Enantioselective Copper-Aminopyridine-catalyzed aza-Henry Reaction with Chelating N-(2-Pyridyl)Sulfonyl Imines

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ABSTRACT This article describes a copper-catalyzed aza-Henry reaction. Copper complexes of camphor-derived aminopyridines catalyze the addition of nitromethane to *N*-(2-pyridyl)sulfonyl aldimines to give the corresponding β-nitrosulfonamides with good yields and variable enantiomeric excesses (up to 83%). An example of transformation of these compounds into *N*-(2-pyridyl)sulfonyl-α-amino acids and deprotection of the sulfonamide with Mg–MeOH is provided. *Chirality 00:000–000, 2012.* © 2012 Wiley Periodicals, Inc.

KEY WORDS: nitro-Mannich reaction; chiral amines; nitro compounds; nucleophilic addition; asymmetric catalysis; copper; chiral ligands

INTRODUCTION

The nucleophilic addition of nitroalkanes to the CN of imines, known as the aza-Henry (or nitro-Mannich) reaction, is a useful carbon-carbon bond-forming reaction of prime importance.^{1–3} The resulting β -nitroamines can be readily transformed into valuable structural motifs such as 1,2-diamines by reduction⁴⁻⁶ or into *α*-amino carbonyl compounds by oxidation (Nef reaction).⁷⁻⁹ Most significantly, up to two new stereogenic centers can be formed, both attached to two different nitrogen functional groups. As a result, much attention has been paid to the catalytic asymmetric version of this reaction over the past several years by either using metal-based procedures^{10-15,6,17-19} or organocatalysis.²⁰⁻³⁹ One of the difficulties that arises when carrying out nucleophilic addition reactions to imines is their lower electrophilicity compared with the carbonyl group. However, this reactivity can be enhanced by attaching strong electron-withdrawing groups to the azomethinic nitrogen atom.^{40,41} In the reported aza-Henry reactions, *N*-carbamoyl imines have been the most commonly used electrophiles.^{11,12,17,19,20,22–31} However, a common However, a common drawback associated with the use of this type of imines is that precautions have to be taken during preparation, handling, and storage because of their substantial hydrolytic lability. N-phosphinoyl imines have been seldom used^{10,21,32} because of their poor stability and relatively low reactivity, although the phosphinoyl moiety can be easily removed by acidic hydrolysis. Similarly, there are few reports on the enantioselective aza-Henry reaction of N-sulfonyl protected imines,^{13,6,16} which are usually bench-stable solids and can be easily prepared. A possible explanation for this is the difficult deprotection of the N-tosyl amines resulting from the nucleophilic addition to the CN bond.

Our group has developed new iminopyridine and aminopyridine ligands (Fig. 1), which in the presence of copper(II) salts catalyzed the Henry reaction with carbonyl compounds very efficiently and with very high enantioselectivity.^{42–49} In this article, we describe the application of these catalysts in the enantioselective aza-Henry reaction with *N*-(2-pyridyl)sulfonyl aldimines.

EXPERIMENTAL Materials and Methods

Commercial reagents were used as purchased. Reactions were monitored by thin layer chromatography (TLC) analysis using Merck © 2012 Wiley Periodicals, Inc.

Silica Gel 60 F-254 thin layer plates. Flash column chromatography was performed on Merck silica gel 60, 0.040-0.063 mm (Merck KGaA, Darmstadt, Germany). Specific optical rotations were recorded on a Perkin-Elmer 241 polarimeter using sodium light (D line 589 nm). NMR spectra were recorded on Bruker Avance spectrometers in deuterated solvents as stated, using residual non-deuterated solvent as internal standard. The carbon type was determined by distortionless enhancement by polarization transfer (DEPT) experiments. Mass spectra (EI) were recorded on a Fisons Instruments VG Autospec GC 8000 series at 70 eV. Electrospray ionization mass spectra (ESI) were recorded on a Waters Q-TOF premier mass spectrometer with an electrospray source. The drying gas as well as the nebulizing gas was nitrogen. Chiral high performance liquid chromatography (HPLC) analyses were performed in a Hitachi Elite Lachrom instrument equipped with a Hitachi UV diode-array L-4500 detector using chiral stationary columns from Daicel and are included in the Supporting information. Retention times (t_r) are given in minutes. 2-Pyridylsulfonamide and 2-methoxyphenylsulfonamide were prepared from the corresponding sulfonyl chlorides according to known procedures. $^{50-52}$ Pyridine-2-sulfonyl chloride was prepared by oxidation of 2-pyridine-2-thiol (Alfa Aesar, Karlsruhe, Germany) with sodium hypochlorite in sulfuric acid.⁵⁰⁻⁵² 2-Methoxybenzenesulfonyl chloride was prepared from 2-methoxybenzenethiol (Sigma-Aldrich, St Louis, MO, USA) following the same procedure in concentrated hydrochloric acid instead of sulfuric acid. Ligands 1-5 were prepared according to our reported procedure.48

Synthesis of Starting Materials

Synthesis of N-sulfonyl imines 6–9. *N*-Sulfonyl imines **6–9** were prepared according to procedures in the literature:⁵² A mixture of the corresponding sulfonamides (11.4 mmol), aldehyde (11.4 mmol), activated 5Å molecular sieves (Sigma-Aldrich, St Louis, MO, USA) (1.70 g), and Amberlyst[®] (Sigma-Aldrich, St Louis, MO, USA) (225 mg) in toluene (35 mL) contained in a round bottom flask provided with a Dean–Stark system was refluxed under nitrogen for 48–60 h. The reaction mixture was cooled to room temperature (rt) and filtered through a sintered-glass

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Fig. 1. Iminopyridine and aminopyridine ligands used in this study.

funnel, washing the solid with dichloromethane (40 mL). The filtrates were evaporated under reduced pressure to give a solid that was washed with 1:1 diethyl ether/hexane (3×7 mL) to give the imine.

(*E*)-*N*-benzylidene-4-methylbenzenesulfonamide (6)⁵³. Obtained in 93% yield: mp 109–111 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.03 (1H, s), 7.97–7.88 (4H, m), 7.62 (1H, tt, *J*=6.6, 1.2 Hz), 7.49 (2H, t, *J*=7.8 Hz), 7.35 (2H, d, *J*=8.1 Hz), 2.44 (3H, s).

(*E*)-*N*-benzylidenethiophene-2-sulfonamide (**7**)⁵². Obtained in 93% yield: mp 108–109 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.01 (1H, s), 7.97–7.94 (2H, m), 7.80 (1H, dd, *J*=3.6, 1.2 Hz), 7.71 (1H, dd, *J*=5.1, 1.2 Hz), 7.64 (1H, tt, *J*=6.9, 1.2 Hz), 7.51 (2H, t, *J*=7.2 Hz), 7.15 (1H, dd, *J*=5.1, 3.9 Hz).

(*E*)-*N*-benzylidene-2-methoxybenzenesulfonamide (8)⁵⁴. Obtained in 63% yield: mp 115–116 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.14 (1H, s), 8.12 (1H, dd, *J*=8.1, 1.7 Hz), 7.97–7.94 (2H, m), 7.66–7.55 (2H, m), 7.50 (2H, t, *J*=7.2 Hz), 7.13 (1H, td, *J*=7.8, 0.6 Hz), 6.97 (1H, d, *J*=8.4 Hz), 3.82 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 173.0 (CH), 157.1 (C), 135.6 (CH), 134.9 (CH), 132.5 (C), 131.3 (CH), 130.7 (CH), 129.2 (CH), 125.3 (C), 120.7 (CH), 112.3 (CH), 56.2 (CH₃).

(*E*)-*N*-benzylidenepyridine-2-sulfonamide (9a)⁵⁰. Obtained in 87% yield: mp 142–144 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 9.31 (1H, s), 8.77 (1H, dq, *J* = 4.5, 1.2 Hz), 8.23–8.15 (2H, m), 8.10–8.08 (2H, m), 7.78–7.72 (2H, m), 7.59 (2H, t, *J* = 7.8 Hz); ¹³C NMR (75.5 MHz, DMSO- d_6) δ 174.9 (CH), 154.9 (C), 150.5 (CH), 139.1 (CH), 135.6 (CH), 132.0 (C), 131.5 (CH), 129.3 (CH), 128.0 (CH), 122.8 (CH); MS (EI) *m/z* (%): 247 [(M⁺+H), 2.8], 181 (14), 79 (100); HRMS: 247.0547 (M⁺+H), C₁₂H₁₁N₂O₂S required 247.0541.

