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SYNTHESIS AND CONFORMATIONAL ANALYSIS OF 1,5-ANHYDRO-2,4-DIDEOXY-p-MANNITOL NUCLEOSIDES

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ABSTRACT: 1,5-Anhydro-4,6-*O*-benzylidene-D-glucitol was used as starting material for the synthesis of 1,5-anhydro-2,4-dideoxy-D-mannitol nucleosides with an adenine and uracil base moiety. The compound with a purine base was obtained by direct nucleophilic substitution of a triflate. The pyrimidine nucleoside could be obtained by epoxide opening followed by inversion of configuration at the 3'-position. Both nucleosides adopt a C1 conformation with an axial base moiety.

INTRODUCTION

Nucleosides with a six membered carbohydrate moiety have several interesting characteristics: a) when compared with natural furanose nucleosides, the presence of an additional carbon atom increases the number of substitution sites from 5 to 6; b) the molecule is mostly frozen in a C_1 or 1C conformation (limiting the possible conformations) so that conformational analysis is easier than with furanose nucleosides and, consequently, the modified nucleoside may be used as a model to investigate structure-function relationship; c) new nucleoside analogues may be obtained without anomeric centre but still having a ring oxygen atom and a 1,4-relationship between the nucleobase and the hydroxymethyl group. This latter arrangement of functional groups is found in anhydrohexitol nucleosides. This means that they are chemically and

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enzymatically more stable than natural nucleosides. Several of these 1,5-anhydrohexitol nucleosides have been found to possess antiviral activity. Some of them (1,5-anhydro-¹ 2,3-dideoxy-D-arabino-hexitol nucleosides with a 5-iodouracil¹ 1, 5-ethyluracil² 2, 5trifluoromethyluracil³ 3, 5-vinyluracil³ 4, 5-propynyluracil³ 5, 5-fluorocytosin² 6 and 2.6-diaminopurine² 7 base moiety in the 2-position) show potent antiherpes activity. Other congeners demonstrate either weak antiherpes acitivity [1,5-anhydro-2-(5iodouracil-1-yl)-2-deoxy-D-mannitol⁴ 8, 1,5-anhydro-2-(cytosin-1-yl)-2,3-dideoxy-Dribohexitol⁵ 9, (1RS,3SR,4RS)-9-[4-(hydroxymethyl)-3-hydroxycyclohexyl]guanine⁶ 10, 1,5-anhydro-2-(thymin-1-yl)-2,3-dideoxy-3-C-hydroxymethyl-D-mannitol⁷ 11] or weak anti-HIV activity (1,5-anhydro-3-ene-2,3,4-trideoxy-D-threo-hexitol with an adenine⁸ 12 and thymine 13 base moiety⁸). When considering the antiherpes (HSV-1) activity, it is of interest to mention that some of these nucleoside analogues (1-5, 11) show clearly different biological activity when evaluated against TK⁺ and TK⁻ strains of HSV-1, which indicates that their anti-HSV activity must rely on the virus encoded thymidine kinase (TK). This difference in biological activity is less pronounced with compounds 6, 7, 9 and 10, indicating that also other cellular/viral mechanisms may play a role in their mode of action. Structure-activity relationship studies, however, only make sense when a series of compounds are envisaged with similar mode of action. The most striking observation is that, till now, all active antiherpes agents have an axially oriented base moiety and an equatorially oriented hydroxymethyl group in a 1,4-relationship⁸. In this conformation the 1,5-anhydrohexitol nucleosides can be considered as mimics of furanose nucleosides in the 2'-exo/3'-endo conformation. This suggests that this conformation might be important for interaction of the nucleoside (or one of its metabolites) with one of the enzymes involved in its mode of action (i.e. kinases and/or In order to further explore this structure-activity relationship we polymerases). synthesized 1,5-anhydro-2,4-dideoxy-D-mannitol nucleosides 14a-b. These molecules lack the equatorial 4-hydroxylgroup, which is replaced by an equatorial 3hydroxylgroup. The gauche effect (O-C3-C2-N) should likewise be favourable for stabilizing the C_1 conformation and this may lead to biological activity. The synthesis of these compounds, additionally, allows us to evaluate the importance of the presence of the 4-hydroxyl group for biological activity.





8 X = I; Y = OH; Z = H11 X = CH₃; Y = CH₂OH; Z = H



12 B = adenin-9-yl **13** B = thymin-1-yl







10



14a B = adenin-9-yl **14b** B = uracil-1-yl



RESULTS AND DISCUSSIONS

(A) Synthesis

Recently, we have reported⁹ the synthesis of 1,5-anhydro-6-O-MMTr-3-O-TBDMS-2-(adenin-9-yl)-2-deoxy-D-mannitol. This compound is similar to 14a, but has an additional hydroxyl group at the 4'-position. For the synthesis of 1,5-anhydro-2,4dideoxy-D-mannitol type nucleosides it seems logical to consider the deoxygenation of the 4-position of the above mentioned 2-deoxy-D-mannitol compound. Thus, 1,5anhydro-6-O-MMTr-3-O-TBDMS-N⁶-benzoyl-2-(adenin-9-yl)-2-deoxy-D-mannitol was with treated phenoxythiocarbonyl chloride in the of N.Npresence dimethylaminopyridine (DMAP) in acetonitrile and, after usual work up, the crude reaction mixture was treated with n-tributyltin hydride in the presence of azobisisobutyronitrile (AIBN). The same reaction was repeated using pyridine both as base and solvent instead of DMAP and acetonitrile. Both deoxygenation reactions, however, led to an intractable mixture of compounds for which the reason is not totally clear. Therefore, we decided to use an alternate reaction scheme starting with another intermediate obtained during the synthesis of 1,5-anhydro-2-deoxy-D-mannitol nucleosides, i.e. 1,5-anhydro-4,6-O-benzylidene-2-O-TBDMS-D-glucitol 15⁹.

Treatment of **15** with pivaloyl chloride in pyridine at room temperature gave **16** in 94% yield. The 4,6-*O*-benzylidene group of **16** was removed by treatment of **16** with trifluoroacetic acid in dichloromethane. This acid treatment, however, also removed the 2-*O*-TBDMS protecting group, giving **17** in 73% yield. Under identical reaction conditions, the *O*-TBDMS group situated in β -position is stable as described previously⁹. The instability of the equatorially oriented 2-*O*-TBDMS group by its stereochemical environment. The primary hydroxyl group of **17** was protected with a monomethoxytrityl (MMTr) group upon treatment with MMTrCl in pyridine, yielding **18** in 76%. Treatment of **18** with phenoxythiocarbonyl chloride (1.2 equiv.) and DMAP in acetonitrile, followed by n-tributyltin hydride and AIBN in toluene at 110 °C, gave **19** in 41% yield. This two step reaction sequence was preferred over a multistep approach using complicated protection and deprotection strategies. At this stage of the reaction scheme, we tried to introduce the nucleobase in the 2-position of **19**. Therefore, compound **19** was treated with trifluoromethanesulphonic anhydride in the



i: Pivaloyl chloride, pyridine, 94%; ii: CF₃COOH, CH₂Cl₂, 73%; iii: MMTr chloride, pyridine, 76%; iv: PhOC(S)Cl, DMAP, CH₃CN; v: nBu₃SnH, AIBN, toluene, Δ , 41% (iv and v); vi: (CF₃SO₂)₂O, pyridine, CH₂Cl₂, vii: tetrabutylammonium salt of adenine, CH₂Cl₂, 28% of **22** and 48% of **23**, Ph=phenyl.

