## Article

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## Pt-Mal-LHRH, a Newly Synthesized Compound Attenuating Breast Cancer Tumor Growth and Metastasis by Targeting Overexpression of the LHRH Receptor

Lindsay E Calderon, Jonathan K Keeling, Joseph D Rollins, Carrie A Black, Kendall Collins, Nova L Arnold, Diane E. Vance, and Margaret W Ndinguri

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### Abstract

A new targeting chemotherapeutic agent, Pt-Mal-LHRH, was synthesized by linking activated cisplatin to 26 27 luteinizing hormone releasing hormone (LHRH). The compound's efficacy and selectivity towards 4T1 breast cancer cells were evaluated. Carboplatin was selected as the comparative platinum complex since 28 29 the Pt-Mal-LHRH malonate linker chelates platinum in a similar manner to carboplatin. Breast cancer 30 and normal cell viability were analyzed by an MTT assay comparing Pt-Mal-LHRH with 31 carboplatin. Cells were also treated with either Pt-Mal-LHRH or carboplatin to evaluate platinum uptake 32 by ICP-MS and cell migration using an *in-vitro* scratch-migration assay. Tumor volume and metastasis 33 were evaluated using an *in-vivo* 4T1 mouse tumor model. Mice were administered Pt-Mal-LHRH (carboplatin molar-equivalent dosage) through IP injection and compared to those treated with carboplatin 34 35 (5 mg/kg/wk), no treatment, and LHRH plus carboplatin (unbound) controls. An MTT assay showed a 36 reduction in cell viability (p<0.01) in 4T1 and MDA-MB-231 breast cancer cells treated with Pt-Mal-LHRH compared to carboplatin. Pt-Mal-LHRH was confirmed to be cytotoxic by flow cytometry using a 37 propidium iodide stain. Pt-Mal-LHRH displayed a 20-fold increase in 4T1 cellular uptake compared to 38 39 carboplatin. There was a decrease (p<0.0001) in 4T1 cell viability compared to 3T3 normal fibroblast cells. Treatment with Pt-Mal-LHRH also resulted in a significant decrease in cell-migration compared to 40 carboplatin. In-vivo testing found a significant reduction in tumor volume (p<0.05) and metastatic tumor 41 colonization in the lungs with Pt-Mal-LHRH compared to carboplatin. There was a slight decrease in 42 43 lung weight and no difference in liver weight between treatment groups. Together, our data indicates that 44 Pt-Mal-LHRH is a more potent and selective chemotherapeutic agent than untargeted carboplatin. 45

## Introduction

In the United States breast cancer afflicts 1 in 8 women during their lifetime.<sup>1</sup> The National
Cancer Institute's SEER database indicates the median overall survival rate for stage IV breast cancer is
23%. Despite recent advancements in the first-line treatments, many patients eventually relapse, their
tumors become chemoresistant, and patients experience numerous debilitating side effects from
chemotherapy treatment. Major issues of chemotherapy administration stem from the resulting severe side
effects, which causes doctors to often resort to dose reduction, treatment delay or suspension of therapy.

The platinum drugs carboplatin and cisplatin have been used to treat breast, prostate, ovarian, bladder, lung, head and neck cancer.<sup>2-3</sup> Cisplatin is the most potent member of the platinum family; however, due to its poor targeting and formation of resistance, it is not an effective treatment option for breast cancer.<sup>4-5</sup> Carboplatin, an analogue of cisplatin has gained greater use due to its lower non-specific toxicity and higher activity against cisplatin-resistant tumors.<sup>6</sup> Platinum anticancer drugs promote cell death by interfering with cellular transcription and DNA replication mechanisms. Serious side effects resulting from the platinum family drugs include nephrotoxicity, myelotoxicity, neurotoxicity, vomiting and nausea.<sup>7</sup> Another possible concern is the ability of certain cancers to develop resistance mechanisms to cisplatin including efflux pumps.<sup>8</sup> To combat acquired and intrinsic multidrug resistance, as well the severe side effects exhibited during high dose treatment, recent cancer research is focused on developing targeted cancer delivery to decrease side effects while improving efficacy. 

Systemic side effects result from anticancer drugs which are primarily designed to destroy rapidly
 dividing cells, including those found in healthy tissues. To decrease adverse side effects and limit
 nonspecific activity several methods of receptor directed chemotherapy have been developed using
 receptor ligands, lectins, antibodies, sugars, hormones and hormone analogs.<sup>9-14</sup> Luteinizing hormone
 releasing hormone (LHRH) receptor, also referred to as gonadotropin releasing hormone (GnRH)
 Page | 3

receptor, has been found to be overexpressed in breast, prostate, endometrial and ovarian cancers in

comparison to normal cells. This makes the LHRH peptide a good candidate for drug targeting.<sup>15-17</sup>