(E)-N-(2-methoxybenzylidene)pyridine-2-sulfonamide (9b)⁵⁵.

Obtained in 83% yield: mp 116–118 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.74 (1H, s), 8.73 (1H, dq, *J*=4.5, 0.9 Hz), 8.23 (1H, dt, *J*=7.8, 0.9 Hz), 8.07 (1H, dd, *J*=7.8, 1.8 Hz), 7.95 (1H, td, *J*=7.8, 1.8 Hz), 7.59 (1H, td, *J*=7.8, 1.8 Hz), 7.51 (1H, ddd, *J*=7.8, 4.8 1.2 Hz), 6.98 (2H, m), 3.93 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 170.2 (CH), 162.1 (C), 156.1 (C), 150.3 (CH), 138.0 (CH), 137.5 (CH), 129.5 (CH), 127.1 (CH), 123.2 (CH), 120.84 (CH), 120.82 (C), 111.5 (CH), 55.7 (CH₃); MS (EI) *m/z* (%): 277 [(M⁺+H), 20], 181 (12), 79 (100); HRMS: 277.0645 (M⁺+H), C₁₃H₁₃N₂O₃S required 277.0647.

(E)-N-(2-methylbenzylidene)pyridine-2-sulfonamide (9c).

Obtained in 75% yield: mp 109–111 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.55 (1H, s), 8.72 (1H, d, *J*=4.5 Hz), 8.24 (1H, d, *J*=7.8 Hz), 8.05 (1H, dd, *J*=8.4, 1.5 Hz), 7.97 (1H, td, *J*=7.8, 1.5 Hz), 7.55–7.47 (2H, m), 7.30–7.25 (2H, m), 2.64 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 172.7 (CH), 162.1 (C), 155.9 (C), 150.3 (CH), 142.8 (C), 138.1 (CH), 135.1 (CH), 131.6 (CH), 130.7 (CH), 130.3 (C), 127.2 (CH), 126.6 (CH), 123.3 (CH), 19.6 (CH₃); MS (EI) *m/z* (%): 261 [(M⁺+H), 0.5], 118 (59), 79 (100); HRMS: 261.0686 (M⁺+H), C₁₃H₁₃N₂O₂S required 261.0698. *Chirality* DOI 10.1002/chir

(E)-N-(2-chlorobenzylidene)pyridine-2-sulfonamide (9d).

Obtained in 81% yield: mp 152–155 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.71 (1H, s), 8.76 (1H, dq, *J*=4.5, 0.9 Hz), 8.25 (1H, d, *J*=8.1 Hz), 8.18 (1H, dd, *J*=8.1, 1.8 Hz), 7.99 (1H, td, *J*=7.8, 1.8 Hz), 7.56 (2H, m), 7.49 (1H, m), 7.34 (1H, t, *J*=7.5 Hz); ¹³C NMR (75.5 MHz, CDCl₃) δ 170.7 (CH), 155.5 (C), 150.5 (CH), 139.4 (C), 138.1 (CH), 136.1 (CH), 130.6 (CH), 130.4 (CH), 129.7 (C), 127.38 (CH), 127.35 (CH), 123.5 (CH); MS (EI) *m/z* (%): 281 [(M⁺+H), 0.7], 79 (100); HRMS: 281.0141 (M⁺+H), C₁₂H₁₀ClN₂O₂S required 281.0152.

(*E*)-*N*-(4-methoxybenzylidene)pyridine-2-sulfonamide (9e)⁵⁵.

Obtained in 77% yield: mp 110–112 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.15 (1H, s), 8.70 (1H, dq, *J*=4.8, 0.9 Hz), 8.22 (1H, dt, *J*=8.1, 0.9 Hz), 7.95 (1H, td, *J*=7.8, 1.8 Hz), 7.93 (1H, d, *J*=8.7 Hz), 7.51 (1H, ddd, *J*=7.8, 4.8, 1.2 Hz), 6.79 (2H, d, *J*=8.7 Hz), 3.88 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 173.2 (CH), 165.7 (C), 156.2 (C), 150.3 (CH), 138.0 (CH), 134.2 (CH), 127.0 (CH), 125.1 (C), 123.1 (CH), 114.7 (CH), 55.7 (CH₃).

(*E*)-*N*-(4-methylbenzylidene)pyridine-2-sulfonamide (9f)⁵⁶. Obtained in 83% yield: mp 144–145 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.20 (1H, s), 8.71 (1H, dq, *J*=4.8, 0.9 Hz), 8.23 (1H, dt, *J*=7.8, 0.9 Hz), 7.96 (1H, td, *J*=7.8, 1.8 Hz), 7.85 (1H, d, *J*=8.4 Hz), 7.52 (1H, ddd, *J*=7.8, 4.8, 0.9 Hz), 7.29 (2H, d, *J*=8.4 Hz), 2.43 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 174.1 (CH), 155.9 (C), 150.4 (CH), 147.0 (C), 138.1 (CH), 131.8 (CH), 130.0 (CH), 129.7 (C), 127.2 (CH), 123.3 (CH), 22.0 (CH₃).

(*E*)-*N*-(4-chlorobenzylidene)pyridine-2-sulfonamide (9g)⁵⁶. Obtained in 83% yield: mp 163–165 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.22 (1H, s), 8.73 (1H, dq, *J*=4.8, 0.6 Hz), 8.25 (1H, dt, *J*=7.8, 0.6 Hz), 7.99 (1H, td, *J*=7.8, 1.8 Hz), 7.92 (1H, d, *J*=8.7 Hz), 7.55 (1H, ddd, *J*=7.8, 4.8, 1.2 Hz), 7.49 (2H, d, *J*=8.7 Hz); ¹³C NMR (75.5 MHz, CDCl₃) δ 172.9 (CH), 155.5 (C), 150.4 (CH), 142.0 (C), 138.2 (CH), 132.7 (CH), 130.6 (C), 129.7 (CH), 127.4 (CH), 123.4 (CH).

(E)-N-(4-nitrobenzylidene)pyridine-2-sulfonamide (9h)⁵⁷. Obtained in 82% yield: mp 192–194 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 10.17 (1H, s), 8.72 (1H, dq, J=4.8, 0.9 Hz), 8.42 (2H, d, J=8.7 Hz), 8.17 (2H, d, J=8.7 Hz), 8.07 (1H, td, J=7.8, 1.8 Hz), 7.93 (1H, dt, J=7.8, 0.9 Hz), 7.64 (1H, ddd, J=7.8, 4.8, 1.2 Hz); ¹³C NMR (75.5 MHz, DMSO- d_6) δ 192.3 (CH), 159.7 (C), 150.6 (C), 149.6 (CH), 140.0 (C), 138.5 (CH), 130.6 (CH), 126.6 (CH), 124.2 (CH), 120.4 (CH).

(E)-N-(3-methoxybenzylidene)pyridine-2-sulfonamide

(9i)⁵⁸. Obtained in 75% yield: mp 95–97°C; ¹H NMR (300 MHz, CDCl₃) δ 9.19 (1H, s), 8.71 (1H, dq, J=4.6, 0.9 Hz), 8.22 (1H, dt, J=7.8, 0.9 Hz), 7.96 (1H, dt, J=6.0, 1.8 H, m), 7.52 (1H, ddd, J=0.9, 4.6, 8.1 Hz), 7.50–7.45 (2H, m), 7.39 (1H, t, J=8.1 Hz), 7.16 (1H, ddd, J=1.3, 2.6, 8.1 Hz), 3.81 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 174.3 (d), 160.1 (s), 155.8 (s), 150.4 (d), 133.6 (s), 130.1 (d), 127.3 (d), 125.8 (d), 123.4 (d), 122.8 (d), 113.4 (d), 55.5 (q).

