BODP – A Versatile Reagent for Phospholipid Synthesis

Pierre-Léonard Zaffalon, Andreas Zumbuehl*

Department of Organic Chemistry, University of Geneva, Quai Ernest-Ansermet 30, 1211 Geneva, Switzerland Fax +41(22)3793215; E-mail: andreas.zumbuehl@unige.ch Received 11 January 2011

Abstract: Benzyloxydichlorophosphine (BODP) has been found to be a convenient reagent for the synthesis of phospholipids. A series of artificial ether and ester phospholipids have been prepared in good to high yields.

Key words: ethers, esters, phospholipids, phosphorylation, protecting group

Despite over 80 years of synthetic efforts, the preparation of phospholipids remains a challenge.¹ We have recently initiated a program for the synthesis of artificial phospholipids and found it necessary to investigate new reactive intermediates for the introduction of a phosphate head group.² Here, we report on a convenient reagent, benzyloxydichlorophosphine (BODP, 1), that was successfully used for the synthesis of various artificial choline phospholipids.

Typical ways to introduce a phosphocholine head group include both phosphorus(V) and phosphorus(III) reagents: for example, bromoethylphosphoric acid dichloride readily reacts with primary alcohols to give a stable intermediate that can be substituted to the choline using trimethylamine.³ Dioxophospholanes can be opened to the corresponding choline.⁴ Successful phosphorus(III) intermediates include H-phosphonates⁵ and reagents such as methyl dichlorophosphite⁶ and phosphoramidites,⁷ which allow a stepwise addition of two different alcohols.

On attempting to phosphorylate a secondary alcohol, we found that phosphorus(V) reagents are generally too unreactive and phosphorus(III) reagents lead to oxidation after the addition of the first alcohol. This and the cost of phosphoramidite reagents led us to look for an alternative, a middle way using a reagent that at the same time was stable towards oxidation, but also reactive enough to allow the stepwise addition of two different alcohols in one pot. Benzyloxydichlorophosphine fulfills these criteria and, to our knowledge, was never before used for the synthesis of phospholipids. The reagent was initially synthesized under rudimentary conditions that lead to explosions during the distillation.⁸ However, the reagent can be used without this purification step, is safe to handle,⁹ and requires only moderately dry working conditions under nitrogen gas (BOPD is routinely used by undergraduate students in our laboratory). The starting materials (benzyl alcohol and PCl₃) are readily available, cheap, and can be used without purification or drying. Furthermore, the benzyl group can be easily removed under various mild conditions.

BODP has been used as a reactive intermediate for the synthesis of ribonucleotides,⁹ phosphoramidates,¹⁰ polyphosphates,¹¹ and in cyclization reactions.¹² When being reacted with phosphonates and amides BODP – like other phosphorus(III) reagents – presumably primarily formed the monochlorophosphine intermediate and not the doubly substituted phosphine,^{9b,11b} making it less useful for symmetrical double substitution, but more interesting for stepwise addition of different nucleophiles. The reactivity is strongly dependent on the solvent used, with tetrahydrofuran being preferred over acetonitrile and dichloromethane.¹³

Here, BODP was successfully used in the synthesis of 3 phospholipids: miltefosine (4), Pet-PC-Pet (7), and Pes-PC-Pes (10). Others and we have recently suggested amendments to the existing IUPAC recommendation for phospholipid nomenclature.^{2,14} The new classification focuses on the type of chemical linkage between the phospholipid tails and the lipid backbone. 'Pes' thus designates a C₁₆ alkyl chain (P = palmitic) connected to the lipid backbone with an ester functionality. In 'Pet', the chemical linker is an ether. The order in which the substituents are attached to the glycerol backbone is represented by the order of the groups in the abbreviation: Pes-PC-Pes is a 1,3-substituted, and Pes-Pes-PC a natural, *sn*-1,2-substituted phospholipid.

Our nomenclature allows easy classification of artificial phospholipids based on their chemical linkage and is complementary to the impressive nomenclature of the International Lipid Classification and Nomenclature Committee.¹⁵

Benzyloxydichlorophosphine (1) was synthesized optimizing a reported procedure:^{12b} benzyl alcohol was reacted with phosphorus trichloride in diethyl ether. Upon completion of the reaction, the excess of diethyl ether was evaporated leaving the BODP that was used without further purification.