Scheme 1

presence of pyridine in dichloromethane at -5 °C, and, after usual work-up procedure⁹, crude **20** was reacted with tetrabutylammonium salt of adenine^{10,11} in dichloromethane. However, this reaction procedure resulted in the isolation of **22** and **23** in a combined yield of 76% for the two steps. The intermediate **21** may give rise to the formation of both compounds. No trace of the desired adenine derivative could be detected.

Apparently, the neighbouring group participation reaction leading to inversion of stereochemistry at position-2 is much faster than the intermolecular nucleophilic substitution reaction. Another protecting group strategy is necessary in order to obtain the 2,4-dideoxy-D-mannitol nucleosides. As shown in Scheme 2, we selected the

i: Pivaloyl chloride, pyridine, 98%; ii: 80% CF₃COOH, CH₂Cl₂, 95%; iii: MMTr chloride, pyridine, 90%; iv: PhOC(S)Cl, DMAP, CH₃CN; v: nBu₃SnH, AIBN, toluene, Δ , 64%; vi: aq. 1N NaOH, dioxane, 92%; vii: TBDMSCl, Imidazole, DMF, 60% of **30** and 18% of **31**; viii: (CF₃SO₂)₂O, pyridine, CH₂Cl₂; ix: tetrabutylammonium salt of adenine, CH₂Cl₂, 18%; x = aq. CF₃COOH, 49%. A = adenin-9-yl.

TBDMS group for protection of the 3-hydroxylgroup. This protecting group is less prone to migration and neighbouring participation reactions.

Treatment of compound 24^9 with pivaloyl chloride in pyridine gave 25 in 98% yield. The 4,6-O-benzylidene protecting group of 25 was removed upon treatment with trifluoroacetic acid in dichloromethane to afford 26 (95%). Treatment of 26 with MMTrCl in pyridine gave 27 in 90% yield. Deoxygenation of the 4-hydroxyl group of 27, via phenoxythiocarbonylation under Barton conditions as described above, yielded

28 in 64% yield (for two steps). Both pivaloyl groups of 28 were removed upon treatment with 1N NaOH in dioxane to give 29 in 92% yield. The 3-hydroxyl group of 29 can now be protected in a rather selective way. When 29 is treated with tbutyldimethylsilyl chloride in the presence of imidazole in DMF¹², 30 was obtained as major compound (60%) together with 31 (18%). The compounds 30 and 31 were differentiated by the use of NMR spectroscopy. For compound **30**, the signal of the OHproton, proved by exchange with D₂O, shows a crosspeak with the C1' signal in a longrange ¹H-¹³C heteronuclear correlation experiment (long-range Hetcor) which clearly indicates that the OH-group is positioned in the 2'-position. Activation of the 2hydroxyl group of 30 as 2-O-triflate, followed by reaction with the tetrabutylammonium salt of adenine^{10,11} in dichloromethane afforded 32 in 18% yield. TLC (CH₂Cl₂-MeOH 95:5) shows the presence of four other compounds in the reaction mixture, which were not identified. Finally, the 6-O-MMTr and 3-O-TBDMS protecting groups of 32 were simultaneously removed using aqueous trifluoroacetic acid, giving 1,5-anhydro-2-(adenin-9-yl)-2,4-dideoxy-D-mannitol 14a in 49% yield. For the preparation of uracil derivatives 14b and 38 (Scheme 3) we started with 31. Thus, 31 was treated with methanesulfonyl chloride (MsCl) in the presence of Et₃N in CH₂Cl₂ to afford 33 which after usual work up and treatment with tetrabutylammonium fluoride (TBAF) in tetrahydrofuran (THF) gave the epoxide 34 in 64% yield (for two steps). The epoxide 34 was treated with uracil in the presence of NaH in DMF to give 35 in 76% yield. The configuration at C3' was inverted using O² neighbouring group participation reaction⁴.

Treatment of **35** with MsCl in the presence of Et_3N in dichloromethane gave the corresponding mesylate **36** which was treated with aqueous 1 N NaOH in ethanol at 60 °C to afford **37** in 83% yield. The 6-*O*-MMTr group in **37** was removed upon treatment with aqueous 80% acetic acid to give **14b** (77%). Finally, **35** was treated with aqueous 80% acetic acid to afford **38** in 85% isolated yield. The determination of the antiviral activity of **14a**, **14b** and **38** is in progress.

(B) Determination of configuration and conformation of 14a, 14b and 38 by high resolution NMR spectroscopy.

The assignment of the resonances of the protons of **14a**, **14b** and **38** was made by using a combination of a DQF-COSY spectrum of **14a** and 1D ¹H NMR spectra of **14a**,

i: MsCl, Et₃N, CH₂Cl₂; ii: TBAF.3H₂O, THF, 64% for two steps; iii: sodium salt of uracil, DMF, 76%; iv: 1N NaOH, H₂O, EtOH, 83% for two steps; v: 80% CH₃COOH, H₂O, 85% for 38 and 77% for 14b.

Scheme 3

14b and 38 at 27 °C at 500 MHz. The ¹H chemical shifts are given in the experimental section.

The configuration and conformation of **14a**, **14b** was determined on the basis of their ${}^{3}J_{HH}$ values. For **14a** and **14b** following ${}^{3}J_{HH}$ values are obtained: (**14a**): ${}^{3}J_{1',2'} = 1.4$, ${}^{3}J_{1'',2'} = 2.3$, ${}^{3}J_{2',3'} = 5.0$, ${}^{3}J_{3',4''} = 12.0$, ${}^{3}J_{3',4''} = 5.0$, ${}^{3}J_{4'',5'} = 11.5$, ${}^{3}J_{4',5'} = 2.1$; (**14b**): ${}^{3}J_{1'',2'} = 1.3$, ${}^{3}J_{1'',2'} = 3.3$, ${}^{3}J_{2',3'} = 5.0$, ${}^{3}J_{3',4''} = 12.3$, ${}^{3}J_{3',4''} = 5.0$, ${}^{3}J_{4'',5'} = 11.5$, ${}^{3}J_{4',5'} = 2.1$. Inspection of these coupling constants reveals the following conclusions: i) the high values found for the vicinal coupling constants ${}^{3}J_{4'',5'} \approx 11$, ${}^{3}J_{3',4''} \approx 12$ implies that H3',

FIG. 2

H4" and H5' are axially oriented which is only possible when the OH group is β positioned and the conformation is C₁ as in A in FIG. 2; ii) the small coupling ${}^{3}J_{H2'-H3'} \approx$ 5 excludes that H2' and H3' are diaxially disposed which implies that the base is axial or in the β -position.