(*E*)-*N*-(3-methylbenzylidene)pyridine-2-sulfonamide (9j)⁵⁰. Obtained in 80% yield: mp 141–143 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.22 (1H, s), 8.73 (1H, dq, *J*=4.6, 0.9 Hz), 8.26 (1H, dt, *J*=7.8, 0.9 Hz), 7.97 (1H, dt, *J*=6.0, 1.8 H, m), 7.82 (1H, br s), 7.76 (1H, br d, *J*=7.5 Hz), 7.53 (1H, ddd, *J*=0.9, 4.6, 7.8 Hz), 7.46 (1H, br d, *J*=7.8 Hz), 7.39 (1H, t, *J*=7.5 Hz), 2.40 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 174.6 (d), 155.9 (s), 150.4 (d), 139.2 (s), 138.1 (d), 136.3 (d), 132.3 (s), 131.6 (d), 129.5 (d), 129.1 (d), 127.2 (d), 123.3 (d), 21.1 (q).

(*E*)-*N*-(3-chlorobenzylidene)pyridine-2-sulfonamide (9k)⁵⁷.

Obtained in 75% yield: mp 133–135°C; ¹H NMR (300 MHz, CDCl₃) δ 9.22 (1H, s), 8.73 (1H, dq, *J*=4.5, 0.9 Hz), 8.26 (1H, dt, *J*=7.8, 0.9 Hz), 8.02–7.97 (2H, m), 7.83 (1H, dt, *J*=7.5, 1.2 Hz), 7.61 (1H, dq, *J*=7.8, 0.9 Hz), 7.55 (1H, ddd, *J*=7.8, 4.8, 1.2 Hz), 7.46 (1H, t, *J*=7.8 Hz); ¹³C NMR (75.5 MHz, CDCl₃) δ 172.9 (CH), 155.5 (C), 150.5 (CH), 138.2 (2xCH), 135.5 (C), 135.1 (CH), 134.4 (C), 130.5 (CH), 130.4 (CH), 123.5 (CH); MS (EI) m/z (%): 281 [(M⁺+H), 10], 215 (12), 154 (13), 111 (26), 79 (100); HRMS: 281.0139 (M⁺+H), C₁₂H₁₀ClN₂O₂S required 281.0152.

(E)-N-(3-nitrobenzylidene)pyridine-2-sulfonamide(91).Obtained in 52% yield: mp 114–116 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 9.49 (1H, s), 8.87 (1H, t, J= 1.9 Hz), 8.79 (1H, dq, J= 4.5, 0.8 Hz), 8.55–8.46(2H, m), 8.25 (dt, J= 6.6, 0.9 Hz), 8.20 (dt, J= 7.8, 1.5 Hz), 7.89 (1H, t,J= 7.8 Hz), 7.79 (ddd, J= 8.7, 3.0, 1.5 Hz); ¹³C NMR (300 MHz, DMSO- d_6) δ 173.0 (d), 155.7 (s), 150.6 (d), 148.2 (s), 139.2 (d), 136.7 (d), 134.9 (s),136.7 (d), 134.9 (s), 130.9 (d), 129.2 (d), 128.3 (d), 125.6 (d), 123.1 (d);HRMS (ESI): 314.0228 (M⁺ + Na), C₁₂H₉N₃NaO₄S required 314.02115.

(E)-N-(thiophen-3-ylmethylene)thiophene-2-sulfonamide (9m).

Obtained in 64% yield: mp 161–163 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.21 (1H, s), 8.71 (1H, dq, J= 4.5, 0.9 Hz), 8.24–8.21 (2H, m), 7.96 (1H, td, J= 7.8, 1.5 Hz), 7.61 (1H, dd, J= 5.9, 0.9 Hz), 7.52 (1H, ddd, J= 7.8, 4.8, 1.2 Hz), 7.39 (1H, ddd, J= 5.1, 2.7, 0.6 Hz); ¹³C NMR (75.5 MHz, CDCl₃) δ 167.0 (CH), 155.9 (C), 150.4 (CH), 139.3 (CH), 138.1 (CH), 137.0 (C), 127.9 (CH), 127.2 (CH), 126.6 (CH), 123.3 (CH); HRMS (ESI): 274.9925 (M⁺ + Na), C₁₀H₈N₂NaO₂S₂ required 274.9925.

(E)-N-(furan-3-ylmethylene)pyridine-2-sulfonamide (9n).

Obtained in 70% yield: mp 146–147 °C; ¹H NMR (300 MHz, CDCl₃) δ 9.20 (1H, s), 8.71 (1H, dq, *J*=4.5, 0.9 Hz), 8.21 (1H, dt, *J*=7.8, 0.9 Hz), 8.16 (1H, t, *J*=0.9 Hz), 7.96 (1H, td, *J*=7.8, 1.5 Hz), 7.54–750 (2H, m), 6.85 (1H, d, *J*=1.8 Hz); ¹³C NMR (75.5 MHz, CDCl₃) δ 165.9 (CH), 155.8 (C), 152.9 (CH), 150.4 (CH), 145.5 (CH), 138.1 (CH), 127.2 (CH), 123.4 (C), 123.3 (CH), 108.0 (CH); HRMS (ESI): 259.0151 (M⁺ + Na), C₁₀H₈N₂NaO₃S required 259.0153.

Asymmetric Reaction

A solution of amino pyridine 2 (9.7 mg, General procedure. 0.04 mmol) in dry diethyl ether (Sigma-Aldrich, St. Louis, MO, USA) (1.2 mL) was added to a suspension of previously dried Cu(OTf)₂ (Alfa Aesar, Karlsruhe, Germany) (14.4 mg, 0.04 mmol) and activated 4Å MS (Sigma-Aldrich, St. Louis, MO, USA) (10 mg) in nitromethane (Sigma-Aldrich, St. Louis, MO, USA) (0.6 mL) contained in a Schlenk tube under nitrogen. The mixture was stirred for 1 h at rt until the formation of a deep blue solution. The imine 9 (0.2 mmol) dissolved in dry diethyl ether: nitromethane (0.24 mL:0.16 mL) was added via syringe, and the reaction flask was introduced in a bath at the reaction temperature. After 10 min, N,N-diisopropylethylamine (DIPEA) (Sigma-Aldrich, St. Louis, MO, USA) (10 µL, 0.06 mmol) dissolved in diethyl ether (0.2 mL) was added, and the reaction mixture was stirred for the indicated time. The mixture was filtered through a short pad of silica gel eluting with hexane/EtOAc (6:4). The filtrate was concentrated under reduced pressure and chromatographed eluting with hexane/EtOAc mixtures to give the corresponding products.

(-)-(*R*)-4-methyl-*N*-(2-nitro-1-phenylethyl)benzenesulfonamide (10). Obtained according to the conditions in Table 1. Enantiomeric excess (17%) was determined by HPLC (Chiralcel OD-H), hexane–*i*-PrOH 90:10, 1 mL/min, major enantiomer (*R*) t_r = 51.8, minor enantiomer (*S*) t_r = 45.0.

mp 155–157 °C; $[\alpha]_D^{25}$ –14.5 (*c* 0.11, CH₂Cl₂, *ee* 18%), Lit.⁶ $[\alpha]_D^{25}$ 73.5 (*c* 0.11, CH₂Cl₂, for the S-enantiomer, *ee* 91%), ¹H NMR (300 MHz, CDCl₃) δ 7.64 (2H, d, *J* = 8.4 Hz), 7.26–7.20 (5H, m), 7.12–7.08 (2H, m), 5.69 (1H, d, *J* = 7.8 Hz), 5.00 (1H, q, *J* = 6.9 Hz), 4.84 (1H, dd, *J* = 12.9, 6.6 Hz), 4.66 (1H, dd, *J* = 12.9, 6.3 Hz), 2.40 (3H, s).

(-)-*N*-(2-nitro-1-phenylethyl)thiophene-2-sulfonamide (11). Obtained according to the conditions in Table 1. Enantiomeric excess (8%) was determined by HPLC (Chiralcel OD-H), hexane–*i*-PrOH 85:15, 1 mL/min, major enantiomer t_r = 44.5, minor enantiomer t_r = 32.0.

