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RESEARCH PAPER



Synthesis and evaluation of anticancer, antiphospholipases, antiproteases, and antimetabolic syndrome activities of some 3H-quinazolin-4-one derivatives

Nahed N. E. El-Sayed^{a,b}, Norah M. Almanei^a, Abir Ben Bacha^c, Omar Al-Obeed^d, Rehan Ahmad^d, Maha Abdulla^d and Ahmed M. Alafeefy^e

^aDepartment of Chemistry, College of Science, King Saud University, Riyadh, Saudi Arabia; ^bNational Organization for Drug Control and Research, Giza, Egypt; ^cBiochemistry Department, College of Sciences, King Saud University, Riyadh, Saudi Arabia; ^dDepartment of Surgery, College of Medicine, King Saud University, Riyadh, Saudi Arabia; ^eDepartment of Chemistry, Kulliyah of Science, International Islamic University Malaysia, Kuantan, Pahang Darul Makmur, Malaysia

ABSTRACT

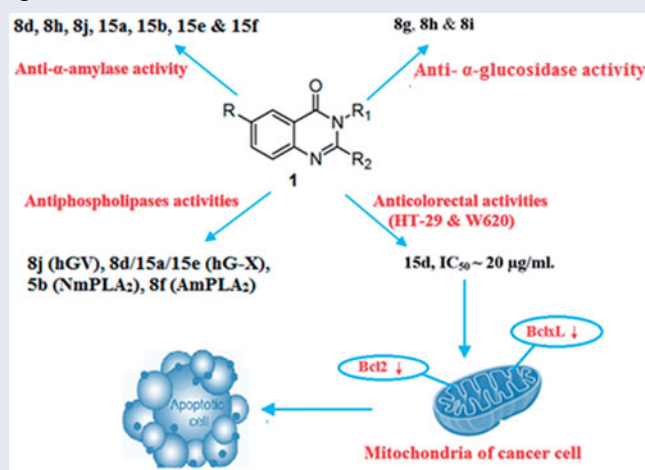
Some new 3H-quinazolin-4-one derivatives were synthesised and screened for anticancer, antiphospholipases, antiproteases, and antimetabolic syndrome activities. Compound **15d** was more potent in reducing the cell viabilities of HT-29 and SW620 cells lines to 38%, 36.7%, compared to 5-FU which demonstrated cell viabilities of 65.9 and 42.7% respectively. The IC₅₀ values of **15d** were ~20 µg/ml. Assessment of apoptotic activity revealed that **15d** decreased the cell viability by down regulating Bcl2 and BclxL. Moreover, compounds, **8j**, **8d/15a/15e**, **5b**, and **8f** displayed lowered IC₅₀ values than oleanolic acid against proinflammatory isoforms of hGV, hG-X, NmPLA₂, and AmPLA₂. In addition, **8d**, **8h**, **8j**, **15a**, **15b**, **15e**, and **15f** showed better anti-α-amylase than quercetin, whereas **8g**, **8h**, and **8i** showed higher anti-α-glucosidase activity than allopurinol. Thus, these compounds can be considered as potential antidiabetic agents. Finally, none of the compounds showed higher antiproteases or xanthine oxidase activities than the used reference drugs.

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Introduction

Colorectal cancer (CRC) is the third-most common cancer and the fourth-most common cause of cancer-related death worldwide¹. One of the key features of cancer is deregulation of apoptosis of the altered cells². Apoptosis is mediated by a complex network of BCL-2 family of proteins, comprising two types of proteins, namely anti-apoptotic proteins, which include Bcl2, BclxL, and Mcl-1, and pro-apoptotic proteins, which include Bax and Bak³. Hence, inhibiting the anti-apoptotic BCL-2 proteins can be established as a mechanism of anticancer defence for the treatment of CRC⁴.

Several *in vitro* and *in vivo* clinical studies have suggested a positive correlation between the activity of certain proteases, such as trypsin⁵, cathepsin B⁶, collagenases⁷, thrombin⁸, and neutrophil elastase⁹ and colorectal carcinogenesis, invasion, proliferation, and metastasis.

Furthermore, three phospholipases, namely (hsPLA₂-IIA), group V, and group X¹⁰ are found to play indirect pro-tumorigenic roles by promoting chronic inflammatory bowel diseases (IBD) that are common risk factors associated with CRC¹¹.

In addition, recent studies have linked the incidence of CRC to certain metabolic syndrome components, especially elevated

CONTACT Nahed N. E. El-Sayed ✉ nelsayed@ksu.edu.sa, nahednas@yahoo.com Department of Chemistry, College of Sciences, King Saud University, P.O. Box 22452, Riyadh 11451, Saudi Arabia

Supplemental data for this article can be accessed [here](#).

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plasma glucose (type 2 diabetes mellitus)¹². Several enzymes are reported to be involved in glucose homeostasis, among them α -amylase, α -glucosidase¹³, and xanthine oxidase¹⁴. Hence, targeting these enzymes has high potential in preventing CRC.

The 3H-quinazolin-4-one scaffold is known to be an important building block of various bioactive natural products¹⁵. In addition to this, many marketed drugs or experimental drug candidates incorporate this pharmacophore as part of the overall topography of the molecules¹⁶ such as ispinesib, nolatrexed, and raltitrexed (the anticancer agents); afloqualone with sedative and muscle relaxant effects, balaglitazone with antidiabetic activities, proquazone (as non-steroidal anti-inflammatory agent), and elinogrel (as reversible P2Y12 receptor antagonist) which is used for the treatment of acute coronary syndrome. Moreover, several new methodologies have been developed for the synthesis of 4-quinazolinone derivatives either using one pot multi-component approaches¹⁷ or *via* multistep routes for investigating their pharmacological potential as dual EGFR/HER2 inhibitors¹⁸, phosphoinositide 3-kinase inhibitors¹⁹, and as anti-hypertensive²⁰, antioxidant²¹, anticonvulsant²², and anti-inflammatory²³ agents.

Based on these observations, and in continuation to our ongoing research programme aims to identify new bioactive molecules^{24,25} herein, we report the synthesis, characterisation, and cytotoxicity screening (including evaluation of apoptotic activity), for a new series of 3H-quinazolin-4-one derivatives against HT-29 and SW620 cell lines derived from CRC. Furthermore, the inhibitory potentials of the synthesised compounds against certain phospholipases and proteases implicated in cancer and inflammatory disorders, in addition to α -amylase, α -glucosidase, and xanthine oxidase, managing metabolic syndrome are investigated with the aim of eradicating the risk factors for CRC or combatting the disease.

Materials and methods

Chemistry

The reactions were monitored and the purity of the final products was checked using thin layer chromatography (TLC) on pre-coated silica gel sheets (60 F254, Merck, Kenilworth, NJ). The developed plates were examined by exposure to ultraviolet light using VL-6.LC (254, 365 nm, 50/60 Hz). All the melting points were determined in open capillary tubes with a Gallenkamp melting point apparatus (°C). The infra-red (IR) spectra were recorded on a Perkin Elmer Fourier transform infra-red (FTIR) spectrophotometer (Spectrum BX 1000) in wave numbers (cm⁻¹) with potassium bromide (KBr) discs. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker NMR spectrometer: 1-Ascend 850 MHz for ¹H and 213 MHz for ¹³C, or on 2-Avance 600 MHz for ¹H and 150 MHz for ¹³C (Nuclear Magnetic Resonance Center, KAU, Jeddah, KSA), or on a Bruker Avance 500 spectrometer operating at 500 MHz for ¹H and 125 MHz for ¹³C at 25 °C (Research Unit, College of Pharmacy, Prince Sattam Bin Abdulaziz University, AlKharij, KSA), or on an Eclipse 300 FT NMR spectrometer operating at 300 MHz spectrometer for ¹H and at 75 MHz for ¹³C at 25 °C (KSU, Riyadh, KSA). The chemical shifts were expressed in (ppm) using tetramethylsilane as the internal standard; coupling constants (*J*) were expressed in Hz. Deuteriochloroform (CDCl₃) was used as solvent. The splitting patterns (multiplicities) in ¹H NMR were designated as: singlet (s), broad singlet (br. s), doublet (d), double of doublet (dd), triplet (t), and multiplet (m). Mass spectra were recorded on Shimadzu Qp-2010 Plus mass spectrometer in

ionisation mode: EI (Micro Analytical Center, Cairo University, Egypt). Elemental analyses were obtained from the Microanalytical Data Unit at Cairo University, Giza, Egypt.

Compounds **3a,b**²⁶, **3c**²⁷, **3d**²⁸, **6a**²⁹, **6b**³⁰, **13a**, **13b**, and **15a**³¹ were reported previously.

General procedure for the preparation of compounds 5a–c^{32,33}

A mixture of the appropriate 2-methyl-benzo[d][1,3]oxazin-4-one derivative **3a–c** (0.003 mol) and 3,4,5-trimethoxyaniline **4** (0.003 mol, 0.55 g) was fused in oil bath at 200 °C for 2 h. After cooling down the temperature to 100 °C, 37% HCl (Conc) (3 ml) was added till all the material was dissolved completely. The reaction mixture was cooled down to room temperature and was made alkaline with diluted NaOH solution (25%). The crude precipitated solid in each case, was filtered, washed with water, dried, and recrystallised from ethanol to yield compounds **5a–c**.

2-Methyl-3-(3,4,5-trimethoxy-phenyl)-3H-quinazolin-4-one (5a)

Black powder, yield (53%), mp 88–90 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3080 (CH-Ar), 2935 & 2837 (CH-aliphatic), 2368, 1682 (C=O), 1600 & 1503 (C=C), 1465, 1422, 1376, 1321, 1233, 1124, 1002, 936, 835, 771, 700, 653, 524; δ_{H} (850 MHz; CDCl₃) 8.26 (1H, d, *J* = 7.6, CH⁵-quinazolin-4-one), 7.77–7.75 (1H, m, CH⁶-quinazolin-4-one), 7.66 (1H, dd, *J* = 3.4, 8.5, CH⁸-quinazolin-4-one), 7.47–7.45 (1H, m, CH⁷-quinazolin-4-one), 6.47 (2H, s, H² and H^{6'}-trimethoxyphenyl), 3.90 (3H, s, OCH₃), 3.84 (6H, s, 2 × OCH₃), 2.32 (3H, s, CH₃); δ_{C} (213 MHz; CDCl₃) 162.36 (C=O), 154.41 (C=N), 154.22, 147.41, 138.46, 134.70, 133.29, 127.04, 126.79, 126.73, 120.69, 105.19 (6 × CH-Ar & 6 × C_q-Ar), 60.96 (OCH₃), 56.27 (2 × OCH₃), 24.04 (CH₃); MS (EI) *m/z* (%) [*M*⁺ + 2] 328.10 (2.50), [*M*⁺ + 1] 327.10 (16.69), [*M*⁺] 326.10 (77.52), 311.10 (46.20), 297.10 (3.85), 279.05 (2.13), 265.05 (1.90), 183.05 (4.87), 168.10 (7.32), 147.65 (8.78), 144.10 (14.54), 143.10 (100.00), 134.10 (4.69), 116.10 (6.24), 102.05 (4.96), 77.00 (6.38), 57.05 (3.38). Anal. calcd. for C₁₈H₁₈N₂O₄ (326.13): C, 66.25; H, 5.56; N, 8.58. Found: C, 66.23; H, 5.46; N, 8.66.