For compound **38**, there was severe overlap of the signals of H1', H1", H2', H3' and the signals of H6' and H6" resonate at the same chemical shift. The signals of H4' and H4" are separated and give a doublet of double doublet as pattern (${}^{3}J \cong 3$ Hz, 10 Hz, 13 Hz) and a double triplet (${}^{3}J \cong 2$ à 3 Hz (2 x), 13 Hz), respectively. Decoupling of H6' and H6" gives a double doublet for H5' with coupling constants of ${}^{3}J_{4',5'} \approx 10$ and ${}^{3}J_{4',5'} \approx 2$ à 3 which implies that H5' and H4' are diaxially disposed as in C₁ (FIG. 2, C).

The compounds **14a,b** and **38** adopt the C_1 conformation as is evident from their NMR spectra. In the light of our previous results this is not unexpected, especially because the base is in 1,3 position with respect to the ring oxygen and they experience then much less steric hindrance when in a axial position. Under these conditions the CH_2OH group will orient itself equatorially and thus determines the conformation as C_1

EXPERIMENTAL SECTION

The ¹H-NMR and ¹³C-NMR spectra were recorded on a Varian Gemini-200 spectrometer using tetramethylsilane as an internal standard. When DMSO- d_6 was used as solvent, the resonance peak at 2.50 ppm was used as internal standard (for ¹H-NMR) and 39.6 ppm (for ¹³C-NMR). (*s*: singlet, *d*: doublet, *dd*: doublet doublet, *ddd*: doublet of double doublet, *t*: triplet, *dt*: double triplet, *m*: multiplet and *br. s*: broad singlet). All chemical shifts are given in δ values. The ¹H and ¹³C NMR spectra of **14a**, **14b** and **38** in D₂O solution have been recorded on a Varian Unity 500 MHz spectrometer at 27 °C. The carbon assignments were based on ATP experiments or 2D ¹H-¹³C heteronuclear correlation experiments (HMQC and long-range Hetcor). The DQF-COSY of **14a** was recorded at 27 °C using 256 experiments of 2K complex data points and a relaxation delay of 1.5 s. The HMQC of **14a** and the long-range Hetcor of **30** were recorded with 128 increments in t₁, 2K data points in t₂. High resolution mass spectra (HRMS) were recorded on a Kratos Concept 1H mass spectrometer. The solvent was concentrated in vacuo. All other technical data were identical to those previously described^{5,9}.

1,5-Anhydro-4,6-O-benzylidene-3-O-pivaloyl-2-O-TBDMS-D-glucitol (16). To a cold solution (ice-water) of **15**⁹ (2.0 g, 5.46 mmol) in pyridine (30 mL), pivaloyl chloride (1.34 mL, 10.92 mmol) was added and the reaction mixture was kept at room temperature for seven days. A cold solution of saturated aqueous sodium bicarbonate (100 mL) was added slowly to the reaction mixture. The reaction mixture was extracted with CH₂Cl₂ (2 x 50 mL). The combined organic layers were washed with water (10 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel column chromatography (0-6% EtOAc in hexane) to give **16** (2.3 g, 94%). ¹H-NMR (DMSO-d₆): 7.38 (s, 5H) arom; 5.60 (s, 1H) benzylidene; 5.05 (t, J_{2.3} \cong J_{3.4} \cong 9.2 Hz, 1H) H3; 4.23 (dd, J = 4.6, 9.8 Hz, 1H); 3.88 (m, 2H); 3.73-3.36 (m, 5H); 1.14 (s, 9H) piv; 0.81 (s, 9H) TBDMS; 0.07 (s, 3H) TBDMS; 0.04 (s, 3H) TBDMS. ¹³C-NMR (DMSO-d₆): 176.9, 137.5, 128.8, 128.1, 125.8, 100.0, 78.8, 75.3, 70.5, 70.0, 69.3, 67.9, 27.1, 25.6, 17.7, -4.3, -5.0 (the quaternary carbon of the piv. group was hidden under the solvent peak). HRMS calcd. for C₂₄H₃₉O₆Si (M+H)⁺ 451.2515, found 451.2535.

1,5-Anhydro-3-O-pivaloyl-D-glucitol (17). Compound **16** (1.9 g, 4.22 mmol) was treated with CF_3COOH (40 mL) in CH_2Cl_2 (40 mL) at room temperature for 24 h.

The solvent was removed, the residue was dissolved in MeOH (20 mL) and treated with NH₄OH (4 mL). After concentration the residue was purified by silica gel column chromatography (0-6% MeOH in CH₂Cl₂) to give 17 (760 mg, 73%). ¹H-NMR (DMSO-*d*₆): 5.15 (d, J = 1.9 Hz, 1H); 4.95 (d, J = 0.7 Hz, 1H); 4.68 (t, J = 9.0 Hz, 1H); 4.65 (t, J = 5.9 Hz, 1H); 3.73 (dd, J = 5.3, 10.8 Hz, 1H); 3.62 (ddd, J = 1.8, 5.8, 11.8 Hz, 1H); 3.40 (m, 2H); 3.28-3.0 (m, 3H); 1.16 (s, 9H). ¹³C-NMR (DMSO-*d*₆): 177.1, 81.6, 79.4, 69.6, 68.1 (two C), 61.1, 38.8, 27.2. HRMS calcd. for C₁₁H₂₁O₆ (M+H)⁺ 249.1338, found 249.1334.