TABLE 1. Addition of nitromethane to *N*-sulfonyl imines according to Scheme 1. Effect of the *N*-sulfonyl group^a

Entry	R	Imine	<i>t</i> (h)	Product	Yield (%)	ее (%) ^ь
1	<i>p</i> -Ts	6	24	10	99	18
2	2-Thiophenyl	7	22	11	80	11
3	2-Methoxyphenyl	8	16	12	75	23
4	2-Pyridyl	9a	5	13a	80	59

^aLigand **2** (20 mol%), Cu(OTf)₂ (20 mol%), DIPEA (30 mol%), 4 Å MS (100 mg/ mmol imine), THF (1.64 mL), nitromethane (0.76 mL), 0 °C. ^bDetermined by HPLC using chiral stationary phases.

mp 116–118 °C; $[\alpha]_{D}^{25}$ –0.69 (*c* 0.51, CH₂Cl₂, *ee* 8%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.98 (1H, br s), 7.74 (1H, dd, *J*=4.8, 1.2 Hz), 7.29–7.20 (6H, m), 6.92 (1H, dd, *J*=4.8, 3.6 Hz), 5.08 (1H, m), 4.86 (1H, dd, *J*=13.2, 5.1 Hz), 4.68 (1H, dd, *J*=13.2, 9.9 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 141.6 (C), 136.0 (C), 132.6 (CH), 131.8 (CH), 128.4 (CH), 128.1 (CH), 127.2 (CH), 126.8 (CH), 78.9 (CH₂), 55.8 (CH); MS (EI) *m/z* (%): 312 (M⁺, 0.1), 251 (17), 146 (100), 104 (22); HRMS: 312.0238, C₁₂H₁₂N₂O₄S required 312.0238.

(-)-2-methoxy-*N*-(2-nitro-1-phenylethyl) benzenesulfonamide (12). Obtained according to the conditions in Table 1. Enantiomeric excess (23%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 80:20, 1 mL/min, major enantiomer t_r = 19.8, minor enantiomer t_r = 16.5.

[α] $_{\rm D}^{25}$ –2.5 (c 0.90, CH₂Cl₂, ee 23%); ¹H NMR (300 MHz, CDCl₃) δ 7.87 (1H, dd, *J*=7.8, 1.8 Hz), 7.46 (1H, td, *J*=8.4, 1.8 Hz), 7.18 (3H, m), 7.02 (3H, m), 6.73 (1H, d, *J*=8.4 Hz), 5.98 (1H, t, *J*=8.4 Hz), 4.88 (2H, m), 4.63 (1H, m); ¹³C NMR (75.5 MHz, CDCl₃) δ 155.8 (C), 135.4 (C), 134.9 (CH), 130.1 (CH), 128.91 (CH), 128.86 (CH), 126.6 (C),126.5 (CH), 120.4 (CH), 111.7 (C), 78.7 (CH₂), 55.9 (CH₃), 55.6 (CH); HRMS (ESI): 359.0676 (M⁺+Na), C₁₅H₁₆N₂NaO₅S required 359.0678.

(-)-*N*-(2-nitro-1-phenylethyl)pyridine-2-sulfonamide (13a). Enantiomeric excess (83%) was determined by HPLC (Chiralpak AD-H), hexane-*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r =23.8, minor enantiomer (*S*) t_r =29.7.

mp 114–116 °C; $[\alpha]_D^{25}$ –17.1 (*c* 0.52, CH₂Cl₂, *ee* 83%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.96 (1H, d, *J*=9.0 Hz), 8.49 (1H, dq, *J*=4.5, 0.9 Hz), 7.86 (1H, td, *J*=7.8, 1.8 Hz), 7.70 (1H, d, *J*=7.8 Hz), 7.48 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.24–7.14 (5H, s), 5.16 (1H, m), 4.85 (1H, dd, *J*=13.2, 5.4 Hz), 4.78 (1H, dd, *J*=13.2, 9.6 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 157.3 (C), 149.6 (CH), 138.1 (CH), 136.4 (C), 128.2 (CH), 127.9 (CH), 126.9 (CH), 126.6 (CH), 121.2 (CH), 78.7 (CH₂), 55.7 (CH); MS (EI) *m/z* (%): 261 [(M⁺-NO₂), 0.1], 149 (30), 102 (58), 94 (89), 91 (57), 78 (91), 77 (73), 51 (100); HRMS: 261.0695, C₁₃H₁₃N₂O₂S required 261.0700.

(-)-*N*-(1-(2-methoxyphenyl)-2-nitroethyl)pyridine-2-sulfonamide (13b). Enantiomeric excess (78%) was determined by HPLC (Chiralpak AD-H), hexane-*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r = 38.0, minor enantiomer (*S*) t_r = 44.2.

mp 134–135 °C; $[\alpha]_{25}^{25}$ –23.6 (*c* 0.48, CH₂Cl₂, *ee* 78%); ¹H NMR (300 MHz, CDCl₃) δ 8.39 (1H, dq, *J*=4.5, 0.9 Hz), 7.81 (1H, dt, *J*=7.8, 0.9 Hz), 7.72 (1H, td, *J*=7.5, 1.5 Hz), 7.29 (1H, ddd, *J*=7.5, 4.8, 1.2 Hz), 7.15 (1H, td, *J*=8.4, 1.8 Hz), 6.93 (1H, dd, *J*=7.2, 1.5 Hz), 6.71 (2H, m), 6.36 (1H, d, *J*=9.9 Hz), 5.28 (1H, m), 4.93 (1H, dd, *J*=12.9, 7.5 Hz), 4.72 (1H, dd, *J*=12.9, 6.6 Hz), 3.84 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 157.1 (C), 156.5 (C), 149.7 (CH), 137.6 (CH), 130.2 (CH), 129.7 (CH), 126.5 (CH), 122.3 (C), 121.8 (CH), 120.8 (CH), 110.7 (CH), 77.6 (CH₂), 55.4 (CH), 55.1 (CH₃); HRMS (ESI): 360.0631 (M⁺+Na), C₁₄H₁₅N₃NaO₅S required 360.0630.

(-)-N-(2-nitro-1-o-tolylethyl)pyridine-2-sulfonamide (13c).

Enantiomeric excess (58%) was determined by HPLC (Chiralpak AD-H), hexane-*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r =21.1, minor enantiomer (*S*) t_r =24.8.

mp 109–101 °C; $[\alpha]_{D}^{25}$ –3.3 (*c* 0.49, CH₂Cl₂, *ee* 58%); ¹H NMR (300 MHz, CDCl₃) δ 8.48 (1H, dq, *J*=5.4, 0.9 Hz), 7.82–7.71 (2H, m), 7.40–7.36 (1H, m), 7.17–7.13 (1H, m), 7.10–6.96 (3H, m), 6.65 (1H, d, *J*=6.9 Hz), 5.54 (1H, q, *J*=6.9 Hz), 4.89 (1H, dd, *J*=13.5, 7.8 Hz), 4.68 (1H, dd, *J*=13.5, 6.6 Hz), 2.27 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 157.0 (C), 149.8 (CH), 138.0 (CH), 135.6 (C), 133.4 (C), 130.8 (CH), 128.7 (CH), 126.8 (CH), 126.7 (CH), 126.1 (CH), 121.9 (CH), 78.3 (CH₂), 51.8 (CH), 19.0 (CH₃); HRMS (ESI): 344.0683 (M⁺+Na), C₁₄H₁₅N₃NaO₄S required 344.0681.

(-)-N-(1-(2-chlorophenyl)-2-nitroethyl)pyridine-2-sulfonamide

(13d). Enantiomeric excess (54%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 70:30, 1 mL/min, major enantiomer (*R*) t_r = 13.7, minor enantiomer (*S*) t_r = 33.1.

mp 127–128 °C; $[\alpha]_{25}^{25}$ –13.5 (*c* 0.27, CH₂Cl₂, *ee* 54%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 9.17 (1H, d, *J*=9.3 Hz), 8.45 (1H, dq, *J*=4.8, 0.9 Hz), 7.87 (1H, td, *J*=7.8, 1.8 Hz), 7.72 (1H, dt, *J*=7.8, 0.9 Hz), 7.46 (1H, ddd, *J*=7.8, 4.8, 1.2 Hz), 7.46–7.42 (1H, m), 7.32–7.29 (1H, m), 7.20–7.13 (2H, m), 5.63 (1H, dt, *J*=9.9, 4.2 Hz), 4.82 (1H, dd, *J*=13.2, 4.2 Hz), 4.65 (1H, dd, *J*=13.2, 9.9 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 156.8 (C), 149.6 (CH), 138.1 (CH), 133.8 (C), 131.3 (C), 129.7 (CH), 129.1 (CH), 128.6 (CH), 127.2 (CH), 126.7 (CH), 121.1 (CH), 77.7 (CH₂), 52.3 (CH); HRMS (ESI): 342.0323 (M⁺+Na), C₁₃H₁₂CIN₃NaO₄S required 342.0315.