Miltefosine (hexadecylphosphocholine, **4**) is an impressive example for an interdisciplinary collaboration between basic and clinical research. Miltefosine was initially identified for its antitumor potency and is now used clinically for the treatment of visceral and cutaneous leishmaniasis.¹⁶ Miltefosine was synthesized by phosphorylation of hexadecan-1-ol with phosphorus oxychloride, ring closure with ethanolamine, and opening of the oxaza-

SYNTHESIS 2011, No. 5, pp 0778–0782 Advanced online publication: 08.02.2011 DOI: 10.1055/s-0030-1258427; Art ID: T10811SS © Georg Thieme Verlag Stuttgart · New York



Scheme 1 Synthesis of miltefosine (hexadecylphosphocholine, 4)

phospholane ring under acidic conditions. The alkylphosphoethanolamine was reacted to form the choline with dimethyl sulfate.¹⁷

Here, BODP (1) was reacted stepwise with hexadecan-1ol (2), followed by bromoethanol, and oxidation to the protected phosphate triester 3. Deprotection with trifluoroacetic acid and substitution with trimethylamine³ lead to the choline derivative 4 in two steps and an overall yield of 37% (Scheme 1).

Surprisingly, changing from a natural 1,2-diacyl phospholipid to a symmetrical 1,3-diacyl phospholipid does not significantly alter the physical properties of the molecule in a membrane.¹⁸ Similarly, changing between diacyl- and dialkylphosphocholines also does not introduce big alterations in the physical characteristics of the molecules.¹⁹ These facts should allow new possibilities to produce membrane probes at more reasonable prices and an efficient synthesis is certainly appreciated.

Pet-PC-Pet (7) had previously been accessed using bromoethyl dichlorophosphate that was substituted with trimethylamine.¹⁹ Here, we started from the corresponding 1,3-dialkylpropan-2-ol **5** that was obtained from the reaction of hexadecan-1-ol with epichlorohydrin.²⁰ The intermediate was reacted with BODP (1) and bromoethanol to give the protected phosphate triester **6**. Again, substitution with trimethylamine led to the product **7** (Scheme 2) in 48% overall yield. 1,3-Diacyl phosphocholines were introduced in order to study the differences between natural 1,2- and artificial 1,3-diacyl phospholipids.^{18,19}

The starting alcohol **8** was available from a reaction of palmitoyl chloride with glycerol.²¹ Reaction with BODP led to the reactive intermediate **9** that was deprotected and further substituted to Pes-PC-Pes (**10**) (Scheme 3). The compound was obtained in 26% yield which is comparable to earlier syntheses of Les-PC-Les (L = lauric).²²

In conclusion, we have presented a rapid and high-yielding approach leading to alkylphosphocholines, 1,3-dialkyl-, and 1,3-diacylphosphocholines. The synthesis is based on benzyloxydichlorophosphine, a reagent rarely used for phosphorylations, but with many advantages over phosphoramidite reagents such as cost, stability and ease of handling. The presented approach for phospholipid synthesis is flexible, allowing the introduction of various head groups. Synthetic efforts in this direction are currently ongoing in our group.

Starting compounds and solvents were purchased from Sigma-Aldrich/Fluka or Acros and were used without further purification. Column chromatographic separations were carried out using 230–400 mesh silica gel. TLC plates were developed with KMnO₄. ¹H, ¹³C, and ³¹P NMR spectra were recorded (as indicated) on either a Bruker 300 MHz, 400 MHz or 500 MHz spectrometer and are reported as chemical shifts (δ) in ppm relative to TMS (δ = 0). Spin multiplicities are reported as a singlet (s), doublet (d), or triplet (t)

Downloaded by: Universite Laval. Copyrighted material



Scheme 3 Synthesis of Pes-PC-Pes (10)

Synthesis 2011, No. 5, 778-782 © Thieme Stuttgart · New York

with coupling constants (*J*) given in Hz, or multiplet (m). Broad peaks are marked as br. HRESI-MS were performed on QSTAR Pulsar (AB/MDS Sciex) spectrometer and are reported as mass-percharge ratio m/z. IR spectra were recorded on a Perkin-Elmer Spectrum One FT-IR spectrometer (ATR, Golden Gate). MCPBA provided by Sigma was 77% pure.