6-Fluoro-2-methyl-3-(3,4,5-trimethoxy-phenyl)-3H-quinazolin-4-one (5b)

Black crystals, yield (55%), mp 193–195 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3079 (CH-Ar), 2931 & 2835 (CH-aliphatic), 2375, 2278, 1673 (C=O), 1601 & 1484 (C=C), 1425, 1374, 1339, 1313, 1235, 1183, 1125 (C-F), 1065, 1008, 966, 878, 843, 778, 752, 723, 674, 626, 558, 526, 458; δ_{H} (500 MHz; CDCl₃) 7.84–7.82 (1H, m, CH⁵-quinazolin-4-one), 7.67–7.65 (1H, m, CH⁷-quinazolin-4-one), 7.49–7.45 (1H, m, CH⁸-quinazolin-4-one), 6.52 (2H, s, H² and H^{6'}-trimethoxyphenyl), 3.91 (3H, s, OCH₃), 3.86 (6H, s, 2 × OCH₃), 2.32 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 161.62 (C=O), 159.65 (C_q-F), 154.20 (C=N), 153.65, 144.06, 138.48, 133.03, 129.23, 123.18, 122.99, 121.87, 105.12 (5 × CH-Ar & 6 × C_q-Ar), 60.89 (OCH₃), 56.24 (2 × OCH₃), 23.87 (CH₃); MS (EI) *m/z* (%) [*M*⁺ + 1] 345.10 (21.69), [*M*⁺] 344.10 (100) 330.10 (11.87), 329.10 (57.89), 313.10 (3.99), 297.05 (2.71), 161.05 (86.15), 162.05 (11.84), 134.10 (7.80), 107.05 (4.64), 94.05 (7.76), 77.00 (1.91), 57.00 (3.40). Anal. calcd. for C₁₈H₁₇FN₂O₄ (344.12): C, 62.79; H, 4.98; N, 8.14. Found: C, 62.68; H, 4.63; N, 8.02.

6-Chloro-2-methyl-3-(3,4,5-trimethoxy-phenyl)-3H-quinazolin-4-one (5c)

Black powder, yield (60%), mp 188–190 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3075 (CH-Ar), 2934 & 2839 (CH-aliphatic), 2369, 1680 (C=O), 1598 & 1504 (C=C), 1467, 1423, 1375, 1312, 1236, 1126, 998, 948, 892, 834 (C-Cl), 771, 738, 692, 613, 524, 452; δ_{H} (500 MHz; CDCl₃) 8.18 (1H, d, J = 2.4, CH⁵-quinazolin-4-one), 7.69 (1H, dd, J = 8.7, 2.4, CH⁷-quinazolin-4-one), 7.60 (1H, d, J = 8.7, CH⁸-quinazolin-4-one), 6.51 (2H, s, H^{2'} and H^{6'}-trimethoxyphenyl), 3.92 (3H, s, OCH₃), 3.88 (6H, s, 2 × OCH₃), 2.32 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 161.28 (C=O), 154.75 (C=N), 154.25, 145.91, 138.56, 135.02, 132.95, 132.36, 128.52, 126.23, 121.73, 105.09 (5 × CH-Ar & 7 × C_q-Ar), 60.93 (OCH₃), 56.28 (2 × OCH₃), 24.02 (CH₃); MS (EI) m/z (%) [M⁺, Cl³⁷] 362.10 (34.88), [M⁺, Cl³⁵] 360.10 (100.00), 347.05 (22.20), 345.10 (60.80), 331.05 (4.80), 329.05 (4.53), 315.05 (1.90), 313.05 (3.09), 301.05 (1.04), 299.10 (1.56), 179.00 (27.46), 177.00 (84.58), 152.05 (3.81), 150.05 (6.54), 136.05 (5.31), 123.05 (3.29), 110.00 (6.17), 75.00 (6.71), 57.10 (3.05). Anal. calcd. for C₁₈H₁₇ClN₂O₄ (360.09): C, 59.92; H, 4.75; N, 7.76. Found: C, 59.88; H, 4.62; N, 7.59.

Synthesis of Schiff's base derivatives (8a–j)³⁴

A mixture of 3-amino-2-methyl-3H-quinazolin-4-one derivatives **6a,b** (0.002 mol) and the appropriate aromatic aldehyde **7a–e**, namely, 2-naphthaldehyde, 4-(pyridin-2-yl) benzaldehyde, 3-methoxy-2-nitrobenzaldehyde, 4-formylbenzonitrile, and 1,1'-biphenyl-4-carbaldehyde (0.002 mol) in absolute ethanol (20 ml) containing (3 ml) glacial acetic acid was heated under reflux for 10 h. The excess solvent was removed under reduced pressure. The obtained solid in each case was washed with water, air dried, and recrystallised from the appropriate solvent to obtain the corresponding Schiff's base.

(E)-2-Methyl-3-[(naphthalen-2-ylmethylene)-amino]-3H-quinazolin-4-one (8a)

White powder (ethanol), yield (74%), mp 163–165 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3062 (CH-Ar), 2929 (CH-aliphatic), 2371, 2340, 1667 (C=O), 1596 & 1468 (C=C), 1426, 1377, 1346, 1320, 1268, 1227, 1156, 1121, 1031, 934, 898, 866, 831, 778, 755, 693, 660, 621, 477; δ_{H} (600 MHz; CDCl₃) 9.18 (1H, s, CH=N), 8.30 (1H, d, J = 8.4, CH-Ar), 8.16–8.14 (2H, m, 2 × CH-Ar), 7.93–7.92 (2H, m, 2 × CH-Ar), 7.89 (1H, d, J = 7.8, CH-Ar), 7.75 (1H, d, J = 7.2, CH-Ar), 7.67 (1H, d, J = 8.4, CH-Ar), 7.59 (1H, t, J = 7.2, CH-Ar), 7.55 (1H, t, J = 7.8, CH-Ar), 7.47 (1H, t, J = 7.8, CH-Ar), 2.70 (3H, s, CH₃); δ_{C} (150 MHz; CDCl₃) 166.32 (C=O), 158.84 (HC=N), 154.21 (C_q=N), 146.49, 135.41, 134.37, 132.93, 132.42, 130.46, 128.97, 128.95, 128.20, 128.02, 127.30, 126.95, 126.94, 126.46, 122.88, 121.58 (11 × CH-Ar & 5 × C_q-Ar), 23.00 (CH₃); MS (EI) m/z (%) [M⁺ + 1] 314.10 (0.44), [M⁺] 313.10 (1.41), 161.05 (10.24), 160.10 (100.00), 153.10 (14.99), 139.10 (4.10), 127.10 (8.79), 118.10 (11.47), 90.05 (11.47), 76.00 (13.65), 63.00 (2.80), 50.00 (3.12). Anal. calcd. for C₂₀H₁₅N₃O (313.12): C, 76.66; H, 4.82; N, 13.41. Found: C, 76.62; H, 4.77; N, 13.29.

(E)-6-Bromo-2-methyl-3-[(naphthalen-2-ylmethylene)-amino]-3H-quinazolin-4-one (8b)

White powder (ethanol), yield (65%), mp 200 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3056 (CH-Ar), 2926 (CH-aliphatic), 1932, 1675 (C=O), 1602 & 1465 (C=C), 1371, 1347, 1313, 1272, 1235, 1208, 1180, 1153, 1122, 1072, 1037, 954, 890, 860, 831, 812, 777, 740, 700, 672 (C-Br), 629, 569, 539, 514, 473; δ_{H} (500 MHz; CDCl₃) 9.15 (1H, s, CH=N), 8.39

(1H, s, CH-Ar), 8.15 (1H, s, CH-Ar), 8.12 (1H, d, J = 8.5, CH-Ar), 7.92 (2H, d, J = 8.2, 2 × CH-Ar), 7.88 (1H, d, J = 7.9, CH-Ar), 7.79 (1H, dd, J = 8.5, 1.8, CH-Ar), 7.61–7.53 (3H, m, 3 × CH-Ar), 2.68 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 166.60 (C=O), 157.65 (HC=N), 154.66 (C_q=N), 145.31, 137.46, 135.46, 132.90, 132.56, 130.26, 129.69, 128.98, 128.80, 128.28, 128.01, 126.98, 122.98, 122.81, 119.86 (10 × CH-Ar & 6 × C_q-Ar), 22.98 (CH₃); MS (EI) m/z (%) [M⁺, ⁸¹Br] 393.05 (2.94), [M⁺, ⁷⁹Br] 391.00 (2.85), 239.95 (100.00), 237.95 (98.69), 197.90 (6.54), 195.95 (4.69), 155.95 (5.37), 153.05 (22.90), 127.10 (21.27), 100.05 (3.19), 75.00 (18.97), 63.00 (3.47), 57.05 (1.26). Anal. calcd. for C₂₀H₁₄BrN₃O (391.03): C, 61.24; H, 3.60; N, 10.71. Found: C, 61.08; H, 3.39; N, 10.65.

(E)-2-Methyl-3-[(4-pyridin-2-yl-benzylidene)-amino]-3H-quinazolin-4-one (8c)

Beige powder (benzene), yield (85%), mp 188–190 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3042 (CH-Ar), 2940 (CH-aliphatic), 1675 (C=O), 1600 & 1468 (C=C), 1436, 1378, 1327, 1230, 1181, 1154, 1108, 987, 939, 875, 847, 772, 718, 688, 663, 626, 589, 479; δ_{H} (500 MHz; CDCl₃) 9.12 (1H, s, CH=N), 8.74 (1H, d, J = 4.7, CH-Ar), 8.30 (1H, d, J = 7.9, CH-Ar), 8.15 (2H, d, J = 8.2, 2 × CH-Ar), 8.00 (2H, d, J = 8.2, 2 × CH-Ar), 7.80–7.73 (3H, m, 3 × CH-Ar), 7.68 (1H, d, J = 8.1, CH-Ar), 7.48 (1H, t, J = 7.5, CH-Ar), 7.36–7.28 (1H, m, CH-Ar), 2.69 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 165.64 (C=O), 158.80 (HC=N), 156.12 (HC=N), 154.18 (C_q=N), 149.93, 146.46, 143.05, 136.92, 134.34, 133.19, 129.24, 127.35, 127.27, 126.93, 126.43, 122.87, 121.56, 120.88 (11 × CH-Ar & 5 × C_q-Ar), 22.94 (CH₃); MS (EI) m/z (%) [M⁺ + 1] 341.10 (0.69), [M⁺] 340.10 (1.39), 181.05 (4.88), 161.05 (10.51), 160.10 (100.00), 145.10 (2.83), 132.10 (3.91), 118.10 (12.08), 90.00 (5.53), 76.00 (16.01), 50.00 (3.71). Anal. calcd. for C₂₁H₁₆N₄O (340.13): C, 74.10; H, 4.74; N, 16.46. Found: C, 73.92; H, 4.63; N, 16.28.

(E)-6-Bromo-2-methyl-3-[(4-pyridin-2-yl-benzylidene)-amino]-3H-quinazolin-4-one (8d)

Beige powder (benzene), yield (77%), mp 209–210 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3149 & 3060 (CH-Ar), 2952 (CH-aliphatic), 1670 (C=O), 1604 & 1465 (C=C), 1375, 1319, 1278, 1240, 1208, 1158, 1124, 1961, 1035, 987, 876, 833, 775, 727, 674 (C-Br), 645, 564; δ_{H} (850 MHz; CDCl₃) 9.10 (1H, s, CH=N), 8.77 (1H, d, J = 4.3, CH-Ar), 8.41 (1H, d, J = 2.6, CH-Ar), 8.17 (2H, d, J = 7.7, 2 × CH-Ar), 8.01 (2H, d, J = 7.7, 2 × CH-Ar), 7.85–7.79 (3H, m, 3 × CH-Ar), 7.55 (1H, d, J = 8.5, CH-Ar), 7.36–7.32 (1H, m, CH-Ar), 2.68 (3H, s, CH₃); δ_{C} (213 MHz; CDCl₃) 165.98 (C=O), 157.67 (HC=N), 155.79 (HC=N), 154.69 (C_q=N), 149.51, 145.26, 137.56, 133.23, 129.74, 129.40, 128.79, 127.54, 123.08, 122.95, 121.23, 119.94 (10 × CH-Ar & 6 × C_q-Ar), 22.95 (CH₃); MS (EI) m/z (%) [M⁺ + 1] 419.05 (1.37), [M⁺] 418.00 (2.03), 240.00 (11.07), 239.95 (100.00%), 237.95 (98.10), 199.90 (2.69), 197.90 (6.83), 181.05 (6.55), 179.05 (4.60), 154.05 (13.02), 152.05 (1.88), 139.10 (3.04), 127.10 (6.15), 116.10 (3.42), 90.05 (4.15), 78.05 (3.66), 75.00 (17.16). Anal. calcd. for C₂₁H₁₅⁷⁹BrN₄O (418.04): C, 60.16; H, 3.61; N, 13.36. Found: C, 60.03; H, 3.43; N, 13.11.

