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Acyclic Nucleosides: Synthesis of 1-[(1-Hydroxy-2-Propoxy) Methyl]Thymine, 6-Azathymine, URACIL, AND 6-Azaauracil as Potential Antiviral Agents

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ACYCLIC NUCLEOSIDES: SYNTHESIS OF 1-[(1-HYDROXY-2-PROPOXY)METHYL]THYMINE, 6-AZATHYMINE, URACIL, AND 6-AZAURACIL AS POTENTIAL ANTIVIRAL AGENTS

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Abstract

A series of acyclic nucleosides have been synthesized. Thymine, 6-azathymine, uracil, and 6-azauracil were silylated with hexamethyldisilazane in the presence of ammonium sulfate and then coupled with 1-benzyloxy-2-chloromethoxypropane to give the corresponding 1-(1-benzyloxy-2-propoxy)methyl derivatives. A minor quantity of benzyloxymethylated product was also obtained in each case. Hydrogenolysis of the protected acyclic nucleosides with palladium(II) hydroxide afforded the title compounds. None of the compounds exhibited significant antiviral activity against human immunodeficiency virus (HIV).

INTRODUCTION

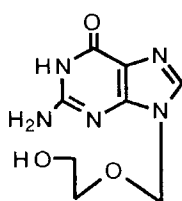
Recent chemotherapeutic approaches toward acquired immunodeficiency syndrome (AIDS) have resulted in some new developments. Besides AZT (3'-azido-3'-deoxythymidine),¹ ddl (2', 3'-dideoxyinosine)² and ddC (2', 3'-dideoxycytidine)³ have also been approved for the treatment of AIDS.⁴ From the viewpoints of the structure of these drugs, they can be classified as

dideoxynucleosides. Other dideoxynucleosides, such as, d4T (2',3'-didehydro-3'-deoxythymidine),⁵ ddU (2',3'-dideoxyuridine),⁶ and ddT (2', 3'-dideoxythymidine)⁷ have also been discovered to possess potential activity against HIV.

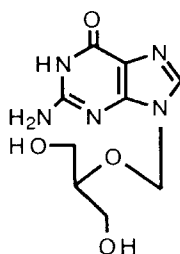
The mechanism of action of these drugs is generally believed to be initial phosphorylation to the corresponding 5'-phosphate, then conversion to the 5'-diphosphate, and finally to the 5'-triphosphate by cellular enzymes, kinases. When this corresponding triphosphate is incorporated into the replicating viral DNA chain by HIV reverse transcriptase, the further chain elongation is inhibited by the lack of a C-3' hydroxy group.⁸

Because of the undesired side effects, the bone marrow inhibition of AZT,⁹ the peripheral neuropathy of ddC and ddI,¹⁰ researchers were prompted to seek more active and less toxic compounds which specifically inhibit AIDS viruses. Several efforts and strategies have been made for these purposes.¹¹ One of the efforts focused on the modification of the sugar moiety leading to the synthesis of 2', 3'-dideoxynucleosides and their analogs (including unsaturated analogs).¹²

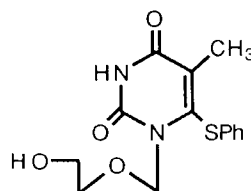
Both 9-[(2-hydroxyethoxy)methyl]guanine (ACV, **1**)¹³ and 9-[(1,3-dihydroxy-2-propoxy)methyl]guanine (DHPG, **2**),¹⁴ which possess an acyclic sugar moiety, have been used clinically to treat herpes simplex virus infections. Other efforts focused on the synthesis of acyclic nucleosides not only for antiherpes but also for anti-HIV purposes. 1-[(2-Hydroxyethoxy)methyl]-6-(phenylthio)-thymine (HEPT, **3**)¹⁵ has been reported to exhibit anti-HIV-1 activity and to be less toxic than AZT for human bone marrow cells *in vitro*. Trinh *et al.*¹⁶ reported thymine acyclic nucleosides (**4-7**) which were regarded as AZT analogs lacking C(1')-C(2') and C(2')-C(3') bonds. Among them, 3'-amino derivatives (**6, 7**) were presumed to interact with the active site of reverse transcriptase, which is a prime target for the development of chemotherapeutic agents for AIDS. These observations led us to initiate a series of studies on the acyclic nucleosides.¹⁷ Herein we would like to report our recent work on the synthesis of 1-[(1-hydroxy-2-propoxy)methyl]thymine, 6-azathymine, uracil, and 6-azauracil (**15a-15d**), which can be regarded as ddU and ddT analogs lacking the C(1')-C(2') and C(2')-C(3') bonds.