In many of these cancers, tumor proliferation can be inhibited by antagonist analogues of LHRH.<sup>18-19</sup> Previous studies have demonstrated that cytotoxic compound attachment at the D-lys moiety of LHRH does not change the intrinsic high binding affinity of the peptide to its receptors.<sup>14, 20-21</sup> For example. Schally and Nagy studied the potent antagonist on conjugates of [DLys6]-LHRH–DOX and [DLvs6]-LHRH–2-pyrrolino-DOX revealing increased efficacy of doxorubicin.<sup>22</sup> The LHRH analogue maintained its high targeted binding affinity while the drug retained its cytotoxic effects on the tumor cells.<sup>14</sup> Aggarwal and coworkers showed that [DLys6]-LHRH–curcumin enhanced apoptosis of tumor tissue and was useful in the treatment of pancreatic cancer.<sup>12</sup> In another study, Hansel and coworkers showed that conjugates of lytic peptides and LHRH are very effective in destroying human breast and prostate cancer xenografts that express LHRH receptors.<sup>23-26</sup> Mudviwa et al. also synthesized a [DLys6]-LHRH conjugate with phthalocyanine; however, the results of the biological testing have not been released.27 

Although numerous breast cancers overexpress LHRH receptors compared to normal breast cells,
few findings have been published regarding the role of LHRH and platinum drugs. This paper
demonstrates the chemical synthesis of the new compound Pt-Mal-LHRH by attaching LHRH (targeting
moiety) with activated cisplatin and reports data on the potency and selectivity of this chemotherapeutic
agent towards breast cancer. Carboplatin was selected as the comparative platinum complex since the PtMal-LHRH malonate linker chelates platinum in a similar manner to carboplatin (Figure 1).



using either an acidic or a neutral buffer system because the platinum complex dissociated. The products were, therefore, purified by gel filtration to remove any excess platinum. Fractions containing the Pt-Mal-LHRH complex in water were identified by ESI-MS and lyophilized to Pt-Mal-LHRH give pure platinum in a 15-17% yield. Mass spectra revealed the expected  $(M + Na)^+$  peak at 1756.7670 for the target compound (see Supporting Information). Any experiment with Pt-Mal-LHRH had to use freshly dissolved conjugate due to precipitates formed after storage, a phenomenon that has been reported before by Ndinguri, et al.<sup>13</sup> The Pt-Mal-LHRH compound was stable after the use of lyophilized powders at -20 °C. Before usage, fresh samples were made by reconstituting the dry samples to the desired concentration. Mass spectrometry was used to verify the integrity of the redissolved Pt-Mal-LHRH. The final

126 region/structure is maintained (Figure 1).

#### 128 Pt-Mal-LHRH increases breast cancer cytotoxicity through increased cellular uptake.

complex yields a chemical structure similar to carboplatin, in which the carboplatin bioactive

To examine the effects of Pt-Mal-LHRH on cell viability, 4T1 breast cancer cells were treated with carboplatin, LHRH, or Pt-Mal-LHRH from 0.1 µM to 100 µM. Pt-Mal-LHRH was found to significantly inhibit 4T1 viability compared to the untargeted carboplatin, concentration-dependently (Figure 2A). Both untargeted carboplatin and Pt-Mal-LHRH at 100 µM was found to significantly attenuate cell viability compared to the untreated control, however, this effect was more pronounced with Pt-Mal-LHRH. In addition, the enhanced potency of Pt-Mal-LHRH was also verified using a second cell line, the MDA-MB-231 breast cancer cell line. Pt-Mal-LHRH was found to significantly attenuate MDA-MB-231 breast cancer cell viability compared to carboplatin (see Supporting Information). Moreover, the induction of cytotoxicity by Pt-Mal-LHRH was confirmed by PI staining in which Pt-Mal-LHRH was found to significantly increase apoptosis compared to carboplatin (Figure 2B). Numerous experimental and clinical studies have shown overexpression of the LHRH receptor on human breast carcinomas, and specifically in the 4T1 and MDA-MB-231 cell lines.<sup>28-29</sup> To show the cytotoxic effect is induced by cellular entry of Pt-Mal-LHRH through Page | 6

#### **Bioconjugate Chemistry**

binding to the LHRH receptor a competition assay was conducted. Treatment with free LHRH antibody
showed a significant attenuation in the ability Pt-Mal-LHRH to induce a cytotoxic effect (Figure 2C).
Subsequently, due to targeting LHRH receptor overexpression, Pt-Mal-LHRH cellular uptake was found to
be 20 fold higher than carboplatin in the 4T1 cell line (Figure 2D). Pt-Mal-LHRH uptake was also found to
be significantly increased in the MDA-MB-231 breast cancer cells (see Supporting Information). These
results suggest that Pt-Mal-LHRH is more potent and efficacious than carboplatin in mediating breast
cancer cytotoxicity due to targeting aided by cellular overexpression of the LHRH receptor.