Benzyloxydichlorophosphine (BODP, 1)

The protocol was optimized from a literature procedure.^{12b} Benzyl alcohol (6.00 mL, 58.0 mmol) was dissolved in anhyd Et₂O (120 mL) and added to PCl₃ (10.1 mL, 116 mmol) in anhyd Et₂O (90 mL) over 5 h. The first 20 min of the addition was performed at 0 °C, and then the ice bath was removed. After the addition, the stirring was continued for 2 h at 20 °C. The solvent was evaporated by distillation at 50 °C and the excess of PCl₃ was removed under reduced pressure. The product was used without further purification; colorless oil; yield: 11.4 g (94%).

¹H NMR (300 MHz, CDCl₃): δ = 7.47 (s, 5 H), 5.32 (d, *J* = 8.0 Hz, 2 H).

¹³C NMR (75 MHz, CDCl₃): δ = 134.8, 128.8, 128.5, 69.8.

³¹P NMR (162 MHz, CDCl₃): $\delta = -177.37$ (s).

Benzyl 2-Bromoethyl Hexadecyl Phosphate (3)

BODP (1; 1.00 mL, 6.10 mmol) and anhyd Et₃N (2.00 mL, 14.0 mmol) were dissolved in anhyd THF (20 mL) under N2. Hexadecan-1-ol (400 mg, 1.65 mmol) in anhyd THF (20 mL) was added dropwise to the solution under stirring at 0 °C over 3 h [completion of the reaction was checked by TLC (MeOH-CH₂Cl₂, 2:98)]. Then, bromoethanol (910 $\mu L,$ 12.8 mmol) was added at 0 $^{\circ}C$ and the mixture was stirred for 3 h (the reaction can be monitored by ³¹P NMR spectroscopy, the signal should shift from 166 ppm to about 140 ppm). MCPBA (1.56 g, 9.60 mmol) was added to the reaction mixture at 0 °C and the mixture was stirred for 1 h (oxidation can be monitored by a signal shift from 140 ppm to about -5 ppm in ³¹P NMR spectrum). The solution was treated with of aq 10% Na₂S₂O₃ (110 mL), sat. aq NaHCO₃ (30 mL), and extracted with CH₂Cl₂ $(3 \times 100 \text{ mL})$. The solvents from the combined organic phases were removed under reduced pressure. The crude product was purified on a silica gel column (eluent: pentane-EtOAc, 4:1) to give the pure product as a brownish oil; yield: 750 mg (88%); $R_f = 0.23$ (pentane– EtOAc, 4:1).

IR (ATR): 2923, 2853, 1457, 1379, 1270, 1215, 1000, 733, 696 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.37 (d, *J* = 7.2 Hz, 5 H), 5.10 (d, *J* = 8.5 Hz, 2 H), 4.25 (dd, *J* = 13.0, 6.5 Hz, 2 H), 4.04 (q, *J* = 6.8 Hz, 2 H), 3.47 (t, *J* = 6.3 Hz, 2 H), 1.63 (dd, *J* = 13.7, 6.7 Hz, 2 H), 1.25 (s, 25 H), 0.88 (t, *J* = 6.5 Hz, 3 H).

¹³C NMR (126 MHz, CDCl₃): δ = 135.7, 128.7, 128.0, 69.5, 68.3, 66.5, 32.0, 30.2, 30.2, 29.7, 29.7, 29.7, 29.6, 29.6, 29.5, 29.5, 29.4, 29.2, 25.4, 22.7, 14.2.

³¹P NMR (162 MHz, CDCl₃): $\delta = -0.63$ (s).

HRMS (ESI+): m/z [M + H]⁺ calcd for C₂₅H₄₅BrO₄P: 519.2236; found: 519.2233.

Hexadecylphosphocholine (4)

To a solution of **3** (292 mg, 0.562 mmol) in CH₂Cl₂ (2 mL) was added CF₃CO₂H (TFA, 98%, 3.00 mL, 40.3 mmol) and the mixture was stirred at r.t. for 20 h. Excess TFA was removed by bubbling N₂ through the mixture. The residue was partitioned between aq HCl (25 mL) and CH₂Cl₂ (4 × 25 mL). The solvent from the combined organic phases were removed under reduced pressure. The white solid obtained (280 mg, 100% conversion of **3**) was used without further purification. A portion of the solid (140 mg) was dissolved in a mixture of CH₂Cl₂ (1 mL), MeCN (1.6 mL), and *i*-PrOH (1.6 mL). Then, Me₃N (45% in H₂O, 2 mL) was added. After 3 h, the solvent was removed under vacuum. The crude product was purified on a silica gel column (eluent: CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3); white solid; yield: 56 mg (42%); $R_f = 0.1$ (CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3).