(-)-N-(1-(4-methoxyphenyl)-2-nitroethyl)pyridine-2-sulfonamide

(13e). Enantiomeric excess (66%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r = 31.1, minor enantiomer (*S*) t_r = 36.4.

mp 110–112 °C; $[\alpha]_D^{25}$ –43.4 (*c* 0.47, CH₂Cl₂, *ee* 66%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.82 (1H, d, *J*=9.0 Hz), 8.51 (1H, dq, *J*=4.5, 0.9 Hz), 7.87 (1H, td, *J*=7.8, 1.8 Hz), 7.68 (1H, dt, *J*=7.8, 0.9 Hz), 7.49 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.12 (2H, d, *J*=8.7 Hz), 6.70 (2H, d, *J*=8.7 Hz), 5.06 (1H, q, *J*=8.1 Hz), 4.86–4.77 (2H, m), 3.67 (3H, s); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 158.8 (C), 157.4 (C), 149.6 (CH), 138.1 (CH), 128.3 (C), 128.2 (CH), 126.5 (CH), 121.3 (CH), 113.6 (CH), 78.8 (CH₂), 55.2 (CH), 55.0 (CH₃); HRMS (ESI): 360.0640 (M⁺+Na), C₁₄H₁₅N₃NaO₅S required 360.0630.

(-)-N-(2-nitro-1-p-tolylethyl)pyridine-2-sulfonamide (13f).

Enantiomeric excess (60%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r =23.2, minor enantiomer (*S*) t_r =27.6.

mp 144–145 °C; $[\alpha]_D^{25}$ –29.8 (*c* 0.58, CH₂Cl₂, *ee* 60%); ¹H NMR (300 MHz, CDCl₃) δ 8.55 (1H, dq, *J*=4.8, 0.9 Hz), 7.87 (1H, dt, *J*=7.5, 0.9 Hz), 7.80 (1H, td, *J*=7.8, 1.5 Hz), 7.43 (1H, ddd, *J*=7.2, 4.8, 1.5 Hz), 7.07–7.00 (4H, m) 6.25 (1H, d, *J*=7.5 Hz), 5.22 (1H, q, *J*=6.6 Hz), 4.92 (1H, dd, *J*=13.2, 6.6 Hz), 4.77 (1H, dd, *J*=13.2, 6.3 Hz), 2.27 (3H, s); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 157.4 (C), 149.6 (CH), 138.1 (CH), 137.2 (C), 133.5 (C), 128.7 (CH), 126.8 (CH), 126.5 (CH), 121.2 (CH), 78.7 (CH₂), 55.4 (CH), 20.5 (CH₃); HRMS (ESI): 344.0675 (M⁺+Na), C₁₄H₁₅N₃NaO₄S required 344.0681.

(-)-N-(1-(4-chlorophenyl)-2-nitroethyl)pyridine-2-sulfonamide

(13g). Enantiomeric excess (66%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 80:20, 1 mL/min, major enantiomer (*R*) t_r = 26.8, minor enantiomer (*S*) t_r = 37.7.

mp 163–165 °C; $[\alpha]_D^{25}$ –35.9 (c 0.48, CH₂Cl₂, ee 66%); ¹H NMR (300 MHz, DMSO- d_6) δ 9.00 (1H, d, J=9.3 Hz), 8.50 (1H, dq, J=4.8, 0.9 Hz), 7.89 (1H, td, J=7.8, 1.5 Hz), 7.71 (1H, dt, J=7.8, 0.9 Hz), 7.51 *Chirality* DOI 10.1002/chir

(1H, ddd, J=7.8, 4.8, 0.9 Hz), 7.27–7.21 (4H, m), 5.16–5.08 (1H, m), 4.86 (1H, dd, J=13.2, 5.4 Hz), 4.77 (1H, dd, J=13.2, 9.6 Hz); ¹³C NMR (75.5 MHz, DMSO- d_6) δ 157.2 (C), 149.7 (CH), 138.2 (CH), 135.5 (C), 132.6 (C), 129.0 (CH), 128.2 (CH), 126.6 (CH), 121.4 (CH), 78.3 (CH₂), 55.0 (CH); HRMS (ESI): 342.0319 (M⁺ + Na), C₁₃H₁₂ClN₃NaO₄S required 342.0315.

(-)-N-(2-nitro-1-(4-nitrophenyl)ethyl)pyridine-2-sulfonamide

(13h). Enantiomeric excess (58%) was determined by HPLC (Chiralpak AD-H), hexane–*i*-PrOH 70:30, 1 mL/min, major enantiomer (*R*) t_r =30.9, minor enantiomer (*S*) t_r =45.5.

mp 192–194 °C; $[\alpha]_{D}^{25}$ –27.2 (*c* 0.14, CH₂Cl₂, *ee* 58%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 9.17 (1H, d, *J*=9.3 Hz), 8.48 (1H, dq, *J*=4.8, 0.9 Hz), 8.06 (2H, d, *J*=9.0 Hz), 7.89 (1H, td, *J*=7.8, 1.8 Hz), 7.73 (1H, dt, *J*=7.8, 0.9 Hz), 7.55 (2H, d, *J*=9.0 Hz), 7.49 (1H, ddd, *J*=7.8, 4.8, 1.2 Hz), 5.29 (1H, m), 4.94 (1H, dd, *J*=13.5, 5.1 Hz), 4.82 (1H, dd, *J*=13.5, 9.6 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 157.0 (C), 149.7 (CH), 147.0 (C), 144.0 (C), 138.3 (CH), 128.6 (CH), 126.8 (CH), 123.2 (CH), 121.3 (CH), 77.9 (CH₂), 54.9 (CH); HRMS (ESI): 353.0556 (M⁺+Na), C₁₃H₁₂N₄NaO₆S required 353.0556.

(-)-*N*-(1-(3-methoxyphenyl)-2-nitroethyl)pyridine-2-sulfonamide (13i). Enantiomeric excess (69%) was determined by HPLC (Chiralpak IC), hexane-*i*-PrOH 70:30, 1 mL/min, major enantiomer (*R*) t_r = 38.8, minor enantiomer (*S*) t_r = 53.3.

mp 123–124 °C; $[\alpha]_D^{25}$ –32.7 (*c* 0.52, CH₂Cl₂, *ee* 69%); ¹H NMR (300 MHz, CDCl₃) δ 8.52 (1H, dq, *J*=4.5, 0.9 Hz), 7.84–7.80 (1H, m), 7.77 (1H, td, *J*=7.2, 1.8 Hz), 7.40 (1H, ddd, *J*=7.2, 4.8, 1.5 Hz), 7.10 (1H, t, *J*=7.8 Hz), 6.84 (1H, d, *J*=7.8 Hz), 6.76–6.68 (3H, m), 5.30–5.22 (1H, m), 4.92 (1H, dd, *J*=13.5, 7.5 Hz), 4.73 (1H, dd, *J*=13.5, 6 Hz), 3.69 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 159.8 (C), 157.2 (C), 149.8 (CH), 138.0 (CH), 136.6 (C), 130.0 (CH), 126.8 (CH), 122.2 (CH), 118.9 (CH), 114.6 (CH), 112.2 (CH), 78.6 (CH₂), 55.8 (CH), 55.2 (CH₃); HRMS (ESI): 338.0805 (M⁺+H), C₁₄H₁₆N₃O₅S required 338.0810.

(-)-N-(2-nitro-1-m-tolylethyl)pyridine-2-sulfonamide (13j).