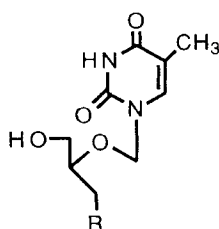
1



2



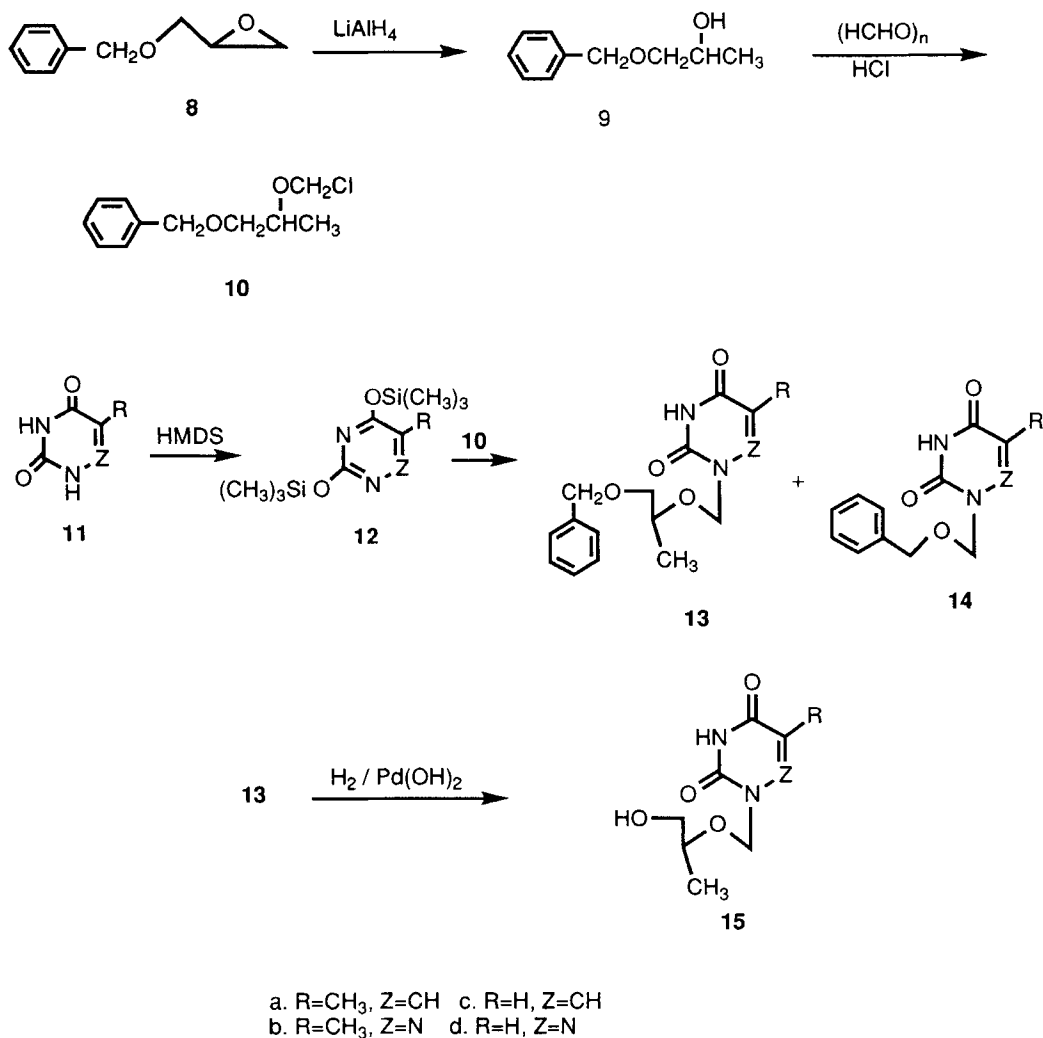
3



4. R=N₃
 5. R=NH₂
 6. R=NHCN
 7. R=NHCHO

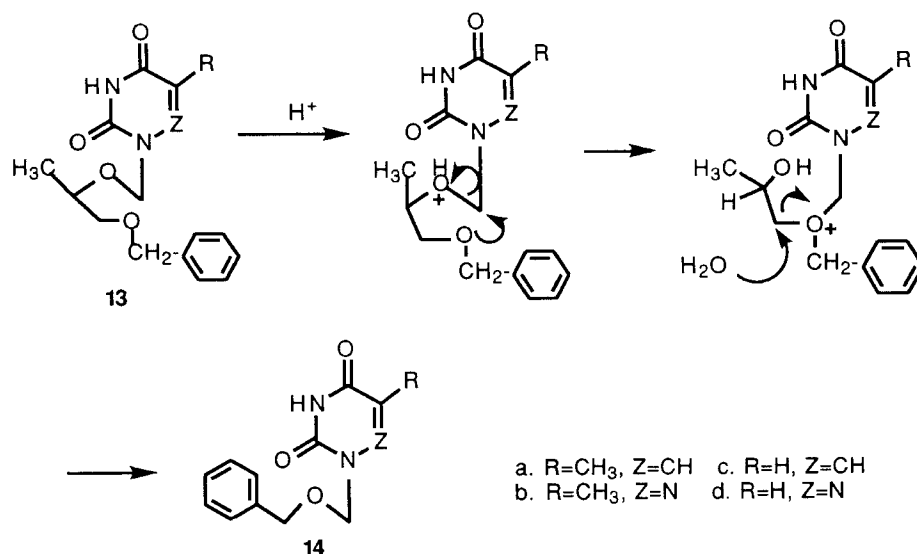
RESULTS AND DISCUSSION

Benzyl glycidyl ether (**8**)¹⁸ was prepared by reacting epichlorohydrin with sodium benzyloxide in dry toluene. Treatment of **8** with lithium aluminum hydride afforded 1-benzyloxy-2-propanol (**9**), which was chloromethylated with paraformaldehyde and dry HCl in anhydrous 1,2-dichloroethane at 0°C to give (1-benzyloxy-2-chloromethoxy)propane (**10**) in a good overall yield. Each of the persilylated intermediates **12a-d**, prepared by silylating the bases **11a-d** with hexamethyldisilazane (HMDS) in the presence of ammonium sulfate, was alkylated with one molar equivalent of **10** and one molar equivalent amount of zinc iodide in dry 1,2-dichloroethane to yield 1-[(1-benzyloxy-2-propoxy)methyl]thymine, 6-azathymine, uracil, and 6-azauracil (**13a-d**) respectively as shown in Scheme 1. In addition to the expected product, a minor product was also isolated. For example, in the case of **12a**, the ¹H nmr spectrum of this minor product showed a one-proton doublet at 7.11 ppm (*J*=1.0 Hz) and a three-proton doublet at 1.90 ppm (*J*= 1.0 Hz) corresponding to the long range coupling of C₆-H and C₅-CH₃. A pair of two-proton singlets at 5.21 ppm and 4.61 ppm indicated the presence of two methylenes. The remaining five-proton singlet at 7.33 ppm was attributed to the phenyl group. The ¹³C nmr spectrum



Scheme 1

again supported the presence of two methylene carbons appeared at 76.52 ppm and 72.07 ppm. The mass spectrum of this minor product showed the molecular ion at m/z 246, corresponding to the molecular formula, C₁₃H₁₄N₂O₃. These results suggested that the minor product was 1-(benzyloxymethyl)thymine (**14a**). Compounds **13a** and **14a** were separated by crystallization from chloroform-ether, **14a** was obtained as colorless crystals while **13a** remained in the solution. Compounds **14b-d** were also obtained as a minor



Scheme 2

product in each case during the reaction of **13b-d**. Hydrogenolysis of **13a-d** with palladium(II) hydroxide afforded the desired title compounds **15a-d** in a fairly good yield.