1,5-Anhydro-6-*O***-MMTr-3-***O***-pivaloyl-D-glucitol (18)**. Compound **17** (750 mg, 3.02 mmol) was treated with MMTrCl (1.11 g, 3.62 mmol) in pyridine (10 mL) at room temperature overnight. A cold solution of saturated aqueous sodium bicarbonate (10 mL) was added and the reaction mixture was extracted with CH_2Cl_2 (2 x 50 mL). The combined organic layer was washed with H_2O (10 mL), dried over Na_2SO_4 , filtered and concentrated. The residue was purified by silica gel column chromatography (0-1.5% MeOH in CH_2Cl_2) to give **18** (1.2 g, 76%). ¹H-NMR (CDCl₃): 7.50-6.78 (m, 14H) arom; 4.71 (t, J \cong 9.1 Hz, 1H) H3; 4.05 (dd, J = 5.6, 11.3 Hz, 1H) H5; 3.79 (s, 3H) OCH₃; 3.78-3.51 (m, 2H) H2, H1'; 3.41-3.18 (m, 2H) H1", H4; 2.78 (d, 1H) H6'; 2.67 (d, 1H) H6"; 1.21 (s, 9H). ¹³C-NMR (CDCl₃): 180.9, 158.6, 144.0, 135.1, 130.3, 128.3, 127.9, 127.0, 113.2, 86.9, 81.0, 78.8, 70.5, 69.9, 69.2, 64.3, 55.2, 39.0, 27.1. HRMS calcd. for $C_{31}H_{36}O_7Na$ (M+Na)⁺ 543.2358, found 543.2354.

1,5-Anhydro-6-O-MMTr-3-O-pivaloyl-4-deoxy-D-glucitol (19). To a cold (icewater) solution of **18** (3.0 g, 5.76 mmol) in acetonitrile was added DMAP (1.4 g, 11.52 mmol) followed by slow addition of phenoxythionocarbonyl chloride (955 μ L, 6.91 mmol). The reaction mixture was kept at room temperature for 24 h, and after being concentrated the residue was dissolved in CH₂Cl₂ (100 mL), and washed successively with saturated aqueous NH₄Cl solution (4 x 15 mL) and H₂O (10 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated. The crude residue was dissolved in toluene (50 mL) and treated with tributyltin hydride (3.09 mL, 11.52 mmol) and AIBN (236 mg, 1.44 mmol) at 110 °C overnight. The reaction mixture was concentrated and the residue was purified by silica gel column chromatography (0-6% EtOAc in hexane) to give **19** (1.20 g, 41% in two steps). ¹H-NMR (CDCl₃): 7.50-6.78 (m, 14H) arom; 4.28 (m, J_{2,3} = 14.2, J_{3,4'} = 9.0, J_{3,4''} = 5.3 Hz, 1H) H3; 3.97 (ddd, J = 1.5, 4.9, 11.8 Hz, 1H) H5; 3.79 (s, 3H) OCH₃; 3.65-3.43 (m, 2H) H2, H1'; 3.38 (m, 3H) H1", H6' and H6"; 2.68 (d, $J_{2,OH} = 3.1$ Hz, 1H) 2OH; 1.98 (m, $J_{3,4'} = 9.0$, $J_{4',5} = 11.8$ Hz, 1H) H4'; 1.65 (m, $J_{3,4''} = 5.3$ Hz, $J_{4',5'} = 4.9$, $J_{4',4''} = 12.7$ Hz, 1H) H4"; 1.20 (s, 9H) piv. ¹³C-NMR (CDCl₃): 177.7, 158.5, 144.2, 135.3, 130.3, 128.3, 127.8, 126.9, 113.1, 86.6, 77.7, 72.0, 67.2, 65.8, 63.5, 55.1, 38.8, 27.1, 25.7. HRMS calcd. for $C_{31}H_{36}O_6Na$ (M+Na)⁺ 527.2409, found 527.2385.

1,5-Anhydro-6-O-MMTr-3-O-pivaloyl-4-deoxy-D-mannitol (22) and 1,5-Anhydro-6-O-MMTr-2-O-pivaloyl-4-deoxy-D-mannitol (23). To a cold (-5 °C) solution of 19 (504 mg, 1.0 mmol) in CH₂Cl₂ (10 mL) was added pyridine (171 µL) followed by slow addition of trifluoromethanesulfonic anhydride (252 µL in CH₂Cl₂, 10 μ L) under nitrogen atmosphere. The reaction mixture was kept at -5 °C for 2 h, quenched with ice-water and diluted by addition of CH₂Cl₂ (30 mL). The two layers were separated. The aqueous layer was extracted with CH_2Cl_2 (2 x 10 mL). The combined organic layers was washed successively with pre-cooled (-5 °C) saturated Na_2HPO_4 (3 x 10 mL) and H_2O (10 mL). The organic layer was dried over MgSO₄, filtered and concentrated at 16 °C. This crude material was dissolved in CH₂Cl₂ (2 mL) and added to a solution of tetrabutylammonium salt of adenine in CH_2Cl_2 (40 mL). The reaction mixture was kept at room temperature for 24 h. The precipitate was filtered off and washed with CH₂Cl₂. The combined filtrate was concentrated and the residue purified by silica gel column chromatography (0-30% EtOAc in hexane) to give 22 (140 mg, 28%) and 23 (240 mg, 48%) in a total yield of 76% for two steps. Compound 22, ¹H-NMR (CDCl₃): 7.50-6.78 (m, 14H) arom; 4.80 (ddd, $J_{2,3} = 2.6$, $J_{3,4'} = 4.9$, $J_{3,4''} = 4.9$ 11.7 Hz, 1H) H3; 4.11-3.94 (m, 2H) H2, H5; 3.79 (s, 3H) OCH₃; 3.58-3.28 (m, 4H) H6', H6", H1' and H1"; 2.19 (d, J_{2.0H} = 4.8 Hz, 1H) 2OH; 2.05 (m, 1H) H4', 1.68 (m, 1H) H4"; 1.21 (s, 9H) piv. ¹³C-NMR (CDCl₃); 177.7, 158.5, 144.2, 135.3, 130.3, 128.3, 127.8, 126.8, 113.1, 86.6, 77.7, 72.0, 67.2, 65.8, 63.5, 55.1, 38.8, 27.1, 25.7. HRMS calcd. for $C_{31}H_{36}O_6Na$ (M+Na)⁺ 527.2409, found 527.2395. Compound 23, ¹H-NMR (CDCl₃): 7.48-6.76 (m, 14H) arom; 5.30 (m, 1H); 4.96 (m, 2H); 3.79 (s, 3H); 3.65-3.40 (m, 2H), 3.28 (dd, J = 5.9, 9.1 Hz, 1H); 3.05 (dd, J = 9.0, 7.2 Hz, 1H); 2.25 (d, J = 2.8Hz, 1H); 1.93-1.63 (m, 2H); 1.10 (s, 9H). ¹³C-NMR (CDCl₃): 179.7, 158.6, 144.0, 135.4, 130.3, 128.4, 127.8, 126.9, 113.0, 86.5, 76.4, 70.0, 69.7, 66.0, 62.4, 55.2, 38.4, 29.3, 27.0. HRMS calcd. for $C_{31}H_{36}O_6Na (M+Na)^+$ 527.2409, found 527.2391.