Enantiomeric excess (70%) was determined by HPLC (Chiralcel OD-H), hexane-*i*-PrOH 85:15, 1 mL/min, major enantiomer (*R*) t_r =29.3, minor enantiomer (*S*) t_r =27.3.

mp 140–141 °C; $[\alpha]_D^{25}$ –51.9 (*c* 0.47, CH₂Cl₂) (*ee* 70%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.89 (1H, d, *J*=8.1 Hz), 8.51 (1H, d, *J*=3.9 Hz), 7.85 (1H, td, *J*=7.5, 1.5 Hz), 7.68 (1H, d, *J*=7.8 Hz), 7.47 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.06–6.94 (4H, m), 5.06 (1H, br s), 4.87–4.74 (2H, m), 2.15 (3H, s); ¹³C NMR (75.5 MHz, DMSO) δ 157.9 (C), 150.0 (CH), 138.5 (CH), 137.9 (C), 136.7 (C), 129.0 (CH), 128.7 (CH), 128.1 (CH), 127.1 (CH), 124.5 (CH), 121.8 (CH), 79.2 (CH₂), 56.2 (CH), 21.3 (CH₃); HRMS (ESI): 344.0676 (M⁺ + Na), C₁₄H₁₅N₃NaO₄S required 344.0681.

(-)-N-(1-(3-chlorophenyl)-2-nitroethyl)pyridine-2-sulfonamide

(13k). Enantiomeric excess (60%) was determined by HPLC (Chiralcel OD-H), hexane–*i*-PrOH 75:25, 1 mL/min, major enantiomer (*R*) t_r =17.4, minor enantiomer (*S*) t_r =15.4.

mp 147–148 °C; $[\alpha]_{25}^{25}$ –29.2 (*c* 0.53, CH₂Cl₂, *ee* 60%); ¹H NMR (300 MHz, MeOD) δ 8.96 (1H, d, *J*=9.0 Hz), 8.49 (1H, dq, *J*=4.5, 0.9 Hz), 7.86 (1H, td, *J*=7.8, 1.8 Hz), 7.70 (1H, d, *J*=7.8 Hz), 7.48 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.24–7.14 (5H, s), 5.16 (1H, m), 4.85 (1H, dd, *J*=13.2, 5.4 Hz), 4.78 (1H, dd, *J*=13.2, 9.6 Hz); ¹³C NMR (75.5 MHz, MeOD) δ 157.3 (C), 149.6 (CH), 138.1 (CH), 136.4 (C), 128.2 (CH), 127.9 (CH), 126.9 (CH), 126.6 (CH), 121.2 (CH), 78.7 (CH₂), 55.7 (CH); HRMS (ESI): 364.0139 (M⁺+Na), C₁₃H₁₂ClN₃NaO₄S required 364.0135.

(-)-N-(2-nitro-1-(3-nitrophenyl)ethyl)pyridine-2-sulfonamide

(131). Enantiomeric excess (66%) was determined by HPLC (Chiralcel AD-H), hexane–*i*-PrOH 70:30, 1 mL/min, major enantiomer (*R*) t_r = 18.6, minor enantiomer (*S*) t_r = 16.7.

mp 166–168 °C; $[\alpha]_{D}^{25}$ –47.2 (c 0.4, CH₂Cl₂, ee 66%); ¹H NMR (300 MHz, DMSO-d₆) δ 9.2 (1H, br s), 8.44 (1H, ddd, *J* = 4.5, 1.5, 0.9 Hz), 8.13 (1H, t, *J* = 2.1 Hz), 8.02 (1H, ddd, *J* = 8.4, 2.4, 1.2 Hz), 7.85 (1H, td, *J* = 7.8, 1.8 Hz), 7.74–7.69 (2H, m), 7.50 (1H, t, *J* = 7.8 Hz), 7 (1H, ddd, *J* = 7.8, 4.8, 1.2 Hz), 5.31 (1H, br s), 4.95 (1H, dd, *J* = 13.8, 5.1 Hz), 4.84 (1H, dd, *J* = 13.8, 9.6 Hz); ¹³C NMR (75.5 MHz, DMSO-d₆) δ 157.5 (C), 150.2 (CH), 147.9 (C), 139.1 (C), 138.8(CH), 134.6 (CH), 130.4 (CH), 127.2 (CH), 123.4 (CH), 122.6 (CH), 121.9 (CH), 78.5 (CH₂), 55.4 (CH); HRMS (ESI): 375.0370 (M⁺+Na), C₁₃H₁₂N₄NaO₆S required 375.0375.

(-)-N-(2-nitro-1-(thiophen-3-yl)ethyl)pyridine-2-sulfonamide

(13m). Enantiomeric excess (77%) was determined by HPLC (Chiralpak AD-H), hexane-*i*-PrOH 75:25, 1 mL/min, major enantiomer (*R*) t_r = 18.5, minor enantiomer (*S*) t_r = 20.5.

mp 135–136 °C; $[\alpha]_D^{25}$ –32.8 (*c* 0.24, CH₂Cl₂, *ee* 77%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.84 (1H, d, *J*=9.3 Hz), 8.53 (1H, dq, *J*=4.5, 0.9 Hz), 7.91 (1H, td, *J*=7.8, 1.5 Hz), 7.74 (1H, d, *J*=7.8 Hz), 7.52 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.33–7.28 (2H, m), 6.94 (1H, dd, *J*=5.1, 1.5 Hz), 5.27–5.19 (1H, m), 4.87 (1H, dd, *J*=13.2, 5.4 Hz), 4.78 (1H, dd, *J*=13.2, 9.3 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 157.3 (C), 149.6 (CH), 138.2 (CH), 137.2 (C), 126.6 (CH), 126.5 (CH), 126.2 (CH), 123.4 (CH), 121.2 (CH), 78.5 (CH₂), 51.5 (CH); HRMS (ESI): 336.0095 (M⁺ + Na), C₁₁H₁₁N₃NaO₄S₂ required 336.0089.

(-)-N-(1-(furan-3-yl)-2-nitroethyl)pyridine-2-sulfonamide

(13n). Enantiomeric excess (60%) was determined by HPLC (Chiralpak AY-H), hexane–*i*-PrOH 70:30, 1 mL/min, major enantiomer (*R*) t_r = 32.8, minor enantiomer (*S*) t_r = 27.9.

mp 152–154 °C; $[\alpha]_D^{25}$ –15.0 (*c* 0.25, CH₂Cl₂, *ee* 60%); ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.72 (1H, d, *J*=9.0 Hz), 8.60 (1H, dq, *J*=4.8, 0.9 Hz), 7.98 (1H, td, *J*=7.8, 1.8 Hz), 7.82 (1H, dt, *J*=7.8, 0.9 Hz), 7.57 (1H, ddd, *J*=7.5, 4.5, 0.9 Hz), 7.43–7.42 (2H, m), 6.32 (1H, s), 5.16–5.08 (1H, m), 4.83 (1H, dd, *J*=12.9, 5.7 Hz), 4.76 (1H, dd, *J*=12.9, 8.7 Hz); ¹³C NMR (75.5 MHz, DMSO-*d*₆) δ 157.4 (C), 149.7 (CH), 143.4 (CH), 140.4 (C), 138.3 (CH), 126.8 (CH), 121.4 (C), 121.3 (CH), 108.9 (CH), 78.4 (CH₂), 48.0 (CH); HRMS (ESI): 320.0316 (M⁺ + Na), C₁₁H₁₁N₃NaO₅S required 320.0317.

Determination of the Absolute Stereochemistry of 13a

Synthesis of (-)-(R)-methyl 2-phenyl-2-(pyridine-2-sulfonamido) acetate (14) by ozonolysis of compound 13a. To a solution of 13a (50.0 mg, 0.16 mmol) in MeOH (3 mL) under nitrogen was added a 30% solution of NaOMe in MeOH (46.3 µL, 0.24 mmol), and the resulting solution was stirred at rt for 10 min. This methanolic solution was cooled to $-78\,^{\circ}$ C, and a stream of O₃ was passed through during 60 min. The reaction was purged with N2 to remove the excess of O3. The resulting solution was allowed to reach rt, BF₃·Et₂O (0.4 mL) was added, and the mixture was heated at reflux temperature for 3.5 h. After cooling to rt, saturated aqueous NaHCO3 (10 mL) was added, the organic solvent was removed under reduced pressure, and the resulting aqueous solution was extracted with dichloromethane $(3 \times 15 \text{ mL})$, washed with brine (10 mL), dried over anhydrous MgSO₄, and concentrated under reduced pressure. The mixture was purified by column chromatography eluting with hexane/EtOAc (7:3) to give 35.7 mg (71%) of (R)-14. Enantiomeric excess (83%) was determined by HPLC (Chiralcel OD-H), hexane: i-PrOH 80:20, 1 mL/min, major enantiomer (R) $t_r = 13.4$, minor enantiomer (S) $t_r = 12.4$.