When the reaction of compounds **12a-d** with **10** was carried out in the presence of aluminium chloride as a catalyst, the rearranged products **14a-d** were predominant along with a trace amount of **13a-d**. A possible mechanism for the formation of **14a-d** from **13a-d** respectively is illustrated in Scheme 2. The rearrangement is believed to be catalyzed by Lewis acid, zinc iodide or aluminium chloride and/or hydrogen chloride.

EXPERIMENTAL SECTION

Melting points (Yanaco micro-melting-point apparatus) are uncorrected. ¹H-NMR and ¹³C-NMR spectra were obtained on a Varian Gemini-200 or VXR-300 spectrometer; chemical shifts are measured in parts per million with respect to TMS. Ultraviolet absorption spectra were measured on a Shimadzu UV-200 spectrophotometer. Elemental analyses were recorded on a Heraeus CHN-O Rapid

analyzer. High-resolution mass spectra were recorded on a VG 70-250 spectrometer and low resolution mass spectra were recorded on a VG Quattro spectrometer. Silica gel (70-230 mesh suitable for column chromatographic use) and thin-layer chromatography on precoated silica gel 60 F-254 plates were purchased from E. Merck. UV light (254 nm) was used to detect the UV-absorption spots on TLC plates after development.

1-Benzylloxy-2-propanol(9)

Benzylglycidyl ether (**8**)¹⁸ (6.56 g; 40 mmol) was dissolved in anhydrous THF(125 mL) and cooled in an ice bath. To this cooled solution, excess LiAlH₄ (0.75 g) was added in portions and stirred under reflux for 6 h. The reaction mixture was then quenched with sat. NH₄Cl and extracted with EtOAc (50 mL x 4), The organic layers were combined, washed with brine, dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under diminished pressure to give an oily residue which was distilled (kugelrohr apparatus) to furnish pure **9** (5.54g, 70% yield), [bp 96-110°C(1-2 mmHg)]; ¹H-NMR (CDCl₃, 200 MHz) δ : 1.12 (d, *J* = 6 Hz, 3H, H-3), 3.0 (br.s, 1H, OH), 3.2-3.5 (m, 2H, H-1), 3.96 (m, 1H, H-2), 4.52 (s, 2H, benzylic-H), 7.32 (m, 5H, aromatic H); ¹³C-NMR(CDCl₃, 50 MHz) δ : 18.75 (C-3), 66.43 (C-2), 73.26 (C-1), 75.85 (benzylic-C), 126.88, 127.74, 128.42, 137.99 (aromatic-C); MS *m/z* 166(M⁺); HRMS for C₁₀H₁₄O₂ Calcd:166.0994. Found: 166.0993.