1,5-Anhydro-4,6-O-benzylidene-2,3-O-bis-pivaloyl-D-glucitol (25). To a cold solution (ice-water) of **24** (10.31 g, 40.91 mmol) in pyridine (200 mL) was added pivaloyl chloride (15.12 mL, 122.76 mmol) and the reaction mixture was kept at room temperature for 7 days. A cold solution of saturated aqueous NaHCO₃ (200 mL) was added slowly to the reaction mixture and the mixture was extracted with CH₂Cl₂ (3 x 100 mL). The combined organic layers were washed with H₂O (25 mL), dried over Na₂SO₄, filtered, concentrated to give pure **25** (17.18 g, 98%). ¹H-NMR (CDCl₃): 7.49-7.32 (m, 5H) arom; 5.51 (s, 1H) benzylidene; 5.37 (t, J = 9.4 Hz, 1H); 5.05 (m, 1H); 4.35 (dd, J = 4.9, 10.4 Hz, 1H); 4.11 (dd, J = 5.8, 11.0 Hz, 1H); 3.78-3.59 (m, 2H); 3.53-3.31 (m, 2H); 1.17 (s, 9H); 1.16 (s, 9H). ¹³C-NMR (CDCl₃): 177.5, 177.2, 137.0, 128.9, 128.2, 125.9, 101.2, 79.2, 72.0, 71.5, 69.3, 68.2, 67.6, 38.8 (two C), 27.1 (two C). HRMS calcd. for C₂₃H₃₃O₇ (M+H)⁺ 421.2226, found 421.2225.

1,5-Anhydro-2,3-*O***-bis-pivaloyl-D-glucitol (26)**. Compound **25** (12.0 g, 28.6 mmol) was treated with CF₃COOH (100 mL) in CH₂Cl₂ (100 mL) at room temperature for 48 h. The reaction mixture was concentrated, the residue dissolved in MeOH (60 mL) and treated with concentrated NH₄OH (6 mL). The residue, after concentration, was purified by silica gel column chromatography (0-50% EtOAc in hexane) to give **26** (9.0 g, 95%). ¹H-NMR (CDCl₃): 5.15 - 4.80 (m, 2H) H2 H3; 4.07 (dd, $J_{1',2} = 5.3$, $J_{1',1''} = 11.3$, 1H)H1'; 3.92 (dd, $J_{5,6'} = 3.3$, $J_{6',6''} = 11.9$ Hz, 1H) H6'; 3.78 (dd, $J_{5,6''} = 4.6$, $J_{6',6''} = 11.9$, 1H) H6''; 3.67 (t, $J_{4',5} \cong J_{3,4'} \cong 9.5$, 1H) H4'; 3.45 - 3.20 (m, 2H) H5, H1'', 2.62 (br. s, 2H) 4OH and 6OH; 1.20 (s, 9H) piv; 1.15 (s, 9H) piv. ¹³C-NMR (CDCl₃): 179.5, 177.4, 80.4, 76.7, 70.0, 68.7, 66.7, 62.4, 39.0, 38.8, 27.1 (two C). HRMS calcd. for $C_{16}H_{29}O_7$ (M+H)⁺ 333.1913, found 333.1922.

1,5-Anhydro-6-O-MMTr-2,3-O-bis-pivaloyl-D-glucitol (27). To a solution of **26** (10.24 g, 30.96 mmol) in pyridine (150 mL) was added MMTrCl (11.46 g, 37.14 mmol) and the reaction mixture was kept at room temperature overnight. A cold solution of saturated aqueous NaHCO₃ (200 mL) was added slowly to the reaction mixture and the mixture was extracted with CH₂Cl₂ (3 x 200 mL). The combined organic layer was washed with H₂O (30 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel column chromatography (0-15% EtOAc in hexane) to give **27** (16.8 g, 90%). ¹H-NMR (CDCl₃): 7.50-6.78 (m, 14H) arom; 5.15 – 4.80 (m, 2H) H2 H3; 4.95 (m, 1H); 4.08 (dd, $J_{1,2} = 5.4$, $J_{1,17} = 11.2$ Hz, 1H) H1'; 3.78 (s, 3H) OCH₃;

3.62 (m, 1H) H4; 3.38 (m, 3H) H5, H6', H6"; 3.23 (t, $J_{1",2} = J_{1',1"} = 11.2$, 1H) H1"; 2.53 (d, $J_{4,OH} = 4.0$, 1H) 4OH; 1.21 (s, 9H) Piv; 1.16 (s, 9H) piv. ¹³C-NMR (CDCl₃): 178.8, 177.4, 158.7, 144.1, 135.3, 130.4, 128.4, 127.9, 127.0, 113.3, 87.0, 79.2, 76.1, 71.2, 68.8, 66.8, 64.1, 55.2, 38.9, 38.8, 27.1 (two C). HRMS calcd. for C₃₆H₄₄O₈Na (M+Na)⁺ 627.2934, found 627.2943.

1,5-Anhydro-6-O-MMTr-2,3-O-bis-pivaloyl-4-deoxy-D-glucitol (28). To a cold (ice-water) solution of 27 (7.0 g, 11.58 mmol) in acetonitrile (100 mL) was added DMAP (3.39 g, 27.79 mmol) followed by slow addition of phenoxythiocarbonyl chloride (1.92 mL, 13.89 mmol) and the reaction mixture was kept at room temperature for 48 h. The reaction mixture was concentrated and the residue dissolved in CH_2Cl_2 (200 mL), washed successively with saturated aqueous solution of NH_4Cl (5 x 20 mL) and H₂O (20 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated. The crude residue was dissolved in dry toluene (60 mL) and treated with n-tributyltin hydride (6.22 mL, 23.16 mmol) in the presence of AIBN (480 mg, 2.89 mmol) at 110 °C for 60 min. The reaction mixture was concentrated and the residue was purified by silica gel column chromatography (0-2.5% EtOAc in hexane) to give 28 (4.34 g, 64% in two steps). ¹H-NMR (CDCl₃): 7.50-6.78 (m, 14H) arom; 5.01 (m, 1H) H3; 4.90 (m, 1H) H2; 4.08 (dd, J_{1',2} = 5.2, J_{1',1"} = 11.1 Hz, 1H) H1'; 3.79 (s, 3H) OCH₃; 3.62 (m, 1H) H5; 3.23 (m, 2H) H6', H1"; 3.04 (dd, $J_{5,6"} = 4.7$, $J_{6',6"} = 9.7$ Hz, 1H) H6"; 2.08 (ddd, $J_{4',5} =$ 1.8, $J_{3,4'} = 4.7$, $J_{4',4''} = 12.5$, 1H) H4'; 1.48 (m, 1H) H4''. 1.18 (s, 18H) 2 x piv. ¹³C-NMR (CDCl₃): 177.7, 177.5, 158.6, 144.3, 135.5, 130.3, 128.4, 127.8, 126.9, 113.1, 86.3, 75.5, 71.4, 69.7, 67.0, 66.0, 55.2, 38.7 (two C), 33.4, 27.1 (two C). HRMS calcd. for $C_{36}H_{44}O_7Na (M+Na)^+ 611.2984$, found 611.2975.

