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Research Article

Synthesis and characterization of new ^{99m}Tc-radiopharmaceuticals with dithiobenzoate derivatives for the study of septic inflammatory processes

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Summary

Improved methods for the preparation of ^{99m}Tc -radiopharmaceuticals containing dithiobenzoate ligands, in sterile and pyrogen free conditions, are described. These procedures are based on the reaction of these ligands either with $[^{99m}Tc]$ pertechnetate in the presence of a strong reducing agent (HCl/tertiary phosphine, $SnCl_2 \cdot 2H_2O)$, or with pre-reduced complexes obtained from different kit formulations. All the preparations led to the high-yield formation of the neutral and lipophilic ^{99m}Tc -complex $[^{99m}Tc][Tc(S_3CPh)_2(S_2CPh)]$, which is analogous to the corresponding compounds obtained with rhenium and the long-lived β -emitting isotope technetium-99 g recently described. HPLC analysis and thin layer chromatography were used to confirm the characterisation of the resulting ^{99m}Tc -radiopharmaceutical which was found to be potentially suitable for blood-cell labelling as applied to the

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Introduction

Nuclear medicine is an area of research having a specific interest in the design, synthesis and clinical application of new radiopharmaceuticals for diagnosis and therapy. 1 In particular, there is a growing interest for the development of new tracers for blood-cell labelling which may be useful in the study of the mechanism of immune response or to visualize inflammatory lesions.² This approach has been successfully applied during the last 20 years using 111 In-oxine and 99m Tc-HMPAO (HMPAO = hexamethylpropyleneamine oximate).³ However, these well-known blood-cell labelling agents suffer from various disadvantages, the most significant being their lack of selectivity, which necessarily requires a first separation step of the target blood cells before labelling. Thus, current efforts are focused on the design and the preparation of more specific blood-cell labelling agents. In our laboratory, we found that the class of nitrido technetium-99m complexes with dithiocarbamate ligands showed promising results for this purpose. Specifically, the complex bis-(N-ethoxy, N-ethyldithiocarbamate)nitridotechnetium(V) [99mTcN(NOET)2] allows in vitro specific polynuclear cells labelling in the whole blood.⁴ whereas similar disubstituted complexes with dithiocarboxylates having aliphatic linear side-chains induce a high lymphocyte selectivity in the same conditions.⁵ [99mTcN(NOET)₂]⁶ is an example of a large class of Tc-99m complexes, including also xanthates⁷ and dithiocarboxylates,⁸ characterised by the presence of a terminal Tc≡N multiple bond. It was found that [99mTcN(NOET)2] exhibits high myocardial localization both in animals and in humans, and it is currently under phase III clinical trials for receiving final approval as heart imaging agent.⁹ The efficient preparation of the Tc≡N group, at no carrier-added level and under sterile and pyrogenic free conditions, has been recently made possible after the introduction by Pasqualini et al. 10 of a suitable kit formulation based on the initial reaction of pertechnetate ($[^{99}\text{m}Tc]TcO_4^-$) with Nmethyl-S-methyl dithiocarbazate $[MDTCZ = H_2N-N(Me)C(=S)SMe]$ or, alternatively, with succinic dihydrazide $[SDH = H_2N-NH-C(=O) (CH_2)_2$ – $(O =)C-NH-NH_2]$ as sources of nitride nitrogen atoms, in the presence of a reducing agent such as SnCl₂·2H₂O or HCl and tertiary phosphines. The initial step is then followed by the addition of the appropriate ligand to afford the final radiopharmaceutical having the nitrido core.

In this paper, we report the synthesis of the complex $[^{99m}\text{Tc}(S_3\text{CPh})_2(S_2\text{CPh})]$ using different kit methods and by varying the nature of the counter ion in the starting ligand PhCS $_2^-\text{X}^+$ <u>L1–L4</u> [X=MgBr, Na, NH $_2$ (CH $_2$) $_5$, NMe $_4$]. The preparation and characterization of the complexes [M(S $_3$ CPh) $_2$ (S $_2$ CPh)] (M= 99g Tc, Re) in macroscopic amounts, using the long-lived isotope Tc-99 g and the stable rhenium congener, has been recently described by our group. These latter species have been utilized here as reference compounds to obtain the full elucidation of the chemical nature of the new 99m Tc-radio-pharmaceutical by HPLC analysis and thin layer chromatography.