¹H NMR (400 MHz, $CDCl_3$): $\delta = 4.27$ (br, 2 H), 3.86 (br, 2 H), 3.78 (dd, J = 13.1, 6.6 Hz, 2 H), 3.41 (s, 9 H), 1.66–1.48 (m, 2 H), 1.25 (s, 26 H), 0.87 (t, J = 6.8 Hz, 3 H).

³¹P NMR (162 MHz, CDCl₃): $\delta = -0.40$ (s).

HRMS (ESI+): m/z [M + H]⁺ calcd for C₂₁H₄₇NO₄P: 408.3237; found: 408.3232.

1,3-Bis(hexadecyloxy)propan-2-ol (5)

NaH (60% in mineral oil, 525 mg, 13.1 mmol) was added in 3 portions to hexadecan-1-ol (6.63 g, 27.3 mmol) at 100 °C. After stirring for 30 min, epichlorohydrin (250 µL, 3.19 mmol) was added dropwise to the hot mixture and maintained at 100 °C overnight, after which H₂O (2 mL) was added to quench the reaction. CH₂Cl₂ (200 mL) was added, followed by THF to facilitate the solubility of the product, and the organic phase was washed with H₂O (3 × 150 mL). The organic phase was dried (Na₂SO₄), filtered, and the solvent removed under reduced pressure. Purification on a silica gel column (eluent: pentane–EtOAc, 5:95) afforded a white solid; yield: 1.06 g (61%); R_f = 0.16 (pentane–EtOAc, 95:5)

¹H NMR (300 MHz, CDCl₃): δ = 3.94 (s, 1 H), 3.60–3.26 (m, 8 H), 2.45 (s, 1 H), 1.57 (s, 4 H), 1.26 (s, 52 H), 0.88 (t, *J* = 5.3 Hz, 6 H). HRMS (ESI+): *m*/*z* [M + H]⁺ calcd for C₃₅H₇₃O₃: 541.5554; found: 541.5565.

Benzyl 1,3-Bis(hexadecyloxy)propan-2-yl 2-Bromoethyl Phosphate (6)

A solution of **5** (400 mg, 739 µmol) in anhyd THF (20 mL) was added dropwise to a solution of BODP (**1**; 405 µL, 2.44 mmol) and anhyd Et₃N (722 µL, 5.18 mmol) in anhyd THF (25 mL) over 3 h at 0 °C. The stirring was continued at 20 °C for 12 h. After the consumption of the starting material (TLC: pentane–EtOAc, 9:1), bromoethanol (370 µL, 5.18 mmol) was added dropwise at 0 °C. The ice-bath was removed and the stirring continued for 45 min. MCPBA (829 mg, 3.70 mmol) was added at 0 °C and the mixture was stirred for 1 h at 20 °C. Then, aq 10% Na₂S₂O₃ (110 mL) and sat. aq NaHCO₃ (50 mL) were added and the aqueous phase was extracted with CH₂Cl₂ (3 × 100 mL). The organic solvent was removed under reduced pressure and the product was purified on a silica gel column (eluent: pentane–EtOAc, 5:2) to give a yellowish solid; yield: 536 mg (89%); $R_f = 0.35$ (pentane–EtOAc, 4:1).

IR (ATR): 2922, 2853, 1462, 1378, 1278, 1117, 1013, 734, 697 cm⁻¹.

¹H NMR (500 MHz, CDCl₃): δ = 7.47–7.31 (m, 5 H), 5.14 (d, *J* = 7.6 Hz, 2 H), 4.70–4.59 (m, 1 H), 4.30 (dd, *J* = 14.8, 6.6 Hz, 2 H), 3.65–3.53 (m, 4 H), 3.53–3.35 (m, 6 H), 1.54 (tt, *J* = 13.7, 6.7 Hz, 4 H), 1.26 (s, 52 H), 0.89 (t, *J* = 7.0 Hz, 6 H).

¹³C NMR (126 MHz, CDCl₃): δ = 181.1, 175.2, 148.1, 135.9, 128.6, 128.5, 127.9, 71.7, 70.2, 69.4, 66.5, 62.8, 31.9, 29.7, 29.7, 29.6, 29.5, 29.5, 29.4, 29.3, 29.2, 26.1, 22.7, 14.1.

