Medicinal Chemistry

Subscriber access provided by IDAHO STATE UNIV

Article

Expanding the Chemical Space of Withaferin A by Incorporating Silicon to Improve its Clinical Potential on Human Ovarian Carcinoma Cells

Nayra Rodríguez Perestelo, Gabriel García Llanos, Carolina Pérez Reyes, Angel Amesty, Kartheek Sooda, Saeed Afshinjavid, Ignacio Antonio Jiménez Díaz, Farideh Javid, and Isabel López Bazzocchi *J. Med. Chem.*, **Just Accepted Manuscript •** DOI: 10.1021/acs.jmedchem.9b00146 • Publication Date (Web): 22 Apr 2019 **Downloaded from http://pubs.acs.org on April 22, 2019**

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



Expanding the Chemical Space of Withaferin A by Incorporating Silicon to Improve its Clinical Potential on Human Ovarian Carcinoma Cells

Nayra R. Perestelo,[†] Gabriel G. LLanos,[†] Carolina P. Reyes,[†] Angel Amesty,[†] Kartheek Sooda,[‡] Saeed Afshinjavid,[§] Ignacio A. Jiménez,[†] Farideh Javid,[‡] and Isabel L. Bazzocchi^{*,†}

[†]Instituto Universitario de Bio-Orgánica Antonio González, Departamento de Química Orgánica, Universidad de La Laguna, Avenida Astrofísico Francisco Sánchez 2, 38206 La Laguna, Tenerife, Spain

[‡] Department of Pharmacy, School of Applied Science, University of Huddersfield, Queensgate, Huddersfield HD1 3DH, United Kingdom

[§] College of Arts, Technology and Innovation (ATI), University of East London, London E16 2RD, United Kingdom

ABSTRACT: Ovarian cancer represents the seventh most commonly diagnosed cancer worldwide. Herein, we report on the development of a withaferin A-silyl ether library with 30 analogues reported for the first time. Cytotoxicity assays on human epithelial ovarian carcinoma cisplatin-sensitive and -resistant cell lines identified eight analogues displaying nanomolar-potency (IC₅₀ ranging from 1 to 32 nM), higher than the lead compound and reference drug. This cytotoxic potency is also coupled with a good selectivity index on a non-tumoral cell line. Cell cycle analysis of two potent analogues revealed cell death by apoptosis without indication of cell cycle arrest in G0/G1 phase. The structure-activity relationship (SAR) and *in silico* ADME studies demonstrated that the incorporation of silicon and a carbonyl group at C-4 in the WA-framework enhances potency, selectivity and drug-likeness. These findings reveal analogues 22, 23 and 25 as potential candidates for clinical translation in patients with relapsed ovarian cancer.

INTRODUCTION

Ovarian cancer (OC) represents the seventh most commonly diagnosed cancer worldwide.¹ Current treatment entails cytoreductive surgery followed by platinum- or taxane-based chemotherapy. Initially, OC responds positively in 70 to 80% of the cases. However, nearly 70% of patients suffer a relapse within 6 months of the last chemotherapeutic cycle, which is attributed to patients eventually developing resistance to carboplatin and paclitaxel adjuvant chemotherapy.³ Therefore, chemotherapy resistance, whether primary (i.e., intrinsic) or secondary (i.e., acquired), represents a major hurdle in OC treatment.⁴ Additionally, platinum-based chemotherapy is associated with multiple severe side effects (e.g. nausea, myelosuppression, neurotoxicity, nephrotoxicity, hepatotoxicity and ototoxicity). ⁵ Thus, there is an urgent need for new second-line therapies to improve the prognosis of patients with relapsed OC. As an alternative treatment strategy to reduce the side effects and resistance caused by cis-platinum-based chemotherapy a number of combinations with other compounds have been explored. In this sense, Natural Products (NPs) are ideal candidates for OC chemoprevention or adjuvants of conventional chemotherapy. Recently, Pistollato and co-workers⁶ have reviewed NPs targeting OC, describing the molecular mechanisms underlying their effects. These NPs, which include curcumin, epigallocatechin 3-gallate, resveratrol, sulforaphane and Withaferin A, are characterized by long-term safety and negligible and/or inexistent side effects, and have been proposed as possible adjuvants to traditional chemotherapy.

Withaferin A (WA), a natural steroidal lactone, is a promising drug candidate multi-targeting various cancer hallmarks. WA down-regulates the Notch, Akt and bcl-2 pathways and causes growth inhibition and apoptosis induction in ovarian carcinoma cell lines, CaOV3 and SKOV3. Studies conducted on various epithelial cancer cell lines (cisplatin-sensitive A2780, cisplatin-resistant variant A2780/CP70, and p53 mutant CaOV3) revealed a

synergetic effect of WA in combination with doxorubicin ⁹ and cisplatin¹⁰ on cell death through the generation of ROS-mediated autophagy, leading to DNA damage and induction of apoptosis. The authors suggested this synergetic therapy could minimize/eliminate the side effects and induction of drug resistance associated with high drug doses. In addition, WA in combination with liposomal preparation of doxorubicin targets aldehyde dehydrogenase I positive cancer stem cells in OC.¹¹ Moreover, WA alone and in combination with cisplatin targets putative cancer stem cells,^{12, 13} suggesting that this may present more efficacious therapy for OC.

A large number of studies have demonstrated the ability of WA to suppress the *in vivo* growth of various human cancer xenograft models, including prostate, breast, lung and colon.⁷ Also experimentally induced carcinogenesis in rodent models strongly suggest the chemopreventive potential of WA.¹⁴ Furthermore, the health benefits of Ashwaganda (*Withania somnifera*), mainly attributed to its most abundant and therapeutically effective component WA, are supported by clinical trials for inflammation, immune modulation, and reducing anxiety and arthritis pain. However, to our knowledge no clinical trials have been carried out with Ashwaganda on cancer or cancer biomarkers as end points.¹⁵ Moreover, in spite of evidence from current preclinical studies suggesting WA to be a promising anticancer drug, its application in clinical oncology is non-existent.

Furthermore, the medicinal applications of organosilicon molecules are particularly interesting because differences in their chemical properties can contribute to enhancing potency and improving pharmacological profile. These differences offer the potential for unique and/or specific interactions between an organosilicon molecule and a biological macromolecule. In fact, there are many examples demonstrating that the incorporation of silyl groups provides a general strategy to increase size and lipophilicity for drug design. With the exception of simethicone, a mixture of polydimethylsiloxane and hidrated silica gel used as

an antifoaming agent,¹⁷ there are no marketed drugs containing silicon, although currently at least nine silicon-containing compounds have entered human clinical trials.¹⁸ Though still a growing area, the incorporation of silicon in drug scaffolds may well offer great potential for enlarging the chemical space of medicinal chemistry. Moreover, computational methods have become a promising tool for identifying active lead compounds and are being used with the pipeline of drug discovery in most pharmaceutical companies.¹⁹ Thus, theoretical prediction of pharmacokinetic properties i.e. ADMET (absorption, distribution, metabolism, excretion, and toxicity) play a key role in drug discovery, since an unfavorable ADMET has been identified as the major cause of failure of candidate molecules in drug development.²⁰

Previously, we have reported that incorporation of silyl ether substituents in the WA-framework enhance its cytotoxic effect on HeLa (carcinoma of the cervix), A-549 (lung carcinoma), and MCF-7 (breast adenocarcinoma) human cancer cell lines, whereas a ketone group at C-4 increases selectivity. Moreover, the induction of apoptosis by 27-O-(tert-butyldimethylsilyl)-4-dehydroxy-4-oxo-withaferin A without necrosis under extreme experimental conditions has drawn our attention to these organosilicon analogues. 21

Therefore, encouraged by previous works highlighting that WA targets various ovarian cancer cell lines, ^{6,8} and the expectation of bio-organosilicon in drug design, efforts to enlarge the chemical space of WA to improve its clinical potential as anticancer agent are continuing. The current study reports the design, synthesis and evaluation of a WA-silyl ether library with enhanced ovarian cancer cell cytotoxicity compared to the lead compound and reference drug. Furthermore, two analogues were investigated for their ability to induce apoptosis, confirming previous studies on this type of scaffold. ²² In addition, extensive structure—activity relationship (SAR) and *in silico* ADME studies were employed to understand the pharmacokinetic properties of this series of WA-analogues.

RESULTS AND DISCUSSION

1. Chemistry. When silicon is incorporated into an organic compound, the chemical and physical differences contributed by the silyl group can provide compounds with unique properties that are relevant for medicinal chemistry.¹⁶

Withaferin A is a C28 ergosterane-type steroid with a δ -lactone ring between C-22 and C-26 in the side chain. To refine structural features and enhance the anticancer profile of WA, a suitable starting material from *Withania aristata*, ²¹ a library of WA-silyl ether analogues (2-22 and 24-34) were designed and synthesized (Schemes 1-4 and detailed in the Experimental Section).

The synthesis of this WA-library was carried out using silyl chloride analogues with different electronic and steric properties, such as hydrophobicity, size and aromaticity. The first step in this task was to investigate the modification of the hydroxyl groups at C-4 and C-27 by converting them into silyl ethers. Thus, 27-silyl ether (2-10) and 4,27-disilyl ether (11-16) analogues were synthesized following the strategy outlined in Scheme 1.

Scheme 1. Synthesis of Withaferin A-silyl Ether Analogues 2-16^a

M, methyl; E, ethyl; IP, iso-propyl; PR, propyl; B, butyl; TB, tert-butyl; H, hexyl; O, octyl; P, phenyl; V, vinyl; S, silicon

^aReagents and conditions: (i) R₁R₂R₃SiCl, imidazole, DMAP, Et₃N, CH₂Cl₂, rt.

It is worth noting that treatment of WA (1) with DMIPSiCl afforded, in addition to the expected silyl ethers 18 and 19, analogues 17 and 20, which were formed by selective silylation of the secondary alcohol at C-4 and Michael addition of imidazole to the enone system in the WA-framework, respectively (Scheme 2).

Scheme 2. Synthesis of Withaferin A-silyl Ether Analogues 17-20^a

WA (1)
$$\stackrel{(i)}{\longrightarrow}$$
 $\stackrel{(i)}{\longrightarrow}$ $\stackrel{(i)}{\longrightarrow}$

^aReagents and conditions: (i) DMIPSiCl, imidazole, DMAP, Et₃N, CH₂Cl₂, rt.

Previously reported structure-activity relationship (SAR) studies on withanolides,²¹ indicated that compounds bearing a ketone at C-4 have a selective pharmacological profile. Encouraged by these results, the synthesis of analogue 23 was carried out following the strategy outlined in Scheme 3. Firstly, selective protection of the primary alcohol in WA (1) with TBDMSiCl yielded the corresponding silyl ether derivative 21, whose oxidation by treatment with Collins reagent afforded the ketone analogue 22. Further cleavage of the protecting group in 22 with carboxylic acid resin led to the 4-dehydro-WA analogue 23.

Scheme 3. Synthesis of Withaferin A-analogues 21-23^a

^aReagents and conditions: (i) TBDMSiCl, imidazole, DMAP, CH₂Cl₂, r.t.; (ii) CrO₃, py, CH₂Cl₂, rt, 5 min; (iii) Dowex (50WX8-200), acetone, rt, 24 h.

Derivatives **24-34** bearing both a 4-ketone and a 27-silyl ether group were prepared from compound **23** by silylation at C-27 with different silyl chloride reagents as shown in Scheme 4.

Scheme 4. Synthesis of 4-Oxo-withaferin A-silyl Ether Analogues 24-34^a

Among the former synthetic analogues, 30 out of 33 are reported for the first time. The structures of the new compounds were elucidated by HRMS and NMR analysis (Supporting information, S2-S31), whereas those of the previous reported analogues, compounds **21-23** were elucidated by comparison of their spectral data with those reported in the literature.²¹

2. Biological Evaluation.

Antiproliferative activity. The *in vitro* antiproliferative activity of lead compound 1 and derivative 23 (4-oxo-WA), and their silyl-analogues 2-22 and 24-34, respectively, were evaluated on two human epithelial ovarian tumor cell lines, cisplatin-sensitive (A2780) and cisplatin-resistant (A2780/CP70) cells, and on the non-carcinoma cell line ARPE-19 (human retinal pigment epithelial), the latter used to test for selectivity.

Cytotoxic evaluation (Table 1, and Supporting Information, S32) against the cisplatinsensitive cell line revealed that the cytotoxicity of 11 analogues (compounds 3-5, 7-9, 13, 15, 20, 22, and 23) was higher than the widely known anticancer WA (1, IC₅₀ 32.7 nM) and the reference drug (carboplatin, IC₅₀ 2.6 μ M), exhibiting IC₅₀ values ranging from 7.3 to 32 nM. Moreover, it is noteworthy that silyl ether analogues bearing a dimethyloctyl (7 and 13, IC₅₀

^a Reagents and conditions: (i) R₁R₂R₃SiCl, imidazole, DMAP, Et₃N, CH₂Cl₂, rt.

10 and 1.5 nM, respectively) or a dimethylphenyl (**15**, IC₅₀ 2.9 nM) moiety as well as oxidation at C-4 (**23**, IC₅₀ 7.3 nM) are favorable trends for optimal cytotoxicity against A2780 cells, improving activity by 3.2- to 21.8-fold compared to lead compound **1**. Curiously, potency of the imidazole derivative **20** (IC₅₀ 20 nM) was slightly higher than the lead compound **1**, which is opposite to previously reported SAR of withanolides in which the enone system is essential for the anticancer activity.²³ On the other hand, functional group interconversion of alcohols in WA by a trihexylsilyl (analogues **6** and **29**) or a tripropylsilyl (derivative **12**) moiety was particularly detrimental furnishing completely inactive analogues.

Drug resistance is a major obstacle for first line chemotherapy in ovarian cancer treatment.⁴ Taking into consideration the promising results obtained for WA-silyl-analogues assayed on A2780 cisplatin-sensitive cell line, this series of compounds was tested for efficacy on a human ovarian carcinoma cisplatin-resistant (A2780/CP70) cell line. The results (Table 1) indicated that 15 analogues showed from similar (7-10, 14, 15, 18, 27 and 30) to slightly improved (5, 13, 17, 20, 25 and 26) profile than the lead compound, WA (IC₅₀ 32 nM). Moreover, potencies of target compounds 2, 3 and 23 on A2780/CP70 cells, exhibiting IC₅₀ values ranging from 1 to 12.8 nM, were significantly improved from 2- to 30-fold compared to compound 1. Regarding analogues with a drastic loss of activity, again analogues 6 and 29 and the disilyl ether derivatives 12 and 16 were from 794- to 364-fold less active than WA.

After confirming that some of the newly-synthesized analogues showed potent activity against both human epithelial ovarian tumor cell lines, cytotoxicity against a non-carcinoma cell line (ARPE19, human retinal pigment epithelial cells) was evaluated for all derivatives to test for selectivity (Table 1). We assume that a selectivity index (SI) value higher than two indicates a good selectivity for inducing cytotoxicity in tumor cell lines as compared to those in non-cancerous cells, according to Suffness.²⁴ Among the evaluated compounds, 25 of them showed selectivity to a some extent (SI > 2) in the non-carcinoma (ARPE19) cell line with

respect to the A2780 cell line. Selectivity was observed for the most potent analogues, **7**, **8**, **13**, **15**, **20**, **22** and **23** (SI ranging from 2.6 to 212.0), but also for those compounds equipotent to WA, analogues **18**, **25**, and **34** (SI 191.2, 53.5 and 58.6, respectively). Similarly, SI was higher than two for 21 analogues regarding the A2780/CP70 cell line. The most active analogues on this cell line, compounds **2**, **3**, **17**, **20**, **23**, **25** and **26**, showed SI values from 2.3 to 93.5. In addition, analogues **10**, **18** and **30** with a similar profile to WA were not cytotoxic on the non-carcinoma cell line (SI 79.1, 185.7 and 62.9, respectively).

The overall results of the biological assays identified analogues 13, 15, 22 and 23 on the cisplatin-sensitive cell line, and even more noteworthy, analogues 3, 17, 23, 25 and 26 on the cisplatin-resistant cell line as having significantly improved activity profiles compared to lead compound 1. These profiles were coupled with remarkable selectivity on the non-tumoral cell line and, therefore, are suitable for further studies.

Table 1. Cytotoxic Activity $(IC_{50}, nM)^a$ of WA-analogues^b on Human Ovarian Carcinoma Cell Lines,^c and on a non-Carcinoma Cancer Cells (ARPE19).