(E)-3-[(3-Methoxy-2-nitro-benzylidene)-amino]-2-methyl-3H-quinazolin-4-one (8e)

Beige powder (ethanol), yield (55%), mp 178–180 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3017 (CH-Ar), 2975 & 2935 (CH-aliphatic), 1680 (C=O), 1615 & 1571 (C=C), 1537, 1464, 1428, 1376, 1283, 1227, 1156, 1107, 1055, 1029, 964, 933, 875, 850, 772, 743, 691, 651, 589, 471, 419;

δ_{H} (850 MHz; CDCl_3) 9.41 (1H, s, CH=N), 8.26 (1H, d, $J = 7.7$, CH-Ar), 7.75 (1H, t, $J = 7.7$, CH-Ar), 7.67 (1H, d, $J = 7.7$, CH-Ar), 7.56 (1H, t, $J = 7.7$, CH-Ar), 7.49–7.46 (2H, m, $2 \times$ CH-Ar), 7.23 (1H, d, $J = 8.5$, CH-Ar), 3.96 (3H, s, OCH_3), 2.6 (3H, s, CH_3); δ_{C} (213 MHz; CDCl_3) 159.22 (C=O), 158.52 (HC=N), 154.87 ($\text{C}_q=\text{N}$), 151.44, 139.74, 134.76, 131.30, 127.39, 126.77, 126.14, 122.03, 121.38, 115.92 ($7 \times$ CH-Ar & $5 \times$ C_q -Ar), 56.78 (OCH_3), 22.65 (CH_3); MS (EI) m/z (%) [M^+] 338.10 (4.91), 321.10 (8.62), 290.10 (8.60), 261.10 (2.25), 186.05 (2.04), 160.10 (100.00), 143.10 (3.47), 132.10 (4.81), 117.10 (18.04), 104.05 (3.96), 76.00 (26.95), 63.00 (4.85), 50.00 (6.06). Anal. calcd. for $\text{C}_{17}\text{H}_{14}\text{N}_4\text{O}_4$ (338.10): C, 60.35; H, 4.17; N, 16.56. Found: C, 60.03; H, 3.94; N, 16.37.

(E)-6-Bromo-3-[(3-methoxy-2-nitro-benzylidene)-amino]-2-methyl-3H-quinazolin-4-one (8f)

Beige powder (ethanol), yield (66%), mp 234–235 °C; ν_{max} (KBr)/ cm^{-1} 3070 & 3012 (CH-Ar), 2977 & 2926 (CH-aliphatic), 1677 (C=O), 1601 & 1533 (C=C), 1464, 1436, 1373, 1303, 1207, 1153, 1104, 1063, 973, 937, 897, 836, 779, 739, 670 (C-Br), 639, 589; δ_{H} (850 MHz; CDCl_3) 9.35 (1H, s, CH=N), 8.38 (1H, d, $J = 2.6$, CH-Ar), 7.82 (1H, dd, $J = 8.5$, 2.6, CH-Ar), 7.58–7.56 (2H, m, $2 \times$ CH-Ar), 7.48 (1H, d, $J = 8.5$, CH-Ar), 7.25 (1H, d, $J = 8.5$, CH-Ar), 3.97 (3H, s, OCH_3), 2.63 (3H, s, CH_3); δ_{C} (213 MHz; CDCl_3) 159.28 (C=O), 157.93 (HC=N), 155.38 ($\text{C}_q=\text{N}$), 151.47, 144.56, 139.68, 137.94, 131.36, 129.85, 128.57, 125.86, 122.76, 122.14, 120.31, 116.16 ($6 \times$ CH-Ar & $6 \times$ C_q -Ar), 56.80 (OCH_3), 22.63 (CH_3); MS (EI) m/z (%) [M^+ , ^{81}Br] 418.00 (4.65), [M^+ , ^{79}Br] 416.00 (4.58), 401.00 (6.90), 399.00 (6.63), 370.00 (5.89), 368.00 (5.13), 341.00 (1.54), 334.00 (1.46), 265.95 (2.41), 263.95 (2.42), 239.95 (97.78), 237.95 (100.00), 197.90 (8.61), 195.95 (6.33), 169.95 (3.24), 167.95 (2.54), 155.95 (7.13), 153.95 (7.22), 133.10 (1.53), 131.10 (3.71), 117.10 (7.38), 116.10 (5.63), 101.10 (4.34), 89.00 (4.73), 76.00 (9.56), 59.05 (13.18), 57.05 (4.20). Anal. calcd. for $\text{C}_{17}\text{H}_{13}^{79}\text{BrN}_4\text{O}_4$ (416.01): C, 48.94; H, 3.14; N, 19.15. Found: C, 48.83; H, 2.97; N, 18.93.

(E)-4-[(2-Methyl-4-oxo-4H-quinazolin-3-ylimino)-methyl]-benzonitrile (8g)

White powder (acetone), yield (55%), mp 222–224 °C; ν_{max} (KBr)/ cm^{-1} 3043 (CH-Ar), 2960 (CH-aliphatic), 2226 (CN), 1674 (C=O), 1599 & 1467 (C=C), 1406, 1376, 1320, 1235, 1145, 1111, 1034, 944, 868, 836, 766, 687, 660, 555; δ_{H} (500 MHz; CDCl_3) 9.37 (1H, s, CH=N), 8.28 (1H, d, $J = 8.0$, CH^5 -quinazolin-4-one), 8.00 (2H, d, $J = 8.2$, $2 \times$ CH-benzonitrile ring), 7.80 (2H, d, $J = 8.2$, $2 \times$ CH-benzonitrile ring), 7.77 (1H, app. t, $J = 7.1$, CH^7 -quinazolin-4-one), 7.68 (1H, d, $J = 8.0$, CH^8 -quinazolin-4-one), 7.49 (1H, app. t, $J = 7.6$, CH^6 -quinazolin-4-one), 2.73 (3H, s, CH_3); δ_{C} (125 MHz; CDCl_3) 162.10 (C=O), 159.10 (HC=N), 154.14 ($\text{C}_q=\text{N}$), 146.23, 137.25, 134.71, 132.69, 128.98, 127.34, 127.07, 126.75, 121.48, 118.16, 115.36 ($8 \times$ CH-Ar, $4 \times$ C_q -Ar & C≡N), 23.02 (CH_3); (EI) m/z (%) [$\text{M}^+ + 2$] 290.10 (3.21), [$\text{M}^+ + 1$] 289.10 (1.58), [M^+] 288.10 (5.65), 273.10 (3.00), 272.10 (6.19), 260.10 (1.60), 186.05 (3.69), 161.10 (10.65), 160.10 (100.00), 145.10 (4.68), 143.10 (3.86), 131.10 (14.47), 130.10 (17.06), 119.10 (11.52), 117.10 (19.54), 102.10 (18.65), 101.10 (5.04), 92.05 (5.14), 90.05 (11.61), 77.00 (20.32), 76.00 (37.53), 60.00 (4.25), 59.05 (19.07), 51.00 (6.85), 50.00 (14.71). Anal. calcd. for $\text{C}_{17}\text{H}_{12}\text{N}_4\text{O}$ (288.10): C, 70.82; H, 4.20; N, 19.43. Found: C, 70.69; H, 4.13; N, 19.27.

(E)-4-[(6-Bromo-2-methyl-4-oxo-4H-quinazolin-3-ylimino)-methyl]-benzonitrile (8h)

White powder (acetone), yield (75%), mp 228–230 °C; ν_{max} (KBr)/ cm^{-1} 3059 (CH-Ar), 2927 (CH-aliphatic), 2230 (CN), 1680 (C=O), 1602 & 1467 (C=C), 1420, 1376, 1328, 1277, 1242, 1211, 1156, 1123, 1071, 981, 899, 876, 831, 778, 723, 675 (C-Br), 644, 555, 433; δ_{H} (500 MHz; CDCl_3) 9.32 (1H, s, CH=N), 8.40 (1H, s, CH^5 -quinazolin-4-one ring), 8.00 (2H, d, $J = 7.7$, $2 \times$ CH-benzonitrile ring), 7.84 (1H, d, $J = 8.5$, CH^7 -quinazolin-4-one ring), 7.81 (2H, d, $J = 7.7$, $2 \times$ CH-benzonitrile ring), 7.55 (1H, d, $J = 8.5$, CH^8 -quinazolin-4-one ring), 2.64 (3H, s, CH_3); δ_{C} (125 MHz; CDCl_3) 162.65 (C=O), 157.83 (HC=N), 154.53 ($\text{C}_q=\text{N}$), 145.027, 137.87, 136.93, 132.67, 129.78, 129.01, 128.9, 122.81, 120.14, 118.04, 115.49 ($7 \times$ CH-Ar, $5 \times$ C_q -Ar & C≡N), 22.98 (CH_3); MS (EI) m/z (%) [M^+ , ^{81}Br] 368.01 (2.60), [M^+ , ^{79}Br] 366.01 (2.56), 354.30 (3.01), 313.30 (12.92), 297.30 (5.50), 239.20 (12.54), 227.10 (2.29), 213.15 (2.33), 185.10 (2.94), 171.10 (2.88), 143.15 (3.48), 129.10 (12.26), 111.15 (14.46), 97.15 (21.74), 83.05 (26.49), 71.05 (43.57), 69.05 (42.50), 59.05 (56.18), 57.05 (100.00). Anal. calcd. for $\text{C}_{17}\text{H}_{11}^{79}\text{BrN}_4\text{O}$ (366.01): C55.61; H, 3.02; N, 15.26. Found: C, 55.49; H, 2.88; N, 15.19.

(E)-3-[(Biphenyl-4-ylmethylene)-amino]-6-bromo-2-methyl-3H-quinazolin-4-one (8j)

White powder (ethanol), yield (97%), mp 208–210 °C; ν_{max} (KBr)/ cm^{-1} 3058 (CH-Ar), 2920 (CH-aliphatic), 1676 (C=O), 1604 & 1467 (C=C), 1373, 1319, 1276, 1236, 1206, 1180, 1154, 1074, 1007, 907, 875, 831, 757, 722, 689 (C-Br), 644, 555, 467; δ_{H} (500 MHz; CDCl_3) 9.03 (1H, s, CH=N), 8.41 (1H, d, $J = 2.3$, CH-Ar), 7.96 (2H, d, $J = 8.3$, $2 \times$ CH-Ar), 7.82 (1H, dd, $J = 8.7$, 2.3, CH-Ar), 7.73 (2H, d, $J = 8.3$, $2 \times$ CH-Ar), 7.66 (2H, d, $J = 7.3$, $2 \times$ CH-Ar), 7.54 (1H, d, $J = 8.7$, CH-Ar), 7.49 (2H, t, $J = 7.3$, $2 \times$ CH-Ar), 7.41 (1H, t, $J = 7.3$ CH-Ar), 2.63 (3H, s, CH_3); δ_{C} (125 MHz; CDCl_3) 166.41 (C=O), 157.64 (HC=N), 154.61 ($\text{C}_q=\text{N}$), 145.45, 145.34, 139.92, 137.49, 131.44, 129.71, 129.43, 129.02, 128.81, 128.25, 127.66, 127.23, 122.97, 119.87 ($12 \times$ CH-Ar & $6 \times$ C_q -Ar), 22.94 (CH_3); MS (EI) m/z (%) [M^+ , ^{81}Br] 419.05 (1.70), [M^+ , ^{79}Br] 417.05 (1.67), 239.95 (100.00), 237.95 (98.77), 197.90 (6.94), 195.95 (4.46), 179.05 (24.10), 165.05 (5.04), 152.10 (13.04), 131.10 (2.31), 116.10 (3.58), 116.10 (3.58), 90.05 (3.62), 75.00 (15.91), 63.00 (3.20), 57.05 (2.43). Anal. calcd. for $\text{C}_{22}\text{H}_{16}^{79}\text{BrN}_3\text{O}$ (417.05): C, 63.17; H, 3.86; N, 10.05. Found: C, 62.97; H, 3.63; N, 9.89.