1-Benzylloxy-2-chloromethoxypropane (10)

1-Benzylloxy-2-propanol (**9**) (6.65 g, 40 mmol) , paraformaldehyde (2.4 g) and anhydrous CaCl₂ (7 g) were added to dry 1,2-dichloroethane (200mL, distilled over P₂O₅). The mixture was cooled in an ice bath and bubbled with dry HCl gas which was generated from NaCl and c-H₂SO₄(95-98 %), and stirred at 0 °C for 6 h; the solution became clear at the end of reaction. The solution was warmed to room temperature and the excess HCl was carefully removed under water aspirator; then the solution was collected by filtration to remove CaCl₂. The filtrate was concentrated at reduced pressure to furnish **10** as a syrup. Due to the instability of this syrup, it was directly used for the next reaction without further purification.

1-[(1-Benzyloxy-2-propoxy)methyl]thymine (13a) and 1-(Benzyloxymethyl)thymine (14a)**Method A:**

Thymine(**11a**) (0.63g; 5 mmol) and ammonium sulfate(530 mg) were added to hexamethyldisilazane (HMDS;100 mL). The mixture was heated at reflux with exclusion of moisture until the solution became clear (4 h). The excess HMDS was removed under reduced pressure to give silylated intermediate (**12a**) which was dissolved in dry 1,2-dichloroethane (20 mL) and to which was added chloromethylether (**10**) (5 mmol in 20 mL dichloroethane). The reaction mixture was cooled to 0°C and anhydrous zinc iodide (1.60g, 5 mmol) was added. The reaction mixture was stirred at room temperature overnight, quenched with ice water and stirred for 30 min. The organic layer was separated and the aqueous layer was extracted with CHCl₃ (50 mL x 4). The combined organic layers were washed with sat. NaHSO₃ (10 mL x 3; the solution became colorless), brine, dried under anhydrous MgSO₄ and filtered. The filtrate was concentrated under reduced pressure and the residue was purified by column chromatography on silica gel using CHCl₃ : MeOH (100 : 5) as an eluent to give a mixture of **13a** and **14a**. The mixture was triturated with chloroform and kept in refrigerator overnight. This gave **14a** as a precipitate. After recrystallization from CHCl₃-Et₂O, **14a**(0.18 g, 15%) was obtained as colorless crystals; mp 130-132°C; UV : HCl(0.1 M) λ_{max} 265 nm (ϵ 8500), NaOH(0.1 M) λ_{max} 265nm (ϵ 6500); ¹H-NMR(CDCl₃) δ : 1.90(d, J = 1 Hz, 3H, CH₃), 4.61 (s, 2H, benzylic H), 5.21(s, 2H, H-1'), 7.11(d, J = 1 Hz, 1H, H-6), 7.33 (s, 5H, aromatic H), 9.57(br. s. 1H, NH); ¹³C-NMR(CDCl₃) δ : 12.80 (CH₃), 72.07 (benzylic C), 76.52 (C-1'), 112.17 (C-5), 128.42, 128.67, 129.03, 137.22 (aromatic C), 139.45 (C-6), 151.85 (C-2), 164.73 (C-4); MS m/z 246(M⁺); Anal. Calcd for C₁₃H₁₄N₂O₃ : C, 63.40; H, 5.73; N, 11.37. Found: C, 63.12; H, 6.09; N, 10.93.

The mother liquor was evaporated and the residue was recrystallized from ethanol to obtain pure **13a** (1.03 g, 68%) as colorless crystals; mp 77-78°C; UV : HCl(0.1 M) λ_{max} 265 nm(ϵ 8000), NaOH(0.1 M) λ_{max} 265nm (ϵ 6500); ¹H-NMR(CDCl₃) δ : 1.49 (d, J = 6.4 Hz, 3H, H-3'), 1.86(d, J = 1.0 Hz, 3H, CH₃), 3.44 (d, J = 5.3 Hz, 2H, H-5'), 3.92 (m, 1H, H-4'), 4.51 (s, 2H, benzylic H), 5.17, 5.27(dd, J =10.5 Hz, 2H, H-1'), 7.17 (d, J = 1.0 Hz, 1H, H-6), 7.31 (m, 5H,

aromatic H), 9.40 (br. s. 1H, NH); ^{13}C -NMR(CDCl_3) δ : 12.72 (C-3'), 17.60 (CH_3), 73.78 (C-5'), 74.55 (C-4'), 74.55 (benzylic C), 75.81 (C-1'), 111.76 (C-5), 128.03, 128.18, 128.89, 138.48 (aromatic C), 139.72 (C-6), 151.67 (C-2), 164.68 (C-4); MS m/z 304(M^+); Anal. Calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_4$, C, 63.14; H, 6.62; N, 9.20. Found : C, 63.17; H, 6.61; N, 9.21.

