

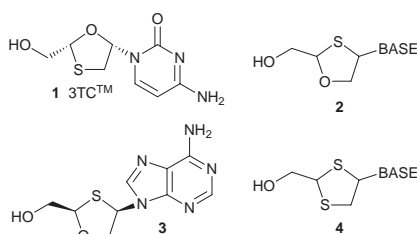
Synthesis and anti-HIV activity of 1,3-dithiolane nucleosides

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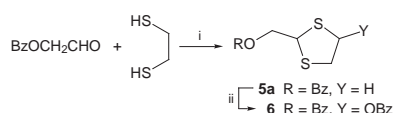
The potent activity displayed by 3'-azido-3'-deoxythymidine (AZT)¹ against human immunodeficiency virus (HIV) provides impetus for the development of novel nucleoside analogues.² Unfortunately, those compounds with the natural stereochemistry possess undesirable pharmacological properties³ and are susceptible to the development of resistant strains of HIV.^{3,4} In an attempt to overcome some of these detrimental side effects, the carbohydrate moiety of 2',3'-dideoxynucleoside analogs has been replaced by other five membered rings.^{3d,4} It has been demonstrated that hetero-substitution of these rings has a profound effect on the biological activity of the resulting nucleoside analogue⁵ as displayed by (–)-2'-deoxy-3'-thiacytidine (3TC, Efavir) **1**.^{5c,6}



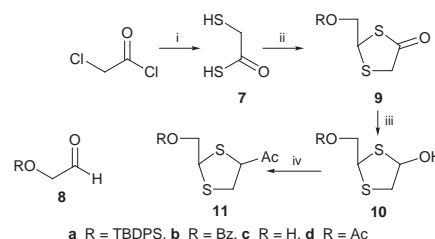
As part of an ongoing search for new anti-AIDS leads, we further explored this class of thioribonucleosides. These compounds possess improved metabolic stability to phosphorolases which cleave the glycosidic bond in nucleosides.⁷ Recently, we reported the anti-HIV activity of 2,4-disubstituted 1,3-oxathiolane nucleosides **2** by transposing the sulfur and oxygen atoms of **1**.^{4a} In this series, the (–)-adenine derivative **3** with the natural configuration was found to be twice as active as ddI in MT-4 cells. We further modified the oxathiolane ring by replacement of the oxygen atom of **2** with sulfur. This is exemplified by the general structure **4**. Here we report the synthesis and anti-HIV activity of this class of compounds.

The synthetic route to (±)-1,3-dithiolane compounds is based upon coupling a persilylated heterocyclic base with a dithiolane moiety **5** bearing a suitable leaving group Y at the 4-position under Vorbruggen's conditions.⁸ Two approaches were considered for the preparation of the ring **5**. The first route is the introduction of a leaving group at position 4 using peroxide derivatives. For example, dithiolane **6** was prepared by treating **5a** with benzoyl peroxide in refluxing benzene (Scheme 1).⁹

The second approach offers a more general route for the synthesis of the key intermediate 1,3-dithiolane **11**† with a variety of displaceable leaving groups at C-4. Our synthetic strategy was based on the preparation of 1,3-dithiolan-4-one **9**, followed by reduction and acylation to give 4-acyloxy-1,3-dithiolane **11** (Scheme 2). Thus, reaction of freshly distilled ClCH₂COCl with excess NaSH (3 equiv.) in absolute ethanol at –10 °C gave **7**¹⁰ in quantitative yield. The crude product was



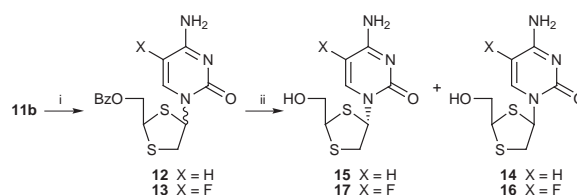
Scheme 1 Reagents and conditions: i, C₆H₆, TSA; ii, BzOOH, C₆H₆, heat.



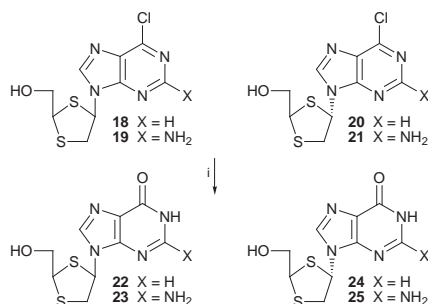
Scheme 2 Reagents and conditions: i, NaSH, absolute EtOH, –10 °C; ii, **8**, ZnI₂, CH₂Cl₂; iii, DIBAL-H, PhMe or BH₃·THF, B(OMe)₃; iv, AcOCl, Py.