1,5-Anhydro-6-*O***-MMTr-4-deoxy-D-glucitol (29)**. Compound **28** (3.3 g, 5.6 mmol) was treated with aqueous (1N) NaOH (50 mL) in dioxane (50 mL) at 55 °C for 24 h. The pH of the reaction mixture was adjusted to 7.0 by slow addition (in ice-water) of aqueous HCl. The volume of the reaction mixture was reduced to 1/3 of it's original volume and extracted with EtOAc (3 x 100 mL). The combined organic layers were concentrated and the residue was purified by silica gel column chromatography (0-3% MeOH in CH₂Cl₂) to give **29** (2.15 g, 92%). ¹H-NMR (CDCl₃): 7.49-6.78 (m, 14H); 3.98 (dd, $J_{1',2} = 4.7$, $J_{1',1''} = 11.0$ Hz, 1H) H1'; 3.79 (s, 3H) OCH₃; 3.65-3.40 (m, 3H) H2, H3, H5; 3.26-2.97 (m, 3H) H1'', H6', H6''; 2.07 (ddd, J = 2.0, 4.2, 12.5 Hz, 1H) H4';

1.43 (m, 1H) H4". ¹³C-NMR (CDCl₃): 158.5, 144.4, 135.5, 130.4, 128.4, 127.8, 126.9, 113.0, 86.3, 75.8, 73.4, 72.3, 69.7, 66.0, 55.2, 36.0. HRMS calcd. for $C_{26}H_{28}O_5Na$ (M+Na)⁺ 443.1834, found 443.1827.

1,5-Anhydro-6-O-MMTr-2-O-TBDMS-4-deoxy-D-glucitol (31)and 1.5-Anhydro-6-O-MMTr-3-O-TBDMS-4-deoxy-D-glucitol (30). To a solution of 29 (2.15 g, 5.1 mmol) in DMF (30 mL), imidazole (416 mg, 6.12 mmol) and TBDMSCI (845 mg, 5.61 mmol) were added in two portions at an interval of 10 h. The reaction mixture was kept at room temperature for 20 h. The reaction mixture was concentrated, the residue was dissolved in CH₂Cl₂ (100 mL), washed successively with saturated aqueous solution of NH₄Cl (3 x 15 mL) and H₂O (10 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was purified by silica gel column chromatography (0-15% EtOAc in hexane) to give **31** (500 mg, 18%) and **30** (1.65 g, 60%). This reaction was also performed using pyridine both as a base and solvent. Thus, to a solution of 29 (2.96 g, 7.03 mmol) in pyridine (35 mL) was added TBDMSCI (1.16 g, 7.73 mmol) in two portions at an interval of 10 h and kept at room temperature for 40 h. The reaction mixture was quenched with cold aqueous saturated NaHCO₃ and extracted with CH₂Cl₂ (3 x 50 mL). The organic layer was concentrated, coevaporated with toluene and the residue was purified by silica gel column chromatography (0-15% ethyl acetate in hexane) to give **30** (2.0 g, 53%) and **31** (1.0 g, 27%). Compound **31**, ¹H-NMR (CDCl₃) 7.50-6.80 (m, 14H) arom; 3.88 (dd, $J_{1',2} = 4.6$, $J_{1',1''} = 11.2$, 1H) H1'; 3.78 (s, 3H) OCH₃; 3.70-3.40 (m, 3H) H2, H3, H5; 3.24-2.99 (m, 3H) H1", H6', H6"; 2.28 (br. s, 1H) 3OH; 2.08 (dd, J = 4.0, 12.0 Hz, 1H) H4'; 1.45 (apparent dt, $J \cong 12$, 1H) H4". ¹³C-NMR (CDCl₃) 158.6, 144.5, 135.6, 130.4, 128.5, 127.8, 126.9, 113.0, 86.2 (MMTr); 76.4 (C5); 73.6 (C2, C3); 70.1 (C1); 66.0 (C6); 55.1 (OCH₃); 35.1 (C4); 25.7, 17.9, -4.5, -4.8 (TBDMS). HRMS calcd. for C₃₂H₄₂O₅SiNa (M+Na)⁺ 557.2699, found 557.2679. Compound 30, ¹H-NMR (CDCl₃) 7.50-6.80 (m, 14H) arom; 4.05 (dd, $J_{1',2} =$ 4.8, $J_{1',1'} = 12.0$ Hz, 1H) H1'; 3.78 (s, 3H) OCH₃; 3.62-3.40 (m, 3H) H2, H3, H5; 3.30-3.10 (m, 2H) H1", H6'; 3.00 (dd, J = 4.0, J = 13.7 Hz, 1H) H6", 2.20 (d, 1H) OH (exchanges with D₂O and shows long-range coupling with C1' in a long-range Hetcor experiment), 1.90 (dd, J = 4.0, 12.0 Hz, 1H) H4'; 1.26 (apparent dt, J ≅ 12 Hz, 1H) H4". ¹³C-NMR (CDCl₃) 158.6, 144.6, 135.7, 130.4, 128.5, 127.8, 126.9, 113.0, 86.2 (MMTr); 75.8 (C5); 74.8 (C3); 72.3 (C2); 69.3 (C1); 66.2 (C6); 55.1 (OCH₃); 36.9

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(C4); 25.7, 18.0, -4.2, -4.6 (TBDMS). HRMS calcd. for $C_{32}H_{42}O_5SiNa$ (M+Na)⁺ 557.2699, found 557.2681.

1,5-Anhydro-6-O-MMTr-3-O-TBDMS-2-(adenin-9-yl)-2,4-dideoxy-D-

mannitol (32). The *O*-triflate of **30** was prepared following identical reaction conditions as described for **20** using **30** (535 mg, 1.0 mmol), trifluoromethanesulfonic anhydride (252 μ L in 10 μ L CH₂Cl₂) in the presence of pyridine (171 μ L) in CH₂Cl₂ (10 mL). The crude triflate was dissolved in CH₂Cl₂ (5 mL) and added to a solution of tetrabutylammonium salt of adenine in CH₂Cl₂ (40 mL). The reaction mixture was kept at room temperature for 48 h. The precipitate was filtered, and washed with CH₂Cl₂. The combined filtrate was concentrated and the residue purified by silica gel column chromatography (0-4% MeOH in CH₂Cl₂) to give **32** (120 mg, 18% overall yield). ¹H-NMR (CDCl₃) 8.30 (s, 1H) H8/H2; 8.25 (s, 1H) H2/H8; 7.50-6.80 (m, 14H) arom; 5.70 (br. s, 2H) NH2; 4.90 (m, 1H) H2; 4.40-3.20 (m, 9H) H1',H1", H3, H5, H6', H6" and OCH₃; 1.70 (m, 2H) H4', H4". ¹³C-NMR (CDCl₃) 158.7, 155.4 (C6), 152.8 (C2), 152.0 (C4), 144.4, 141.2 (C8), 135.5,130.4, 128.5, 127.9, 127.0, 119.5 (C5), 113.2, 86.5, 76.0, 68.8, 68.1, 66.2, 55.2, 53.3, 33.2, 25.3, -5.0. HRMS calcd. for C₃₇H₄₅N₅O₄SiNa (M+Na)⁺ 674.3138, found 674.3099.