Method B:

Compound **11a** (0.63g; 5 mmol) was silylated as described above to give **12a** as an oil after the evaporation of excess HMDS. **12a** was dissolved in dry 1,2-dichloroethane (20 mL) and to which was added chloromethylether **10** (5 mmol in 20 mL dichloroethane). The reaction mixture was cooled to 0°C and anhydrous aluminium chloride (0.69g; 5 mmol) was added. The reaction mixture was stirred at room temperature overnight, quenched with ice water and stirred for 30 min. The organic layer was separated and the aqueous layer was extracted with CHCl_3 (50 mL x 4). The combined organic layers were extracted with saturated sodium bicarbonate solution (50 mL x 6). The aqueous solution was immersed in ice-bath and carefully acidified with concentrated hydrochloride until the congo red paper change from red to blue, and then extracted with chloroform (100mL X 5). The organic solution was dried with anhydrous MgSO_4 and filtered. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography using CHCl_3 : MeOH (100 : 5) as an eluent to give the mixture of **14a** as major and **13a** as minor component. After recrystallization from CHCl_3 - Et_2O , pure **14a** (0.74 g, 60%) was obtained. The mother liquor was evaporated and the residue was recrystallized from ethanol to obtain pure **13a** (0.15 g, 10%).

The same procedures (method A) were adopted to convert each of the compounds **11b-d** to the respective **13b-d** as major ,and **14b-d** as minor compounds.

1-[(1-Benzyloxy-2-propoxy)methyl]-6-azathymine(**13b**) and 1-(Benzyloxymethyl)-6-azathymine (**14b**)

The alkylated product of **12b** was purified by silica gel column to give a mixture of **13b** and **14b** which was triturated with CHCl_3

to give **14b** as colorless crystals. Evaporation of the mother liquor gave **13b** (0.95 g, 62%) as a colorless oil; UV : HCl(0.1 M) λ_{max} 265 nm (ϵ 8100), NaOH(0.1 M) λ_{max} 251 nm (ϵ 6100); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 1.17 (d, J = 6.4 Hz, 3H, H-3'), 2.20(s, 3H, CH₃), 3.42-3.46(m, 2H, H-5'), 4.04(m, 1H, H-4'), 4.50 (s, 2H, benzylic H), 5.34, 5.42(dd, J = 10.6 Hz, 2H, H-1'), 7.29 (m, 5H, aromatic H), 10.36(br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 16.70 (C-3'), 17.91 (CH₃), 73.72 (C-5'), 74.77 (benzylic C), 75.37 (C-4'), 79.02 (C-1'), 128.00, 128.09, 128.86, 138.61, (aromatic C), 144.75 (C-5), 149.87 (C-2), 157.37 (C-4); MS m/z 305(M⁺); HR Mass for C₁₅H₁₉N₃O₄ Calcd : 305.1374. Found : 305.1378.

Pure **14b** (0.12 g, 10%) was obtained as colorless crystals; mp 125-127°C UV : HCl(0.1 M) λ_{max} 260 nm (ϵ 8500), NaOH(0.1 M) λ_{max} 251 nm (ϵ 6100); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 2.44 (s, 3H, CH₃), 4.92 (s, 2H, benzylic H), 5.59 (s, 2H, H-1'), 7.54 (m, 5H, Aromatic H), 10.22 (br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 16.72 (CH₃), 72.49 (benzylic C), 79.44 (C-1'), 128.22, 128.40, 128.86, 137.77 (aromatic C), 149.72 (C-2), 157.05 (C-4); MS m/z 247(M⁺); Anal. Calcd for C₁₂H₁₃N₃O₃ : C, 58.29; H, 5.30; N, 17.00; found: C, 58.06; H, 5.35; N, 16.88.