HRMS (Turbo Spray): m/z [M + H]⁺ calcd for C₄₄H₈₃BrO₆P: 817.5105; found: 817.5113.

Pet-PC-Pet (7)

To a solution of **6** (503 mg, 615 μ mol) in CH₂Cl₂ (7 mL) was added TFA (98%, 7.00 mL, 93.3 mmol). After stirring the mixture for 24 h, the TFA was removed by bubbling N₂ through the solution. The residual crude oil was partitioned between aq 1 M HCl (25 mL) and CH₂Cl₂ (4 × 25 mL). The solvent from the combined organic phases was removed under reduced pressure to give 315 mg (70% conver-

sion of **6**) of a white solid, which was used without further purification. A portion of the crude solid (285 mg) was dissolved in a mixture of CHCl₃ (8 mL), MeCN (5 mL), and *i*-PrOH (8 mL). Then, Me₃N (45% in H₂O, 4 mL) was added. After stirring for 48 h, the solvents were removed under reduced pressure and the crude product was purified on a silica gel column (eluent: CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3); white solid; yield: 213 mg (54%); R_f = 0.27 (CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3).

¹H NMR (500 MHz, CDCl₃): δ = 4.33 (s, 3 H), 3.81 (s, 2 H), 3.55 (d, *J* = 5.1 Hz, 4 H), 3.39 (s, 13 H), 1.61–1.43 (m, 4 H), 1.25 (s, 52 H), 0.87 (t, *J* = 6.8 Hz, 6 H).

HRMS (Turbo Spray): m/z [M + H]⁺ calcd for C₄₀H₈₅NO₆P: 706.6109; found: 706.6126.

2-Hydroxypropane-1,3-diyl Dipalmitate (8)¹⁸

A solution of palmitoyl chloride (1.70 mL, 5.45 mmol) in anhyd THF (12 mL) was added dropwise to a solution of glycerol (200 µL, 2.72 mmol) and pyridine (900 µL, 11.1 mmol) in anhyd THF (18 mL) at 0 °C under N₂ over 90 min. The stirring was continued at 20 °C for 1 h. Then, the solution was partitioned between aq 1 M HCl (100 mL) and Et₂O (3 × 150 mL). The product was recrystallized from Et₂O at 6 °C overnight. The white powder was filtered, dried, and used without further purification; yield: 540 mg (35%); $R_f = 0.7$ (CH₂Cl₂–MeOH, 49:1).

¹H NMR (400 MHz, CDCl₃): δ = 4.20–4.11 (m, 5 H), 2.41 (s, 1 H), 2.34 (t, *J* = 7.3 Hz, 2 H), 1.63–1.61 (m, 4 H), 1.25 (s, 48 H), 0.88 (t, *J* = 8.0 Hz, 6 H).

HRMS (ESI–): m/z calcd [M + MeCOO[–]][–] for $C_{37}H_{71}O_7$: 627.5205; found: 627.5205

2-[Benzyloxy(2-bromoethoxy)phosphoryloxy]propane-1,3-diyl Dipalmitate (9)

A solution of **8** (365 mg, 0.64 mmol) in anhyd THF (20 mL) was added dropwise to a solution of BODP (1; 460 µL, 2.76 mmol) and anhyd Et₃N (820 µL, 5.90 mmol) in anhyd THF (10 mL) over 90 min at 0 °C. The stirring continued at 20 °C for 5 h. After the consumption of the starting material (TLC: pentane–EtOAc, 5:2), bromoethanol (420 µL, 5.90 mmol) was added dropwise at 0 °C. Then the ice-bath was removed and the stirring continued for 45 min. MCPBA (733 mg, 3.27 mmol) was added at 0 °C and the solution was stirred for 1 h at 20 °C. Aq 10% Na₂S₂O₃ (110 mL) was added followed by sat. aq NaHCO₃ (50 mL). The aqueous phases were extracted with CH₂Cl₂ (4 × 100 mL). The solvents were removed from the combined organic phases and the crude product was purified on a silica gel column (eluent: pentane–EtOAc, 5:2) to give a yellow solid; yield: 275 mg (51%); $R_f = 0.5$ (pentane–EtOAc, 4:1).