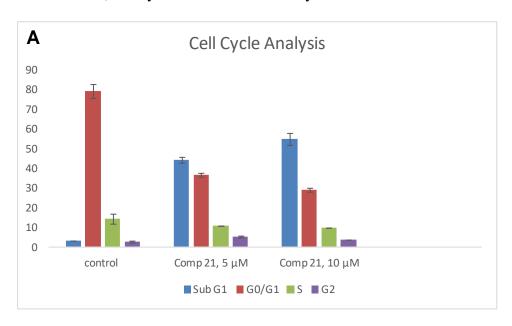
compd	A2780	A2780/CP70	ARPE19	SI^d A2780	SI ^d A2780/CP70
1	32.7 ± 0.2	32.0 ± 2.0	37.0 ± 14.0	1.1	1.2
2	> 100	12.8 ± 2.0	30.0 ± 10.0	-	2.3
3	32.0 ± 2.0	3.6 ± 1.4	30.0 ± 0.6	0.9	8.3
4	30.0 ± 2.0	62.0 ± 20.0	62.0 ± 30.0	2.1	1.0
5	27.0 ± 4.0	22.0 ± 0.3	260.0 ± 210.0	9.6	11.8
7	10.0 ± 0.7	33.0 ± 10.0	32.0 ± 5.0	3.2	1.0
8	27.0 ± 10.0	31.0 ± 2.0	86.0 ± 16.0	3.2	2.8
9	22.0 ± 5.0	28.3 ± 0.3	70.0 ± 10.0	3.2	2.5
10	46.0 ± 9	29.0 ± 6.0	2295.0 ± 25.0	49.9	79.1
13	1.5 ± 0.5	24.9 ± 10.0	318.0 ± 61.0	212.0	12.8
14	33.0 ± 0.5	27.5 ± 5.0	92.0 ± 30.0	2. 8	3.4
15	2.9 ± 1.0	29 ± 0.005	309.0 ± 40.0	106.6	10.7
17	34.0 ± 1.0	23.0 ± 9.0	310.0 ± 200.0	9.1	13.5
18	34.0 ± 0.6	35.0 ± 0.4	6500.0 ± 150.0	191.2	185.7
20	20.0 ± 7.0	19.0 ± 10.0	52.0 ± 10.0	2.6	2.7
21	47.5 ± 19.0	> 100	12.4 ± 1.0	0.3	0.1

22	17.0 ± 10.0	> 100	1660.0 ± 90.0	97.7	11.3
23	7.3 ± 6.0	< 1	32.0 ± 2.0	4.3	>32.0
24	41.0 ± 5.0	68.8 ± 8.0	820.0 ± 300.0	20.0	11.9
25	35.0 ± 10.0	20.0 ± 0.2	1870.0 ± 70.0	53.5	93.5
26	69.0 ± 20.0	21.0 ± 6.0	1740.0 ± 340.0	25.2	82.9
27	35.0 ± 6.0	35.0 ± 1.0	37.5 ± 4.0	1.1	1.1
28	53.0 ± 40.0	> 100	617.0 ± 500.0	11.6	-
30	43.0 ± 0.2	34.0 ± 0.2	2140.0 ± 100.0	49.8	62.9
33	59.0 ± 10.0	> 100	2305.0 ± 270.0	39.1	-
34	35.0 ± 10.0	> 100	2050.0 ± 350.0	58.6	-

 $[^]a$ IC₅₀ values (nM) of WA-silyl analogues were determined as described in the Biological Studies section. Carboplatin was used as a reference drug (IC₅₀ 2.6, 44.9 and 4.6 μM on A2780, A2780/CP70 and ARPE19 cell lines, respectively). Results are expressed as the mean \pm standard deviation of three independent experiments performed in duplicate. b WA-analogues exhibiting IC₅₀ values \leq 100 nM. c Human ovarian carcinoma cisplatin-sensible (A2780) and cisplatin-resistant (A2780/CP70) cell lines d SI, Selectivity Index.

Cell Cycle Assay. Our previous studies have indicated an apoptotic effect associated with withanolide-type steroids, and reported the first examples of WA-silyl ether analogues, 21 and 22²¹ in HeLa cells. Moreover, WA has been reported to trigger the apoptotic cascade by extrinsic or intrinsic pathways, e.g., in promyelocytic leukemia HL-6022²⁵ and U937 cells, ²⁶ prostate cancer cells, ²⁷ head and neck squamous carcinoma cells (HNSCC)²⁸ and Caki cells. ²⁹ In the present study, the cell cycle event mediated by two potent analogues, 21 and 22, was investigated on human ovarian cancer cell line A2780. It should be noted that 22 showed a very good selectivity on non-cancerous cells (SI 97.68) (Table 1, and Supporting Information, Table S32). Cell cycle analysis was carried out using NucleoCounter® NC-3000TM system by rapid quantification of DNA content which was measured using fluorescent 4',6-diamidino-2-phenylindole dihydrochloride (DAPI) stained cells. This assay will determine cell sorting at different phases of cell cycle. Results showed that both analogues, 21 and 22, induce a dose-dependent increase in DNA fragmentation as evidenced by the increase in the number of cells with low intensity DAPI signal in sub-G0/G1 as compared to control cells (Figure 1, and Supporting Information, Figure S34). This is

suggestive of cell death by apoptosis without indication of a cell cycle arrest in G0/G1 over 48 h contact time with the compounds. It is well known that apoptosis is often associated with growth arrest. However, cell death can be induced without the initiation of cell cycle arrest. Indeed, cell cycle arrest does not always lead to cell death.³⁰



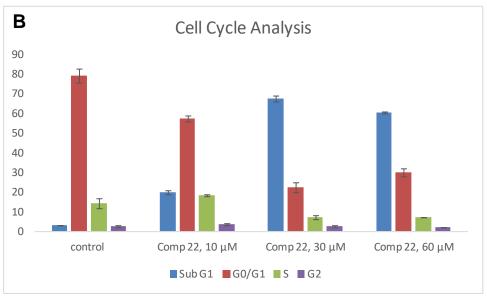


Figure 1. Percentage of cells in sub-G1, G0/G1, S and G2 phases after performing two-step cell cycle assay on human ovarian cancer cells (A2780 cell line) with compounds **21** (A) and **22** (B) at different concentrations. Data are based on 48 h exposure to compounds or vehicle control. Each column represents mean \pm s. e. mean of n = 2.

3. Structure-Activity Relationship Analysis. The previous reported SAR studies agree that an α , β -unsaturated ketone on ring A, a 5β , 6β -epoxide in ring B and an α , β -unsaturated δ -lactone on side-chain in the WA-framework²³ are antitumor structural feature requirements, and more recently, acylation is reported to enhance cytotoxicity.²²

In this work, chemical modulation by incorporation of silicon on the WA-framework was investigated. Thus, taking into consideration the IC₅₀ values against both cancer cell lines assayed, the effect of silvl ether substituents was analyzed for each synthesized analogue (2-34), according to the nature of the group attachment to silicon. The trends of the SAR study from this series of WA-silyl analogues on the A2789 cell line were as follows. Compounds carrying heterogeneous alkyl substituents on the silyl ether (7-10) displayed higher cytotoxicity than those with a homogeneous silvl ether (2-6). Indeed, replacement of dimethyloctyl or dimethylphenyl groups in the potent analogues 13 and 15 by a trihexyl, triisopropyl, or methyldiphenyl moiety led to the loss of activity (6, 12 and 29). Previous work revealed that oxidation of the secondary alcohol at C-4 of the WA-framework plays an important role in cytotoxicity. 21 Oxidation changes H-bonding ability, and lipophylicity and confers a pseudo-planarity spatial arrangement of the A-ring. These features seem to modulate the 4-oxo-analogues cytotoxicity profile. Surprisingly, the 4-oxo-WA derivative 23 (7.3 nM) showed a 4.4-fold increase in activity as compared to 1 (32.7 nM). Furthermore, to explore the effect of replacement the primary hydroxyl group at C-27 by a silyl ether, derivatives 24-34 were prepared from 23 (Scheme 4). These analogues showed a broad profile of inhibitory activities with IC₅₀ values ranging from 35 nM to 25.17 µM, although all of them were significantly less potent than the congener 23 (IC₅₀ 7.3 nM) and lead compound 1 (IC₅₀ 32.7 nM). In general, silyl-analogues with a hydroxyl group at C-4 (3-9 and 18) are more potent than those with a ketone group (26-29, 31 and 32) at this position.

Regarding the cisplatin-resistant cell line (A2780/CP70), SAR studies of this series of analogues revealed that compounds carrying a heterogeneous alkyl substituent on the silyl ether (7-10 and 13-15) displayed similar profiles (IC₅₀ 24.9-33.0 nM) to 1, indicating that activity was not greatly influenced by their corresponding silyl ether moiety on the with an olide skeleton. Analogues with a homogeneous alkyl substituent on the silyl ether (2-6, 11 and 12) showed a great range of cytotoxic activity. In fact, replacement of the triethyl or tripropyl subtituent in potent compounds 2 and 3 by a trihexyl, di-tripropyl, or dimethyldiphenyl moiety (compounds 6, 12, and 16) led to significant loss of activity. Moreover, analogue 23 was 30-fold more potent than the parent against the cisplatin-resistant cell line, whereas silyl-analogues 24-34 (Scheme 4) were significantly less potent than their congener 23, as occurred on the A2780 cell line. Therefore, a heterogeneous alkyl substituent on the silvl ether is favorable versus a homogenous one for the A2789 cell line, with the DMOS and DMPS the best functional groups. On the other hand, the homogenous alkyl substituents, TES and TPRS are the best functional groups on the A2789/CP70 cell line. Compounds with a hydroxyl group at C-4 are favorites versus those oxidized at C-4 on both cell lines, except for compound 23.

These results reveal that silylation of the WA-framework leads to a wide range of cytotoxic activity, since minor modifications on the silyl ether substituent had noteworthy repercussions on compound activity. Furthermore, silylation of the WA-framework leads to selectivity on both tumor cell lines. The potential benefit of silicon in medicinal chemistry is due to its physical-chemical properties. The atomic size and covalent radius of silicon alters the bond lengths and bond angles having an important influence on the conformation and reactivity. Moreover, the silicon enhanced cell distribution due to the increase in lipophilicity, whereas the electropositive nature of silicon enhances H-bonding ability. ^{16,18,31} Furthermore, additional lipophilicic, steric and stereoelectronic synergetic effects contributed by the

substituted-silyl ether moiety seem to guide cytotoxicity, selectivity and ADME profiles of analogues. Taking into consideration these features, WA-silyl analogues could present advantages as drug candidates.

4. *In silico* **ADME Predictions.** Understanding pharmacokinetic properties (ADME, absorption, distribution, metabolism, and excretion molecular properties) is an important step in drug discovery to select new lead/drug candidates, since potent *in vitro* activity along with enhanced ADME profiles increase the probability of clinical success. Moreover, despite a great deal of research conducted on the potential anticancer properties of WA-related withanolides, there are only two reports of ADME studies. 33,34

The QikProp module of Schrödinger software³⁵ was used for analyzing physicochemical and pharmacokinetic descriptors (ADME properties) of selected compounds (IC₅₀ values ≤ 1 μ M on A2780 and A2780/CP70 cell lines) with the aim of increasing the success rate of compounds reaching further stages of development. A detailed account of these parameters is given in Table S35 of the Supporting Information. These parameters provide insights into key aspects such as drug likeness, solubility, permeability, bioavailability predictions, oral absorption, metabolism, etc. One of the primary descriptors that were taken into account was #stars. The #stars descriptor informs about the number of properties of each compound that fail to remain within the recommended ranges, therefore, a lower number of #stars denotes a better drug-like molecule.³⁵

Thus, taking into consideration the IC₅₀ values of the assayed series of analogues with higher cytotoxic effect than WA (IC₅₀ < 32.7 nM on A2780 cell line), and #stars values (0 or 1), compounds **2**, **8**, **17**, **20**, **22**, **23** and **25** were selected to analyze their predicted pharmacokinetic properties (Table 2). Analogue **23** and the silyl ether analogue **25** as well as WA lie within the recommended range of known drugs for all the analyzed parameters (#

stars = 0), whereas silyl ether analogue 2, 8, 17, 20 and 22 fail in the QPlogS displaying low aqueous solubility, although values for these analogues were near the upper limit of the recommended range. Nevertheless, these silyl analogues showed predicted intestinal absorption rate (QPPCaco) and apparent cell permeability (QPPMDK) greater than the lead compound WA, thus predicting good oral bioavailability. The lipophilicity is also an important physicochemical property requirement for a potential drug. It is expressed as QPlogPo/w and play a crucial role in absorption, bioavailability, hydrophobic drug-receptor interactions, metabolism, and toxicity. All the selected compounds log Po/w values lay within the permissible range. The predicted values for properties such as octanol/water partition (QPlogPo/w), gut-blood barrier permeability (QPPCaco, QPlogBB and QPPMDCK), human serum albumin binding (QPlogKhsa) and percent of human oral absorption (> 83%) were within ideal ranges. Therefore, compounds under study were predicted to have good drug-likeness, since they have mostly favorable pharmacokinetic properties, especially regarding membrane permeability and oral absorptivity.

Table 2. In silico ADME profile predictions of selected WA-analogues^a and their range/recommended values.^b

Property/ Descriptor	1	2	8	17	20	22	23	25	Range/ recommended values
#stars	0	1	1	1	1	1	0	0	0-5
QPlogBB	-1.311	-1.072	-1.113	-1.253	-1.258	-087	-1.267	-1.156	-3.0 to 1.2
QPPCaco	258.201	1060.23	759.53	550.181	600.427	996.046	243.685	711.663	<25 poor, >500 great
QPPMDCK	114.484	526.993	367.48	259.339	285.031	492.595	107.544	342.512	<25 poor, >500 great
QPlogKhsa	0.329	1.189	1.017	1.136	1.1	0.892	-0.13	0.827	-1.5 to 1.5
QPlogPo/w	3.046	5.959	5.291	5.752	5.573	5.464	2.519	5.391	-2.0 to 6.5
QPlogKp	-3.842	-2.271	-2.635	-3.075	-2.816	-2.615	-3.977	-2.74	-8.0 to -1.0
QPlogS	-4.999	-7.586	-7.336	-7.379	-7.844	-6.713	-3.974	-6.489	-6.5 to 0.5
#metab	4	4	4	4	5	3	3	3	1 to 8
%HOA	87.948	90.068	83.565	83.76	83.39	86.682	84.413	83.643	>80% high <25% poor
PSA	96.36	85.36	85.36	85.36	103.18	82.2	93.2	82.2	7.0 to 200.0

SASA	718.234	906.765	874.31	88.418	949.884	877.558	711.717	882.911	300.0 to 1,000.0
Mol MW	470.605	584.867	554.8	570.84	638.918	582.851	468.589	582.851	130.0 to 725.0
#rotor	5	9	7	7	7	6	4	8	0 to 15
donorHB	1	1	1	0	1	0	0	0	0.0 to 6.0
accptHB	9.4	9.55	9.55	8.55	11.55	9.85	9.7	9.85	2.0 to 20.0
volume	1396.689	1803.47	1709.1	1757.66	1903.72	1778.773	1383.64	1785.65	500.0 to 2.000.0

^a WA-analogues exhibiting IC₅₀ values lower than those for WA on A2780 and/or A2780 cell lines (IC₅₀ < 32.7 nM), and #stars values between 0-1. ^b #star (number of property values that fall outside the 95% range of similar values for known drugs), QPlogBB (predicted brain/blood partition coefficient), QPPCaco2 (predicted human epithelial colorectal adenocarcinoma cell line permeability in nm/s), QPPMDCK (predicted Madin-Darby canine kidney permeability in nm/s), QPlogKhsa (predicted binding to human serum albumin), QPlogPo/w (predicted octanol/water partition coefficient), QPlogKp (skin permeability), QPlogS (predicted aqueous solubility), #metab (number of likely metabolic reactions), % HOA (predicted human oral absorption on 0 to 100%), PSA (Van der Waals surface area of polar nitrogen and oxygen atoms and carbonyl atoms), SASA (total solvent accessible surface area), MW (molecular weight), #rotor (number of non-trivial, non-hindered rotable bonds).

CONCLUSIONS

The current study reports on our efforts to find new drug candidates for OC resistant to current treatments. Therefore, the synthesis and evaluation of a WA-silyl ether library, together with structure-activity relationship and *in silico* ADME studies, were employed to find drug candidates for the treatment of ovarian cancer. We have successfully identified a new generation of potent and selective WA-analogues with a significantly improved cytotoxic profile. In fact, ten WA-analogues exhibited higher potency than the lead compound and reference drug on the cisplatin-sensitive cell line, and more notable, fifteen analogues enhanced cytotoxic profile on the cisplatin-resistant cell line. Cell cycle analysis of two potent analogues revealed cell death by apoptosis without cell cycle arrest in G0/G1. Furthermore, the predicted pharmacokinetic properties highlight three analogues with great potential to become drug candidates: the 4-oxo-WA (23) exhibiting single-digit nanomolar potency on both cancer cell lines, and two potent silyl-ether analogues, 22 and 25, on A2780 and A2780/CP70 cells, respectively. This potency is accompanied by an excellent selectivity

index and favorable drug-likeness. Thus, analogues reported herein are promising candidates in anticancer drug development for OC that deserve further investigation.

In summary, the current study provides an insight into the anticancer potential of WAanalogues on OC, whether alone or in combination with clinical drugs, and support the increasingly important role that silicon will play in drug design.