[α] $_{\rm D}^{25}$ –50.2 (c 0.53, CHCl₃, ee 83%). ¹H NMR (300 MHz, CDCl₃) δ 8.59 (1H, dq, J=4.5, 0.9 Hz), 7.82–7.74 (2H, m), 7.43–7.39 (1H, m), 7.29–7.23 (5H, m), 6.13 (1H, d, J=8.1 Hz), 5.40 (1H, d, J=8.1 Hz), 3.67 (3H, s); ¹³C NMR (75.5 MHz, CDCl₃) δ 170.5, (C), 157.8 (C), 149.7 (CH), 137.8 (CH), 135.4 (C), 128.7 (CH), 128.5 (CH), 127.2 (CH), 126.5 (CH), 121.7 (CH), 60.1 (CH), 53.0 (CH₃); HRMS (ESI): 329.0576 (M⁺+Na), C₁₄H₁₄N₂NaO₄S required 329.0572. Synthesis of (+)-(S)-methyl 2-phenyl-2-(pyridine-2-sulfonamido) acetate (14) from (S)-(+)-phenylglycine methyl ester hydrochloride. A solution of pyridine-2-sulfonyl chloride (195 mg, 1.1 mmol) in CH₂Cl₂ (10 mL) was added to a solution of (*S*)-(+)-phenylglycine methyl ester hydrochloride (15, 208 mg, 1 mmol) and Et₃N (277 µL, 2 mmol) in CH₂Cl₂ (40 mL) cooled to 0 °C. The resulting mixture was allowed to reach rt and was stirred for 2 h. The reaction mixture was washed with saturated aqueous NaHCO₃ (2 × 25 mL) water (25 mL). The organic layer was dried over anhydrous MgSO₄ and evaporated under reduced pressure. Column chromatography eluting with hexane/EtOAc (6:4) gave 281 mg (92%) of (S)-14. Enantiomeric excess (99%) was determined by HPLC (Chiralcel OD-H), hexane:*i*-PrOH 80:20, 1 mL/min, major enantiomer (*S*) t_r = 12.3, minor enantiomer (*R*) t_r = 13.9. [α]²⁵ +113.7 (*c* 0.59, CHCl₃, *ee* 99%).

(-)-(*R*)-methyl 2-amino-2-phenylacetate (16). To a solution of (*R*)-14 (30.0 mg, 0.098 mmol, *ee* 83%) in MeOH (2.5 mL) and THF (0.65 mL) under nitrogen at 0 °C was added Mg powder (23.6 mg, 0.98 mmol), and the resulting suspension was stirred at rt for 4 h. Then, diethyl ether (3.8 mL) and a saturated aqueous NH₄Cl (3.8 mL) were added, and the resulting solution was stirred for additional 2 h at rt. The reaction mixture was diluted with water (15 mL) and extracted with diethyl ether (3 × 15 mL). The combined organic layers were dried over anhydrous MgSO₄ and concentrated under reduced pressure. Chromatography on a short pad of silica gel gave 14.5 mg (87%) of compound 16.

 $[\alpha]_{\rm D}^{25}$ –90.0 (c 0.05, MeOH, ee 83%), Lit 59 $[\alpha]_{\rm D}^{25}$ +130 (c 0.02, MeOH, for the S-enantiomer); $^1{\rm H}$ NMR (300 MHz, CDCl₃) δ 7.37–7.30 (5H, m), 4.64 (1H, s), 3.70 (3H, m), 2.56 (2H, br s).

RESULTS AND DISCUSSION

In the onset of our research, we tested the effect of different *N*-sulfonyl groups. Thus, *N*-tosyl- (**6**), *N*-(2-thiophenyl)sulfonyl- (**7**), *N*-(2-methoxyphenyl) sulfonyl- (**8**), and *N*-(2-pyridyl)sulfonyl- (**9**) imines derived from benzaldehyde were prepared, and their reaction with nitromethane in the presence of aminopyridine **2** (the best ligand for the enantioselective Henry reaction⁴⁴) and Cu(II) was studied (Scheme 1).

Initially, the reaction was carried out under conditions similar to those described by Feng,⁶ using ligand **2**, Cu (OTf)₂, catalytic DIPEA in THF-nitromethane, and 4Å molecular sieves. The results are gathered in Table 1.

The best results were obtained with the N-(2-methoxyphenyl) sulforyl imine **8** and especially with the N-(2-pyridyl)



Scheme 1. Copper-catalyzed addition of nitromethane to different N-sulfonyl imines.

 TABLE 2. Addition of nitromethane to N-sulfonyl imine 9a
 according to Scheme 1. Effect of ligand, amount of base

 and solvent^a
 and solvent^a

Entry	L	Solvent	Base	Yield (%)	ee (%) ^b
1	2	THF	DIPEA (30 mol%)	80	59
2	1	THF	DIPEA (30 mol%)	65	6
3	3	THF	DIPEA (30 mol%)	70	13
4	4	THF	DIPEA (30 mol%)	70	6
5	5	THF	DIPEA (30 mol%)	50	0
6	2	THF	DIPEA (20 mol%)	54	55
7	2	THF	DIPEA (10 mol%)	_	_
8	2	THF	DIPEA (50 mol%)	85	50
9	2	THF	DIPEA (100 mol%)	85	39
10	2	EtOH	DIPEA (30 mol%)	60	3
11	2	CH_2Cl_2	DIPEA (30 mol%)	65	26
12	2	Toluene	DIPEA (30 mol%)	85	57
13	2	CH_3NO_2	DIPEA (30 mol%)	95	38
14	2	Dioxane	DIPEA (30 mol%)	90	55
15	2	<i>t</i> -BuOMe	DIPEA (30 mol%)	90	64
16	2	<i>i</i> -Pr ₂ O	DIPEA (30 mol%)	90	64
17	2	Et_2O	DIPEA (30 mol%)	85	74

^aLigand (20 mol%), Cu(OTf)₂ (20 mol%), base, 4Å MS (100 mg/mmol imine), 0 °C.

^bDetermined by HPLC using chiral stationary phases.

sulfonyl imine **9a**. It is remarkable that both compounds **8** and **9a** have a potential coordinating group at the sulfone substituent, which may indicate that the reaction proceed through a chelated transition state.¹ As a matter of fact, the chelating character of the pyridine nitrogen has been shown to play an essential role in other metal-catalyzed enantioselective reactions with *N*-(2-pyridyl)sulfonyl imines.^{50–52}

Further optimization was carried out with N-(2-pyridyl)sulfonyl imine 9a. First, we tested the iminopyridine ligand 1 under the same conditions that allowed obtaining compound 13a with 65% yield but only 6% ee (Table 2, entry 2). Then, we focused our study on aminopyridine ligands 3-5 (entries 3-5), but none of them improved the results obtained with ligand 2. The optimal amount of base was found to be 30 mol%. Higher base loads had a deleterious effect on the enantioselectivity (entries 8-9), whereas when using a too low amount of base (10 mol%), the reaction did not take place (entry 7). The effect of the different solvents was also tested. Protic solvents (EtOH) provided the expected product almost in racemic form (entry 10). Toluene (entry 12) gave almost the same results as THF, whereas dichloromethane gave lower yield and ee (entry 11). Ether-type solvents gave the best performance, the best result being obtained in diethyl ether (entry 17).

Once the amount of the base (30 mol%) and the solvent (diethyl ether) had been established, we studied the reaction

TABLE 3. Addition of nitromethane to *N*-sulfonyl imine 9a according to Scheme 1. Effect of base and MS^a

Entry ^ь	Base	MS	Т (°С)	<i>t</i> (h)	Yield (%)	ee (%)°
1 ^d	DIPEA	4Å MS	0	5	85	74
2^{d}	DIPEA	3Å MS	0	5	80	56
3 ^d	DIPEA	5Å MS	0	5	85	49
4	DIPEA	4ÅMS	0	4	90	82
5	Et ₃ N	4ÅMS	0	4	90	79
6	Na ₂ CO ₃	4ÅMS	0	3	65	63
7	0BProton	4ÅMS	0	24	50	72
	Sponge [®]					
8	Bu ₃ N	4ÅMS	0	22	70	44
9	DIPEA	4ÅMS	-15	3	85	83
10	DIPEA	4ÅMS	-25	3	80	78
11	DIPEA	4Å MS	-40	3	80	79

^a**2** (20 mol%), Cu(OTf)₂ (20 mol%), base (30 mol%), MS (50 mg/mmol imine), Et₂O.