IR (ATR): 2922, 2853, 1743, 1457, 1379, 1276, 1163, 1011, 735, 697 $\rm cm^{-1}.$

¹H NMR (500 MHz, CDCl₃): δ = 7.45–7.31 (m, 5 H), 5.11 (dd, J = 8.1, 1.7 Hz, 2 H), 4.81–4.74 (m, 1 H), 4.35–4.24 (m, 4 H), 4.18 (ddd, J = 12.1, 7.7, 5.9 Hz, 2 H), 3.47 (tdd, J = 6.4, 1.7, 0.5 Hz, 2 H), 2.37–2.29 (m, 2 H), 2.26 (td, J = 7.4, 1.6 Hz, 2 H), 1.78–1.40 (m, 4 H), 1.25 (s, 48 H), 0.88 (t, J = 7.0 Hz, 6 H).

¹³C NMR (126 MHz, CDCl₃): δ = 173.2, 135.4, 128.7, 128.7, 128.0, 74.2, 69.7, 66.8, 62.7, 34.0, 33.9, 31.9, 29.6, 29.3, 29.1, 24.7, 22.7, 14.1.

³¹P NMR (121 MHz, CDCl₃): $\delta = -2.60$ (s).

HRMS (ESI+): $m/z = [M + H]^+$ calcd for $C_{44}H_{79}BrO_8P$: 845.469; found: 845.4682.

Pes-PC-Pes (10)

To a solution of the phosphate triester **9** (55.0 mg, 65.0 μ mol) in CH₂Cl₂ (3 mL) was added TFA (98%, 3.00 mL, 40.0 mmol). After 36 h, the solvent was removed by bubbling N₂ through the vial and

the crude oil obtained was used directly in the next step. The oil was dissolved in a mixture of CHCl₃ (1.5 mL), *i*-PrOH, MeCN (1.5 mL), and Me₃N (45% in H₂O, 3 mL; 2 mL of additional Me₃N were added after 24 h) and the mixture was stirred for 36 h. The solvents were removed under reduced pressure and the crude product was purified on a silica gel column (eluent: CH₂Cl₂–MeOH–25% NH₄OH, 175:22:3, then CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3) to give the product as a white powder; yield: 24 mg (50%); R_f = 0.2 (CH₂Cl₂–MeOH–25% NH₄OH, 75:22:3).

¹H NMR (300 MHz, CDCl₃): δ = 4.49 (br s, 1 H), 4.33 (br s, 2 H), 4.24 (d, *J* = 4.9 Hz, 4 H), 3.80 (s, 2 H), 3.34 (s, 9 H), 2.29 (t, *J* = 7.6 Hz, 4 H), 1.71–1.46 (m, 4 H), 1.25 (s, 48 H), 0.88 (t, *J* = 6.7 Hz, 6 H).

HRMS (ESI+): m/z [M + H]⁺ calcd for C₄₀H₈₁O₈NP: 734.5694; found: 734.5728.

Acknowledgment

We would like to thank Mr. Frédéric Loosli for his help on the project during an undergraduate student internship. The authors thank the University of Geneva and the Swiss National Science Foundation for the financial support (SNF 200021-121718) and acknowledge the contributions of the Mass Spectrometry platform at the Faculty of Sciences, University of Geneva, for mass spectrometry services and A. Pinto for NMR measurements.