EXPERIMENTAL SECTION

General Methods for Chemistry. Optical rotations were measured on a Perkin Elmer 241 automatic polarimeter, in CHCl₃ at 25 °C, the $[\alpha]_D$ values are given in units of 10^{-1} deg cm² g⁻¹ ¹. ¹H (500 MHz) and ¹³C (125 MHz) NMR spectra were recorded on a Bruker Avance 400 spectrometer; chemical shifts are given in ppm and coupling constants in Hz. Solutions were typically prepared in CDCl₃ with chemical shifts referenced to deuterated solvent as an internal standard. EIMS and HREIMS were measured on a Micromass Autospec spectrometer, and ESIMS and HRESIMS (positive mode) were measured on a LCT Premier XE Micromass Autospec spectrometer. Silica gel 60 for column chromatography (particle size 15-40 and 63-200 μm), Polygram Sil G/UV₂₅₄ used for analytical and preparative TLC, and HPTLC-Platten Nano-Sil 20 UV₂₅₄ were purchased from Macherey-Nagel. Reactions were monitored by TLC, the spots were visualized by UV light and heating silica gel plates sprayed with H₂O-H₂SO₄-AcOH (1:4:20). Varian high-performance liquid chromatography (HPLC) equipment consisted of a ProStar 210 solvent delivery module, ProStar 335 photodiode array detector, using an analytical Pursuit C18 column (2.0 x 100 mm, 3 µm) with a flow rate of 0.3 mL/min, and mixtures of acetonitrile-H₂O as eluent. The degree of purity of the compounds was over 95% as indicated by the appearance of a single peak using HPLC. Unless otherwise noted, solvents and reagents were obtained from commercial suppliers and used without further purification. Anhydrous THF and Cl₂CH₂ were distilled from sodium/ benzophenone and calcium hydride ketyl under nitrogen, respectively. All solvents used were analytical grade from Panreac, and the reagents were purchased from Sigma Aldrich. Withaferin A (WA, 1), used as starting material, was isolated from the leaves of *W. aristata* as previously described.²¹

General procedure for the preparation of silyl-ether derivatives. Compounds 2-21 and 24-34. To a solution of 1 or 23 in dry CH₂Cl₂ (5-10 ml) were added imidazole (7-9 mg, 0.1-0.13 mmol), 4-(*N*,*N*-dimethylamino)-pyridine (6-10 mg, 0.05-0.08 mmol), two drops of triethylamine, and the corresponding silyl chloride. The reaction mixture was stirred at room temperature and under an argon atmosphere until all starting material was consumed. The progress of the reaction was monitored by TLC using CH₂Cl₂/acetone (9:1). After the mixture was concentrated to dryness under reduced pressure, the residue was purified by column chromatography on silica gel and eluted with CH₂Cl₂/acetone mixtures of increasing polarity (from 10:0 to 7:3) affording the desired compounds 2-21 and 24-34.

Preparation of 27-*O*-(Triethylsilyl)withaferin A (2). A solution of 1 (20 mg, 0.04 mmol) and triethylsilyl chloride (15 μL, 0.09 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 2 (8.5 mg, 37%). [α]²⁰_D +61.7 (c 0.65, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.71 (3H, s, Me-18), 0.95 (1H, m, H-14), 0.99 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-9), 1.08 (1H, m, H-17), 1.12 (1H, m, H-12), 1.15 (1H, m, H-15), 1.28 (1H, dd, J = 11.2, 14.2 Hz, H-7), 1.38 (1H, m, H-16), 1.42 (3H, s, Me-19), 1.47 (1H, m, H-11), 1.52 (1H, dd, J = 3.9, 11.2 Hz, H-8), 1.62 (1H, m, H-15), 1.68 (1H, m, H-16), 1.83 (1H, ddd, J = 3.4, 6.9, 13.9 Hz, H-11), 1.96 (1H, dd, J = 3.3, 17.5 Hz, H-23α), 1.97 (1H, m, H-12), 2.08 (3H, s, Me-28), 2.01 (1H, m, H-20), 2.16 (1H, m, H-7β), 2.47 (1H, dd, J = 13.5, 17.5 Hz, H-23β), 3.24 (1H, br s, H-6), 3.77 (1H, d, J = 5.8, H-4), 4.39 (1H, dt, J = 3.5, 13.4 Hz, H-22), 4.39, 4.52 (2H, d_{AB}, J = 11.6 Hz, H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H,

dd, J = 5.8, 10.0 Hz, H-3), OTES [0.64 (6H, t, J = 8.0 Hz), 0.96 (9H, t, J = 8.0 Hz); ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.5 (CH₃, C-19), 20.6 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 56.7 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 125.9 (C, C-25), 132.3 (CH, C-2), 141.9 (CH, C-3), 154.6 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), OTES [4.3 (3 x CH₃), 6.8 (3 x CH₂)]; EIMS m/z 584 [M]⁺ (2), 555 (100), 537 (1), 417 (1), 299 (3), 255 (21) 211 (8), 123 (11), 103 (20), 95 (12), 75 (15); HREIMS m/z 584.3509 [M]⁺ (calcd for C₃₄H₅₂O₆Si, 584.3533).

Preparation of 27-O-(tripropylsilyl)withaferin A (3). A solution of 1 (20 mg, 0.04 mmol) and tripropylsilyl chloride (10 µL, 0.05 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 3 (19.6 mg, 78%) as an amorphous solid. $[\alpha]^{20}_{D}$ +66.1 (c 0.41, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.70 (3H, s, Me-18), 0.95 (1H, H-14), 0.99 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-9), 1.10 (1H, m, H-17), 1.13 (1H, m, H-12), 1.16 (1H, m, H-15), 1.26 (1H, dd, J = 11.6, 14.9 Hz, H-7), 1.39 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.48, (1H, m, H-11), 1.53 (1H, m, H-8), 1.65 (1H, m, H-15), 1.70 (1H, m, H-16), 1.84 (1H, m, H-11), 1.93 (1H, m, H-12), 1.94 (2H, dd, J = 3.4, 17.5 Hz, H-23 α), 2.01 $(1H, H-20), 2.06 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.5, 3.9, 14.9 Hz, H-7\beta), 2.46 (1H, dd, J$ = 13.4, 17.7 Hz, H-23 β), 2.62 (1H, t, J = 2.5 Hz, OH-4), 3.24 (1H, br s, H-6), 3.76 (1H, dd, J $= 2.5, 5.9 \text{ Hz}, \text{H-4}, 4.39 \text{ (1H, dt, } J = 3.5, 13.2 \text{ Hz}, \text{H-22}), 4.37, 4.50 \text{ (2H, d}_{AB}, J = 11.7 \text{ Hz},$ H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.9, 10.0 Hz, H-3), OTPRS [0.63] (6H, br t, J = 8.5 Hz), 0.96 (9H, t, J = 7.3 Hz), 1.37 (6H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.4 (CH₃, C-19), 20.5 (CH₃, C-28), 22.1 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-

17), 56.1 (CH, C-14), 56.6 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 125.9 (C, C-25), 132.3 (CH, C-2), 141.9 (CH, C-3), 154.5 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), OTPRS [16.3 (3 x CH₂), 16.8 (3 x CH₂), 18.4 (3 x CH₃)]; ESIMS *m/z* 649 [M + Na]⁺ (100); HRESIMS *m/z* 649.3906 [M + Na]⁺ (calcd for C₃₇H₅₈O₆NaSi, 649.3900).

Preparation of 27-O-(triisopropylsilyl)withaferin A (4). A solution of 1 (20 mg, 0.04 mmol) and triisopropylsilyl chloride (10 µL, 0.05 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 4 (7.9 mg, 21%) as an amorphous solid. $[\alpha]^{20}_{D}$ +64.1 (c 0.49, CHCl₃); ¹H NMR (CDCl₃ 500 MHz) δ 0.71 (3H, s, Me-18), 0.94 (1H, m, H-14), 1.00 (3H, d, J = 6.7 Hz, Me-21), 1.11 (1H, m, H-9), 1.13 (1H, m, H-17), 1.16 (1H, m, H-12), 1.19 (1H, m, H-15), 1.27 (1H, ddd, J = 3.5, 11.0, 14.3 Hz, H-7), 1.40 (1H, m, H-16), 1.42 (3H, s, Me-19), 1.47 (1H, m, H-11), 1.53 (1H, m, H-8), 1.60 (1H, m, H-15), 1.68 $(1H, m, H-16), 1.84 (1H, ddd, J = 3.5, 6.9, 13.9 Hz, H-11), 1.97 (1H, d, J = 17.5 Hz, H-23\alpha),$ 1.98 (1H, m, H-12), 2.01 (1H, m, H-20), 2.09 (3H, s, Me-28), 2.16 (1H, dt, J = 2.6, 14.9, H- β), 2.47 (1H, dd, J = 13.2, 17.5 Hz, H-23 β), 3.25 (1H, br s, OH-4), 3.36 (1H, br s, H-6), 3.77 (1H, d, J = 5.8 Hz, H-4), 4.38 (1H, dt, J = 3.1, 13.2 Hz, H-22), 4.49, 4.60 (2H, d_{AB}, J = 11.7Hz, H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.8, 10.0 Hz, H-3), OTIPS [0.90-1.20 (3H, m), 1.07 (18H, d, J = 6.5 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.5 (CH₃, C-19), 20.6 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.2 (CH, C-9), 47.7 (C, C-10), 52.1 (CH, C-17), 56.1 (CH, C-14), 57.4 (CH₂, C-27), 62.7 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.2 (CH, C-22), 126.1 (C, C-25), 132.3 (CH, C-2), 141.8 (CH, C-3), 154.6 (C, C-24), 165.9 (C, C-26), 202.3 (C, C-1), OTIPS [12.0 (6 x CH₃), 18.0 (3 x CH)]; ESIMS m/z 649 [M + Na]⁺ (100); HRESIMS m/z $649.3893 \text{ [M + Na]}^+ \text{ (calcd for } C_{37}H_{58}O_6\text{NaSi}, 649.3900).$

Preparation of 27-*O***-(tributylsilyl)withaferin A (5).** A solution of **1** (22 mg, 0.05 mmol) and tributylsilyl chloride (15 µL, 0.06 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 5 (6.1 mg, 18%) as an amorphous solid. $[\alpha]^{20}$ _D +64.3 (c 0.35, CHCl₃); ¹H NMR (CDCl₃ 500 MHz) δ 0.71 (3H, s, Me-18), 0.95 (1H, m, H-14), 1.00 (3H, d, J = 6.8 Hz, Me-21), 1.04 (1H, m, H-9), 1.09 (1H, m, H-17), 1.14 (1H, m, H-12), 1.17 (1H, m, H-15), 1.28 (1H, m, H-7), 1.36 (1H, m, H-16), 1.42 (3H, s, Me-19), 1.47 (1H, m, H-11), 1.53 (1H, m, H-8), 1.63 (1H, m, H-15), 1.68 (1H, m, H-16), 1.84 (1H, ddd, J = 3.4, 7.1, 14.1 Hz, H-11), 1.95 (1H, m, H-12), 1.96 (1H, dd, <math>J = 2.4, 17.5 Hz, H-7), 2.01 $(1H, m, H-20), 2.07 (3H, s, Me-28), 2.16 (1H, ddd, J = 2.6, 3.9, 14.7 Hz, H-7\beta), 2.47 (1H, ddd, J = 2.6, 3.9, 14.7 Hz, H-7\beta)$ dd, J = 13.6, 17.7 Hz, H-23 β), 3.24 (1H, br s, H-6), 3.77 (1H, d, J = 5.8, H-4), 4.39 (1H, dt, J = 5.8, H-4), 4.39 = 3.5, 13.3 Hz, H-22), 4.38, 4.51 (2H, d_{AB} , J = 11.7 Hz, H-27), 6.21 (1H, d_{AB} , J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.8, 10.0 Hz, H-3), OTBS [0.63 (6H, br t, J = 7.3 Hz), 0.89 (9H, t, J = 7.3 Hz) 6.9 Hz), 1.32 (12H, m)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.3 (CH₃, C-21), 17.4 (CH₃, C-19), 20.5 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.2 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 56.7 (CH₂, C-27), 62.7 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 126.0 (C, C-25), 132.3 (CH, C-2), 141.8 (CH, C-3), 154.4 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), OTBS [13.2 (3 \times CH₃), 13.7 (3 x CH₂), 25.4 (3 x CH₂), 26.6 (3 x CH₂)]; ESIMS m/z 691 [M + Na]⁺ (100); HRESIMS m/z 691.4382 [M + Na]⁺ (calcd for C₄₀H₆₄O₆NaSi, 691.4370).

Preparation of 27-*O*-(trihexylsilyl)withaferin A (6). A solution of 1 (20.0 mg, 0.04 mmol) and trihexylsilyl chloride (15 μ L, 0.4 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 6 (17.7 mg, 59%) as an amorphous solid. [α]²⁰_D +55.1 (c 1.50, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.70 (3H, s, Me-18), 0.94 (1H, m, H-14), 0.99 (3H, d, J = 6.7 Hz, Me-21), 1.01 (1H, m, H-9), 1.07 (1H, m, H-17), 1.10 (1H,

m, H-12), 1.11 (1H, m, H-7), 1.15 (1H, m, H-15), 1.26 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.46 (1H, m, H-11), 1.52 (1H, m, H-8), 1.63 (1H, m, H-15), 1.69 (1H, m, H-16), 1.83 (1H, dd, J = 2.9, 14.2 Hz, H-11), 1.95 (1H, d, J = 17.1 Hz, H-23 α), 1.97 (1H, m, H-12), 2.00 (1H, m, H-20), 2.06 (3H, s, Me-28), 2.15 (1H, br d, J = 14.6 Hz, H-7 β), 2.46 (1H, dd, J = 13.3, 17.1 Hz, H-23 β), 2.61 (1H, d, J = 2.5 Hz, OH-4), 3.24 (1H, br s, H-6), 3.76 (1H, d, J = 5.9Hz, H-4), 4.38 (1H, dt, J = 3.2, 13.2 Hz, H-22), 4.36, 4.50 (2H, d_{AB}, J = 11.7 Hz, H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.9, 10.0 Hz, H-3), OTHS [0.62 (6H, br t, J = 7.7Hz), 0.88 (9H, t, J = 6.7 Hz), 1.29 (24H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.4 (CH₃, C-19), 20.5 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 56.7 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 126.0 (C, C-25), 132.3 (CH, C-2), 141.9 (CH, C-3), 154.4 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), OTHS [13.5 (3 x CH₂), 14.2 (3 x CH₂), 22.6 (3 x CH₃), 23.1 (3 x CH₂), 31.6 $(3 \times CH_2)$, 33.4 $(3 \times CH_2)$]; ESIMS m/z 775 $[M + Na]^+$ (100); HRESIMS m/z 775.5315 [M + Nal^+ (calcd for C₄₆H₇₆O₆NaSi, 775.5309).

Preparation of 27-*O*-(dimethyloctylsilyl)withaferin A (7). A solution of 1 (20.0 mg, 0.04 mmol) and dimethyloctylsilyl chloride (10 μL, 0.04 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 7 (5.2 mg, 20%) as an amorphous solid. [α]²⁰_D +85.0 (c 0.30, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.71 (3H, s, Me-18), 0.96 (1H, m, H-14), 0.99 (3H, d, J = 6.6 Hz, Me-21), 1.03 (1H, m, H-9), 1.08 (1H, m, H-17), 1.14 (1H, m, H-12), 1.17 (1H, m, H-15), 1.28 (1H, m, H-7), 1.39 (1H, m, H-16), 1.42 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.51 (1H, m, H-8), 1.62 (1H, m, H-15), 1.67 (1H, m, H-16), 1.84 (1H, ddd, J = 3.3, 6.9, 14.3 Hz, H-11), 1.96 (1H, m, H-12), 2.01 (1H, m, H-20), 2.06 (3H, s, Me-28), 2.16 (1H, m, H-7β), 2.47 (1H, dd, J = 13.3, 17.2 Hz, H-23β), 2.51 (1H, d, J = 2.4 Hz,

OH-4), 3.24 (1H, br s, H-6), 3.77 (1H, dd, J = 2.4, 5.8 Hz, H-4), 4.41 (1H, dt, J = 3.3, 13.2 Hz, H-22), 4.36, 4.49 (2H, d_{AB}, J = 11.6 Hz, H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.8, 10.0 Hz, H-3), ODMOS [0.13 (6H, s), 0.63 (2H, m), 0.88 (3H, t, J = 7.8 Hz), 1.23-1.35 (12H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.3 (CH₃, C-21), 17.4 (CH₃, C-19), 20.4 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.7 (CH, C-8), 30.0 (CH₂, C-23), 31.1 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.5 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 56.4 (CH₂, C-27), 62.6 (CH, C-6), 63.8 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 125.8 (C, C-25), 132.3 (CH, C-2), 141.8 (CH, C-3), 154.5 (C, C-24), 165.7 (C, C-26), 202.3 (C, C-1), ODMOS [-2.10 (2 x CH₃), 14.1 (CH₃), 16.3 (CH₂), 22.7 (CH₂), 23.1 (CH₂), 29.3 (CH₂), 29.2 (CH₂), 31.9 (CH₂), 33.5 (CH₂)]; ESIMS m/z 663 [M + Na]⁺ (100); HRESIMS m/z 663.4055 [M + Na]⁺ (calcd for C₃₈H₆₀O₆NaSi, 663.4057).