Figure 2: Pt-Mal-LHRH attenuates breast cancer cell viability through increased cellular uptake.
 4TI breast cancer cells were treated with Pt-Mal-LHRH, LHRH, or carboplatin (carbo) from a range of
 0.1 μM to 100 μM for 24 hours. Viability rates were analyzed by a MTT assay after the cells were
 Page | 7

incubated for 48 hours (A). Pt-Mal-LHRH was Flow cytometric analysis of cell apoptosis was conducted by treating cells with Pt-Mal-LHRH or carboplatin (100 µM) for 4 hours, then washing the collected cells followed by incubating the cells in 20 mg/ml PI for 15 minutes (B). Free LHRH antibody decreases the cytotoxic effect of Pt-Mal-LHRH. Cells were pretreated with free LHRH antibody (100  $\mu$ M) for 30 minutes and then concurrently with Pt-Mal-LHRH (100 µM) for 24 hours. Cellular viability was analyzed by MTT assay after 48 hours (C). To measure drug uptake  $1 \times 10^{6}$  4T1 cells were treated with Pt-Mal-LHRH or carboplatin (100 µM) for 24 hours. Cells were collected and metal (platinum) concentration mg/L was measured by ICP-MS (**D**). (n=3); \* p<0.05; \*\* p<0.01; \*\*\* p<0.001; \*\*\*\* p<0.0001; two-way ANOVA, one-way ANOVA and T-test. 

12 161 14 162

#### 162 Enhanced breast cancer cell selectivity is mediated by Pt-Mal-LHRH.

To determine the selectivity of Pt-Mal-LHRH towards breast cancer compared to normal fibroblast

164 cells, a cell viability assay was conducted. Pt-Mal-LHRH was found to significantly attenuate cell viability

in 4T1 cells compared to 3T3 normal fibroblast cells. This data indicates that the targeting ability of Pt-Mal-

166 LHRH mediates selectivity towards breast cancer than normal cells (Figure 3).



48 173 

## **Pt-Mal-LHRH** decreases breast cancer cell migration *in-vitro*.

The foremost cause of mortality in breast cancer patients is metastasis, in which 10-15% of
women develop distant tissue colonization within 3 years after a primary tumor is diagnosed. <sup>30</sup> For this

Page | 8

Figure 3: Pt-Mal-LHRH attenuates 4T1 breast cancer cell viability compared to normal cells (3T3).

4T1 and 3T3 cells were treated with Pt-Mal-LHRH from a range of 0.1 µM to 100 µM. Viability rates

were analyzed by a MTT assay after 72 hours of treatment. (n=3); \*\*\*\* p<0.0001; Two-way ANOVA.

 

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177 reason we conducted an *in-vitro* scratch assay to investigate the ability of Pt-Mal-LHRH to retard cell

178 migration. We found that Pt-Mal-LHRH significantly attenuated gap closure compared to untargeted

179 carboplatin and LHRH (100  $\mu$ M) treatments (Figure 4).



Figure 4: Breast cancer cell migration is decreased by Pt-Mal-LHRH. Cultured 4T1 cells were scratched
 under normal conditions (saline), carboplatin (carbo), LHRH, or Pt-Mal-LHRH (100 μM). Representative
 10x images indicate Pt-Mal-LHRH treatment attenuated breast cancer cell migration (A). Cell migration
 was quantitatively evaluated by measuring the distance between the scratch edges after 6 and 24 hours
 (B). (n=3); \*\*\*\* p<0.0001; One-way ANOVA. Images were adjusted for brightness and contrast.</li>

39 186 

## 187 Pt-Mal-LHRH attenuates breast cancer tumor growth and lung metastasis *in-vivo*.

188 To support our *in-vitro* results, we examined the antitumor activity and the effects on body and

189 liver weights of Pt-Mal-LHRH via *in-vivo* studies. Orthotopic 4T1 tumors grown in the right abdominal

190 mammary fat pad were treated with saline, carboplatin, carboplatin plus LHRH (unbound), or Pt-Mal-

191 LHRH by intraperitoneal injection of molar equivalent doses for two weeks. We purposefully chose a low

192 treatment dose to optimally discern differences in the therapeutic effect of Pt-Mal-LHRH to carboplatin.

5354 193 Previous publications highlight the use of higher treatment doses of carboplatin to mediate a significant

194 therapeutic effect on tumor growth.<sup>31-34</sup> As predicted no significant decrease in tumor growth was found

with carboplatin treatment compared to control mice; however, there was a significant decrease in tumor volume with Pt-Mal-LHRH treatment compared to control tumors (Figure 5A-B). Additionally, no difference in liver weights were found (Figure 5C) and cage-side observations did not detect any noticeable side effects from Pt-Mal-LHRH treatment. Lung examination revealed a significant decrease in lung tumor colonization in mice treated with Pt-Mal-LHRH compared to control and carboplatin treated mice (Figure 5D). Subsequently, Pt-Mal-LHRH mice showed a slight decrease in lung weight compared to control mice; however, significance was not reached (Figure 5E). Our *in-vivo* results indicate that Pt-Mal-LHRH results in regression of the breast cancer volume and metastasis. 