immediately treated with aldehyde **8** in the presence of ZnI₂ as catalyst in CH₂Cl₂¹¹ to give the desired (±)-1,3-dithiolan-4-one **9** in moderate yield (50–60%). The initial synthetic procedure was based upon using the TBDPS protecting group for the hydroxy function of **9**. Reduction of **9a** with DIBAL-H (1.1 equiv.) in toluene gave thiolactol **10a** which was subsequently acylated to give the key intermediate **11a** in high yield. The silyl protecting group was later replaced by a benzoate group in order to facilitate the separation of the *cis* and *trans* nucleoside isomers. Applying the same conditions to reduce the benzoate **9b** did not result in any reduction product. However, using excess of DIBAL-H (3 equiv.) was successful and both the thiolactone and the benzoate function were reduced to give diol **10c** in 40% yield. Compound **10c** was then bis-acylated giving intermediate **11d** in high yield. Efforts were then directed to scale-up this procedure. Unfortunately, the DIBAL-H reduction proved particularly intractable. We therefore investigated other reducing agents that are selective and require little work-up. Only BH₃·THF (1.2 equiv.) catalyzed by B(OMe)₃ (1 equiv.) gave satisfactory results. The reduction was completed in 16 h and the product **10b** was obtained in 95% yield. This compound was then acylated to give the expected product **11b**. Following the same procedure, a number of different leaving groups (Bz, *m*-ClC₆H₄CH₂ and *p*-O₂NC₆H₄CH₂) were successfully introduced at C₄ of the sugar moiety **11**.

Compound **11b** is suitable for coupling with silylated cytosine or 5-fluorocytosine under refluxing conditions in CH₂Cl₂ and in the presence of SnCl₄ (Scheme 3). This gave the desired nucleoside analogue **12** or **13** as a 1 : 2 mixture of *cis* and *trans* isomers in moderate yields. Replacement of SnCl₄ with TMSI altered the ratio of the isomers. For example, compound **6** reacted with silylated N-acetylcytosine to give a mixture of the *cis* and *trans* nucleosides **12** in 62% yield with a slight predominance of the *cis* isomer.⁹ Similar results were obtained using other leaving groups at C₄. This did not improve the yield



Scheme 3 Reagents and conditions: i, 2,4-bis(trimethylsilyloxy)pyrimidine, ClCH₂CH₂Cl, SnCl₄, reflux, 16 h; ii, NH₃, MeOH.



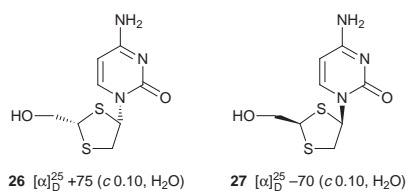
Scheme 4 Reagents and conditions: i, 25% aq. $\text{Me}_3\text{N}\cdot\text{H}_2\text{O}$, 55–65%.

or the *cis:trans* ratio. The next step was the separation of the isomers **12** or **13**. This was achieved by flash chromatography on silica gel by prior acetylation of the amino group (cytosine) or by reverse chromatography HPLC after deprotection (5-fluorocytosine). The protecting groups were then removed by treatment with methanolic ammonia to give the desired nucleosides **14–17** in high yields. The relative stereochemistry of these products was assigned by difference NOE spectra.

Similarly, uracil, thymine, adenine and guanine derivatives were produced from **11b** using the same conditions. However, in the case of hypoxanthine and guanine analogs **22–25**, synthesis was undertaken by treating the corresponding 6-chloropurine or 2-amino-6-chloropurine derivatives **18–21** with 20 equiv. of an aqueous Me_3N solution in water (Scheme 4).

The anti-HIV activity of (\pm)-1,3-dithiolane nucleoside analogues was evaluated in MT-4 (human T helper) cells at concentrations up to $100\ \mu\text{g ml}^{-1}$ and compared with 3TCTM (Epivir)¹² and the 5-fluoro derivative (FTC).¹³ In this assay, only *cis* cytosine and 5-fluorocytosine derivatives **14** and **16** displayed inhibitory activity at ID_{50} of 9.3 and $4.8\ \mu\text{g ml}^{-1}$ and were not cytotoxic at $100\ \mu\text{g ml}^{-1}$, whereas 3TCTM and FTC showed anti-HIV activity at 0.3 and $0.14\ \mu\text{g ml}^{-1}$, respectively. All the other nucleosides did not exhibit antiviral activity with no cytotoxicity up to $100\ \mu\text{g ml}^{-1}$. In contrast, *cis* and *trans* 6-chloropurine derivatives **18** and **20** showed cytotoxicity at CD_{50} of $10\ \mu\text{g ml}^{-1}$.