Preparation of 27-*O*-(Dimethylvinylsilyl)withaferin A (8). A solution of 1 (20 mg, 0.04 mmol) and dimethylvinylsilyl cholride (10 μL, 0.07 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 8 (12.0 mg, 47%). [α]²⁰_D +76.3 (c 0.94, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.70 (3H, s, Me-18), 0.94 (1H, m, H-14), 0.99 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-9), 1.08 (1H, m, H-17), 1.13 (1H, m, H-12), 1.16 (1H, m, H-15), 1.28 (1H, dd, J = 3.7, 11.3 Hz, H-7), 1.37 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.52 (1H, m, H-8), 1.63 (1H, m, H-15), 1.67 (1H, m, H-16), 1.83 (1H, ddd, J = 3.6, 7.3, 14.3 Hz, H-11), 1.96 (1H, dd, J = 3.3, 17.8 Hz, H-23α), 1.97 (1H, m, H-12), 2.00 (1H, m, H-20), 2.05 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.6, 4.0, 14.9 Hz, H-7β), 2.47 (1H, dd, J = 13.5, 17.8 Hz, H-23β), 2,57 (1H, d, J = 2.5 Hz, OH-4), 3.24 (1H, br s, H-6), 3.77 (1H, dd, J = 2.5, 5.9 Hz, H-4), 4.40 (1H, dt, J = 3.5, 13.3 Hz, H-22), 4.36, 4.50 (2H, d_{AB}, J = 11.6 Hz, H-27), 6.21 (1H, d, J = 9.9 Hz, H-2), 6.94 (1H, dd, J = 5.9, 9.9 Hz, H-3), ODMVS [0.22 (6H, s), 5.80 (1H, dd, J = 3.9, 20.6 Hz), 6.03 (1H, dd, J = 3.9, 15.2 Hz), 6.17 (1H, dd, J = 15.2,

20.6 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.3 (CH₃, C-21), 17.4 (CH₃, C-19), 20.5 (CH₃, C-28), 22.1 (CH₂, C-11), 24.2 (CH₂, C-15), 27.2 (CH₂, C-16), 29.7 (CH, C-8), 30.0 (CH₂, C-23), 31.1 (CH₂, C-7), 38.7 (CH, C-20), 39.3 (CH₂, C-12), 42.5 (C, C-13), 44.1 (CH, C-9), 47.6 (C, C-10), 52.0 (CH, C-17), 56.0 (CH, C-14), 56.5 (CH₂, C-27), 62.6 (CH, C-6), 63.8 (C, C-5), 69.9 (CH, C-4), 78.0 (CH, C-22), 125.7 (C, C-25), 132.3 (CH, C-2), 141.8 (CH, C-3), 154.6 (C, C-24), 165.7 (C, C-26), 202.3 (C, C-1), ODMVS [-2.2 (2 x CH₃), 133.3 (CH), 137.3 (CH₂)]; ESIMS m/z 577 [M + Na]⁺ (100); HRESIMS m/z 577.2957 [M + Na]⁺ (calcd for C₃₂H₄₆O₆NaSi, 577.2961).

Preparation of 27-O-(Methyldiphenylsilyl)withaferin A (9). A solution of 1 (20 mg, 0.04 mmol) and methyldiphenylsilyl chloride (20 µL, 0.09 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 9 (13.8 mg, 52%). $[\alpha]^{20}$ _D +48.4 (c 1.13, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.70 (3H, s, Me-18), 0.90 (1H, m, H-14), 0.96 (3H, d, J = 6.6 Hz, Me-21), 0.99 (1H, m, H-9), 1.03 (1H, m, H-17), 1.10 (1H, m, H-12), 1.16 (1H, m, H-15), 1.27 (1H, m, H-7), 1.36 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.52 (1H, m, H-8), 1.62 (1H, m, H-15), 1.65 (1H, m, H-16), 1.83 (1H, ddd, J $= 3.4, 6.9, 14.2 \text{ Hz}, \text{H-}11), 1.87 \text{ (1H, dd, J} = 3.0, 17.6 \text{ Hz}, \text{H-}23\alpha), 1.94 \text{ (1H, m, H-}12), 1.95$ (1H, m, H-20), 1.96 (3H, s, Me-28), 2.15 (1H, dt, J = 3.5, 14.5 Hz, H-7 β), 2.36 (1H, dd, J =13.7, 17.6 Hz, H-23 β), 2.57 (1H, br s, OH-4), 3.24 (1H, br s, H-6), 3.76 (1H, d, J = 6.2 Hz, H-4), 4.25 (1H, dt, J = 3.4, 13.2 Hz, H-22), 4.48, 4.61 (2H, d_{AB}, J = 11.7 Hz, H-27), 6.21 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.9, 10.0 Hz, H-3), OMDPS [0.70 (3H, s), 7.34-7.43 (6H, m), 7.60 (4H, d, J = 7.6 Hz)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.3 (CH₃, C-21), 17.5 (CH₃, C-19), 20.5 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.4 (CH₂, C-16), 29.8 (CH, C-8), 29.9 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 57.3 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.0 (CH, C-22), 125.5

(C, C-25), 132.3 (CH, C-2), 141.9 (CH, C-3), 154.8 (C, C-24), 165.6 (C, C-26), 202.3 (C, C-1), OMDPS [-3.2 (CH₃), 127.8 (4 x CH), 129.8 (2 x CH), 134.4 (4 x CH), 135.9 (C), 136.0 (C)]; ESIMS m/z 689 [M + Na]⁺ (100); HRESIMS m/z 689.3283 [M + Na]⁺ (calcd for $C_{41}H_{50}O_6NaSi$, 689.3274).

Preparation of 27-O-(tertbutyldiphenylsilyl)withaferin A (10). A solution of 1 (20.0 mg, 0.04 mmol) and tertbutyldiphenylsilyl chloride (100 µL, 0.04 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 10 (18.7 mg, 66%) as an amorphous solid. $[\alpha]^{20}_{D}$ +48.4 (c 1.60, CHCl₃); ¹H NMR (CDCl₃ 500 MHz) δ 0.71 (3H, s, Me-18), 0.95 (1H, m, H-14), 0.99 (3H, d, J = 6.8 Hz, Me-21), 1.02 (1H, m, H-9), 1.07 (1H, m, H-17), 1.12 (1H, m, H-12), 1.18 (1H, m, H-15), 1.29 (1H, dd, J = 11.6, 14.6 Hz, H-7), 1.42 (3H, s, Me-19), 1.47 (1H, m, H-11), 1.53 (1H, m, H-8), 1.69 (1H, m, H-16), 1.84 (1H, ddd, J = 3.5, 7.2, 14.3 Hz, H-11), 1.89 (1H, m, H-23 α), 1.95 (1H, m, H-12), 1.92 (3H, s, Me-28), 1.97 (1H, m, H-20), 2.17 (1H, dt, J = 3.3, 14.9 Hz, H-7 β), 2.41 (1H, dd, J = 13.2, 17.6 Hz, H-23 β), 2.56 (1H, d, J = 2.4 Hz, OH-4), 3.25 (1H, s, H-6), 3.77 (1H, dd, J = 2.4, 5.8 Hz, H-4), 4.27 (1H, dt, J = 3.3, 13.2 Hz, H-22), 4.44, 4.57 (2H, d_{AB}, J = 11.8 Hz, H-27), 6.22 (1H, d, J = 10.0 Hz, H-2), 6.94 (1H, dd, J = 5.8, 10.0 Hz, H-3), OTBDPS [1.05 (9H, s), 7.36-7.45 (6H, m), 7.71 (4H, m)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.5 (CH₃, C-19), 20.5 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.0 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.2 (CH, C-9), 47.7 (C, C-10), 52.1 (CH, C-17), 56.1 (CH, C-14), 57.9 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 125.8 (C, C-25), 132.3 (CH, C-2), 141.9 (CH, C-3), 154.1 (C, C-24), 165.6 (C, C-26), 202.3 (C, C-1), OTBDPS [19.3 (C), 26.9 (3 x CH₃), 127.6 (2 x CH), 127.7 (2 x CH), 129.6 (CH), 129.7 (CH), 133.5 (C), 133.6 (C), 135.6 (2 x CH), 135.7 (2 x CH)]; ESIMS m/z 731 [M + Na]⁺ (100); HRESIMS m/z 731.3754 [M + Na]⁺ (calcd for C₄₄H₅₆O₆NaSi, 731.3744).

Preparation of 4,27-O-Di-(triethylsilyl)withaferin A (11). A solution of 1 (20 mg, 0.04 mmol) and triethylsilyl chloride (15 µL, 0.09 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 11 (5.6 mg, 20%). $\left[\alpha\right]_{D}^{20}$ +64.0 (c 0.35, CHCl₃); ¹H NMR (CDCl₃ 500 MHz) δ 0.69 (3H, s, Me-18), 0.80 (1H, dt, J = 4.4, 12.0 Hz, H-9), 0.89 (1H, m, H-14), 0.98 (3H, d, J = 6.7 Hz, Me-21), 1.02 (1H, m, H-12), 1.07 (1H, m, H-17), 1.13 (1H, m, H-15), 1.23 (1H, dd, J = 11.6, 13.9 Hz, H-7), 1.37 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.46 (1H, m, H-11), 1.49 (1H, m, H-8), 1.62 (1H, m, H-15), 1.66 (1H, m, H-16), $1.70 \text{ (1H, m, H-11)}, 1.95 \text{ (2H, dd, } J = 3.4, 17.7 \text{ Hz, H-}23\alpha), 1.96 \text{ ((1H, m, H-12), 1.99 (1H, m, H-12), 1.99 (1$ m, H-20), 2.07 (3H, s, Me-28), 2.15 (1H, m, H-7 β), 2.46 (1H, dd, J = 13.1, 17.7 Hz, H-23 β), 3.08 (1H, br s, H-6), 3.58 (1H, d, J = 6.2, H-4), 4.39 (1H, dt, J = 3.8, 13.1 Hz, H-22), 4.39, 4.52 (2H, d_{AB} , J = 11.6 Hz, H-27), 6.15 (1H, d_{AB} , J = 9.8 Hz, H-2), 6.91 (1H, d_{AB} , J = 6.2, 9.8Hz, H-3), OTES [0.58 (6H, dq, J = 3.3, 8.1 Hz), 0.64 (6H, q, J = 7.8 Hz), 0.94 (9H, t, J = 7.9Hz), 0.96 (9H, t, J = 7.9 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.4 (CH₃, C-21), 16.3 (CH₃, C-19), 20.5 (CH₃, C-28), 21.1 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.4 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.6 (C, C-13), 44.3 (CH, C-9), 48.1 (C, C-10), 52.0 (CH, C-17), 56.2 (CH, C-14), 56.7 (CH₂, C-27), 59.8 (CH, C-6), 63.5 (C, C-5), 71.1 (CH, C-4), 78.1 (CH, C-22), 125.9 (C, C-25), 132.1 (CH, C-2), 143.7 (CH, C-3), 154.6 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), OTES [4.3 (3 x CH₃), 4.9 (3 x CH₃), 6.7 (3 x CH₂), 6.8 (3 x CH₂)]; EIMS m/z 698 [M]⁺ (4), 669 (96), 641 (1), 555 (5), 385 (5), 265 (3), 239 (6), 149 (4), 103 (100), 95 (11), 75 (85); HREIMS m/z 698.4370 [M]⁺ (calcd for C₄₀H₆₆O₆Si₂, 698.4398).

Preparation of 4,27-*O***-di-(tripropylsilyl)withaferin A (12).** A solution of **1** (20.0 mg, 0.04 mmol) and tripropylsilyl chloride (20 μ L, 0.09 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound **12** (22.4 mg, 72%) as an amorphous solid. [α]²⁰_D +93.1 (c 1.95, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.68 (3H, s, Me-18), 0.79

(1H, m, H-9), 0.89 (1H, m, H-14), 0.96 (3H, d, J = 6.7 Hz, Me-21), 1.00 (1H, m, H-12), 1.06(1H, m, H-17), 1.23 (1H, dd, J = 11.9, 14.9 Hz, H-7), 1.33 (1H, m, H-15), 1.35 (1H, m, H-16), 1.39 (3H, s, Me-19), 1.42 (1H, m, H-11), 1.47 (1H, m, H-8), 1.62 (1H, m, H-15), 1.66 (1H, m, H-16), 1.68 (1H, m, H-11), 1.94 (1H, dd, J = 3.3, 17.6 Hz, H-7), 1.95 (1H, m, H-12),1.99 (1H, m, H-20), 2.05 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.3, 4.1, 14.9 Hz, H-7 β), 2.45 $(1H, dd, J = 13.3, 17.6 Hz, H-23\beta), 3.07 (1H, br s, H-6), 3.56 (1H, d, J = 6.1 Hz, H-4), 4.36,$ 4.50 (2H, d_{AB} , J = 11.7 Hz, H-27), 4.38 (1H, dt, J = 3.5, 13.3 Hz, H-22), 6.15 (1H, d, J = 9.8Hz, H-2), 6.90 (1H, dd, J = 6.1, 9.8 Hz, H-3), OTPRS [0.55-0.65 (12H, m), 0.94 (9H, t, J =7.4 Hz), 0.95 (9H, t, J = 7.4 Hz), 1.29-1.42 (12H, m)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.3 (CH₃, C-21), 17.8 (CH₃, C-19), 20.5 (CH₃, C-28), 21.1 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.5 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.6 (C, C-13), 44.3 (CH, C-9), 48.1 (C, C-10), 52.0 (CH, C-17), 56.2 (CH, C-14), 56.6 (CH₂, C-27), 59.7 (CH, C-6), 63.4 (C, C-5), 71.1 (CH, C-4), 78.1 (CH, C-22), 126.0 (C, C-25), 132.1 (CH, C-2), 143.7 (CH, C-3), 154.4 (C, C-24), 165.8 (C, C-26), 202.2 (C, C-1), OTPRS [16.3 (3 x CH₂), 16.6 (3 x CH₂), 16.7 (3 x CH₂), 16.8 (3 x CH₂), 18.4 $(3 \times CH_3)$, 18.4 $(3 \times CH_3)$]; ESIMS m/z 805 $[M + Na]^+$ (100); HRESIMS m/z 805.5240 [M + Na^{+} (calcd for $\text{C}_{46}\text{H}_{78}\text{O}_6\text{NaSi}_2$, 805.5235).

Preparation of 4,27-*O*-di-(dimethyloctylsilyl)withaferin A (13). A solution of 1 (20.0 mg, 0.04 mmol) and dimethyloctylsilyl chloride (15 μL, 0.06 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 13 (27.2 mg, 84%) as an amorphous solid. [α]²⁰_D +84.0 (*c* 2.50, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.68 (3H, s, Me-18), 0.80 (1H, m, H-9), 0.89 (1H, m, H-14), 0.97 (3H, d, J = 6.6 Hz, Me-21), 1.01 (1H, m, H-12), 1.07 (1H, m, H-17), 1.23 (1H, m, H-7), 1.34 (2H, m, H-15, H-16), 1.38 (3H, s, Me-19), 1.44 (1H, m, H-11), 1.49 (1H, m, H-8), 1.61 (1H, m, H-15), 1.66 (1H, m, H-16), 1.70 (1H, m, H-11), 1.93 (1H, dd, J = 3.0, 17.7 Hz, H-23α), 1.94 (1H, m, H-12), 1.99 (1H, m, H-16)

20), 2.05 (3H, s, Me-28), 2.14 (1H, br d, J = 15.1 Hz, H-7β), 2.46 (1H, dd, J = 13.2, 17.7 Hz, H-23β), 3.08 (1H, br s, H-6), 3.56 (1H, d, J = 6.1 Hz, H-4), 4.35, 4.48 (2H, d_{AB}, J = 11.6 Hz, H-27), 4.39 (1H, dt, J = 3.3, 13.2 Hz, H-22), 6.14 (1H, d, J = 9.8 Hz, H-2), 6.88 (1H, dd, J = 6.1, 9.8 Hz, H-3), ODMOS [0.08 (3H, s), 0.09 (3H, s), 0.11 (6H, s), 0.55 (2H, t, J = 7.4 Hz), 0.61 (2H, t, J = 7.6 Hz), 0.88 (6H, t, J = 7.5 Hz), 1.18-1.33 (24H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.3 (CH₃, C-21), 16.7 (CH₃, C-19), 20.4 (CH₃, C-28), 21.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.0 (CH₂, C-23), 31.4 (CH₂, C-7), 38.7 (CH, C-20), 39.2 (CH₂, C-12), 42.5 (C, C-13), 44.3 (CH, C-9), 48.1 (C, C-10), 51.9 (CH, C-17), 56.1 (CH, C-14), 56.4 (CH₂, C-27), 59.9 (CH, C-6), 63.2 (C, C-5), 71.0 (CH, C-4), 78.0 (CH, C-22), 125.8 (C, C-25), 132.1 (CH, C-2), 143.6 (CH, C-3), 154.5 (C, C-24), 165.7 (C, C-26), 202.2 (C, C-1), ODMOS [-2.1 (2 x CH₃), -1.5 (CH₃), -1.4 (CH₃), 14.1 (2 x CH₃), 16.3 (2 x CH₂), 22.6 (2 x CH₂), 23.0 (CH₂), 23.2 (CH₂), 29.1 (CH₂), 29.2 (2 x CH₂), 29.3 (CH₂), 31.9 (CH₂), 32.0 (CH₂), 33.3 (CH₂), 33.4 (CH₂)]; ESIMS m/z 833 [M + Na]⁺ (100), HRESIMS m/z 833.5538 [M + Na]⁺ (calcd for C₄₈H₈₂O₆NaSi₂, 833.5548).