1-[(1-Benzylloxy-2-propoxy)methyl]uracil (**13c**) and 1-(Benzylloxymethyl)uracil (**14c**)

The alkylated product of **12c** was purified by silica gel column to give a mixture of **13c** and **14c** which was crystallized with EtOH to give **13c** (1.09 g, 75%) as colorless crystals. mp 68-70°C; UV : HCl(0.1 M) λ_{max} 259 nm (ϵ 8500), NaOH(0.1 M) λ_{max} 260 nm (ϵ 7000); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 1.16 (d, J = 6.4 Hz, 3H, H-3'), 3.43 (m, 2H, H-5'), 3.92 (m, 1H, H-4'), 4.51 (s, 2H, benzylic H), 5.20, 5.28 (dd, J = 10.5 Hz, 2H, H-1'), 5.68 (d, J = 8 Hz, 1H, H-5), 7.31 (m, 5H, aromatic H), 7.35 (d, J = 8 Hz, 1H, H-6), 9.71(br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 17.17 (C-3'), 73.39 (C-5'), 74.14 (benzylic C), 74.50 (C-4'), 75.72 (C-1'), 102.74 (C-5), 127.65, 127.78, 128.45, 137.93 (aromatic C), 143.37 (C-6), 150.90 (C-2), 163.32 (C-4); MS m/z 290(M⁺); Anal. Calcd for C₁₅H₁₈N₂O₄ : C, 62.06; H, 6.25; N, 9.65; found: C, 62.09; H, 6.20; N, 9.63.

Evaporation of the mother liquor to give a residual mass which was crystallized with CHCl₃-Et₂O to obtain pure **14c** (0.17 g, 15%)

as colorless crystals; mp 138-139°C, UV : HCl(0.1 M) λ_{max} 260 nm (ϵ 8500), NaOH(0.1 M) λ_{max} 260nm (ϵ 6500); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 4.63 (s, 2H, benzylic H), 5.24 (s, 2H, H-1'), 5.75 (d, J = 8 Hz, 1H, H-5), 7.30(d, J = 8 Hz, 1H, H-6), 7.31 (m, 5H, aromatic H), 9.10(br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 72.18 (benzylic C), 76.68 (C-1'), 103.72 (C-5), 128.43, 128.76, 129.08, 137.04 (aromatic C), 143.61 (C-6), 151.62 (C-2), 164.02 (C-4); MS m/z 232(M^+); Anal. Calcd for $\text{C}_{12}\text{H}_{12}\text{N}_2\text{O}_3$: C, 62.06; H, 5.21; N, 12.06; found: C, 61.77; H, 5.25; N, 12.19.

1-[(1-Benzyloxy-2-propoxy)methyl]-6-azauracil (13d) and 1-(Benzyloxymethyl)-6-azauracil (14d)

The alkylated product of **12d** was purified by silica gel column to give a mixture of **13d** and **14d** which was crystallized with $\text{CHCl}_3\text{-Et}_2\text{O}$ to give **14d** as colorless crystals. Evaporation of the mother liquor gave **13d** (1.02 g, 70%) as colorless oil; UV : HCl(0.1 M) λ_{max} 260 nm (ϵ 8100), NaOH(0.1 M) λ_{max} 251nm (ϵ 6700); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 1.17 (d, J = 6.5 Hz, 3H, H-3'), 3.43-3.46 (m, 2H, H-5'), 4.06 (m, 1H, H-4'), 4.50 (s, 2H, benzylic H), 5.34, 5.46 (dd, J = 10.6 Hz, 2H, H-1'), 7.30 (m, 5H, aromatic H), 7.37 (s, 1H, H-5), 10.02 (br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 17.88 (C-3'), 73.82 (C-5'), 74.81 (benzylic C), 75.62 (C-4'), 79.42 (C-1'), 128.10, 128.17, 128.89, 136.05 (aromatic C), 138.50 (C-6), 148.91 (C-2), 156.83 (C-4); MS m/z 291(M^+); HRMS for $\text{C}_{14}\text{H}_{17}\text{N}_3\text{O}_4$, Calcd : 291.1218. Found : 291.1212.

Pure **14d** (0.12 g, 10%) was obtained as colorless crystals; mp 92-94°C; UV : HCl(0.1 M) λ_{max} 260 nm (ϵ 8500), NaOH(0.1 M) λ_{max} 255nm (ϵ 6000); $^1\text{H-NMR}(\text{CDCl}_3)$ δ : 4.92 (s, 2H, benzylic H), 5.61 (s, 2H, H-1'), 7.54 (s, 1H, H-6), 7.62 (s, 5H, aromatic H), 9.57 (br. s. 1H, NH); $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ : 72.59 (benzylic C), 79.70 (C-1'), 128.22, 128.53, 128.94, 136.19 (aromatic C), 137.48 (C-5), 148.71 (C-2), 156.47 (C-4); MS m/z 233(M^+); Anal. Calcd for $\text{C}_{11}\text{H}_{11}\text{N}_3\text{O}_3$: C, 56.65; H, 4.75; N, 18.02; found: C, 56.44; H, 4.83; N, 17.74.