Experimental

Materials

Tetrahydrofuran (THF) was distilled twice under nitrogen from sodium/benzophenone before use, while diethyl ether, dichloromethane and petroleum ether were distilled using the same procedure from calcium chloride. Bromobenzene, magnesium turnings, carbon disulfide, tetramethylammonium hydroxyde, piperidine, triphenylphosphine and stannous chloride dihydrate were commercially available (Aldrich, St Quentin Fallavier, France) and used as purchased. Sodium tris (m-sulfonatophenyl)phosphine $[P(m-C_6H_4SO_3)_3]Na_3$ (TPPTS) was synthesized as reported previously. ¹² Sodium [99mTc]pertechnetate, in physiological solution, was obtained from a ⁹⁹Mo/^{99m}Tc generator ELU III (CIS bio international/Schering SA, Gif sur Yvette, France). All other chemicals were of laboratory grade and used without further purification. 1,2-diaminopropane-*N-N-N'-N'*-tetraacetic acid (PDTA) and kits for the preparation of the Tc≡N group (99mTc-MDTCZ and ^{99m}Tc-SDH) and of ^{99m}Tc-Gluconate were generously offered by CIS bio international/Schering SA (Gif-sur-Yvette, France).

Ligands

The magnesium bromide salt, $PhCS_2^-MgBr^+$ <u>L1</u>, was prepared in anhydrous THF as reported previously, ¹³ and isolated by filtration at

Scheme 1. Synthesis of dithiobenzoic ligands

Table 1. Ligand synthesis

| Ligands | Conter ion | Rfª | M.p. (°C) | yield(%) |
|----------------------|------------------|------|------------------|----------|
| L1 | MgBr | 0.85 | Oxidation in air | 82 |
| $\overline{L2}$ | Na | 0.81 | 92-94 | 94 |
| L3 | $H_2N(CH_2)_5$ | 0.83 | 96-98 | 76 |
| L1 L2 L3 L4 | NMe ₄ | 0.74 | 98-100 | 52 |

a eluent : acetone

−15°C, washed with cold diethyl ether and dried under reduced pressure. The tetramethylammonium salt L4, PhCS₂-NMe₄⁺, was precipitated by addition of an equivalent amount of tetramethylammonium hydroxide to a pink solution of dithiobenzoic acid, under vigorous stirring, and then isolated by filtration, washed with diethyl ether and dried under reduced pressure. PhCS₂H was previously obtained by addition of 20 ml of HCl (1 mol⁻¹) to the reaction solution containing the magnesium bromide salt at 0°C and then isolated after extraction with diethyl ether. The sodium salt L2, PhCS₂-Na⁺, was prepared using a similar procedure by addition of an equivalent amount of aqueous sodium hydroxide (1.25 mol dm⁻³), and precipitated after concentration of the resulting solution. Similarly, the piperidinium L3, salt PhCS₂(C₅H₁₀)NH₂⁺, was precipitated from PhCS₂H, by addition of an equivalent amount of piperidine, and then isolated by filtration, washed with diethyl ether and dried under vacuum. The preparations of the various salts of the ligand PhCS₂ are summarized in Scheme 1. Analytical data are reported in Tables 1–3. All the products were stored at -12°C under a nitrogen atmosphere.