Preparation of 4,27-*O*-Di-(dimethylvinylsilyl)withaferin A (14). A solution of 1 (20 mg, 0.04 mmol) and dimethylvinylsilyl cholride (10 μL, 0.07 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 14 (2.4 mg, 9%). [α]²⁰_D +79.7 (*c* 0.36, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.69 (3H, s, Me-18), 0.82 (1H, dt, J = 4.3, 12.0 Hz, H-9), 0.91 (1H, m, H-14), 0.98 (3H, d, J = 6.6 Hz, Me-21), 1.03 (1H, m, H-12), 1.06 (1H, m, H-17), 1.14 (1H, m, H-15), 1.24 (1H, ddd, J = 1.2, 11.7, 14.5 Hz, H-7), 1.37 (1H, m, H-16), 1.40 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.50 (1H, m, H-8), 1.62 (1H, m, H-15), 1.66 (1H, m, H-16), 1.72 (1H, ddd, J = 3.6, 7.3, 14.4 Hz, H-11), 1.95 (1H, dd, J = 3.2, 17.6 Hz, H-23α), 1.96 (1H, m, H-12), 2.00 (1H, m, H-20), 2.05 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.4, 4.3, 15.0 Hz, H-7β), 2.47 (1H, dd, J = 13.2, 17.5 Hz, H-23β), 3.09 (1H, br s, H-6), 3.60 (1H, d, J = 6.2 Hz, H-4), 4.40 (1H, dt, J = 3.4, 13.2 Hz, H-22), 4.36, 4.50 (2H, d_{AB}, J

= 11.6 Hz, H-27), 6.16 (1H, d, J = 9.9 Hz, H-2), 6.88 (1H, dd, J = 6.2, 9.9 Hz, H-3), ODMVS [0.18 (3H, s), 0.21 (3H, s), 0.23 (6H, s), 5.78 (1H, dd, J = 4.1, 20.1 Hz), 5.81 (1H, dd, J = 4.0, 20.2 Hz), 6.01 (1H, dd, J = 4.1, 14.9 Hz), 6.03 (1H, dd, J = 4.0, 14.9 Hz), 6.10 (1H, dd, J = 14.9, 20.1 Hz), 6.18 (1H, dd, J = 14.9, 20.3 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.4 (CH₃, C-21), 16.4 (CH₃, C-19), 20.5 (CH₃, C-28), 21.3 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.0 (CH₂, C-23), 31.4 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.6 (C, C-13), 44.4 (CH, C-9), 48.1 (C, C-10), 52.0 (CH, C-17), 56.2 (CH, C-14), 56.6 (CH₂, C-27), 60.1 (CH, C-6), 63.2 (C, C-5), 71.2 (CH, C-4), 78.1 (CH, C-22), 125.7 (C, C-25), 132.2 (CH, C-2), 143.5 (CH, C-3), 154.6 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), ODMVS [-1.2 (CH₃), -1.6 (CH₃), -2.1 (CH₃), -2.2 (CH₃), 133.4 (CH), 133.6 (CH), 137.4 (CH₂), 137.4 (CH₂)]; ESIMS m/z 661 [M + Na]⁺ (100); HRESIMS m/z 661.3365 [M + Na]⁺ (calcd for C₃₆H₅₄O₆NaSi₂, 661.3357).

Preparation of 27-*O*-(dimethylphenylsilyl)withaferin A (15). A solution of 1 (20.0 mg, 0.04 mmol) and dimethylphenylsilyl chloride (10 μL, 0.06 mmol) was stirred at room temperature for 4 h. The residue was purified affording compound 15 (9.7 mg, 33%) as an amorphous solid. $[\alpha]^{20}_D$ +88.5 (*c* 0.75, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.69 (3H, s, Me-18), 0.81 (1H, m, H-14), 0.96 (3H, d, J = 6.7 Hz, Me-21), 1.01 (1H, m, H-9), 1.05 (1H, m, H-17), 1.14 (1H, m, H-12), 1.21 (1H, m, H-7), 1.33 (2H, m, H-15, H-16), 1.43 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.50 (1H, m, H-8), 1.61 (1H, m, H-15), 1.64 (1H, m, H-16), 1.72 (1H, ddd, J = 23.4, 7.0, 14.4 Hz, H-11), 1.87 (1H, dd, J = 3.2, 17.6 Hz, H-23α), 1.88 (1H, m, H-12), 1.94 (1H, m, H-20), 1.96 (3H, s, Me-28), 2.14 (1H, ddd, J = 2.3, 4.0, 14.9 Hz, H-7β), 2.37 (1H, dd, J = 13.4, 17.6 Hz, H-23β), 3.02 (1H, br s, H-6), 3.55 (1H, d, J = 6.2 Hz, H-4), 4.28 (1H, dt, J = 3.5, 13.4 Hz, H-22), 4.37, 4.50 (2H, d_{AB}, J = 11.6 Hz, H-27), 6.14 (1H, d, J = 9.9 Hz, H-2), 6.77 (1H, dd, J = 6.2, 9.9 Hz, H-3), ODMPS [0.37 (3H, s), 0.42 (6H, s), 0.43 (3H, s), 7.35-7.42 (6H, m), 7.55-7.62 (4H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-

18), 13.3 (CH₃, C-21), 16.3 (CH₃, C-19), 20.4 (CH₃, C-28), 21.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 29.9 (CH₂, C-23), 31.4 (CH₂, C-7), 38.7 (CH, C-20), 39.2 (CH₂, C-12), 42.5 (C, C-13), 44.3 (CH, C-9), 48.1 (C, C-10), 51.9 (CH, C-17), 56.1 (CH, C-14), 56.8 (CH₂, C-27), 60.0 (CH, C-6), 63.2 (C, C-5), 71.2 (CH, C-4), 77.9 (CH, C-22), 125.6 (C, C-25), 132.2 (CH, C-2), 143.3 (CH, C-3), 154.6 (C, C-24), 165.6 (C, C-26), 202.2 (C, C-1), ODMPS [-0.63 (CH₃), -1.38 (CH₃), -1.85 (CH₃), -2.00 (CH₃), 127.8 (CH), 127.8 (2 x CH), 127.9 (CH), 129.6 (CH), 129.7 (CH), 132.9 (CH), 133.5 (CH), 133.6 (2 x CH), 137.4 (C), 137.8 (C)]; ESIMS *m/z* 761 [M + Na]⁺ (100); HRESIMS *m/z* 761.3668 [M + Na]⁺ (calcd for C₄₄H₅₈O₆NaSi₂, 761.3670).

Preparation of 4,27-O-Di-(methyldiphenylsilyl)withaferin A (16). A solution of 1 (20 mg, 0.04 mmol) and methyldiphenylsilyl chloride (20 µL, 0.09 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 16 (10.5 mg, 30%). $[\alpha]^{20}$ _D +76.5 (c 0.80, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.68 (3H, s, Me-18), 0.80 (1H, m, H-8), 0.88 (1H, m, H-14), 0.94 (3H, d, J = 6.7 Hz, Me-21), 1.01 (1H, m, H-12), 1.12 (1H, m, H-17), 1.19 (1H, m, H-15), 1.33 (1H, m, H-7), 1.44 (1H, m, H-16), 1.46 (3H, s, Me-19), 1.47 (1H, m, H-11), 1.50 (1H, m, H-8), 1.60 (1H, m, H-15), 1.62 (1H, m, H-16), 1.70 (1H, ddd, J $= 4.0, 7.6, 14.6 \text{ Hz}, \text{H-}11), 1.85 \text{ (1H, dd, } J = 3.3, 17.7 \text{ Hz}, \text{H-}23\alpha), 1.91 \text{ (1H, m, H-}12), 1.94$ $(1H, m, H-20), 1.95 (3H, s, Me-28), 2.14 (1H, ddd, J = 2.3, 3.9, 14.9Hz, H-7\beta), 2.34 (1H, dd, J=2.3, 3.9, 3.9, 3.9, 3.9, 3.9)$ $J = 13.2, 17.7 \text{ Hz}, \text{H-}23\beta$), 2.98 (1H, br s, H-6), 3.61 (1H, d, J = 6.2 Hz, H-4), 4.23 (1H, dt, J = 6 $= 3.5, 13.3 \text{ Hz}, H-22), 4.46, 4.60 (2H, d_{AB}, J = 12.3 \text{ Hz}, H-27), 6.15 (1H, d, J = 9.9 \text{ Hz}, H-2),$ 6.69 (1H, dd, J = 6.2, 9.9 Hz, H-3), OMDPS [0.67 (3H, s), 0.68 (3H, s), 7.33-7.43 (12H, m), 7.53-7.61 (8H, m)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.3 (CH₃, C-21), 16.4 (CH₃, C-19), 20.5 (CH₃, C-28), 21.2 (CH₂, C-11), 24.2 (CH₂, C-15), 27.4 (CH₂, C-16), 29.8 (CH, C-8), 29.9 (CH₂, C-23), 31.3 (CH₂, C-7), 38.7 (CH, C-20), 39.2 (CH₂, C-12), 42.5 (C, C-13), 44.3 (CH, C-9), 48.1 (C, C-10), 51.9 (CH, C-17), 56.1 (CH, C-14), 57.2 (CH₂, C-27),

60.0 (CH, C-6), 63.2 (C, C-5), 71.5 (CH, C-4), 77.9 (CH, C-22), 125.5 (C, C-25), 132.3 (CH, C-2), 143.1 (CH, C-3), 154.7 (C, C-24), 165.6 (C, C-26), 202.1 (C, C-1), OMDPS [-3.2 (CH₃), -2.10 (CH₃), 127.7 (4 x CH), 127.8 (2 x CH), 1279 (2 x CH), 129.8 (2 x CH), 129.9 (CH), 130.0 (CH), 134.3 (2 x CH), 134.4 (4 x CH), 134.5 (2 x CH), 135.5 (C), 135.7 (C), 135.9 (C), 136.0 (C)]; ESIMS *m/z* 885 [M + Na]⁺ (100); HRESIMS *m/z* 885.3983 [M + Na]⁺ (calcd for C₅₄H₆₂O₆NaSi₂, 885.3983).

Preparation of derivatives 17-20. A solution of **1** (40 mg, 0.08 mmol) and dimethylisopropylsilyl chloride (20 μ L, 0.12 mmol) was stirred at room temperature for 1 h. The residue was purified affording compounds **17** (4.6 mg, 20%), **18** (8.0 mg, 18%), **19** (25.4 mg, 47%) and **20** (6.5 mg, 12%).

4-*O***-(Dimethylisopropylsilyl)withaferin A (17).** [α]²⁰_D +43.3 (*c* 0.30, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.70 (3H, s, Me-18), 0.91 (1H, m, H-14), 1.00 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-9), 1.11 (1H, m, H-12), 1.15 (1H, m, H-15), 1.24 (1H, dt, J = 2.2, 13.5 Hz, H-7), 1.38 (1H, m, H-16), 1.40 (3H, s, Me-19), 1.45 (1H, m, H-11), 1.48 (1H, m, H-8), 1.61 (1H, m, H-15), 1.64 (1H, m, H-16), 1.72 (1H, m, H-11), 1.95 (1H, m, H-12), 2.02 (1H, m, H-20), 2.04 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.3, 4.0, 14.8 Hz, H-7β), 2.50 (1H, dd, J = 13.6, 17.6 Hz, H-23β), 2.87 (1H, t, J = 6.6 Hz, OH-27), 3.09 (1H, br s, H-6), 3.57 (1H, d, J = 6.1, H-4), 4.42 (1H, dt, J = 3.4, 13.2 Hz, H-22), 4.36, 4.38 (2H, dd_{AB}, J = 6.6, 11.6 Hz, H-27), 6.16 (1H, d, J = 9.9 Hz, H-2), 6.90 (1H, dd, J = 6.1, 9.9 Hz, H-3), ODMIPS [0.05 (3H, s), 0.09 (3H, s), 0.80 (1H, m), 0.92 (3H, d, J = 6.6 Hz), 0.94 (3H, d, J = 6.5 Hz]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.3 (CH₃, C-21), 16.3 (CH₃, C-19), 20.0 (CH₃, C-28), 21.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8; CH₂, C-23), 31.4 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.6 (C, C-13), 44.4 (CH, C-9), 48.1 (C, C-10), 51.9 (CH, C-17), 56.2 (CH, C-14), 57.5 (CH₂, C-27), 59.9 (CH, C-6), 63.3 (C, C-5), 71.2 (CH, C-4), 78.8 (CH, C-22), 125.7 (C, C-25), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 132.1 (CH, C-2), 143.6 (CH, C-3), 152.7 (C, C-20), 143.6 (CH, C-3), 152.7 (C, C-20), 143.6 (CH, C-3), 152.7 (C, C-20), 143.6 (CH, C-3), 152.7 (C, C-20),

24), 167.0 (C, C-26), 202.3 (C, C-1), ODMIPS [-3.9 (CH₃), -3.6 (CH₃), 14.7 (CH), 16.7 (2 x CH₃)]; ESIMS *m/z* 593 [M + Na]⁺ (100); HRESIMS *m/z* 593.3273 [M + Na]⁺ (calcd for C₃₃H₅₀O₆NaSi, 593.3274).

27-O-(Dimethylisopropylsilyl)withaferin A (18). $[\alpha]^{20}$ _D +77.1 (c 0.56, CHCl₃); ¹H NMR (CDCl₃ 500 MHz) δ 0.70 (3H, s, Me-18), 0.92 (1H, m, H-14), 0.99 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-9), 1.07 (1H, m, H-17), 1.11 (1H, m, H-12), 1.16 (1H, m, H-15), 1.28 (1H, dd, J = 11.1 14.4 Hz, H-7), 1.37 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.46 (1H, m, H-16), 1.41 (3H, s, Me-19), 1.41 (3H, s, Me-19), 1.40 (1H, m, H-16), 1.41 (1H, m,11), 1.52 (1H, m, H-8), 1.63 (1H, m, H-15), 1.66 (1H, m, H-16), 1.83 (1H, ddd, J = 3.4, 7.5,14.3 Hz, H-7), 1.96 (1H, m, H-12), 1.97 (1H, dd, J = 3.3, 17.7 Hz, H-23 α), 2.01 (1H, m, H-20), 2.06 (3H, s, Me-28), 2.15 (1H, m, H-7 β), 2.47 (1H, dd, J = 13.4, 17.7 Hz, H-23 β), 2.55 (1H, br s, OH-4), 3.24 (1H, br s, H-6), 3.76 (1H, d, J = 5.8 Hz, H-4), 4.40 (1H, dt, J = 3.5, 13.4 Hz, H-22), 4.37, 4.50 (2H, d_{AB} , J = 11.5 Hz, H-27), 6.21 (1H, d_{AB} , J = 10.0 Hz, H-2), 6.93 (1H, dd, J = 5.8, 10.0 Hz, H-3), ODMIPS [0.09 (6H, s), 0.89 (1H, m), 0.95 (6H, d, J = 6.4Hz)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃, C-18), 13.4 (CH₃, C-21), 17.5 (CH₃, C-19), 20.5 (CH₃, C-28), 22.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.2 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 44.1 (CH, C-9), 47.7 (C, C-10), 52.0 (CH, C-17), 56.1 (CH, C-14), 56.6 (CH₂, C-27), 62.6 (CH, C-6), 63.9 (C, C-5), 69.9 (CH, C-4), 78.1 (CH, C-22), 125.9 (C, C-25), 132.3 (CH, C-2), 141.8 (CH, C-3), 154.5 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), ODMIPS [-4.4 (CH₃), -4.5 (CH₃), 14.4 (CH), 16.9 (2 x CH₃)]; ESIMS m/z 593 [M + Na]⁺ (100); HRESIMS m/z $593.3265 \text{ [M + Na]}^+ \text{ (calcd for } C_{33}H_{50}O_6\text{NaSi}, 593.3274).$

4,27-*O***-Di-(dimethylisopropylsilyl)withaferin A (19).** [α]²⁰_D +59.6 (c 0.50, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.69 (3H, s, Me-18), 0.83 (1H, H-9), 0.91 (1H, H-14), 0.98 (3H, d, J = 6.6 Hz, Me-21), 1.06 (1H, H-12), 1.08 (1H, H-17), 1.15 (1H, H-15), 1.24 (1H, H-7), 1.39 (3H, s, Me-19), 1.40 (1H, H-16), 1.48 (1H, H-11), 1.51 (1H, H-8), 1.64 (1H, H-15), 1.69 (1H,

H-16), 1.73 (1H, H-11), 1.96 (1H, H-12), 1.95 (1H, dd , J = 3.2, 17.7Hz, H-23α), 2.00 (1H, H-20), 2.06 (3H, s, Me-28), 2.15 (1H, ddd, J = 2.1, 4.0, 14.8 Hz, H-7β), 2.47 (1H, dd, J = 3.5, 13.2 Hz, H-23β), 4.39 (1H, dt, J = 3.5, 13.2 Hz, H-22), 4.37, 4.50 (2H, d_{AB}, J = 11.5 Hz, H-27), 6.15 (1H, d, J = 9.8 Hz, H-2), 6.90 (1H, dd, J = 6.1, 9.8 Hz, H-3), ODMIPS [0.05 (3H, s), 0.08 (3H, s), 0.09 (6H, s), 0.80 (1H, m), 0.90 (1H, m), 0.92 (3H, d, J = 7.0 Hz), 0.94 (3H, d, J = 6.6 Hz), 0.96 (6H, d, J = 6.6 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.5 (CH₃, C-18), 13.4 (CH₃, C-21), 16.3 (CH₃, C-19), 20.5 (CH₃, C-28), 21.2 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.8 (CH, C-8), 30.1 (CH₂, C-23), 31.5 (CH₂, C-7), 38.8 (CH, C-20), 39.3 (CH₂, C-12), 42.6 (C, C-13), 44.4 (CH, C-9), 48.1 (C, C-10), 52.0 (CH, C-17), 56.2 (CH, C-14), 56.6 (CH₂, C-27), 59.9 (CH, C-6), 63.3 (C, C-5), 71.2 (CH, C-4), 78.1 (CH, C-22), 125.9 (C, C-25), 132.1 (CH, C-2), 143.6 (CH, C-3), 154.5 (C, C-24), 165.8 (C, C-26), 202.3 (C, C-1), ODMIPS [-4.5 (CH₃), -4.4 (CH₃), -3.9 (CH₃), -3.6 (CH₃), 14.4 (CH), 14.7 (CH), 16.7 (2 x CH₃), 16.9 (2 x CH₃)]; ESIMS m/z 693 [M + Na]⁺ (100); HRESIMS m/z 693.3975 [M + Na]⁺ (calcd for C₃₈H₆₂O₆NaSi₂, 693.3983).

