub>21</sub> INSb	C12 C <sub>18</sub> H <sub>21</sub> FNSb
Compound Empirical formula Formula weight	<b>C7</b> C <sub>20</sub> H <sub>17</sub> INSb 520.00	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11
Compound Empirical formula Formula weight Temperature/K	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic	<b>C11</b> C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i>	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i>
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8)	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9)
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i> <i>b/Å</i>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)`	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6)
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i> <i>b/Å</i> <i>c/Å</i>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11)	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i> 9.7692(12) 15.933(2) 11.4491(14)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10)
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i> <i>b/Å</i> <i>c/Å</i> <i>a/°</i>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group a/Å b/Å c/Å a/° β/°	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1)	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2)
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group a/Å b/Å c/Å α/° β/° γ/°	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8) <sup>°</sup> 15.7161(11) 90.00 101.559 (1) 90.00	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i> <i>b/Å</i> <i>c/Å</i> <i>a/°</i> β/° γ/° V/Å <sup>3</sup>	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11)	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2)	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic <i>P2</i> (1)/ <i>n</i> 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4)	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11)	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10)	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3)
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group a/Å b/Å c/Å α/° β/° γ/° V/Å <sup>3</sup> Z	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8) <sup>°</sup> 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8
CompoundEmpirical formulaFormula weightTemperature/KCrystal systemSpace group $a/Å$ $b/Å$ $c/Å$ $a/°$ $\beta/°$ $\gamma/°$ $V/Å^3$ ZNo. of reflections collected	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4 9422	<b>C9</b> C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4 9317	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989
Compound Empirical formula Formula weight <i>Temperature/K</i> Crystal system Space group <i>a/Å</i> <i>b/Å</i> <i>c/Å</i> <i>α/°</i> β/° γ/° V/Å <sup>3</sup> <i>Z</i> No. of reflections collected No. of unique reflections	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4 9422 3560	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4 9317 3889	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232
Compound         Empirical formula         Formula weight         Temperature/K         Crystal system         Space group $a/Å$ $b/Å$ $c/Å$ $b/Å$ $c/Å$ $\beta/°$ $\gamma/°$ $V/Å^3$ Z         No. of reflections collected         No. of unique reflections $R_{int}$	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510 0.055	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4 9422 3560 0.063	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4 9317 3889 0.027	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978 0.0437	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058 0.0353	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232 0.0229
Compound         Empirical formula         Formula weight         Temperature/K         Crystal system         Space group $a/Å$ $b/Å$ $c/Å$ $a/°$ $\beta/°$ $\gamma/°$ $V/Å^3$ Z         No. of reflections collected         No. of unique reflections $R_{int}$ $R_1(reflections)$	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510 0.055 0.0354	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8)` 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4 9422 3560 0.063 0.0323	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4 9317 3889 0.027 0.0326	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978 0.0437 0.0143	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058 0.0353 0.0237	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232 0.0229 0.0419
CompoundEmpirical formulaFormula weightTemperature/KCrystal systemSpace group $a/Å$ $b/Å$ $c/Å$ $a'^{\circ}$ $\beta/^{\circ}$ $\gamma/^{\circ}$ $V/Å^3$ ZNo. of reflections collectedNo. of unique reflections $R_{int}$ $R_i(reflections)$ $wR_2(reflections)$	C7 C <sub>20</sub> H <sub>17</sub> INSb 520.00 298 monoclinic P2(1)/c 9.1637(3) 10.9833(4) 18.0957(7) 90.00 95.784(1) 90.00 1812.02(11) 4 9043 3510 0.055 0.0354 0.0912	C8 C <sub>20</sub> H <sub>17</sub> FNSb 412.10 293 monoclinic P2(1)/n 10.2974(8) 10.3031(8) <sup>°</sup> 15.7161(11) 90.00 101.559 (1) 90.00 1633.6(2) 4 9422 3560 0.063 0.0323 0.0840	C9 C <sub>18</sub> H <sub>21</sub> CINSb 408.56 273 Monoclinic P2(1)/n 9.7692(12) 15.933(2) 11.4491(14) 90.00 110.103(2) 90.00 1 673.5(4) 4 9317 3889 0.027 0.0326 0.0860	C10 C <sub>18</sub> H <sub>21</sub> BrNSb 453.02 273 monoclinic P2(1)/n 9.8178(4) 16.1278(5) 11.4078(5) 90.00 109.895(1) 90.00 1698.50(11) 4 49645 2978 0.0437 0.0143 0.0541	C11 C <sub>18</sub> H <sub>21</sub> INSb 500.01 100 monoclinic P2(1)/c 12.3974(4) 9.4136(3) 14.8803(5) 90.00 92.218(3) 90.00 1735.29(10) 4 11089 3058 0.0353 0.0237 0.0516	C12 C <sub>18</sub> H <sub>21</sub> FNSb 392.11 273 monoclinic <i>Cc</i> 17.7432(9) 10.6409(6) 17.8551(10) 90.00 96.996(2) 90.00 3346.0(3) 8 31989 8232 0.0229 0.0419 0.1205