1,5-Anhydro-2'-(adenin-9-yl)-2,4-dideoxy-D-mannitol (14a). Compound **32** (100 mg, 0.15 mmol) was treated with aqueous 80% CF₃COOH (10 mL) at room temperature overnight. The reaction mixture was concentrated. The residue was dissolved in MeOH (10 mL) and treated with NH₄OH (2 mL), concentrated and crystallized from MeOH to give **14a** (20 mg, 49%). ¹H-NMR (D₂O) 8.51 (s, 1H) H2/H8; 8.15 (s, 1H) H2/H8; 4.84 (m, 1H) H2'; 4.31 (dt, $J_{2',3'} = J_{3',4'} = 5.0, J_{3',4''} = 12.0, 1H)$ H3'; 4.22 (dd, $J_{1',2'} = 1.4, J_{1',1''} = 13.2, 1H)$ H1'; 3.99 (dd, $J_{1'',2'} = 2.3, J_{1',1''} = 13.2, 1H)$ H1'; 3.75-3.68 (m, 2H) H5', H6'; 3.63 (dd, $J_{5',6''} = 6.5, J_{6',6''} = 12.4, 1H)$ H6''; 1.83-1.70 (m, after decoupling of H2', multiplet becomes ddd, $J_{4',5'} \cong 2.1, J_{3',4''} = 5.0, J_{4',4''} \cong 13.2$ HZ, 1H) H4'; 1.49 (apparent dt, $J_{3'4''} = 12.1, J_{4'',5'} = 11.5, J_{4',4''} \cong 13.2, 1H)$ H4''. ¹³C-NMR (D₂O) 156.4 (C6), 153.2 (C2), 151.2 (C4), 142.6 (C8), 119.0 (C5), 77.8 (C5'), 68.8 (C1'), 67.5 (C3'), 64.7 (C6'), 54.2 (C2'), 30.7 (C4'). HRMS calcd. for C₁₁H₁₆N₅O₃ (M+H)⁺ 266.1253, found 266.1297. Elem. anal. calcd. for C₁₁H₁₅N₅O_{3.2}H₂O: C: 43.83, H: 6.36, N: 23.25. Found C: 43.75, H : 6.27, N: 23.17.

1,5;2,3-Di-anhydro-6-O-MMTr-D-allitol (34). To a cold (ice-water) solution of 31 (950 mg, 1.77 mmol) in CH₂Cl₂ (15 mL), Et₃N (368 μ L, 2.65 mmol) was added,

followed by MsCl (164 µL, 2.13 mmol) and the reaction mixture was kept at 5 °C for 2 h. The reaction mixture was quenched with cold aqueous saturated NaHCO₃ and after addition of CH₂Cl₂ (30 mL), the two layers were separated. The aqueous layer was reextracted with CH₂Cl₂ (2 x 10 mL). The combined organic layers were washed with water (1 x 10 mL), dried over Na₂SO₄, filtered and concentrated to give crude **33**. The crude residue was dissolved in THF (20 mL), TBAF.3H₂O (1.24 g, 3.54 mmol) was added and kept at 50 °C for 48 h. The solvent was removed and the residue was purified by silica gel column chromatography (0-10% ethyl acetate in hexane) to give **34** (460 mg, 64%). ¹H-NMR (CDCl₃) 7.50-6.80 (m, 14H) arom; 4.22 (dd, $J_{1',2} = 4.0$ Hz, $J_{1',1''} = 13.5$ Hz, 1H) H1'; 3.93 (d, $J_{1',1''} = 13.5$, 1H) H1''; 3.65-3.52 (m, 1H) H5; 3.40-3.30 (m, 1H) H3; 3.23 (t, $J_{1',2} \cong J_{2,3} = 4.0$, Hz, 1H) H2; 3.08 (dd, $J_{5,6''} = 5.9$, $J_{6',6''} = 9.7$, 1H) H6'; 2.94 (dd, $J_{5,6''} = 4.7$, $J_{6',6''} = 9.7$, 1H) H6''; 2.05 (m, 1H) H4'; 1.76 (ddd, J = 2.9, 11.1, 14.5, 1H) H4''. ¹³C-NMR (CDCl₃) 158.6, 144.5, 135.7, 130.4, 128.5, 127.8, 126.9, 113.0, 86.1, 69.6, 66.2, 65.7, 55.1, 51.2, 50.7, 28.2. HRMS calcd. for C₂₆H₂₆O₄Na (M+Na)⁺ 425.1729, found 425.1771.

1,5-Anhydro-6-O-MMTr-2-(uracil-1-yl)-2,4-dideoxy-D-altritol (35). A mixture of uracil (233 mg, 2.08 mmol) and NaH 80% in oil (125 mg, 4.16 mmol) in DMF (10 mL) was heated at 100 °C for 60 min. To that mixture 34 (420 mg, 1.04 mmol) in DMF (5 mL) was added and kept at 120 °C for 48 h. The reaction mixture was cooled to room temperature and concentrated. A mixture of ethyl acetate (50 mL) and H_2O (10 mL) was added and the two layers were separated. The aqueous layer was extracted with ethyl acetate (2 x 30 mL). The combined organic layer was dried over Na₂SO₄, filtered, concentrated and purified by silica gel column chromatography (0-2.5% MeOH in CH₂Cl₂) to give **35** (410 mg, 76%). ¹H-NMR (CDCl₃) 10.25 (s, 1H) NH; 8.44 (d, J_{5,6} = 8.4Hz, 1H) H6; 7.50-6.80 (m, 14H) arom; 5.78 (d, J_{5,6} = 8.4 Hz, 1H) H5; 4.42 (m, 1H) H1'; 4.16 (m, 4H) H1", H2', H3', H5'; 3.78 (s, 3H) OCH₃; 3.24 (dd, 1H) H6'; 3.03 (dd, 1H) H6"; 1.90 (t, 1H) H4'; 1.62 (m, 2H) H4", 3'OH. ¹³C-NMR (CDCl₃) 163.6, 158.7, 151.8, 144.5, 143.8, 135.5, 130.5, 128.4, 127.9, 127.0, 113.1, 102.6, 86.2, 70.9, 65.9, 64.6, 64.0, 55.2, 54.7, 29.5. HRMS calcd. for C₃₀H₂₉N₂O₆Na₂ $(M+2Na-H)^+$ 559.1821, found 559.1844.