Described herein is a novel class of anti-HIV (\pm)-1,3-dithiolane nucleoside analogues. The biological results demonstrate that replacement of an oxygen atom of the oxathiolane with sulfur causes reduction in antiviral activity. It should be noted that compounds **14** and **16** are racemic. Resolution of the enantiomers may improve the activity. Therefore, enantiomeric separations of the racemic *cis* **14** was undertaken by chiral HPLC.¹⁴ This gave the two enantiomers **26** and **27** in a



reasonable yield. It was found that isomer **27** possesses the natural configuration as evidenced by enzymatic resolution of the racemic mixture. Thus treatment of the mixture of the two enantiomers with cytidine deaminase converted only **27** to its corresponding uracil derivative. However, the unnatural enantiomer **26** was recovered and characterized by comparison of HPLC retention time and optical rotation with the previously isolated isomer. Both enantiomers were submitted for anti-HIV evaluation. Neither of the two compounds displayed improved antiviral activity.

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Notes and references

† Selected data for **10b**: colorless oil; δ_{H} (CDCl_3): 8.03 (m, 2H), 7.57 (dt, 1H, J 7, 1), 7.44 (t, 2H, J 7), 5.85 (m, 1H), 4.84 and 4.76 (t's, 1H, J 7), 4.68 and 4.39 (m's, 1H), 4.48 and 4.27 (m's, 1H), 3.30 (m, 2H), 2.85 (m, 1H). For **16**: δ_{H} ($\text{DMSO}-d_6$) 8.34 (d, 1H, J 7.5), 7.83 (br s, 1H), 7.59 (br s, 1H), 6.39 (m, 1H), 5.57 (t, 1H, J 5.5), 4.62 (t, 1H, J 6), 3.75 (t, 2H, J 6), 3.59 (m, 2H). For **17**: δ_{H} ($\text{DMSO}-d_6$) 8.10 (d, 1H, J 7 Hz), 7.78 (br s, H), 7.54 (br s, 1H), 6.41 (t, 1H, J 2), 5.36 (t, 1H, J 5.5), 4.81 (t, 1H, J 7 Hz), 3.45 (m, 4H). For **19**: δ_{H} ($\text{DMSO}-d_6$) 8.45 (s, 1H), 7.03 (br s, 2H), 6.40 (t, 1H, J 4), 5.53 (t, 1H, J 6), 4.73 (t, 1H, J 7), 3.80 (dd, 1H, J 13, 4.5), 3.74 (t, 2H, J 6), 3.63 (dd, 1H, J 13, 5); δ_{C} ($\text{DMSO}-d_6$) 159.80, 153.49, 150.01, 141.95, 123.61, 65.57, 64.52, 56.23, 42.93; HRMS (FAB): M^+ calc. for $\text{C}_9\text{H}_{11}\text{ClN}_5\text{OS}_2$ 304.00937, found 304.00880. For **21**: δ_{H} ($\text{DMSO}-d_6$) 8.34 (s, 1H), 7.02 (br s, 1H), 6.44 (t, 1H, J 3), 5.41 (t, 1H, J 6), 4.86 (t, 1H, J 7), 3.58 (m, 3H), 3.49 (m, 1H); HRMS (FAB): M^+ calc. for $\text{C}_9\text{H}_{11}\text{ClN}_5\text{OS}_2$ 304.00937, found 304.00840. For **26**: mp 108–110 °C; δ_{H} ($\text{DMSO}-d_6$) 8.06 (d, 1H, J 6), 7.20 (br d, 2H, NH_2), 6.47 (t, 1H, J 4.4), 5.74 (d, 1H, J 7.42), 5.50 (t, 1H), 4.61 (t, 1H, J 6.5 Hz), 3.74 (t, 2H, J 6.00 Hz), 3.46 (dd, 1H, J 4.30, 12.9), and 3.36 (dd, 1H, J 4.1, 10.4); HRMS (FAB): M^+ calc. for $\text{C}_8\text{H}_{12}\text{N}_3\text{O}_2\text{S}_2$ 246.03709, found 246.03610. For **27**: mp 200–202 °C (decomp.); δ_{H} ($\text{DMSO}-d_6$) 7.90 (d, 1H, J 7.35), 7.17 (br d, 2H), 6.48 (d, 1H, J 3.72), 5.69 (d, 1H, J 7.47), 5.38 (t, 1H), 4.74 (t, 1H, J 6.83), 3.43 (m, 4H); HRMS (FAB): M^+ calc. for $\text{C}_8\text{H}_{12}\text{N}_3\text{O}_2\text{S}_2$ 246.03709, found 246.034640.