Table 2. ¹H NMR and ¹³C NMR of ligands

| Ligands (solvent) | ¹ H NMR [ppm, peak multiplicity, (number of hydrogens)] ^a | | | | |
|---|---|--|--|--|--|
| $\underline{\mathbf{L1}}$: dmso-d ⁶ | Aromatic hydrogens: 7.19, t, $J = 7.6 \text{ Hz}(2 \text{ H})$ 7.29, t, $J = 7.6 \text{ Hz}(1 \text{ H})$ | | | | |
| $\underline{\mathbf{L2}}: \mathrm{D_2O}$ | 8.15, d, $J = 7.6 \text{ Hz}(2 \text{ H})$ Aromatic hydrogens: 7.28, t, $J = 7.1 \text{ Hz}$ (2 H) 7.39, t, $J = 7.1 \text{ Hz}$ (1 H) 7.89, d, $J = 7.1 \text{ Hz}$ (2 H) | | | | |
| $\underline{\mathbf{L3}}$: CDCl ₃ | CH ₂ : p, $J = 5.0 \text{ Hz}(2 \text{ H})$ CH ₂ : p, $J = 6.0 \text{ Hz}(4 \text{ H})$ NCH ₂ : 3.22, s (12 H) | | | | |
| $\underline{\mathbf{L4}}$: dmso-d ⁶ | Aromatic hydrogens: 7.35, t, J =7.5 Hz(2 H) 7.46, t, J =7.0 Hz(1 H) 8.46, d, J =7.5 Hz(2 H) NH: 8.12,s(1 H) NCH ₃ : 3.10, s, 12 H Aromatic hydrogens: 7.14, t, J =7.1 Hz(2 H) 7.29, t, J =7.1 Hz(1 H) 8.15, d, J =7.1 Hz(2 H) | | | | |
| $\begin{array}{l} \underline{\textbf{L1}}: dmso\text{-}d^6 \\ \underline{\textbf{L2}}: D_2O \\ \underline{\textbf{L3}}: CDCl_3 \end{array}$ | ¹³ C NMR (ppm) Aromatic carbons: 125.8, 125.9, 128.5, 152.7 CS ₂ : 231.0 Aromatic carbons: 126.3, 127.9, 130.6, 153.9 CS ₂ : 228.4 CH ₂ : 22.2; 22.8 NCH ₂ : 45.4 Aromatic carbons: 126.9, 127.3, 130.8, 150.8 CS ₂ : 254.8 | | | | |
| $\underline{\mathbf{L4}}$: dmso-d ⁶ | NCH ₃ : 54.3 Aromatic carbons: 126.2, 126.3, 128.0, 152.9 CS ₂ : 250.2 | | | | |

 $^{^{}a}$ m = multiplet, t = triplet, d = doublet and p = pentuplet

Methods

¹H and ¹³C NMR spectra were recorded in CDCl₃, D₂O or dmso-d⁶ solutions on a Bruker ARX 400 instrument. Values for the chemical shift were determined using CHCl₃ as a reference. Elemental analysis of C, H and S were performed by I.C.S.N. (Gif-sur-Yvette, France) on a Carlo Erba 1106 analyser. FT IR spectra were recorded on a Nicolet 205 instrument in KBr pellets within the range 4000–500 cm⁻¹.

Radiochemical purity (RCP) of radioactive compounds was measured by thin-layer chromatography (TLC) on aluminium-backed silicagel plates (F_{254} , Merck) using a mixture of petroleum ether and dichloromethane (60:40) as mobile phase. Quantitative evaluation of radioactivity profiles was obtained by applying the following method. After development of the chromatogram, the plates were dried under an air stream and protected with an adhesive tape to avoid contamination. The chromatograms were then placed in close contact with a Fuji imaging plate (BAS-IIIS), in a dark box, for 5–10 min depending on the initial activity. Location of the radioactivity as a dark spot was accomplished with a Fujix Bas 1000 bio-imaging analyser (Fujix Bas 1000). The original chromatographic strip was then cut in correspondence of each spot and radioactivity was measured with a Kontron GAMMAMATIC γ -counter. RCP values were calculated as the ratio of migrated radioactivity to the total radioactivity on the strip.

| Table 3. Elemental analysis and Infrared of dithiobenzoate ligand | Table 3. | Elemental | analysis | and | Infrared | of | dithiobenzoate | ligands |
|---|----------|-----------|----------|-----|-----------------|----|----------------|---------|
|---|----------|-----------|----------|-----|-----------------|----|----------------|---------|