Synthesis of (2-methyl-4-oxo-4H-quinazolin-3-yl)-carbamic acid isobutyl ester (10)

A mixture of 3-amino-2-methyl-3H-quinazolin-4-one **6a** (0.0028 mol, 0.5 g) and isobutyl chloroformate **9** (13.6 equiv., 0.038 mol, 5.2 g, 4.94 ml) was refluxed for 10 h³⁵. The excess reagent was evaporated under reduced pressure and the remaining residue was treated with diethyl ether. The precipitated solid was filtered, washed with water, and purified by recrystallisation from benzene to obtain the *title compound* as a beige powder with 50% yield, mp 168–170 °C; ν_{max} (KBr)/ cm^{-1} 3223 (NH), 3080 (CH-Ar), 2963 & 2875 (CH-aliphatic), 1755 (C=O), 1694 (C=O), 1651 (C=N), 1606 & 1508 (C=C), 1468, 1380, 1232, 1108, 1046, 988, 942, 872, 772, 719, 695, 633, 520, 462; δ_{H} (850 MHz; CDCl_3) 8.19 (1H, d, $J = 7.7$, CH-Ar), 7.75 (1H, app. t, $J = 7.7$, CH-Ar), 7.64 (1H, d, $J = 8.5$, CH-Ar), 7.62 (1H, br. s, NH), 7.44 (1H, t, $J = 7.7$, CH-Ar), 4.00 (2H, apparent s, O- CH_2 -CH-(CH_3)₂), 2.60 (3H, s, CH_3), 1.96–1.93 (1H, m, O- CH_2 -CH-(CH_3)₂), 0.94 (6H, d, $J = 6.0$, O- CH_2 -CH-(CH_3)₂); δ_{C} (125 MHz; CDCl_3) 160.78 (C=O), 156.28

(C=O), 156.15 (C=N), 146.83, 134.89, 126.94, 126.85, 126.58, 120.60 (4 × CH-Ar, 2 × C_q-Ar), 72.71 (OCH₂), 27.78 (CH), 21.52 (CH₃), 18.63 (2 × CH₃); MS (EI) *m/z* (%) [*M*⁺ + 1] 276.10 (21.00), [*M*⁺] 275.10 (100.00), 260.10 (1.46), 220.05 (4.41), 202.00 (11.98), 175.05 (95.02), 160.05 (9.43), 146.10 (85.56), 131.10 (4.53), 131.10 (4.53), 117.10 (27.22), 90.00 (23.15), 76.00 (30.46), 63.00 (8.24), 57.05 (86.01). Anal. calcd. for C₁₄H₁₇N₃O₃ (275.13): C, 61.08; H, 6.22; N, 15.26. Found: C, 61.00; H, 6.01; N, 15.17.

Synthesis of (E)-N,N-dimethyl-N'-(2-methyl-4-oxo-4H-quinazolin-3-yl)-formamidine (12)

A mixture of 3-amino-2-methyl-3H-quinazolin-4-one **6a** (0.0028 mol, 0.5 g) and N,N-dimethyl formamide dimethyl acetal **11** (5.39 equiv. 0.0151 mol, 1.79 g, 2 ml) in ethanol (15 ml) was refluxed for 2 h³⁶. The excess solvent was removed under reduced pressure. The remaining solid was washed with *n*-hexane and recrystallised from ethanol to obtain the *title compound* as a yellow powder with 50% yield, mp 100–102 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3010 (CH-Ar), 2924 & 2807 (CH-aliphatic), 2145, 1660 (C=O), 1621 (C=N), 1609 & 1589 (C=C), 1475, 1418, 1366, 1323, 1237, 1174, 1120, 1069, 944, 874, 783, 701, 660, 626, 551, 509; δ_{H} (500 MHz; CDCl₃) 8.23 (1H, dd, *J* = 7.7, 1.7, CH-Ar), 7.74 (1H, s, HC=N), 7.69–7.67 (1H, m, CH-Ar), 7.62 (1H, dd, *J* = 8.5, 2.6, CH-Ar), 7.41–7.39 (1H, m, CH-Ar), 3.08 (3H, s, N-CH₃), 3.04 (3H, s, N-CH₃), 2.52 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 162.22 (C=O), 160.08 (HC=N), 155.08 (C_q=N), 146.39, 133.39, 126.41, 125.61, 121.04 (4 × CH-Ar & 2 × C_q-Ar), 40.88, 34.53 (N(CH₃)₂), 22.56 (CH₃); MS (EI) *m/z* (%) [*M*⁺ + 1] 231.10 (5.52), [*M*⁺] 230.10 (34.52), 215.05 (1.48), 186.00 (10.30), 161.10 (11.53), 160.10 (100.00), 145.10 (5.33), 132.10 (7.42), 118.10 (21.02), 104.10 (1.60), 90.00 (9.13), 76.00 (16.94), 63.00 (5.01), 50.00 (6.27). Anal. calcd. for C₁₂H₁₄N₄O (230.12): C, 62.59; H, 6.13; N, 24.33. Found: C, 62.47; H, 5.98; N, 24.18.

Synthesis of 2-(4-chloro-benzylsulfanyl)-6-methyl-3-phenyl-3H-quinazolin-4-one (15a³¹, 15b) and 2-(4-oxo-3-phenyl-3,4-dihydro-quinazolin-2-ylsulfanyl)-acetamide derivatives (15c-f)

A mixture of the appropriate derivative of 2-mercapto-3-phenyl-3H-quinazolin-4-one (**13a'**, **b'**) (0.0018 mol) and 4-chlorobenzyl chloride **14a** or chloroacetamide derivatives **14b,c**^{37,38} (1 equiv., 0.0018 mol) and potassium carbonate (2.5 equiv., 0.0045 mol, 0.62 g) in dry acetone (20 ml) was refluxed for 6 h³⁹. The hot reaction mixture was filtered and the filtrate was concentrated under reduced pressure. The resulting solid in each case was washed with water, air dried, and recrystallised from ethanol to obtain compounds **15a-f**.

2-(4-Chloro-benzylsulfanyl)-6-methyl-3-phenyl-3H-quinazolin-4-one (15b)

White powder, yield (96%), mp 164–165 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3058 (CH-Ar), 2915 (CH-aliphatic), 1689 (C=O), 1617 & 1575 (C=C), 1548, 1487, 1401, 1399, 1335, 1299, 1259, 1204, 1128, 1089, 1014, 963, 824 (C-Cl), 742, 692, 641, 539, 506; δ_{H} (500 MHz; CDCl₃) 8.01 (1H, s, CH⁵-quinazolin-4-one), 7.54–7.48 (5H, m, 5 × CH-Ar), 7.29–7.18 (6H, m, 6 × CH-Ar), 4.32 (2H, s, CH₂), 2.44 (3H, s, CH₃); δ_{C} (125 MHz; CDCl₃) 161.83 (C=O), 155.60 (C=N), 145.82, 136.17, 136.12, 135.90, 135.38, 133.28, 130.77, 130.00, 129.72, 129.26, 128.68, 126.76, 126.05, 119.72 (12 × CH-Ar & 6 × C_q-Ar), 36.22 (CH₂), 21.33 (CH₃); MS (EI) *m/z* (%) [*M*⁺, ³⁷Cl] 394.05 (27.71), [*M*⁺ + 1, ³⁵Cl] 393.10 (19.45), [*M*⁺, ³⁵Cl] 392.10 (69.22), 375.05 (1.96),

361.10 (35.50), 359.10 (100.00), 324.10 (11.09), 301.00 (12.62), 283.05 (37.48), 267.05 (81.50), 249.10 (51.70), 235.05 (33.05), 201.00 (89.80), 193.05 (3.31), 180.05 (4.87), 166.05 (14.31), 150.05 (8.84), 133.10 (77.88), 104.10 (28.46), 89.00 (25.12), 77.00 (63.94), 63.00 (10.37), 51.00 (11.84). Anal. calcd. for C₂₂H₁₇³⁵ClN₂OS (392.08): C, 67.25; H, 4.36; N, 7.13. Found: C, 67.20; H, 4.32; N, 7.06.

N-(2,6-Dimethyl-phenyl)-2-(4-oxo-3-phenyl-3,4-dihydro-quinazolin-2-ylsulfanyl)-acetamide (15c)

White crystals, yield (55%), mp 218–220 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3331 (NH), 3065 (CH-Ar), 2971 & 2917 (CH-aliphatic), 2366, 1664 (C=O), 1601 (C=O), 1552 & 1493 (C=C), 1467, 1362, 1335, 1299, 1264, 1205, 1112, 1030, 962, 881, 768, 695, 641, 595, 568, 523, 468; δ_{H} (850 MHz; CDCl₃) 8.66 (1H, br. s, NH), 8.27 (1H, d, *J* = 7.7, CH-Ar), 7.76 (1H, t, *J* = 7.7, CH-Ar), 7.62 (1H, d, *J* = 7.7, CH-Ar), 7.59–7.58 (2H, m, 2 × CH-Ar), 7.47 (1H, t, *J* = 7.7, CH-Ar), 7.33–7.32 (2H, m, 2 × CH-Ar), 7.11–7.07 (2H, m, 2 × CH-Ar), 7.04 (2H, d, *J* = 6.8, 2 × CH-Ar), 3.99 (2H, s, CH₂), 2.18 (6H, s, 2 × CH₃); δ_{C} (312 MHz; CDCl₃) 166.94 (C=O), 161.33 (C=O), 157.78 (C=N), 146.99, 135.26, 135.23, 135.19, 133.55, 130.55, 130.05, 129.02, 128.27, 127.66, 127.38, 126.75, 125.73, 119.91 (12 × CH-Ar, 6 × C_q-Ar), 35.49 (CH₂), 18.33 (2 × CH₃); MS (EI) *m/z* (%) [*M*⁺ + 1] 416.15 (1.05), [*M*⁺] 415.15 (2.86), 400.10 (9.45), 296.05 (20.13), 295.05 (100.00), 268.05 (28.58), 253.05 (10.77), 235.10 (30.01), 221.05 (5.97), 177.05 (7.25), 162.00 (2.47), 148.05 (3.93), 132.10 (20.59), 119.05 (11.21), 105.10 (2.87), 91.05 (12.43). Anal. calcd. for C₂₄H₂₁N₃O₂S (415.14): C, 69.37; H, 5.09; N, 10.11. Found: C, 69.18; H, 4.92; N, 10.00.

N-(2,6-Dimethyl-phenyl)-2-(6-methyl-4-oxo-3-phenyl-3,4-dihydro-quinazolin-2-ylsulfanyl)-acetamide (15d)

White powder, yield (57%), mp 229–230 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3269 (NH), 3024 (CH-Ar), 2961 & 2919 (CH-aliphatic), 2372, 2341, 1688 (C=O), 1651 (C=O), 1574 & 1547 (C=C), 1486, 1408, 1336, 1296, 1260, 1199, 1103, 1032, 963, 912, 836, 774, 748, 692, 648, 604, 543, 504; δ_{H} (500 MHz; CDCl₃) 8.70 (1H, br. s, NH), 8.05 (1H, s, CH-Ar), 7.57–7.49 (5H, m, 5 × CH-Ar), 7.32–7.30 (2H, m, 2 × CH-Ar), 7.07–7.02 (3H, m, 3 × CH-Ar), 3.96 (2H, s, CH₂), 2.46 (3H, s, CH₃), 2.16 (6H, s, 2 × CH₃); δ_{C} (125 MHz; CDCl₃) 166.99 (C=O), 161.39 (C=O), 156.66 (C=N), 145.07, 136.98, 136.57, 135.42, 135.18, 133.64, 130.46, 130.01, 129.08, 128.25, 127.32, 127.05, 125.59, 119.64 (11 × CH-Ar, 7 × C_q-Ar), 35.49 (CH₂), 21.30 (CH₃), 18.35 (2 × CH₃); MS (EI) *m/z* (%) [*M*⁺ + 1] 430.10 (1.24), [*M*⁺] 429.15 (3.40), 414.10 (9.75), 309.05 (100.00), 282.05 (35.11), 267.05 (10.35), 249.10 (38.79), 235.05 (5.16), 205.05 (1.86), 177.05 (5.60), 160.05 (2.87), 146.10 (16.74), 133.10 (10.36), 121.10 (4.97), 104.05 (10.15), 91.05 (11.20), 77.00 (21.32), 59.00 (3.71). Anal. calcd. for C₂₅H₂₃N₃O₂S (429.15): C, 69.91; H, 5.40; N, 9.78. Found: C, 69.76; H, 5.32; N, 9.64.