3β-(Imidazol-1-yl)-27-*O*-(dimethylisopropylsilyl)withaferin A (20). [α]²⁰_D +3.8 (c 0.39, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.66 (3H, s, Me-18), 0.97 (3H, d, J = 6.6 Hz, Me-21), 1.01 (1H, m, H-14), 1.12 (1H, m, H-17), 1.16 (1H, m, H-12), 1.21 (1H, m, H-15), 1.25 (1H, m, H-9), 1.36 (1H, m, H-16), 1.37 (3H, s, Me-19), 1.41 (1H, m, H-11), 1.44 (1H, m, H-8), 1.63 (1H, m, H-15), 1.66 (1H, m, H-16), 1.70 (1H, m, H-11), 1.93 (1H, m, H-12), 1.96 (1H, dd, J = 3.3, 14.0 Hz, H-8), 1.97 (1H, m, H-20), 2.06 (3H, s, Me-28), 2.15 (1H, m, H-7β), 2.46 (1H, dd, J = 14.0, 17.5 Hz, H-23β), 2.82 (1H, br s, H-6), 2.98 (1H, dd, J = 6.6, 15.7 Hz, H-2α), 3.21 (1H, dd, J = 8.8, 15.7 Hz, H-2β), 3.50 (1H, d, J = 4.0 Hz, H-4), 4.37 (1H, dd, J = 3.3, 13.3 Hz, H-22), 4.38, 4.49 (2H, d_{AB}, J = 11.7 Hz, H-27), 4.70 (1H, ddd, J = 4.0, 6.6, 10.5 Hz, H-3), imidazole [6.93 (1H, s), 7.07 (1H, s), 7.58 (1H, s)], ODMIPS [0.07 (3H, s), 0.08 (3H, s), 0.88 (6H, d, J = 6.6 Hz), 0.90 (1H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.6 (CH₃,

C-18), 13.4 (CH₃, C-21), 15.7 (CH₃, C-19), 20.5 (CH₃, C-28), 21.4 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.2 (CH, C-8), 30.1 (CH₂, C-23), 31.1 (CH₂, C-7), 38.7 (CH, C-20), 39.4 (CH₂, C-2), 42.7 (CH, C-9), 43.2 (C, C-13), 50.4 (CH, C-17), 51.9 (C, C-10), 56.1 (CH, C-3), 56.2 (CH, C-14), 57.1 (CH₂, C-27), 58.5 (CH, C-6), 63.2 (C, C-5), 76.6 (CH, C-4), 78.1 (CH, C-22), 126.0 (C, C-25), 154.6 (C, C-24), 165.9 (C, C-26), 208.1 (C, C-1), imidazole [117.4 (CH), 129.8 (CH), 136.2 (CH)], ODMIPS [-5.3 (CH₃), -5.2 (CH₃), 18.4 (CH), 25.9 (2 x CH₃)]; ESIMS *m/z* 653 [M + 1]⁺ (100); HRESIMS *m/z* 653.3994 [M + 1]⁺ (calcd for C₃₇H₅₇N₂O₆Si, 653.3986).

Preparation of 27-*O*-(tert-butyldimethylsilyl)withaferin A (21). To a solution of 1 (108.0 mg, 0.25 mmol) in dry dichloromethane (10 mL) were added imidazole (18.0 mg, 0.24 mmol), 4-(*N*,*N*-dimethylamino)pyridine (28.0 mg, 0.24 mmol) and *tert*-butylchlorodimethylsilyl chloride (56.0 mg, 0.36 mmol). The reaction was stirred at room temperature for 3 h. The residue was then purified by column chromatography (dichloromethane/acetone, 9:1) to give 21²¹ (131.1 mg, 94%):

Preparation of 27-*O*-(*tert*-butyldimethylsilyl)-4-dehydroxy-4-oxowithaferin A (22). To a solution of 21 (105.0 mg, 0.18 mmol) in dry CH₂Cl₂ (20 mL) was added drop wise a solution of chromium trioxide/pyridine complex in CH₂Cl₂, previously prepared by addition of CrO₃ (700 mg, 7.1 mmol) to dry pyridine (1 mL) and dry CH₂Cl₂ (20 mL). The reaction mixture was stirred under an argon atmosphere for 5 min. The progress of the reaction was monitored by TLC using CH₂Cl₂/acetone (95:5). The reaction was subsequently quenched with 2-propanol (0.5 mL) and the resulting black suspensions were filtered through florisil. The filtrate was concentrated under reduced pressure, and the residue was purified by preparative TLC using CH₂Cl₂/acetone (9/1) to give the corresponding derivative 22²¹ (102.0 mg, 97%).

Preparation of 4-dehydroxy-4-oxowithaferin A (23). To a solution of **22** (96 mg, 0.16 mmol) in dry acetone (12 mL) was added a suspension of Dowex 50WX8-200 (600 mg) in dry acetone (16 mL). The reaction mixture was stirred at room temperature for 24 h. The progress of the reaction was monitored by TLC using CH₂Cl₂/acetone (95:5). The suspension were filtered through a pad of celite, and the filtrate was concentrated under reduced pressure. The residue was purified by preparative TLC using CH₂Cl₂/acetone (95/5) to give the corresponding derivative **23**²¹ (74.0 mg, 99%).

Preparation of 27-O-(trimethylsilyl)-4-dehydroxy-4-oxowithaferin A (24). A solution of 23 (10.0 mg, 0.02 mmol) and trimethylsily chloride (10 µL, 0.08 mmol) was stirred at room temperature for 10 min. The residue was purified affording compound 24 (8.8 mg, 41%) as an amorphous solid. $[\alpha]^{20}_D$ +59.7 (c 0.70, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.01 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-14), 1.14 (1H, m, H-17), 1.19 (1H, m, H-15), 1.26 (1H, m, H-12), 1.34 (1H, m, H-16), 1.37 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.44 (1H, m, H-11), 1.46 (1H, m, H-9), 1.58 (1H, m, H-8), 1.62 (1H, m, H-15), 1.67 (1H, m, H-16), 1.97 (1H, m, H-23α), 1.99 (1H, m, H-20), 2.01 (1H, m, H-12), 2.05 (1H, m, H-11), 2.07 (3H, s, Me-28), 2.17 (1H, dt, J = 3.2, 15.1 Hz, H-7 β), 2.49 (1H, dd, J =13.2, 17.4 Hz, H-23 β), 3.43 (1H, d, J = 1.9 Hz, H-6), 4.42 (1H, dt, J = 3.4, 13.2 Hz, H-22), 4.36, 4.49 (2H, d_{AB} , J = 11.3 Hz, H-27), 6.86 (1H, d, J = 10.5 Hz, H-2), 6.88 (1H, d, J = 10.5Hz, H-3), OTMS [0.16 (9H, s)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.7 (CH₃, C-18), 13.3 (CH₃, C-21), 19.1 (CH₃, C-19), 20.0 (CH₃, C-28), 23.4 (CH₂, C-11), 24.2 (CH₂, C-15), 27.1 (CH₂, C-16), 29.6 (CH, C-8), 29.8 (CH₂, C-23), 30.5 (CH₂, C-7), 38.7 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 43.6 (CH, C-9), 49.8 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 57.4 (CH₂, C-27), 63.5 (CH, C-6), 63.9 (C, C-5), 78.7 (CH, C-22), 125.7 (C, C-25), 139.1 (CH, C-2), 141.6 (CH, C-3), 152.7 (C, C-24), 166.9 (C, C-26), 193.8 (C, C-4), 202.1 (C, C-1), OTMS [1.90 (3 x CH₃)]; EIMS m/z 540 [M]⁺ (53), 525 (100), 509 (12), 468 (9), 342 (5), 292 (15),

213 (24), 200 (67), 124 (17), 118 (21), 110 (27), 95 (26), 75 (33), 68 (73); HREIMS *m/z* 540.2909 [M]⁺ (calcd for C₃₁H₄₄O₆Si, 540.2907).

Preparation of 27-O-(triethylsilyl)-4-dehydroxy-4-oxowithaferin A (25). A solution of 23 (10.0 mg, 0.02 mmol) and triethylsilyl chloride (10 µL, 0.06 mmol) was stirred at room temperature for 2 h. The residue was purified affording compound 25 (4.8 mg, 41%) as an amorphous solid. $[\alpha]^{20}_D$ +82.4 (c 0.25, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.00 (3H, d, J = 6.8 Hz, Me-21), 1.03 (1H, m, H-14), 1.13 (1H, m, H-17), 1.19 (1H, m, H-15), 1.27 (1H, m, H-12), 1.33 (1H, m, H-16), 1.37 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.44 (1H, m, H-11), 1.47 (1H, m, H-9), 1.61 (1H, m, H-8), 1.69 (1H, m, H-15), 1.97 (1H, dd, $J = 3.3, 17.4 \text{ Hz}, \text{H-}23\alpha$), 1.98 (1H, m, H-20), 2.00 (1H, m, H-12), 2.03 (1H, m, H-11), 2.08 $(3H, s, Me-28), 2.17 (1H, dt, J = 3.2, 15.0 Hz, H-7\beta), 2.37 (1H, dd, J = 13.1, 17.4 Hz, H-7\beta)$ β), 3.43 (1H, d, J = 2.3 Hz, H-6), 4.40 (1H, dt, J = 3.4, 13.1 Hz, H-22), 4.39, 4.52 (2H, d_{AB}, J = 11.7 Hz, H-27), 6.85 (1H, d, J = 10.4 Hz, H-2), 6.88 (1H, d, J = 10.4 Hz, H-3), OTES [0.64 (6H, q, J = 7.9 Hz), 0.96 (9H, t, J = 7.8 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.7 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 126.0 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.5 (C, C-24), 165.8 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), OTES [4.3 (3 x CH₃), 6.8 (3 x CH₂)]; ESIMS m/z 647 [M + Na]⁺ (100); HRESIMS m/z 647.3751 [M + Na]⁺ (calcd for $C_{37}H_{56}O_6NaSi$, 647.3744).

Preparation of 27-*O*-(tripropylsilyl)-4-dehydroxy-4-oxowithaferin A (26). A solution of 23 (10.0 mg, 0.02 mmol) and tripropylsilyl chloride (10 μ L, 0.05 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 26 (10 mg, 80%) as an amorphous solid. [α]²⁰_D +87.2 (c 0.78, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s,

Me-18), 1.00 (3H, d, J = 6.8 Hz, Me-21), 1.04 (1H, m, H-14), 1.13 (1H, m, H-17), 1.18 (1H, m, H-15), 1.28 (1H, m, H-12), 1.33 (1H, m, H-16), 1.37 (1H, m, H-7), 1.38 (3H, s, Me-19), 1.42 (1H, m, H-11), 1.46 (1H, m, H-9), 1.62 (1H, m, H-8), 1.66 (1H, m, H-15), 1.70 (1H, m, H-16), 1.97 (1H, dd, J = 3.5, 17.5 Hz), 2.00 (1H, m, H-20), 2.02 (1H, m, H-12), 2.04 (1H, m, H-11), 2.07 (3H, s, Me-28), 2.17 (1H, dt, J = 3.0, 14.8 Hz, H-7 β), 2.48 (1H, dd, J = 13.4, 17.5 Hz, H-23 β), 3.44 (1H, d, J = 2.1 Hz, H-6), 4.40 (1H, dt, J = 3.4, 13.4 Hz, H-22), 4.37, 4.50 $(2H, d_{AB}, J = 11.6 \text{ Hz}, H-27), 6.86 (1H, d, J = 10.3 \text{ Hz}, H-2), 6.89 (1H, d, J = 10.3 \text{ Hz}, H-3),$ OTPRS [0.63 (6H, t, J = 8.2 Hz), 0.96 (9H, t, J = 7.2 Hz), 1.33-1.41 (6H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.6 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 126.0 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.4 (C, C-24), 165.8 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), OTPRS [16.4 (3 x CH₂), 16.8 (3 x CH₂), 18.4 (3 x CH₃)]; ESIMS m/z 647 [M + Na]⁺ (100); HRESIMS m/z 647.3751 [M + Na]⁺ (calcd for C₃₇H₅₆O₆NaSi, 647.3744).

Preparation of 27-*O*-(triisopropylsilyl)-4-dehydroxy-4-oxowithaferin A (27). A solution of 23 (10.0 mg, 0.02 mmol) and triisopropylsilyl chloride (10 μL, 0.05 mmol) was stirred at room temperature for 2 h. The residue was purified affording compound 27 (8.4 mg, 67%) as an amorphous solid. [α]²⁰_D +60.5 (c 0.57, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.02 (3H, d, J = 6.7 Hz, Me-21), 1.14 (1H, m, H-14), 1.18 (1H, m, H-17), 1.27 (1H, m, H-15), 1.32 (1H, m, H-12), 1.35 (1H, m, H-16), 1.37 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.42 (1H, m, H-11), 1.47 (1H, m, H-9), 1.62 (1H, m, H-8), 1.66 (1H, m, H-15), 1.70 (1H, m, H-16), 1.98 (1H, dd, J = 3.3, 17.5 Hz, H-23α), 1.99 (1H, m, H-20), 2.01 (1H, m, H-12), 2.02 (1H, m, H-11), 2.10 (3H, s, Me-28), 2.18 (1H, dt, J = 3.2, 15.08 Hz, H-7β), 2.49 (1H, dd, J =

13.5, 17.5 Hz, H-23 β), 3.44 (1H, d, J = 2.3 Hz, H-6), 4.39 (1H, dt, J = 3.5, 13.2 Hz, H-22), 4.49, 4.60 (2H, d_{AB}, J = 11.6 Hz, H-27), 6.86 (1H, d, J = 10.4 Hz, H-2), 6.89 (1H, d, J = 10.4 Hz, H-3), OTIPS [1.07 (18H, d, J = 7.2 Hz), 2.00 (3H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.6 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.5 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.2 (CH, C-17), 55.6 (CH, C-14), 57.4 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 78.2 (CH, C-22), 126.1 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.6 (C, C-24), 165.9 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), OTIPS [12.0 (6 x CH₃), 18.0 (3 x CH)]; ESIMS m/z 647.3758 [M + Na]⁺ (calcd for C₃₇H₅₆O₆NaSi, 647.3744).