| Ligands | As | sayed valu | ies (%) | Cal | Calculated values (%) | | | |
|---|---|------------|---------|-----|-----------------------|---|--|--|
| | С | Н | S | С | Н | S | | |
| $\begin{array}{ c c }\hline \underline{L1}: C_7H_5BrMgS_2\\ \underline{L2}: C_7H_5NaS_2\\ \underline{L3}: C_{12}H_{15}NS_2\\ \underline{L4}: C_{11}H_7NS_2\\ \end{array}$ | 32.54 2.12 24.81 32.70 2.00 24.90 47.73 2.92 36.38 47.70 2.90 36.40 60.17 7.16 26.71 60.20 7.16 26.70 58.34 7.54 27.13 58.10 7.50 28.20 | | | | | | | |
| | Infrared (KBr pellets) : cm ⁻¹ (intensity) | | | | | | | |
| <u>L1</u> | 1609(m), 1444(m), 1304(w), 1216(s), 1173(m), 1077(m), 1008(s, | | | | | | | |
| <u>L2</u> | v_{C-S}), 980(s), 967(m), 904(s), 757(s), 683(s), 653(m). 1629(s), 1438(m), 1316(w), 1223(s), 1168(m), 1077(m), 1014(s, v_{C-S}), 910(m), 906(m), 765(s), 692(m), 655(m) | | | | | | | |
| <u>L3</u> | 2955(w), 1584(f), 1443(m), 1384(w), 1305(w), 1263(w), 1208(m), 1177(w), 1082(m), 1004(s, v_{C-S}), 988(s), 944(w), 926(w), 906(w), | | | | | | | |
| <u>L4</u> | 767(m), 694(m), 655(m) 1487(s), 1437(m), 1403(w), 1216(w), 1168(m), 1077(m), 1014(s, v_{C-S}), 956(s), 948(s), 906(m), 759(s), 694(m) | | | | | | | |

The complexes $[M(S_3CPh)_2(S_2CPh)]$ $(M = {}^{99m}Tc, {}^{99g}Tc$ and Re) chromatograms were performed by HPLC (System Gold, Beckman), equiped with a Spectra-Physics SP 8800 ternary pump. Injections were performed automatically with a Spectra-Physics autosampler with 20-µl Rheodine injector. Elutions were run in the isocratic mode on a ODS Beckman Column $(4.6 \times 250 \,\mathrm{mm}, \,\mathrm{d.p.} \,5\,\mu\mathrm{m})$ equiped with a ODS precolumn $(4.6 \times 45 \,\text{mm}, \,\text{dp} \,\,5\,\mu\text{m})$ using He-degassed tetrahydrofuran (THF)/H₂O gradient (0-3 min 70% H₂O; 3.1-17 min 100% THF; 17.1–30 min 70% H₂O). The flow was 1 ml/min and the injection volume varied between 10 and 100 µL depending on the activity containing in the vial. The detector consist of a ABI Kratos UV-Vis detector (Spectroflow 783) coupled to a 10-µl loop flow-through gamma detector (LB2040 Berthold Spectrometer). Output signals were analysed by a Spectra-Physics SP4290 dual-channel integrator. The program controlling the apparatus is from Beckman (System Gold Package). Retention time differences (16.1 and 15.6 min) between gamma emitting compounds and UV-Vis complexes is due to the answer delay resulting from the coupling through a loop-flow of two different detectors.

Elution yields of the different radioactive entities were satisfactory (>95%), thus indicating that these compounds were not retained in the column or the HPLC system (Table 4).

[^{99m}TcO₄] $\int_{0.01}^{99} Tc(S_3CPh)_2$ $\int_{0.01}^{99} Tc(S_3CPh)_2$ $[Re(S_3CPh)_2(S_2CPh)]$ (S_2CPh) (S_2CPh) RCP (%)a >95 >95 >90 >90 $R_{\rm t} \, ({\rm min})^{\rm b}$ 3.44 16.1 15.6 15.6 $R_{\rm f}^{\ c}$ 0.62 0.62 0.62

Table 4. HPLC and TLC analysis of $[M(S_3CPh)_2(S_2CPh)]$ (M = ^{99m}Tc , ^{99g}Tc , $^{185/187}$ Re)

Preparation of $\lceil^{99m}Tc(S_2CPh)(S_3CPh)_2\rceil$

The title compound was prepared by reacting the dithiobenzoate ligand DTCX [PhCS₂X <u>L1</u>–<u>L4</u>, X = MgBr, NMe₄, Na (C₅H₁₀)NH₂] with two different kit formulations for producing the Tc \equiv N triple bond at carrier-free level which have been described previously.^{6, 9, 10, 14} The two formulations, ^{99m}Tc-MDTCZ and ^{99m}Tc-SDH, differ in the nature of the donor of N³⁻ groups. In ^{99m}Tc-MDTCZ this reagent is *N*-methyl *S*-methyl dithiocarbazate [MDTCZ=H₂N–N(Me)C(=S)SMe] while in ^{99m}Tc-SDH it is succinic dihydrazide [SDH=H₂N.NH–C(=O)–(CH₂)₂–(O=)C–NH–NH₂].