N-(2,3-Dimethyl-phenyl)-2-(6-methyl-4-oxo-3-phenyl-3,4-dihydro-quinazolin-2-ylsulfanyl)-acetamide (15f)

White powder, yield (76%), mp 218–220 °C; $\nu_{\max}$ (KBr)/cm⁻¹ 3259 (NH), 3000 (CH-Ar), 2925 (CH-aliphatic), 2371, 2341, 1692 (C=O), 1648 (C=O), 1574 & 1545 (C=C), 1487, 1300, 1263, 1204, 1147, 963, 834, 787, 750, 694, 651; δ_{H} (500 MHz; CDCl₃) 8.90 (1H, s, NH), 8.06 (1H, s, CH-Ar), 7.57–7.53 (5H, m, 5 × CH-Ar), 7.49 (1H, d, *J* = 7.4, CH-Ar), 7.32–7.25 (2H, m, 2 × CH-Ar), 7.08 (1H, apparent t, *J* = 7.3, CH-Ar), 6.97 (1H, d, *J* = 6.9, CH-Ar), 3.97 (2H, s, CH₂), 2.48

(3H, s, CH₃), 2.23 (3H, s, CH₃), 2.01 (3H, s, CH₃); δ_C (125 MHz; CDCl₃) 167.11 (C=O), 161.38 (C=O), 156.74 (C=N), 145.11, 137.41, 136.94, 136.50, 135.39, 135.31, 130.46, 129.96, 129.10, 128.95, 127.31, 127.05, 125.93, 125.67, 121.93, 119.66 (11 $\times$ CH-Ar & 7 $\times$ C_q-Ar), 36.14 (CH₂), 21.30 (CH₃), 20.55 (CH₃), 13.82 (CH₃); MS (EI) m/z (%) [M⁺] 429.15 (0.15), 414.10 (0.65), 309.05 (100.00), 282.05 (5.52), 269.10 (6.32), 249.10 (13.83), 235.05 (3.23), 189.05 (1.52), 161.10 (1.76), 146.10 (10.05), 133.10 (6.23), 113.10 (7.77), 101.10 (4.81), 77.00 (13.43), 59.00 (21.59). Anal. calcd. for C₂₅H₂₃N₃O₂S (429.15): C, 69.91; H, 5.40; N, 9.78. Found: C, 69.85; H, 5.35; N, 9.75.

Biological evaluation

Cell culture

Human CRC cell lines, HT-29 and SW620 cells were grown in RPMI (Invitrogen) containing 10% heat-inactivated foetal bovine serum, 100 μ g/ml streptomycin, 100 units/ml penicillin and 2 mmol/L L-glutamine.

Cell viability assay

Cell viability was determined using the 3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide) MTT assay according to Mosmann⁴⁰. 5-Fluorouracil (5-FU) was used as positive control. Briefly, after culturing cells in 96 well plates for 24 h, they were treated with different compounds (30 μ g/ml) for 24 h. Freshly prepared MTT (5 mM) solutions were added to the cells and were further incubated for 2 h at 37 °C in 5% CO₂. One hundred microliters of dimethyl sulphoxide (DMSO) was added in each well to dissolve the formazan crystal, formed in the reaction during incubation by careful pipetting. The absorbance of the product was measured at 540 nm using a microplate reader. The experiments were performed in triplicate for each condition. The graph illustrates the mean and standard deviation (SD) values of three independent experiments.

Cytotoxicity assay using the xCELLigence system

The optimal seeding concentration for proliferation of HT-29 was determined. HT-29 cells (5000 cell in 150 μ l medium/well) and SW620 cells (12,000 cell in 150 μ l medium/well) were seeded in 16 well plates (E-plate 16-ACEA Biosciences Inc., San Diego, CA) as described in the xCELLigence Real Time Cell Analyzer (RTCA) DP instrument manual provided by the manufacturer. After 24 h, 20 and 40 μ g/ml of compound **15d** were added and the experiments were allowed to run for another 3 d. Baseline cell indexes were calculated for at least two measurements from three replicate experiments.

Western blotting

Whole cell lysates were prepared using radioimmunoprecipitation assay (RIPA) lysis buffer as described by Al-Khayal and co-workers⁴¹. Total protein concentration was determined using Bradford protein reagent (Bio-Rad, Hercules, CA). Soluble proteins were loaded on precast TGX gels and were analysed using immunoblotting with anti-Bcl2, anti-BclxL (dilution 1:1000; Santa Cruz Biotechnology, Dallas, TX), and anti- β -actin (1:10,000 Sigma, St. Louis, MO). Reactivity was detected using horseradish peroxidase-conjugated secondary antibodies and chemiluminescence by Clarity Western ECL substrate (Bio-Rad). Membrane was developed using C-Digit Blot Scanner (LI-COR, Hamburg, Germany).

In vitro evaluation of antiphospholipases activities

The inhibitory activities of the synthesised 3H-quinazolin-4-one derivatives against a group of phospholipases were investigated according to the method of De Aranjó and Radvány⁴² using five sPLA₂s: human group IIA (hG-IIA), human group V (hG-V), human group X (hG-X), *Apis mellifera* bee venom (AmPLA₂) and *Naja mossambica mossambica* sPLA₂ (NmPLA₂). Ten microliters of each sPLA₂ solution (20 μ g/ml) was mixed with 10 μ l of each compound at different concentrations (0–50 μ g/ml) and the mixture was incubated for 20 min at room temperature. Then, 1 ml of the PLA₂ substrate (3.5 mM lecithin suspended in 100 mM NaCl, 10 mM CaCl₂, 3 mM NaTDC, and 0.055 mM red phenol, pH 7.6) was added. The hydrolysis kinetics was followed spectrophotometrically for 5 min at 558 nm. The results were reported as the inhibition percentage that was calculated by comparison with a control experiment (absence of compound) and the IC₅₀ values were determined from the curve. Oleonic acid (Sigma) was used as the positive control.

In vitro evaluation of antiproteases activities

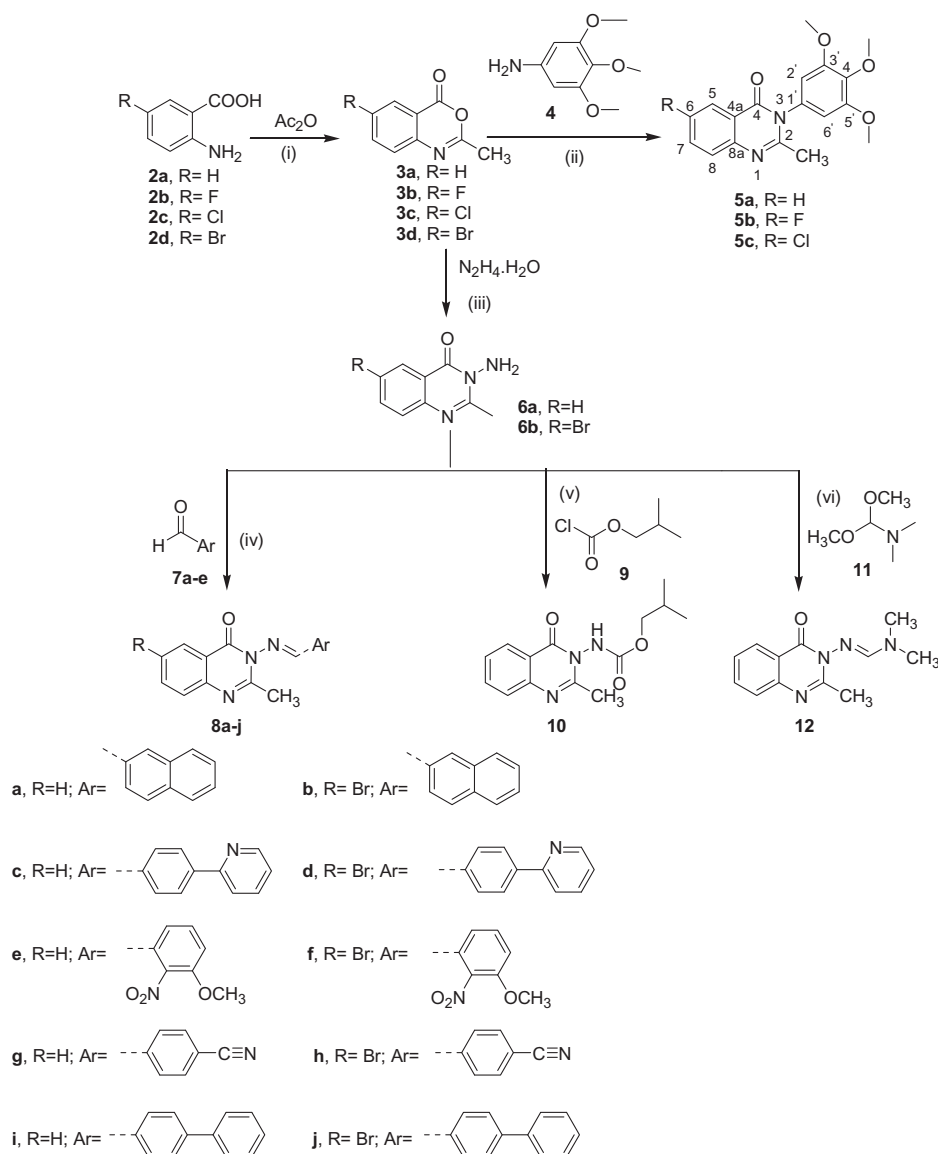
The inhibitory effects of 21 3H-quinazolin-4-one derivatives on several available therapeutically important proteases such as cathepsin-B, collagenase, elastase, thrombin, and trypsin were investigated. The inhibition of the enzymes in the presence of test compounds was evaluated by adding different concentrations of each compound (0–75 μ g/ml) to the respective reaction mixture, preincubating for 15 min and then measuring the remaining enzyme activity as previously described⁴³. The protease inhibitory activity was expressed as inhibition percentage, which was determined by comparison with a control experiment. The results were expressed as IC₅₀ values that were determined from the standard curve. A protease inhibitor cocktail (Sigma) was used as the positive control.

In vitro inhibition of α -amylase activity

The α -amylase inhibitory activity of the tested compounds was investigated according to the reported method by Subranian⁴⁴. Briefly, 10 μ l of α -amylase (3,3 U) (Sigma) was mixed with 10 μ l of different concentrations (20–200 μ g/ml) of each compound, the appropriate solvent, or quercetin (positive control), and incubated at 37 °C for 5 min. After adding 180 μ l of Labtest (amylase substrate), the samples were incubated for 8 min and the absorbance of first reaction was measured at 620 nm. Then, 100 μ l of the reaction mixture was incubated for more five additional minutes at 37 °C and the absorbance of the second reaction was measured. Labtest was diluted in distilled water (1:1) before being added to the microplate. The concentration of quercetin used was same as those of the test compounds. The α -amylase inhibition percentage was calculated as follows: % inhibition = $100 - \frac{(X2 \text{ sample} - X1 \text{ sample})}{(X2 \text{ control} - X1 \text{ control})} \times 100$ where X1 is the absorbance of the initial reading and X2 is the absorbance of the final reading. The results were expressed as IC₅₀ determined from the curve.