#### **Bioconjugate Chemistry**



Figure 5: Breast cancer tumor growth is attenuated by Platinum-LHRH treatment. Female BALB/c mice were implanted with 4T1 cells  $(1x10^6)$  in the right second mammary fat pad. Tumors were grown for 7 days, distributed into treatment groups, and treated with carboplatin (carbo), carboplatin and LHRH, or Pt-Mal-LHRH (5mg/kg/wk) by intraperitoneal injection for 2 wks. Tumor volume was measured over the 2 wk injection period (A) along with end tumor weight (B). Lung weight (C) Lung tumor nodule formation (D) and liver weight (E) was measured. (n=7); \* p<0.05; \*\* p<0.01; \*\*\*\* p<0.0001; Two and One-way ANOVA.

1 2 3	212	
4 5	213	Taken together, synthesis of Pt-Mal-LHRH produced a targeting chemotherapeutic agent which is
6 7	214	more potent and selective towards breast cancer cells than carboplatin shown by our <i>in-vitro</i> results.
8 9	215	Subsequently, our in-vivo results suggest Pt-Mal-LHRH attenuates breast cancer tumor growth and
10 11	216	metastasis compared to control and carboplatin.
12 13 14	217	
14 15 16 17	218	Discussion
18 19	219	
20 21	220	The platinum family anticancer drugs are commonly used for chemotherapy of advanced
22 23	221	carcinomas. Our research is to develop the platinum drugs into targeting agents, in order to increase the
24 25	222	potency and efficacy while reducing systemic side effects. We synthesized the targeting
26 27	223	chemotherapeutic agent Pt-Mal-LHRH to target the overexpression of the LHRH receptor on cancer cells
28 29	224	relative to normal tissue. Studies have shown that LHRH receptors are overexpressed in breast, prostate,
30 31 22	225	endometrial and ovarian cancers in comparison to normal cells making the LHRH receptor a good
33 34	226	candidate for drug targeting. <sup>14-17</sup> Specifically, numerous experimental and clinical studies have shown
35 36	227	overexpression of the LHRH receptor on approximately 50% of human breast carcinomas <sup>35</sup> and
37 38	228	specifically in the 4T1 cell line. <sup>28-29</sup> Additionally, various non-reproductive cancers have been shown to
39 40	229	overexpress the LHRH receptor including lung, bladder, pancreatic, among others. <sup>36-38</sup>
41 42	230	
43 44 45	231	To form Pt-Mal-LHRH we have combined the two moieties (LHRH and activated cisplatin) using
46 47	232	a malonate linker making the new conjugate target specific. In our synthesized platinum conjugate, we
48 49	233	introduce an ester bond between the malonate linker and platinum. Prior published reports have shown
50 51	234	that such a bond is hydrolyzed by cellular esterases, leaving the drug free to act at the cellular level. <sup>12, 39-40</sup>
52 53	235	In our study we chose carboplatin as our untargeted platinum complex due to the fact that the malonate
54 55 56	236	linker chelates platinum in a manner similar to that of carboplatin, as has been previously reported. <sup>13</sup> We
57 58 59		Page   12

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have conjugated activated cisplatin using the malonate linker on position 6 to ensure that the intrinsic
properties of the LHRH peptide are maintained which would allow the delivery of the cytotoxic aquated
platinum to the tumor cells.

One would expect that binding a platinum-based chemotherapeutic compound with a targeting agent should induce greater tumor toxicity with a concomitant reduction in systemic side effects. Chemotherapeutic agents which are given systemically without a significant targeting advantage should ideally have a low systemic toxicity or administered at a reduced dose to avoid complications. An example of this would be the use of BEL for the treatment of breast tumors, which in the animal model, has shown statistically significant reductions of tissue growth in both nicotine stimulated (p < 0.01) and non-nicotine stimulated breast cancer implants (p < 0.05).<sup>41</sup> We hypothesize an increase in the concentration of the drug Pt-Mal-LHRH in tumor cells as demonstrated in our *in-vitro* results would spare the normal cells from unnecessary exposure and advantageously allow for a higher dose of Pt-Mal-LHRH to be administered.

An examination of the current literature involving the use of LHRH with platinum drugs as a targeted treatment mechanism yields few results constituting effective or applicable targeted delivery. The few published studies utilizing platinum derivatives employ combinatory methods on cisplatin resistant tumors with LHRH and differ significantly in architecture and mode of delivery from the Pt-Mal-LHRH complex reported in this research.<sup>42-43</sup> For instance, LHRH-targeted nanogels show complications with free drug loading and slow release of the encapsulated cisplatin resulting in diminished cytotoxicity compared to free cisplatin.<sup>43-44</sup> However, our data demonstrated the successful synthesis of Pt-Mal-LHRH and its effectiveness on breast cancer. Additional research examining the effectiveness of Pt-Mal-LHRH on other cancers that express the LHRH receptor including but not limited to lung, bladder, and pancreatic will need to be performed. 