 $^{99m}Tc\text{-}MDTCZ$. [^{99m}Tc]Pertechnetate (0.4–0.8 GBq) was added to a vial containing 1.0 mg MDTCZ, 0.1 mg of $SnCl_2 \cdot 2H_2O$ and 10.0 mg of PDTA in a freeze-dried form. The mixture was heated at $100^{\circ}C$ for 30 min and then cooled to room temperature. 20.0 mg of the appropriate dithiobenzoate ligand DTCX, dissolved in 1.0 mL of physiological serum, were finally added and the solution was heated at $100^{\circ}C$ for 45 min.

 $^{99m}Tc-SDH$. [99m Tc]Pertechnetate (0.4–0.8 GBq) was added to a vial containing 5.0 mg of SDH, 0.1 mg of SnCl₂·2H₂O and 5.0 mg of PDTA in a freeze-dried form. The mixture was stirred at room temperature for 30 min. 20.0 mg of the appropriate DTCX ligand, dissolved in 1.0 mL of physiological serum were finally added, and the solution was heated at 100° C for 45 min.

The following procedures have been also used to prepare the same complex [$^{99m}Tc(S_2CPh)(S_3CPh)_2$] using different reducing agents and from the commercial kit ^{99m}Tc -Gluconate. 15

^aChromatographic purity determined by HPLC analysis.

^b retention time.

^cTLC analysis on SiO₂ normal plates eluated with CH₂Cl₂/petroleum ether 30/70.

 PPh_3/HCl . [99m Tc]Pertechnetate (0.4–0.8 GBq) was added to a vial containing 0.2 ml of an ethanolic solution of triphenylphosphine ($2 \times 10^{-2} \, \text{mol dm}^{-3}$), 0.2 mL of aqueous HCl (0.1 mol dm $^{-3}$) and 0.6 ml of physiological serum. The mixture was vortexed and then heated at 100° C for 15 min. 10.0 mg of the appropriate ligand DTCX, dissolved in 1.0 ml of physiological serum, were finally added to the hot solution and the heating was continued for 30 min.

TPPTS/HCl. [^{99m}Tc]Pertechnetate (0.4–0.8 GBq) was added to a vial containing 0.2 mL of an aqueous solution of TPPTS (2×10^{-2} mol dm⁻³), 0.2 mL of aqueous HCl (0.1 mol dm⁻³) and 0.6 mL of physiological serum. The mixture was vortexed and then heated at 100° C for 15 min. 10.0 mg of the appropriate ligand DTCX, dissolved in 1.0 ml of physiological serum, were finally added to the hot solution and the heating was further continued for 30 min.

 $SnCl_2$. [99mTc]Pertechnetate (0.4–0.8 GBq) was added to a vial 0.1 mg of $SnCl_2 \cdot 2H_2O$ and 10 mg of 1,2-diaminopropane-N-N-N'-N'-tetraacetic acid (PDTA). The mixture was vortexed and then heated at $100^{\circ}C$ for 15 min. 20.0 mg of the appropriate DTCX ligand, dissolved in 1.0 mL of physiological serum, were finally added to the hot solution and the heating was further continued for 45 min.

^{99m}Tc-Gluconate. [^{99m}Tc]Pertechnetate (0.4–0.8 GBq) was added to a vial containing 75.0 mg of calcium gluconate dissolved in 0.1 ml of water, 0.75 mg of SnCl₂2H₂O and 25.0 mg of sodium chloride. The mixture was stirred for 10 min at room temperature. 20.0 mg of the appropriate DTCX ligand, dissolved in 1.0 mL of physiological serum, were finally added and the solution was heated at 100°C for 15 min.

Results and discussion

Ligands

Various salts of dithiobenzoic acid PhCS₂X <u>L1</u>–<u>L4</u> (DTCX, X = MgBr, Na, NH₂(C₅H₁₀), NMe₄) were synthesized using well-established procedures. Analytical, spectral and chromatographic data for these compounds are given in Tables 1–3.