In vitro inhibition of α -glucosidase activity

The α -glucosidase inhibitory activity of the tested compounds was evaluated according to the method described by Andrade-Cetto and collaborators⁴⁵ based on the release of 4-nitrophenol α -D-glucopyranoside (4 NPGP). Briefly, 20 μ l of different concentrations (ranging from 0 to 50 μ g/ml) of each compound, the appropriate solvent, or quercetin (positive control) was mixed with 180 μ l of the α -glucosidase enzyme from *Saccharomyces cerevisiae* (Sigma)



Scheme 1. (i) Heating in domestic microwave at 540 watt for 8 min, cooling, and washing with petroleum ether; (ii) Fusion in oil bath at 200 °C, for 2 h, cooling to 100 °C, treatment with 37% HCl (Conc) till dissolution of the reaction mixture; cooling to room temperature and alkalisalation with dilute NaOH (25%); (iii) ethanol, reflux for 8 h; (iv) ethanol and acetic acid catalytic, reflux for 10 h; (v) reflux for 10 h; (vi) ethanol, reflux for 2 h.

and the obtained mixture was incubated at 37 °C for 2 min, followed by additional incubation for more than 15 min at 37 °C after the addition of 150 μ l of the colour reagent NPGP. The colorimetric assay included 2 U of α -glucosidase, 5 mM of 4-NPGP, and 10 mM potassium phosphate buffer, pH 6.9. The concentration of quercetin used was identical to that of the tested compounds. The reading assay was performed using a microplate reader at 405 nm. The α -glucosidase inhibition percentage was calculated as follows: % inhibition = $100 - (X2 \text{ sample} - X1 \text{ sample} / X2 \text{ control} - X1 \text{ control}) \times 100$ where X1 is the absorbance of the initial reading and X2 is the absorbance of the final reading, control is the absorbance of the assay with the appropriate solvent. The results were expressed as IC_{50} determined from the curve.

In vitro inhibition of xanthine oxidase

The xanthine oxidase inhibitory activities of the examined chemical compounds were determined by following the formation of

uric acid from xanthine according to the reported method by Bondet and co-workers⁴⁶. Forty microliters xanthine oxidase (667 mM) and 15 μ l of each compound (0–150 μ g/ml), allopurinol (positive control), or appropriate solvent (negative control) were added to each microplate well and preincubated for 5 min at 37 °C. Then, 95 μ l of reagent 1 (mixture of hydroxylamine (0.2 mM), EDTA (0.1 mM), and xanthine oxidase (667 mM) in 50 mM phosphate buffer solution (pH 7.5) were added to the reaction mixture and incubated at the same temperature for 30 min. Subsequently, the absorbance was measured at 295 nm using a microplate reader. Finally, after addition of 150 μ l of uric acid reagent, the absorbance was measured again. The concentration of allopurinol used was same as that of the tested compounds. Xanthine oxidase inhibition was calculated as follows: % inhibition = $100 - (X2 \text{ sample} - X1 \text{ sample} / X2 \text{ control} - X1 \text{ control}) \times 100$ where X1 is the absorbance of the initial reading and X2 is the absorbance of the final reading. The results were expressed as IC_{50} determined from the curve.

Results and discussion

Chemistry

The main targets of this investigation were 3*H*-quinazolin-4-one derivatives possessing general structure **1**.

Two synthetic approaches were used for the synthesis of these scaffolds.

The first synthetic approach featured the preparation and chemical transformations of the key intermediates benzo[*d*][1,3]oxazin-4-one derivatives **3a–d** to various 3*H*-quinazolin-4-one derivatives as depicted in Scheme 1.

The conventional methods available for the synthesis of compounds **3a–d** via cyclocondensation of anthranilic acid derivatives **2a–d** with acetic anhydride^{26–28} were time consuming, therefore, these compounds were synthesised using the more efficient microwave-assisted method. Fusion of oxazinones **3a–c** with 3,4,5-trimethoxyaniline **4**^{32,33} yielded 2-methyl-3-(3,4,5-trimethoxyphenyl)-3*H*-quinazolin-4-one derivatives **5a–c**. The assigned molecular structures of these new quinazolin-4-one derivatives were in full agreement with their spectroscopic and elemental analyses. Thus the IR spectrum (KBr) of compound **5c**, for example, displayed absorption bands at $\nu_{\max}$ 3075, 2934; 2839, 1680, 1598; 1504 and 834 cm^{-1} for aromatic-CH, aliphatic-CH, the carbonyl group, C=C and C-Cl, respectively. The ^1H NMR spectrum of compound **5c** revealed the presence of five aromatic protons, which were displayed as; a one proton doublet signal at δ 8.18 ppm with coupling constant *J* of 2.4 Hz corresponding to CH^5 -quinazolin-4-one, a one-proton double-doublet signal at δ 7.69 ppm with coupling constant *J* of 8.7, 2.4 Hz due to CH^7 -quinazolin-4-one, a one-proton doublet signal at δ 7.60 ppm with coupling constant *J* of 8.7 Hz corresponding to CH^8 -quinazolin-4-one, and a two-protons singlet peak at δ 6.51 ppm, which was attributed to the two methine protons of 3,4,5-trimethoxyphenyl moiety (H^2 and $\text{H}^{6'}$). The nine protons of the three methoxy groups were displayed as a three-protons singlet at δ 3.92, and a six-protons singlet at δ 3.88 ppm. Finally, the three protons of the methyl group were characterised by a three-protons singlet signal at δ 2.32 ppm. Moreover, the ^{13}C NMR spectrum also confirmed the assigned structure based on the emergence of fifteen spectral lines at δ_{C} 161.28 (CO), 154.75 (C=N), 154.25, 145.91, 138.56, 135.02, 132.95, 132.36, 128.52, 126.23, 121.73, 105.09 ($5 \times \text{CH-Ar}$ & $7 \times \text{C}_q\text{-Ar}$), 60.93 (OCH_3), 56.28 ($2 \times \text{OCH}_3$) and 24.02 (CH_3 -quinazolin-4-one). The mass spectrum (EI) of compound **5c** indicated the presence of the expected molecular ion peak [M^+] at *m/z* 360.10 (100%) for $\text{C}_{18}\text{H}_{17}^{35}\text{ClN}_2\text{O}_4$.

Refluxing of oxazinones **3a,d** with excess hydrazine hydrate in ethanol yielded 3-aminoquinazolinones **6a,b**^{29,30}, which were converted to certain novel Schiff bases **8a–j** via condensation with a series of aryl aldehydes **7a–e** in absolute ethanol containing catalytic amount of glacial acetic acid³⁴. The structures of the new Schiff's bases were verified by their IR, ^1H NMR, ^{13}C NMR, mass spectroscopic (MS) data, and CHN analyses. The IR spectra of compounds **8a–j** indicated disappearance of the stretching bands of the amino groups in 3-aminoquinazolin-4-one derivatives **6a,b**. The ^1H NMR spectra were characterised by the disappearance of the two-protons singlet signals corresponding to the amino protons of starting materials, which resonated at δ 5.79 and 4.89 ppm, accompanied by emergence of new one-proton singlet signals attributed to azomethine protons ($\text{CH}=\text{N}$) at δ 9.41–8.70 ppm range. In addition, the ^{13}C NMR spectra displayed the expected spectral lines for each compound with the signals

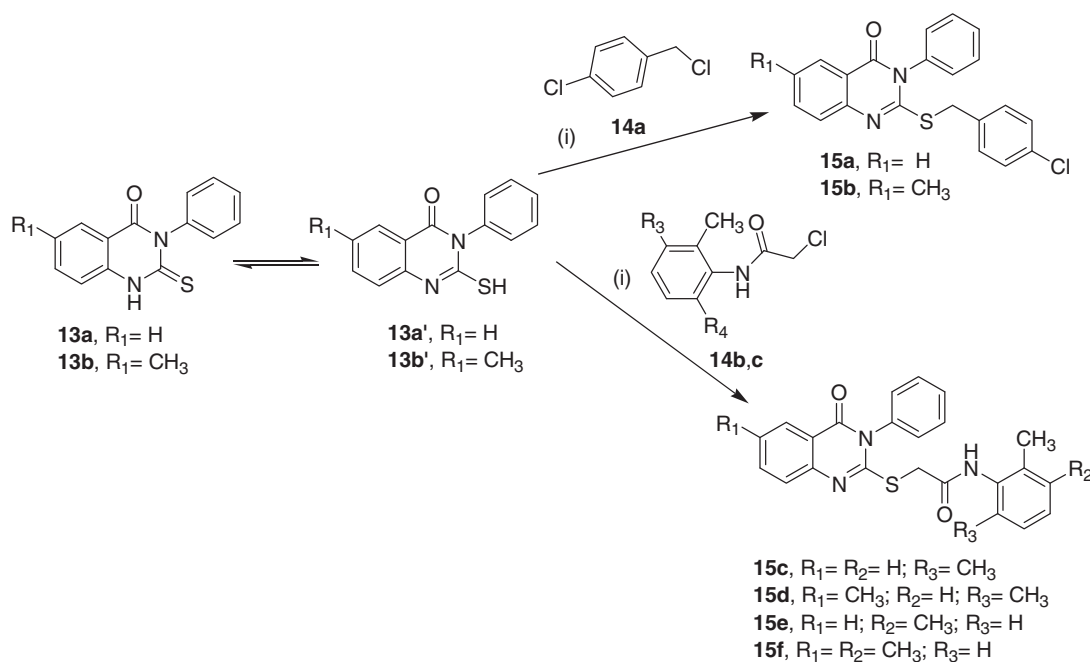
corresponding to ($\text{CH}=\text{N}$) groups resonating at δ_{C} 159.10–156.46 ppm.

Furthermore, the amino group of 3-amino-2-methyl-3*H*-quinazolin-4-one **6a** was transformed to the corresponding carbamate group via direct nucleophilic substitution reaction for isobutyl chloroformate **9**³⁵ to yield the new carbamate derivative **10**. The structure of the latter product was established on the basis of its IR, ^1H NMR, ^{13}C NMR, MS and HCN elemental analyses. The IR spectrum displayed the characteristic stretching absorption bands at $\nu_{\max}$ 3223, 1755, and 1694 cm^{-1} indicating the presence of NH and two C=O groups. The ^1H NMR spectrum of compound **10** was characterised by the presence of a one-proton broad singlet peak at δ 7.62 ppm due to NH group, in addition to the presence of a two-protons apparent singlet signal, a one-proton multiplet, and a six-protons doublet signal with coupling constant value *J* of 6.0 Hz at δ 4.00, 1.96–1.93 and 0.94 ppm, respectively corresponding to ($\text{O-CH}_2\text{-CH-(CH}_3)_2$) isobutoxy group. Moreover, the ^{13}C NMR spectrum of carbamate **10** revealed the emergence of a new spectral line at δ_{C} 156.28 ppm attributable to the C=O of the carbamate functional group. The methylene, methine and two methyl groups of isobutoxy moiety demonstrated three spectral lines at δ_{C} 72.71, 27.78, and 18.72 ppm, respectively. Finally, the mass spectrum (EI) of compound **10** showed the anticipated molecular ion peaks [$\text{M}^+ + 1$] at *m/z* 276.10 (21.00%) for $\text{C}_{14}\text{H}_{17}\text{N}_3\text{O}_3$ and [M^+] 275.10 (100.00%).

Next, 3-amino-2-methyl-3*H*-quinazolin-4-one **6a** was transformed to the corresponding *N,N*-dimethylformamidine derivative **12** by refluxing with excess *N,N*-dimethyl formamide dimethyl acetal (DMF-DMA) **11** in absolute ethanol³⁶. The structure of this new compound was deduced from its IR, $^1\text{HNMR}$, $^{13}\text{CNMR}$, MS data, and elemental analyses. The IR spectrum revealed the presence of absorption bands at $\nu_{\max}$ 3010, 2924; 2807, 1660, 1621, 1589; 1475 cm^{-1} which were attributed to CH-aromatic, CH-aliphatic, C=O, C=N, and C=C, respectively. In addition, the ^1H NMR spectrum of formamidine **12** was characterised by the presence of a new one-proton singlet signal at δ 7.74 ppm attributed to azomethine proton ($\text{HC}=\text{N}$) and two singlet signals, each integrating to three protons resonating at δ 3.08 and 3.04 ppm, corresponding to the two methyl groups in dimethylamino $\text{N}(\text{CH}_3)_2$ moiety. Furthermore, the ^{13}C NMR spectrum of compound **12** demonstrated three new peaks at δ_{C} 160.08, 40.88, and 34.53 ppm which were attributed to $\text{HC}=\text{N-N}(\text{CH}_3)_2$ and two carbons of the methyl groups in $\text{N}(\text{CH}_3)_2$ moiety, respectively. The mass spectrum (EI) of compound **12** showed the anticipated molecular ion peak [M^+] at *m/z* 230.00 (34.52%) for $\text{C}_{12}\text{H}_{14}\text{N}_4\text{O}$.