^bExperiments in entries 1–5 were not checked before the annotated times.

^cDetermined by HPLC using chiral stationary phases.

^d100 mg MS/mmol imine was used.

with different bases and with different kinds and amounts of molecular sieves (Table 3). Changing 4Å MS to 3 or 5Å MS led to a decrease in the enantiomeric excess of the product (entries 1–3). The amount of 4Å MS was also adjusted, so when the reaction was carried out with 50 mg MS per mmol of imine, an appreciable increase in enantioselectivity was observed with respect to the reaction in the presence of 100 mg MS per mmol of imine (entry 1 versus entry 4). Although the use of MS as an additive in several enantioselective reactions involving nitrocompounds^{6,12,13,22} has been shown to have a beneficial effect on the yield and stereoselectivity, the mechanism of this effect is not clear. It has been suggested that, further to act as scavenger for water and proton contaminants that may have a deleterious effect,^{12,60} MS could facilitate the enolization of nitromethane.^{61,62} It may also be possible that a heterogeneous catalyst is formed from the copper complex and molecular sieves, which may help to improve the stability of the catalyst and its stereodifferentiation ability.63,64 Other bases were tested, but none of them performed better than DIPEA. Finally, we tested the reaction at lower temperatures. A slight increase in the enantioselectivity was observed when the temperature was decreased from 0 to $-15 \,^{\circ}\text{C}$ (entry 9); however, further decrease of the temperature (entries 10-11) did not bring about any noticeable improvement of the reaction.

Once the best reaction conditions have been established, we studied the scope of the reaction with several N-(2-pyridyl)sulfonyl aldimines (Scheme 2). The results are gathered in Table 4.

In general, the reactions with aromatic and heteroaromatic⁶⁵ sulfonamides **9a–n** were completed in less than 3 h, providing the expected nitro sulfonamides **13a–n** with fair yields and moderate enantiomeric excesses. The highest *ee* values were obtained with the imines derived from benzaldehyde (entry 1), 2-methoxybenzaldehyde (entry 2), and thiophene-3-carbaldehyde (entry 10) with *ee* close to 80%. However, with the rest of imines, the *ee* values were lower, being around 60% in most cases.

Compounds **13** can be easily transformed into *N*-(2-pyridyl)sulfonyl amino acid derivatives. Thus, upon ozonolysis

¹During the reviewing process, one of the referees suggested that the reaction may not proceed through a chelated transition state, because imine **7**, bearing a thiophene ring with a potentially coordinating sulfur atom, gave poor results. Although the chelating character of N-(2-pyridyl)sulfonyl imines has been demonstrated in other enantioselective reactions, it cannot be excluded for the thiophene derivative. However, the size and bite angle of the potential chelate formed with the thiophene derivative may largely differ from those of the pyridine derivative, which may explain the different enantioselectivities obtained in entries 2 and 4 of Table 1.



Scheme 2. Enantioselective addition of nitromethane to N-sulfonyl imines 9.

TABLE 4. Addition of nitromethane to *N*-sulfonyl imines 9^a

Entry	9	R	<i>t</i> (h)	13	Yield (%)	ee (%) [•]
1	9a	Ph	3	13a	77	83
2	9b	2-MeOC ₆ H ₄	3	13b	88	78
3	9c	$2-MeC_6H_4$	3.5	13c	70	58
4	9d	$2-ClC_6H_4$	4	13d	64	54
5	9e	4-MeOC ₆ H ₄	3	13e	60	64
6	9f	$4 - MeC_6H_4$	3	13f	71	60
7	9g	$4-ClC_6H_4$	3	13g	67	66
8	9h	$4-NO_2C_6H_4$	3	13h	54	58
9	9i	$3-MeOC_6H_4$	3	13i	55	69
10	9j	$3-MeC_6H_4$	3	13j	63	70
11	9k	$3-ClC_6H_4$	3	13k	68	60
12	91	$3-NO_2C_6H_5$	3	131	50	66
13	9m	3-Thiophenyl	3	13m	70	77
14	9n	3-Furanyl	3	13n	72	60

 $^{\rm a}2$ (20 mol%), Cu(OTf)_2 (20 mol%), DIPEA (30 mol%), 4 Å MS (50 mg/mmol imine), Et_2O, $-15\,^{\circ}{\rm C}.$

^bDetermined by HPLC using chiral stationary phases.

of the nitronate of **13a** followed by esterification with methanol in the presence of BF_3 ·Et₂O, we could obtain directly the *N*-sulfonyl amino ester **14** (Scheme 3).

The absolute stereochemistry of compound 14, and hence that of compound 13a, was determined by comparison with the same product prepared by sulfonylation of commercially available (*S*)-(+)-methyl phenylglycinate (15) with 2-pyridylsulfonyl chloride (Scheme 3). The product prepared in this way showed opposite optical rotation sign to that prepared from 13a. In this way, the aza-Henry product 13a was assigned the *R* configuration at the stereogenic center. The rest of compounds 13b–n were tentatively assigned as *R* on the assumption of a uniform mechanistic pathway.²

One of the advantages of the *N*-(2-pyridyl)sulfonyl group is that it can be removed under milder conditions than other *N*-sulfonyl groups. Thus, compound **14** could be transformed into methyl phenylglycinate upon treatment with magnesium in MeOH–THF with good yield and without loss of optical purity (Scheme 4).

The results obtained with different amounts of base (Table 2, entries 1, 6–9) can be rationalized (Scheme 5) in terms very similar to those described by Jørgensen^{66,67} and for us⁴⁵ for the Cu(II)-catalyzed addition of nitromethane to α -keto esters. The enantioselective catalytic pathway requires



Scheme 3. Determination of the absolute stereochemistry of compound 13a.



Scheme 4. Deprotection of the amine.

the coordination of the *N*-(2-pyridyl)sulfonyl imine to the copper atom of the **2**-Cu(OTf)₂ complex and the presence of a deprotonated molecule of nitromethane (a nitronate). A competitive equilibrium between DIPEA and the initial **2**-Cu(OTf)₂ with an inactive **2**-CuOTf(DIPEA) is established. The reaction requires the presence in the solution of enough base to deprotonate the nitromethane. If the base is in excess with respect to the Lewis acid, the equilibrium is shifted toward the inactive complex, hence trapping the chiral Lewis acid. The remaining base induces a nonenantioselective pathway between the nitronate and the uncoordinated imine. If the Lewis acid is in excess with respect to the base, then the low concentration of free amine brings about a slow reaction with low conversion.

With respect to the stereochemical mechanism, the results indicate a preference for the nitronate to attack from the *Si* face of the CN double bond. According to our previous studies on the Henry reaction, $^{42-49}$ we propose a plausible stereochemical model that would account for the observed stereochemistry (Fig. 2). This model is similar to that proposed for the Henry reaction with aldehydes catalyzed by **2**-Cu(OAc)₂, ⁴⁴ but with the imine coordinating the two more acidic equatorial positions of the Cu(II) ion by the azomethine and pyridine nitrogen atoms, for a maximum electrophilic activation. The nitronate would occupy the less *Chirality* DOI 10.1002/chir

²All compounds **13** showed identical optical rotation signs (–). Compounds **13a-h** and **13n** also showed identical behavior during HPLC analysis with the Chiralpak AD-H column. Compounds **13j** and **13k** showed identical behavior with the Chiralcel OD-H column.





Scheme 5. Competitive catalytic and non-catalytic pathways.



Fig. 2. Proposed transition state model for the copper-catalyzed enantioselective aza-Henry reaction of imines 9 with ligand 2.

hindered apical position from which it would be transferred to the *Si* face of the imine.

CONCLUSIONS

In conclusion, we have described a new catalytic enantioselective aza-Henry reaction. The reaction expands the application of camphor-derived aminopyridine ligands in asymmetric catalysis. This is the first example of aza-Henry reaction with *N*-(2-pyridyl)sulfonyl imines, which have the advantage of an easy removal of the *N*-protecting group under mild reductive conditions.

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