References

- (1) Grün, A.; Limpächer, R. Chem. Ber. **1926**, 59, 1350.
- (2) Fedotenko, I. A.; Zaffalon, O. L.; Favarger, F.; Zumbuehl, A. *Tetrahedron Lett.* 2010, *51*, 5382.
- (3) (a) Hirt, R.; Berchtold, R. *Pharm. Acta Helv.* **1958**, *33*, 349.
 (b) Eibl, H.; Nicksch, A. *Chem. Phys. Lipids* **1978**, *22*, 1.
- (4) Thuong, N.; Charbier, P. Bull. Soc. Chim. Fr. 1974, 667.
- (5) Liu, X.; Stocker, B. L.; Seeberger, P. H. J. Am. Chem. Soc. 2006, 128, 3638.
- (6) Martin, S. F.; Josey, J. A.; Wong, Y. L.; Dean, D. W. J. Org. Chem. 1994, 59, 4805.
- (7) Liu, H.; Zhu, Z.; Kang, H.; Wu, Y.; Sefan, K.; Tan, W. *Chem. Eur. J.* **2010**, *16*, 3791.
- (8) Razumov, A. I.; Rispolozhenskii, N. I. Trans. Kirov Inst. Chem. Tech. Kazan 1940, 8, 42; Chem. Abstr. 1941, 35, 15654.
- (9) (a) Ogilvie, K. K.; Theriault, N. Y.; Seifert, J. M.; Pon, R. T.; Nemer, M. J. *Can. J. Chem.* **1980**, *58*, 2686. (b) Ding, Y.; Wang, J.; Schuster, S. M.; Richards, N. G. J. *J. Org. Chem.* **2002**, *67*, 4372.
- (10) (a) Kitas, E. A.; Knorr, R.; Trzeciak, A.; Bannwarth, W. *Helv. Chim. Acta* **1991**, *74*, 1314. (b) Prestwich, G. D.; Marecek, J. F.; Mourey, R. J.; Theibert, A. B.; Ferris, C. D.; Danoff, S. K.; Snyder, S. H. *J. Am. Chem. Soc.* **1991**, *113*, 1822.
- (11) (a) Lowe, G.; Semple, G. J. Chem. Soc., Chem. Commun. 1988, 377. (b) Mons, S.; Klein, E.; Mioskowski, C.; Lebeau, L. Tetrahedron Lett. 2001, 42, 5439.
- (12) (a) Saady, M.; Valleix, A.; Lebeau, L.; Mioskowski, C. *J. Org. Chem.* **1995**, *60*, 3685. (b) Amigues, E. J.; Greenberg, M. L.; Ju, S.; Chen, Y.; Migaud, M. E. *Tetrahedron* **2007**, *63*, 10042.
- (13) (a) Beld, A.; Claesen, C. A. A.; Roersma, E. S.; Schippers,
 W. J. M.; Keizer, L. M.; Tesser, G. I. *Recl. Trav. Chim. Pays-Bas* 1984, *103*, 196. (b) Claesen, C. A.; Segers, R. P. A. M.;
 Tesser, G. I. *Recl. Trav. Chim. Pays-Bas* 1985, *104*, 119.
- (14) Huang, Z.; Szoka, F. C. J. Am. Chem. Soc. 2008, 130, 15702.
- (15) (a) Fahy, E.; Subramaniam, S.; Brown, H. A.; Glass, C. K.; Merrill, A. H. J. r.; Murphy, R. C.; Raetz, C. R. H.; Russell,

Synthesis 2011, No. 5, 778-782 © Thieme Stuttgart · New York

D. W.; Seyama, Y.; Shaw, W.; Shimizu, T.; Spener, F.; van Meer, G.; VanNieuwenhze, M. S.; White, S. H.; Witztum, J. L.; Dennis, E. A. *J. Lipid Res.* **2005**, *46*, 839. (b) Fahy, E.; Subramaniam, S.; Murphy, R. C.; Nishijima, M.; Raetz, C. R. H.; Shimizu, T.; Spener, F.; van Meer, G.; Wakelam, M. J. O.; Dennis, E. A. *J. Lipid Res.* **2009**, *50*, S9.

- (16) (a) Eibl, H.; Unger, C. *Cancer Treat. Rev.* **1990**, *17*, 233.
 (b) Jha, T. K.; Sundar, S.; Thakur, C. P.; Bachmann, P.; Karbwang, J.; Fischer, C.; Voss, A.; Berman, J. *New Engl. J. Med.* **1999**, *341*, 1795.
- (17) Seifert, K.; Duchene, M.; Wernsdorfer, W. H.; Kollaritsch, H.; Scheiner, O.; Wiedermann, G.; Hottkowitz, T.; Eibl, H. *Antimicrob. Agents Chemother.* **2001**, *45*, 1505.
- (18) Seelig, J.; Dijkman, R.; De Haas, G. H. *Biochemistry* **1980**, *19*, 2215.
- (19) Kunitake, T.; Okahata, Y.; Tawaki, S. I. J. Colloid Interface Sci. 1985, 103, 190.
- (20) Li, T.; Hamdi, J.; Hawthorne, M. F. *Bioconjugate Chem.* 2006, 17, 15.
- (21) Rose, W. G. J. Am. Chem. Soc. 1947, 69, 1384.
- (22) Haftendorn, R.; Ulbrich-Hofmann, R. *Tetrahedron* **1995**, *51*, 1177.