The second synthetic approach for preparation of 3*H*-quinazolin-4-one derivatives involved the synthesis of compounds **15a–f** via nucleophilic attack by 2-mercapto-3-phenyl-3*H*-quinazolin-4-one derivatives **13a',b'** on 4-chlorobenzyl chloride **14a** or chloroacetamide derivatives **14b,c**^{37,38} to yield compounds **15a–f** by adapting modified procedure to what was reported by Salman et al.³⁹ as illustrated in Scheme 2.

Structures of the new quinazolinone derivatives **15b** and **15d,f** were verified based on their respective IR, ^1H NMR, ^{13}C NMR, MS data, and elemental analyses. The IR spectra (KBr) of these products were characterised by the disappearance of the absorption bands at $\nu_{\max}$ 3245 and, 3169 cm^{-1} for the NH group (NH, SH exchangeable) and at $\nu_{\max}$ 1195 cm^{-1} for the thiocarbonyl group ($\text{C}=\text{S}$) of starting substrates **13a,b**. Furthermore, their ^1H NMR spectra indicated the disappearance of the signals at δ 13.05 and 12.99 ppm due to the thiol protons of the starting materials. In addition, the spectrum of compound **15b** showed characteristic two-protons singlet signal at δ 4.32 corresponding to the benzylic methylene



Scheme 2. Reagents and conditions: (i) K_2CO_3 and dry acetone, reflux for 6–8 h.

group. Analogously, the spectra of compounds **15d,f** were characterised by the emergence of new one-proton broad singlet signals at δ 8.70 and 8.90 ppm corresponding to NH groups, and two-protons singlet signals at δ 3.96 and, 3.97 belong corresponding to methylene groups. In addition, the two methyl groups of the acetamide moieties were represented by a six-protons singlet signal at δ 2.16 ppm for compound **15d**, and two singlet signals each integrating to three-protons at δ 2.23 and, 2.01 ppm for compound **15f**.

Furthermore, the ^{13}C NMR spectra of compounds **15b,d,f** revealed the absence of spectral lines at δ_{C} 176.05 and 176.00 ppm corresponding to the thiocarbonyl group in the starting materials **13a,b** and the presence of new peaks at δ_{C} 36.22, 35.49, and 36.14 ppm attributed to the new methylene groups. The carbonyl and methyl groups of the acetamide moieties in compounds **15d,f** were observed at δ_{C} 166.99, 167.11, 18.35, 20.55, and 13.82, respectively. In addition, the mass spectra (EI) of compounds **15b,d,f** displayed the anticipated molecular ion peaks.

Biological evaluation

Effects of the synthesised 3H-quinazolin-4-one derivatives on cell viability of human colorectal cancer cell lines HT-29 and SW620

As CRC starts with initial benign stages advanced to late stage leading to metastasis. Therefore, two human CRC cell lines which belong to two different stages namely, HT-29 which is derived from adenocarcinoma CRC and SW620 which is derived from metastatic site were selected for *in vitro* evaluation of the cytotoxicity (30 $\mu\text{g}/\text{ml}$) of twenty-one compounds incorporating the 3H-quinazolin-4-one scaffold.

Cell viability was monitored using the MTT assay and the results are presented in [Supplementary Figure 1](#). Compared to the negative control, and the positive control (5-FU), the synthesised 3H-quinazolin-4-one derivatives displayed varied degrees of cytotoxicity against the examined colorectal cell lines. Eight out of all the examined compounds showed significant decrease in cell viability. In one hand, compound **15c** inhibited only HT-29 cell line (47.5% viability) whereas compound **5b** inhibited only SW620 cell line (46.6% viability). On the other hand, six compounds namely,

8a, 8f, 8j, 15a, 15b, and 15d, were potently capable of inhibiting cell viability of both human CRC cell lines. Compound **15d** was the most active in this set reducing the cell viabilities of HT-29 and SW620 cells lines to 38% and, 36.7%, compared to 5-FU which demonstrated cell viabilities of 65.9 and 42.7%, respectively. Hence, compound **15d** was selected for further mechanistic studies. The results indicated that compound **15d** inhibited cell viability in a dose-dependent manner. The IC_{50} for **15d** was $\sim 20 \mu\text{g}/\text{ml}$ for HT-29 and SW620 cells ([Supplementary Figures 2 and 3](#)). These results were again confirmed using the xCELLigence RTCA system, which measures the cytotoxic effect of inhibitors in real time at different time points and **15d** was found to inhibit the cell proliferation in human CRC HT-29 and SW620 cell lines in dose- and time-dependent manner ([Supplementary Figures 4 and 5](#)).

Evaluation of anti-apoptotic protein expression by compound 15d

SW620 cells were treated with various concentrations of **15d** for 24 h. Subsequently, cell lysates were immunoblotted with the indicated antibodies in order to investigate the mechanism via which this compound inhibited cell viability of human CRC cells. Results shown in [Supplementary Figure 6](#) indicated that **15d** inhibited the anti-apoptotic proteins, Bcl2 and BclxL. These findings demonstrate that compound **15d** inhibited the cell viability of human CRC cells by down regulating the expression of Bcl2 and BclxL and triggering the apoptotic pathway.

In vitro evaluation of antiphospholipase A_2 (PLA_2) and antiproteases activities of the synthesised 3H-quinazolin-4-one derivatives

As inflammation is associated with several types of cancers via influencing growth, apoptosis, and proliferation of cancer and stromal cells, the synthesised 3H-quinazolin-4-one derivatives were further screened to evaluate their antiphospholipases activities against proinflammatory enzymes implicated in cancer namely, hGIIA, hGV, and X^{10} . In addition, the inhibitory activity of these compounds towards the allergen and inflammatory PLA_2 from honey bee venom (*Apis mellifera*, Am PLA_2)⁴⁷ and proinflammatory, hemolytic, myotoxic and neurotoxic Cobra (*Naja mosambique*

mosambique) venom PLA₂ (NmPLA₂)⁴⁸ were evaluated by determining their IC₅₀ values. Results (Supplementary Table 1) revealed that six compounds, namely, **5a**, **8c**, **8d**, **8j**, **15a**, and **15d** displayed the most promising potency in inhibiting the catalytic activity of the studied hG-IIA, with IC₅₀ values ranging from 5.75 ± 1.06 to 8.60 ± 0.56 µg/ml compared to that recorded with oleanolic acid which was used as a standard drug (5.25 ± 0.35 µg/ml). Amongst these compounds, **8d** exhibited the lowest IC₅₀ value (5.75 ± 1.06 µg/ml).

The anti-PLA₂ hGV data showed that three compounds; **5a**, **8d**, and **8j** inhibited the enzyme with IC₅₀ values ranging from 6.25 ± 0.78 to 9.60 ± 0.85 µg/ml compared to the value recorded with the standard drug (oleanolic acid, IC₅₀; 7.50 ± 0.71 µg/ml). Amongst them, compound **8j** (IC₅₀; 6.25 ± 0.78 µg/ml), was being more potent than the reference drug.

The analysis of the inhibitory potency of the examined compounds against PLA₂ hG-X showed that compounds; **5a**, **5b**, **5c**, **8d**, **8j**, **10**, **15a**, and **15e** inhibited the enzymatic activity with IC₅₀ values ranging from 6.60 ± 0.85 to 9.50 ± 0.71 µg/ml. It worth to note that compounds **8d**, **15a**, and **15e** exhibited lowered IC₅₀ values than the reference drug (IC₅₀; 7.55 ± 0.64 µg/ml).

Amongst the tested 3H-quinazolin-4-one derivatives which exhibited inhibitory activity against NmPLA₂ were compounds; **5a**, **5b**, **5c**, **8c**, **8d**, **8e**, **8f**, **15a**, and **15d** with IC₅₀ values ranging from 4.15 ± 0.50 to 8.50 ± 2.12 µg/ml compared to oleanolic acid (IC₅₀; 4.90 ± 0.42 µg/ml). Interestingly, compound **5b** demonstrated the lowest IC₅₀ which is also lower than that of the reference drug fustpot.

Finally, 13 compounds namely, **5a**, **5b**, **5c**, **8b**, **8c**, **8d**, **8e**, **8f**, **8i**, **10**, **15a**, **15d**, and **15e** effectively suppressed AmPLA₂ with IC₅₀ values ranging from 4.20 ± 0.57 to 9.40 ± 0.85 µg/ml compared to the IC₅₀ 4.20 ± 0.42 µg/ml value of oleanolic acid. Based on these results, AmPLA₂ appears to be the maximally affected phospholipase by the synthesised compounds with compound **8f** (IC₅₀; 4.20 ± 0.57 µg/ml) demonstrating inhibitory activity equivalent to that of the reference drug.

Similarly, the inhibitory potency of the selected 3H-quinazolin-4-one derivatives against five proteases namely, cathepsin-B, collagenase, thrombin, elastase, and trypsin was quantified by measuring and comparing the IC₅₀ values produced by these compounds relative to the value produced by the protease inhibitor cocktail. The results of anti-proteases shown in Supplementary Table 2 indicated that compounds **8j** and **15e** were being the most active inhibitors against protease cathepsin-B with IC₅₀ values of 1.40 ± 0.06 and 1.60 ± 0.07 µg/ml respectively compared to cocktail (IC₅₀; 0.175 ± 0.04 µg/ml).

Furthermore, compounds **8b**, **8d**, and **15e** were found to be the most active in suppressing the collagenase activity with IC₅₀ values ranging from 3.00 ± 0.01 to 4.50 ± 0.71 µg/ml compared to cocktail (IC₅₀; 0.15 ± 0.07 µg/ml) with compound **8d** exhibiting the lowest IC₅₀ value.

The highest anti-thrombin activity was demonstrated by compounds **5b**, **5c**, **8e**, **10**, and **15b**, with IC₅₀ values ranging from 0.90 ± 0.14 to 2.50 ± 0.71 µg/ml, with **5b** demonstrating the lowest IC₅₀ compared to cocktail (IC₅₀; 0.25 ± 0.07 µg/ml).

The highest anti-elastase activity was demonstrated by compounds **5b**, **5c**, **8e**, **10**, and **15b**, with IC₅₀ values ranging from 0.65 ± 0.02 µg/ml (compound **15b**) to 1.75 ± 0.05 µg/ml (compound **5c**) compared to that of cocktail (IC₅₀; 0.125 ± 0.04 µg/ml).

Finally, the lowest IC₅₀ values ranging from 0.50 ± 0.00 to 2.25 ± 0.04 µg/ml against trypsin were demonstrated by compounds **5b**, **5c**, **8e**, **10**, and **15b**, with compound **15b** displaying

the lowest IC₅₀ compared to that of the reference drug (IC₅₀; 0.215 ± 0.04 µg/ml).

In vitro evaluation of inhibitory activities of 3H-quinazolin-4-one derivatives against enzymes implicated in metabolic syndrome

Considering an interactive relationship between diabetes and cancers, the *in vitro* inhibitory potential of the prepared 3H-quinazolin-4-one derivative against α -amylase, α -glucosidase, and xanthine oxidase (enzymes related to metabolic syndrome) was investigated.

The IC₅₀ values exhibited by the chemical compounds against α -amylase activity are shown in Supplementary Table 3, which indicated that compounds **8d**, **8h**, **8j**, **15a**, **15b**, **15e**, and **15f**, were more potent in suppressing the enzyme with IC₅₀ values ranging from 94.00 ± 2.83 µg/ml (compound **15b**) to 121.00 ± 9.90 µg/ml (compound **15e**) than quercetin the reference drug (IC₅₀ 123.0 ± 2.83 µg/ml).

In addition, compounds **8g** (IC₅₀ 3.25 ± 0.35 µg/ml), **8h** (IC₅₀ 3.05 ± 0.21 µg/ml), and **8i** (IC₅₀ 3.45 ± 0.64 µg/ml) were more potent in inhibiting α -glucosidase activity than quercetin (IC₅₀ 3.80 ± 0.28 µg/ml) (Supplementary Table 4).