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263	Other modes of targeting platinum drugs that have been explored yielding limited results.
264	Targeted platinum sugar complexes have been shown to have differences in activities between D and L -
265	glucose conjugates; however, no increase in activity over the analogous carboplatin was found in either of
266	the complexes. <sup>45-47</sup> Other complexes formed from a range of different sugars have been prepared, but the
267	results of the biological testing have not been released. <sup>47-48</sup> Other studies targeting strategies for drug
268	delivery vehicles against the estrogen receptor (ER) in breast cancer have included steroids as targeting
269	units; however, these compounds did not fare better than cisplatin. <sup>46, 49-52</sup> In one study, the platinum
270	conjugates cis-dichloro[N-(4-(17-ethynylestradiolyl)-benzyl)-ethylenediamine]platinum(II) and cis-
271	diamino[2-(4-(17-ethynylestradiolyl)-benzoylamino)-malonato]platinum-(II) enhanced proliferation
272	which made things worse and rendered them unsuitable for further investigation. <sup>53</sup> It has been shown that
273	various cancer cell lines overexpress a glycoprotein that acts as a folate receptor. Cisplatin and
274	carboplatin derivatives bearing folate units were prepared; however, the conjugates had low solubility in
275	water which prohibited their use in biological experiments. <sup>46, 54</sup> Aronov et al. used a PEG spacer to
276	enhance water solubility but the conjugate was found to be less cytotoxic than carboplatin. <sup>55</sup> In regards to
277	peptide-based targeting, instances in which targeting has been successfully achieved with platinum(II)
278	conjugates are few. One successful example is the use of cyclic peptide CNGRC sequence that targets the
279	CD13 receptor overexpressed on the surface of certain cancer cells. <sup>13</sup>

The cytotoxic mechanism of action of Pt-Mal-LHRH mediates enhanced selectivity and efficacy 281 282 through use of the LHRH moiety to bind to the LHRH receptors which leads to internalization of the compound. Our in-vitro data utilizing 4T1 cells supports the theory that Pt-Mal-LHRH increases 283 cytotoxicity through enhanced cellular uptake by over 20- fold increasing the bioavailability of 284 carboplatin and further augmenting its *in-vivo* tumoricidal properties. The increased tumor concentration 285 may also allow a decrease in the systemic dosage of the chemotherapeutic agent with a concomitant 286 287 decrease in system toxicity and side effects. This is an important consideration because of the 288 carboplatin's narrow therapeutic index. Moreover, there are three cellular mechanisms which may impart Page | 14

#### **Bioconjugate Chemistry**

a resistance to platinum based drugs. These include decreased uptake, increased repair of DNA damage and increased drug deactivation.<sup>56</sup> It would be expected that the enhanced uptake of platinum with the Pt-Mal-LHRH compound may result in mitigating these resistance mechanisms, either directly or if saturation can be obtained. Subsequently, once, Pt-Mal-LHRH is internalized it will be cleaved releasing the aquated platinum to bind to the DNA forming DNA adducts resulting in cytotoxicity. We postulate the DNA adduct formation will be similar to carboplatin which preferentially attacks the sequences of GG >> AG > GA > GXG (X=an undefined residue), in that order.<sup>57-58</sup> One possible effect of Pt-Mal-LHRH is the targeting of the ovaries, adrenal and pituitary gland. Chemical oophorectomy, adrenalectomy and hypophysectomy have been used in the treatment of advanced breast cancer.<sup>59</sup> If targeting of these organs take place, it is possible that it will be selective and possibly affect only some of the endocrine functions. Interestingly, one study demonstrates that the damaging effect to the pituitary gland by a targeted cytotoxic analog of LHRH was found to be reversible after 2 wks of treatment.<sup>60</sup> Further research into this area along with determining the exact mechanisms of tumorcidal activity, and potential risks and benefits is needed. In addition, we observed that LHRH treatment modestly stimulated tumor growth and migration. This finding is expected since it is known that LHRH antagonists are effective breast cancer treatments.<sup>61</sup> It has been shown that LHRH treatment promotes proliferation through reducing the number of resting cells in the G0-phase.<sup>62</sup> Subsequently, our results demonstrate that tumor growth and metastasis stimulated by LHRH is slightly greater than the control. However, we were unable to demonstrate tumor cytotoxicity augmentation with the administration of un-linked carboplatin with LHRH treatment; therefore, the optimal timing, dosage and frequency of administration was not investigated. We found Pt-Mal-LHRH significantly enhanced cellular cytotoxicity in 4T1 cells compared to the 

3T3 normal cell line. Taken together, both our *in-vitro and in-vivo* data suggest Pt-Mal-LHRH elicits
tumor-targeted drug delivery with increased potency, efficacy, and a possible reduction in