Preparation of 27-*O***-(tributylsilyl)-4-dehydroxy-4-oxowithaferin A (28).** A solution of **23** (10.0 mg, 0.02 mmol) and tributylsilyl chloride (10 μL, 0.04 mmol) was stirred at room temperature for 5 min. The residue was purified affording compound **28** (13.0 mg, 97%) as an amorphous solid. [α]²⁰_D +61.2 (c 1.30, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.00 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-14), 1.14 (1H, m, H-17), 1.19 (1H, m, H-15), 1.28 (1H, m, H-12), 1.31 (1H, m, H-16), 1.33 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.43 (1H, m, H-11), 1.46 (1H, m, H-9), 1.59 (1H, m, H-8), 1.63 (1H, m, H-15), 1.68 (1H, m, H-16), 1.97 (1H, dd, J = 3.4, 17.7 Hz, H-23α), 1.99 (1H, m, H-20), 2.01 (1H, m, H-12), 2.03 (1H, m, H-11), 2.07 (3H, s, Me-28), 2.17 (1H, dt, J = 3.1, 14.8 Hz, H-7β), 2.47 (1H, dd, J = 13.4, 17.7 Hz, H-23β), 3.44 (1H, d, J = 2.2 Hz, H-6), 4.40 (1H, dt, J = 3.5, 13.3 Hz, H-22), 4.38, 4.51 (2H, d_{AB}, J = 11.7 Hz, H-27), 6.85 (1H, d, J = 10.4 Hz, H-2), 6.88 (1H, d, J = 10.4 Hz, H-3), OTBS [0.63 (6H, br t, J = 8.1 Hz), 0.89 (9H, t, J = 6.8 Hz), 1.32 (12H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.5 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9),

49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.7 (CH₂, C-27), 63.5 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 126.0 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.4 (C, C-24), 165.7 (C, C-26), 193.8 (C, C-4), 202.1 (C, C-1), OTBS [13.2 (3 x CH₃), 13.8 (3 x CH₂), 25.4 (3 x CH₂), 26.6 (3 x CH₂)]; ESIMS *m/z* 689 [M + Na]⁺ (100); HRESIMS *m/z* 689.4219 [M + Na]⁺ (calcd for C₄₀H₆₄O₆NaSi, 689.4213).

Preparation of 27-O-(trihexylsilyl)-4-dehydroxy-4-oxowithaferin A (29). A solution of 23 (10.0 mg, 0.02 mmol) and trihexylsilyl chloride (10 µL, 0.03 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 29 (2.3 mg, 15%) as an amorphous solid. $[\alpha]^{20}_D$ +70.4 (c 0.26, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.01 (3H, d, J = 6.6 Hz, Me-21), 1.03 (1H, m, H-14), 1.13 (1H, m, H-17), 1.19 (1H, m, H-15), 1.28 (1H, m, H-12), 1.35 (1H, m, H-16), 1.38 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.43 (1H, m, H-11), 1.48 (1H, m, H-9), 1.65 (1H, m, H-15), 1.70 (1H, m, H-16), 1.97 (1H, dd, J = 3.3, 17.7 Hz, H-7), 2.00 (1H, m, H-20), 2.02 (1H, m, H-12), 2.03 (1H, m, H-11), 2.07 $(3H, s, Me-28), 2.17 (1H, dt, J = 3.2, 14.9 Hz, H-7\beta), 2.48 (1H, dd, J = 13.4, 17.5 Hz, H-7\beta)$ β), 3.44 (1H, d, J = 2.3 Hz, H-6), 4.40 (1H, dt, J = 3.5, 13.4 Hz, H-22), 4.37, 4.51 (2H, d_{AB}, J = 11.8 Hz, H-27), 6.86 (1H, d, J = 10.4 Hz, H-2), 6.89 (1H, d, J = 10.4 Hz, H-3), OTHS [0.62 (6H, br t, J = 7.9 Hz), 0.96 (9H, t, J = 7.2 Hz), 1.24-1.34 (24H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.6 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.2 (CH, C-17), 55.6 (CH, C-14), 56.7 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 126.0 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.4 (C, C-24), 165.8 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), OTHS [13.5 (3 x CH₃), 14.2 (3 x CH₂), 22.6 (3 x CH₂), 23.2 (3 x CH₂), 31.6 (3 x CH₂), 33.4 (3 x CH₂)]; ESIMS m/z 773 [M + Na]⁺ (100); HRESIMS m/z 773.5162 [M + Na]⁺ (calcd for C₄₆H₇₄O₆NaSi, 773.5152).

Preparation of 27-O-(dimethylisopropylsilyl)-4-dehydroxy-4-oxowithaferin A (30). A solution of 23 (10.0 mg, 0.02 mmol) and dimethylisopropylsilyl chloride (10 µL, 0.06 mmol) was stirred at room temperature for 30 min. The residue was purified affording compound 30 (8.8 mg, 15%) as an amorphous solid. $[\alpha]^{20}_{D}$ +89.4 (c 0.63, CHCl₃); ¹H NMR (CDCl₃ 500) MHz) δ 0.72 (3H, s, Me-18), 1.01 (3H, d, J = 6.7 Hz, Me-21), 1.03 (1H, m, H-14), 1.13 (1H, m, H-17), 1.19 (1H, m, H-15), 1.27 (1H, m, H-12), 1.31 (1H, m, H-15), 1.36 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.43 (1H, m, H-11), 1.46 (1H, m, H-9), 1.63 (1H, m, H-8), 1.65 (1H, m, H-15), 1.69 (1H, m, H-15), 1.97 (1H, dd, J = 3.4, 17.7 Hz, H-7), 1.98 (1H, m, H-20), 2.01 (1H, m, H-12), 2.03 (1H, m, H-11), 2.07 (3H, s, Me-28), 2.17 (1H, dt, J = 3.3, 15.0 Hz, H- β), 2.48 (1H, dd, J = 13.5, 17.5 Hz, H-23 β), 3.44 (1H, d, J = 2.3 Hz, H-6), 4.40 (1H, dt, J = 2.3 Hz, H-6), 4.50 (1H, dt, J = 2.33.6, 13.3 Hz, H-22), 4.38, 4.50 (2H, d_{AB} , J = 11.5 Hz, H-27), 6.86 (1H, d_{AB} , J = 10.3 Hz, H-2), 6.88 (1H, d, J = 10.3 Hz, H-3), ODMIPS [0.10 (6H, s), 0.90 (1H, m), 0.96 (6H, d, J = 6.5Hz)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.1 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.6 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 125.9 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.5 (C, C-24), 165.8 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), ODMIPS [-4.4 (CH₃), -4.5 (CH₃), 14.4 (CH), 16.9 (2 x CH₃)]; ESIMS m/z 591 [M + Na]⁺ (100); HRESIMS m/z $591.3107 \text{ [M + Na]}^+ \text{ (calcd for } C_{33}H_{48}O_6\text{NaSi}, 591.3118).$

Preparation of 27-*O*-(dimethyloctylsilyl)-4-dehydroxy-4-oxowithaferin A (31). A solution of 23 (10.0 mg, 0.02 mmol) and dimethyloctylsilyl chloride (10 μ L, 0.04 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 31 (4.4 mg, 34%) as an amorphous solid. [α]²⁰_D +81.1 (c 0.44, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.00 (3H, d, J = 6.6 Hz, Me-21), 1.03 (1H, m, H-14), 1.13 (1H, m, H-

17), 1.18 (1H, m, H-15), 1.28 (1H, m, H-12), 1.34 (1H, m, H-16), 1.38 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.44 (1H, m, H-11), 1.47 (1H, m, H-9), 1.66 (1H, m, H-15), 1.69 (1H, m, H-16), 1.97 (1H, dd, J = 3.3, 17.5 Hz, H-23 α), 1.99 (1H, m, H-20), 2.01 (1H, m, H-12), 2.03 (1H, m, H-11), 2.07 (3H, s, Me-28), 2.17 (1H, dt, J = 3.0, 15.0 Hz, H-7 β), 2.48 (1H, dd, J =13.6, 17.5 Hz, H-23 β), 3.44 (1H, d, J = 2.1 Hz, H-6), 4.41 (1H, dt, J = 3.4, 13.3 Hz, H-22), 4.36, 4.49 (2H, d_{AB} , J = 11.5 Hz, H-27), 6.86 (1H, d, J = 10.3 Hz, H-2), 6.88 (1H, d, J = 10.3Hz, H-3), ODMOS [0.13 (6H, s), 0.62 (2H, br t, J = 8.4 Hz), 0.88 (3H, t, J = 6.9 Hz), 1.23-1.34 (12H, m)]; 13 C NMR (CDCl₃, 125 MHz) δ 11.7 (CH₃, C-18), 13.3 (CH₃, C-21), 19.2 (CH₃, C-19), 20.4 (CH₃, C-28), 23.4 (CH₂, C-11), 24.2 (CH₂, C-15), 27.2 (CH₂, C-16), 30.5 (CH, C-8), 31.9 (CH₂, C-23), 33.4 (CH₂, C-7), 38.7 (CH, C-20), 39.4 (CH₂, C-12), 42.6 (C, C-13), 43.6 (CH, C-9), 49.8 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.4 (CH₂, C-27), 63.5 (CH, C-6), 63.9 (C, C-5), 78.0 (CH, C-22), 125.8 (C, C-25), 139.1 (CH, C-2), 141.6 (CH, C-3), 154.5 (C, C-24), 165.7 (C, C-26), 193.9 (C, C-4), 202.1 (C, C-1), ODMOS [-2.11 (2 x CH₃), 14.1 (CH₃), 16.3 (CH₂), 22.4 (CH₂), 23.2 (CH₂), 29.2 (CH₂), 29.3 (CH₂), 31.9 (CH₂), 33.4 (CH₂)]; ESIMS m/z 661 [M + Na]⁺ (100); HRESIMS m/z 661.3909 [M + Na]⁺ (calcd for C₃₈H₅₈O₆NaSi, 661.3900).

Preparation of 27-*O*-(dimethylvinylsilyl)-4-dehydroxy-4-oxowithaferin **A** (32). A solution of 23 (10.0 mg, 0.02 mmol) and dimethylvinylsilyl chloride (10 μL, 0.07 mmol) was stirred at room temperature for 30 min. The residue was purified affording compound 32 (4.4 mg, 40%) as an amorphous solid. [α]²⁰_D +88.8 (c 0.25, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.72 (3H, s, Me-18), 1.00 (3H, d, J = 6.7 Hz, Me-21), 1.02 (1H, m, H-14), 1.13 (1H, m, H-17), 1.18 (1H, m, H-15), 1.27 (1H, m, H-12), 1.32 (1H, m, H-16), 1.37 (1H, m, H-7), 1.39 (3H, s, Me-19), 1.43 (1H, m, H-11), 1.46 (1H, m, H-9), 1.60 (1H, m, H-8), 1.64 (1H, m, H-15), 2.06 (3H, s, Me-28), 1.68 (1H, m, H-16), 1.96 (1H, dd, J = 3.1, 17.5 Hz, H-23α), 1.99 (1H, m, H-20), 2.02 (1H, m, H-12), 2.05 (1H, m, H-11), 2.17 (1H, dt, J = 3.2, 14.9 Hz, H-7β),

2.48 (1H, dd, J = 13.3, 17.5 Hz, H-23β), 3.44 (1H, d, J = 2.2 Hz, H-6), 4.41 (1H, dt, J = 3.3, 13.1 Hz, H-22), 4.36, 4.50 (2H, d_{AB}, J = 11.5 Hz, H-27), 6.86 (1H, d, J = 10.4 Hz, H-2), 6.88 (1H, d, J = 10.4 Hz, H-3), ODMVS [0.23 (6H, s), 5.81 (1H, dd, J = 3.9, 20.3 Hz), 6.04 (1H, dd, J = 3.9, 14.9 Hz), 6.17 (1H, dd, J = 14.9, 20.3 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.4 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 30.0 (CH₂, C-23), 30.6 (CH₂, C-7), 38.8 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.6 (CH₂, C-27), 63.5 (CH, C-6), 64.0 (C, C-5), 78.1 (CH, C-22), 125.8 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.6 (C, C-24), 165.7 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), ODMVS [-2.1 (CH₃), -2.2 (CH₃), 133.4 (CH), 137.4 (CH₂)]; ESIMS m/z 575 [M + Na]⁺ (100); HRESIMS m/z 575.2804 [M + Na]⁺ (calcd for C₃₂H₄₄O₆NaSi, 575.2805).

Preparation of 27-*O*-(dimethylphenylsilyl)-4-dehydroxy-4-oxowithaferin A (33). A solution of 23 (10.0 mg, 0.02 mmol) and dimethylphenylsilyl chloride (10 μL, 0.06 mmol) was stirred at room temperature for 20 min. The residue was purified affording compound 33 (3.8 mg, 32%) as an amorphous solid. [α]²⁰_D +83.8 (c 0.21, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 0.71 (3H, s, Me-18), 0.98 (3H, d, J = 6.7 Hz, Me-21), 1.02 (1H, m, H-14), 1.10 (1H, m, H-17), 1.18 (1H, m, H-15), 1.27 (1H, m, H-12), 1.32 (1H, m, H-16), 1.35 (1H, m, H-7), 1.38 (3H, s, Me-19), 1.44 (1H, m, H-12), 1.46 (1H, m, H-9), 1.59 (1H, m, H-8), 1.62 (1H, m, H-15), 1.65 (1H, m, H-16), 1.89 (1H, dd, J = 3.2, 17.7 Hz, H-23α), 1.96 (1H, m, H-20), 1.97 (3H, s, Me-28), 1.99 (1H, m, H-12), 2.01 (1H, m, H-11), 2.17 (1H, dt, J = 3.0, 15.0 Hz, H-7β), 2.39 (1H, dd, J = 13.3, 17.5 Hz, H-23β), 3.44 (1H, d, J = 2.2 Hz, H-6), 4.40 (1H, dt, J = 3.5, 13.1 Hz, H-22), 4.37, 4.50 (2H, d_{AB}, J = 11.5 Hz, H-27), 6.86 (1H, d, J = 10.3 Hz, H-2), 6.88 (1H, d, J = 10.3 Hz, H-3), ODMPS [0.42 (3H, s), 0.43 (3H, s), 7.37 (3H, m), 7.59 (2H, m)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.3 (CH₃, C-21), 19.2 (CH₃, C-19),

20.4 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.2 (CH₂, C-16), 29.6 (CH, C-8), 29.9 (CH₂, C-23), 30.6 (CH₂, C-7), 38.7 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 56.8 (CH₂, C-27), 63.5 (CH, C-6), 64.0 (C, C-5), 77.9 (CH, C-22), 125.6 (C, C-25), 139.1 (CH, C-2), 141.6 (CH, C-3), 154.6 (C, C-24), 165.7 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), ODMPS [-2.0 (CH₃), -1.8 (CH₃), 127. 6 (2 x CH), 129.8 (CH), 133.6 (2 x CH), 137.8 (C)]; ESIMS *m/z* 625 [M + Na]⁺ (100); HRESIMS *m/z* 625.2956 [M + Na]⁺ (calcd for C₃₆H₄₆O₆NaSi, 625.2961).

Preparation of 27-O-(methyldiphenylsilyl)-4-dehydroxy-4-oxowithaferin A (34). A solution of 23 (10.0 mg, 0.02 mmol) and methyldiphenylsilyl chloride (10 µL, 0.05 mmol) was stirred at room temperature for 1 h. The residue was purified affording compound 34 (12.3 mg, 93%) as an amorphous solid. $[\alpha]^{20}$ _D +73.8 (c 0.32, CHCl₃); ¹H NMR (CDCl₃, 500) MHz) δ 0.72 (3H, s, Me-18), 0.97 (3H, d, J = 6.6 Hz, Me-21), 1.02 (1H, m, H-14), 1.10 (1H, m, H-17), 1.18 (1H, m, H-15), 1.27 (1H, m, H-12), 1.35 (1H, m, H-7), 1.35 (1H, m, H-7), 1.36 (1H, m, H-16), 1.39 (3H, s, Me-19), 1.47 (2H, m, H-9, H-11), 1.61 (1H, m, H-8), 1.65 $(1H, m, H-15), 1.66 (1H, m, H-16), 1.90 (1H, dd, J = 3.2, 17.8 Hz, H-23\alpha), 1.97 (3H, s, Me-$ 28), 1.98 (1H, m, H-20), 1.99 (1H, m, H-12), 2.02 (1H, m, H-11), 2.17 (1H, dt, J = 3.0, 14.9 Hz, H-7 β), 2.37 (1H, dd, J = 13.2, 17.7 Hz, H-23 β), 3.44 (1H, d, J = 2.3 Hz, H-6), 4.26 (1H, dt, J = 3.4, 13.2 Hz, H-22), 4.48, 4.61 (2H, d_{AB}, J = 11.8 Hz, H-27), 6.86 (1H, d, J = 10.3 Hz, H-2), 6.88 (1H, dd, J = 10.3 Hz, H-3), OMDPS [0.70 (3H, s), 7.38 (4H, t, J = 7.5 Hz), 7.40 (2H, t, J = 7.6 Hz), 7.61 (4H, d, J = 7.5 Hz)]; ¹³C NMR (CDCl₃, 125 MHz) δ 11.8 (CH₃, C-18), 13.3 (CH₃, C-21), 19.2 (CH₃, C-19), 20.5 (CH₃, C-28), 23.5 (CH₂, C-11), 24.3 (CH₂, C-15), 27.3 (CH₂, C-16), 29.6 (CH, C-8), 29.9 (CH₂, C-23), 30.5 (CH₂, C-7), 38.7 (CH, C-20), 39.4 (CH₂, C-12), 42.7 (C, C-13), 43.7 (CH, C-9), 49.9 (C, C-10), 52.1 (CH, C-17), 55.6 (CH, C-14), 57.3 (CH₂, C-27), 63.6 (CH, C-6), 64.0 (C, C-5), 77.9 (CH, C-22), 125.6 (C, C-25), 139.2 (CH, C-2), 141.6 (CH, C-3), 154.7 (C, C-24), 165.6 (C, C-26), 193.9 (C, C-4), 202.2 (C, C-1), OMDPS [-3.2 (CH₃), 127.8 (4 x CH), 129.8 (2 x CH), 134.4 (4 x CH), 135.9 (C), 136.0 (C)]; ESIMS m/z 687 [M + Na]⁺ (100); HRESIMS m/z 687.3118 [M + Na]⁺ (calcd for C₄₁H₄₈O₆NaSi, 687.3118).