Although none of the tested compounds was more effective than allopurinol (reference drug) in inhibiting xanthine oxidase (XO) activity (Supplementary Table 5) (IC₅₀ 0.65 ± 0.07 µg/ml), compound **8f** (IC₅₀ 3.00 ± 1.41 µg/ml) was the most active among the tested compounds.

Conclusions

A series of 25 compounds incorporating the 3H-quinazolin-4-one scaffold were synthesised, 20 of which were new and fully characterised. Twenty-one compounds were screened for their cytotoxicity on human CRC cell lines HT-29 and SW620 using the MTT assay. Six compounds namely, **8a**, **8f**, **8j**, **15a**, **15b**, and **15d** potentially reduced viability of both cell lines, with **15d** the most active in this set displaying residual viability 38%, 36.7% against HT-29 and SW620 cells lines, respectively. Further mechanistic studies using **15d** revealed that this compound inhibited the cell proliferation of HT-29 and SW620 cell lines in a dose- and time-dependent manner with IC₅₀ values ~20 µg/ml. Evaluation of anti-apoptotic protein expression by compound **15d** revealed that it reduced the viability of the used human CRC cells by down regulating the expression of Bcl2 and BclxL.

Based on these results, **15d** could be considered as an attractive candidate for further investigations as a potential anticancer agent. Furthermore, the prepared compounds were screened for their inhibitory activities against certain phospholipases including, hG-IIA, hG-V, hG-X, NmPLA₂, and AmPLA₂, and five proteases namely, cathepsin-B, collagenase, thrombin, elastase, and trypsin, in addition to three enzymes involved in metabolic syndrome namely, α -amylase, α -glucosidase, and xanthine oxidase. Although none of the tested compounds showed good antiprotease activity, six compounds **8j**, **8d/15a/15e**, **5b**, and **8f** demonstrated higher antiphospholipases potential against proinflammatory isoforms hGV, hG-X, NmPLA₂ and AmPLA₂ than oleanolic acid, which was used as reference drug.

Therefore, these compounds would be of interest as preclinical candidates in trials for anticancer and anti-inflammatory drugs.

All the tested compounds were inactive against xanthine oxidase, whereas compounds **8d**, **8h**, **8j**, **15a**, **15b**, **15e**, and **15f** elicited higher α -amylase inhibition than the reference drug quercetin. Furthermore, compounds **8g**, **8h**, and **8i** were more

potent in inhibiting α -glucosidase activity than the reference drug allopurinol. Consequently, these compounds can be considered as potential targets that can be further investigated for developing novel anti-diabetes mellitus (DM) drugs.

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Disclosure statement

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References

1. Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015;136:E359–86.
2. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000;100:57–70.
3. Adams JM, Cory S. The Bcl-2 apoptotic switch in cancer development and therapy. *Oncogene* 2007;26:1324–37.
4. Vogler M, Dinsdale D, Dyer MJS, Cohen GM. Bcl-2 inhibitors: small molecules with a big impact on cancer therapy. *Cell Death Differ* 2009;16:360–7.
5. Soreide K, Janssen EA, Korner H, Baak JPA. Trypsin in colorectal cancer: molecular biological mechanisms of proliferation, invasion and metastasis. *J Pathol* 2006;209:145–56.
6. Rakashanda S, Rana F, Rafiq S, et al. Role of proteases in cancer: a review. *Biotechnol Mol Biol Rev* 2012;7:90–101.
7. Zucker S, Vacirca J. Role of matrix metalloproteinases (MMPs) in colorectal cancer. *Cancer Metastasis Rev* 2004;23:101–17.
8. Nierodzik ML, Karparkin S. Thrombin induces tumor growth, metastasis, and angiogenesis: evidence for a thrombin regulated dormant tumor phenotype. *Cancer Cell* 2006;10:355–62.
9. Ho AS, Chen CH, Cheng CC, et al. Neutrophil elastase as a diagnostic marker and therapeutic target in colorectal cancers. *Oncotarget* 2014;5:473–80.
10. Dennis EA, Cao J, Hsu YH, et al. Phospholipase A2 enzymes: physical structure, biological function, disease implication, chemical inhibition, and therapeutic intervention. *Chem Rev* 2011;111:6130–85.
11. Luo C, Zhang H. The role of proinflammatory pathways in the pathogenesis of colitis-associated colorectal cancer. *Mediators Inflamm* 2017;2017:1.
12. Vulcan A, Manjer J, Ohlsson B. High blood glucose levels are associated with higher risk of colon cancer in men: a cohort study. *BMC Cancer* 2017;17:842–9.
13. Ojo OA, Afon AA, Ojo AB, et al. Inhibitory effects of solvent-partitioned fractions of two nigerian herbs (spondias mombin linn. and mangifera indica L.) on α -amylase and α -glucosidase. *Antioxidants* 2018;7:73.
14. Li X, Meng X, Gao X, et al. Elevated serum xanthine oxidase activity is associated with the development of type 2 diabetes: a prospective cohort study. *Diabetes Care* 2018;41:88–90.
15. Kshirsagar UA. Recent developments in the chemistry of quinazolinone alkaloids. *Org Biomol Chem* 2015;13:9336–52.
16. Tiwary BK, Pradhan K, Nanda AK, Chakraborty R. Implication of quinazoline-4 (3H)-ones in medicinal chemistry: a brief review. *J Chem Biol Ther* 2015;1:1000104.
17. Vavsari VF, Ziarani GM. Synthesis of 4-quinazolinones by transition metal-catalyzed processes (microreview). *Chem Heterocycl Comp* 2018;54:317–9.
18. Ghorab MM, Alsaid MS, Soliman AM, Al-Mishari AA. Benzo[g]quinazolin-based scaffold derivatives as dual EGFR/HER2 inhibitors. *J Enzyme Inhib Med Chem* 2018;33:67–73.
19. Mekala N, Buddepu SR, Dehury SK, et al. A novel strategy for the manufacture of idelalisib: controlling the formation of an enantiomer. *RSC Adv* 2018;8:15863–9.
20. Seema R, Pathak Malhotra V, Nath R, et al. Synthesis and antihypertensive activity of novel quinazolin-4(3H)-one derivatives. *Cent Nerv Syst Agents Med Chem* 2014;14:34–8.
21. Al-Omar MA, El-Azab AS, El-Obeid HA, Abdel Hamide SG. Synthesis of some new 4-(3H)-quinazoline analogs as potential antioxidant agents. *J Saudi Chem Soc* 2006;10:113–30.
22. Jain N, Jaiswal J, Pathak A, Singour PK. Synthesis, molecular docking and evaluation of 3-{4-[2-amino-4-(substituted-phenyl)-2H-[1, 3] oxazin/thiazin-6-yl] 2-phenyl-3H-quinazolin-4-one derivatives for their anticonvulsant activity. *Cent Nerv Syst Agents Med Chem* 2018;18:63–73.
23. Osarumwense PO, Edema MO, Usifoh O. Synthesis and anti-inflammatory activity of 4(3H)-quinazolinone and its 2-methyl and 2-phenyl-4 (3H)- quinazolinone derivatives. *IOSR J Appl Chem* 2018;11:12–5.
24. El-Sayed NN, Alafeefy AM, Bakht MA, et al. Synthesis, anti-phospholipase A2, antiprotease, antibacterial evaluation and molecular docking analysis of certain novel hydrazones. *Molecules* 2016;21:1664–80.
25. El-Sayed NN, Al-Balawi NA, Alafeefy AM, et al. Synthesis, characterization and antimicrobial evaluation of some thiazole-derived carbamates, semicarbazones, amides and carboxamide. *J Chem Soc Pak* 2016;38:358–68.
26. Nagase T, Mizutani T, Ishikawa S, et al. Synthesis, structure-activity relationships, and biological profiles of a quinazolinone class of histamine H3 receptor inverse agonists. *J Med Chem* 2008;51:4780–9.
27. Nagase T, Mizutani T, Sekino E, et al. structurally constrained quinazolinone derivatives as potent and selective histamine H3 receptor inverse agonists. *J Med Chem* 2008;51:6889–901.
28. Kumar S, Kaur H, Singh I, et al. Synthesis, characterization and biological activity of various substituted quinazolinone derivatives containing dopamine moiety. *World J Chem* 2009;4:195–200.
29. Kumar P, Shrivastava B, Stables JP, Pandeya SN. Design, synthesis and potential 6 Hz psychomotor seizure test activity of some novel 2-(substituted)-3-[[substituted]amino]quinazolin-4(3H)-one. *Eur J Med Chem* 2011;46:1006–18.
30. Kumar A, Singh S, Saxena AK, Shanker K. Synthesis of quinazolinyl pyrimidinediones and their anti-inflammatory activity. *Indian J Chem* 1988;27B:443–7.
31. Kim H, Lee HJ, Kim DP. Integrated one-flow synthesis of heterocyclic thioquinazolinones through serial microreactions

- with two organolithium intermediates. *Angew Chem Int Ed Engl* 2015;54:1877–80.
32. van Zyl EF. A survey of reported synthesis of methaqualone and some positional and structural isomers. *Forensic Sci Int* 2001;122:142–9.
 33. Soliman FSG, Shafik RM, Elnenaey EA. Synthesis of methaqualone and its diphasic titration in pure and tablet forms. *J Pharm Sci* 1978;67:411–3.
 34. Rajasekaran S, GopalKrishna R, Sanjay PPN. Gurpreet SS. Synthesis, antibacterial and *in vitro* antioxidant activity of 2,3-substituted quinazolin-4(3*H*)-ones. *J Chem Pharm Res* 2010;2:482–8.
 35. El-Baih FEM, Al-Blowy HAS, Al-Hazimi HM. Synthesis of some thienopyrimidine derivatives. *Molecules* 2006;11:498–513.
 36. Dineen TA, Zajac MA, Myers AG. Efficient transamidation of primary carboxamides by in situ activation with *N,N*-dialkylformamide dimethyl acetals. *J Am Chem Soc* 2006;128:16406–9.
 37. Reilly TJ. The preparation of lidocaine. *J Chem Educ* 1999;76:1557.
 38. Lofgren NM, Lundquist BJ. Alkyl glycinanilides, Patent 2441498A, USA; 1948.
 39. Salman AS, Abdel-Aziem A, Alkubbat MJ. Design, synthesis of some new thio-substituted imidazole and their biological activity. *Am J Org Chem* 2015;5:57–72.
 40. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assay. *J Immunol Methods* 1983;65:55–63.
 41. Al-Khayal K, Alafeefy A, Vaali-Mohammed MA, et al. Novel derivative of aminobenzenesulfonamide (3c) induces apoptosis in colorectal cancer cells through ROS generation and inhibits cell migration. *BMC Cancer* 2017;17:4–15.
 42. Lôbo de Araújo A, Radvanyi F. Determination of phospholipase A₂ activity by a colorimetric assay using a pH indicator. *Toxicon* 1987;25:1181–8.
 43. Kunitz M. Crystalline soyabean trypsin inhibitor: II. General properties. *J Gen Physiol* 1947;30:291–310.
 44. Subramanian R, Asmawi MZ, Sadikun A. In vitro alpha-glucosidase and alpha-amylase enzyme inhibitory effects of *Andrographis paniculata* extract and andrographolide. *Acta Biochim Pol* 2008;55:391–8.
 45. Andrade-Cetto A, Becerra-Jiménez J, Cardenas VR. Alfa-glucosidase-inhibiting activity of some Mexican plants used in the treatment of type 2 diabetes. *J Ethnopharmacol* 2008;116:27–32.
 46. Bondet V, Brand-Williams W, Berset C. Kinetics and mechanisms of antioxidant activity using the DPPH• free radical method. *LWT Food Sci Technol* 1997;30:609–15.
 47. Zolfagharian H, Mohajeri M, Babaie M. Honey bee venom (*Apis mellifera*) contains anticoagulation factors and increases the blood-clotting time. *J Pharmacopuncture* 2015;18:7–11.
 48. Wacklin H. Interfacial mechanism of phospholipase A₂: pH-dependent inhibition and ME-B-cyclodextrin activation. *Biochemistry* 2009;48:5874–81.