- H. Mitsuya, J. K. Weinhold, P. A. Furman, M. H. St-Clair, S. Nusinoff-Lehrman, R. C. Gallo, D. Bolognesi, D. W. Barry, S. Broder, *Proc. Natl. Acad. Sci. U.S.A.*, 1985, **82**, 7096.
- H. Mitsuya and S. Broder, *Proc. Natl. Acad. Sci. U.S.A.*, 1986, **83**, 1911; R. Yarchoan, H. Mitsuya, R. V. Thomas, J. M. Pluda, N. R. Hartman, C.-F. Perno, K. S. Marczyk, J.-P. Allain, D. G. Johns and S. Broder, *Science*, 1989, **245**, 412; T.-S. Lin, R. F. Schinazi and W. H. Prusoff, *Biochem. Pharmacol.*, 1987, **36**, 2713.
- (a) R. W. Klecker, J. M. Collins, R. Yarchoan, R. Thomas, J. F. Jenkins, S. Broder, C. E. Myers, *Pharmacol. Ther.*, 1987, **47**, 407; (b) M. S. Hirsch and J. C. Kaplan, *Antimicrob. Agents Chemother.*, 1987, **31**, 939; (c) R. Yarchoan, *Lancet*, 1988, **i**, 76; (d) M. L. Peterson and R. Vince, *J. Med. Chem.*, 1991, **34**, 2787 and references cited therein.
- (a) B. Belleau, L. Brasili, L. Chan, M. D. DiMarco, B. Zacharie, N. Nguyen-Ba, H. J. Jenkinson, J. A. V. Coates, J. M. Cameron, *Bioorg. Med. Chem. Lett.*, 1993, **3**, 1723; (b) M. J. Bamford, D. C. Humber, R. Storer, *Tetrahedron Lett.*, 1991, **32**, 271 and references cited therein; (c) J. Branalt and I. Kvarnstrom, *J. Org. Chem.*, 1996, **61**, 3604 and references cited therein.
- (a) T. S. Mansour, C. A. Evans, M. A. Siddiqui, M. Charron, B. Zacharie, N. Nguyen-Ba, N. Lee and B. Korba, *Nucleosides Nucleotides*, 1997, **16**, 993; (b) H. Soudey, X. J. Yao, Q. Gao, B. Belleau, J.-L. Kraus, N. Nguyen-Ba, B. Spira and M. A. Wainberg, *Antimicrob. Agents Chemother.*, 1991, **35**, 1386; (c) J. A. V. Coates, N. Cammack, H. J. Jenkinson, I. M. Mutton, B. A. Pearson, R. Storer, J. M. Cameron and C. R. Penn, *Antimicrob. Agents Chemother.*, 1992, **36**, 202; (d) M. W. Chun, D. H. Shin, H. R. Moon, J. Lee, H. Park and L. S. Jeong, *Bioorg. Med. Chem. Lett.*, 1997, **7**, 1475.
- U.S. Pat. 05047407, BioChem Pharma Inc.; M. A. Nowak, S. Bonhoeffer, A. M. Hill, R. Boehrme, H. C. Thomas and H. McDade, *Proc. Natl. Acad. Sci. U.S.A.*, 1996, **93**, 4398.
- J. A. Secrist III, K. N. Tiwari, A. T. Shortnacy-Fowler, L. Messini, J. M. Riordan and J. A. Montgomery, *J. Med. Chem.*, 1998, **41**, 3865; M. R. Dyson, P. L. Coe and R. T. Walker, *J. Med. Chem.*, 1991, **34**, 2782.
- H. Vorbruggen, K. Krolkiewicz and B. Bennua, *Chem. Ber.*, 1981, **114**, 1234.
- N. Nguyen-Ba, W. Brown, N. Lee and B. Zacharie, *Synthesis*, 1998, 759.
- M. Therien, J. Y. Gauthier and R. N. Young, *Tetrahedron Lett.*, 1988, **29**, 6733.
- J. Y. Gauthier, T. Henien, L. Lo, M. Therien and R. N. Young, *Tetrahedron Lett.*, 1988, **29**, 6729.
- J. M. Cameron, P. Collis, M. Daniel, R. Storer and P. Wilcox, *Drugs Future*, 1993, **18**, 319 and references cited therein.
- L. W. Frick, L. St-John, L. C. Taylor, G. R. Painter, P. A. Furman, D. C. Liotta, E. S. Furfine and D. J. Nelson, *Antimicrob. Agents Chemother.*, 1993, **37**, 2285.
- Chiral Column: Cyclobond I 2000 Beta-RSP 4.6 mm ID \times 250 mm; mobile phase 10% MeCN–0.05% (AcOH– Et_3N , pH 6.74); pressure 965 psi and flow rate $0.50\ \text{ml min}^{-1}$. For **26**: t_{R} = 19.880 min; for **27**: t_{R} = 22.201 min.

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