chemotherapeutic side effects which would allow a full or higher dose of chemotherapy to be used in a patient, as compared to other platinum drugs. Subsequently, our *in-vitro* scratch assay data demonstrated a reduction in migration. Importantly, *in-vivo* metastasis was investigated since the major cause of mortality in breast cancer patients is metastasis to distant sites including the lungs. Our in-vivo data supports our cellular data, as we found a significant decrease in tumor volume and lung tumor colonization through Pt-Mal-LHRH treatment. The advantage of the Pt-Mal-LHRH conjugate is selectivity toward tumors that overexpress LHRH receptors locally at the site of their growth, while avoiding systemic distribution. Our approach to tumor-targeted drug/delivery will not be limited specifically to breast cancer, but could, in general, have applications in other metastatic cancers that overexpress the LHRH receptor such as bladder, melanoma, and pancreatic cancers, among others. Conclusion In summary, we have demonstrated the synthesis of a new chemotherapeutic agent, Pt-Mal-LHRH, designed to selectively target cancer cells overexpressing the LHRH receptor. Our *in-vitro* results using 4T1 breast cancer cells indicate increased cytotoxicity and uptake compared to carboplatin, along with selectivity towards breast cancer compared to normal cells. In addition, our *in-vivo* results indicate Pt-Mal-LHRH treatment significantly decreases tumor volume and metastasis to the lung. Our results indicate that Pt-Mal-LHRH has increased potency, efficacy, and selectivity towards breast cancer cells overexpressing the LHRH receptor compared to carboplatin. Pt-Mal-LHRH has the potential to be a clinically used chemotherapeutic agent. Page | 16

1 2 3 4	339	Experimental Procedures		
5 6	340	Reagents		
7 8	341	All starting reagents listed below were obtained from commercial sources and used without further		
9 10 11	342	purification: N,N-diisopropylethylamine (DIEA), acetic acid, dichloromethane (CH <sub>2</sub> Cl <sub>2</sub> ), trifluoroacetic		
12 13	343	acid (TFA), N-methylmorpholine (NMM), triisopropylsilane (Tips), dry solvents, AgNO <sub>3</sub> , cis-		
14 15	344	diammineplatinum(II) dichloride (Cisplatin), were obtained from Fisher Scientific or Sigma Aldrich; all		
16 17 18	345	amino acids and activators were from Novabiochem or AGTC Bioproducts.		
19 20	346			
21 22	347	NMR Spectroscopy		
23 24 25	348	The NMR spectroscopy was used to analyze and confirm the products obtained in each steps of		
25 26 27	349	linker formation. All <sup>1</sup> H NMR spectra were recorded on 400 MHz JEOL Eclipse+ NMR Spectrometer		
28 29	350	and processed with Delta <sup>TM</sup> NMR software.		
30 31	351			
32 33	352	Mass spectrometry		
34 35	353	Analysis was done on a Thermo Scientific LTQ XL Mass Spectrometer using a DART (Direct		
36 37 29	354	Analysis in Real Time) or (Electrospray Ionization) ESI on an Agilent Technologies instrument processed		
39 40	355	with Analyst QS1.1 (Applied Biosystems) or Mass Hunter (Agilent)		
41 42	356			
43 44	357	HPLC		
45 46	358	Analytical high performance liquid chromatography (HPLC) was performed to confirm purity of		
47 48	359	LHRH-Mal using an Agilent 1100 series multisolvent delivery system with a DAD detector controlled by		
49 50 51	360	Agilent Chem Station Plus with detection at 220 nm.		
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#### **Inductive Coupled Plasma-Optical Emission Spectroscopy**

Platinum uptake was analyzed using a Varian, Vista MPX CCD simultaneous spectrometer. PlasmaCal Pt (1000  $\mu$ g/mL) from SCP Science was used for calibration by diluting it to standard solutions of known concentration.

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#### 368 Synthesis of Malonate (Mal) Linker

The malonate linker, Di-tert-butyl 2-(3-(4-Carboxybutanamido)propyl)-malonate (Mal) was synthesized
as previously described by Ndinguri et al.<sup>13</sup> All analyses matched the reported <sup>1</sup>H NMR and mass spectra
values.<sup>13</sup>

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#### 373 Synthesis of Malonate- LHRH (Mal-LHRH)

374 The Mal-LHRH decapeptide was synthesized using Fmoc solid phase chemistry techniques and then chelated to activated cisplatin. Briefly, Fmoc-Rink Amide-AM resin (0.4 mmol) was placed onto a 375 376 reaction vessel. The resin was then washed with DMF and DCM in continuous-flow mode using a PS3 peptide synthesizer. All couplings employed four equivalents of amino acid and (2-(6-Chloro-1H-377 378 benzotriazole-1-yl)-1,1,3,3-tetramethylaminium hexafluorophosphate (HCTU) and sometimes (7-Azabenzotriazol-1-vloxy)tripyrrolidinophosphoniumhexafluorophosphate (PyAOP) dissolved in 0.4 M N-379 methylmorpholine (NMM) in DMF at room temperature. Coupling involved minimal preactivation times. 380 The side chain-protected amino acid derivatives Fmoc-Gly-OH, Fmoc-Pro-OH, Fmoc-Arg(Pbf)-OH, 381 Fmoc-Leu-OH, Fmoc-DLys(Alloc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Trp(Boc)-OH, 382 Fmoc-His(Trt)-OH and Boc-Glu(OtBu)-OH were used. The Alloc protecting group was removed using 383 384 tetrakis(triphenylphosphine)palladium(0) along with a 37:2:1 mixture of methylene chloride, acetic acid, and NMM for 2 hours, followed by washing and double coupling of malonate linker (Mal). 385 386 Intermediate products were washed between reactions with DMF. The Fmoc group was deprotected with 20% piperidine in DMF for 3 min. Using a cocktail of trifluoroacetic acid: water: triisopropylsilane 387 388 (TFA:H<sub>2</sub>O:TIPS) (90:5:5:) (15 mL), the peptide was cleaved from the resin, precipitated and dried under Page | 18