 $[^{99m}Tc(S_2CPh)(S_3CPh)_2]$

The preparation of the complex bis(trithioperoxybenzoate)(dithiobenzoate)technetium-99m, [99mTc(S₂CPh)(S₃CPh)₂], was accomplished using various procedures and different salts of the ligand PhCS₂⁻ L. This study was undertaken with the aim to find the more suitable procedure for obtaining this radiopharmaceutical in high yield and in conditions as close as possible to physiological conditions. The main interest in the compound [99mTc(S₂CPh)(S₃CPh)₂] comes from the fact that in vitro experiments showed that it is very efficiently and selectively retained into leukocytes when incubated with whole blood. Further, it was found that the activity was stably incorporated thus suggesting that this tracer has a peculiar affinity for these blood constituents. The solid structure of the analogous complexes, [M(S₂CPh)(S₃CPh)₂], prepared in macroscopic amounts respectively, with the long-lived Tc-99 isotope and cold rhenium, has been determined by X-ray diffraction methods. 11 The central metal lies in a distorted trigonal prism environment made up by the six terminal sulfur atoms. These complexes were utilized to establish the chemical identity of the corresponding Tc-99m compound through the comparison of their chromatographic properties. HPLC and TLC chromatographic data are illustrated in Table 4.

The need to investigate different reaction procedures for obtaining an efficient method for the preparation of complex the [99mTc(S₂CPh)(S₃CPh)₂], at no carried-added level, was dictated by the very complicated reactivity of dithiocarboxylate ligands which may give rise to a wide variety of final products. ¹² A first attempt was undertaken by reacting the ligands L1-L4 with the $[^{99m}Tc \equiv N]^{2+}$ core prepared through a well-established procedure described a few years ago. 10 Such an attempt was justified by the fact that it was previously observed that thiol and dithiol ligands were able to remove easily the Tc≡N multiple bond to afford the corresponding S-substituted complexes. 16 Thus, the reaction was carried out by first producing the Tc≡N group through the reaction of [99mTc]pertechnetate with S-methyl-N-methyldithiocarbazate (MDTCZ), or alternatively succinic dihydrazide (SDH), as source of nitride nitrogen atoms (N³⁻), in the presence of a reducing agent such as SnCl₂. 2H₂O or a combination of HCl and a tertiary phosphine.

Another approach to the synthesis of the complex [^{99m}Tc(S₂CPh) (S₃CPh)₂] was undertaken when ^{99m}Tc-gluconate was used as prereduced substrate in the reaction with DTCX. In this preparation, a commercial kit formulation for ^{99m}Tc-gluconate¹⁵ was utilized, and successively mixed with the ligands DTCX. Finally,

Scheme 2. Synthesis of [99mTc(S₃CPh)₂(S₂CPh)]

Table 5. Influence of the chemical nature of the ligand on RCP using the MDTCZ/PDTA Kit

| Ligand | NOET ^a | $C_{10}MgBr^{a}$ | <u>L1</u> | <u>L2</u> | <u>L3</u> | <u>L4</u> |
|---|-------------------|-------------------|-----------|-------------------|-----------|-------------------|
| $ \frac{R_{\rm f}}{\text{RCP}^{\rm d}} (\%) $ | 0.62 ^b | 0.75 ^b | 0.62° | 0.62 ^c | 0.62° | 0.62 ^c |
| | > 90 | 70 | 70 | 90 | 70 | 40 |

^a Ligands used as reference (NOET: Et(OEt)NCS₂Na and C₁₀MgBr: CH₃(CH₂)₈CS₂MgBr).

 $[^{99m}Tc(S_2CPh)(S_3CPh)_2]$ was prepared through a simpler route involving the reaction of DTCX with $[^{99m}Tc]$ pertechnetate in the presence of $SnCl_2$ or HCl/tertiary phosphine as reductants. All the procedures are summarised in Scheme 2. It should be noted that, in all preparations using stannous chloride, 1,2-diaminopropane-N-N-N'-N'-tetraacetic acid (PDTA) was always added as complexing agent to avoid precipitation of Sn^{2+} ions co-ordinated to the ligand DTCX.