#### Table 3

Cytotoxic effect of organoantimony(III) halide complexes C1-C12, nitrogen-containing precursors P1-P3 and cisplatin on various cancerous and nonmalignant cell lines after 24 h

Cell lines	HepG2		MDA-MB-231		MCF-7		HeLa		HEK-293
Compound	$IC_{50} \pm SD^{a}$	$SI^{b}$	$\rm IC_{50}\pm SD$	SI <sup>c</sup>	$\text{IC}_{50}\pm\text{SD}$	$SI^d$	$\rm IC_{50}\pm SD$	SI <sup>e</sup>	$\rm IC_{50}\pm SD$
C1	$1.84\pm0.42$	5.05	$1.40\pm0.32$	6.64	$6.08 \pm 1.38$	1.53	$1.63\pm0.49$	5.70	$9.29 \pm 1.16$
C2	$4.86\pm0.91$	2.10	$3.63\pm0.87$	2.82	$6.39 \pm 1.45$	1.60	$3.12\pm0.45$	3.28	$10.23\pm0.55$
C3	$1.61\pm0.33$	5.63	$1.28\pm0.54$	7.08	$5.79 \pm 1.56$	1.56	$1.13\pm0.21$	8.02	$9.06 \pm 1.31$
C4	$1.06\pm0.17$	7.15	$0.91\pm0.22$	8.33	$2.14\pm0.93$	3.54	$0.93\pm0.15$	8.15	$7.58\pm0.89$
C5	$2.78\pm0.59$	3.07	$1.72\pm0.33$	4.97	$3.14\pm0.81$	2.72	$3.32\pm0.50$	2.57	$8.54 \pm 1.88$
C6	$1.90\pm0.12$	4.10	$1.05\pm0.13$	7.42	$2.61\pm0.60$	2.98	$2.34\pm0.32$	3.33	$7.79 \pm 1.36$
C7	$1.87\pm0.11$	3.33	$1.04\pm0.11$	5.98	$2.39 \pm 1.67$	2.60	2.06 ± 0.29	3.02	$6.22 \pm 1.69$
C8	$0.88\pm0.32$	4.41	$0.52\pm0.02$	7.46	$2.31\pm0.32$	1.68	$1.12\pm0.21$	3.46	$3.88 \pm 1.01$
С9	$1.41\pm0.06$	4.03	$0.84\pm0.23$	6.76	$3.41\pm0.30$	1.67	$1.98\pm0.54$	2.87	$5.68 \pm 1.32$
C10	$2.74\pm0.63$	2.27	$1.85\pm0.25$	3.36	$6.34 \pm 1.31$	0.98	$4.78\pm0.83$	1.30	$6.22 \pm 1.07$
C11	1.84 ?0.28	4.05	$0.97\pm0.19$	7.69	$4.89 \pm 1.13$	1.53	$2.09\pm0.62$	3.57	$7.46\pm0.66$
C12	$0.71\pm0.06$	3.51	$0.51\pm0.12$	4.88	$0.96\pm0.83$	2.59	1.08 ?0.47	2.31	$2.49\pm0.67$
P1	> 50.00	-	> 50.00	-	> 50.00		> 50.00		> 50.00
P2	> 50.00	-	> 50.00	_	> 50.00	-	> 50.00		> 50.00
P3	> 50.00	-	> 50.00	-	> 50.00	2	> 50.00		> 50.00
cisplatin	$13.19 \pm 4.51$	3.71	$12.63 \pm 1.22$	3.88	$22.75\pm8.59$	2.15	$17.35\pm3.64$	2.82	$48.96 \pm 8.75$

Cancerous Cell Lines: HepG2 (human liver hepatocellular carcinoma cell line), MDA-MB-231 (human breast cancer cell line), MCF-7 (human breast adenocarcinoma cell line) and HeLa (human cervical carcinoma cell line). Nonmalignant Cell Line: HEK-293 (human embryonic kidney cell line). IC<sub>50</sub>: concentration that is cytotoxic against 50% of cell lines. SI: selectivity index. <sup>*a*</sup> The IC<sub>50</sub> values were determined through non-linear regression analysis; Each well was triplicated and each experiment was repeated at least three times. IC<sub>50</sub> values quoted are mean  $\pm$  SD ( $\mu$ M). <sup>*b*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7.<sup>*e*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7.<sup>*e*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7.<sup>*e*</sup> Calculated as the ratio between IC<sub>50</sub> in HEK-293 cells and IC<sub>50</sub> in MCF-7.<sup>*e*</sup>

# Highlights

- Cytotoxicity against human cancerous cell lines of Sb(III) halide complexes was evaluated.
- The cytotoxicity was closely related to the  $N \rightarrow Sb$  coordinate bond length.
- Sb(III) complex C4 exhibited the highest selectivity index.
- C4 induced S phase cell cycle arrest and necrosis in MDA-MB-231 cells.
- The cytotoxicity was dependent on the production of intracellular reactive oxygen species.

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