Materials for Biological Studies. MTT and media for growing cell lines and all supplements were purchased from Sigma Aldrich, UK, and cell lines were purchased from ATCC. Non-cancerous cells were kindly donated by Prof Roger Phillips and Dr Simon Allison at the University of Huddersfield. Phosphate buffered saline (PBS), 50 μg/mL Annexin V-CF488A conjugate, Annexin V binding buffer (10 x concentrate), Solution 15 (500 μg/mL Hoechst 33342), Solution 16 (500 μg/mL Propidium Iodide), Solution 10 (Lysis buffer), Solution 11 (stabilization buffer), Solution 12 (500 μg/mL DAPI), NC-Slide A8TM, NC-Slide A2TM glass slides and via-1 cassettes were bought from ChemoMetec, Denmark. NC-3000TM image cytometer was used to perform the assays.

Cell Culture and Viability Assay. Cells were grown and maintained in DMEM or RPMI 1640 medium supplemented with 10% fetal bovine serum, 5% penicillin streptomycin at 37 °C, 5% CO₂/95% air as instructed by the suppliers. The cells were plated in 96-well culture plates at a density of 1 x 10⁴ cells mL⁻¹ and allowed to adhere at 37 °C for 24 h. The following day, different doses of the compounds or vehicle were added to the cells in varying concentrations of compounds and were further incubated for 96 hours. Following the aforementioned incubation time the supernatant was removed and MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added for 4 h. The ability of cells to form formazan crystals by active mitochondrial respiration was determined using a Microplate reader after dissolving the crystals in DMSO. Cytotoxicity was expressed as a

relative percentage of the absorbance measured at 540 nm in the control and extract-treated cells. Data were presented as the mean \pm s.e. mean.

Two-step Cell Cycle Assay. Cells were seeded into T25 flasks containing 5 mL of complete media and were incubated for 24 h at 37 °C. After 24 h elapsed cells were treated with vehicle control or compounds at different concentrations and left in the incubator for a further 48 h. Cells were then subjected to two-step cell cycle assay according to the manufacture's instructions. Briefly, 1 mL of cells (1 x 10⁶ cells/mL) was transferred to eppendorf tubes. In a separate eppendorf a mixture of 1960 μL of Lysis buffer (Solution 10) plus 40 μL of 500 μg/mL DAPI (Solution 12) was prepared. Eppendorfs containing cells were centrifuged at 400 g for 5 minutes, supernatant was removed and cells were resuspended in 250 μL of the above mixture, mixed well and incubated at 37 °C for 5 minutes. 250 μL of stabilization buffer (Solution 11) was then added to the cells and mixed well. 10 μL of each sample was then loaded on A8 slide and subjected to the two-step cell cycle assay using NC-3000TM.

Statistical Analysis. All results were expressed as means \pm s.e. mean. Significant differences between groups were determined using unpaired Student's *t*-test. Significance was set at p < 0.05.

ADME Properties Predictions of WA-analogues. Prediction of descriptors related to adsorption, distribution, metabolism and excretion (ADME) properties of the compounds were predicted using the QikProp program (QikProp, version 5.3)³⁶ in Fast mode and based on the method of Jorgensen.^{37,38} Preparation of compounds and the 2D-to-3D conversion was performed using LigPrep tool, a module of the Small-Molecule Drug Discovery Suite in

Schrödinger software package, followed by a MacroModel 12.01 Monte Carlo conformational search to locate the lowest energy conformation of each ligand. The program computes pharmacokinetic relevant properties such as octanol/water partitioning coefficient, aqueous solubility, brain/blood partition coefficient, Caco-2 cell permeability, serum protein binding, number of likely metabolic reactions, and others. Drug likeness (#stars), number of property descriptors from the full list of descriptors computed by the QikProp that fall outside the range of values determined for 95% of known drugs, was used as additional compound selection filter.

ASSOCIATED CONTENT

Supporting Information. ¹H and ¹³C NMR spectra of novel withaferin A-silyl derivatives **2-20** and **24-34**; cytotoxic activity of WA-silyl analogues on human ovarian carcinoma cisplatin-sensible and cisplatin-resistant cell lines, and on a non-carcinoma cancer cell line; representative scatter plot indicating the percentage of cells in G0/G1 (M1), S (M2), and G2 (M3) and Sub-G1 (M4) phases in human ovarian carcinoma cells treated with compounds **21** and **22**; *in silico* ADME profile of WA (**1**) and selected WA-analogues; Molecular formula strings with biochemical and biological data.

AUTHOR INFORMATION

Corresponding Author

*I. L.B.: phone, +34-922-318594; E-mail, ilopez@ull.edu.es.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This study was supported by the SAF2015-65113-C2-1-R Spanish MINECO, and Gobierno Autónomo de Canarias (ProID2017010071) projects. These projects are also co-funded by the European Regional Development Fund (FEDER). CPR thanks the Cabildo de Tenerife (Agustín de Betancourt Program).

ABBREVIATIONS USED

SAR, structure-activity relationship; NPs, natural products; WA, withaferin A; Et3N, triethylamina; DMAP, 4-(N,N-dimethylamino)pyridine; rt, room temperature; DMIPSiCl, dimethylisopropylsilyl chloride; py, pyridine; TBDMSCl, tert-butyldimethylsilyl chloride; MTT, 3-(4,5-dimethyl thiazol-2-yl)-2,5-diphenyltetrazolium bromide; DMSO, dimethyl sulfoxide; PBS, phosphate buffered saline.

REFERENCES

- (1) Reid, B. M; Permuth, J. B; Sellers, T. A. Epidemiology of ovarian cancer: a review. *Cancer Biol. Med* **2017**, *14*, 9-32.
- (2) Bowtell, D. D.; Böhm, S.; Ahmed, A. A.; Aspuria, P. J.; Bast Jr, R. C.; Beral, V.; Berek, J. S.; Birrer, M. J.; Blagden, S.; Bookman, M. A.; Brenton, J. D.; Chiappinelli, K. B.; Martins, F. C.; Coukos, G.; Drapkin, R.; Edmondson, R.; Fotopoulou, C.; Gabra, H.; Galon, J.; Gourley, C.; Heong, V.; Huntsman, D. G.; Iwanicki, M.; Karlan, B. Y.; Kaye, A.; Lengyel, E.; Levine, D. A.; Lu, K. H.; McNeish, I. A.; Menon, U.; Narod, S. A.; Nelson, B. H.; Nephew, K. P.; Pharoah, P.; Powell Jr, D. J.; Ramos, P.; Romero, I. L.; Scott, C. L.; Sood, A.

- K.; Stronach, E. A.; Balkwill, F. R. Rethinking ovarian cancer II: reducing mortality from high-grade serous ovarian cancer. *Nat. Rev. Cancer* **2015**, *15*, 668-679.
- (3) Siegel, R. L.; Miller, K. D.; Jemal, A. Cancer statistics 2016. CA Cancer J. Clin. 2016, 66, 7-30.
- (4) Galluzzi1, L.; Senovilla, L.; Vitale, I.; Michels, J.; Martins, I.; Kepp, O.; Castedo, M.; Kroemer, G. Molecular mechanisms of cisplatin resistance. *Oncogene* **2012**, *3*1, 1869-1883.
- (5) Benkafadar, N.; Menardo, J.; Bourien, J.; Nouvian, R.; François, F.; Decaudin, D.; Maiorano, D.; Puel, J.; Wang, J. Reversible p53 inhibition prevents cisplatin ototoxicity without blocking chemotherapeutic efficacy. *EMBO Mol. Med.* **2017**, *9*, 7-26
- (6) Pistollato, F.; Iglesias, R. C.; Ruiz, R.; Aparicio, S.; Crespo, J.; Lopez, L. D.; Giampieri, F.; Battino, M. The use of natural compounds for the targeting and chemoprevention of ovarian cancer. *Cancer Lett.* **2017**, *411*, 191-200.
- (7) Lee, I-Ch.; Choi, B. Y. Withaferin-A a natural anticancer agent with pleitropic mechanisms of action. *Int. J. Mol. Sci.* **2016**, *17*, 290-312.
- (8) Zhang, X.; Samadi, A. K.; Roby, K. F.; Timmermann, B.; Cohen, M. S. Inhibition of cell growth and induction of apoptosis in ovarian carcinoma cell lines CaOV3 and SKOV3 by natural withanolide Withaferin A. *Gynecol. Oncol.* **2012**, *124*, 606-612.
- (9) Fong, M. Y.; Jin, S.; Rane, M.; Singh, R. K.; Gupta, R.; Kakar, S. S. Withaferin A synergizes the therapeutic effect of doxorubicin through ROS-mediated autophagy in ovarian cancer. *PloS One* **2012**, *7*, e42265.

- (10) Kakar, S. S.; Jala, V. R.; Fong, M. Y. Synergistic cytotoxic action of cisplatin and withaferin A on ovarian cancer cell lines. *Biochem. Biophys. Res. Commun.* **2012**, *423*, 819-825.
- (11) Kakar, S. S.; Parte, S.; Carter, K.; Joshua, I. G.; Worth, C.; Rameshwar, P.; Ratajczak, M. Z. Withaferin A (WFA) inhibits tumor growth and metastasis by targeting ovarian cancer stem cells. *Oncotarget* **2017**, *8*, 74494-74505.
- (12) Kakar, S. S.; Ratajczak, M. Z.; Powell, K. S.; Moghadamfalahi, M.; Miller, D. M.; Batra, S. K.; Sing, S. K. Withaferin A alone and in combination with cisplatin suppresses growth and metastasis of ovarian cancer by targeting putative cancer stem cells. *PloS One* **2014**, *9*, e107596.
- (13) Kakar, S. S.; Worth, C. A; Gunjal, P.; Wang, Z.; Carter, K.; Ratajczak, M. DOXIL when combined with Withaferin A (WFA) targets ALDH1 positive cancer stem cells in ovarian cancer. JCSCR 2016, 4, e1002.
- (14) Vyas, A.R.; Singh, S.V. Molecular targets and mechanisms of cancer prevention and treatment by withaferin-A, a naturally occurring steroidal lactone. *AAPS J.* **2014**, *16*, 1–10.
- (15) Palliyaguru, D. L.; Singh, S. V.; Kensler, T. W. *Withania somnifera*: From prevention to treatment of cancer. *Mol. Nutr. Food Res.* **2016**, *60*, 1342-1353.
- (16) Franz, A. K.; Wilson, S. O. Organosilicon molecules with medicinal applications. *J. Med. Chem.* **2013**, *56*, 388-405.
- (17) Yoo, K.; Jeen, Y. T.; Kang, S. H.; Lee, J. H.; Kim, S. H.; Lee, J. M.; Choi, H. S.; Kim, E. S.; Keum, B.; Chun, H. J.; Lee, H. S.; Kim, Ch. D. Improving of bowel cleansing effect for polyethylene glycol with ascorbic acid using simethicone. A randomized controlled trial. *Medicine* **2016**, *95*, 1-6.

- (18) Ramesshi, R.; Reddy, D. S. Quest for novel chemical entities through incorporation of silicon in drug scaffolds. *J. Med. Chem.* **2018**, *61*, 3779-3798.
- (19) van de Waterbeemd, H.; Gifford, E. ADMET *in silico* modeling: Towards prediction paradise? *Nat. Rev. Drug. Discov.* **2003**, *2*, 192-204.
- (20) Sanders, J. M.; Beshore, D. C.; Culberson, J. C.; Fells, J. I.; Imbriglio, J. E.; Gunaydin, H.; Haidle, A. M.; Labroli, M.; Mattioni, B. E.; Sciammetta, N.; Dhipe, W. D.; Sherdan, R. P.; Suen, L. M.; Verras, A.; Walji, A.; Joshi, E. M.; Bueters, T. Informing the selection of screening hit series with *in silico* absorption, distribution, metabolism, excretion, and toxicity profiles. *J. Med. Chem.* **2017**, *60*, 6771-6780.
- (21) LLanos, G. G.; Araujo, L. M.; Jiménez, I. A.; Moujir, L. M.; Bazzocchi, I. L. Withaferin A-related steroids from *Withania aristata* exhibit potent antiproliferative activity by inducing apoptosis in human tumor cells. *Eur. J. Med. Chem.* **2012**, *54*, 499-511.
- (22) LLanos, G. G.; Araujo, L. M.; Jiménez, I. A.; Moujir, L. M.; Rodríguez, J.; Jiménez, C.; Bazzocchi, I. L. Structure-based design, synthesis, and biological evaluation of withaferin A-analogues as potent apoptotic inducers. *Eur. J. Med. Chem.* **2017**, *140*, 52-64.
- (23) Zhang, H.; Samadi, A. K.; Cohen, M. S.; Timmermann, B. N. Antiproliferative withanolides from the Solanaceae: a structure-activity study. *Pure Appl. Chem.* **2012**, *84*, 1353-1367.
- (24) Suffness, M.; Pezzuto, J. M. Assays related to cancer drug discovery. In Methods in Plant Biochemistry: Volume 6. Assays for Bioactivity, K. Hostettmann, Ed., Academic Press: London, UK, 1991, pp. 71-133.
- (25) Senthil, V.; Ramavedi, S.; Venkatakrishnan, V.; Giridharan, P.; Lakshmi, B. S.; Vishwakarma, R. A.; Balakrishnan, A. Withanolide induces apoptosis in HL-60 leukemia

cells via mitochondria mediated cytochrome c release and caspase activation. *Chem. Biol. Interact.* **2007**, *167*, 19-30.

- (26) Oh, J. H.; Lee, T. J.; Kim, S. H.; Choi, Y. H.; Lee, S. H.; Lee, J. M.; Kim, Y. H.; Park, J. W.; Kwon, T. K. Induction of apoptosis by withaferin A in human leukemia U937 cells through down-regulation of Akt phosphorylation. *Apoptosis* **2008**, *13*, 1494-1504.
- (27) Srinivasan, S.; Ranga, R. S.; Burikhanov, R.; Han, S. S.; Chendil, D. Par-4-dependent apoptosis by the dietary compound with aferin A in prostate cancer cells. *Cancer Res.* **2007**, *67*, 246-253.
- (28) Samadi, A. K.; Tong, X.; Mukerji, R.; Zhang, H.; Timmermann, B. N.; Cohen, M. S. Withaferin A, a cytotoxic steroid from *Vassobia breviflora*, induces apoptosis in human head and neck squamous cell carcinoma. *J. Nat. Prod.* **2010**, 73, 1476-1481.
- (29) Yang, E. S.; Choi, M. J.; Ki, J. H.; Choi, K. S.; Kwon, T. K. Whitaferin A enhances radiation-induced apoptosis in Caki cells through induction of reactive oxygen species, Bcl-2 downregulation and Akt inhibition. *Chem. Biol. Interact.* **2011**, *190*, 9-15.
- (30) Weng, L.-P.; Brown, J. L.; Eng, Ch. PTEN induces apoptosis and cell cycle arrest through phosphoinositol-3-kinase/Akt-dependent and -independent pathways. *Hum. Mol. Genet.* **2001**, *10*, 237-242.
- (31) Fujii, S.; Miyajima, Y.; Masuno, H.; Kagechika, H. Increased hydrophobicity and estrogenic activity of simple phenols with silicon and germanium-containing substituents. *J. Med. Chem.* **2013**, *56*, 160-166.
- (32) Sharma, A.; Sharma, S.; Gupta, M.; Fatima, S.; Saini, R.; Agarwal, S. M. Pharmacokinetic profiling of anticancer phytocompounds using computational approach. *Phytochem. Analysis* **2018**, *29*, 559-568.

- (33) Gill, B. S.; Navgeet; Kumar, S. Ganoderic acid targering multiple receptors in cancer: in silico and in vitro study. *Tumor Biol.* **2016**, *37*, 14271-14290.
- (34) Yadav, D. K; Kumar, S; Saloni; Singh, H.; Kim, M.; Sharma, P.; Misra, S.; Khan, F. Molecular docking, QSAR and ADMET studies of withanolide analogs against breast cancer. *Drug Des. Dev. Ther.* **2017**, *11*, 1859-1870.
 - (35) Schrödinger Release 2018-2: QikProp, Schrödinger, LLC, New York, NY, 2018.
- (36) Zhou, W.; Wang, Y.; Lu, A.; Zhang, G. Systems pharmacology in small molecular drug discovery. *Int. J. Mol. Sci.* **2016**, *17*, 246.
- (37) Duffy, E. M.; Jorgensen, W. L. Prediction of properties from simulations: Free energies of solvation in hexadecane, octanol, and water. *J. Am. Chem. Soc.* **2000**, *122*, 2878-2888.
- (38) Jorgensen, W. L.; Duffy, E. M. Prediction of drug solubility from structure. *Adv. Drug Deliv. Rev.* **200**2, *54*, 355-366.

Table of Contents Graphic