The results are reported in Tables 5 and 6. The complex $[^{99m}Tc(S_2CPh)(S_3CPh)_2]$ was obtained as the main product in all preparations, with a radiochemical purity (RCP) depending on both the chemical nature of the ligand DTCX and the reaction procedure performed. Starting with a precursor containing the $Tc\equiv N$ multiple bond, the highest yield (approximately 90%) was obtained with MDTCZ as donor of N^{3-} groups and using the sodium salt $\underline{L2}$ of the ligand DTCX (X = Na). Others salts gave lower RCP values. Similarly, the preparation containing SDH as N^{3-} donor produced unsatisfactory results with a final yield lower than 60%.

The complex $[^{99m}Tc(S_2CPh)(S_3CPh)_2]$ could not be easily obtained starting from $[^{99m}Tc]$ pertechnetate and $SnCl_2$ (RCP < 20%). Nevertheless, the use of a tertiary phosphine and aqueous hydrochloric acid

^bTLC (SiO₂), eluent CH₂Cl₂, R_f (\pm 0.12).

^cTLC (SiO₂), eluent petroleum ether/CH₂Cl₂: 70/30, R_f (\pm 0.12).

^d Radiochemichal purity estimated.

Table 6. Different kit formulations for the synthesis of [99mTc(S₃CPh)₂(S₂CPh)] with PhCS2Na L2

| KIT | medium / pH | T (°C) | t (min) | $R_{ m f}^{\ m g}$ | PRC (%) |
|---|-------------|--------|---------|---------------------|---------|
| MDTCZ/PDTA ^a | рН4 | 100 | 45 | 0.62 | 85 |
| HCl/PPh ₃ ^b | Serum φ | 100 | 30 | 0.62 | 71 |
| HCl/TPPTS ^c | Serum φ | 100 | 30 | 0.62 | 94 |
| SnCl ₂ /Gluconate ^d | Serum φ | 100 | 15 | 0.62 | > 95 |
| SDHe | Serum φ | 100 | 45 | 0.62 | 55 |
| SnCl ₂ / PDTA ^f | Serum φ | 100 | 45 | 0.62 | < 20 |

^a MDTCZ/PDTA kit (CIS bio international) and PhCS₂NH₂(CH₂)₅ L3.

gave the ^{99m}Tc-radiopharmaceutical in higher yield (71% for PPh₃ and 94% for TPPTS). The most quantitative reaction was obtained using the ^{99m}Tc-gluconate kit. [^{99m}Tc(S₃CPh)₂(S₂CPh)] was prepared with the highest radiochemical yield (> 95%) through the reaction of ^{99m}Tcgluconate with PhCS₂Na at 100°C for 15 min. In this reaction, the use of the other salts of the same ligand produced lower yields.

Conclusion

In this work, the ^{99m}Tc-labelled compound [^{99m}Tc(S₃CPh)₂(S₂CPh)] analogous to the compounds $[M(S_3CPh)_2(S_2CPh)]$ $(M = {}^{99}Tc, Re)$. previously isolated and characterized in macroscopic amounts, was prepared using several procedures. The most efficient method was based on the reaction of a lyophilized formulation of ^{99m}Tc-gluconate with the sodium salt L2 of phenyldithiocarboxylic acid. The neutral and lipophilic [99mTc(S₃CPh)₂(S₂CPh)] was obtained in very high yield (> 95%) and characterized by TLC and HPLC chromatography. The similarity between its chromatographic behaviour and that of the analogous ^{99g}Tc and Re complexes clearly established the chemical nature of this new radiopharmaceutical. The aim of these experiments will be in carefully evaluating the possibility to introduce the new ^{99m}Tclabelled compound into the clinical practice as an attractive alternative to the current employed leukocyte labelling agents.

^bHCl 0.1 N (0.2 ml), PPh₃ 2.10⁻² M / EtOH (0.2 ml). ^cHCl 0.1 N (0.2 ml), TPPTS 2.10⁻² M / H₂O (0.2 ml).

^dSnCl₂/gluconate kit (CIS bio international) diluted 1/10 (1.0 ml).

^eSDH kit (CIS bio international).

fSnCl₂/ PDTA kit.

 $^{^{\}rm g}$ TLC (SiO₂), eluent petroleum ether/CH₂Cl₂: 70/30, $R_{\rm f} \pm 0.12$.

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