1-[(1-Hydroxy-2-propoxy)methyl]thymine (15a)

Compound **13a** (0.456 g; 1.5mmol), palladium(II) hydroxide (300 mg), cyclohexene (4 mL), an ethanol (16 mL) were refluxed for 4 h (monitored by TLC). The resulting solution was filtered and the filtrate evaporated to give a residual solid which was crystallized

with EtOH-Et₂O to afford pure **15a** (0.29 g 89%) as colorless crystals; mp 108 - 109 °C; UV : HCl(0.1M) λ_{max} 265 nm (ϵ 8300), NaOH(0.1M) λ_{max} 266nm (ϵ 6300); ¹H-NMR(CDCl₃) δ : 1.15 (d, J = 6.4 Hz, 3H, H-3'), 1.92 (d, J = 1.0 Hz, 3H, CH₃), 2.64 (br.s. 1H, OH), 3.46-3.65 (m, 2H, H-5'), 3.78-3.86 (m, 1H, H-4'), 5.16, 5.27 (dd, J = 10.3 Hz, 2H, H-1'), 7.18 (d, J = 1 Hz, 1H, H-6), 9.35 (br.s. 1H, NH); ¹³C-NMR(DMSO-d₆) δ : 12.79 (C-3'), 16.92 (CH₃), 66.81 (C-5'), 75.89 (C-1'), 76.16 (C-4'), 112.22 (C-5), 139.75 (C-6), 151.89 (C-2), 164.64 (C-4); MS m/z 214(M⁺); Anal. Calcd for C₉H₁₄N₂O₄: C 50.46, H 6.59, N 13.08. found : C, 50.24; H, 6.51; N, 13.06.

The same procedure was used to convert each of the compounds **13b-d** to the respective **15b-d**.

1-[(1-Hydroxy-2-propoxy)methyl]-6-azathymine (**15b**)

Pure **15b** (0.27 g 85%) was obtained as an oil; UV : HCl(0.1M) λ_{max} 253 nm (ϵ 5000), NaOH(0.1M) λ_{max} 265nm(ϵ 6300); ¹H-NMR(CDCl₃) δ : 1.10 (d, J = 6.2 Hz, 3H, H-3'), 2.20 (s, 3H, CH₃), 3.41-3.59 (m, 2H, H-5'), 3.83-3.91 (m, 1H, H-4'), 5.33 (s, 2H, H-1'); 10.85 (br.s. 1H, NH). ¹³C-NMR(DMSO-d₆) δ : 16.59 (C-3'), 16.69 (CH₃), 66.65 (C-5'), 76.37 (C-1'), 78.52 (C-4'), 145.15 (C-5), 150.07 (C-2), 157.51 (C-4); MS m/z 215(M⁺); HRMS for C₈H₁₃N₃O₄, Calcd : 215.0905. Found : 215.0902.

1-[(1-Hydroxy-2-propoxy)methyl]uracil (**15c**)

Pure **15c** (0.27 g 85%) was obtained as colorless crystals; mp 97-98°C(EtOH-Et₂O); UV : HCl(0.1M) λ_{max} 259 nm (ϵ = 8700), NaOH(0.1M) λ_{max} 260nm (ϵ 6100); ¹H-NMR(CDCl₃) δ : 0.94 (d, J = 6.4 Hz, 3H, H-3'), 2.70 (br.s. 1H, OH), 3.22-3.39 (m, 2H, H-5'), 3.53-3.61 (m, 1H, H-4'), 4.99, 5.08(dd, J = 10.3 Hz, 2H, H-1'), 5.49 d, J = 7.88 Hz, 1H, H-5), 7.26 (d, J = 1 Hz, 1H, H-6), 10.51 (br.s. 1H, NH); ¹³C-NMR(DMSO-d₆) δ : 16.92 (CH₃), 66.00 (C-5'), 75.52 (C-1'), 75.84 (C-4'), 102.74 (C-5), 144.04 (C-6), 151.62 (C-2), 164.42 (C-4); MS m/z 214(M⁺); Anal. Calcd for C₉H₁₄N₂O₄: C 48.00, H 6.04, N 13.99. found : C, 48.06; H, 6.10; N, 14.28.

1-[(1-Hydroxy-2-propoxy)methyl]-6-azauracil (**15d**)

Pure **15d** (0.27 g 89%) was obtained as an oil; UV : HCl(0.1M) λ_{max} 252 nm (ϵ 5400), NaOH(0.1M) λ_{max} 261nm (ϵ 4600); ¹H-

NMR(CDCl₃) δ : 1.12 (d, J = 6.3 Hz, 3H, H-3'), 3.42-3.60 (m, 2H, H-5'), 3.83-3.91 (m, 1H, H-4'), 5.36 (s, 2H, H-1'), 7.39 (s, 1H, H-5); ¹³C-NMR(DMSO-d₆) δ : 16.59 (CH₃), 66.58 (C-5'), 76.50 (C-1'), 78.81 (C-4'), 136.51 (C-5), 149.54 (C-2), 157.55 (C-4); MS m/z 201(M⁺); HRMS for C₇H₁₁N₃O₄, Calcd : 201.0749. Found : 201.0743.