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#### **Bioconjugate Chemistry**

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vacuum to give the crude peptide conjugate Mal-LHRH. Mal-LHRH was purified by HPLC or gel filtration using Sephadex G-10. The purified samples were lyophilized before use in the next step. The crude peptide was purified using HPLC on reverse phase C18 column with a linear gradient from 5% to 80% B eluent in 15 min;  $t_R$  5.5 min (see Supporting Information). The purity of the compound was also confirmed using mass spectrometry. Yield, 39%, ESI-MS (M+H)<sup>+</sup>, calculated for C<sub>70</sub>H<sub>101</sub>N<sub>19</sub>O<sub>20</sub>, 1528.69 found 1528.7304 (see Supporting Information).

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## 396 Synthesis of Pt-Mal-LHRH

397Reaction of Mal-LHRH and activated cisplatin yielded Pt-Mal-LHRH, see Figure 6. Mal-LHRH398was dissolved in water, and the pH of the solution was adjusted to 7 by titration with 1 M NaOH. A399solution of cis-[Pt-(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)<sub>2</sub>]<sup>2+</sup>(NO<sub>3</sub>)<sub>2</sub> was made by stirring cis-PtCl<sub>2</sub>(NH<sub>3</sub>)<sub>2</sub> and AgNO<sub>3</sub> vigorously400overnight in water in the dark. A precipitate of AgCl formed was filtered to give cis-[Pt-401 $(NH_3)_2(H_2O)_2]^{2+}(NO_3)_2$  (35, 36). A 1.2 fold excess of cis-[Pt-(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)<sub>2</sub>]<sup>2+</sup>(NO<sub>3</sub>)<sub>2</sub> (20 mmol) was402reacted with Mal-LHRH (30 mg, 20 mmol) to form the Pt-Mal-LHRH. Yield, 15-17%, ESI-MS (TOF)403gave a signal at 1756.760 (M + Na)<sup>+</sup> calculated for C<sub>70</sub>H<sub>101</sub>N<sub>21</sub>O<sub>19</sub>Pt 1756.72



#### **Bioconjugate Chemistry**

412 Cell Culture
413 The 4T1 mouse mammary tumor cell line and 3T3 mouse embryonic fibroblast cell line was
414 obtained from ATCC and maintained as previously described.<sup>41</sup> In addition, the MDA-MB-231 cell line
415 was provided by Dr. Michael Kilgore at the University of Kentucky (purchased from ATCC which
416 authenticates the cell line through short tandem repeat (STR) profiling). The cells were maintained in
417 Dulbecco's Modified Eagle Medium (DMEM) containing 10% Fetal Bovine Serum and 100 U/ml
418 penicillin and 100 µg/ml streptomycin.

#### 419 Cell Viability Assay

To assess the ability of Pt-Mal-LHRH to affect cell viability an MTT Assay was utilized. 4T1 or
MDA-MB-231 cells were seeded 2000 cells/100µl in a 96-well plate. The wells were treated with PBS,
carboplatin, LHRH, or Pt-Mal-LHRH from a range of 0.1 µM to 100 µM for 24 hours. Cells were then
washed with PBS and cultured in DMEM supplemented with 10% FBS for 48 hours. After this the cells
were incubated at 37°C for 4 hours in 10 µl MTT solution obtained from Vybrant MTT Cell Proliferation
Assay Kit (Life Technologies). Cells were solubilized and mixed with SDS (sodium dodecyl sulfate) and
the absorbance read at 595 nm on a Phenix Genios Tecon 96 well plate reader.

#### 427 Cytotoxicity Assay

428 Apoptotic cells were detected by flow cytometry using propidium Iodide (PI) staining of DNA 429 fragments to determine drug cytotoxicity. 4T1 cells were seeded  $1X10^6$  cells/well and treated with 100 430  $\mu$ M concentration of Pt-Mal-LHRH or carboplatin for 4 hours. Cells were collected, washed 3 times with 431 PBS and incubated with 20  $\mu$ g/ml PI for 15 minutes. Next, flow cytometry was carried out using a 432 FACSCalibur<sup>TM</sup> flow cytometer (Becton-Dickinson) at the University of Kentucky flow cytometry and 433 cell sorting core facility. A total of 50,000 events were acquired and analyzed.