1,5-Anhydro-6-O-MMTr-2-(uracil-1-yl)-2,4-dideoxy-D-mannitol (37). To a cold (ice-water) solution of **35** (230 mg, 0.45 mmol) in CH_2Cl_2 (10 mL) was added Et_3N

(93 μ L, 0.67 mmol) followed by addition of MsCl (42 μ L, 0.54 mmol) and the mixture was kept at 5 °C for 2 h. The reaction mixture was guenched with cold aqueous saturated NaHCO₃. After addition of CH₂Cl₂ (20 mL), the two layers were separated. The aqueous layer was reextracted with CH_2Cl_2 (2 x 20 mL). The combined organic layers were washed with H_2O (10 mL), dried over Na_2SO_4 , filtered and concentrated to give crude 36. Crude 36 in EtOH (20 mL) was heated until it became a clear solution and treated with aqueous (1N) NaOH (3 mL) at 60 °C for 5 h. The reaction mixture was cooled to 5 °C and the pH was adjusted to 7.0 by addition of dil. HCl. The reaction mixture was concentrated, the residue dissolved in CH2Cl2 (50 mL), washed with aqueous saturated NaHCO₃ (10 mL), H₂O (10 mL), dried, filtered and concentrated. The residue was purified by silica gel column chromatography (0-2.5% MeOH in CH₂Cl₂) to give **37** (190 mg, 83%). ¹H-NMR (CDCl₃) 9.46 (br. s, 1H) NH; 8.43 (d, J_{5.6} = 8.4 Hz, 1H); 7.50-6.80 (m, 14H) arom; 5.71 (d, J_{5.6} = 8.4 Hz, 1H) H5; 4.78 (m, 1H) H2'; 4.26 (d, $J_{1',1''} = 13.7$ Hz, 1H) H1'; 4.25-4.18 (m, 1H) H3'; 3.90 (dd, $J_{1'',2'} = 3.0$, $J_{1',1''} = 13.7$ Hz, 1H) H1"; 3.78 (s, 3H) OCH₃; 3.65-3.60 (m, 1H) H5'; 3.50 (s, 1H) 3'OH; 3.33 (dd, $J_{5',6'} = 3.7$, $J_{6',6''} = 9.9$ Hz, 1H) H6'; 3.17 (dd, $J_{5',6''} = 3.8$, $J_{6',6''} = 9.9$ Hz, 1H) H6"; 1.85-1.80 (m, 2H) H4', H4". ¹³C-NMR (CDCl₃) 163.2 (C4); 158.7 (MMTr); 153.0 (C2); 144.3 (MMTr); 144.1 (C6); 135.3, 130.4, 128.4, 127.8, 127.0, 113.1 (MMTr); 102.4 (C5); 86.4 (MMTr); 76.0 (C5'); 68.5 (C1'); 67.4 (C3'); 65.7 (C6'); 55.2 (OCH3); 53.3 (C2'); 30.4 (C4'). HRMS calcd. for $C_{30}H_{29}N_2O_6Na_2$ (M+2Na-H)⁺ 559.1821, found 559.1802.

1,5-Anhydro-2-(uracil-1-yl)-2,4-dideoxy-D-altritol (38). Compound **35** (100 mg, 0.19 mmol) was treated with aqueous 80% acetic acid (10 mL) at room temperature overnight. The solvent was removed, coevaporated with methanol and toluene. The residue was purified by silica gel column chromatography (0-7% MeOH in CH₂Cl₂) to give **38** (40 mg, 85%). ¹H-NMR (D₂O) 8.24 (d, J_{5,6} = 8.0 Hz, 1H) H6; 5.88 (d, J_{5,6} = 8.0 Hz, 1H) H5; 4.28-4.16 (m, 4H) H1', H1", H2' and H3'; 3.95-4.05 (m, 1H) H5'; 3.70 (d, 2H) H6', H6"; 1.88-1.80 (ddd, J_{3',4'} = 3.2, J_{4',5'} = 10.1, J_{4'4"} = 13.0 Hz, 1H) H4'; 1.73 (dt, ³J \cong 2 à 3 Hz, J_{4'4"} = 13 Hz, 1H) H4". ¹³C-NMR (D₂O) 167.1 (C4); 153.0 (C2), 145.6 (C6), 102.3 (C5), 73.0 (C5'), 64.7 (C1'), 64.1 (C3'), 63.5 (C6'), 55.7 (C2'), 30.1 (C4'). HRMS calcd. for C₁₀H₁₅N₂O₅ (M+H)⁺ 243.0980, found 243.1008. Elem. anal. calcd. for C₁₀H₁₄N₂O₅: C: 49.58, H: 5.83, N: 11.56. Found C: 49.51, H: 5.97, N: 11.62.

1,5-Anhydro-2'-(uracil-1-yl)-2,4-dideoxy-D-mannitol (14b). Compound 37 (165 mg, 0.32 mmol) was treated with aqueous 80% CH₃COOH (10 mL) at room temperature overnight. The reaction mixture was concentrated, coevaporated with methanol and toluene. The residue was purified by silica gel column chromatography (0-7% MeOH in CH₂Cl₂) to give **14b** (60 mg, 77%). ¹H-NMR (D₂O) 8.42 (d, J_{5,6} = 8.2 Hz, 1H) H6; 5.88 (d, J_{5,6} = 8.2 Hz, 1H) H5; 4.88 (m, 1H) H2'; 4.36 (dt, J_{2',3'} = J_{3',4'} = 5.0, J_{3',4''} = 12.3 Hz, 1H) H3'; 4.33 (dd, J_{1',2'} = 1.3, J_{1',1''} = 13.6 Hz, 1H) H1'; 4.05 (dd, J_{1'',2'} = 3.3, J_{1',1''} = 13.6 Hz, 1H) H1'; 3.80-3.75 (m, 2H) H5', H6'; 3.70 (dd, J_{5',6''} = 6.8, J_{6',6''} = 12.7 Hz, 1H) H6''; 1.86 (m, after decoupling of H2', multiplet becomes ddd, J_{4',5'} \cong 2.1, J_{3',4'} = 5.0, J_{4',4''} \cong 13.3 Hz, 1H) H4''. ¹³C-NMR (D₂O) 166.9 (C4), 154.0 (C2), 146.6 (C6), 102.0 (C5), 77.6 (C5'), 68.0 (C1'), 67.5 (C3'), 64.6 (C6'), 53.1 (C2'), 30.3 (C4'). HRMS calcd. for C₁₀H₁₅N₂O₅ (M + H)⁺ 243.0980, found 243.1041.

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