ANTIVIRAL SCREENING

Human T cell leukemia cell lines, MT-4 and MOLT-4 cells were used in this study. The reverse transcriptase assay was carried out as described by Sarin et al.²¹ The results indicated that the compounds (**15a-d**) described in this manuscript were inactive against HIV-1.

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REFERENCES

1. a). Horwitz, J. P.; Chua, J.; Noel, M. *J. Org. Chem.* **1964**, 29, 2076.
b). Chu, C. K.; Beach, J. W.; Ullas, G. V.; Kosugi, Y. *Tetrahedron Lett.* **1988**, 29, 5349.
2. Yarchoan, R.; Mitsuya, H.; Thomas R. V.; Pluda, J. M.; Haryman, N. R.; Perno, C.F.; Marczyk, K. S.; Allain, J. P.; Johns, D. G.; Broder, S. *Science*, **1989**, 245, 412.
3. Mitsuya, H.; Broder, S. *Proc. Natl. Acad. Sci. U.S.A.* **1986**, 83, 1911.
4. Hurn, D. M.; Okabe, M. *Chem. Rev.* **1992**, 92, 1745. and literatures cited therein.
5. Hamamoto, Y.; Nakashima, H.; Matsui, T.; Matsuta, A.; Ueda, T.; Yamamoto, N. *Antimicrob. Agents Chemother.* **1987**, 31, 907.
6. Shiragami, H.; Irie, Y.; Shirae, H.; Yokozaki, K.; Yasuda, N. *J. Org.Chem.* **1988**, 53, 5170.
7. Baba, M.; Pauwels, R.; Herdewijn, P.; De Clercq, E. ; Desmyter, J.; Vandeputte, M. *Biochem. Biophys. Res. Commun.* **1987**, 142, 128.
8. De Clercq, E. *J. Med. Chem.* **1986**, 29, 1561.

9. a). Matsuya, H.; Weinhold, K.J.; Furman, P. A.; St. Clair, M. H.; Lehrman, S. N.; Gallo, R. L.; Bolognesi, D. P.; Barry, D. W.; Broder, S. *Proc. Natl. Acad. Sci. U.S.A.* **1985**, 82, 7096. b). Fischl, M. A.; Richman, D. D.; Grieco, M. H.; Gottlieb, M. S.; Volberding, P. A.; Laskin, O. L.; Leedom, J. M.; Groopman, J. E.; Mildvan, D.; Schooley, R. T.; Jackson, G. G.; Durack, D. T.; King, D. *New Engl. J. Med.* **1987**, 317, 185.
10. Herdewijn, P.; De Clercq, E. *In Design of Anti-AIDS Drugs; Elsevier Science: Amsterdam*, **1990**; pp 141-171.
11. Chu, C. K.; Cutler, S. J. *J. Heterocyclic Chem.* **1986**, 23, 289.
12. Chu, C. K.; Bhadti, V. S.; Doboszewski, B.; Gu, Z. P.; Kosugi, Y.; Pullaiah, K. C.; Van Roey, P. *J. Org. Chem.* **1989**, 54, 2217.
13. Schaeffer, H. J.; Beauchamp, L.; de Miranda, P. Elion, G. B.; Bauer, D. J.; Collins, P. *Nature.* **1978**, 272, 583.
14. Ogilvie, K. K.; Cheriyan, U. O.; Radatus, B. K.; Smith, K. O.; Galloway, K. S.; Kennell, W. L. *Can. J. Chem.* **1982**, 60, 3005.
15. Miyasaka, T.; Tanaka, H.; Baba, M.; Hayakawa, H.; Walker, R. T.; Balzarini, J.; De Clercq, E. *J. Med. Chem.* **1989**, 32, 2507.
16. Trinh, M. C., Florent, J. C., Grierson, D. S., Monneret, C. *Tetrahedron Lett.* **1991**, 32, 1447.
17. a) Han, C. H.; Chen, Y. L.; Tzeng, C. C. *Nucleosides & Nucleotides* **1991**, 10, 1391. b) Lee, K. H.; Chen, Y. L.; Huang B. R.; Tzeng, C. C. *Nucleosides & Nucleotides* **1991**, 10, 1407. c). Wang, E. C.; Chen, H. Y.; Tzeng, C. C. *J. Chin. Chem. Soc* **1993**, 40, 73.
18. Bacon, J. R.; Collis, M. J. *Chem & Ind.* **1971**, 930.
19. Martin, J. C.; McGee, D. P. C.; Jeffrey, G. A.; Hobbs, D. W.; Smee, D. F.; Matthews, T. R.; Verheyden, J. P. H. *J. Med. Chem.* **1986**, 29, 1384.
20. Kim, Y. H.; Kim, J. Y. *Heterocycles*, **1988**, 27, 71.
21. Sarin, P. S.; Sun, D., Thomson, A.; Muller, W. E. *J. Natl. Cancer* **1987**, 78, 663.

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