434 Competition Assay

435 To determine binding of Pt-Mal-LHRH to the LHRH receptor for entry into the cell a competition
436 assay was conducted. 4T1 cells were seeded 2000 cells/100μl in a 96-well plate. Cells were pretreated
437 with free LHRH antibody (EMD Millipore Corp. AB1567) 100 μM for 30 minutes and then concurrently
Page | 21

**ACS Paragon Plus Environment** 

2 3	438	with Pt-Mal-LHRH 100 $\mu$ M for 24 hours. Cells were then washed with PBS and cultured in DMEM
4 5	439	supplemented with 10% FBS for 48 hours. A cell viability assay was then performed as described above.
6 7	440	Drug Uptake Assay
8 9	441	To determine cellular uptake of Pt-Mal-LHRH, 4T1 cells were seeded 1X10 <sup>6</sup> in 6 well plates and
10 11	442	treated with either carboplatin (100 $\mu$ M) or Pt-Mal-LHRH (100 $\mu$ M) for 24 hours. Cells were washed 3
12 13 14	443	times with PBS, collected, and metal (platinum) concentration mg/L as measured by ICP-Mass
14 15 16	444	spectrometry at Louisiana State University as previously described. <sup>13</sup>
17 18	445	Cell Migration
19 20	446	Cell migration was examined using a scratch assay. 4T1 cells were cultured in a 6-well plate
21 22	447	until a confluent monolayer was formed. The wells were scratched with a 20 µl pipet tip, rinsed with PBS,
23 24 25	448	and cultured in DMEM supplemented with 10% FBS. The wells were treated with PBS, carboplatin,
25 26 27 28 29	449	LHRH, or Pt-Mal-LHRH. A concentration of 100 $\mu$ M was used for all compounds. Four representative
	450	10x images were taken at 0 and 24 hours and the gap width was quantified using an average of three
30 31	451	leading edge measurements for each image.
32 33	452	In-vivo Syngeneic Isograft Model of Breast Cancer
34 35	453	Female BALB/c mice, 10 weeks old with an average weight of 19.80g, were purchased from
36 37 28	454	Jackson Laboratory (Bar Harbor, ME) and housed as previously described. <sup>41</sup> Animal protocols were
30 39 40	455	approved by the committee on animal research care and use at Eastern Kentucky University.
41 42	456	
43 44	457	4T1 cells ( $1X10^{6}$ ) were suspended in 100 µl of DMEM not supplemented with FBS and injected
45 46	458	into the right second mammary fat pad of female BALB/c mice as previously described in literature. <sup>63</sup>
47 48	459	After 7 days of orthotopic tumor initiation (~100mm <sup>3</sup> ) the mice were treated with saline, carboplatin,
49 50 51	460	carboplatin plus LHRH, or Pt-Mal-LHRH by intraperitoneal injection for two weeks. A molar equivalent
52 53	461	dosage of 5mg/kg/wk was used for all compounds according to their molecular weights. Tumor growth
54 55	462	was monitored daily and tumor volumes (mm <sup>3</sup> ) were calculated using the formula: $(width)^2 \times length/2$ ,
56 57 58 59 60		Page   22

## **Bioconjugate Chemistry**

1 2	463	where width is the smaller of the two measurements. After 2 weeks of treatments the mice were sacrificed		
3 4	464	and tumor volume and organ weights were measured.		
5 6 7	465	Statistics		
7 8 9	466	Data are illustrated as mean $\pm$ SEM and statistical analyses were carried out using GraphPad.		
10				
11 12 13 14	467	Prism 6 (San Diego, CA) using t-tests. One- or two-way ANOVAs were used where appropriate.		
	468			
15 16	469	Acknowledgements: We would like to thank Dr. Thomas Blanchard for his help with the ICP analysis.		
17 18 19 20	470	Funding Sources: Kentucky Biomedical Research Infrastructure Network (KBRIN) grants		
	471	(P20GMI04489 and P20GM103436); EKU University Research Grant and National Institute of General		
21 22	472	Medical Sciences at the National Institutes of Health (grant # 1R25GM102776-01)		
23 24	473			
25 26 27	474	Supporting information: Supporting Information Available: MDA-MB-231 cytotoxicity graph,		
27 28 29 30 31	475	MDA-MB-231 uptake graph, HPLC analysis spectra and mass spectra analysis.		
	476	This material is available free of charge via the Internet at <u>http://pubs.acs.org</u>		
32	477			
33 34 35 36 37 38 39 40 41	478	Abbreviations: N,N-diisopropylethylamine (DIEA), dichloromethane (CH <sub>2</sub> Cl <sub>2</sub> ), trifluoroacetic acid		
	479	(TFA), N-methylmorpholine (NMM), triisopropylsilane (Tips), Dimethylformamide (DMF), cis-		
	480	diammineplatinum(II) dichloride (Cisplatin), 2-(6-Chloro-1H-benzotriazole-1-yl)-1,1,3,3-		
	481	tetramethylaminium hexafluorophosphate (HCTU), 7-Azabenzotriazol-1-		
42 43	482	yloxy)tripyrrolidinophosphoniumhexafluorophosphate (PyAOP), Dulbecco's Modified Eagle Medium		
44 45 46	483	(DMEM), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), American Type Culture		
40 47 48	484	Collection (ATCC), Fetal Bovine Serum (FBS)		
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